Lab Log

Daniel Padfield

18/10/2017

- Autoclaved the soil again
- Autoclaved 1 L of DI millicule water
- Autoclaved 1 L of 0.6% soft agar
- Autoclaved a spoon to weigh out soil
- Got some $SBW25\phi2$ from Floh. Around 1 mL at 7.26 x 10^8 .
 - Grow up overnight
 - Transfer 60 μl of bacteria and 10 μl of phage (around 7 x 10 6 pfus) into 6 mL of KB agar
 - Should give a concentration of around 10⁸ phage/mL
 - Done this in triplicate
- Grow up lacZ and WT strains overnight. Should give concentration of around ~10⁸ cells in 60 µl.
- Do these in triplicate
- Added 60 μL of frozen overnight culture from first experiment (18/08/2017 lacZ and WT)

Retrospectively work out density of the overnight stocks and phage

19/10/2017

- Put 80g of soil into each 10cm x 10cm microcosm
 - Used autoclaved spoon
 - Placed scale in laminar flow hood (cleaned with ethanol before and after)
- Placed 5 mL ($\sim 200~\mu$ l per microcosm) of lacZ and WT into separate 12 mL centrifuge tubes
 - Centrifuged for 15 minutes at max speed (~4500 r.p.m) on big centrifuge
 - Want to get to 5 mL per microcosm for inoculating (~ 125 mL in total)
 - Resuspended pellet into 2250 μ l, vortexed and placed 620 μ l, 620 μ l and 810 μ l into three different falcon tubes
 - Filled these three falcon tubes up to 40 mL, 40 mL and 45 mL respectively
 - * This guaranteed the same concentration of sample in each falcon tube
 - Placed 5 mL of lacZ or WT strain into each microcosm
 - Froze (-80 °C) 900 μl of inoculate in 900 μl of glycerol (25% final concentration)
- In the no phage treatments, we added 5 mL of M9
- Added 5 mL of phage to phage treatments
 - Place 900 μl of bacteria + phage into three centrifuge tubes
 - Add 100 ul (10%) chloroform into each tube (under fume hood)
 - Vortex rigorously
 - Centrifuge for 2-3 minutes at full speed (minifuge)
 - Take out supernatant and placed in a single tube (took out 800 μl of each tube)
 - Put 40 mL of M9 into 6 tubes
 - Added 400 µl into each tube (~100 fold dilution from the initial stock)
 - Shake each tube and add 5 mL into each microcosm
- Place microcosms into the 26 °C incubator (Level 1 incubator room)

22/10/2017

- setup 2 WT microcosms up for spot assays of phage
- used a crystal from T₀
- autoclaved 2 L of KB agar. Put in small autoclave

23/10/2017

- $\bullet~$ phage spot test $10_{\text{-}1}$ to $10_{\text{-}8}$
 - Floh's phage (7.26×10^8)
 - Phage we inoculated with
 - Extra phage tube (should have lower concentration)
 - blank plate
- plated T_0 of WT and $lacZ\ P.\ fluorescens$

24/10/2017

- phage spot test was no good (streaky)
- likely that we did not wait long enough for the spot to dry
- set up more T_0 bacteria overnight

25/10/2017

- phage spot assay again
- plating only 4 spots per plate instead of 8
- counted T_0 counts (plated 30 μ l)
 - $-\ lac Z$ @ 10^{-4} : 18
 - − WT @ 10^{-4} : 64