CG3

Bromus erectus grassland

Synonymy

Chalk grassland auct. angl. p.p.; Brometum auct. angl.; Zernetum auct. angl.; Cirsio-Brometum Shimwell 1968a p.p.; Cirsio-Brometum Shimwell 1968a emend. Willems 1978 p.p.

Constant species

Bromus erectus, Carex flacca, Festuca ovina, Lotus corniculatus, Plantago lanceolata, Sanguisorba minor.

Rare species

Aceras anthropophorum, Astragalus danicus, Carex humilis, Hypochoeris maculata, Linum perenne ssp. anglicum, Phyteuma tenerum, Polygala calcarea, Pulsatilla vulgaris, Senecio integrifolius ssp. integrifolius, Thesium humifusum, Thymus pulegioides.

Physiognomy

This community comprises all those swards in which B. erectus makes up more than 10% of the cover and where other grasses of similar physiognomy, notably Brachypodium pinnatum and Avenula pubescens, make a usually negligible contribution. It includes vegetation which is similar in floristics and structure to the Festuca-Avenula grassland as well as some where the dense tussocks of B. erectus have a much more dominating effect. Taking the community as a whole, however, the most obvious general feature is the markedly reduced frequency here of most chamaephytes (Thymus praecox, Hieracium pilosella, Helianthemum nummularium), those hemicryptophytes which cannot produce extensive bushy sprawls of shoots, large rosettes or tall leafy stems (Asperula cynanchica, Koeleria macrantha, Briza media, Scabiosa columbaria, Campanula rotundifolia, Plantago media), certain therophytes (Linum catharticum, Gentianella amarella) and bryophytes (Pseudoscleropodium purum, Homalothecium lutescens and acrocarps of patches of bare soil). The most frequent associates overall are all hemicryptophytes which can persist by depressing or growing through the coarse, bulky foliage of the dominant and its accumulating litter: Sanguisorba minor, Carex flacca, Plantago lanceolata, Cirsium acaule and Lotus corniculatus. Festuca ovina seems to survive the presence of substantial amounts of B. erectus, though it often persists patchily and, in the rankest swards, it is commonly replaced by F. rubra in a tussocky form.

The rare species which are such a distinctive feature of certain kinds of Festuca-Avenula grassland are also, for the most part, reduced here. Some of those which are more frequent and widespread in that community are chamaephytes (Polygala calcarea, Thesium humifusum) or diminutive geophytes (Orchis ustulata, Herminium monorchis, Spiranthes spiralis) which are readily overwhelmed. Others fare better. Phyteuma tenerum, for example, can persist in numbers and with a splendid stature in these coarser swards (e.g. Wells 1975). Senecio integrifolius ssp. integrifolius can also grow tall and perhaps survive here as a short-lived perennial (Smith 1979) and Linum perenne ssp. anglicum may benefit by the long life of its individuals (Ockendon 1968). Hypochoeris maculata has basal leaves which, growing larger, can depress the surrounding herbage (Wells 1976). Pulsatilla vulgaris now survives mainly in this community (Wells & Barling 1971) and has been observed actually to increase in numbers by vegetative growth from lateral buds on the rootstock when B. erectus invades, though with a very marked reduction in the proportion of plants flowering. Much-etiolated survivors seem to be able to resume flowering when grazing is re-instated (Wells 1968). Among the orchids, taller species such as Aceras anthropophorum and Himantoglossum hircinum seem able to persist well and these are sometimes joined by more widespread robust species such as Anacamptis pyramidalis and Gymnadenia conopsea which are generally less common in the short turf of the Festuca-Avenula grassland.

Sub-communities

Typical sub-community: Zerna erecta grassland Wells 1975 p.p. This sub-community includes those swards which are closest to the Festuca-Avenula grassland. B.

erectus and F. ovina (with occasional F. rubra) are present in roughly equal proportions and there are frequently smaller amounts of Koeleria macrantha, Briza media and Avenula pratensis, together with some of the characteristic dicotyledons and bryophytes of that vegetation type. Hemicryptophytes, such as Leontodon hispidus, Scabiosa columbaria and Plantago media continue to occur quite commonly, though there is an obvious reduction in Asperula cynanchica and most of the chamaephytes. Moreover, the sward often has a generally coarser or markedly patchy appearance, being organised around clonal tussocks of B. erectus, between which there are stretches of shorter and more finegrained turf.

Centaurea nigra sub-community: Chalk grassland Tansley & Adamson 1926 p.p. The general floristics here are quite similar to those above but there are some slight shifts in composition and physiognomy. B. erectus is more abundant (generally with more than 50% cover) and the sward is taller and ranker with a further reduction in the frequency and cover of some Festuca-Avenula species. Most obviously, Centaurea nigra becomes constant and there are occasional records for a variety of coarse dicotyledons such as Senecio jacobaea, Rhinanthus minor, Galium mollugo, Achillea millefolium, Daucus carota and Hypochoeris radicata.

Knautia arvensis-Bellis perennis sub-community: Brometum Cornish 1954 p.p.; Zerna erecta grassland Wells 1975 p.p.; Cirsio-Brometum anthoxanthetosum Willems 1978. Again, in this sub-community, B. erectus tends to be much the most abundant species in swards which are rather poor in many Festuca-Avenula plants, but the most noticeable feature is the rise to prominence of species characteristic of ranker mesotrophic grasslands. These include grasses such as Dactylis glomerata, Cynosurus cristatus, Phleum pratense ssp. bertolonii and, less frequently, Holcus lanatus, Lolium perenne, Trisetum flavescens and Agrostis stolonifera and perennial dicotyledons like Prunella vulgaris, Taraxacum officinale agg., Bellis perennis, Trifolium pratense, T. repens and, especially distinctive here, Knautia arvensis. The sprawling legumes Vicia cracca and Lathyrus pratensis are quite common and there are occasional to frequent records for the pauciennials Medicago lupulina, Blackstonia perfoliata and Centaurium erythraea.

Festuca rubra-Festuca arundinacea sub-community. In the tall, rank and much more species-poor swards of this sub-community, B. erectus is generally very much the dominant species, frequently with somewhat smaller amounts of tussocky F. rubra and F. arundinacea. F. ovina and the other fine-leaved grasses of the Festuca-Avenula grassland, together with many of its characteristic dicotyledons are here no more than occasional.

Leontodon hispidus and Centaurea nigra remain common and are here joined by another hemicrypto-phyte, Centaurea scabiosa.

Habitat

Like many species with a Continental distribution through Europe, *Bromus erectus* reaches a general north-western limit in Britain around the Humber–Severn line, extending northwards more locally and, then, often on warmer, south-facing slopes. Within these climatically-related limits, it is typically a dominant in lightly-grazed or ungrazed grasslands on calcareous, base-rich soils mostly, in the south-eastern part of the country, over Chalk and Oolite. Floristic variation within the community seems to be related partly to the degree of expansion of *B. erectus* in the sward and partly to the edaphic conditions prevailing when it invades or extends its cover.

B. erectus is palatable to sheep, cattle and rabbits and, though areas of shorter, mixed and more nutritious turf may be preferred to its coarse and fibrous herbage (Hope-Simpson 1940b), there is little doubt that it is held at low levels by heavy grazing and that, when this is relaxed, it can show a vigorous and rapid vegetative expansion (e.g. Tansley & Adamson 1926, Hope-Simpson 1941b, Watt 1957, 1962, Thomas 1960, 1963, Wells 1967a, 1968, 1971). Established tussocks grow by tillering around their margins and expanding radially and they may eventually coalesce and totally dominate a sward (e.g. Austin 1968a). Many of the stands included here have almost certainly originated in this way with the reduction of grazing of Festuca-Avenula grassland in which B. erectus occurs occasionally at low cover values (and perhaps also from some types of Festuca-Hieracium-Thymus grassland where it is present at very low frequencies). The drop in frequency of many of the characteristic Festuca-Avenula species can be readily understood as a response to the shading effect of the tall and bulky foliage of B. erectus and the accumulation of its coarse and resistant litter (Wells 1968, 1971, Austin 1968a). The differential loss of such species between the different sub-communities (and between different stands of the same sub-community) may also reflect to some degree the extent to which B. erectus expansion has proceeded since the relaxation of grazing. Thus, the swards of the Typical sub-community may have been more recently or heavily grazed than the ranker vegetation of the Centaurea nigra sub-community, though we have no information which might test this view. Also, there may be some local and temporary waning of the dominance of the B. erectus which could complicate matters: concentric rings of the grass are a common sight and Austin (1968a) has shown how tussocks can degenerate centrally, leaving the way open for a resurgence of a more varied calcicolous flora within.

However, some of the floristic differences between the

sub-communities are probably related to edaphic variation. The Typical sub-community, for example, is characteristically a vegetation type of shallower and more calcicolous grey and humic rendzinas (especially over Chalk) while the Knautia-Bellis sub-community is more frequent on the Oolite, where this softer rock often weathers to produce deeper and more mesotrophic brown rendzinas (e.g. Findlay 1976). The Centaurea sub-community may be intermediate in this respect: it is especially typical of deeper colluvial soils towards the eastern end of the South Downs (e.g. Tansley & Adamson 1926). Some of this floristic variation may be inherited from the swards in which B. erectus has expanded because exactly the same edaphically-related contrast is seen within the Festuca-Avenula grassland where it marks the difference between the Cirsium-Asperula and Holcus-Trifolium sub-communities.

How far B. erectus can invade such vegetation types rather than expand within them from established plants is a difficult question. When ungrazed, it can certainly fruit prolifically and, though its seeds are heavy (Lloyd 1964, Lloyd & Pigott 1967), they are awned and readily caught in animal hair or on clothing. Where there are still some grazing animals about and where the sward is not yet too rank, it is possible that some spread may occur by dispersal from fruiting tussocks nearby. The invasion of abandoned arable land, a common feature of the Chalk in certain periods with changes in the agricultural economy, is a more complicated matter. Though B. erectus has an ample food store in its large seeds, it seems strikingly unable to gain a hold where such exposed soils are markedly oligotrophic as, for example, where very shallow rendzinas have been ploughed and left and have lost any nitrogen through leaching (Cornish 1954, Lloyd 1964, Lloyd & Pigott 1967). The addition of nitrogen in field trials has also been shown to produce no response, which suggests that a poor water supply may also be involved (Lloyd & Pigott 1967). Disturbed areas of deeper and more mesotrophic soils, however, can be invaded: at Porton Down, on the Hampshire/Wiltshire border, for example, Wells et al. (1976) reported patchy B. erectus grassland on land ploughed 50-100 years before. These swards seem mostly to be of the kind included here in the Festuca rubra-F. arundinacea sub-community and it is possible that this distinctively rank and species-poor vegetation has developed primarily by such direct invasion of open ground with no resumption of pasturing.

Zonation and succession

Zonations involving the *Bromus* grassland are most often reflections of seral changes related to the intensity of grazing and sometimes complicated histories of ploughing and abandonment. Occasionally, the community may show an abrupt transition to disturbed, oligotrophic soils that cannot, apparently, be invaded

by *B. erectus* (Lloyd 1964, Lloyd & Pigott 1967). More usually, it occurs in irregular patchworks with surviving areas of *Festuca-Avenula* grassland, other rank swards that have developed with a demise of grazing (e.g. the *Brachypodium pinnatum* and *Avenula pubescens* grasslands) and scrub. Such patterns have been described in detail from Lullington Heath in Sussex (Grubb *et al.* 1969), Barton Hills in Bedfordshire (Wells & Morris 1970) and Porton Down (Wells *et al.* 1976) and they are a common feature of many areas of the southern Chalk and Oolite (e.g. Ratcliffe 1977).

In principle, the fact that *B. erectus* will be eaten by stock and rabbits means that its expansion in swards is reversible with a resumption of grazing (Hope-Simpson 1940b). In practice, this may be more complicated. For example, Wells (1971) showed how *B. erectus* maintained its cover in this kind of vegetation over shallow, rendziniform soils with quite heavy grazing, even increasing in extent on deeper loamy soils. It is therefore possible that swards may stabilise with a higher *B. erectus* cover and that really hard grazing, with no alternative, more palatable herbage, is necessary to effect any marked reduction.

The high cost of this kind of pastoral control has focused attention on mowing as an alternative means of retaining or recovering the species-richness and diversity of those calcicolous swards where B. erectus can gain a hold. Wells (1971) demonstrated that cutting in spring, shortly after growth commenced in March, was effective in holding steady or reducing the cover of B. erectus, perhaps by depleting the newly-mobilised food reserves. Regrowth was then poor and other species were able to capitalise on the provision of space. Summer cutting sometimes encouraged the expansion of B. erectus, perhaps because of stimulating the production of tillers when growth was in full swing. Though mowing is unselective (and not always practical on very steep slopes), it can be precisely timed and this may have advantages in allowing selected species to flower and set seed (e.g. Wells 1969, 1971, 1973).

Burning, though it has been more traditionally practised on grasslands with *Brachypodium pinnatum*, has sometimes been used as a means of controlling *B. erectus* which, in dry weather, is readily ignited with its accumulations of litter. It may not always be successful. Indeed, it may stimulate fresh growth in the cleaned sward and, though new shoots may be more eagerly eaten by stock, without grazing the *B. erectus* may simply resume its dominance.

The subsequent successional development of *Bromus* grassland with the continued absence of grazing has never been followed in detail. Though stands of the community often occur in close association with scrub, we do not know how readily shrubs and trees can invade the ranker swards. Certainly, some stands, after rapid establishment following myxomatosis, seem to have

remained substantially unchanged for 30 years. Large populations of field voles (*Microtus agrestis*) often become established in such litter-choked tussocky grasslands and these may play an important role in eating and barking seedlings of woody species (Smith 1980). Deliberate or accidental burning may also repeatedly set back any invasion and help maintain the vegetation as a plagioclimax. Where colonisation by *B. erectus* is more gradual or patchy, it is possible that shrubs and trees may become established in the more open turf or on anthills which are often abandoned as they become increasingly shaded by the tall grasses (Wells *et al.* 1976, King 1977c).

Distribution

Bromus grasslands occur throughout the range of the species but they are especially frequent over the Chalk in the North and South Downs, through Hampshire and Wiltshire and into the Chilterns, on the Oolite of the Cotswolds and Northamptonshire and over the Magnesian Limestone of West Yorkshire. The Typical and Centaurea sub-communities occur throughout the range while the Knautia-Bellis and Festuca rubra-F. arundinacea sub-communities have been more frequently encountered in Wiltshire. B. erectus also occurs with Brachypodium pinnatum in mixed swards, especially on the Cotswold and Northamptonshire Oolite.

Affinities

Traditionally, the more calcicolous kinds of *Bromus*-dominated grasslands in Britain have been treated as part of a broadly-defined 'Chalk grassland' or its phytosociological equivalent, the *Cirsio-Brometum* Shimwell 1968a emend. Willems 1978. Where ranker swards have been described, they have usually been separated off into various kinds of *Bromus* grassland (e.g. Cornish 1954, Wells 1975, Wells et al. 1976, Smith 1980). Here, all such swards have been united within a single community which, though it has a reduction overall in the representation of calcicoles, still has clear affinities with the Mesobromion. The community can be regarded as the major natural ungrazed counterpart of the mainstream *Festuca-Avenula* grassland over most of the southern part of its range.

Similar Mesobromion grasslands in which *B. erectus* plays an important role have been described from France (e.g. Allorge 1921–2), Belgium (LeBrun *et al.* 1949), The Netherlands (Westhoff & den Held 1969) and Germany (e.g. Ellenberg 1978, Oberdorfer 1978). On the Continent, however, this grass is also a prominent species within the xeric swards of the Xerobromion and certain of the steppe-grasslands of the Festucetalia valesiacae.

Floristic table CG3

Sanguisorba minor IV (1-6) V (2-6) IV (1-5) V (2-6) V (1-6) Carex flacca IV (1-6) IV (1-5) II (1-6) III (1-8) III (1-8) III (1-8) II (1-9) II (1-10) II (1-10) II (1-10) II (1-10) II (1-10) II (1-10)<		a	b	c	đ	3
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Fissidens cristatus II (1-3) I (2) I (1-2) I (1-3) I (1-3) Centaurea nigra I (1-4) IV (1-4) I (2-4) II (1-4) II (1-4) Thymus praecox II (1-6) III (2-5) II (1-5) I (2) II (1-6) Polygala vulgaris I (1-3) II (1-5) I (1) I (1) I (1-5) Senecio jacobaea I (1-2) II (1-3) I (2-3) I (2) I (1-3) Rhinanthus minor I (1-4) II (2-3) I (5) I (1-2) I (1-5) Galium mollugo I (1-3) II (2-5) I (1-3) I (1-2) I (1-5) Achillea millefolium I (2-4) II (1-4) I (2-3) I (2-3) I (1-2) I (1-4) Daucus carota I (1-3) II (1-4) I (2-3) I (2-3) I (1-4) Hypochoeris radicata II (1-4) II (1-4) IV (1-4) I (1-2) II (1-4) Prunella vulgaris III (1-4) II (1-4) IV (1-4) I (1-3) III (1-4) Trifolium pratense <td>Homalothecium lutescens</td> <td>II (1–4)</td> <td>I (1–3)</td> <td>I (1)</td> <td>I (2)</td> <td>I (1-4)</td>	Homalothecium lutescens	II (1–4)	I (1–3)	I (1)	I (2)	I (1-4)
Centaurea nigra I (1-4) IV (1-4) I (2-4) II (1-4) II (1-4) II (1-4) II (1-5) II (1-6) III (1-6) III (2-5) II (1-5) I (2) II (1-6) Polygala vulgaris I (1-3) II (1-5) I (1) I (1) I (1) I (1-5) Senecio jacobaea I (1-2) II (1-3) I (2-3) I (2) I (1-3) Rhinanthus minor I (1-4) II (2-3) I (5) I (1-5) I (1-6)	Asperula cynanchica	II (1–2)	I (2–4)	I (1)		I (1-4)
Thymus praecox II (1-6) III (2-5) II (1-5) I (2) III (1-6) Polygala vulgaris I (1-3) II (1-5) I (1) I (1) I (1-5) Senecio jacobaea I (1-2) II (1-3) I (2-3) I (2) I (1-3) Rhinanthus minor I (1-4) II (2-3) I (5) I (1-2) I (1-5) Galium mollugo I (1-3) II (2-5) I (1-3) I (1-2) I (1-5) Achillea millefolium I (2-4) II (1-4) I (2-3) I (2-3) I (1-2) I (1-5) Achillea millefolium I (2-4) II (1-4) I (2-3) I (1-2) I (1-5) Achillea millefolium I (2-4) II (1-4) I (2-3) I (1-2) I (1-5) Achillea millefolium I (1-3) II (1-4) I (2-3) I (1-2) I (1-5) Achillea millefolium I (1-3) II (1-4) I (2-3) I (1-2) I (1-4) Daucus carota I (1-1) II (1-4) I (1-4) I (1-4) I (1-4) I (1-4) I (1-4)	Fissidens cristatus	II (1–3)	I (2)	I (1–2)		I (1-3)
Polygala vulgaris	Centaurea nigra	I (1-4)	IV (1-4)	I (2-4)	II (1-4)	II (1–4)
Senecio jacobaea I (1-2) II (1-3) I (2-3) I (2) I (1-3) Rhinanthus minor I (1-4) II (2-3) I (5) I (1-2) I (1-5) Galium mollugo I (1-3) II (2-5) I (1-3) I (1-2) I (1-5) Achillea millefolium I (2-4) II (1-4) I (2-3) I (2-3) I (1-4) Daucus carota I (1-3) II (1-4) I (1-2) I (1-4) Hypochoeris radicata II (1-4) IV (1-4) I (1-2) I (1-4) Prunella vulgaris III (1-4) II (1-4) IV (1-4) I (1-3) III (1-4) Prunella vulgaris II (1-4) II (1-4) IV (1-4) I (1-3) III (1-4) Prunella vulgaris II (1-4) II (2-4) IV (1-3) II (1-3) III (1-3) Bellis perennis I (1-3) I (2-4) IV (1-3) II (1-3) II (1-4) Trifolium pratense II (1-8) II (1-5) V (1-6) I (1) II (1-8) Medicago lupulina II (1-3) I (2-4) <t< td=""><td>Thymus praecox</td><td>II (1–6)</td><td>III (2–5)</td><td>II (1–5)</td><td>I (2)</td><td>II (1–6)</td></t<>	Thymus praecox	II (1–6)	III (2–5)	II (1–5)	I (2)	II (1–6)
Rhinanthus minor I (1-4) II (2-3) I (5) I (1-2) I (1-5) Galium mollugo I (1-3) II (2-5) I (1-3) I (1-2) I (1-5) Achillea millefolium I (2-4) II (1-4) I (2-3) I (2-3) I (1-4) Daucus carota I (1-3) II (1-4) I (2-3) I (1-2) I (1-4) Hypochoeris radicata III (1-4) II (1-4) IV (1-4) I (1-3) III (1-4) Prunella vulgaris III (1-4) II (1-4) IV (1-4) I (1-3) III (1-4) Taraxacum officinale agg. I (2-3) I (2) IV (1-3) II (1-3) II (1-3) Bellis perennis I (1-3) I (2-4) IV (1-3) I (1) II (1-4) Trifolium pratense II (1-8) II (1-5) V (1-6) I (1) II (1-8) Medicago lupulina II (1-4) I (2-4) V (1-5) II (1-8) II (1-5) Knautia arvensis I (1-3) I (2-3) IV (1-3) II (1-8) II (1-8) Viola hirta <td< td=""><td>Polygala vulgaris</td><td>I (1–3)</td><td>II (1-5)</td><td>I (1)</td><td>I (1)</td><td>I (1-5)</td></td<>	Polygala vulgaris	I (1–3)	II (1-5)	I (1)	I (1)	I (1-5)
Galium mollugo I (1-3) II (2-5) I (1-3) I (1-2) I (1-5) Achillea millefolium I (2-4) III (1-4) I (2-3) I (2-3) I (1-4) Daucus carota I (1-3) III (1-4) I (2-3) I (1-2) I (1-4) Hypochoeris radicata III (1-4) IV (1-4) I (1-2) I (1-4) Prunella vulgaris III (1-4) II (1-4) IV (1-4) I (1-3) III (1-4) Taraxacum officinale agg. I (2-3) I (2) IV (1-3) II (1-3) II (1-3) Bellis perennis I (1-3) I (2-4) IV (1-3) I (1) II (1-4) Trifolium pratense II (1-8) II (1-5) V (1-6) I (1) II (1-8) Medicago lupulina II (1-4) I (2-4) V (1-5) II (1-5) II (1-5) Knautia arvensis I (1-3) I (2-3) IV (1-3) II (1-8) II (1-8) Viola hirta II (1-5) II (1-5) III (1-5) II (1-6) II (1-6) Vicia cracca I (1-3) <	Senecio jacobaea	I (1–2)	II (1–3)	I (2–3)	I (2)	I (1-3)
Achillea millefolium I (2-4) II (1-4) I (2-3) I (2-3) I (1-4) Daucus carota I (1-3) II (1-4) I (1-4) I (1-2) I (1-4) Hypochoeris radicata II (1-4) II (1-4) IV (1-4) I (1-3) III (1-4) Prunella vulgaris III (1-4) II (1-4) IV (1-4) I (1-3) III (1-4) Taraxacum officinale agg. I (2-3) I (2) IV (1-3) II (1-3) II (1-3) Bellis perennis I (1-3) I (2-4) IV (1-3) I (1) II (1-4) Trifolium pratense II (1-8) II (1-5) V (1-6) I (1) II (1-8) Medicago lupulina II (1-4) I (2-4) V (1-5) II (1-5) II (1-5) Knautia arvensis I (1-3) I (2-3) IV (1-3) II (1-8) II (1-8) Dactylis glomerata II (1-5) II (1-5) III (1-6) III (1-6) II (1-7) Viola hirta II (1-3) I (2-4) III (1-5) I (5) II (1-4) Vicia cracca I (1-3) I (3) III (1-4) I (1) I (1-4) </td <td>Rhinanthus minor</td> <td>I (1–4)</td> <td>II (2–3)</td> <td>I (5)</td> <td></td> <td>I (1-5)</td>	Rhinanthus minor	I (1–4)	II (2–3)	I (5)		I (1-5)
Daucus carota I (1-3) II (1-4) I (1-2) I (1-4) Hypochoeris radicata II (1-4) II (1-4) IV (1-4) I (1-3) III (1-4) Prunella vulgaris III (1-4) II (1-4) IV (1-4) I (1-3) III (1-4) Taraxacum officinale agg. I (2-3) I (2) IV (1-3) II (1-3) II (1-3) Bellis perennis I (1-3) I (2-4) IV (1-3) I (1) II (1-4) Trifolium pratense II (1-8) II (1-5) V (1-6) I (1) II (1-8) Medicago lupulina II (1-4) I (2-4) V (1-5) II (1) II (1-5) Knautia arvensis I (1-3) I (2-3) IV (1-3) II (1-8) II (1-3) Dactylis glomerata II (1-5) II (1-5) III (1-4) II (1-8) II (1-8) Viola hirta II (1-5) I (2-4) III (1-5) I (5) II (1-5) Vicia cracca I (1-3) I (3) III (1-4) I (1) I (1-4)	Galium mollugo	I (1–3)	II (2–5)	I (1–3)	I (1-2)	I (1-5)
Hypochoeris radicata	Achillea millefolium	I (2–4)	II (1–4)	I (2-3)	I (2-3)	I (1-4)
Prunella vulgaris III (1-4) II (1-4) IV (1-4) I (1-3) III (1-4) Taraxacum officinale agg. I (2-3) I (2) IV (1-3) II (1-3) II (1-3) Bellis perennis I (1-3) I (2-4) IV (1-3) I (1) II (1-4) Trifolium pratense II (1-8) II (1-5) V (1-6) I (1) II (1-8) Medicago lupulina II (1-4) I (2-4) V (1-5) II (1-5) Knautia arvensis I (1-3) I (2-3) IV (1-3) II (1-8) Dactylis glomerata II (1-5) II (1-5) III (1-4) II (1-8) II (1-8) Viola hirta II (1-5) I (2-4) III (1-5) I (5) II (1-5) Vicia cracca I (1-3) I (3) III (1-4) I (1) I (1-4)	Daucus carota	I (1–3)	II (1–4)		I (1-2)	I (1-4)
Taraxacum officinale agg. I (2-3) I (2) IV (1-3) II (1-3) II (1-3) Bellis perennis I (1-3) I (2-4) IV (1-3) I (1) II (1-4) Trifolium pratense II (1-8) II (1-5) V (1-6) I (1) II (1-8) Medicago lupulina II (1-4) I (2-4) V (1-5) II (1-5) II (1-5) Knautia arvensis I (1-3) I (2-3) IV (1-3) II (1-3) II (1-8) II (1-8) Dactylis glomerata II (1-5) II (1-5) III (1-5) III (1-8) II (1-8) II (1-8) Viola hirta II (1-5) I (2-4) III (1-5) I (5) II (1-5) Vicia cracca I (1-3) I (3) III (1-4) I (1) I (1-4)	Hypochoeris radicata		II (1 -4)			I (1–4)
Bellis perennis I (1-3) I (2-4) IV (1-3) I (1) II (1-4) Trifolium pratense II (1-8) II (1-5) V (1-6) I (1) II (1-8) Medicago lupulina II (1-4) I (2-4) V (1-5) II (1-5) Knautia arvensis I (1-3) I (2-3) IV (1-3) II (1-3) Dactylis glomerata II (1-5) II (1-5) III (1-4) II (1-8) II (1-8) Viola hirta II (1-5) I (2-4) III (1-5) I (5) II (1-5) Vicia cracca I (1-3) I (3) III (1-4) I (1) I (1-4)	Prunella vulgaris	III (1–4)	II (1 -4)	IV (1-4)	I (1-3)	III (1-4)
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Taraxacum officinale agg.		I (2)	IV (1–3)	II (1-3)	II (1–3)
Medicago lupulina II (1-4) I (2-4) V (1-5) II (1-5) Knautia arvensis I (1-3) I (2-3) IV (1-3) II (1-3) Dactylis glomerata II (1-5) II (1-5) III (1-4) II (1-8) II (1-8) Viola hirta II (1-5) I (2-4) III (1-5) I (5) II (1-5) Vicia cracca I (1-3) I (3) III (1-4) I (1) I (1-4)	Bellis perennis	I (1–3)	I (2–4)	IV (1–3)	I (1)	II (1–4)
Knautia arvensis I (1-3) I (2-3) IV (1-3) II (1-3) Dactylis glomerata II (1-5) II (1-5) III (1-4) II (1-8) II (1-8) Viola hirta II (1-5) I (2-4) III (1-5) I (5) II (1-5) Vicia cracca I (1-3) I (3) III (1-4) I (1) I (1-4)	Trifolium pratense	II (1–8)	II (1–5)	V (1-6)	I (1)	II (1–8)
Dactylis glomerata II (1-5) II (1-5) III (1-4) II (1-8) II (1-8) Viola hirta II (1-5) I (2-4) III (1-5) I (5) II (1-5) Vicia cracca I (1-3) I (3) III (1-4) I (1) I (1-4)	Medicago lupulina	II (1–4)	I (2–4)	V (1-5)		II (1-5)
Viola hirta II (1-5) I (2-4) III (1-5) I (5) II (1-5) Vicia cracca I (1-3) I (3) III (1-4) I (1) I (1-4)	Knautia arvensis	I (1–3)	I (2–3)	IV (1-3)		II (1-3)
$Vicia\ cracca \hspace{1.5cm} I\ (1-3) \hspace{1.5cm} I\ (3) \hspace{1.5cm} III\ (1-4) \hspace{1.5cm} I\ (1) \hspace{1.5cm} I\ (1-4)$	Dactylis glomerata		, ,	, ,	II (1–8)	II (1–8)
	Viola hirta	II (1–5)	I (2-4)	III (1-5)	I (5)	II (1–5)
<i>Blackstonia perfoliata</i> I (1–4) I (2–3) III (1–3) I (1) I (1–4)	Vicia cracca			, ,		I (1-4)
	Blackstonia perfoliata	I (1–4)	I (2-3)	III (1–3)	I (1)	I (1–4)

Trifolium repens	I (1-3)	I (1-3)	III (1-5)	I (1–4)	I (1-5)
Cynosurus cristatus	I (1-5)	I (1–6)	III (2-5)		I (1–6)
Centaurium erythraea	I (1-3)	I (3)	III (1–3)		I (1-3)
Phleum pratense bertolonii	I (1–2)	I (2–4)	III (1–4)		I (1–4)
Holcus lanatus	I (1-5)	I (2-5)	II (1–4)	I (2–6)	I (1–6)
Lolium perenne	I (1)	I (2–6)	II (1–4)	I (1–3)	I (1–6)
Ranunculus bulbosus	I (1–4)	I (2)	II (1–3)	I (1)	I (1–4)
Cerastium fontanum	I (1-3)	I (2-3)	II (1–3)	I (1)	I (1–3)
Trisetum flavescens	I (1–6)	I (2-5)	II (1–4)	I (1)	I (1-6)
Lathyrus pratensis	I (1)	I (3)	II (1–3)	I (1)	I (1-3)
Primula veris	I (1–2)	I (1–6)	II (1–3)	I (2-3)	I (1–6)
Anthyllis vulneraria	I (1–3)	I (1–4)	II (1–3)	I (1)	I (1–4)
Leucanthemum vulgare	I (1–4)	I (1–6)	II (1-3)		I (1–6)
Agrostis stolonifera	I (1-3)	I (2-5)	II (1-6)		I (1–6)
Ononis spinosa	I (1-3)		II (1-3)	I (1)	I (1-3)
Cirsium eriophorum			II (1-3)		I (1-3)
Festuca rubra	II (2-9)	I (2–8)	II (4–6)	IV (1-6)	II (1-9)
Festuca arundinacea	I (1–6)		I (1)	III (2–6)	I (1-6)
Centaurea scabiosa	I (1–4)	II (2–4)	I (1–4)	III (1–6)	II (1–6)
Cirsium acaule	IV (1–8)	III (1–7)	IV (1–6)	II (1–3)	III (1-8)
Briza media	III (1–7)	IV (1–6)	IV (1–6)	I (1)	III (1-7)
Koeleria macrantha	III (1-5)	III (1–5)	II (1-4)	II (1–4)	III (1-5)
Pimpinella saxifraga	III (1–4)	III (1–4)	II (1–3)	II (1 -4)	III (1–4)
Avenula pratensis	III (1–6)	II (1-4)	II (2–3)	III (1-5)	II (1–6)
Linum catharticum	III (1–4)	II (1–3)	III (1-4)	I (2)	II (1–4)
Helianthemum nummularium	II (1–6)	II (1–6)	II (1–6)	III (1 -4)	II (1–6)
Plantago media	III (1–5)	I (1–4)	III (1–4)	I (1-2)	II (1-5)
Scabiosa columbaria	III (1–4)	III (1–4)	I (1-2)	I (1–4)	II (1–4)
Galium verum	II (1–4)	II (1–4)	II (1–4)	I (1-3)	II (1–4)
Hieracium pilosella	II (1–4)	I (2-4)	II (1-6)	I (1)	II (1–6)
Carex caryophyllea	II (1–4)	I (1–2)	II (1-5)	I (1-3)	II (1-5)
Hippocrepis comosa	II (1–5)	II (14)	I (3)	I (1-3)	II (1-5)
Euphrasia officinalis agg.	II (1–4)	I (2-3)	II (1–4)		II (1-4)
Avenula pubescens	II (1–7)	I (1-5)		II (1 -4)	II (1–7)
птении риссиень	(- ')	- ()		()	

Floristic table CG3 (cont.)

	a	b
Brachypodium pinnatum	I (4)	I (2-4)
Picris hieracioides	I (1-3)	I (1)
Weissia cf. microstoma	I (1-3)	I (1)
Campylium chrysophyllum	I (1-5)	I (1)
Crataegus monogyna sapling	I (1-2)	I (1-3)
Filipendula vulgaris	I (1-7)	I (1-3)
Tragopogon pratensis	I (1–3)	I (3)
Campanula glomerata	I (1-4)	I (2-3)
Calliergon cuspidatum	I (2–6)	I (4)
Anthoxanthum odoratum	I (1)	I (2-5)
Agrimonia eupatoria	I (1)	I (3)
Phleum pratense pratense	I (3)	I (2-3)
Brachypodium sylvaticum	I (2-4)	I (2-5)
Potentilla reptans	I (1-3)	I (3)
Ctenidium molluscum	I (2–8)	I (2-4)
Gentianella amarella	I (1-3)	I (3)
Thymus pulegioides	I (1-4)	I (3-5)
Hypnum cupressiforme	I (1-5)	I (3)
Cruciata laevipes	I (1)	I (3)
Anacamptis pyramidalis	I (1-3)	I (1-4)
Leontodon taraxacoides	I (4)	I (2-3)
Ophrys apifera	I (2)	I (1-2)
Gymnadenia conopsea	I (1)	I (1-3)
Ononis repens	I (2-5)	I (3-5)
Veronica chamaedrys	I (1)	I (1-3)
Thesium humifusum	I (3)	I (2)
Origanum vulgare	I (4–5)	I (2-5)
Pastinaca sativa	I (1–2)	I (2-4)
Clematis vitalba	I (1-3)	I (3-4)
Cirsium arvense	I (1-3)	I (2-3)
Ulex europaeus	I (1)	I (1)
Hypochoeris maculata	I (1-2)	I (1-2)

c	d	3
I (2–4)	I (4)	I (2-4)
I (2)	I (1-2)	I (1-3)
I (1)	I (1–3)	I (1-3)
I (1–2)	I (2-3)	I (1-5)
I (1)	I (1-2)	I (1-3)
I (1–3)	I (3)	I (1–7)
I (1)	I (1)	I (1-3)
I (2)	I (3)	I (1-4)
I (1–4)	I (2)	I (1–6)
I (1–3)	I (1)	I (1-5)
I (1-2)	I (1)	I (1-3)
I (1)	I (1)	I (1-3)
I (3)	I (4)	I (2-5)
I (1)		I (1-3)
I (2-5)		I (2–8)
I (3)		I (1-3)
I (2-3)		I (1-5)
I (1)		I (1-5)
I (1)		I (1-3)
I (1–3)		I (1–4)
I (1–6)		I (1-6)
I (1–3)		I (1-3)
I (1-2)		I (1-3)
I (2)		I (2-5)
I (1)		I (1-3)
I (1)		I (1–3)
I (1)		I (1-5)
I (1)		I (1–4)
I (1)		I (1–4)
I (4)		I (1–4)
I (1)		I (1)
I (1-2)		I (12)

Number of species/sample	21 (4–35)	22 (9–36)
Number of samples	105	76
Ranunculus repens	-	I (2-4)
Onobrychis viciifolia		I (3–6)
Rumex acetosa		I (3)
Dactylorhiza fuchsii		I (1-3)
Clinopodium vulgare		I (2-4)
Fraxinus excelsior sapling		I (1)
Convolvulus arvensis		I (3)
Pulsatilla vulgaris	I (2-5)	
Dicranum scoparium	I (1)	
Arctium minus agg.	I (1–3)	
Coeloglossum viride	I (1–3)	
Rhytidiadelphus triquetrus	I (1–4)	
Crepis capillaris	I (1-2)	
Rosa canina agg.	I (1)	I (1-3)
Rhytidiadelphus loreus	I (4)	I (3-5)
Agrostis capillaris	I (5)	I (3–4)
Carlina vulgaris	I (1-3)	I (2-5)
Reseda lutea		I (2)
Carduus nutans		I (1)
Poa pratensis		I (3)
Arrhenatherum elatius		I (3–8)
Orobanche elatior		I (2)
Phyteuma tenerum	I (2-3)	I (1-4)
Danthonia decumbens	I (1-3)	

a Typical sub-community

b Centaurea nigra sub-community

c Knautia arvensis-Bellis perennis sub-community

d Festuca rubra-Festuca arundinacea sub-community

³ Bromus erectus grassland (total)

I (1)	I (1)	I (1-3)
	I (3–4)	I (1–4)
I (1)	I (1)	I (1–2)
I (2)	I (1–7)	I (1–8)
I (1-3)	I (1)	I (1-3)
I (1)	I (1)	I (1)
I (1)	I (1)	I (1-2)
		I (1-5)
		I (3-5)
		I (3-5)
		I (1-3)
I (1)		I (1-2)
I (1)		I (1-4)
I (1-2)		I (1-3)
I (1)		I (1-3)
I (1-2)		I (1-2)
` ,	I (1-5)	I (1-5)
I (1)	` ,	I (1–3)
I (1)		I (1)
I (2–3)		I (2-4)
I (1)		I (1–3)
I (1)		I (1-3)
I (2)		I (2–6)
I (1)		I (1–4)
34	48	263
28 (6-39)	13 (6–22)	20 (4–39)



