

# Lab Log

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**18/10/2017**

- Autoclaved the soil again
- Autoclaved 1 L of DI millicule water
- Autoclaved 1 L of 0.6% soft agar
- Autoclaved a spoon to weigh out soil
- Got some *SBW25* from Floh. Around 1 mL at  $7.26 \times 10^8$ .
  - Grow up overnight
  - Transfer 60  $\mu$ L of bacteria and 10  $\mu$ L of phage (around  $7 \times 10^6$  pfus) into 6 mL of KB agar
  - Should give a concentration of around  $10^8$  phage/mL
  - Done this in triplicate
- Grow up *lacZ* and *WT* strains overnight. Should give concentration of around  $\sim 10^8$  cells in 60  $\mu$ L.
- Do these in triplicate
- Added 60  $\mu$ L of frozen overnight culture from first experiment (18/08/2017 *lacZ* and *WT*)

**Retrospectively work out density of the overnight stocks and phage**

**19/10/2017**

- Put 80g of soil into each 10cm x 10cm microcosm
  - Used autoclaved spoon
  - Placed scale in laminar flow hood (cleaned with ethanol before and after)
- Placed 5 mL ( $\sim 200$   $\mu$ L per microcosm) of *lacZ* and *WT* into separate 12 mL centrifuge tubes
  - Centrifuged for 15 minutes at max speed ( $\sim 4500$  r.p.m) on big centrifuge
  - Want to get to 5 mL per microcosm for inoculating ( $\sim 125$  mL in total)
  - Resuspended pellet into 2250  $\mu$ L, vortexed and placed 620  $\mu$ L, 620  $\mu$ L and 810  $\mu$ L into three different falcon tubes
  - Filled these three falcon tubes up to 40 mL, 40 mL and 45 mL respectively
    - \* This guaranteed the same concentration of sample in each falcon tube
  - Placed 5 mL of *lacZ* or *WT* strain into each microcosm
  - Froze ( $-80$   $^{\circ}$ C) 900  $\mu$ L of inoculate in 900  $\mu$ L of glycerol (25% final concentration)
- In the no phage treatments, we added 5 mL of M9
- Added 5 mL of phage to phage treatments
  - Place 900  $\mu$ L of bacteria + phage into three centrifuge tubes
  - Add 100  $\mu$ L (10%) chloroform into each tube (under fume hood)
  - Vortex rigorously
  - Centrifuge for 2-3 minutes at full speed (minifuge)
  - Take out supernatant and placed in a single tube (took out 800  $\mu$ L of each tube)
  - Put 40 mL of M9 into 6 tubes
  - Added 400  $\mu$ L into each tube ( $\sim 100$  fold dilution from the initial stock)
  - Shake each tube and add 5 mL into each microcosm
- Place microcosms into the 26  $^{\circ}$ C incubator (Level 1 incubator room)

**22/10/2017**

- setup 2 *WT* microcosms up for spot assays of phage
- used a crystal from  $T_0$
- autoclaved 2 L of KB agar. Put in small autoclave

**23/10/2017**

- phage spot test 10<sub>-1</sub> to 10<sub>-8</sub>
  - Floh's phage ( $7.26 \times 10^8$ )
  - Phage we inoculated with
  - Extra phage tube (should have lower concentration)
  - blank plate
- plated T<sub>0</sub> of WT and *lacZ P. fluorescens*

**24/10/2017**

- phage spot test was no good (streaky)
- likely that we did not wait long enough for the spot to dry
- set up more T<sub>0</sub> bacteria overnight

**25/10/2017**

- phage spot assay again
- plating only 4 spots per plate instead of 8
- counted T<sub>0</sub> counts (plated 30 µl)
  - *lacZ* @ 10<sup>-4</sup>: 18
  - WT @ 10<sup>-4</sup>: 64

**26/10/2017**

- check phage spot, 10<sup>-6</sup> looks to be the correct dilution
- further counts are done by putting the 10 µl phage into the 1% bacteria soft agar
  - vortex and plate
  - leave to dry
  - incubate overnight

**30/10/2017**

- grow up some T<sub>0</sub> bacteria overnight

**31/10/2017**

- serial dilution of phage to 10<sup>-6</sup>
- 50 µl bacteria + 10 µl phage + 5 ml soft agar
  - vortex
  - pour
- set samples up for inoculated phage, floh's phage stock and our phage dregs (each in triplicate)
- in 28 °C incubator overnight

**01/11/2017**

- looked at phage spots (10<sup>-6</sup> dilution)
- calculated how many phage are there

$$\text{Phage concentration} = \frac{\text{number of plaques}}{\text{dilution factor} \times \text{volume added}}$$

Volume added is almost always 10 µl. I wrote a mini function to calculate this

```
d <- data.frame(dregs = c(3,4,6), phage = c(6,2,4))

# back calculate number in there
# PFU/ml = plaque number / (dil fac * volume added)
plaq_num <- function(x, dilfac){return(x/(dilfac*0.01))}

d <- dplyr::mutate_at(d, c('dregs', 'phage'), plaq_num, dilfac = 10^-6)

knitr::kable(d)
```

dregs	phage
3e+08	6e+08
4e+08	2e+08
6e+08	4e+08

```
# concentration of phage added
mean(d$phage)
```

```
## [1] 4e+08
```

```
# concentration of other phage
mean(d$dregs)
```

```
## [1] 433333333
```

- sampled all soil microcosms (after 13 days)
  - ~2g of soil into a 12 mL centrifuge tube (with ~ 7 glass beads)
    - \* sample using the big end of a 5 mL pipette and weigh on scales under hood
  - add 10 mL of M9
  - vortex for ~ 1 minute
  - add 900 µl of sample to 900 µl of 50% glycerol and freeze at -80 °C (bacteria + phage)
  - make phage suspension for each tube
    - \* add 900 µl of sample to centrifuge tube
    - \* add 100 µl of chloroform under fume hood
    - \* vortex
    - \* centrifuge at full speed for 4 minutes
    - \* put supernatant into separate tube
      - samples 1-17 800 µl, samples 18-48 750 µl
- names of samples
- 1-12: WT no phage
- 13-24: *lacZ* no phage
- 25-36: WT + phage
- 37-48: *lacZ* + phage
- Autoclaved things
  - 2 empty 500 mL bottles
  - M9 salts x10 (500 mL)
    - \* converted the weight of  $Na_2HPO_4$  to the amount of  $Na_2HPO_4 \cdot 7H_2O$  needed.
    - \* MW of  $Na_2HPO_4 \cdot 7H_2O$  is 268. MW of  $Na_2HPO_4$  is 142.
    - \* to convert from weight of  $Na_2HPO_4$  to  $Na_2HPO_4 \cdot 7H_2O$  is to multiply by  $\frac{268}{142}$

- 1 L of DI water
- 2 L hard agar
- centrifuge tubes.

sample	weight_g
1	2.1
2	2.0
3	1.9
4	2.0
5	2.0
6	2.0
7	2.0
8	2.0
9	2.1
10	1.9
11	2.2
12	2.1
13	2.1
14	1.9
15	2.2
16	2.2
17	1.9
18	2.2
19	2.0
20	2.3
21	2.3
22	2.2
23	1.9
24	2.1
25	2.1
26	2.2
27	1.9
28	2.0
29	2.2
30	2.3
31	2.0
32	2.2
33	2.2
34	2.2
35	2.0
36	2.0
37	2.3
38	1.9
39	2.2
40	1.9
41	2.2
42	2.0
43	2.0
44	2.2
45	2.1
46	2.2
47	2.2
48	2.1

**02/11/2017**

- set up phage spot tests of all soil phage suspensions against ancestral WT on soft agar.
  - just checking for presence/absence of phage
  - 5 mL per plate, 4 spots per plate, 12 spots in total
  - important to let the spot dry before moving
- poured plates (all Xgal)

**03/11/2017**

- all no phage treatments had no phage and all phage treatments had phage
- plated all the bacteria + phage treatments at  $10^{-4}$  and  $10^{-5}$  (30  $\mu$ l)
  - left on bench over the weekend

**06/11/2017**

- counted all the plates
  - all still had bacteria in and were not obviously contaminated (Yays)
- picked 20 colonies from each sample and placed in KB media in 48 well plates
  - 750  $\mu$ l of KB media
  - used matchsticks
- grown statically for 2 days at 26 °C

**08/11/2017**

- put 500  $\mu$ l of 50% glycerol into each well (final concentration 20%)
- froze in the -80 °C
- autoclaved
  - 2x 800 mL hard agar
  - 2x 800 mL soft agar

**13/11/2017**

- prep for sampling soil microcosms tomorrow. Labelled samples. Booked downstairs hood with power access.

**14/11/2017**

- sampled all 48 microcosms
- froze 900  $\mu$ l of bacteria + phage in 900  $\mu$ l of 50% glycerol (final concentration equals 25%)
- phage sample in fridge
  - 1-12: 750  $\mu$ l
  - 13-48: 600  $\mu$ l
- setup soft agar plates of ancestral bacteria and did phage spot assays

sample	weight_g
1	2.2
2	2.2
3	2.3
4	2.1

sample	weight_g
5	2.2
6	2.1
7	2.1
8	2.0
9	2.0
10	2.0
11	2.2
12	2.1
13	2.0
14	2.2
15	2.2
16	2.0
17	2.1
18	2.3
19	2.0
20	2.0
21	2.1
22	2.0
23	2.1
24	2.2
25	2.0
26	2.0
27	2.2
28	2.0
29	2.0
30	2.0
31	2.3
32	2.1
33	2.2
34	2.2
35	2.1
36	2.1
37	2.1
38	2.3
39	2.0
40	2.0
41	2.3
42	2.0
43	2.2
44	2.2
45	2.0
46	2.2
47	2.0
48	2.0

**15/11/2017**

- all phageless samples are still phageless, all phage samples still contain phage

16/11/2017

- plated all T<sub>2</sub> samples
  - 10<sup>-3</sup> and 10<sup>-4</sup>
  - 30 µl of on plate
- **used undiluted M9 x10 on Tuesday!!!**
- made up 400 mL of M9 salts x10 for autoclave

20/11/2017

- counted plates.
  - Most are ok at 10<sup>-3</sup>. Counts were much lower. Want to be certain that the reduction in abundance is not due to using concentrated M9.
- full sample again, T<sub>3</sub>.
  - Took 700 µL samples of phage
- setup overnight stock of ancestral bacteria
- autoclaved 2400 mL of KB agar

sample	weight_g
1	2.0
2	2.0
3	1.9
4	2.3
5	2.0
6	2.2
7	2.0
8	2.3
9	2.0
10	2.1
11	2.2
12	2.1
13	2.0
14	2.1
15	2.0
16	2.3
17	2.3
18	2.1
19	2.2
20	2.0
21	2.1
22	2.1
23	2.1
24	2.2
25	2.1
26	2.0
27	2.2
28	2.1
29	2.1
30	2.0
31	2.2
32	2.0
33	2.1
34	2.1

sample	weight_g
35	2.2
36	2.2
37	2.0
38	2.1
39	2.1
40	2.0
41	2.1
42	2.2
43	2.0
44	2.3
45	2.2
46	2.1
47	2.1
48	2.0

### 21/11/2017

- plated all 48 T<sub>3</sub> samples at 10<sup>-2</sup> and 10<sup>-3</sup>
- re-plated all T<sub>2</sub> replicates that did not contain > 20 colonies at 10<sup>-2</sup>
- poured ~150 Xgal plates. Stored in the cold room
- autoclaved two boxes of ependorfs
- put beads in Virkon to clean
- phage spot assay on phage extractions from T<sub>3</sub>. Used ancestral bacteria as the bacterial lawn

### 23/11/2017

- all T<sub>3</sub> samples were ok (> 30 colonies at 10<sup>-3</sup>) apart from 12 and 37
  - 12 and 37 were re-plated at 10<sup>0</sup> and 10<sup>-1</sup>
- picked 20 colonies from each sample and placed in KB media in 48 well plates
  - 750 µl of KB media
  - used matchsticks
- grown statically for 2 days at 26 °C
- Checked phage spot assays

### 25/11/2017

- picked colonies from 12 (10<sup>0</sup>) and 37 (10<sup>-1</sup>)
  - incubate for two days
- put 500 µl of 50% glycerol into each well of the other replicates that had been growing for two days (final concentration 20%)
- froze in the -80 °C

### 27/11/2017

- autoclaved 4L of KB hard agar



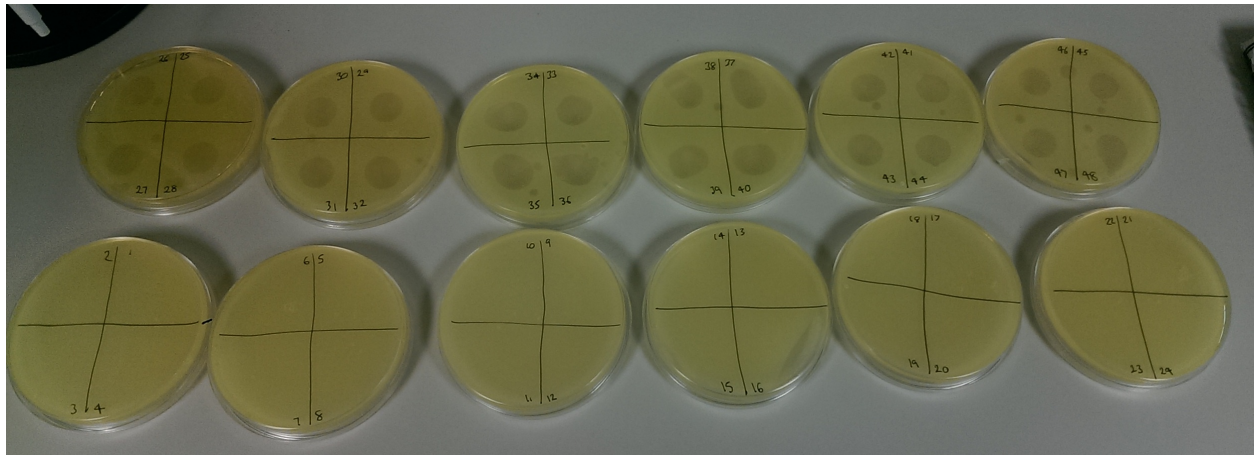


Figure 1: **Figure 1. phage spot assays are on point**

**28/11/2017**

- poured 2L of KB agar into square plates (Xgal)
- put 180  $\mu$ l of KB media into 96 well plates
- grew T<sub>F</sub> clones up overnight from frozen isolates (phage present microcosms only, 25-48)
  - placed in 28 °C incubator

**29/11/2017**

- look at coevolution of phage and host using streak plates
- streak each clone against contemporary phage, ancestral phage, a paired WT phage and a paired *lacZ* phage
- spot 30  $\mu$ l of each phage onto a square plate, let run down the plate then let it dry
- 1 plate per microcosm
- streak clones across the line of phage using matchsticks and the template

**30/11/2017**

- looked like it didnt work
- everything grew everywhere, even the controls
- set up an ancestral overnight

**01/12/2017**

- set up some phage spot assays of random phage against the ancestral phage
- did some streak assays

**02/12/2017**

- phage streak worked
- all phage spots worked
- **current hypothesis of problem is that we did not mix the phage properly**

**05/12/2017**

- grew up all clones overnight
- poured lots of square plates

**06/12/2017**

- did another phage streak day

**08/12/2017**

- counted the phage streaks. Processed these but they do not look as expected.
- autoclaved 4 L of hard KB agar
- autoclaved 1 L of KB media

**10/12/2017**

- setup isolated clones overnight in 180  $\mu$ l of KB media
- poured many square plates
- amplifying  $T_F$  phage in ancestral WT overnight
  - 60  $\mu$ l of ancestor and 10  $\mu$ l phage
- setup some phage spot assays (P.27, P.38, P.41)
  - spotted 10  $\mu$ l from  $10^{-1}$  to  $10^{-8}$

**11/12/2017**

- Unfortunately some of the lids of the phage amplifications had come off
- Reset these today
- Plated all the phage with ancestral bacteria at  $10^{-3}$

**12/12/2017**

- Counted phage

**13/12/2017**

- Did phage streaks using comb

**14/12/2017**

- Marked phage streaks