M19

Calluna vulgaris-Eriophorum vaginatum blanket mire

Synonymy

Eriophoretum vaginati Smith & Moss 1903, Lewis 1904, Lewis & Moss 1911, Moss 1913, Adamson 1918, Watson 1932, Tansley 1939, all p.p.; Mixed moor Pearsall 1938; Sphagnum community, Type B 'Moss' Pearsall 1941; Calluna & Calluna-Eriophorum Moss Pearsall 1941; Betula nana bogs Poore & McVean 1957; Blanket bogs Ratcliffe 1959; Calluneto-Eriophoretum McVean & Ratcliffe 1962, Eddy et al. 1969, Birks 1973, Meek 1976, Evans et al. 1977; Empetreto-Eriophoretum McVean & Ratcliffe 1962; Vaccinio-Ericetum tetralicis Moore 1962, Birse & Robertson 1976, Dierssen 1982; Trichophoreto-Eriophoretum Eddy et al. 1969 p.p.; Eriophorum vaginatum bog Edgell 1969 p.p.; Erica tetralix-Vaccinium oxycoccos & Juncus squarrosus-Deschampsia flexuosa Series Tallis 1969 p.p.; Rhytidiadelphus loreus-Sphagnum fuscum Community Birse & Robertson 1976; Vaccinio-Eriophoretum Hill & Evans 1978 p.p.; Mire noda 10 & 11 Daniels 1978; Rhytidiadelpho-Sphagnetum fusci Birse 1980; Erico-Sphagnetum papillosi Moore (1964) 1968 emend. Birse 1980 p.p.; Calluna-Pleurozium & Calluna-Cladonia noda Bignal & Curtis 1981.

Constant species

Calluna vulgaris, Eriophorum angustifolium, E. vaginatum, Rubus chamaemorus, Pleurozium schreberi, Sphagnum capillifolium.

Rare species

Arctostaphylos alpinus, Betula nana, Vaccinium microcarpum, Kiaeria starkei.

Physiognomy

The Calluna vulgaris-Eriophorum vaginatum mire comprises blanket bog vegetation that is generally dominated by mixtures of Eriophorum vaginatum and ericoid sub-shrubs. Sphagna can be prominent over wetter ground but typically this element is not so rich or luxuriant as in the Scirpus-Eriophorum or Erica-Sphag-

num mires. And the true hummock/hollow relief that can commonly be found in those communities is only rarely developed here, though the surface of the ground is often uneven because of the marked tussockiness of E. vaginatum, a structural feature of some importance for the strong contingent of hypnoid mosses, which constitute a further distinctive component over drier surfaces.

E. vaginatum is consistently more important here than in the Scirpus-Eriophorum and Erica-Sphagnum mires: among our Sphagnetalia communities, it shows a peak of abundance in this kind of vegetation and in the Eriophorum mire, which seems often to have been derived from it by impoverishment of the associated flora and with which it was often grouped in early schemes under the heading of Eriophoretum vaginati (e.g. Smith & Moss 1903, Lewis 1904a, b, Lewis & Moss 1911, Moss 1913, Adamson 1918, Watson 1932, Tansley 1939). E. vaginatum is usually at least a co-dominant here, contributing a dull-green colour to the vegetation through much of the year and giving it a particularly striking aspect in June with its cottony fruits. The tussocks, which grow by intravaginal tillering, acquire a robust hemispherical shape and can come to stand proud of the bog surface by 2-3 dm, crowned by the densely-packed living shoots and the remains of the foliage, which dies back annually by February but persists intact for many years (Goodman & Perkins 1968, Polozova 1970, Wein 1973, Chapin et al. 1979). The tussocks thus provide a relatively dry microhabitat, though one which is heavily shaded until senescence sets in, which, under normal circumstances, may not be for many decades (Polozova 1970).

The proportion of *E. vaginatum* to the ericoid subshrubs is very variable and, though it shows some consistency in the different sub-communities, it is very much affected by certain kinds of treatment, particularly by burning and grazing which can help convert the vegetation into the *Eriophorum* mire, where the cotton-grass is overwhelmingly dominant and the sub-shrubs of much more patchy occurrence. Variation between the

two communities is continuous and replacement of the richer by the more impoverished can be temporary, so transitions are common. In general, however, the two can be separated by the very frequent occurrence here of Calluna, Vaccinium myrtillus and Empetrum nigrum ssp. nigrum and, at higher altitudes, of the Arctic-Alpine V. vitis-idaea, V. uliginosum and E. nigrum ssp. hermaphroditum. Overall, Calluna is the most common co-dominant of E. vaginatum, but diverse mixtures of these species occur very frequently and typically the subshrubs are so abundant and so vigorous, especially over the drier ground, as to give the vegetation the appearance of a heathy moorland, an aspect recognised in Pearsall's (1938, 1968) epithet of 'mixed moor' and in the names Calluneto-Eriophoretum and Empetreto-Eriophoretum coined by McVean & Ratcliffe (1962). Two other woody Arctic-Alpines, Arctostaphylos alpinus and Betula nana, both national rarities, can also be found here, preferentially among our Sphagnetalia mires, though they are very local and generally both rather inconspicuous, Arctostaphylos being a prostrate plant and Betula usually growing low and gnarled.

The distinctly montane character of this element in the vegetation provides a sharp contrast with both the *Scirpus-Eriophorum* and *Erica-Sphagnum* mires, something reinforced by the absence here of *Myrica gale*, essentially a lowland plant. One further difference, which perhaps reflects the generally drier nature of the peats in this community, is the rather restricted occurrence of *Erica tetralix*: this is commoner in stands to the west and at usually lower altitudes, as in parts of Wales (Ratcliffe 1959a, Tallis 1969, Edgell 1969), south-west Scotland (Birse & Robertson 1976, Birse 1980) and through Strathclyde (Bignal & Curtis 1981), where its frequency helps define the *Erica* sub-community, but it is distinctly local through much *Calluna-Eriophorum* mire.

Further contrasts with these other kinds of ombrogenous mire can be seen among the vascular associates. Compared with the strongly oceanic blanket bog vegetation of the Scirpus-Eriophorum mire, Scirpus cespitosus often plays a subordinate role here, Molinia caerulea is strikingly scarce and herbs such as *Potentilla erecta*, Polygala serpyllifolia and Pedicularis sylvatica are very infrequent. The first two tend to follow E. tetralix in their pattern of occurrence, thus helping to give the *Erica* sub-community a transitional floristic character. It is there, too, that Narthecium ossifragum and Drosera rotundifolia, which can be so prominent in the wetter lawns of both the Scirpus-Eriophorum and Erica-Sphagnum mires, have most of their relatively sparse occurrences in this community. Andromeda polifolia and Vaccinium oxycoccos, both good preferentials for the Erica-Sphagnum mire, are very uncommon here, although high-altitude Calluna-Eriophorum mire provides the British locus for the rare small cranberry V. microcarpum.

On the positive side, the commonest vascular associates in the community are *Eriophorum angustifolium*, a very frequent plant throughout British Oxycocco-Sphagnetea vegetation, and, much more distinctive here, Rubus chamaemorus. This Arctic-Subarctic herb is strongly preferential to the Calluna-Eriophorum mire, though it is not universally present: it is decidedly scarce in the lower-altitude Erica sub-community and sometimes of patchy occurrence elsewhere, being affected by particular treatments. Early or late sampling may also under-estimate its abundance, because it is not evergreen: it passes the winter as dormant buds brought close to the peat surface by rhizome extension, does not put up its shoots until May and shows early senescence of its aerial parts, with their quick death in the first frosts of the autumn. When in bloom, in late May and early June, the large white flowers provide a delightful relief to what is for the most part a rather dreary scene, but flowering is somewhat sporadic and local and, since the flowering period coincides with lactation in deer and hill-sheep, the flowers often get eaten off providing a nutritious bite (Taylor 1971).

Few other features of the vascular flora are distinctive, though *Deschampsia flexuosa* and *Juncus squarrosus* occur occasionally throughout and each can be locally abundant and, at higher altitudes, *Carex bigelowii* becomes frequent. *Nardus stricta*, *Carex nigra*, *Melampyrum pratense* and *Galium saxatile* can be found at low frequencies and the community also provides an occasional locus for the Arctic-Subarctic *Cornus suecica* and the Northern Montane *Listera cordata*. By and large, however, herbs here are few in number and occur as scattered individuals.

The bryophyte flora, by contrast, is rich and often extensive, frequently covering more than 50% of the ground, a further difference between this community and the *Eriophorum* mire. As noted above, though, varied and luxuriant carpets of Sphagna are not the rule in this community. Sphagnum capillifolium is sufficiently frequent throughout to qualify as a constant and it can be locally abundant, but even this species is somewhat patchy and others, like S. papillosum, S. subnitens and S. tenellum, prominent in the wet lawns of other ombrogenous mires, and the semi-aquatics S. cuspidatum and S. auriculatum, which can figure elsewhere in transitions to bog pools, are generally uncommon and rather uneven in their occurrence. Some of these species are better represented in the transitional vegetation of the Erica sub-community, where there can be a semblance of hummock/hollow relief; and some higher-altitude Calluna-Eriophorum mires also show locally extensive Sphagnum carpets, in which the montane S. fuscum has its best representation among British Sphagnetalia communities. In some places, too, generally in the Erica subcommunity, though also in the *Empetrum* sub-community, *S. recurvum* can be prominent, perhaps in response to some soligenous enrichment.

Typically, however, it is hypnaceous mosses which provide the consistency to the bryophyte element in the Calluna-Eriophorum mire. Among these, Pleurozium schreberi, Rhytidiadelphus loreus, Hypnum cupressiforme/jutlandicum and Plagiothecium undulatum are all very frequent throughout, with Hylocomium splendens becoming common at higher altitudes, and mixtures of these can form extensive mats over drier areas of the ground, particularly over the tops of old E. vaginatum hummocks and among the stools of sub-shrubs where the bushes have opened up somewhat. Other mosses recorded frequently are Dicranum scoparium, Polytrichum commune and Aulacomnium palustre, with Racomitrium lanuginosum, Polytrichum alpestre and Pohlia nutans attaining occasional levels in some of the subcommunities and Campylopus paradoxus and Rhytidiadelphus squarrosus being found sparsely throughout. High-altitude stands sometimes have the rare montane moss Kiaeria starkei.

Leafy hepatics are typically not so conspicuous as they are in more oceanic ombrogenous mires and some of the most characteristic species there are either absent from this community (*Pleurozia purpurea*) or much less common (*Mylia anomala*, *Odontoschisma sphagni*). But others occur fairly frequently, such as *Lophozia ventricosa*, *Barbilophozia floerkii*, *Diplophyllum albicans*, *Calypogeia fissa*, *C. muellerana*, *C. trichomanis*, *Cephalozia bicuspidata*, *Mylia taylori* and, particularly at higher altitudes, *Ptilidium ciliare*, and some of these can attain a measure of local abundance.

Finally, among the general features of the community, there is the frequent occurrence of lichens, especially in the higher-altitude *Vaccinium-Hylocomium* subcommunity. Larger *Cladonia* spp., such as *C. impexa*, *C. uncialis* and *C. arbuscula*, can be particularly abundant crowning the tops of old *Eriophorum* hummocks and among senescent *Calluna*, but peat-encrusting species, such as *C. squamosa*, *C. bellidiflora*, *C. floerkeana* and *C. gracilis*, also occur and there can be records for *Cetraria islandica*, *Hypogymnia physodes* and *Cornicularia aculeata*.

Sub-communities

Erica tetralix sub-community: Blanket bogs Ratcliffe 1959 p.p.; Vaccinio-Ericetum tetralicis Moore 1962 p.p., Birse & Robertson 1976 p.p.; Eriophorum vaginatum bog, low-level facies Edgell 1969 p.p.; Erica tetralix-Vaccinium oxycoccos series, Plagiothecium-Hylocomium & Racomitrium-Cladina noda Tallis 1969; Trichophoreto-Eriophoretum Eddy et al. 1969 p.p.; Calluneto-Eriophoretum Meek 1976; Calluneto-Erio-

phoretum typicum Evans et al. 1977; Erico-Sphagnetum papillosi, Rhytidiadelphus loreus subassociation Moore (1964) 1968 emend. Birse 1980 p.p.; Calluna-Pleurozium & Calluna-Cladonia noda Bignal & Curtis 1981. This kind of Calluna-Eriophorum mire shows a number of floristic features transitional to more oceanic ombrogenous bogs. Among the vascular plants, for example, E. vaginatum can be co-dominant with the subshrubs, though it is generally less abundant and less prominent structurally than it is in the other subcommunities. Calluna often predominates among the woody species, frequently with some E. nigrum ssp. nigrum and a little V. myrtillus, but E. tetralix is strongly preferential and is sometimes of moderately high cover. Then, there is frequently some *Scirpus* and this, too, can be locally abundant with prominent tussocks. Molinia also occurs occasionally and there can be scattered plants of *Narthecium* and, more sparsely, of *Drosera* rotundifolia or Vaccinium oxycoccos. The great infrequency of Rubus chamaemorus, which is very local here, is a further distinctive feature.

The bryophyte element is less strikingly peculiar but Sphagna tend to be more consistently abundant than in many tracts of Calluna-Eriophorum mire, certainly than in much of the Empetrum sub-community, with S. capillifolium being quite commonly accompanied by S. papillosum and sometimes by S. tenellum, S. subnitens or S. compactum. And there can be a slight suggestion of the development of hummocks and hollows with these species disposed over stretches of wetter or drier ground. Other stands can have an abundance of S. recurvum, sometimes with S. palustre, in fairly extensive lawns.

The difference between this kind of vegetation and drier types of *Scirpus-Eriophorum* and *Erica-Sphagnum* mire, where hypnaceous mosses and herbs such as *Juncus squarrosus* and *Deschampsia flexuosa* (all quite common in this sub-community) show some increase, is thus a fine one, and especially hard to discern if individual samples are examined in isolation. The context of the samples, that is the overall character of the mire from which they are taken, sometimes helps but the similarity of the vegetation types is a real reflection of the convergence of ombrogenous mire floristics in certain environmental conditions (see below).

Empetrum nigrum ssp. nigrum sub-community: Eriophoretum vaginati Smith & Moss 1903, Lewis 1904, Lewis & Moss 1911, Moss 1913, Adamson 1918, Watson 1932, Tansley 1939, all p.p.; Mixed moor Pearsall 1938; Sphagnum community, Type B 'Moss' Pearsall 1941; Calluna & Calluna-Eriophorum Moss Pearsall 1941; Blanket bogs Ratcliffe 1959 p.p.; Eriophorum vaginatum bog, high-level facies Edgell 1969; Calluneto-Eriophoretum Eddy et al. 1969 p.p., Birks 1973; Calluneto-Eriophoretum deschampsietosum & myrtillosum Evans

et al. 1977; Vaccinio-Eriophoretum Hill & Evans 1978 p.p.; Mire noda 10 & 11 Daniels 1978. This subcommunity, long familiar as the richer kind of 'Pennine blanket bog', generally preserves all the typical floristic features of the Calluna-Eriophorum mire, though it is very variable in the proportions of the major structural elements and thus in its gross appearance, a feature very well described from Moor House in Cumbria (Welch & Rawes 1966, Eddy et al. 1969, Rawes & Hobbs 1979). E. vaginatum is usually abundant and structurally prominent and, indeed, on uneroded mires, it can be dominant for some time after episodes of burning, and become more permanently ascendant under intensive grazing. Generally, however, sub-shrubs play a fairly consistent part in the vegetation. Calluna is usually the leading species among these and, on virgin mires, this forms the bulk of an open canopy, 2-3 dm tall, the bushes alternating with healthy Eriophorum tussocks or growing on moribund ones, and the Calluna itself sometimes showing its familiar stages of growth from pioneer through to degenerate where burning or grazing are absent (Watt 1955, Gimingham 1972). There is also often some V. myrtillus and/or E. nigrum ssp. nigrum and both of these, particularly the latter, can increase to very great abundance for some time after burning or where grazing has been withdrawn, or more permanently where drier peat surfaces have been bared by erosion: under these last conditions both E. vaginatum and Calluna can show much less vigorous growth than usual. At the high altitudes characteristic of this vegetation, such erosion, often aggravated by generations of burning and grazing and, more recently, by drainage schemes, is widespread, the peats often deeply scored by systems of haggs, which introduce a characteristic patterning into the vegetation (e.g. Tansley 1939, Tallis 1964b, 1965, 1973b, 1985b). This sub-community also provides some of the more southerly stations for the Arctic-Alpine sub-shrubs V. vitis-idaea and V. uliginosum, though these become very much more frequent in the Vaccinium-Hylocomium subcommunity. In sharp contrast to the Erica subcommunity, E. tetralix is very scarce here, and any increase is often an indication of the local occurrence of a tract of that kind of *Calluna-Eriophorum* mire within the Empetrum sub-community (as at Moor House: Eddy et al. 1969).

Scirpus and Molinia both decline here to their usual low levels of frequency in the community and Rubus chamaemorus increases greatly, though it is absent from some far-flung localities for this sub-community, as in North Wales (Ratcliffe 1959a, Edgell 1969) and on Skye (Birks 1973) and can be decidedly patchy elsewhere, as on Stainmore (Pearsall 1941) and parts of Moor House (Eddy et al. 1969), this local variation sometimes being obviously related to treatment. Eriophorum angustifolium occurs very commonly, usually at low covers, and

there is occasionally some *Deschampsia flexuosa*, but other herbs are few and there are no preferentials among this element.

Bryophytes, too, show little that is distinctive, though there is a typically rather impoverished *Sphagnum* flora. *S. capillifolium* remains frequent and it can be quite abundant on the wetter ground around the *E. vaginatum* and beneath the sub-shrubs, though burning and erosion can have a drastic effect on its extent. *S. papillosum* is especially scarce and *S. subnitens* occurs only very occasionally though, as in the *Erica* sub-community, some stands show a local abundance of *S. recurvum* (as in a facies distinguished by Eddy *et al.* 1969).

Over somewhat drier ground, the hypnaceous mosses are particularly abundant, forming extensive mats over the decumbent branches and litter among older Calluna bushes and over the crowns of ageing Eriophorum tussocks. Where bare peat is exposed, acrocarpous species such as Dicranum scoparium, Campylopus paradoxus and Pohlia nutans become frequent and these are commonly accompanied by such hepatics as Mylia taylori, Kurzia pauciflora, Ptilidium ciliare, Cephalozia spp. and Calypogeia spp., together with lichens like Cladonia impexa, C. squamosa, C. bellidiflora, C. floerkeana, Cornicularia aculeata and Hypogymnia physodes.

Vaccinium vitis-idaea-Hylocomium splendens subcommunity: Betula nana bogs Poore & McVean 1957; Calluneto-Eriophoretum McVean & Ratcliffe 1962; Empetreto-Eriophoretum McVean & Ratcliffe 1962, Dierssen 1982; Juncus-Deschampsia Series Tallis 1969 p.p.; Calluneto-Eriophoretum Eddy et al. 1969 p.p. This sub-community embraces all of the high-montane blanket mire first comprehensively described by McVean & Ratcliffe (1962). It preserves all the general floristic features of the community but is especially distinctive in the consistent presence of the Arctic-Alpine sub-shrubs V. vitis-idaea and E. nigrum ssp. hermaphroditum, which are frequent throughout, and V. uliginosum, which is of somewhat more restricted occurrence. Various mixtures of these species, usually with abundant Calluna and generally smaller amounts of V. myrtillus, and sometimes with some E. nigrum ssp. nigrum, are typically co-dominant with Eriophorum vaginatum in the familiar kind of mixed canopy, though one which is often less tall than at lower altitudes. The montane character of the vegetation is reinforced by the occurrence of the rarities Arctostaphylos alpinus, Betula nana and Vaccinium microcarpum in some stands.

Among the vascular associates, Eriophorum angustifolium is rather less common here than usual but Rubus chamaemorus remains very frequent, Juncus squarrosus occurs quite often, usually at low covers, and in many stands there is some Carex bigelowii. Scirpus cespitosus also shows something of a resurgence in frequency here.

The remaining distinctive features of this subcommunity are found among the ground layer. First, Sphagna can be quite prominent, with S. capillifolium being accompanied in some stands by S. papillosum and S. subnitens, in others by S. fuscum; S. russowii is also found very occasionally. Then, the hypnaceous mosses are consistently joined by Hylocomium splendens and there is quite commonly some Racomitrium lanuginosum and Polytrichum alpestre; and the rare Kiaeria starkei can sometimes be found. Among the hepatics, Ptilidium ciliare is strongly preferential, and Mylia anomala a little more common than usual, with Anastrepta orcadensis figuring occasionally. The lichen flora, too, is usually well developed, Cladonia arbuscula and C. uncialis being especially frequent, C. impexa, C. rangiferina and Cetraria islandica occurring somewhat less commonly. Among the available stands, three variants have been recognised.

Betula nana variant: Betula nana bogs Poore & McVean 1957; Calluneto-Eriophoretum, shrub-rich facies McVean & Ratcliffe 1962. Superficially, this kind of Calluna-Eriophorum mire is virtually identical with the typical Vaccinium-Hylocomium form, mixtures of E. vaginatum and Calluna generally being co-dominant, with smaller amounts of V. myrtillus, V. vitisidaea, E. nigrum ssp. nigrum and/or E. nigrum ssp. hermaphroditum, and sometimes a little Erica tetralix, forming a cover 2 dm or so high. Closer inspection, however, reveals the presence of *Betula nana* generally growing low among or beneath the other woody species, sometimes showing more conspicuously where their cover is a little thinner. Some stands, from north of the Great Glen, can also have small amounts of Arctostaphylos alpinus growing low over the ground and others, in the north-west Highlands, contain Vaccinium microcarpum. Rubus chamaemorus is generally of low cover in this variant but, among the other vascular associates, Juncus squarrosus can be locally abundant. Carex bigelowii, in contrast to the other variants, is absent here.

Bryophytes have extensive cover but, though Sphagnum growth seems vigorous, the carpet is made up almost entirely of S. capillifolium, very occasionally with some S. fuscum. Hypnaceous mosses are more varied, with Pleurozium schreberi, Hylocomium splendens, Plagiothecium undulatum and Hypnum cupressiforme/jutlandicum all well represented. Ptilidium ciliare is the commonest hepatic, though Mylia anomala and Lophozia ventricosa occur occasionally. Lichens tend to be less conspicuous than in the other variants but C. arbuscula and C. impexa can be locally abundant and C. rangiferina is found in some stands.

Typical variant: Calluneto-Eriophoretum, Sphagnum type and lichen-rich facies McVean & Ratcliffe 1962; Juncus-Deschampsia series Tallis 1969 p.p.; Calluneto-Eriophoretum, Empetrum facies Eddy et al. 1969 p.p.

Calluna and E. vaginatum are generally co-dominant in this variant, with some V. myrtillus and V. vitis-idaea and, rarely, a little V. uliginosum, and E. nigrum ssp. hermaphroditum usually better represented than E. nigrum ssp. nigrum. Rubus chamaemorus and Juncus squarrosus can both be quite abundant and Carex bigelowii becomes frequent, though generally at low covers.

Although the contingent of hypnaceous mosses remains an important element in the ground cover, Sphagna can be quite abundant in wetter places, with S. capillifolium the best represented species (S. quinquefarium sometimes being distinguished) but S. subnitens and S. fuscum also figuring with local prominence. Cladonia spp., too, are common with C. arbuscula, C. uncialis and C. rangiferina occurring throughout and sometimes attaining high cover.

Vaccinium uliginosum-Polytrichum alpestre variant: Empetreto-Eriophoretum McVean & Ratcliffe 1962. This is the most distinctive kind of high-montane Calluna-Eriophorum mire in which E. nigrum ssp. hermaphroditum almost totally replaces ssp. nigrum, becoming the usual co-dominant of Eriophorum vaginatum in place of Calluna, which is only very occasional and of low cover. Vaccinia are also prominent, with V. uliginosum frequently joining V. myrtillus and V. vitis-idaea. Mixtures of the sub-shrubs and cotton-grass form a very low canopy, often scarcely 1 dm high. Scattered through this are frequent plants of Rubus chamaemorus and Juncus squarrosus and, now becoming very common, Carex bigelowii and Scirpus cespitosus. Vaccinium microcarpum has also been recorded very occasionally.

In the ground layer, S. capillifolium and S. papillosum can both have high cover and, less frequently, there can be some S. fuscum, S. subnitens, S. tenellum or S. russowii. Hypnaceous mosses are a little less conspicuous, though Pleurozium schreberi, Rhytidiadelphus loreus and Hylocomium splendens all occur quite commonly in small amounts. Dicranum scoparium is also frequent, but more striking here is the constancy of Racomitrium lanuginosum and Polytrichum alpestre and the occasional occurrence of Kiaeria starkei. Ptilidium ciliare and Lophozia ventricosa are the commonest hepatics. Lichens also remain a prominent feature, with frequent records for Cladonia arbuscula and C. uncialis, and occasional C. impexa and Cetraria islandica.

Habitat

The Calluna-Eriophorum mire is the typical blanket bog vegetation of high-altitude ombrogenous peats that have accumulated in the wet and cold climate of the uplands of northern Britain. The harsh nature of the montane environment is reflected in the floristics of the community, though the vegetation often takes some of its character from the effects of treatments, notably

burning, grazing and draining; and these have contributed, perhaps with climatic change, to its modification and, in some places, to the erosion of the peats.

On the mantle of blanket peat that has developed over gentler slopes in those parts of Britain with more than 1200 mm annual precipitation (Climatological Atlas 1952) or in excess of 160 wet days yr⁻¹ (Ratcliffe 1968), the Calluna-Eriophorum mire is characteristic of higher altitudes where high precipitation/evaporation ratios coincide with low temperatures throughout the year. In fact, the climate is not quite so consistently humid as within the range of the Scirpus-Eriophorum mire, which is concentrated within the 2000 mm isohyet, with more than 200 wet days yr⁻¹. The Calluna-Eriophorum mire does extend into this zone, in the higher reaches of the north-west Highlands, but, for the most part, annual rainfall is between 1200 and 2000 mm, with 160-200 wet days yr⁻¹. But the greater difference between the distributions of the two communities is in relation to temperature. The Scirpus-Eriophorum mire is essentially an oceanic blanket bog community, largely confined to lower altitudes with relatively mild winters and a comparatively small annual temperature range. The Calluna-Eriophorum mire, by contrast, is characteristic of areas with harsh winters, with February minima for the most part below freezing, and cool summers. Overall, its range coincides closely with those parts of Britain which have annual accumulated temperatures of less than 830 day-degrees C yr⁻¹ (Page 1982), an area which takes in most of the higher ground through the Scottish Highlands, in the Southern Uplands, in the Lake District and down the Pennines, but which excludes most of Wales and also Dartmoor, where the climate is wet enough to support blanket mire development, but insufficiently cool for this community to be more than local. The altitudinal separation between the Scirpus-Eriophorum and Calluna-Eriophorum mires is fairly well maintained throughout their ranges, though its absolute level depends on the degree of oceanicity. In general, the former is restricted to sites below 500 m, with a mean altitude in the available samples of about 300 m, while the latter is generally found above 300 m, with a mean of 550 m, but in the wetter west of Scotland, the dividing line tends to be set rather higher, whereas, in the more continental east, the Calluna-Eriophorum mire extends down virtually to sea-level and the Scirpus-Eriophorum mire is almost totally excluded.

At these higher altitudes in northern Britain, the Calluna-Eriophorum mire is confined to deeper peats, usually more than 2 m thick (at least when uneroded) and sometimes considerably more, on flat or gently-sloping ground, generally 0–10° (mean 4°), though extending on to somewhat steeper slopes in places. It is thus most extensively developed on high-level plateaus and broad watersheds, such as dominate much of the

landscape of the eastern Highlands, in the Monadhliath, the Forest of Atholl and the Angus Hills (McVean & Ratcliffe 1962), parts of the Southern Uplands (Birse & Robertson 1976, Birse 1980) and the high Cheviot, and the Pennine summits right down to north Derbyshire (e.g. Moss 1913, Tansley 1939, Pearsall 1941, Eddy et al. 1969), with a few outlying stands in Wales (Ratcliffe 1959a, Edgell 1969, Tallis 1969). In more rugged country, where the climate is suitable but the terrain often too steep for the development of this kind of vegetation, its occurrence is more restricted: although the community is the major kind of blanket bog at higher altitudes in the western Highlands, it is thus rather patchy there, and it is decidedly scarce in the Lake District.

The peats are usually well-humified, at least above, highly acidic (with a surface pH often less than 4) and very oligotrophic. But, typically, they are not so thoroughly or consistently waterlogged as in the Scirpus-Eriophorum mire and, indeed, can become surface-dry and oxidised in the summer. This is probably partly due to the drier climate characteristic of the Calluna-Eriophorum mire over most of its range, but it is also perhaps accentuated by the fact that the community often occurs on broadly convex summits and slopes which shed water quite readily. Very frequently, too, such drainage has been sharpened by various kinds of treatment (see below). Thus, although the Calluna-Eriophorum mire is clearly a Sphagnetalia community, floristically distinct from the Ericetalia wet heaths of thinner, drier peats, Sphagna are less varied and luxuriant than in the Scirpus-Eriophorum or Erica-Sphagnum mires, the major peat-builders S. papillosum and S. magellanicum especially being of restricted occurrence. And the Calluna-Eriophorum mire only rarely shows the development of the hummock/hollow structure associated with active differentiation of surface relief within wet Sphagnum-dominated carpets. The nearest the community comes to this kind of composition and physiognomy is in the *Erica* sub-community, where lawn species like E. tetralix, Narthecium, Drosera rotundifolia and Vaccinium oxycoccos are preferential, and this kind of Calluna-Eriophorum mire is characteristic of habitats that are climatically or topographically transitional to those typical of our other ombrogenous mires. It is concentrated towards the more oceanic west, often at lower altitudes (mean about 400 m), including most of the stands of the community in Wales (Ratcliffe 1959a, Edgell 1969, Tallis 1969) and in Strathclyde (Bignal & Curtis 1981) and it occurs at higher levels where blanket mire of this general kind extends over flat or concave areas of relief (Tallis 1969, Eddy et al. 1969) where a higher water-table can be maintained. The more consistent occurrence of Scirpus in the Erica sub-community may also reflect the frequency of surface-waterlogging there: it occurs patchily in some other kinds of *Calluna-Eriophorum* mire, but in stands on more surface-dry peats is often only abundant along tracks, where aeration is reduced by trampling.

In the Erica sub-community, Eriophorum vaginatum often shows some reduction in abundance and vigour but, for the most part, it is the prominence of this species, rather than the luxuriance of Sphagna, which gives the Calluna-Eriophorum mire its Sphagnetalian character. The combination of a generally cool climate with a firm, acidic peat substrate, kept generally moist below but not surface-waterlogged, seems to provide this plant with very favourable conditions for its ascendancy (Tansley 1939, Godwin 1975). It can therefore be a major contributor to the accumulating peats under this vegetation, though it has not always been pre-eminent in their development. Its fibrous tussock bases are very resistant to decay and tend to survive in disproportionate abundance as fibrous clods, but earlier views that the deposits here are mainly E. vaginatum peat (e.g. Woodhead & Erdtman 1926) have not been borne out by subsequent studies: very often profiles show a transition to peat with abundant Sphagnum remains below and layers rich in E. vaginatum frequently occur in situations suggestive of climatic or edaphic dryness, so the cottongrass has probably waxed and waned in abundance since the inception of the development of this kind of blanket mire (e.g. Conway 1954, Tallis 1965, Godwin 1975).

The present climatic and soil conditions experienced by the Calluna-Eriophorum mire are reflected in other major features of the vegetation. First, the very poor representation here of Molinia, which is virtually confined to the transitional Erica sub-community and, even there, not very prominent, and the absence or scarcity of Myrica, Potentilla erecta, Polygala serpyllifolia and Pedicularis sylvatica, betoken the shift away from the lowland oceanic environment of the Scirpus-Eriophorum mire, where they are all well represented, to a harsher montane habitat. Second, there is the complementary increase in associates whose European distribution is either Arctic-Subarctic, like Rubus chamaemorus, or Arctic-Alpine, as with Vaccinium vitis-idaea, V. uliginosum, Empetrum nigrum ssp. hermaphroditum and Carex bigelowii, all of which occur frequently, and Betula nana and Arctostaphylos alpinus, which are less common but preferential to this community among our Sphagnetalia mires. The more structurally important of these species are, incidentally, all plants which thrive on acidic but well-drained substrates (e.g. Ritchie 1955a, Taylor 1971, Gimingham 1972) and their prominence here, together with Calluna, Vaccinium myrtillus and Empetrum nigrum ssp. nigrum, which have a wider representation on ombrogenous mires, is further testimony to the freedom from surface-waterlogging in the peats.

Much of the floristic variation between the subcommunities and variants of the Calluna-Eriophorum mire can be related to the different sensitivities of these species to the temperature regimes within the range of altitudes occupied by the community as a whole. At lower levels, this kind of blanket mire extends some way into areas where the climate is a little more equable, with February minima sometimes above freezing and mean annual maxima over 25 °C. In the Erica sub-community, therefore, the montane contingent is very poorly represented and the sub-shrub canopy consists largely of more broadly temperature-tolerant species such as Calluna, the cool oceanic E. nigrum ssp. nigrum and the Continental Northern V. myrtillus, though these show somewhat depressed vigour on the wetter peats, with E. tetralix being correspondingly more frequent. The woody element thus approaches that found in more oceanic ombrogenous mires, reinforcing the transitional character of the Erica sub-community with its more luxuriant Sphagnum component, frequent Scirpus and occasional Molinia.

With the shift to higher altitudes and a cooler climate, the Empetrum sub-community becomes the usual form of Calluna-Eriophorum mire, sporadically in Wales, but very extensively all up the Pennines, over the Cheviot and in south-east Scotland and into the eastern Highlands. The mean altitude of this sub-community is around 600 m and, at these levels, the winters are harsher and the mean annual maximum temperature generally between 21 and 25 °C. The sub-shrub component remains much as before, except that E. tetralix, which retains some measure of physiological activity in the winter (Gimingham 1972), is now much reduced in occurrence, with balance shifting to the other species, all of which perform well on the somewhat drier peats. More strikingly, Rubus chamaemorus appears as a constant: the British range of this plant coincides closely with the distribution of the more montane kinds of Calluna-Eriophorum mire and is closely confined within the 25 °C mean annual maximum isotherm (Taylor 1971).

V. vitis-idaea has a very similar range in Britain to R. chamaemorus (Ritchie 1955a) and it, too, shows a modest rise in frequency in the Empetrum sub-community, but it becomes really common with a further rise in altitude and drop in temperatures in the Vaccinium-Hylocomium sub-community. This kind of Calluna-Eriophorum mire occurs almost exclusively in Scotland, being especially extensive in the Central Highlands with more patchy occurrences in the more rugged country of the north-west Highlands, but it has outliers on the high Cheviot and in the Pennines and transitional stands can be found on the Berwyns. The mean altitude in this sub-community is almost 700 m and virtually everywhere the mean annual maximum temperature is less than 21 °C.

The area bounded by this isotherm coincides closely with the distribution of E. nigrum ssp. hermaphroditum, which becomes frequent here and gradually replaces ssp. nigrum with increasing altitude: the exact upper limit of the latter is hard to fix because of confusion between the two when in the vegetative state, but it is probably around 750 m (Bell & Tallis 1973). Certainly, in the Vaccinium-Polytrichum variant, which attains the highest altitudes reached by the Calluna-Eriophorum mire, occurring on summits in the Central Highlands with a mean height of over 850 m, it is very much the predominant sub-species and quite often the leading sub-shrub, being frequently co-dominant with E. vaginatum, itself a circumpolar plant that retains its abundance and vigour fairly well throughout the whole altitudinal range. Here, too, V. uliginosum, another species generally confined within the 21 °C isotherm, becomes constant with the other Vaccinia, and Calluna is almost extinguished, reaching its altitudinal limit as a vigorous plant at around 850 m (Gimingham 1960).

In the very harsh climate experienced by the Vaccinium-Hylocomium sub-community, the woody cover can be reduced to a very low canopy and the vegetation provides a locus for some other high-montane species. Carex bigelowii has a geographical distribution which closely matches those of E. nigrum ssp. hermaphroditum and V. uliginosum, and it follows them here in becoming gradually more prominent in moving to the higher altitudes of the Vaccinium-Polytrichum variant. Vaccinium microcarpum and Sphagnum fuscum occur occasionally in both this and other variants, and with Betula nana bring the composition of the vegetation close to that of the North European Sphagnion fusci mires. B. nana, however, and Arctostaphylos alpinus, seem both to be better represented at not quite the highest altitudinal extreme of the Vaccinium-Hylocomium sub-community: they help characterise a lowerlevel Betula variant which is found patchily through the central and north-west Highlands, and where E. nigrum ssp. nigrum and Calluna still retain a frequent presence. The floristic differences between this Betula variant and the typical variant may be due partly to treatment (Poore & McVean 1957): the possibility of soligenous influence in the Betula variant (Poore & McVean 1957) was discounted by McVean & Ratcliffe (1962).

Treatments, particularly burning and grazing, are of considerable importance in influencing the composition and structure of the *Calluna-Eriophorum* mire throughout its range and particularly where stands form part of unenclosed hill-grazings or grouse-moors. Thus, although the qualitative differences between the subcommunities are set largely by climatic variation, the actual appearance of the vegetation within each altitudinal zone is often the result of human activity. Since all the major structural components of the community, *E*.

vaginatum, the sub-shrubs, R. chamaemorus, the bryophytes and the lichens, can be affected by such treatments, and often affected in rather different ways, the range of quantitative variation is considerable. And, because treatments have often been applied in complex combinations within a small compass, spatial diversity in individual stands is frequently high. This is shown very clearly in the accounts of the Calluna-Eriophorum mire from Moor House, where apparently uniform tracts of the community are revealed on closer inspection to comprise much smaller and quite varied blocks. (e.g. Eddy et al. 1969, Rawes & Heal 1978, Rawes & Hobbs 1979). This kind of pattern is typical of many stretches of this type of blanket mire.

It is from Moor House that we have the most detailed studies of the impact of burning and grazing on the Calluna-Eriophorum mire and, although work there has been largely on the Empetrum sub-community (which, with the impoverished Eriophorum mire, makes up the bulk of the blanket bog on the reserve), the general results of the investigations are of some significance for the community as a whole. With burning, the immediate effects are to destroy a proportion of the above-ground parts of the vegetation, produce fertilising ash and increase light penetration (Rawes & Hobbs 1979). Intense fires may consume all standing material, living or dead, but, if burning occurs in the winter, plants with some measure of dormancy, like E. vaginatum and R. chamaemorus, may escape the worst effects and the ground layer be better protected against severe damage by the increased wetness of the peat surface. And, indeed, it is E. vaginatum that typically shows the first response when the sub-shrubs have been burned off, coming to dominate for up to two decades after a fire (Eddy et al. 1969, Rawes & Hobbs 1979). R. chamaemorus, too, can become very abundant in these early stages, increasing its standing crop and fruit production (Taylor 1971, Taylor & Marks 1971). Over this period, however, the sub-shrubs gradually recover (provided there is no grazing): at Moor House, the mean age of stems of Calluna, the usual woody dominant in the blanket bog there, increased through the first decade after a burn and then levelled out. New growth from stools and seed diversified the age-structure of the population with time, though the degenerate phase of growth did not develop because of a smothering of the older stems by the Sphagnum carpet (Rawes & Hobbs 1979). E. nigrum ssp. nigrum and V. myrtillus, typical sub-shrubs of the *Empetrum* sub-community, did not figure prominently in post-burn seres at Moor House, but both they and, at higher altitudes, V. vitis-idaea, can attain great abundance after fires, provided burning has not been so intense as to destroy their rhizomes: each of these species can thus dominate for some time before being overtopped by Calluna (Ritchie 1955a, Ratcliffe 1959a, Gimingham 1964b, 1972, Bell & Tallis 1973). Recovery of *Calluna*, both vegetatively and from seed, proved better at Moor House under short-rotation (10-year) burning than long-rotation (20-year). Indeed, Rawes & Hobbs (1979) reported that most bog species seemed to benefit from the former kind of regime, which appeared to maintain some sort of steady state. Very frequent burning, however, leads to degeneration of the vegetation (see below).

Moderate levels of grazing, too, can maintain a stable diversity in the *Calluna-Eriophorum* mire. The major herbivores on this kind of blanket bog are sheep, with some contribution from deer (mostly red) and smaller mammals (rabbits, hares and voles) and, of particular importance for *Calluna*, grouse and, at higher altitudes, ptarmigan. At Moor House, Rawes & Hobbs (1979) considered that the community could support about 1 sheep per 2.5 ha, without any burning, and continue to produce sufficient food for both stock and grouse.

The most obvious effect of grazing is on the balance between E. vaginatum and the palatable ericoids Calluna and V. myrtillus. E. vaginatum is eaten, but its growing points are well protected within the tussocks (Wein 1973), whereas the sub-shrubs can be grazed to extinction under sustained heavy stocking levels (Eddy et al. 1969, Rawes & Hobbs 1979) or, conversely, encouraged by enclosure (Rawes 1981, 1983). R. chamaemorus also benefits greatly from protection from grazing provided the surrounding vegetation does not become too dense (Taylor 1971, Rawes & Hobbs 1979, Rawes 1983). Less palatable sub-shrubs, on the other hand, like E. nigrum ssp. nigrum (Bell & Tallis 1973) and V. vitis-idaea (Ritchie 1955a), can continue to make a more persistent contribution to the cover on grazed Calluna-Eriophorum mire. On shallower peats grazing may also increase the proportion of Juncus squarrosus and, where trampling decreases aeration along pathways, Scirpus cespitosus may spread. Trampling can also disrupt the Sphagnum cover, destroy larger Cladonia spp. and favour an increase in encrusting lichens, acrocarpous mosses like Campylopus flexuosus and some leafy hepatics on exposed peat surfaces. Such developments produce considerable fine variation between stands of the community and, where grazing is particularly heavy and longsustained, presage a more substantial change in the character of the vegetation (see below).

It is also possible for episodes of burning and grazing to contribute to the development of erosion within stretches of *Calluna-Eriophorum* mire, a process which can ultimately destroy the vegetation and its substrate, but which, in its less severe manifestations, introduces a further measure of patterning into the community. In fact, erosion of the kind of blanket peat of which the *Calluna-Eriophorum* mire is characteristic is a complex of processes, probably with varied causes (e.g. Johnson

1957, Radley 1962, Tallis 1964b, c, 1965, 1985b). At the higher altitudes to which the community penetrates, wind, rain and frost contribute to an element of natural climatic erosion on exposed summits and ridges, producing networks of bare areas, sometimes showing colonisation by encrusting lichens and bryophytes (Radley 1962, Tallis 1965). More dramatic is the marginal fretting of these bogs, in which extensive systems of gullies run back into the peat, separating upstanding remnant haggs, and sheet-erosion with extensive wastage of the upper layers (e.g. Lewis & Moss 1911, Tansley 1939, McVean & Ratcliffe 1962, Tallis 1965, Eddy et al. 1969). Severe erosion of this kind can be seen throughout the range of the Calluna-Eriophorum mire, being much more generally widespread than in the Scirpus-Eriophorum mire, where the bog margins are not so exposed, but it is especially pronounced through the southern Pennines, affecting about 75% of the blanket peat there (Anderson & Tallis 1981; see also Bower 1960,

This kind of erosion, too, may have a natural and long-standing component. In a detailed study of a small, but probably quite typical, area, Tallis (1985b) developed an original suggestion of Conway (1954) that mass movement might occur around the bog margins, restoring stability where accumulation had reached some kind of critical limit but exposing bare peat for removal with enhanced local drainage. With the early initiation of peat development in the southern Pennines, such a stage might have been reached a considerable time ago (Conway 1954, Tallis 1985a, b). More recently, erosion seems to have been markedly exacerbated by a variety of biotic influences responsible for the exposure of areas of bare peat, over which surface run-off can be channelled, with increased erosive power. Burning, particularly deep, catastrophic fires, and heavy grazing can both contribute to such developments (Radley 1962, Shimwell 1974, Tallis 1981, Tallis & Yalden 1984) and sheep-tracks (Tallis 1973b) and footpaths (Shimwell 1981) may also disrupt the cover of surface vegetation. In the southern Pennines, too, atmospheric pollution from the great industrial conurbations of Lancashire and Yorkshire (Conway 1949, Tallis 1964b, c, Ferguson & Lee 1983) has undoubtedly played a major part, together with enhanced drying of the peats as a result of water extraction, in the loss of Sphagnum from blanket mire vegetation there over the past two centuries, and this must also have exposed extensive areas of bare peat (Tallis 1985b).

Quite often, the bog vegetation in such seve elyeroded areas has become itself so impoverished as to fall within the *Eriophorum* mire, but eroded tracts can retain sufficient floristic richness as to still qualify as *Calluna-Eriophorum* mire. Typically, however, they exhibit a distinctive mosaic with *R. chamaemorus* often especially

abundant around the gully heads, and *E. nigrum* ssp. *nigrum* and *V. myrtillus* becoming more prominent along the sides of the gullies, over the deeply-fretted haggs and on tumbling masses of peat around the bog margins. More gently-sloping gullies with a measure of stagnation may show a regeneration of the mire vegetation but commonly other communities appear here and over thin redistributed peat and any exposed underlying drift (see below).

Zonation and succession

The Calluna-Eriophorum mire is characteristically found in zonations and mosaics with wet and dry heaths and grasslands over sequences of increasingly better-drained soils, with soligenous mires often interrupting the patterns where there is local flushing and sometimes relieving the prevailingly calcifuge nature of the vegetation cover. Almost universally, such transitions have been affected by treatments, which can blur the boundaries between the vegetation types, encourage some successional changes among them and introduce new elements of variation, particularly striking where these factors contribute to gross erosion of the peat.

The basic edaphically-related pattern is best seen where there is a gradual thinning of the ombrogenous peat cover to a humic top over gleyed podzols and then a thin layer of mor over freely-draining podzols, where the Calluna-Eriophorum mire gives way first to Ericetalia heath, then to Calluno-Ulicetalia heath and/or Nardetalia grasslands. Typically, such sequences are disposed over ground of progressively increasing slope, running downhill where the blanket bog occupies the summits of hills (a frequent occurrence here), running uphill where it clothes valley bottoms and plateaus which give way above to a fringe of steeper ground. The transitions may be on a grand scale, where more or less flat land is a prevailing element in the scenery, as is the case in parts of the Central Highlands, over the rounded summits of the Cheviot and on the high Pennines, where the communities can show a crude altitudinal pattern of replacement one by the other. But, in more rugged terrain, the zonations are on a finer scale, with repeated sequences over the stepped landscapes of grits and shales, for example, or complex mosaics disposed over craggy ground with small boulders. And, as in similar transitions with the Scirpus-Eriophorum mire, regional and local climate can influence the proportions of the communities that are represented, with the wetter elements of the sequence extended on to somewhat steeper slopes in areas with higher rainfall or on northern aspects, but contracted in drier sites.

The particular kinds of *Calluna-Eriophorum* mire found in these zonations, and the types of heath and grassland, vary with altitude and, where the bog gives way both above and below to such sequences, different

heaths and grasslands may be represented in each, though the general physiognomic character of the transitions is very similar. The shift to Ericetalia wet heath is thus usually marked by a fairly rapid loss of E. vaginatum and a further reduction in the often already low cover of Sphagna such as S. papillosum in the ground carpet. Where the Calluna-Eriophorum mire extends to lower altitudes in the Erica sub-community, such transitions can be fairly gradual and very like those found around the Scirpus-Eriophorum mire, with the Sphagnum sub-community of the Scirpus-Erica wet heath forming a marginal zone. But, at higher altitudes, where the Calluna-Eriophorum mire is represented by other sub-communities, it is usually drier forms of Ericetalia heath that replace the community as the peat cover thins: often, in such situations, the Vaccinium subcommunity of the Scirpus-Erica wet heath can be found, but at lower altitudes in eastern Scotland, the Juncus-Dicranum sub-community of the Ericetum tetralicis can mark the transition. In these vegetation types, the most obvious floristic changes, apart from the reduction in E. vaginatum and the Sphagna, are increased vigour among the sub-shrubs, an abundance of Juncus squarrosus and/ or Nardus stricta and the frequent occurrence of acrocarpous mosses such as Dicranum scoparium and Campylopus paradoxus, with Racomitrium lanuginosum and lichens often prominent on bare peat.

Sometimes, this wet-heath zone is of considerable extent but quite frequently, where the margin of the bog is steep, it is reduced in width or totally absent, with an abrupt switch to dry heath, with overwhelming dominance of Calluna, or one of the Empetra or Vaccinia. Usually, such vegetation can be included within the Calluna-Vaccinium or Vaccinium-Deschampsia heaths, but, at higher altitudes, the Calluna-Cladonia or Calluna-Arctostaphylos heaths can replace the Calluna-Eriophorum mire on steeper, drier slopes, and these in turn may give way to lichen- or moss-dominated vegetation on very exposed summits. At lower altitudes, the zonation often continues beyond the heaths into some kind of Nardetalia grassland and, quite often, there are direct transitions from the community to these vegetation types with the increasing influence of pastoral agriculture (see below).

General patterns of the kinds described above are frequently complicated by the local occurrence of soligenous mires, and sometimes small topogenous basins associated with them. Again, the particular character of this vegetation varies with altitude. At lower levels, through the range of the *Erica* and *Empetrum* subcommunities, various kinds of *Carex echinata-Sphagnum* mire, dominated by small sedges or Junci, can be found around flushes and along water-tracks through the bog and into the zones of fringing vegetation. At higher altitudes, this community is replaced in analo-

gous situations by the Carex-Sphagnum russowii mire. Where the flushing waters are somewhat less base-poor, the Carex-Sphagnum warnstorfii mire occurs, or where there is a marked enrichment, as where the blanket bog has encroached over calcareous substrates, the Pinguiculo-Caricetum, Carici-Saxifragetum or Caricetum saxatilis can mark out sites with soligenous influence, with the Carex-Calliergon mire in small waterlogged hollows.

Such variegation often provides the floristically most interesting element of internal patterning on the Calluna-Eriophorum mire, because true hummock/hollow relief, with its distinctive zoning of plants and transitions to Rhynchosporion pools, is only rarely encountered. It is sometimes developed in the Erica sub-community, which can occur within tracts of the Empetrum type where a higher water-table is locally maintained, and then the Rhynchosporion component is generally represented by the Sphagnum cuspidatum/recurvum community. In other cases, the Carex rostrata-Sphagnum recurvum or Eriophorum angustifolium communities can mark out pools.

Much more commonly, internal variation within tracts of the *Calluna-Eriophorum* mire comprises quantitative differences in the major structural elements developed in response to burning and grazing. If such treatments are drastic and frequent or long-maintained, they can induce more substantial qualitative changes in the vegetation: in some cases, such changes can be reversed but they may initiate a run-down of the community and contribute to the destruction of the underlying peats.

Most obviously, frequent burning and heavy grazing contribute to the conversion of the Calluna-Eriophorum mire into the Eriophorum mire, where ericoids, Sphagna and hypnaceous mosses are of very patchy occurrence and E. vaginatum overwhelmingly dominant. Locallyextensive tracts of this impoverished vegetation can be found through much of the range of the Calluna-*Eriophorum* mire, though reversion to the richer blanket bog has been demonstrated after only 15 years of enclosure and freedom from burning at Moor House (Rawes 1983). In the southern Pennines, however, where injudicious treatments have been combined with draining and aerial pollution over the past two centuries, the degeneration of the Calluna-Eriophorum mire is particularly widespread and perhaps irremediable. Here, too, erosion of the peats is especially severe, with much bare peat even among the remnants of the richer blanket bog, patchy Eriophorum angustifolium pools occurring where water collects and soligenous vegetation approximating to the Carex echinata-Sphagnum mire marking out gullies with some moderate water movement. Similar mosaics are very well seen in the map of Moor House in Eddy et al. (1969).

In other cases, treatments can convert the Calluna-

Eriophorum mire into heath or grassland. The peats here are already fairly dry, so it is often a small step, with artificial drainage, to encourage a succession to wet heath, a change marked most obviously by a reduction in the vigour of E. vaginatum, and then to dry heath, with its overwhelming dominance of ericoids. Combined with grazing, such treatment can produce a Nardetalia sward or, with liming and fertiliser application, even something approximating to a Cynosurion pasture. With less assiduous improvement, Juncus squarrosus often gains an opportunity to spread in such situations, though enclosure of a Juncus-dominated pasture at Moor House showed that such a trend could be reversed, the vegetation beginning to develop into Calluna-Eriophorum mire after 25 years (Rawes 1981). The recent subsidising of improvement of hill-grazings has greatly speeded the reclamation of this kind of blanket bog for agriculture and rendered such changes more permanent. In other places, draining has been a prelude to afforestation.

Distribution

The Calluna-Eriophorum mire is centred on the higher ground in the Pennines and the Central Highlands of Scotland. The Empetrum sub-community is especially extensive in the former area, extending northwards through Cheviot and the Borders into eastern Scotland, but most stands in the Highlands are of the Vaccinium-Hylocomium sub-community, which extends to the altitudinal limit of this kind of blanket bog in central Scotland as the Vaccinium-Polytrichum variant, with the Betula and Typical variants represented at somewhat lower levels and penetrating patchily into the north-west Highlands. At lower altitudes, with a rather more oceanic climate, as through Wales and Strathclyde, the Erica sub-community is the usual form.

Affinities

The Calluna-Eriophorum mire in this scheme unites the richer kinds of Eriophoretum vaginati or 'Pennine blanket bog' (e.g. Smith & Moss 1903, Lewis 1904a, b, Lewis & Moss 1911, Moss 1913, Adamson 1918, Watson 1932, Tansley 1939) with the Calluneto-Eriophoretum common in Scotland (Poore & McVean 1957, McVean & Ratcliffe 1962, Birks 1973, Evans et al. 1977), also taking in related forms of blanket mire described from Wales (Ratcliffe 1959a, Edgell 1969, Tallis 1969) and southwest Scotland (Meek 1976, Birse & Robertson 1976, Birse 1980, Bignal & Curtis 1981) under a variety of names. The essential differences between these types are preserved here in the Empetrum, Vaccinium-Hylocomium and Erica sub-communities, though the first includes what Eddy et al. (1969) described as Calluneto-Eriophoretum (and some of their Trichophoro-Eriophoretum). The Vaccinium-Hylocomium sub-community also sub-

sumes the *Empetreto-Eriophoretum* of McVean & Ratcliffe (1962) and Dierssen (1982) as a distinctive variant.

In the scheme proposed by Moore (1968) for the classification of European mires, this kind of vegetation was termed *Vaccinio-Ericetum tetralicis* (see also Moore 1962) and Birse & Robertson (1976) adopted this name for their lower altitude blanket bog of this kind. But Moore's account was very much based on Irish stands, with some of Pearsall's (1941) data from Stainmore, and the character of this vegetation was prevailingly of the type included here in the *Erica* sub-community. In Birse's (1980) account of Scottish vegetation, his *Vaccinio-Ericetum* became subsumed in the *Erico-Sphagnetum*, though he retained a separate *Rhytidiadelpho-Sphagnetum fusci* for the high-montane vegetation contained in this scheme in the *Vaccinium-Hylocomium* sub-community.

Despite these varied treatments of the different types of this kind of blanket bog, there is sufficient in common among them for them to be retained within a single community. It shows obvious similarities to both the Scirpus-Eriophorum and Erica-Sphagnum mires and is probably best grouped with them in the Erico-Sphagnion (as Moore (1968) proposed). However, among our ombrogenous communities, it comes closest to the Sphagnion fusci bogs of boreal peatlands, particularly in the Vaccinium-Hylocomium sub-community, where Rubus chamaemorus and Empetrum nigrum ssp. nigrum are joined by E. nigrum ssp. hermaphroditum, Vaccinium vitis-idaea, V. uliginosum, Betula nana, V. microcarpum and Sphagnum fuscum, all of them species which become prominent to varying degrees through Poland, northeast USSR, Finland and Sweden.

Floristic table M19

	a	b	c	19
Eriophorum vaginatum	V (1–9)	V (1–9)	V (1-8)	V (1-9)
Calluna vulgaris	V (1–9)	V (1–9)	V (1-9)	V (1-9)
Pleurozium schreberi	IV (1–6)	IV (1–6)	V (1-5)	IV (1-6)
Sphagnum capillifolium	IV (1-8)	IV (1-7)	IV (1-9)	IV (1–9)
Eriophorum angustifolium	IV (1-8)	V (1-7)	III (1 -4)	IV (1–8)
Rubus chamaemorus	I (1-6)	V (1–9)	V (1-4)	IV (1–9)
Erica tetralix	V (1-4)	I (1–4)	I (1–2)	II (1–4)
Scirpus cespitosus	IV (1-6)	II (1 -4)	II (1-5)	II (1–6)
Hypnum jutlandicum	IV (1–8)	I (1–4)		II (1–8)
Narthecium ossifragum	II (1-5)	I (1-2)	I (1-2)	I (1-5)
Molinia caerulea	II (1–7)	I (1-4)		I (1-7)
Drosera rotundifolia	I (1–3)			I (1-3)
Sphagnum compactum	I (1–4)			I (1–4)
Vaccinium oxycoccos	I (1–2)			I (1-2)
Empetrum nigrum nigrum	III (1–5)	V (1-6)	II (1–6)	III (1-6)
Mylia taylori	I (1–3)	II (1–3)	I (1–3)	I (1–3)
Pohlia nutans	I (1)	II (1–3)	I (1-2)	I (1-3)
Cladonia impexa	I (1–4)	II (1–6)		I (1–6)
Cephalozia bicuspidata	I (1-3)	II (1-3)		I (1–3)
Calypogeia trichomanis		II (1–3)	I (1–3)	I (1–3)
Hylocomium splendens	I (1-4)	I (1–2)	V (1-6)	II (1–6)
Cladonia arbuscula	I (4)	II (1–3)	IV (1–8)	II (1–8)
Vaccinium vitis-idaea	I (1–4)	II (1–4)	IV (1-4)	II (1–4)
Juncus squarrosus	II (1–4)	I (1–7)	III (1–5)	II (1-7)
Ptilidium ciliare	I (1–4)	II (1–3)	III (1–5)	II (1–5)
Cladonia uncialis	I (1-4)	I (1)	III (14)	II (1–4)
Empetrum nigrum hermaphroditum	I (1)		III (1-6)	II (1–6)
Carex bigelowii		I (1–4)	III (1-5)	II (1-5)

Racomitrium lanaginosum	Vaccinium uliginosum	I (1)	I (1-2)	II (1-4)	I (1–4)
Sphagnum papillosum	Racomitrium lanuginosum	I (1-5)	I (1-3)	II (1–6)	I (1-6)
Sphagnum subnitiens	Polytrichum alpestre	I (1)	I (1)	II (1-4)	I (1-4)
Certaria islandica I (1-3) II (1-3) I (1-3) Sphagnum fuscum I (1-3) II (1-4) I (1-4) Betula nama I (1) II (1-4) I (1-4) Polyrichum alpinum I (1-4) I (1-4) I (1-4) Arctostaphylos alpinus I (1-3) I (1-3) I (1-3) Vaccinium microcarpum III (1-8) III (1-9) IV (1-5) III (1-6) Dicramm scoparium III (1-4) III (1-6) III (1-5) III (1-8) III (1-9) Plagiothecium undulatum III (1-6) III (1-5) III (1-8) III (1-10) III (1-7) II (1-6) Hypnum cupressiforme II (1-6) III (1-8) II (1-7) II (1-1) II (1-8) II (1-1) II (1-1)<	Sphagnum papillosum	I (1-4)	I (1–6)	II (1-7)	I (1-7)
Sphagnum fuscum	Sphagnum subnitens	I (1–6)	I (4-5)	II (1–8)	I (1-8)
Betula nana	Cetraria islandica		I (1-3)	II (1-3)	I (1-3)
Polytrichum alpinum	Sphagnum fuscum		I (1-3)	II (1-4)	I (1-4)
Arctostaphylos alpinus	Betula nana		I (1)	II (1-4)	I (1-4)
Vaccinium microcarpum	Polytrichum alpinum			I (1–4)	I (1-4)
Vaccinium myrtillus III (1-8) III (1-9) IV (1-5) III (1-6) Dicranum scoparium III (1-4) III (1-6) III (1-5) III (1-6) Rhytidiadelphus loreus II (1-4) III (1-5) III (1-8) III (1-8) Plagiothecium undulatum III (1-6) III (1-5) II (1-2) III (1-6) Hypnum cupressiforme II (1-6) III (1-4) II (1-3) II (1-8) Polagiothecium undulatum III (1-6) III (1-4) II (1-3) II (1-8) Polagiothecium undulatum III (1-6) III (1-4) II (1-13) II (1-8) Polagiothecium undulatum III (1-6) III (1-4) II (1-14) II (1-15) II (1-14) II (1-15) II (1-14) II (1-16) II (1-18) II (1-14) II (1-16) II (1-18)	Arctostaphylos alpinus			I (1–4)	I (1–4)
Dicranum scoparium III (1-4) III (1-6) III (1-5) III (1-6) Rhytidiadelphus loreus II (1-4) III (1-4) III (1-8) III (1-8) Plagiothecium undulatum III (1-6) III (1-5) II (1-7) III (1-7) Hypnum cupressiforme II (1-8) III (1-4) II (1-7) II (1-7) Deschampsia flexuosa II (1-8) II (1-4) II (1-3) II (1-8) Polytrichum commune II (1-4) II (1-4) II (1-4) II (1-8) Polytrichum commune II (1-5) II (1-4) II (1-4) II (1-5) Lophozia ventricosa I (1-2) II (1-3) II (1-3) II (1-4) Lophozia ventricosa I (1-2) II (1-3) II (1-3) II (1-3) Sphagnum palustre I (1-6) I (1-8) I (4) I (1-8) Sphagnum palustre I (1-5) I (1-6) I (1-4) I (1-4) Potentilla erecta I (1-2) I (1-4) I (1-4) I (1-4) Rhytidiadelphus squarrosus I (1-3) I (1-4) I (1-4)	Vaccinium microcarpum			I (1–3)	I (1-3)
Rhytidiadelphus loreus II (1-4) III (1-4) III (1-8) III (1-8) III (1-8) III (1-8) III (1-8) III (1-6) III (1-5) II (1-2) III (1-6) III (1-5) II (1-2) III (1-6) III (1-6) III (1-4) III (1-3) II (1-8) III (1-3) II (1-13) II (1-14) II (1-15) II (1-16) II (1-16) II (1-18) II (1-16) II (1-18) II (1-18) II (1-16) II (1-18)	Vaccinium myrtillus	III (1–8)	III (1-9)	IV (1-5)	III (1–9)
Plagiotheclum undulatum III (1-6) III (1-5) II (1-2) III (1-6) Hypnum cupressiforme II (1-6) III (1-4) II (1-7) II (1-7) Deschampsia flexuosa II (1-8) III (1-8) II (1-3) II (1-7) Polytrichum commune II (1-4) III (1-4) II (1-4) II (1-4) Aulacomnium palustre II (1-5) II (1-4) I (1) II (1-5) Lophozia ventricosa I (1-2) II (1-3) II (1-3) II (1-3) Sphagnum recurum I (1-6) I (1-8) I (4) I (1-4) I (1-6) Sphagnum palustre I (1-5) I (1-6) I (1-4) I (1-4) I (1-6) Potentilla erecta I (1-2) I (1-4) I (1-4) I (1-4) Nardus stricta I (1-8) I (1-4) I (1-6) I (1-8) Rhytidiadelphus squarrosus I (1-13) I (1-4) I (1-6) I (1-8) Barbilophozia floerkii I (1-2) I (1-4) I (1-2) I (1-4) Listera cordata I (1-3) I (1-3)	Dicranum scoparium	III (1–4)	III (1–6)	III (1-5)	III (1–6)
Hypnum cupressiforme II (1-6) III (1-4) II (1-7) II (1-7) Deschampsia flexuosa II (1-8) II (1-8) II (1-3) II (1-4) Polytrichum commune II (1-4) II (1-4) II (1-4) II (1-1) II (1-4) Aulaconnium palustre II (1-5) II (1-3) II (1-3) II (1-5) Lophozia ventricosa I (1-2) II (1-3) II (1-3) II (1-3) Sphagnum recurvum I (1-6) I (1-8) I (4) I (1-8) Sphagnum palustre I (1-5) I (1-6) I (1-4) I (1-4) Potentilla erecta I (1-2) I (1-4) I (1-4) I (1-6) Potentilla erecta I (1-8) I (1-4) I (1-6) I (1-8) Rhytidiadelphus squarrosus I (1-3) I (1-4) I (1-6) I (1-8) Rhytidiadelphus squarrosus I (1-3) I (1-4) I (1-1) I (1-4) Barbilophozia floerkii I (1-2) I (1-4) I (1-2) I (1-4) Barbilophozia floerkii I (1-2) I (1-3) I (1-	Rhytidiadelphus loreus	II (1-4)	III (1–4)	III (1–8)	III (1–8)
Deschampsia flexuosa II (1-8) II (1-8) II (1-3) II (1-8) Polytrichum commune II (1-4) II (1-1) II (1-5) Lophozia ventricosa I (1-2) II (1-3) II (1-3) II (1-3) II (1-3) Sphagmum recurvum I (1-6) I (1-8) I (4) I (1-8) Sphagmum palustre I (1-5) I (1-6) I (1-4) I (1-4) I (1-4) I (1-6) I (1-4) I (1-10-2) I (1-4) I (1-2) I (1-4) I (1-2) I (1-4) I (1-2) I (1-4) I (1-3) I (1-2) I (1-3) I (1-3) I (1-3) I (1-3)	Plagiothecium undulatum	III (1–6)	III (1-5)	II (1-2)	III (1–6)
Polytrichum commune	Hypnum cupressiforme	II (1–6)	III (1–4)	II (1-7)	II (1–7)
Aulacomnium palustre II (1-5) II (1-4) I (1) II (1-5) Lophozia ventricosa I (1-2) II (1-3) II (1-3) II (1-3) Sphagnum recurvum I (1-6) I (1-8) I (4) I (1-8) Sphagnum palustre I (1-5) I (1-6) I (1-4) I (1-6) Potentilla erecta I (1-2) I (1-4) I (1-4) I (1-4) Nardus stricta I (1-8) I (1-4) I (1-6) I (1-8) Rhytidiadelphus squarrosus I (1-3) I (1-4) I (1) I (1-4) Barbilophozia floerkii I (1-2) I (1-4) I (1-2) I (1-4) Barbilophozia floerkii I (1-2) I (1-4) I (1-2) I (1-4) Barbilophyllum albicans I (1-2) I (1-4) I (1-2) I (1-4) Chiplophyllum albicans I (1-2) I (1-4) I (1-2) I (1-4) Cardata I (1-2) I (1-3) I (1-2) I (1-4) Campylopus paradoxus I (1-4) I (1-3) I (1-3) I (1-4)	Deschampsia flexuosa	II (1–8)	II (1–8)	II (1–3)	II (1–8)
Lophozia ventricosa I (1-2) II (1-3) II (1-3) II (1-3) Sphagnum recurvum I (1-6) I (1-8) I (4) I (1-8) Sphagnum palustre I (1-5) I (1-6) I (1-4) I (1-4) Potentilla erecta I (1-2) I (1-4) I (1-4) I (1-4) Nardus stricta I (1-8) I (1-4) I (1-6) I (1-8) Rhytidiadelphus squarrosus I (1-3) I (1-4) I (1-2) I (1-4) Barbilophozia floerkii I (1-2) I (1-4) I (1-2) I (1-4) Diplophyllum albicans I (1-3) I (1-3) I (1-3) I (1-4) Listera cordata I (1-2) I (1-4) I (1-2) I (1-4) Camera cordata I (1-2) I (1-4) I (1-2) I (1-4) Camera cordata I (1-2) I (1-4) I (1-2) I (1-4) Camera nigra I (1-4) I (1-3) I (1-2) I (1-4) Camera nigra I (1-4) I (1-3) I (1-3) I (1-4) Melampyrum pratense </td <td>Polytrichum commune</td> <td>II (1-4)</td> <td>II (1-4)</td> <td>I (1–4)</td> <td>II (1–4)</td>	Polytrichum commune	II (1-4)	II (1-4)	I (1–4)	II (1–4)
Sphagnum recurvum I (1-6) I (1-8) I (4) I (1-8) Sphagnum palustre I (1-5) I (1-6) I (1-4) I (1-6) Potentilla erecta I (1-2) I (1-4) I (1-4) I (1-4) Nardus stricta I (1-8) I (1-4) I (1-6) I (1-8) Rhytidiadelphus squarrosus I (1-3) I (1-4) I (1) I (1-4) Barbilophozia floerkii I (1-2) I (1-4) I (1-2) I (1-4) Diplophyllum albicans I (1-3) I (1-3) I (1-3) I (1-3) Listera cordata I (1-2) I (1-4) I (1-2) I (1-4) Campylopus paradoxus I (1-4) I (1-3) I (1-2) I (1-4) Campylopus paradoxus I (1-4) I (1-3) I (1-2) I (1-4) Campylopus paradoxus I (1-4) I (1-3) I (1-2) I (1-4) Campylopus paradoxus I (1-4) I (1-3) I (1-3) I (1-4) Campylopus paradoxus I (1-4) I (1-3) I (1-3) I (1-4) <	Aulacomnium palustre	II (1–5)	II (1–4)	I (1)	II (1–5)
Sphagnum palustre I (1–5) I (1–6) I (1–4) I (1–6) Potentilla erecta I (1–2) I (1–4) I (1–4) I (1–4) Nardus stricta I (1–8) I (1–4) I (1–6) I (1–8) Rhytidiadelphus squarrosus I (1–3) I (1–4) I (1) I (1–4) Barbilophozia floerkii I (1–2) I (1–4) I (1–2) I (1–4) Diplophyllum albicans I (1–3) I (1–3) I (1–3) I (1–3) Listera cordata I (1–2) I (1–4) I (1–2) I (1–4) Campylopus paradoxus I (1–4) I (1–3) I (1–2) I (1–4) Campylopus paradoxus I (1–4) I (1–3) I (1–2) I (1–4) Carex nigra I (1–4) I (1–3) I (1–2) I (1–4) Melampyrum pratense I (1–2) I (1–3) I (1–3) I (1–4) Cladonia squamosa I (1–2) I (1–3) I (1–3) I (1–3) Cladonia squamosa I (1–2) I (1–3) I (1–3) I (1–3) Cladonia	Lophozia ventricosa	I (1–2)	II (1–3)	II (1–3)	II (1–3)
Potentilla erecta	Sphagnum recurvum	I (1–6)	I (1–8)	I (4)	I (1–8)
Nardus stricta I (1-8) I (1-4) I (1-6) I (1-8) Rhytidiadelphus squarrosus I (1-3) I (1-4) I (1) I (1-4) Barbilophozia floerkii I (1-2) I (1-4) I (1-2) I (1-4) Diplophyllum albicans I (1-3) I (1-3) I (1-3) I (1-3) Listera cordata I (1-2) I (1-4) I (1-2) I (1-4) Campylopus paradoxus I (1-4) I (1-3) I (1-2) I (1-4) Odontoschisma sphagni I (1-4) I (1-3) I (1-2) I (1-4) Carex nigra I (1-4) I (1-3) I (1-3) I (1-4) Melampyrum pratense I (1-2) I (1-3) I (1-3) I (1-4) Cladonia squamosa I (1-2) I (1-3) I (1-2) I (1-3) Cladonia squamosa I (1-2) I (1-2) I (1) I (1-2) Sphagnum tenellum I (1-2) I (1-2) I (1) I (1-2) Cladonia bellidiflora I (1-3) I (1-3) I (1-3) I (1-3) Calypog	Sphagnum palustre	I (1-5)	I (1–6)	I (1–4)	I (1–6)
Rhytidiadelphus squarrosus I (1-3) I (1-4) I (1) I (1-4) Barbilophozia floerkii I (1-2) I (1-4) I (1-2) I (1-4) Diplophyllum albicans I (1-3) I (1-3) I (1-3) I (1-3) Listera cordata I (1-2) I (1-4) I (1-2) I (1-4) Campylopus paradoxus I (1-4) I (1-3) I (1-2) I (1-4) Odontoschisma sphagni I (1-4) I (1-3) I (1-3) I (1-4) Carex nigra I (1-4) I (1-3) I (1-3) I (1-4) Melampyrum pratense I (1-2) I (1-3) I (1-2) I (1-3) Cladonia squamosa I (1-2) I (1-3) I (1-4) I (1-4) Sphagnum tenellum I (1-2) I (1-2) I (1) I (1-2) Cladonia bellidiflora I (1-3) I (1-3) I (1-3) I (1-3) Callypogeia fissa I (1-3) I (1-3) I (1-3) I (1-3) Calypogeia muellerana I (1-4) I (1-3) I (1-3) I (1-3) <t< td=""><td>Potentilla erecta</td><td>I (1-2)</td><td>I (1–4)</td><td>I (1–4)</td><td>I (1–4)</td></t<>	Potentilla erecta	I (1-2)	I (1–4)	I (1–4)	I (1–4)
Barbilophozia floerkii I (1-2) I (1-4) I (1-2) I (1-4) Diplophyllum albicans I (1-3) I (1-3) I (1-3) I (1-3) Listera cordata I (1-2) I (1-4) I (1-2) I (1-4) Campylopus paradoxus I (1-4) I (1-3) I (1-2) I (1-4) Odontoschisma sphagni I (1-4) I (1-3) I (1-3) I (1-4) Carex nigra I (1-4) I (1-3) I (1-3) I (1-4) Melampyrum pratense I (1-2) I (1-3) I (1-2) I (1-3) Cladonia squamosa I (1-2) I (1-3) I (1-4) I (1-3) I (1-4) I (1-3) I (1-4) I (1-4) I (1-3) I (1-4) I (1-4) I (1-3) I (1-4) I (1-3) I (1-4) I (1-2) I (1-3) I (1-4) I (1-2) I (1-3) I (1-4) I (1-2) I (1-3) I (1-2) I (1-3) I (1-2) I (1-3)	Nardus stricta	I (1–8)	I (1–4)	I (1–6)	I (1–8)
Diplophyllum albicans I (1-3) I (1-3) I (1-3) I (1-3) I (1-3) I (1-3) I (1-4) I (1-2) I (1-4) I (1-2) I (1-4) I (1-2) I (1-4) I (1-2) I (1-4) I (1-3) I (1-2) I (1-4) Odontoschisma sphagni I (1-4) I (1-3) I (1-3) I (1-4) I (1-3) I (1-2) I (1-3) I (1-4) I (1-2) I (1-2) I (1-3) I (1-2) I (1-3) I (1-2) I (1-3)	Rhytidiadelphus squarrosus	I (1-3)	I (1–4)	I (1)	I (1–4)
Listera cordata I (1-2) I (1-4) I (1-2) I (1-4) Campylopus paradoxus I (1-4) I (1-3) I (1-2) I (1-4) Odontoschisma sphagni I (1-4) I (1-3) I (1-3) I (1-4) Carex nigra I (1-4) I (1-3) I (1-3) I (1-4) Melampyrum pratense I (1-2) I (1-3) I (1-2) I (1-3) Cladonia squamosa I (1-2) I (1-3) I (1-4) I (1-4) Sphagnum tenellum I (1-2) I (1-2) I (1) I (1-2) Cladonia bellidiflora I (1-3) I (1-3) I (1-3) I (1-3) Galium saxatile I (1-3) I (1-3) I (1-3) I (1-3) Calypogeia fissa I (1-3) I (1-3) I (1-3) I (1-3) Pinguicula vulgaris I (1-2) I (1-3) I (1-2) I (1-2) Calypogeia fissa I (1-1) I (1-3) I (1-2) I (1-2) Cladonia coccifera I (1-4) I (1-3) I (1-2) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) I (1-3)	Barbilophozia floerkii	I (1-2)	I (1–4)	I (1–2)	I (1–4)
Campylopus paradoxus I (1-4) I (1-3) I (1-2) I (1-4) Odontoschisma sphagni I (1-4) I (1-3) I (1-3) I (1-4) Carex nigra I (1-4) I (1-3) I (1-3) I (1-4) Melampyrum pratense I (1-2) I (1-3) I (1-2) I (1-3) Cladonia squamosa I (1-2) I (1-3) I (1-4) I (1-4) Sphagnum tenellum I (1-2) I (1-2) I (1) I (1-2) Cladonia bellidiflora I (1-3) I (1-3) I (1-3) I (1-3) Galium saxatile I (1-3) I (1-3) I (1-3) I (1-3) Calypogeia fissa I (1-3) I (1-3) I (1-3) I (1-3) Pinguicula vulgaris I (1-2) I (1-3) I (1-2) I (1-2) Calypogeia muellerana I (1-2) I (1-3) I (1-2) I (1-2) Cladonia coccifera I (1-4) I (1-3) I (1-3) I (1-3) Hypogymnia physodes I (1-3) I (1-3) I (1-3) I (1-3) Cladonia t	Diplophyllum albicans	I (1-3)	I (1-3)	I (1-3)	I (1-3)
Odontoschisma sphagni I (1-4) I (1-3) I (1-3) I (1-4) Carex nigra I (1-4) I (1-3) I (1-3) I (1-4) Melampyrum pratense I (1-2) I (1-3) I (1-2) I (1-3) Cladonia squamosa I (1-2) I (1-3) I (1-4) I (1-4) Sphagnum tenellum I (1-2) I (1-2) I (1) I (1-2) Cladonia bellidiflora I (1-3) I (1-3) I (1-3) I (1-3) Galium saxatile I (1-3) I (1-3) I (1-3) I (1-3) Calypogeia fissa I (1-3) I (1-3) I (1-3) I (1-3) Pinguicula vulgaris I (1-2) I (1-3) I (1-2) I (1-2) Calypogeia muellerana I (1-4) I (1-3) I (1-2) I (1-2) Cladonia coccifera I (1-4) I (1-3) I (1-3) I (1-3) Hypogymnia physodes I (1-3) I (1-3) I (1-3) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) I (1-3) Kurzia pauc	Listera cordata	I (1-2)	I (1–4)	I (1–2)	I (1–4)
Carex nigra I (1-4) I (1-3) I (1-3) I (1-4) Melampyrum pratense I (1-2) I (1-3) I (1-2) I (1-3) Cladonia squamosa I (1-2) I (1-3) I (1-4) I (1-4) Sphagnum tenellum I (1-2) I (1-2) I (1) I (1-2) Cladonia bellidiflora I (1-3) I (1-3) I (1-3) I (1-3) Galium saxatile I (1-3) I (1-3) I (1-3) I (1-3) Calypogeia fissa I (1-3) I (1-3) I (1-3) I (1-3) Pinguicula vulgaris I (1-2) I (1-3) I (1-2) I (1-2) Calypogeia muellerana I (1-4) I (1-3) I (1-2) I (1-2) Caldonia coccifera I (1-4) I (1-3) I (1-2) I (1-3) Hypogymnia physodes I (1-3) I (1-3) I (1-3) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) I (1-3) Kurzia pauciflora I (1-3) I (1-3) I (1-3) I (1-3) Cladonia tenuis	Campylopus paradoxus	I (14)	I (1–3)	I (1–2)	I (1-4)
Melampyrum pratense I (1-2) I (1-3) I (1-2) I (1-3) Cladonia squamosa I (1-2) I (1-3) I (1-4) I (1-4) Sphagnum tenellum I (1-2) I (1-2) I (1) I (1-2) Cladonia bellidiflora I (1-3) I (1-3) I (1-3) I (1-3) Galium saxatile I (1-3) I (1-3) I (1-3) I (1-3) Calypogeia fissa I (1-3) I (1-3) I (1-3) I (1-3) Pinguicula vulgaris I (1-2) I (1-3) I (1-2) I (1-2) Calypogeia muellerana I (1-4) I (1-3) I (1-2) I (1-2) Cladonia coccifera I (1-2) I (1-3) I (1-3) I (1-3) Hypogymnia physodes I (1-3) I (1-3) I (1-3) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) I (1-3) Kurzia pauciflora I (1-3) I (1-3) I (1-3) I (1-3) Cladonia tenuis I (1-3) I (1-3) I (1-3) I (1-3) Cornus suec	Odontoschisma sphagni	I (1-4)	I (1-3)	I (1-3)	I (1–4)
Cladonia squamosa I (1-2) I (1-3) I (1-4) I (1-4) Sphagnum tenellum I (1-2) I (1-2) I (1) I (1-2) Cladonia bellidiflora I (1-3) I (1-3) I (1-3) I (1-3) Galium saxatile I (1-3) I (1-3) I (1-3) I (1-3) Calypogeia fissa I (1-3) I (1-3) I (1-3) I (1-3) Pinguicula vulgaris I (1-2) I (1-3) I (1-2) I (1-2) Calypogeia muellerana I (1-4) I (1-3) I (1-2) I (1-2) Cladonia coccifera I (1-4) I (1-3) I (1-3) I (1-3) Hypogymnia physodes I (1-3) I (1-3) I (1-3) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) I (1-3) Kurzia pauciflora I (1-3) I (1-3) I (1-3) I (1-3) Cladonia tenuis I (4) I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) I (1-3) Festuca vivipara </td <td>Carex nigra</td> <td>I (1-4)</td> <td>I (1-3)</td> <td>I (1-3)</td> <td>I (1–4)</td>	Carex nigra	I (1-4)	I (1-3)	I (1-3)	I (1–4)
Sphagnum tenellum I (1-2) I (1-2) I (1) I (1-2) Cladonia bellidiflora I (1-3) I (1-3) I (1-3) I (1-3) Galium saxatile I (1-3) I (1-3) I (1-3) I (1-3) Calypogeia fissa I (1-3) I (1-3) I (1-3) I (1-3) Pinguicula vulgaris I (1-2) I (1-3) I (1-2) I (1-2) Calypogeia muellerana I (1-4) I (1-3) I (1-2) I (1-3) Cladonia coccifera I (1-2) I (1-3) I (1-3) I (1-3) Hypogymnia physodes I (1-3) I (1-3) I (1-3) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) I (1-3) Kurzia pauciflora I (1-3) I (1-3) I (1-3) I (1-3) Cladonia tenuis I (4) I (1-3) I (1-3) Mylia anomala I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-3) I (1-3)	Melampyrum pratense	I (1–2)	I (1–3)	I (1–2)	I (1–3)
Cladonia bellidiflora I (1-3) I (1-2) I (1-2) I (1-2) I (1-3) I (1-4) I (1-3) I (1-3) </td <td>Cladonia squamosa</td> <td>I (1–2)</td> <td>I (1–3)</td> <td>I (1–4)</td> <td>I (1–4)</td>	Cladonia squamosa	I (1–2)	I (1–3)	I (1–4)	I (1–4)
Galium saxatile I (1-3) I (1-3) I (1-3) I (1-3) Calypogeia fissa I (1-3) I (1-3) I (1-3) I (1-3) Pinguicula vulgaris I (1-2) I (1-3) I (1-2) I (1-2) Calypogeia muellerana I (1-4) I (1-3) I (1-3) I (1-4) Cladonia coccifera I (1-2) I (1-3) I (1-3) I (1-3) Hypogymnia physodes I (1-3) I (1-3) I (1-3) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) I (1-3) Kurzia pauciflora I (1-3) I (1-3) I (1-3) I (1-3) Cladonia tenuis I (4) I (1-3) I (1-4) Mylia anomala I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Sphagnum tenellum	I (1–2)	I (1-2)	I (1)	I (1-2)
Calypogeia fissa I (1-3) I (1-3) I (1-3) Pinguicula vulgaris I (1-2) I (1-2) I (1-2) Calypogeia muellerana I (1-4) I (1-3) I (1-3) Cladonia coccifera I (1-2) I (1-3) I (1-3) Hypogymnia physodes I (1-3) I (1-3) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) Kurzia pauciflora I (1-3) I (1-3) I (1-3) Cladonia tenuis I (4) I (1-3) I (1-4) Mylia anomala I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Cladonia bellidiflora	I (1-3)	I (1-3)	I (1–3)	I (1-3)
Pinguicula vulgaris I (1-2) I (1-2) I (1-2) Calypogeia muellerana I (1-4) I (1-3) I (1-4) Cladonia coccifera I (1-2) I (1-3) I (1-3) Hypogymnia physodes I (1-3) I (1-3) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) Kurzia pauciflora I (1-3) I (1-3) I (1-3) Cladonia tenuis I (4) I (1-3) I (1-4) Mylia anomala I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Galium saxatile	I (1-3)	I (1-3)	I (1-3)	I (1-3)
Calypogeia muellerana I (1-4) I (1-3) I (1-4) Cladonia coccifera I (1-2) I (1-3) I (1-3) Hypogymnia physodes I (1-3) I (1-3) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) Kurzia pauciflora I (1-3) I (1-3) I (1-3) Cladonia tenuis I (4) I (1-3) I (1-4) Mylia anomala I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Calypogeia fissa	I (1–3)		I (1–3)	I (1–3)
Cladonia coccifera I (1-2) I (1-3) I (1-3) Hypogymnia physodes I (1-3) I (1-3) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) Kurzia pauciflora I (1-3) I (1-3) I (1-3) Cladonia tenuis I (4) I (1-3) I (1-4) Mylia anomala I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Pinguicula vulgaris	I (1–2)		I (1–2)	I (1–2)
Hypogymnia physodes I (1-3) I (1-3) I (1-3) Cladonia floerkeana I (1-3) I (1-3) I (1-3) Kurzia pauciflora I (1-3) I (1-3) I (1-3) Cladonia tenuis I (4) I (1-3) I (1-4) Mylia anomala I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Calypogeia muellerana	I (1–4)	I (1–3)		I (1-4)
Cladonia floerkeana I (1-3) I (1-3) I (1-3) Kurzia pauciflora I (1-3) I (1-3) I (1-3) Cladonia tenuis I (4) I (1-3) I (1-4) Mylia anomala I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Cladonia coccifera	I (1–2)	I (1–3)		I (1–3)
Kurzia pauciflora I (1-3) I (1-3) I (1-3) Cladonia tenuis I (4) I (1-3) I (1-4) Mylia anomala I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Hypogymnia physodes	I (1-3)	I (1-3)		I (1–3)
Cladonia tenuis I (4) I (1-3) I (1-4) Mylia anomala I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Cladonia floerkeana	I (1-3)	I (1–3)		I (1-3)
Mylia anomala I (1-3) I (1-3) I (1-3) Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Kurzia pauciflora		I (1-3)	I (1-3)	I (1-3)
Cornus suecica I (1-3) I (1-3) I (1-3) Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Cladonia tenuis		I (4)	I (1–3)	I (1-4)
Festuca vivipara I (1-6) I (1-2) I (1-6) Huperzia selago I (1-3) I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3) I (1-3)	Mylia anomala		, ,	I (1-3)	I (1–3)
Huperzia selago I (1-3) I (1-3) Cornicularia aculeata I (1-3) I (1-3)	Cornus suecica		I (1-3)	I (1-3)	I (1-3)
Cornicularia aculeata I (1–3) I (1–3) I (1–3)	Festuca vivipara		I (1-6)	I (1–2)	I (1–6)
	Huperzia selago		I (1-3)	I (1-3)	I (1-3)
Dicranum fuscescens I $(1-3)$ I $(1-3)$				I (1–3)	
	Dicranum fuscescens		I (1-3)	I (1-3)	I (1-3)

Floristic table M19 (cont.)

	a	b	c	19
Sphagnum robustum		I (1-2)	I (1-4)	I (1-4)
Cladonia gracilis		I (1-3)	I (1-3)	I (1–3)
Blechnum spicant		I (1-2)	I (1–2)	I (1-2)
Dicranum majus		I (4)	I (1–2)	I (1-4)
Carex binervis		I (1–4)	I (1–2)	I (1-4)
Luzula sylvatica		I (1–4)	I (1-3)	I (1-4)
Number of samples	61	53	56	181
Number of species/sample	17 (11–33)	19 (7–33)	20 (10–33)	19 (7–33)
Herb height (cm)	21 (8–40)	23 (11–60)	17 (5–45)	21 (5–60)
Herb cover (%)	83 (6–100)	89 (65–100)	96 (70–100)	89 (6-100)
Bryophyte height (mm)	38 (5–80)	40 (10–100)	33 (10–60)	37 (5–100)
Bryophyte cover (%)	56 (20–100)	39 (2–80)	75 (60–100)	53 (2–100)
Altitude (m)	409 (28-820)	601 (355–978)	691 (457–923)	568 (28–978)
Slope (°)	4 (0–12)	4 (0–18)	6 (0–25)	5 (0-25)
Soil pH	4.3 (3.2–6.2)	3.8 (3.1–4.7)	4.6 (4.1–5.5)	4.2 (3.1–6.2)

a Erica tetralix sub-community

Floristic table M19c, variants

	ci	cii	ciii
Eriophorum vaginatum	V (1-5)	V (1-8)	V (1-8)
Calluna vulgaris	V (6-9)	V (2–9)	II (1–4)
Pleurozium schreberi	V (1-4)	V (1-5)	IV (1-4)
Sphagnum capillifolium	IV (1–9)	IV (1-6)	IV (1-4)
Eriophorum angustifolium	II (1–4)	II (1-3)	V (1-4)
Rubus chamaemorus	V (1–3)	V (1-4)	V (1-4)
Vaccinium myrtillus	V (1-2)	V (1–5)	V (1-4)
Hylocomium splendens	V (1–6)	V (1-6)	III (1–3)
Vaccinium vitis-idaea	III (1–3)	V (1-3)	IV (1-4)
Cladonia arbuscula	III (1–5)	V (1-8)	IV (1-4)
Betula nana	V (1-4)	I (1)	
Empetrum nigrum nigrum	III (1 -4)	II (1-5)	I (1-2)
Hypnum cupressiforme	III (1–4)	II (1–8)	
Arctostaphylos alpinus	III (1–4)	I (4)	
Erica tetralix	III (1–3)	• •	
Cladonia rangiferina	II (1–3)	I (1-3)	
Mylia anomala	II (1-3)	I (1-3)	I (1-3)

b Empetrum nigrum nigrum sub-community

c Vaccinium vitis-idaea-Hylocomium splendens sub-community

¹⁹ Calluna vulgaris-Eriophorum vaginatum blanket mire (total)

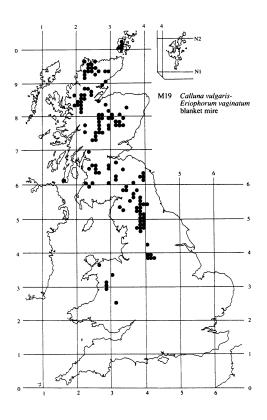
Vaccinium microcarpum	II (1–3)		
Vaccinium uliginosum	I (1-3)	I (2-4)	V (1-4)
Carex bigelowii		III (1–6)	V (1–6)
Racomitrium lanuginosum		II (1–4)	V (1-5)
Polytrichum alpestre		I (1–4)	V (1-4)
Scirpus cespitosus	II (1-2)	II (1-5)	IV (1-4)
Sphagnum papillosum		I (1-4)	III (4–6)
Kiaeria starkei			I (1–2)
Empetrum nigrum hermaphroditum	III (1–3)	III (1-4)	IV (4-6)
Juncus squarrosus	IV (1-5)	II (1–5)	III (1–3)
Rhytidiadelphus loreus	III (1 -4)	IV (1-8)	III (1–3)
Ptilidium ciliare	III (1–6)	III (1–2)	III (1–3)
Dicranum scoparium	II (1–3)	III (1–6)	III (1–5)
Cladonia uncialis	I (1-3)	IV (1-4)	IV (1-4)
Cladonia impexa	II (1-2)	II (1–4)	II (1–3)
Lophozia ventricosa	II (1-3)	I (1–2)	II (1-3)
Plagiothecium undulatum	II (1–3)	II (1–3)	, ,
Deschampsia flexuosa	, ,	II (1–3)	II (1-3)
Cetraria islandica		II (1–3)	II (1–3)
Sphagnum subnitens		II (1–8)	II (1–4)
Sphagnum fuscum	I (1-2)	II (1–4)	I (4–5)
Anastrepta orcadensis	I (1-3)	II (1–2)	I (1)
Melampyrum pratense	I (1-2)	II (1)	
Rhytidiadelphus squarrosus	I (1–3)	, ,	II (1-3)
Polytrichum commune	I (1-2)	I (1-4)	I (1-3)
Diplophyllum albicans	I (1-3)	I (1-3)	I (1-3)
Campylopus paradoxus	I (1-2)	I (1-3)	I (1-2)
Aulacomnium palustre	I (1-2)	I (1-2)	I (1-3)
Pinguicula vulgaris	I (1-2)	I (2)	I (2)
Barbilophozia floerkii	I (1-3)	I (1-3)	I (1)
Cornus suecica	I (1-3)	I (1–3)	I (1-3)
Potentilla erecta	I (1-2)	I (1–4)	I (1)
Erica cinerea	I (1-3)	I (1-3)	I (1-3)
Sphagnum robustum	I (1–4)	I (1–2)	I (1-4)
Cladonia gracilis	I (1-3)	I (1-3)	I (1-3)
Cladonia squamosa	I (1–3)	I (1–4)	I (1-3)
Mylia taylori	I (1–3)	I (1)	
Kurzia pauciflora	I (1-3)	I (1-3)	
Cladonia tenuis	I (1-2)	I (2)	
Listera cordata	I (1-3)	I (1-3)	
Calypogeia fissa	I (1-3)	I (1–3)	
Solidago virgaurea	I (1)	I (1-2)	
Carex binervis	I (1–2)	I (1-3)	
Sphagnum palustre	I (1–3)	I (4)	
Trientalis europaea		I (1)	I (1)
Cladonia bellidiflora		I (1-2)	I (1-3)
Sphagnum tenellum		I (1-3)	I (2)
Cornicularia aculeata		I (1-3)	I (1-3)
Festuca vivipara		I (1–3)	I (1-3)
Galium saxatile		I (1-2)	I (2)

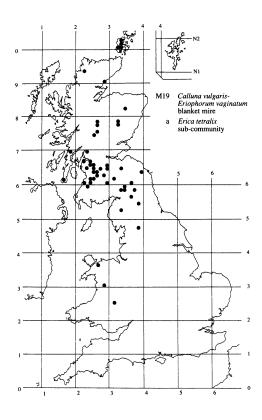
Floristic table M19c, variants (cont.)

	ci	cii	ciii
Dicranum fuscescens		I (1-3)	I (1-3)
Polytrichum alpinum		I (1–4)	I (1–3)
Number of samples	20	22	12
Number of species/sample	20 (16–27)	21 (12–33)	23 (18–33)
Herb height (cm)	24 (12–45)	15 (5–30)	9 (5–11)
Herb cover (%)	92 (70–100)	98 (85–100)	100
Bryophyte height (mm)		29 (20–40)	40 (10-60)
Bryophyte cover (%)		75	, ,
Altitude (m)	569 (457–766)	705 (495–853)	856 (762–923)
Slope (°)	4 (0–15)	8 (2–25)	5 (2–15)
Soil pH		4.7 (4.1–5.5)	4.3 (4.2–4.3)

ci Betula nana variant

ciii Vaccinium uliginosum-Polytrichum alpestre variant





cii Typical variant

