

Lab Log

Daniel Padfield

18/10/2017

- Autoclaved the soil again
- Autoclaved 1 L of DI millicule water
- Autoclaved 1 L of 0.6% soft agar
- Autoclaved a spoon to weigh out soil
- Got some *SBW25* from Floh. Around 1 mL at 7.26×10^8 .
 - Grow up overnight
 - Transfer 60 μ L of bacteria and 10 μ L of phage (around 7×10^6 pfus) into 6 mL of KB agar
 - Should give a concentration of around 10^8 phage/mL
 - Done this in triplicate
- Grow up *lacZ* and *WT* strains overnight. Should give concentration of around $\sim 10^8$ cells in 60 μ L.
- Do these in triplicate
- Added 60 μ L of frozen overnight culture from first experiment (18/08/2017 *lacZ* and *WT*)

Retrospectively work out density of the overnight stocks and phage

19/10/2017

- Put 80g of soil into each 10cm x 10cm microcosm
 - Used autoclaved spoon
 - Placed scale in laminar flow hood (cleaned with ethanol before and after)
- Placed 5 mL (~ 200 μ L per microcosm) of *lacZ* and *WT* into separate 12 mL centrifuge tubes
 - Centrifuged for 15 minutes at max speed (~ 4500 r.p.m) on big centrifuge
 - Want to get to 5 mL per microcosm for inoculating (~ 125 mL in total)
 - Resuspended pellet into 2250 μ L, vortexed and placed 620 μ L, 620 μ L and 810 μ L into three different falcon tubes
 - Filled these three falcon tubes up to 40 mL, 40 mL and 45 mL respectively
 - * This guaranteed the same concentration of sample in each falcon tube
 - Placed 5 mL of *lacZ* or *WT* strain into each microcosm
 - Froze (-80 $^{\circ}$ C) 900 μ L of inoculate in 900 μ L of glycerol (25% final concentration)
- In the no phage treatments, we added 5 mL of M9
- Added 5 mL of phage to phage treatments
 - Place 900 μ L of bacteria + phage into three centrifuge tubes
 - Add 100 μ L (10%) chloroform into each tube (under fume hood)
 - Vortex rigorously
 - Centrifuge for 2-3 minutes at full speed (minifuge)
 - Take out supernatant and placed in a single tube (took out 800 μ L of each tube)
 - Put 40 mL of M9 into 6 tubes
 - Added 400 μ L into each tube (~ 100 fold dilution from the initial stock)
 - Shake each tube and add 5 mL into each microcosm
- Place microcosms into the 26 $^{\circ}$ C incubator (Level 1 incubator room)

22/10/2017

- setup 2 *WT* microcosms up for spot assays of phage
- used a crystal from T_0
- autoclaved 2 L of KB agar. Put in small autoclave

23/10/2017

- phage spot test 10₋₁ to 10₋₈
 - Floh's phage (7.26×10^8)
 - Phage we inoculated with
 - Extra phage tube (should have lower concentration)
 - blank plate
- plated T₀ of WT and *lacZ P. fluorescens*

24/10/2017

- phage spot test was no good (streaky)
- likely that we did not wait long enough for the spot to dry
- set up more T₀ bacteria overnight

25/10/2017

- phage spot assay again
- plating only 4 spots per plate instead of 8
- counted T₀ counts (plated 30 µl)
 - *lacZ* @ 10⁻⁴: 18
 - WT @ 10⁻⁴: 64

26/10/2017

- check phage spot, 10⁻⁶ looks to be the correct dilution
- further counts are done by putting the 10 µl phage into the 1% bacteria soft agar
 - vortex and plate
 - leave to dry
 - incubate overnight

30/10/2017

- grow up some T₀ bacteria overnight

31/10/2017

- serial dilution of phage to 10⁻⁶
- 50 µl bacteria + 10 µl phage + 5 ml soft agar
 - vortex
 - pour
- set samples up for inoculated phage, floh's phage stock and our phage dregs (each in triplicate)
- in 28 °C incubator overnight

01/11/2017

- looked at phage spots (10⁻⁶ dilution)
- calculated how many phage are there

$$\text{Phage concentration} = \frac{\text{number of plaques}}{\text{dilution factor} \times \text{volume added}}$$

Volume added is almost always 10 µl. I wrote a mini function to calculate this

```
d <- data.frame(dregs = c(3,4,6), phage = c(6,2,4))

# back calculate number in there
# PFU/ml = plaque number / (dil fac * volume added)
plaq_num <- function(x, dilfac){return(x/(dilfac*0.01))}

d <- dplyr::mutate_at(d, c('dregs', 'phage'), plaq_num, dilfac = 10^-6)

knitr::kable(d)
```

dregs	phage
3e+08	6e+08
4e+08	2e+08
6e+08	4e+08

```
# concentration of phage added
mean(d$phage)
```

```
## [1] 4e+08
```

```
# concentration of other phage
mean(d$dregs)
```

```
## [1] 433333333
```

- sampled all soil microcosms (after 13 days)
 - ~2g of soil into a 12 mL centrifuge tube (with ~ 7 glass beads)
 - * sample using the big end of a 5 mL pipette and weigh on scales under hood
 - add 10 mL of M9
 - vortex for ~ 1 minute
 - add 900 µl of sample to 900 µl of 50% glycerol and freeze at -80 °C (bacteria + phage)
 - make phage suspension for each tube
 - * add 900 µl of sample to centrifuge tube
 - * add 100 µl of chloroform under fume hood
 - * vortex
 - * centrifuge at full speed for 4 minutes
 - * put supernatant into separate tube
 - samples 1-17 800 µl, samples 18-48 750 µl
- names of samples
- 1-12: WT no phage
- 13-24: *lacZ* no phage
- 25-36: WT + phage
- 37-48: *lacZ* + phage
- Autoclaved things
 - 2 empty 500 mL bottles
 - M9 salts x10 (500 mL)
 - * converted the weight of Na_2HPO_4 to the amount of $Na_2HPO_4 \cdot 7H_2O$ needed.
 - * MW of $Na_2HPO_4 \cdot 7H_2O$ is 268. MW of Na_2HPO_4 is 142.
 - * to convert from weight of Na_2HPO_4 to $Na_2HPO_4 \cdot 7H_2O$ is to multiply by $\frac{268}{142}$

- 1 L of DI water
- 2 L hard agar
- centrifuge tubes.

sample	weight_g
1	2.1
2	2.0
3	1.9
4	2.0
5	2.0
6	2.0
7	2.0
8	2.0
9	2.1
10	1.9
11	2.2
12	2.1
13	2.1
14	1.9
15	2.2
16	2.2
17	1.9
18	2.2
19	2.0
20	2.3
21	2.3
22	2.2
23	1.9
24	2.1
25	2.1
26	2.2
27	1.9
28	2.0
29	2.2
30	2.3
31	2.0
32	2.2
33	2.2
34	2.2
35	2.0
36	2.0
37	2.3
38	1.9
39	2.2
40	1.9
41	2.2
42	2.0
43	2.0
44	2.2
45	2.1
46	2.2
47	2.2
48	2.1

02/11/2017

- set up phage spot tests of all soil phage suspensions against ancestral WT on soft agar.
 - just checking for presence/absence of phage
 - 5 mL per plate, 4 spots per plate, 12 spots in total
 - important to let the spot dry before moving
- poured plates (all Xgal)

03/11/2017

- all no phage treatments had no phage and all phage treatments had phage
- plated all the bacteria + phage treatments at 10^{-4} and 10^{-5} (30 μ l)
 - left on bench over the weekend

06/11/2017

- counted all the plates
 - all still had bacteria in and were not obviously contaminated (Yays)
- picked 20 colonies from each sample and placed in KB media in 48 well plates
 - 750 μ l of KB media
 - used matchsticks
- grown statically for 2 days at 26 °C

08/11/2017

- put 500 μ l of 50% glycerol into each well (final concentration 20%)
- froze in the -80 °C
- autoclaved
 - 2x 800 mL hard agar
 - 2x 800 mL soft agar

13/11/2017

- prep for sampling soil microcosms tomorrow. Labelled samples. Booked downstairs hood with power access.

14/11/2017

- sampled all 48 microcosms
- froze 900 μ l of bacteria + phage in 900 μ l of 50% glycerol (final concentration equals 25%)
- phage sample in fridge
 - 1-12: 750 μ l
 - 13-48: 600 μ l
- setup soft agar plates of ancestral bacteria and did phage spot assays

sample	weight_g
1	2.2
2	2.2
3	2.3
4	2.1

sample	weight_g
5	2.2
6	2.1
7	2.1
8	2.0
9	2.0
10	2.0
11	2.2
12	2.1
13	2.0
14	2.2
15	2.2
16	2.0
17	2.1
18	2.3
19	2.0
20	2.0
21	2.1
22	2.0
23	2.1
24	2.2
25	2.0
26	2.0
27	2.2
28	2.0
29	2.0
30	2.0
31	2.3
32	2.1
33	2.2
34	2.2
35	2.1
36	2.1
37	2.1
38	2.3
39	2.0
40	2.0
41	2.3
42	2.0
43	2.2
44	2.2
45	2.0
46	2.2
47	2.0
48	2.0

15/11/2017

- all phageless samples are still phageless, all phage samples still contain phage

16/11/2017

- plated all T₂ samples
 - 10⁻³ and 10⁻⁴
 - 30 µl of on plate
- **used undiluted M9 x10 on Tuesday!!!**
- made up 400 mL of M9 salts x10 for autoclave

20/11/2017

- counted plates.
 - Most are ok at 10⁻³. Counts were much lower. Want to be certain that the reduction in abundance is not due to using concentrated M9.
- full sample again, T₃.
 - Took 700 µL samples of phage
- setup overnight stock of ancestral bacteria
- autoclaved 2400 mL of KB agar

21/11/2017

- plated all 48 T₃ samples at 10⁻² and 10⁻³
- re-plated all T₂ replicates that did not contain > 20 colonies at 10⁻²
- poured ~150 Xgal plates. Stored in the cold room
- autoclaved two boxes of ependorfs
- put beads in Virkon to clean
- phage spot assay on phage extractions from T₃. Used ancestral bacteria as the bacterial lawn