

# Bioinformática-introducción

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Bioinformática

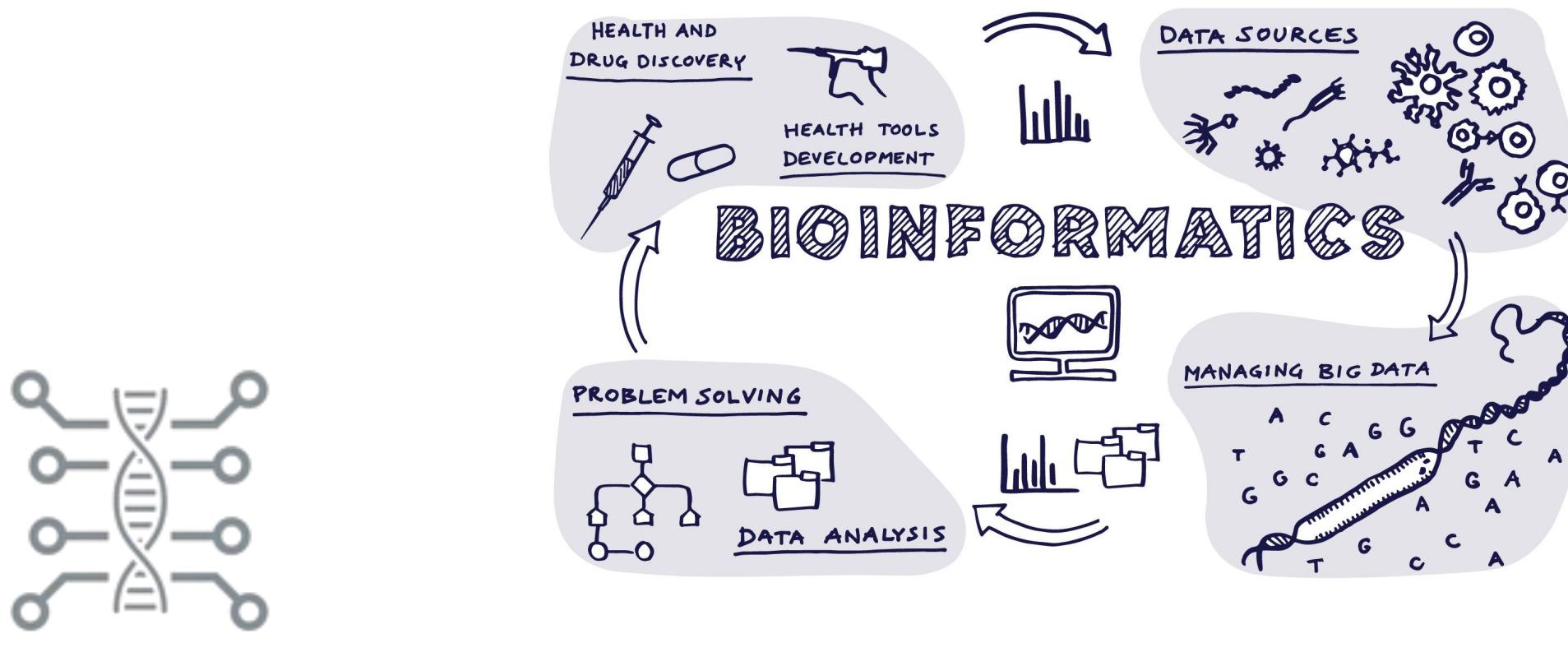
Semestre 2026-1

# ¿Qué es bioinformática?



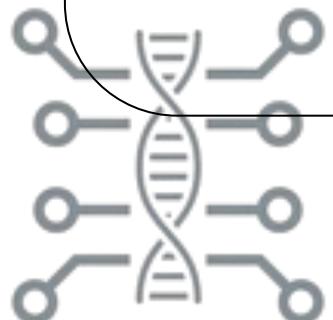
## ¿Qué es bioinformática?

“Bioinformatics is a field that combines biology and computer science to study biological information. It involves using computers to analyze and interpret large amounts of data related to genetics, DNA, and proteins to understand how living organisms work.”

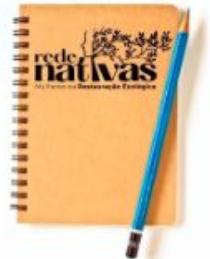


# Biologists starter pack

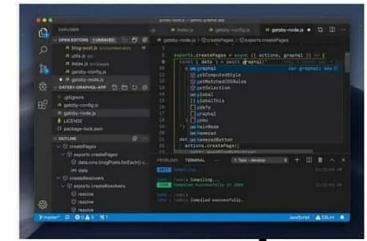
## De bata



## De Bota



## De BigData



macbook

vs code



glasses must



False promises

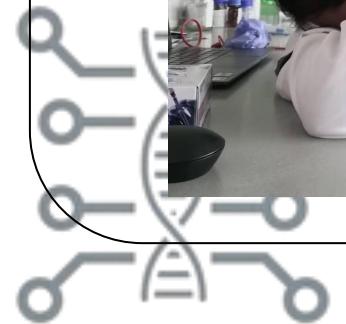


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# Biologists starter pack

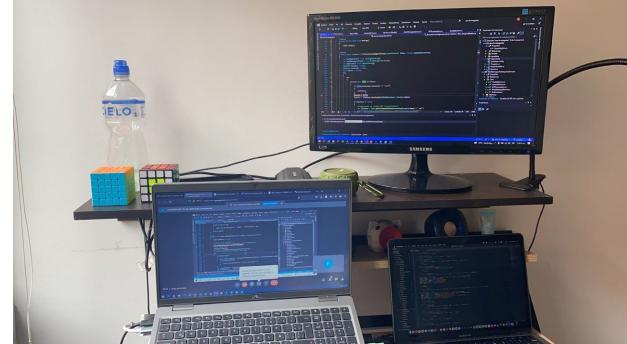
**De bata**



**De Bota**



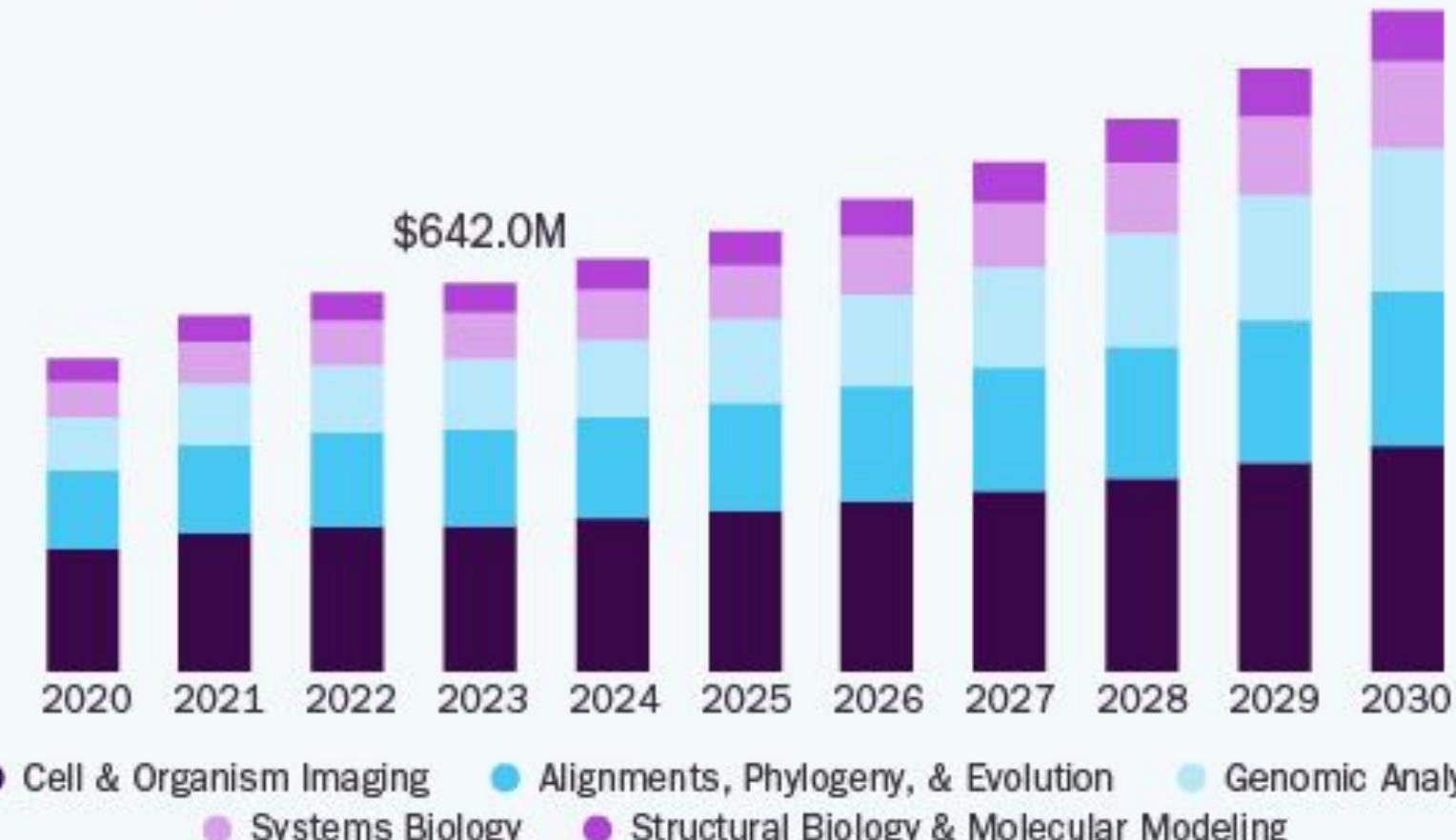
**De BigData**



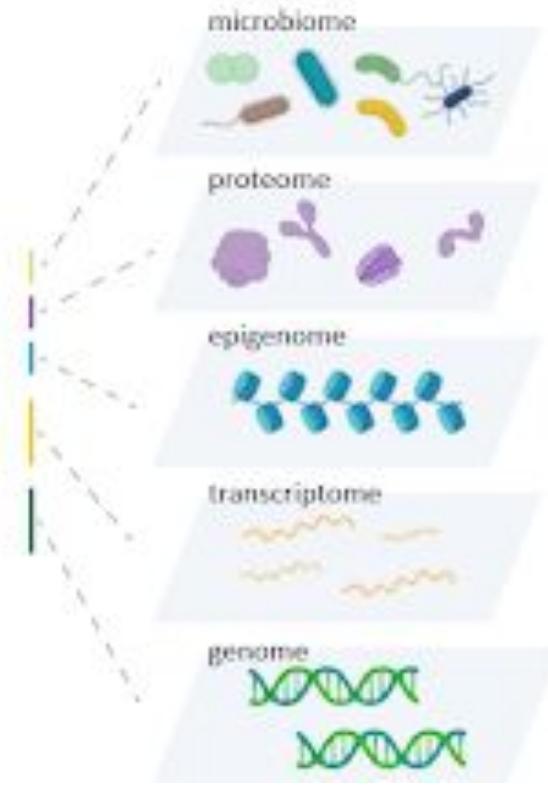
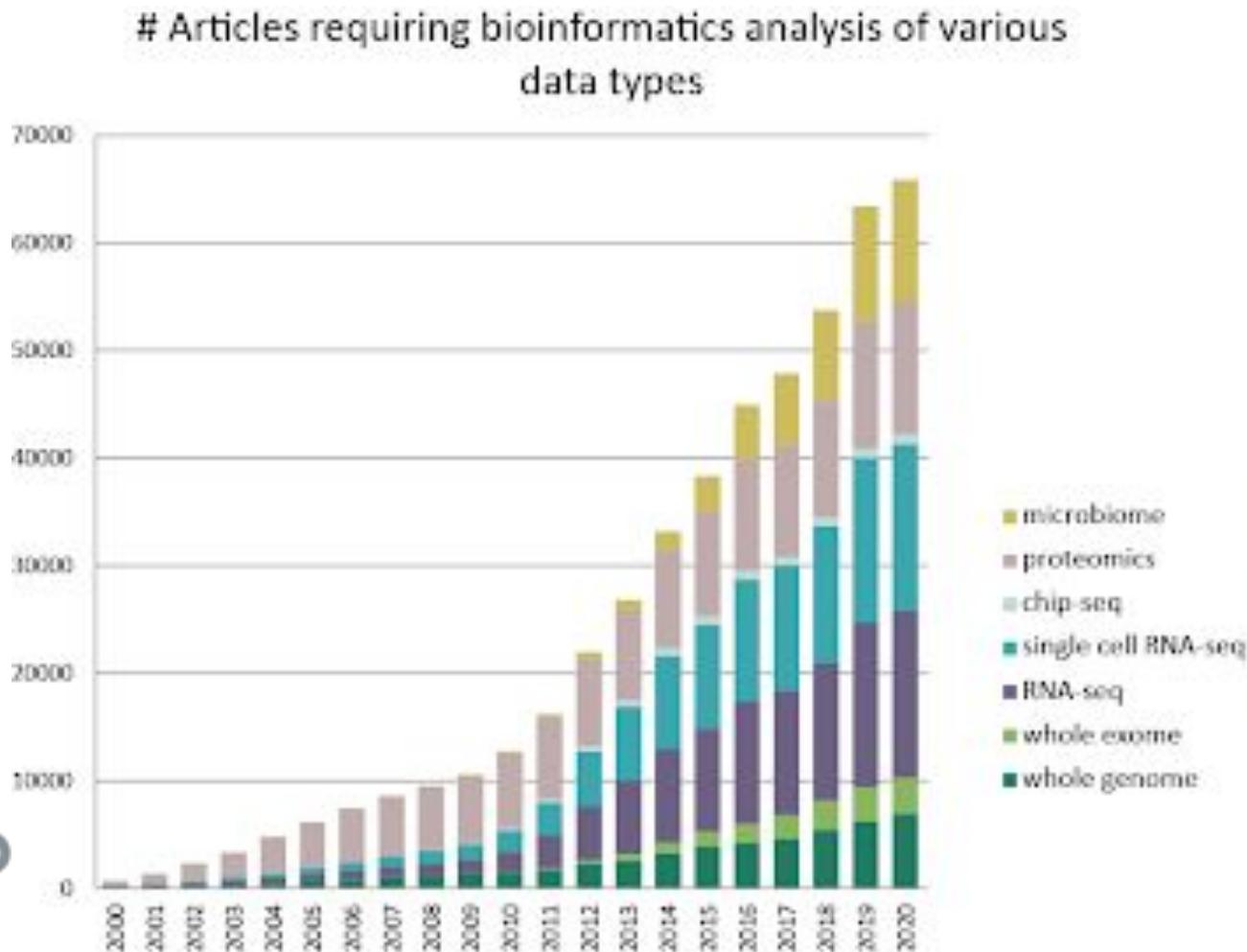
# ¿Por qué Bioinformática?

## Biological Data Visualization Market

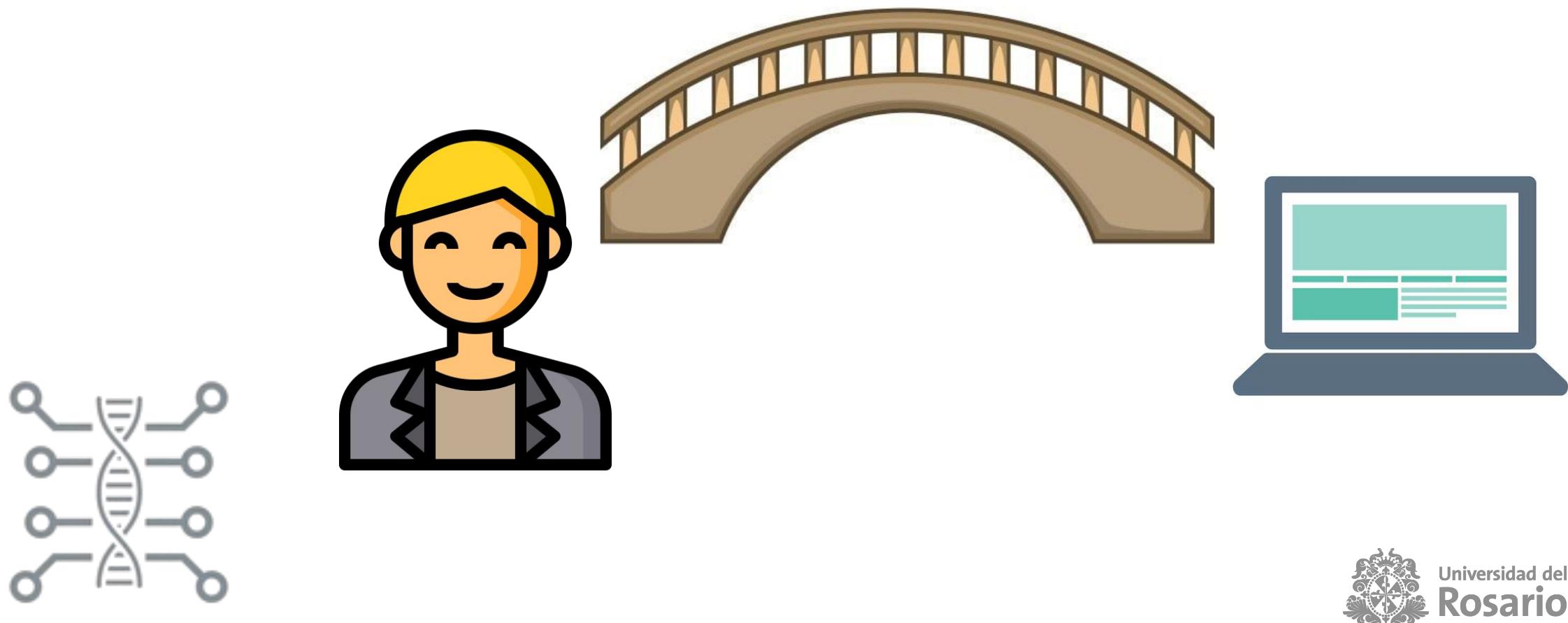
Size, by Application, 2020 - 2030 (USD Million)



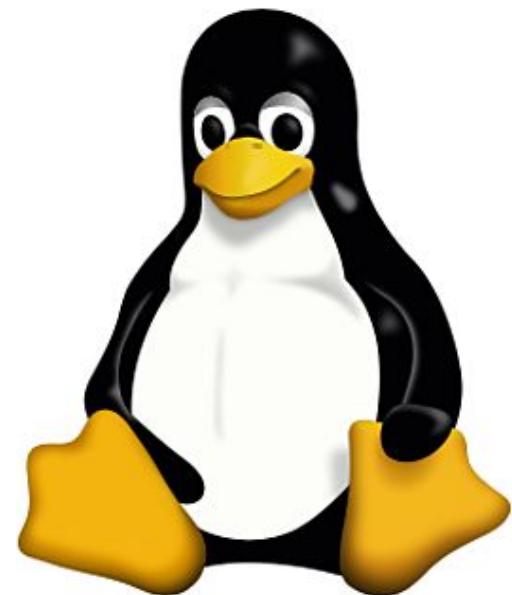
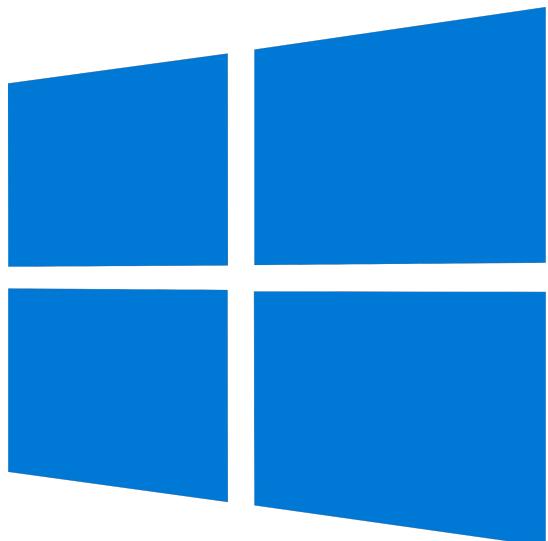
# ¿Por qué Bioinformática?



# ¿Qué es un sistema operativo?



# ¿Qué es un sistema operativo?



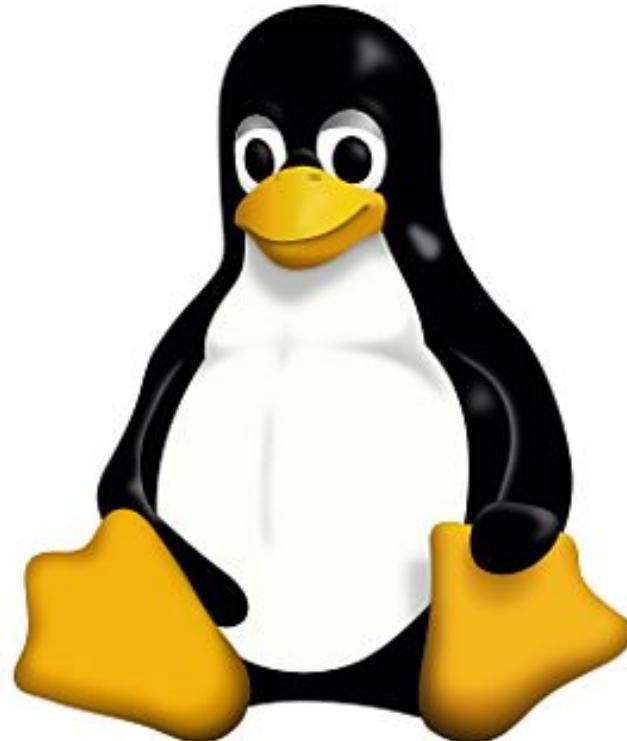
iOS



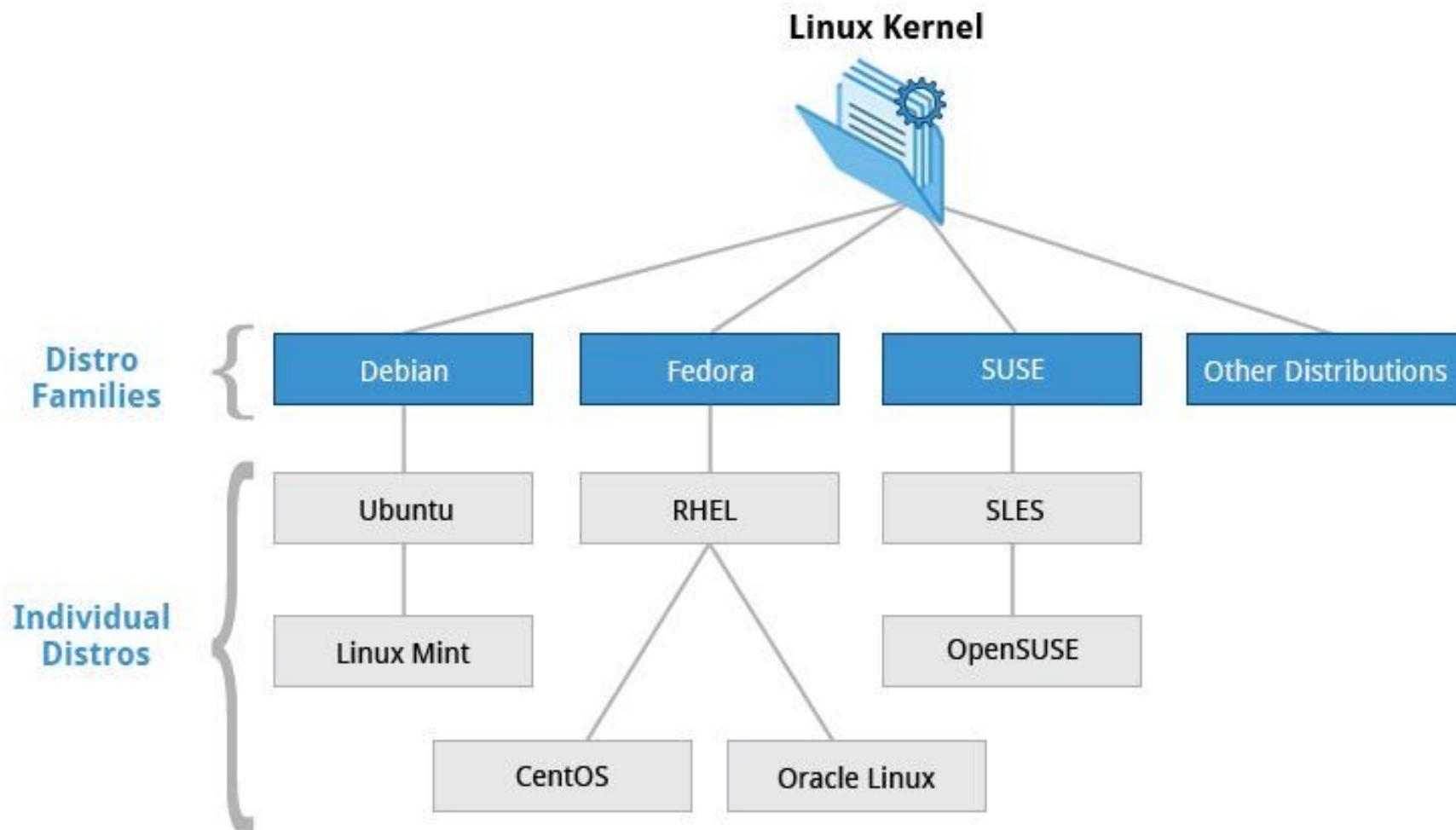
# **¿Por qué algunos biólogos no usan Windows?**



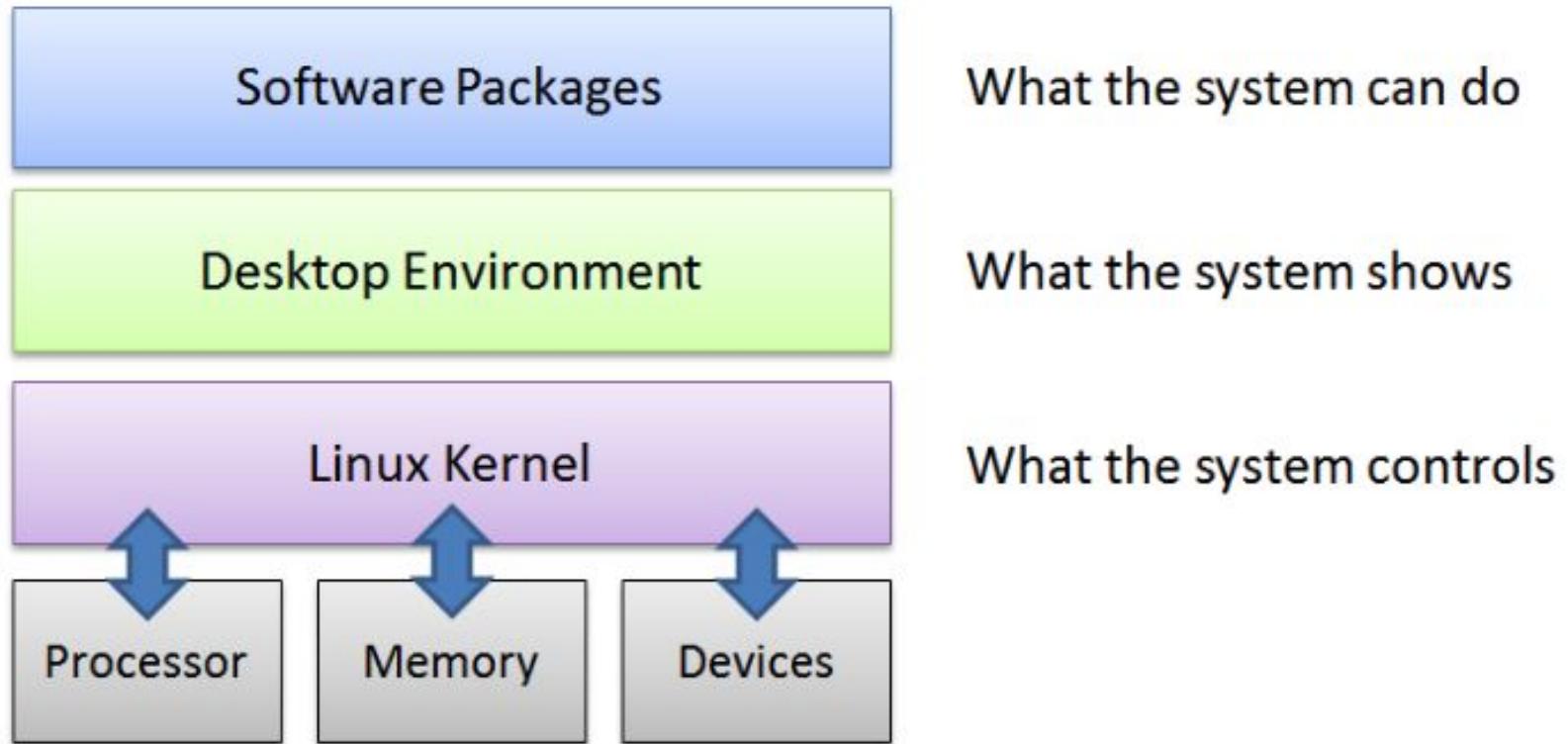
# ¿Por qué algunos biólogos no usan Windows?



# Distribuciones de Linux



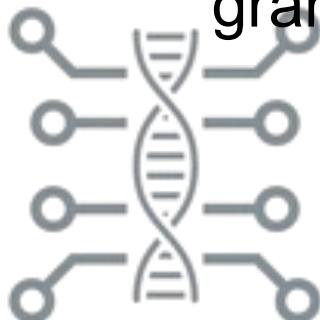
# Distribuciones de Linux



# ¿Por qué algunos biólogos no usan Windows?

La mejor parte!!

- ¡Es gratuito!
- Código abierto
- Hay mucho software en biología escrito
- Permite organizarse muy fácil
- Facilita el procesamiento de datos grandes.

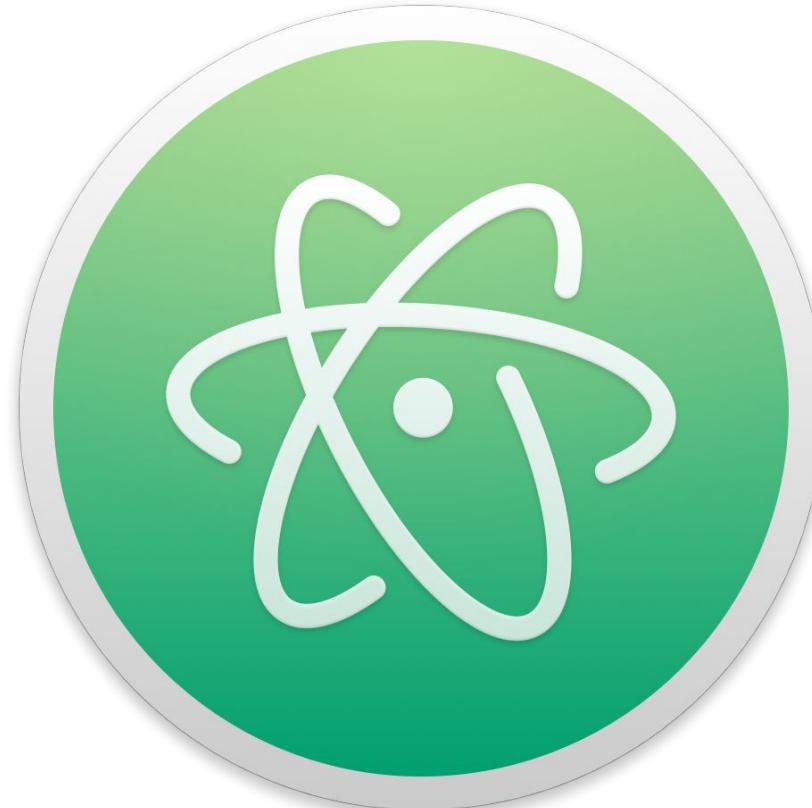
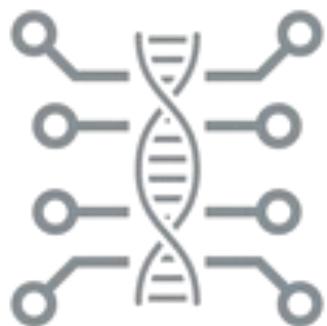
A screenshot of a terminal window titled "ruvelro@ruvelro-vm: ~". The window shows a root prompt (\$). The text in the terminal reads:

```
To run a command as administrator (user "root"), use "sudo <command>".
See "man sudo_root" for details.

ruvelro@ruvelro-vm:~$
```

# Para esta clase necesitamos

- Un computador
- Un editor de texto
- Y un sistema operativo Unix

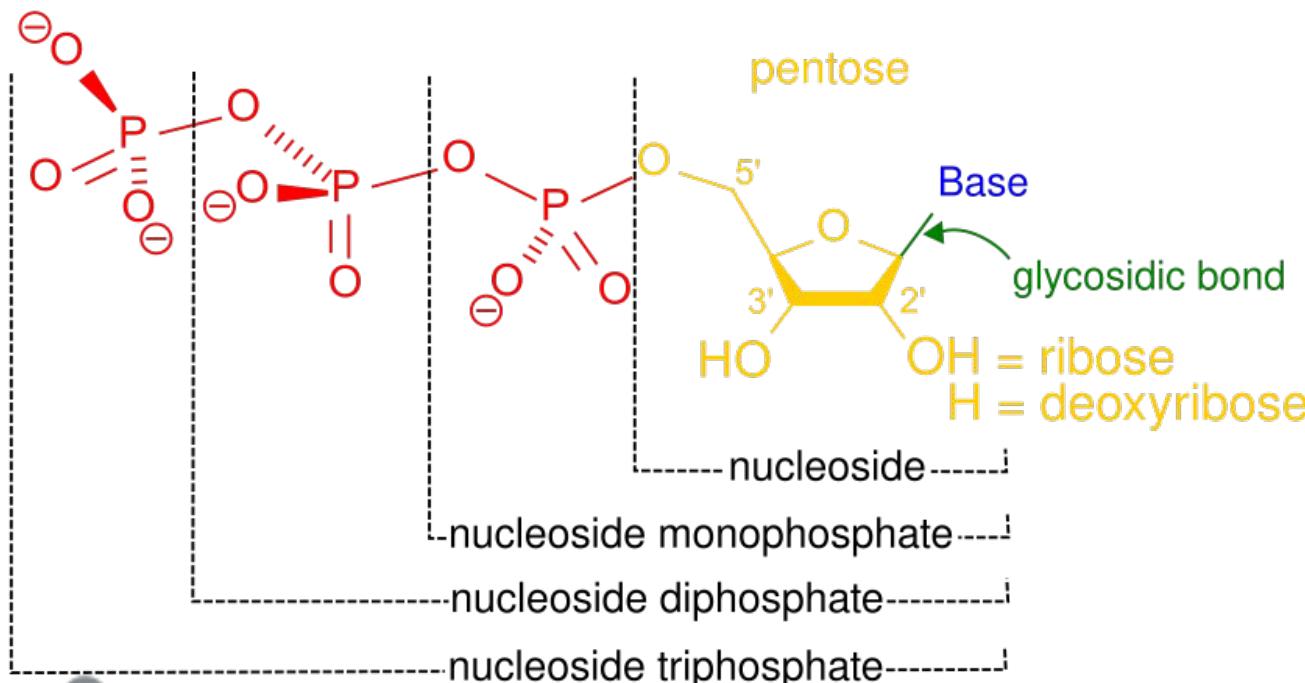


# ¿Cómo estamos de términos?

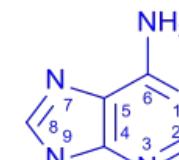
- Base/nucleotide
- Read
- Contig
- Scaffold
- Chromosome
- FASTA
- FASTQ
- GFF
- GTF
- VCF
- SAM
- BAM



# Base/nucleotide



## Purines

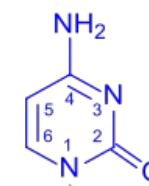


Adenine

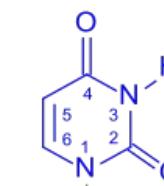


Guanine

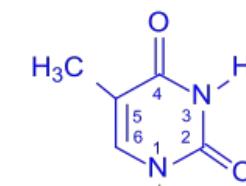
## Pyrimidines



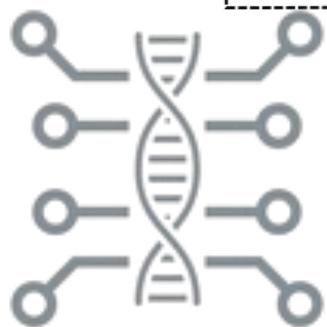
Cytosine



Uracil



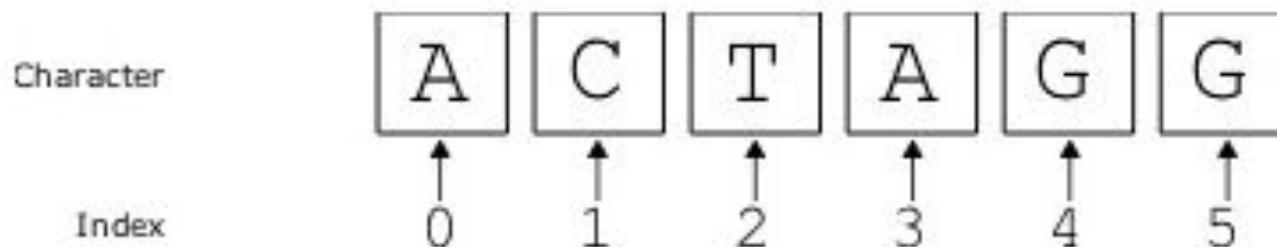
Thymine



Unidades estructurales y funcionales básicas de los ácidos nucleicos

# Base/nucleotide

TTAACCTTGGTTTGAACTTGAACACTTAGGGGATTGAAGAGATTCAACAACCCTAAAGCTGGGGTAAAAC



*Figure 1.1: A simple representation of a character string. Each position of the string can be accessed by its index number, which goes from 0 to the length minus one.*

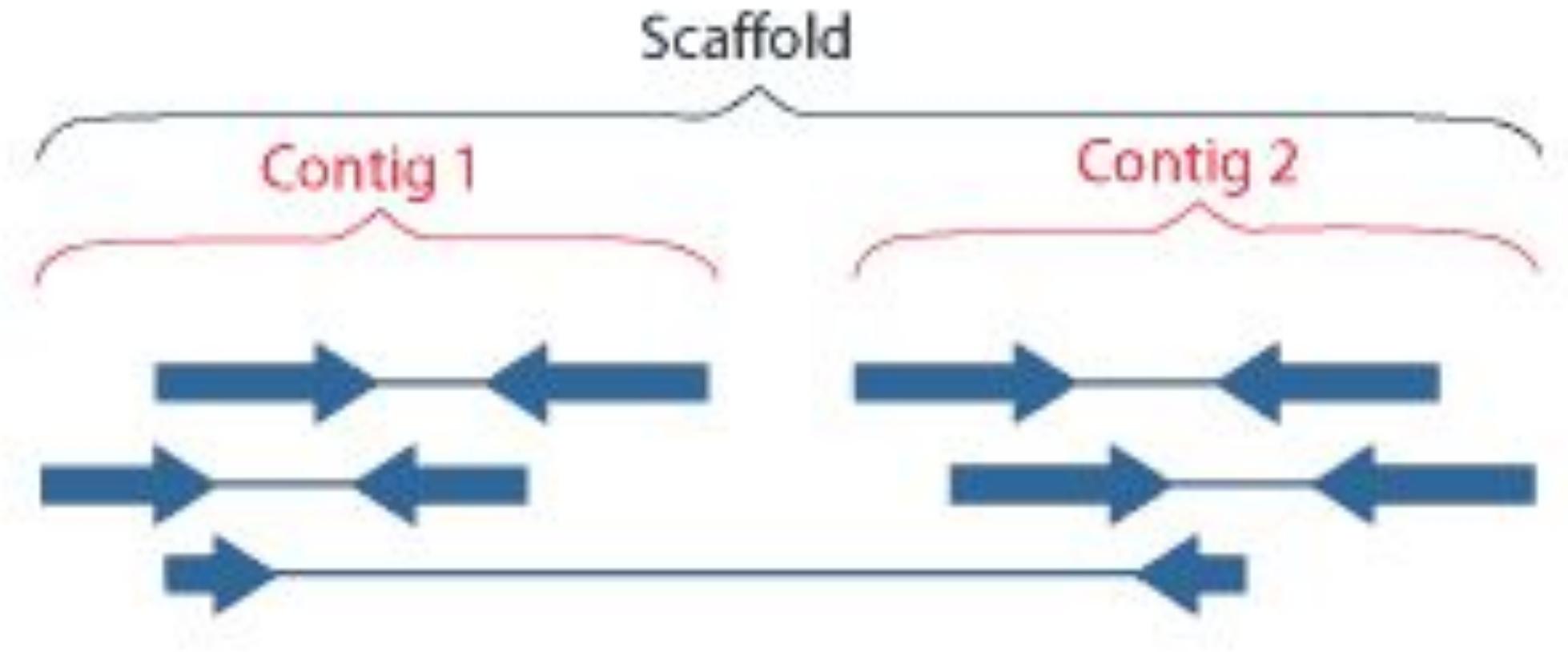


# Contig

|        |   |
|--------|---|
| Read   | TTAACCTTGGTTTGAACTTGAACACTTAGGGGATTG                                  |
| Read   | TGGTTTGAACTTGAACACTTAGGGGATTGAAGATTCAACAA                             |
| Read   | AACTTGAACACTTAGGGGATTGAAGATTCAACAACCCCTAAAGCT                         |
| Read   | CACTTAGGGGATTGAAGATTCAACAACCCCTAAAGCTTGGG                             |
| Read   | ATTGAAGATTCAACAACCCCTAAAGCTTGGGT                                      |
| Read   | ACCCTAAAGCTTGGGTAA  |
| Read   | AGCTTGGGTAAAAC  |
| Contig | TTAACCTTGGTTTGAACTTGAACACTTAGGGGATTGAAGATTCAACAACCCCTAAAGCTTGGGTAAAAC |



Un contig es la secuencia de consenso generada al alinear las lecturas entre sí.



Fragment



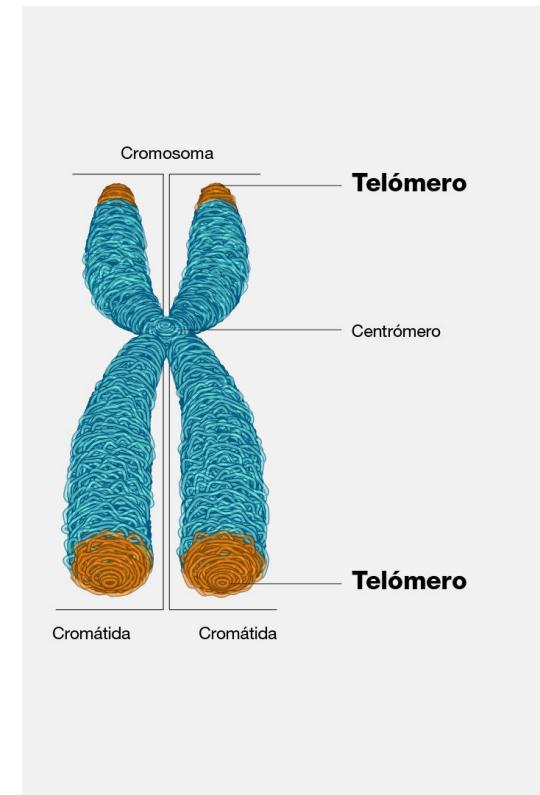
Read (known sequence)



Roughly known length but not known sequence

# Cromosoma

- Moléculas de ADN más grandes de una célula.
- Se pueden ordenar y orientar mediante un mapa genético o datos de Hi-C.
- El objetivo final de un proyecto de ensamblaje genómico es ensamblar lecturas en cromosomas en fase que representen a un individuo real.
- La mayoría de los ensamblajes cromosómicos producidos hoy en día no están en fase.



## FASTA

```
1 >gi|425153|gb|L26238.1|MUSHOME Mus domesticus (lbx) homeodomain mRNA, partial cds
2 CCATTTCAACAAGTACCTGACCAGGGCTCGCGAGTGGAGTTGCCGTATTCTCGAGCTAACGAAACT
3 CAAGTGAAAATT
```

- 1 Line 1: starts with ">" followed by ID
- 2 Line 2: Sequence data

## FASTQ

1 Line 1: starts with "@" followed by ID  
2 Line 2: Sequence data  
3 Line 3: Starts with "+" rest of the description is optional  
4 Line 4: Quality score for each base in the sequence



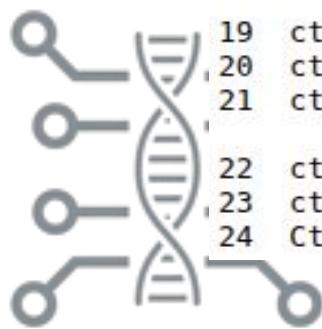
# GFF-General Feature Format

Column 1 seqID (e.g. chromosome/scaffold, genome id, etc..)  
Column 2 Source (program used to generate or location of download)  
Column 3 Feature type (gene, mRNA, CDS, exon, etc.)  
Column 4 Start position of feature  
Column 5 End position of feature  
Column 6 Score (some program outputs will have a score of confidence for feature)  
Column 7 Strand (+,-,..)  
Column 8 Phase  
Column 9 List of attributes in the format tag=value. Multiple attributes are separated by ";"



# GFF-General Feature Format

```
0 ##gff-version 3
1 ##sequence-region ctg123 1 1497228
2 ctg123 . gene    1000 9000 . + . ID=gene00001;Name=EDEN
3 ctg123 . TF_binding_site 1000 1012 . + . ID=tfbs00001;Parent=gene00001
4 ctg123 . mRNA    1050 9000 . + . ID=mRNA00001;Parent=gene00001;Name=EDEN.1
5 ctg123 . mRNA    1050 9000 . + . ID=mRNA00002;Parent=gene00001;Name=EDEN.2
6 ctg123 . mRNA    1300 9000 . + . ID=mRNA00003;Parent=gene00001;Name=EDEN.3
7 ctg123 . exon    1300 1500 . + . ID=exon00001;Parent=mRNA00003
8 ctg123 . exon    1050 1500 . + . ID=exon00002;Parent=mRNA00001,mRNA00002
9 ctg123 . exon    3000 3902 . + . ID=exon00003;Parent=mRNA00001,mRNA00003
10 ctg123 . exon   5000 5500 . + . ID=exon00004;Parent=mRNA00001,mRNA00002,mRNA00003
11 ctg123 . exon   7000 9000 . + . ID=exon00005;Parent=mRNA00001,mRNA00002,mRNA00003
12 ctg123 . CDS    1201 1500 . + 0 ID=cds00001;Parent=mRNA00001;Name=edenprotein.1
13 ctg123 . CDS    3000 3902 . + 0 ID=cds00001;Parent=mRNA00001;Name=edenprotein.1
14 ctg123 . CDS    5000 5500 . + 0 ID=cds00001;Parent=mRNA00001;Name=edenprotein.1
15 ctg123 . CDS    7000 7600 . + 0 ID=cds00001;Parent=mRNA00001;Name=edenprotein.1
16 ctg123 . CDS    1201 1500 . + 0 ID=cds00002;Parent=mRNA00002;Name=edenprotein.2
17 ctg123 . CDS    5000 5500 . + 0 ID=cds00002;Parent=mRNA00002;Name=edenprotein.2
18 ctg123 . CDS    7000 7600 . + 0 ID=cds00002;Parent=mRNA00002;Name=edenprotein.2
19 ctg123 . CDS    3301 3902 . + 0 ID=cds00003;Parent=mRNA00003;Name=edenprotein.3
20 ctg123 . CDS    5000 5500 . + 2 ID=cds00003;Parent=mRNA00003;Name=edenprotein.3
21 ctg123 . CDS    7000 7600 . + 2 ID=cds00003;Parent=mRNA00003;Name=edenprotein.3
22 ctg123 . CDS    3391 3902 . + 0 ID=cds00004;Parent=mRNA00003;Name=edenprotein.4
23 ctg123 . CDS    5000 5500 . + 2 ID=cds00004;Parent=mRNA00003;Name=edenprotein.4
24 Ctg123 . CDS    7000 7600 . + 2 ID=cds00004;Parent=mRNA00003;Name=edenprotein.4
```



# GTF: Gene Transfer Formats

GTF es una ligera variación de GFF. Las primeras ocho columnas son iguales. La novena columna tiene una sintaxis diferente.

## Example for GTF file

```
AB000381 Twinscan CDS      380  401  .  +  0 gene_id "001"; transcript_id "001.1";
AB000381 Twinscan CDS      501  650  .  +  2 gene_id "001"; transcript_id "001.1";
AB000381 Twinscan CDS      700  707  .  +  2 gene_id "001"; transcript_id "001.1";
AB000381 Twinscan start_codon 380  382  .  +  0 gene_id "001"; transcript_id "001.1";
AB000381 Twinscan stop_codon 708  710  .  +  0 gene_id "001"; transcript_id "001.1";
```

## GFF for comparison

```
1 | ctg123 . mRNA          1050 9000 . + . ID=mRNA00001;Parent=gene00001;Name=EDEN.1
2 | ctg123 . mRNA          1050 9000 . + . ID=mRNA00002;Parent=gene00001;Name=EDEN.2
3 | ctg123 . mRNA          1300 9000 . + . ID=mRNA00003;Parent=gene00001;Name=EDEN.3
4 |
```



# VCF: Variant Call Format

Archivo de texto para almacenar variantes de secuencia, SNP e InDels. A diferencia de otros formatos, VCF no almacena todos los datos genéticos redundantes que se comparten entre los genomas.

```
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT sample1 sample2 ...
```

Column 1: CHROM – chromosome name

Column 2: POS – position in the chromosome

Column 3: ID – identifier

Column 4: REF – reference base(s) in the reference genome

Column 5: ALT – alternate base(s) in the comparing sequence

Column 6: QUAL – quality score

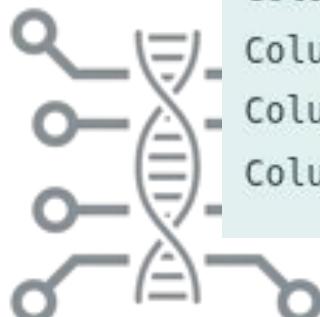
Column 7: FILTER – filter status

Column 8: INFO – additional information

Column 9: FORMAT – genotype information

Column 10: sample-1

Column 11: sample-2 and so on ...



# VCF: Variant Call Format

```
##fileformat=VCFv4.2
##contig=<ID=2,length=51304566>
##INFO=<ID=AC,Number=A,Type=Integer,Description="Allele count in genotypes">
##INFO=<ID=AN,Number=1,Type=Integer,Description="Total number of alleles in called genotypes">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Read Depth">
##FORMAT=<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT SAMPLE1 SAMPLE2 SAMPLE3 SAMPLE4 SAMPLE5 SAMPLE6
2 81170 . C T . . AC=9;AN=7424 GT:DP:GQ 0/0:4:12 0/0:3:9 0/1:1:3 0/1:9:24 1/0:4:12 0/0:5:15
2 81171 . G A . . AC=6;AN=7446 GT:DP:GQ 0/1:4:12 0/0:3:9 0/0:1:3 0/0:9:24 0/1:4:12 0/1:5:15
2 81182 . A G . . AC=5;AN=7506 GT:DP:GQ 0/0:5:15 0/0:4:12 0/0:5:15 0/0:9:24 0/0:4:12 0/0:4:12
2 81204 . T G . . AC=2;AN=7542 GT:DP:GQ 1/0:5:15 0/0:9:27 0/0:10:30 0/0:15:39 0/0:9:27 1/0:13:39
```



# SAM-Sequence Alignment/Map

|  |                |
|--|----------------|
| QHD VN:1.5 SO:coordinate   | Header section |
| @SQ SN:ref LN:45   |                |
| r001 99 ref 7 30 8M2I4M1D3M = 37 39 TTAGATAAAGGATACTG *            |                |
| r002 0 ref 9 30 3S6M1P1I4M * 0 0 AAAAGATAAGGATA *                  |                |
| r003 0 ref 9 30 5S6M * 0 0 GCCTAACGCTAA * SA:Z:ref,29,-,6H5M,17,0; |                |
| r004 0 ref 16 30 6M14N5M * 0 0 ATAGCTTCAGC *                       |                |
| r003 2064 ref 29 17 6H5M * 0 0 TAGGC * SA:Z:ref,9,+,5S6M,30,1;     |                |
| r001 147 ref 37 30 9M = 7 -39 CAGCGGCAT * NM:i:1                   |                |

**Optional fields** in the format of TAG:TYPE:VALUE

**FLAG:** indicates alignment information about the read, e.g. paired, aligned, etc.

**QNAME:** query template name, aka. read ID

**RNAME:** reference sequence name, e.g. chromosome/transcript id

**POS:** 1-based position

**MAPQ:** mapping quality

**CIGAR:** summary of alignment, e.g. insertion, deletion

**TLEN:** the number of bases covered by the reads from the same fragment. Plus/minus means the current read is the leftmost/rightmost read. E.g. compare first and last lines.

**PNEXT:** Position of the primary alignment of the NEXT read in the template. Set as 0 when the information is unavailable. It corresponds to POS column.

**RNEXT:** reference sequence name of the primary alignment of the NEXT read. For paired-end sequencing, NEXT read is the paired read, corresponding to the RNAME column.

**SEQ:** read sequence

**QUAL:** read quality; \* meaning such information is not available



# SAM-Sequence Alignment/Map

## FLAGS

| Bit  | Description   |
|------|---|
| 1    | 0x1 template having multiple segments in sequencing                     |
| 2    | 0x2 each segment properly aligned according to the aligner              |
| 4    | 0x4 segment unmapped  |
| 8    | 0x8 next segment in the template unmapped                               |
| 16   | 0x10 SEQ being reverse complemented                                     |
| 32   | 0x20 SEQ of the next segment in the template being reverse complemented |
| 64   | 0x40 the first segment in the template                                  |
| 128  | 0x80 the last segment in the template                                   |
| 256  | 0x100 secondary alignment   |
| 512  | 0x200 not passing filters, such as platform/vendor quality controls     |
| 1024 | 0x400 PCR or optical duplicate  |
| 2048 | 0x800 supplementary alignment   |

<https://broadinstitute.github.io/picard/explain-flags.html>

