

Week 09: Comparative Methods

Due April 3rd

I. General

Type up all of your work in a text editor.

Before you begin, make a new folder in Tasks called Task_08, and save an empty file named task08.r in that folder.

You must also make a separate “Project” folder within Tasks. Inside your Project folder, make “code”, “data”, and “text” subfolders. By Friday, you should have some data in the data folder, some code to read the data in to R in the code folder, and a document with a *single sentence* describing your hypothesis.

When you’re **done** with this assignment, turn it in by (1) saving your text document, (2) opening your Terminal or GitBash, (3) navigating to the appropriate directory using `cd` and (4) typing:

```
git add -A (enter)
git commit -m "Task 08" (enter)
git push -u origin master (enter)
```

The main goal this week is a bit meta. Almost everyone is going to end up analyzing data along a phylogeny for their final project. So we’re going to walk through some functions that allow you to do that clearly and cleanly.

II. Your Code!

```
# We're going to unite data and phylogenies into one grand thing

# First, set your working directory to your working folder
# Now, we need to tell R that we want to use functions from the phytools
library
# Next, read the phylogeny into R using the read.tree() function (see the
last R assignment for how to do this)

plot(tree, type="fan")
# QUESTION 1: How many tips are there in the tree, and are branch lengths
present?

# Now, let's read in the data. It's in the file svl.csv. SVL stands for "
snout-vent length", and it's a standard measure of how long a vertebrate
is
data <- read.csv("https://jonsmitchell.com/data/svl.csv", stringsAsFactors=F
, row.names=1)

# QUESTION 2: What kind of object is "data"? What are its dimensions?
```

```

39
40 # We're going to convert the object data into a vector
41 svl <- setNames(data$svl, rownames(data))
42
43 # Now, we have an evolutionary tree of these lizards, and the body size of
44 the living species. Let's RECONSTRUCT the ancestral states. That is, let's
45 estimate how large the ancestors were using the phylogeny and the data.
46
47 Ancestors <- fastAnc(tree, svl, vars=TRUE, CI=TRUE)
48
49 # Examine the Ancestors object and the help file for fastAnc
50 # QUESTION 3: Where are the estimated values stored? What is the CI95
51 element?
52 # QUESTION 4: What are two assumptions made in the estimation of the
53 ancestral states using fastAnc?
54
55 # Let's plot all of this together!
56
57 # First, we'll plot the tree
58 par(mar=c(0.1,0.1,0.1,0.1))
59 plot(tree, type="fan", lwd=2, show.tip.label=F)
60
61 # Now we'll put points instead of names at the tips. The size of the points
62 will be proportional to the size of the lizard. To do this, we'll use
63 tiplabels(), but tell it to plot points instead of words (using the pch
64 parameter), and to scale the points by the size of each lizard using the
65 parameter cex. We'll make sure the tips match the points by reorganizing
66 the svl vector using the tip labels of the tree using square brackets.
67 tiplabels(pch=16, cex=0.25*svl[tree$tip.label])
68
69 # Now, we'll add the ancestral states to the tree using nodelabels(), as
70 each node
71 nodelabels(pch=16, cex=0.25*Ancestors$ace)
72
73 # Looking at the points, you can see SOME variation in size, but it's hard
74 to tell how large or small any given tip is relative to its ancestors or
75 even to other tips. So let's try visualizing this data in a different way
76 .
77
78 obj <- contMap(tree, svl, plot=F)
79 plot(obj, type="fan", legend=0.7*max(nodeHeights(tree)), sig=2, fsize=c(0.7,
80 0.9))
81
82 # Now, we're going to add some fossils in using data from ‘‘Amber fossils
83 demonstrate deep-time stability of Caribbean lizard communities’’ by Emma
84 Sherratt et al. 2015 in PNAS
85
86 # We'll take the measurements and the node closest to each fossil species
87 fossilData <- data.frame(svl=log(c(25.4, 23.2, 17.7, 19.7, 24, 31)), tip1=c(
88 "aliniger", "aliniger", "occultus", "christophe", "cristatellus", "

```

```

89     occultus"), tip2=c("chlorocyanus", "coelestinus", "monticola", "cybotes",
90     "angusticeps", "angusticeps"))
91
92 # What we'll do is: for each fossil, find what node corresponds to the Most
93     Recent Common Ancestor (MRCA) of the pair of tips in the dataframe.
94
95 # QUESTION 5: I've set up most of the for loop. You'll need to add the
96     actual for() line, and close the loop in the appropriate places.
97 fossilNodes <- c()
98 nodeN <- c()
99
100 Node <- fastMRCA(tree, fossilData[i, "tip1"], fossilData[i, "tip2"])
101 fossilNodes[i] <- fossilData[i, "svl"]
102 nodeN[i] <- Node
103
104 names(fossilNodes) <- nodeN
105
106 # Now we estimate the ancestral states, but this time use the fossils to
107     constrain the estimates at the relevant nodes
108 Ancestors_withFossils <- fastAnc(tree, svl, anc.states=fossilNodes, CI=TRUE,
109     var=TRUE)
110
111 # QUESTION 7: How do fossils, in general, change the estimated ancestral
112     sizes? (Hint, making a plot of the two different ancestral state
113     estimates against one another will help!)
114
115 # QUESTIONS 8 – 10: Now, I want you to determine if the assumptions you
116     identified in question 4 hold for these data. The way you'll want to go
117     about this is by installing a library called "geiger", and use a
118     function in that package called "fitContinuous". I want you to 1) fit
119     several different models of evolution to these data, 2) determine which
120     model fits the data best, and 3) describe whether or not that model is
121     different from what fastAnc assumes (look at the help page for fastAnc).
122
123 # This will require that you read the help pages for the pertinent functions
124     , and possibly some other functions. Google will be your friend here.
125     Akaike Information Criteria (AIC) will be an important facet. We'll talk
126     about what AIC is in class, but for this assignment all you need to know
127     is that the model with the LOWEST AIC is the best-fitting model.

```