

VAR-Seq project report template: Some Descriptive Title

Project ID: VARseq_PI_Name_Organism_Mar2015

Project PI: First Last (first.last@inst.edu)

Author of Report: First Last (first.last@inst.edu)

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1 Introduction

This report describes the analysis of an VAR-Seq project from Dr. First Last's lab which studies the genetic differences of ... in *organism* The experimental design is as follows...

2 Sample definitions and environment settings

2.1 Environment settings and input data

Typically, the user wants to record here the sources and versions of the reference genome sequence along with the corresponding annotations. In the provided sample data set all data inputs are stored in a data subdirectory and all results will be written to a separate results directory, while the `systemPipeVARseq.Rnw` script and the `targets` file are expected to be located in the parent directory. The R session is expected to run from this parent directory.

To run this sample report, mini sample FASTQ and reference genome files can be downloaded from [here](#). The chosen data set [SRP010938](#) contains 18 paired-end (PE) read sets from *Arabidopsis thaliana* [Howard et al. \(2013\)](#). This data set comes from a different NGS application area, but it is sufficient to demonstrate the analysis steps of this workflow. To minimize processing time during testing, each FASTQ file has been subsetting to 90,000-100,000 randomly sampled PE reads that map to the first 100,000 nucleotides of each chromosome of the *A. thaliana* genome. The corresponding reference genome sequence (FASTA) and its GFF annotation files (provided in the same download) have been truncated accordingly. This way the entire test sample data set is less than 200MB in storage space. A PE read set has been chosen for this test data set for flexibility, because it can be used for testing both types of analysis routines requiring either SE (single end) reads or PE reads.

2.2 Required packages and resources

The `systemPipeR` package needs to be loaded to perform the analysis steps shown in this report ([Girke, 2014](#)).

```
> library(systemPipeR)
```

If applicable load custom functions not provided by `systemPipeR`

```
> source("systemPipeVARseq_Fct.R")
```

2.3 Experiment definition provided by targets file

The `targets` file defines all FASTQ files and sample comparisons of the analysis workflow.

```
> targetspath <- system.file("extdata", "targetsPE.txt", package="systemPipeR")
> targets <- read.delim(targetspath, comment.char = "#")[,1:5]
> targets
```

	FileName1	FileName2	SampleName	Factor	SampleLong
1	./data/SRR446027_1.fastq	./data/SRR446027_2.fastq	M1A	M1	Mock.1h.A
2	./data/SRR446028_1.fastq	./data/SRR446028_2.fastq	M1B	M1	Mock.1h.B
3	./data/SRR446029_1.fastq	./data/SRR446029_2.fastq	A1A	A1	Avr.1h.A
4	./data/SRR446030_1.fastq	./data/SRR446030_2.fastq	A1B	A1	Avr.1h.B
5	./data/SRR446031_1.fastq	./data/SRR446031_2.fastq	V1A	V1	Vir.1h.A

6	./data/SRR446032_1.fastq	./data/SRR446032_2.fastq	V1B	V1	Vir.1h.B
7	./data/SRR446033_1.fastq	./data/SRR446033_2.fastq	M6A	M6	Mock.6h.A
8	./data/SRR446034_1.fastq	./data/SRR446034_2.fastq	M6B	M6	Mock.6h.B
9	./data/SRR446035_1.fastq	./data/SRR446035_2.fastq	A6A	A6	Avr.6h.A
10	./data/SRR446036_1.fastq	./data/SRR446036_2.fastq	A6B	A6	Avr.6h.B
11	./data/SRR446037_1.fastq	./data/SRR446037_2.fastq	V6A	V6	Vir.6h.A
12	./data/SRR446038_1.fastq	./data/SRR446038_2.fastq	V6B	V6	Vir.6h.B
13	./data/SRR446039_1.fastq	./data/SRR446039_2.fastq	M12A	M12	Mock.12h.A
14	./data/SRR446040_1.fastq	./data/SRR446040_2.fastq	M12B	M12	Mock.12h.B
15	./data/SRR446041_1.fastq	./data/SRR446041_2.fastq	A12A	A12	Avr.12h.A
16	./data/SRR446042_1.fastq	./data/SRR446042_2.fastq	A12B	A12	Avr.12h.B
17	./data/SRR446043_1.fastq	./data/SRR446043_2.fastq	V12A	V12	Vir.12h.A
18	./data/SRR446044_1.fastq	./data/SRR446044_2.fastq	V12B	V12	Vir.12h.B

3 Read preprocessing

3.1 FASTQ quality report

The following `seeFastq` and `seeFastqPlot` functions generate and plot a series of useful quality statistics for a set of FASTQ files including per cycle quality box plots, base proportions, base-level quality trends, relative k-mer diversity, length and occurrence distribution of reads, number of reads above quality cutoffs and mean quality distribution. The results are written to a PDF file named `fastqReport.pdf`.

```
> args <- systemArgs(sysma="bwa.param", mytargets="targets.txt")
> fqlist <- seeFastq(fastq=infile1(args), batchsize=100000, klength=8)
> pdf("./results/fastqReport.pdf", height=18, width=4*length(fqlist))
> seeFastqPlot(fqlist)
> dev.off()
```

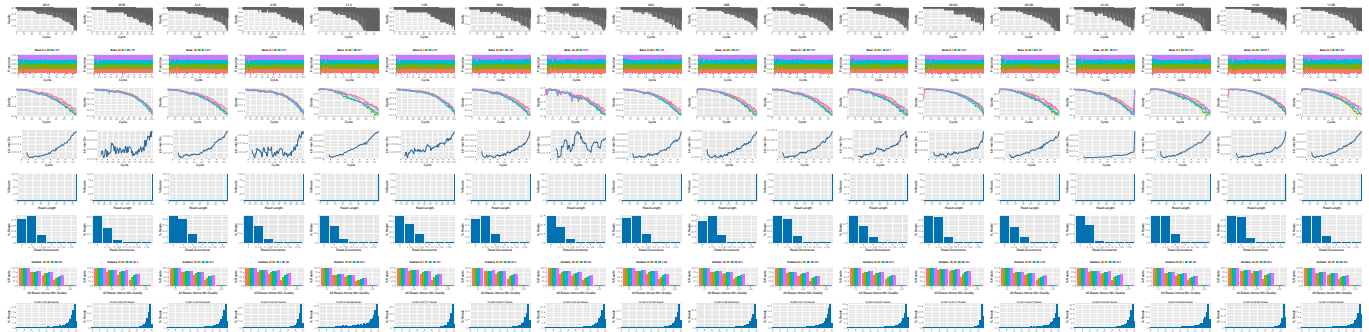


Figure 1: QC report for 18 FASTQ files.

4 Alignments

4.1 Read mapping with BWA

The NGS reads of this project are aligned against the reference genome sequence using the highly variant tolerant short read aligner BWA (Li, 2013; Li and Durbin, 2009). The parameter settings of the aligner are defined in the `bwa.param` file.

```
> args <- systemArgs(sysma="bwa.param", mytargets="targets.txt")
> sysargs(args)[1] # Command-line parameters for first FASTQ file
```

Runs the alignments sequentially (e.g. on a single machine)

```
> bampaths <- runCommandline(args=args)
```

Alternatively, the alignment jobs can be submitted to a compute cluster, here using 72 CPU cores (18 qsub processes each with 4 CPU cores).

```
> moduleload(modules(args))
> system("bwa index -a bwtsw ./data/tair10.fasta")
> resources <- list(walltime="20:00:00", nodes=paste0("1:ppn=", cores(args)), memory="10gb")
> reg <- clusterRun(args, conffile=".BatchJobs.R", template="torque.tmpl", Njobs=18, runid="01",
+                   resourceList=resources)
```

Check whether all BAM files have been created

```
> file.exists(outpaths(args))
```

4.2 Read and alignment stats

The following generates a summary table of the number of reads in each sample and how many of them aligned to the reference.

```
> read_statsDF <- alignStats(args=args)
> write.table(read_statsDF, "results/alignStats.xls", row.names=FALSE, quote=FALSE, sep="\t")
```

4.3 Create symbolic links for viewing BAM files in IGV

The symLink2bam function creates symbolic links to view the BAM alignment files in a genome browser such as IGV. The corresponding URLs are written to a file with a path specified under urlfile, here [IGVurl.txt](#).

```
> symLink2bam(sysargs=args, htmldir=c("~/html/", "somedir/"),
+             urlbase="http://biocluster.ucr.edu/~tgirke/",
+             urlfile="./results/IGVurl.txt")
```

5 Variant calling

The following performs variant calling with GATK and BCFtools in parallel mode on a compute cluster ([McKenna et al., 2010](#); [Li, 2011](#)). If a cluster is not available, the runCommandline() function can be used to run the variant calling for each sample sequentially on a single machine. Typically, the user would choose here one variant caller rather than running several ones.

5.1 Variant calling with GATK

The following creates in the initial step a new targets file (targets_bam.txt). The first column of this file gives the paths to the BAM files created in the alignment step. The new targets file and the parameter file gatk.param are used to create a new SYSargs instance for running GATK. Since GATK involves many processing steps, it is executed by a bash script gatk_run.sh where the user can specify the detailed run parameters. All three files are expected to be located in the current working directory. Samples files for gatk.param and gatk_run.sh are available in the subdirectory ./inst/extdata/ of the source file of the systemPipeR package. Alternatively, they can be downloaded directly from [here](#).

```
> writeTargetsout(x=args, file="targets_bam.txt")
> system("java -jar CreateSequenceDictionary.jar R=./data/tair10.fasta O=./data/tair10.dict")
> # system("java -jar /opt/picard/1.81/CreateSequenceDictionary.jar R=./data/tair10.fasta O=./data/tair10.dict")
> args <- systemArgs(sysma="gatk.param", mytargets="targets_bam.txt")
> resources <- list(walltime="20:00:00", nodes=paste0("1:ppn=", 1), memory="10gb")
> reg <- clusterRun(args, conffile=".BatchJobs.R", template="torque.tmpl", Njobs=18, runid="01",
+                   resourceList=resources)
```

5.2 Variant calling with BCFtools

The following runs the variant calling with BCFtools. This step requires in the current working directory the parameter file sambcf.param. A bash script is not required in this case.

```
> args <- systemArgs(sysma="sambcf.param", mytargets="targets_bam.txt")
> resources <- list(walltime="20:00:00", nodes=paste0("1:ppn=", 1), memory="10gb")
> reg <- clusterRun(args, conffile=".BatchJobs.R", template="torque.tmpl", Njobs=18, runid="01",
+                   resourceList=resources)
```

6 Filter variants

The function `filterVars` filters VCF files based on user definable quality parameters. It sequentially imports each VCF file into R, applies the filtering on an internally generated `VRanges` object and then writes the results to a new subsetted VCF file. The filter parameters are passed on to the corresponding argument as a character string. The function applies this filter to the internally generated `VRanges` object using the standard subsetting syntax for two dimensional objects such as: `vr[filter,]`. The parameter files (`filter_gatk.param` and `filter_sambcf.param`), used in the filtering steps, define the paths to the input and output VCF files which are stored in new `SYSargs` instances.

6.1 Filter variants called by GATK

The below example filters for variants that are supported by $\geq x$ reads and $\geq 80\%$ of them support the called variants. In addition, all variants need to pass $\geq x$ of the soft filters recorded in the VCF files generated by GATK. Since the toy data used for this workflow is very small, the chosen settings are unreasonably relaxed. A more reasonable filter setting is given in the line below (here commented out).

```
> library(VariantAnnotation)
> args <- systemArgs(sysma="filter_gatk.param", mytargets="targets_gatk.txt")
> filter <- "totalDepth(vr) >= 2 & (altDepth(vr) / totalDepth(vr) >= 0.8) & rowSums(softFilterMatrix(vr)) >= 2"
> # filter <- "totalDepth(vr) >= 20 & (altDepth(vr) / totalDepth(vr) >= 0.8) & rowSums(softFilterMatrix(vr)) >= 2"
> filterVars(args, filter, varcaller="gatk", organism="A. thaliana")
> writeTargetsout(x=args, file="targets_gatk_filtered.txt")
```

6.2 Filter variants called by BCFtools

The following shows how to filter the VCF files generated by BCFtools using similar parameter settings as in the previous filtering of the GATK results.

```
> args <- systemArgs(sysma="filter_sambcf.param", mytargets="targets_sambcf.txt")
> filter <- "rowSums(vr) >= 2 & (rowSums(vr[,3:4])/rowSums(vr[,1:4]) >= 0.8)"
> # filter <- "rowSums(vr) >= 20 & (rowSums(vr[,3:4])/rowSums(vr[,1:4]) >= 0.8)"
> filterVars(args, filter, varcaller="bcftools", organism="A. thaliana")
> writeTargetsout(x=args, file="targets_sambcf_filtered.txt")
```

7 Annotate filtered variants

The function `variantReport` generates a variant report using utilities provided by the *VariantAnnotation* package. The report for each sample is written to a tabular file containing genomic context annotations (e.g. coding or non-coding SNPs, amino acid changes, IDs of affected genes, etc.) along with confidence statistics for each variant. The parameter file `annotate_vars.param` defines the paths to the input and output files which are stored in a new `SYSargs` instance.

7.1 Annotate filtered variants called by GATK

```
> library("GenomicFeatures")
> args <- systemArgs(sysma="annotate_vars.param", mytargets="targets_gatk_filtered.txt")
> txdb <- loadDb("./data/tair10.sqlite")
> fa <- FaFile(systemPipeR::reference(args))
> variantReport(args=args, txdb=txdb, fa=fa, organism="A. thaliana")
```

7.2 Annotate filtered variants called by BCFtools

```
> args <- systemArgs(sysma="annotate_vars.param", mytargets="targets_sambcf_filtered.txt")
> txdb <- loadDb("./data/tair10.sqlite")
> fa <- FaFile(systemPipeR::reference(args))
> variantReport(args=args, txdb=txdb, fa=fa, organism="A. thaliana")
```

8 Combine annotation results among samples

To simplify comparisons among samples, the `combineVarReports` function combines all variant annotation reports referenced in a `SYSargs` instance (here `args`). At the same time the function allows to consider only certain feature types of interest. For instance, the below setting `filtercol=c(Consequence="nonsynonymous")` will include only nonsynonymous variances listed in the `Consequence` column of the annotation reports. To omit filtering, one can use the setting `filtercol="All"`

8.1 Combine results from GATK

```
> args <- systemArgs(sysma="annotate_vars.param", mytargets="targets_gatk_filtered.txt")
> combinedF <- combineVarReports(args, filtercol=c(Consequence="nonsynonymous"))
> write.table(combinedF, "./results/combinedF_nonsyn_gatk.xls", quote=FALSE, row.names=FALSE, sep="\t")
```

8.2 Combine results from BCFtools

```
> args <- systemArgs(sysma="annotate_vars.param", mytargets="targets_sambcf_filtered.txt")
> combinedF <- combineVarReports(args, filtercol=c(Consequence="nonsynonymous"))
> write.table(combinedF, "./results/combinedF_nonsyn_sambcf.xls", quote=FALSE, row.names=FALSE, sep="\t")
```

9 Summary statistics of variants

The function `varSummar` counts the number of variances for each feature type included in the annotation reports.

9.1 Summary for GATK

```
> args <- systemArgs(sysma="annotate_vars.param", mytargets="targets_gatk_filtered.txt")
> write.table(varSummary(args), "./results/variantStats_gatk.xls", quote=FALSE, col.names = NA, sep="\t")
```

9.2 Summary for BCFtools

```
> args <- systemArgs(sysma="annotate_vars.param", mytargets="targets_sambcf_filtered.txt")
> write.table(varSummary(args), "./results/variantStats_sambcf.xls", quote=FALSE, col.names = NA, sep="\t")
```

10 Venn diagram of variants

The venn diagram utilities defined by the *systemPipeR* package can be used to identify common and unique variants reported for different samples and/or variant callers. The below generates a 4-way venn diagram comparing four samples for each of the two variant callers.

```
> args <- systemArgs(sysma="annotate_vars.param", mytargets="targets_gatk_filtered.txt")
> varlist <- sapply(names(outpaths(args))[1:4], function(x) as.character(read.delim(outpaths(args)[x])$VAR))
> vennset_gatk <- overLapper(varlist, type="vennsets")
> args <- systemArgs(sysma="annotate_vars.param", mytargets="targets_sambcf_filtered.txt")
> varlist <- sapply(names(outpaths(args))[1:4], function(x) as.character(read.delim(outpaths(args)[x])$VAR))
> vennset_bcf <- overLapper(varlist, type="vennsets")
> pdf("./results/vennplot_var.pdf")
> vennPlot(list(vennset_gatk, vennset_bcf), mymain="", mysub="GATK: red; BCFtools: blue", colmode=2, ccol=)
> dev.off()
```

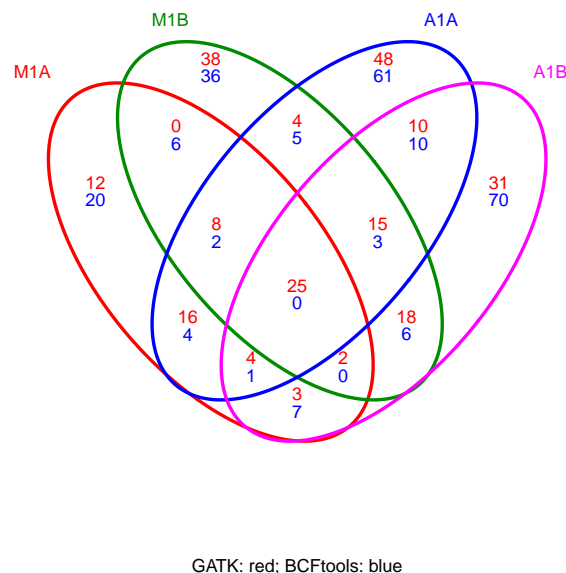


Figure 2: Venn Diagram for 4 samples from GATK and BCFtools.

11 Version Information

```
> toLatex(sessionInfo())
```

- R version 3.1.2 (2014-10-31), x86_64-unknown-linux-gnu
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, parallel, stats, stats4, utils
- Other packages: AnnotationDbi 1.28.2, Biobase 2.26.0, BiocGenerics 0.12.1, BiocParallel 1.0.2, Biostrings 2.34.1, DBI 0.3.1, GenomeInfoDb 1.2.4, GenomicAlignments 1.2.1, GenomicRanges 1.18.4, IRanges 2.0.1, RSQLite 1.0.0, Rsamtools 1.18.2, S4Vectors 0.4.0, ShortRead 1.24.0, XVector 0.6.0, systemPipeR 1.0.12
- Loaded via a namespace (and not attached): AnnotationForge 1.8.2, BBmisc 1.9, BatchJobs 1.5, BiocStyle 1.4.1, Category 2.32.0, GO.db 3.0.0, GOSTats 2.32.0, GSEABase 1.28.0, MASS 7.3-37, Matrix 1.1-5, RBGL 1.42.0, RColorBrewer 1.1-2, Rcpp 0.11.4, XML 3.98-1.1, annotate 1.44.0, base64enc 0.1-2, bitops 1.0-6, brew 1.0-6, checkmate 1.5.1, codetools 0.2-11, colorspace 1.2-6, digest 0.6.8, edgeR 3.8.5, fail 1.2, foreach 1.4.2, genefilter 1.48.1, ggplot2 1.0.0, graph 1.44.1, grid 3.1.2, gtable 0.1.2, hwriter 1.3.2, iterators 1.0.7, lattice 0.20-30, latticeExtra 0.6-26, limma 3.22.4, munsell 0.4.2, pheatmap 1.0.2, plyr 1.8.1, proto 0.3-10, reshape2 1.4.1, rjson 0.2.15, scales 0.2.4, sendmailR 1.2-1, splines 3.1.2, stringr 0.6.2, survival 2.38-1, tools 3.1.2, xtable 1.7-4, zlibbioc 1.12.0

12 Funding

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13 References

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