

methylation

Introduction

Install the following packages:

```
## Loading required package: Biobase

## Loading required package: BiocGenerics

## Loading required package: parallel

##
## Attaching package: 'BiocGenerics'

## The following objects are masked from 'package:parallel':
##
##   clusterApply, clusterApplyLB, clusterCall, clusterEvalQ,
##   clusterExport, clusterMap, parApply, parCapply, parLapply,
##   parLapplyLB, parRapply, parSapply, parSapplyLB

## The following objects are masked from 'package:stats':
##
##   IQR, mad, sd, var, xtabs

## The following objects are masked from 'package:base':
##
##   anyDuplicated, append, as.data.frame, basename, cbind, colnames,
##   dirname, do.call, duplicated, eval, evalq, Filter, Find, get, grep,
##   grepl, intersect, is.unsorted, lapply, Map, mapply, match, mget,
##   order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank,
##   rbind, Reduce, rownames, sapply, setdiff, sort, table, tapply,
##   union, unique, unsplit, which.max, which.min

## Welcome to Bioconductor
##
##   Vignettes contain introductory material; view with
##   'browseVignettes()'. To cite Bioconductor, see
##   'citation("Biobase)", and for packages 'citation("pkgname)".

## Setting options('download.file.method.GEOquery'='auto')

## Setting options('GEOquery.inmemory.gpl'=FALSE)
```

```
##
## Attaching package: 'dplyr'

## The following object is masked from 'package:Biobase':
##
## combine

## The following objects are masked from 'package:BiocGenerics':
##
## combine, intersect, setdiff, union

## The following objects are masked from 'package:stats':
##
## filter, lag

## The following objects are masked from 'package:base':
##
## intersect, setdiff, setequal, union

## Loading required package: usethis

##
## Attaching package: 'limma'

## The following object is masked from 'package:BiocGenerics':
##
## plotMA

##
## Attaching package: 'kableExtra'

## The following object is masked from 'package:dplyr':
##
## group_rows
```

Importing the data

```
## Warning: One or more parsing issues, see 'problems()' for details
## One or more parsing issues, see 'problems()' for details
```

Some datasets on GEO may be derived from different microarray platforms. Therefore the object `gse` is a list of different datasets. You can find out how many were used by checking the length of the `gse` object. Usually there will only be one platform and the dataset we want to analyse will be the first object in the list (`gse[[1]]`).

```
length(gse)
```

```
## [1] 1
```

Extract the data

```
gse <- gse[[1]]
gse
```

```
## ExpressionSet (storageMode: lockedEnvironment)
## assayData: 27578 features, 55 samples
##   element names: exprs
## protocolData: none
## phenoData
##   sampleNames: GSM813533 GSM813534 ... GSM813587 (55 total)
##   varLabels: title geo_accession ... tissue:ch1 (58 total)
##   varMetadata: labelDescription
## featureData
##   featureNames: cg00000292 cg00002426 ... cg27665659 (27578 total)
##   fvarLabels: ID Name ... ORF (38 total)
##   fvarMetadata: Column Description labelDescription
## experimentData: use 'experimentData(object)'
##   pubMedIds: 22613842
## Annotation: GPL8490
```

Exploratory analysis

The `exprs` function can retrieve the expression values as a data frame; with one column per-sample and one row per-gene.

```
pdata= pData(gse) #sample information
edata= exprs(gse) #expression data
fdata = fData(gse) #gene annotation
```

Inspect the clinical variables

Data submitted to GEO contain sample labels assigned by the experimenters, and some information about the processing protocol. All these data can be extracted by the `pData` function.