



Ricardo Henriques

Instituto Gulbenkian de Ciência
MRC-Laboratory for Molecular Cell Biology, UCL
rjhenriques@igc.gulbenkian.pt; @HenriquesLab
Born 20/05/1980; Portuguese

OVERVIEW

My laboratory focuses on advancing the boundaries of optical microscopy, with the aim of establishing novel technologies to address cell biology and biophysical questions, both in health and disease. We are recognized by our open-source and widely available contributions to the optical microscopy community, as well as our translational work with industry. For example, we have established the novel SRRF super-resolution approach that underpins Andor Technology's latest super-resolution spinning disk microscopes. In cell biology, we tackle broad questions through links with collaborating laboratories, in fields such as virology, host-pathogen interactions, immunology, cell signaling and evolution. We do so by establishing new classes of fluorescent probes, high-speed cell friendly super-resolution methods and computational modelling approaches that, although designed to answer questions of interest in the lab, have extensive applications in cell biology.

EDUCATION

PhD in Biophysics 2008-2011

Faculdade de Medicina Universidade de Lisboa

"Beyond Rayleigh's limit: achieving real-time super-resolution fluorescence microscopy"

Thesis advisor: Dr. Musa Mhlanga (CSIR, Pretoria, South Africa; IMM, Lisbon, Portugal)

Diploma in Physics 1998-2005

Faculdade de Ciências Universidade de Lisboa

Research advisors: Dr. Nuno Moreno and Prof. José Feijó (IGC, Portugal)

PROFESSIONAL HISTORY

Research Group Leader 2020-

Instituto Gulbenkian de Ciência, Oeiras, Portugal

- Research: technology development for super-resolution microscopy, computational bioimaging, machine learning, structural modelling, viral host-pathogen interactions
- Honorary Professor at *University College London*
- Affiliate Group Leader at *The Francis Crick Institute* with a Satellite Laboratory

Professor Chair of Computational and Optical Biophysics 2019-2020

MRC-Laboratory for Molecular Cell Biology, University College London

Associate Professor 2013-2019

MRC-Laboratory for Molecular Cell Biology, University College London

Postdoctoral Fellow 2011-2013

Institut Pasteur Paris, Department of Cell Biology and Infection

- Research at Zimmer Lab: super-resolution microscopy, T-cell immunological synapse formation, HIV-1 intracellular trafficking and uncoating.

Systems Developer and Consultant 2009-2019
Andor Technology (US and Northern Ireland)

- Scientific advisor, software developer, optical design consultant

Bioimaging Facility Manager 2005-2008
Instituto de Medicina Molecular and Instituto Gulbenkian de Ciência

- Teaching-on and maintenance-of optical microscopy equipment

OTHER APPOINTMENTS [since 2013]

Affiliate Group Leader at the Francis Crick Institute, Satellite Lab	2017-2020
Co-Director, Wellcome Trust-UCL Optical Biology PhD Programme	2020-
Wellcome Trust Multi-user Equipment Grants Committee Member	2020-2023
<i>bioRxiv</i> affiliate and advocate	2019-
Advisory board member of FocalPlane by Company of Biologists	2019-
MRC-LMCB Athena Swan Committee	2017-2018
UCL Advanced Microscopy Strategy Board	2017-2020
BBSRC Grants Committee, panel D and TRDF	2016-
Royal Microscopy Society Light Microscopy Committee	2014-2016
UCL Super-Resolution Microscopy Steering Board	2013-2020
MRC-LMCB Microscopy Committee	2013-2020
MRC-LMCB IT Committee	2013-2020

Academic Editor for: Scientific Reports (editorial board), Journal of Physics D [1] (guest)
Reviewer for: Nature Biotech, Nature Methods, Nature Communication, Nature Protocols, PNAS, Scientific Reports, PloS One, Journal of Microscopy, Optics Express, Traffic, Journal of Biophotonics, Light: Science & Application, Methods, Bioinformatics, Nanoscale
Grant reviewer for: ANR, BBSRC, CRUK, EPSRC, ERC, FCT, la Caixa Foundation, Leverhulm Trust, MRC, Netherlands Org. Sci. Res., Royal Society, Wellcome Trust

PRIZES, AWARDS AND HONOURS

Research highlighted by <i>Nature</i> [1]	2023
Research highlighted by <i>90-segundos-ciencia</i> (Portuguese news) [1]	2023
Research highlighted by <i>The Microscopists</i> (Podcast) [1]	2022
Research highlighted by <i>Nature Methods</i> [1]	2021
Research highlighted by <i>Expresso</i> (Portuguese printed news)[1]	2021
ERC CoG and EMBO IA awards highlighted by <i>Publico</i> (Portuguese news) [1][2]	2021
Research highlighted by <i>Nature News</i> ("Deep learning takes on tumours") [1]	2020
Research highlighted by <i>MRC</i> (UK's Medical Research Council) [1]	2019
Research highlighted by <i>Clubic</i> (French Technology Magazine) [1]	2019
Made Fellow of the Royal Microscopy Society	2018
Research highlighted by <i>The Times</i> [1]	2018
Research highlighted by <i>The Scientist</i> twice [1][2]	2018
Research highlighted by <i>This Week in Virology (TWiV)</i> [1]	2018
Spirit of SLMS award for Scientific Excellence, Nils Gustafsson (PhD student)	2018
Ref. case study for UCL-Consultancy, " UCL pioneers algorithm in microscopy "	2017
Cirklo Prize Best Concept for Scientific Facilities, Pedro Almada (PhD student)	2015
Pasteur Roux Post-doctoral Fellowship	2013
FCT Doctoral Research Fellowship	2010

COLLABORATIONS

P = Joint Authorship; G = Joint Funding; R = Joint Researchers. This list is often outdated, please visit our [laboratory website](#) for recent collaborations with joint publications

Andela Šarić, UCL, UK [1P,1G]; **Ann-Christin Lindas**, Stockholm Univ., Sweden [2P,1G]; **Ashley Cadby**, Univ. Sheffield, UK; **Buzz Baum**, UCL, UK [5P,2G,3R]; **Christophe Leterrier**, Aix University, France [3P]; **Christophe Zimmer**, Pasteur, France [5P,1G]; **Dylan Owen**, KCL, UK [1P]; **Ed Cohen**, Imperial College, UK [1P]; **Ethan Garner**, Harvard Univ., USA [1G]; **Eva Frickel**, Crick Institute, UK [1P]; **Ewa Paluch**, Cambridge Univ., UK [1R]; **Fabrice Agou**, Pasteur, France [1P]; **Florian Jug**, MPI-CBG, Germany [2P]; **Gabriel Martins**, IGC, Portugal; **Giuseppe Battaglia**, UCL, UK; **Guillaume Charras**, UCL, UK [1P,1R]; **Guillaume Jacquemet**, Åbo Akademi University, Finland [2P]; **Jake Baum**, Imperial College, UK; **Jan Löwe**, LMB, UK [1G]; **Jason Mercer**, UCL, UK [5P,1G,3R]; **Joe Grove**, Royal Free, UK [1P]; **Johanna Ivaska**, University of Turku, Finland [1P]; **Jonas Ries**, EMBL, Germany [2P]; **Jost Enninga**, Pasteur, France [1P]; **Loïc Royer**, CZ-Biohub, USA [2P]; **Maria Carmo-Fonseca**, IMM, Portugal [1P]; **Mary Collins**, Okinawa IST, Japan [1P]; **Maria Mota**, IMM, Portugal [1P]; **Mariana Pinho**, ITQB, Portugal [1G,1R]; **Mark Marsh**, UCL, UK [2P,2R]; **Maximiliano Gutierrez**, Crick Institute, UK [1R]; **Mike Heilemann**, Goethe University Frankfurt, DE [1P,1R]; **Mohan Balasubramanian**, Warwick Univ., UK [1G]; **Musa Mhlanga**, UCT, South Africa [7P,2G]; **Nick Robinson**, Lancaster University, UK [1P]; **Nuno Moreno**, IGC, Portugal [1G]; **Pavel Tomancak**, MPI-CBG, Germany [1P]; **Ralf Jungmann**, MPI Biochemistry, Germany; **Serge Mostowy**, Imperial College, UK [2P]; **Seamus Holden**, Newcastle Univ., UK [2P]; **Steven Lee**, Cambridge Univ., UK [1P]; **Simon Foster**, Univ. Sheffield, UK [1P]; **Thijs Ettema**, Univ. of Uppsala, Sweden [1G];

GRANTS AND FUNDING [since 2013, ≈25M/10yr]

26. **CZI** Applications: napari Plugin Foundations grants (Cycle 2), “[Cutting-edge super-resolution image analysis in napari through NanoJ](#)”, £25K (PI, 01/23 - 12/23).
25. **H2022** EIC Pathfinder Open, “Real-Time high-content Super-Resolution Imaging of ES Cell States”, £3.5M (Co-PI, 04/23 - 03/27).
24. **H2021** INFRA, “[Artificial Intelligence for Image Data Analysis in the Life Sciences](#)”, £3.6M (Co-PI, 09/22 - 10/25).
23. **CZI** Visual Proteomics Imaging, “[VP-CLEM-KIT: a pipeline for democratising volumetric visual proteomics](#)”, £3M (Co-PI, 12/21 - 06/24).
22. **EMBO** Installation Grant, “[Unveiling live-cell viral replication at the nanoscale](#)”, £150K (PI, 01/21 - 01/24).
21. **ERC** Consolidator, “[Enabling Live-Cell 4D Super-Resolution Microscopy Guided by Artificial Intelligence](#)”, £2M (PI, 09/21 - 09/26).
20. **BBSRC** ALERT, “[Benchtop, turnkey super-resolution microscopy for biology, biophysics and biotechnology](#)”, £200K (Co-PI, 05/20 - 04/21).
19. **Wellcome Trust** 4-year PhD Programme in Science, “Optical Biology”, £6M (Co-Director, 08/21 - 08/25).
18. **Wellcome Trust**, “[Understanding cellular organisation: from archaea to eukaryotes](#)”, £1.1M out of £4M (Co-PI, 12/16 - 12/21).

17. **Royal Society** International Exchanges 2019 (UK-Ireland), “An international joint collaboration to develop and democratise high-accessible open-source AI controlled microfluidics to enable unprecedented nanoscale cell biology research”, £12K (Henriques and Reynaud labs partnership, 08/19 - 08/21).
16. **UCL-Osaka** Strategic Partner Fund, “Establishing collaborative research between UCL and Osaka University”, £10K (Henriques and Nagai labs partnership, 08/19-08/20).
15. **UCL** Cities Partnership Programme & **EMBO** Short-Term Fellowship, “Establishing collaborative research between UCL and Institut Curie”, £7K (Application by Dr. Romain Laine - PDRA, 06/19).
14. **UCL** Capital Equipment Call (CEF3), “4D Super-Resolution Proteomics: Establishing a unique Super-Resolution Microscope capable of automatically mapping a theoretically unlimited number of proteins in space-and-time”, £150K (PI, 06/19 - 07/20).
13. **MRC** Skills Development Fellowship (Sponsor), “Dr. Romain Laine”, £288K (PI, 01/20 - 01/23).
12. **BBSRC** iCASE Studentship, “[Content Aware AI Driven Driven Super Resolution Microscopy](#)”, £107K (PI, 10/18 - 09/22).
11. **BBSRC** TRDF, “[Democratising Live-Cell Adaptive Super-Resolution Microscopy based on SRRF](#)”, £151K (PI, 01/19 - 02/20).
10. **BBSRC** TRDF, “[An accessible framework to achieve multi-dimensional live-cell super-resolution high-content screening](#)”, £151K (PI, 12/17 - 12/18).
9. **BBSRC** ALERT, “[Enabling Live-Cell Super Resolution Imaging Through Lattice Light Sheet Microscopy](#)”, £513K (Co-PI - main writer, 01/17 - 05/18).
8. **BBSRC** NIRG, “[Super-Beacons and Beacon-STORM: a new generation of small tunable photoswitching probes and Super-Resolution approaches.](#)”, £364K (PI, 01/16 - 12/18).
7. **MRC** Next Generation Optical Microscopy Initiative, “[Super Resolution Imaging for Cell Biology and Neuroscience at UCL](#)”, £220K out of £1.1M (not named PI but main contributor to grant impact and outcomes, 02/13 - 11/18).
6. **FCT** Research and Development Projects, “[Imaging the structure and dynamics of molecules and complexes in living organisms](#)”, £500K (Co-PI, 01/13 - 01/16).
5. Industrial R&D Collaboration with **3i**, “Adapting of SRRF to light-sheet”, £300K (PI, 09/16 - 12/19).
4. **NVidia** GPU Grant Programme, “Developing AI for Microscopy”, £5K (PI, 12/18).
3. **Marie-Curie** Postdoctoral Fellowship (Sponsor), “Dr David Albrecht”, £150K (Co-PI, 05/17 - 05/19).
2. Sir Henry **Wellcome** Postdoctoral Fellowship (Sponsor), “Dr Theo Sanderson”, £250K (Co-PI, 06/17 - 05/21).
1. **UK-SA Commonwealth** PhD Studentship (Sponsor), “Caron Jacobs”, £112K (PI, 09/14 - 03/18).

RECENT INVITED TALKS [showing selected]

Annually Invited: Advanced Imaging Course, EMBL Heidelberg, Germany	2012-
Annually Invited: ESRIC Super-Resolution Summer School, Edinburg, UK	2017-
Keynote: SPAOM, Valencia, Spain	2020
Keynote: Lifetime Unconference 2, Montpellier, France	2019
Keynote: Microscopy Society of Ireland Symposium, Dublin, UK	2019
Keynote: RMS Frontiers in BioImaging, Glasgow, UK	2018
Keynote: Single Mol. Approaches in Imaging, Ghent, Belgium (declined)	2018
Keynote: Scott. Microscopy Group Annual Symposium, Glasgow, UK	2017
Keynote: Spanish-Portug. Meeting Advanced Optical Microscopy, Bilbao, Spain	2016
Webinar: Global Bioimaging [1]	2021
Webinar: Crick-EMBL PostDoc symposium [1]	2021
Webinar: Cell Press [1]	2021
Webinar: EMBO YIP meeting	2021
Webinar: Physics of Life - University of York [1]	2021
Webinar: Physics Department - Faculdade de Ciências Universidade de Lisboa [1]	2021
Webinar: Living Systems Institute - University of Exeter [1]	2021
Webinar: CFCT - Faculdade de Ciências Universidade de Lisboa [1]	2021
Webinar: GDR - Imaging viruses, from single molecule to diagnosis [1]	2021
Webinar: CFCT - Faculdade de Ciências Universidade de Lisboa [1]	2021
Webinar: EuroBioImaging Virtual Pub [1]	2021
Webinar: Abbelight Academia Webinar [1] , Paris, France	2020
Webinar: Labroots Cell and developmental Biology Webinar [1]	2020
Webinar: Invited speaker for Science/AAAS Technology Webinar Series [1]	2018
Invited: LS4Future inaugural meeting, Oeiras, Portugal	2023
Invited: Engineering Biology, Cambridge University, UK [1]	2023
Invited: Koç Üniversitesi, Istanbul, Turkey	2023
Invited: Nikon Centre of Excellence Inauguration, Marseille, France [1]	2022
Invited: i3S annual retreat, Porto, Portugal	2022
Invited: France BioImaging Meeting, Institut Curie Paris, France [1]	2022
Invited: ELMI Meeting, Turku, Finland [1]	2022
Invited: SHIFT2022 Meeting, Tenerife, Spain [1]	2022
Invited: EMBO Computational Optical Biology, Oeiras, Portugal [1]	2022
Invited: Translational BioImaging Symposium, Wurzburg, Germany [1]	2022
Invited: Institut Pasteur, Seul, South Korea	2022
Invited: EMBO 3D Developmental Biology, Oeiras, Portugal [1]	2022
Invited: Single Molecule Localization Microscopy Symposium, Paris, France [1]	2022
Invited: Alexander Fleming Institute, Paris, France [1]	2021
Invited: Institut Curie, Paris, France [1]	2021
Invited: Center for Research in Myology, Paris, France	2021
Invited: Living Systems Institute, University of Exeter, UK	2021
Invited: From Images to Knowledge with ImageJ & Friends, Janelia Farm, US	2020
Invited: University of Oxford, Oxford, UK	2020
Invited: 3D Single-Mol. Localization Workshop, The Francis Crick Institute, UK	2020
Invited: Data Science in Cell Imaging Workshop, Company of Biologists, UK	2020
Invited: Quantitative Methods in Biology, Imperial College, UK	2019
Invited: Vlaams Instituut voor Biotechnologie, Ghent, Belgium	2019
Invited: MRC Weatherall Institute of Molecular Medicine, Oxford, UK	2019
Invited: University of Birmingham, Birmingham, UK	2019
Invited: University of Oxford, Oxford, UK	2019

Invited: University of Cambridge, Cambridge, UK	2019
Invited: UZH and ETH Advanced Microscopy Winter School, Zurich, Switzerland	2019
Invited: The Institute of Cancer Research	2018
Invited: MiFoBio - Functional Microscopy in Biology, Seignosse, France	2018
Invited: 84th Harden Conference: Single-Molecule Bacteriology, Oxford, UK	2018
Invited: First UK/Japan Super-resolution Bioimaging Meeting	2018
Invited: EMBO Course 3D Developmental Imaging, IGC, Portugal	2018
Invited: Focus on Microscopy international meeting, Singapore	2018
Invited: Biochem. Society Harden Conf. Single Mol. Bacteriology, Oxford, UK	2018
Invited: University of Bern, Bern, Switzerland	2018
Invited: University of Cambridge, Cambridge, UK	2018
Invited: Institut Pasteur, Paris, France	2018
Invited: 7th Single Molecule Localization Microscopy Symposium, London, UK	2017
Invited: ICFO, Barcelona, Spain	2017
Invited: Queen's College London, UK	2017
Invited: University of Liverpool, UK	2017
Invited: UK Membrane-Trafficking Meeting, London, UK	2016
Invited: Pharmac. Summer Course, Univer. Menéndez Pelayo, Santander, Spain	2016
Invited: Summer School on Molecular-Scale Engineering, Sheffield, UK	2016
Invited: University of Edinburgh, UK	2016
Invited: University of Sussex, UK	2016
Invited: Royal Society UK-SA Imaging in Host-Path. Interact., South Africa	2014
<u>PI position interview</u> : University of Oxford (offered), UK	2019
<u>PI position interview</u> : University of Birmingham (offered), UK	2019
<u>PI position interview</u> : Crick satellite programme (offered), UK	2016
<u>PI position interview</u> : MRC-LMCB at UCL (offered), UK	2013
<u>PI position interview</u> : MRC-LMB (offered), UK	2013

INDUSTRIAL PARTNERSHIPS

R&D with Andor Technology : developed the SRRF-Stream technology	2016-2019
R&D with 3i : host lab of UKs eng. team, developed SRRF for Lattice Light-Sheet	2016-
R&D with Abbelight : implementation of microfluidics in super-resolution	2018-
Reference lab for Cairn Research : test of prototype equipment	2017-
Reference lab for Mizar Imaging : test of prototype equipment	2018-

SCIENTIFIC MEETINGS ORGANISED [since 2013]

Bi-monthly London Super-Resolution Group Meetings, London, UK	2013-2020
EMBO Practical Course "Computational Optical Biology", Oeiras, Portugal	2022
EMBO Practical Course "3D development(all) imaging", Oeiras, Portugal	2022
EMBO Practical Course "3D development(all) imaging", Oeiras, Portugal	2020
ASCB Workshop "Optogenetics Imaging Techniques", Washington DC, USA	2020
7th Single Molecule Localization Microscopy Symposium, London, UK	2017
Super-Res. Microscopy in Infection and Immunity Symposium, IGC, Portugal	2015
UCL Super-Resolution Symposium, London, UK	2015
Royal Society UK-SA Imaging in Host-Path. Interactions, South Africa	2014

THESES SUPERVISED

8. [Neza Vadjnal](#) – PhD (Co-PI, 2017-22) - “Investigate the molecules controlling actin network architecture and their influence on cortex tension generation.” *Now postdoc researcher in Paluch lab.*
7. [Lucas Von Chamier](#) – PhD (PI, 2017-22) - “Using artificial intelligence to achieve smart super-resolution microscopy” *Now postdoc researcher.*
6. [Yue \(Julie\) Yuan](#) – PhD (PI, 2017-21) - “Super-resolution mapping of receptor engagement during HIV entry.” *Now postdoc researcher.*
5. [Robert Gray](#) – PhD (PI, 2015-18) - “Understanding vaccinia virus entry by super-resolution and particle averaging.” *Now computational biologist at Sixfold Bioscience.*
4. [Jerzy Samolej](#) – PhD (Co-PI, 2015-18) - “Identification of anti-poxviral agents by high-throughput image-based screening.” *Now postdoc researcher in Mercer lab.*
3. [Caron Jacobs](#) – PhD (PI, 2014-18) - “The nanoscale organisation of HIV cell surface receptors CD4 and CCR5.” *Now postdoc at University of Cape Town, South Africa.*
2. [Pedro Bento Almada](#) – PhD (PI, 2014-17) - “[Developing highly multiplexed technology for high-throughput Super-resolution Fluorescence Microscopy.](#)” *Now scientific consultant for Almada Scientific Services, UK.*
1. [Nils Gustafsson](#) – PhD (PI, 2014-17) - “[Enabling live-cell super-resolution microscopy by computational analysis and fluorescent probe design.](#)” *Now postdoc at Ludwig-Maximilians-Universität, Germany.*

PhD examinations: 1) [Garth Burn](#) - Andrew Cope and Dylan Own Lab, KCL, UK [2014]; 2) [Frederico Leon](#) - Achillefs Kapanidis Lab, Univ. Oxford, UK [2015]; 3) [Timothée Verdier](#) - Martin Castelnovo, ENS - Lyon, France [2015]; 4) [Adela Staszowska](#) - Susan Cox Lab, KCL, UK [2016]; 5) [Samuel Barnett](#) - Neil Hunter and Ashley Cadby Lab, Univ. Sheffield, UK [2017]; 6) [Pedro Silva](#) - Jorge Carneiro Lab, IGC, Portugal [2017]; 7) [Anna Bove](#) - Guillaume Charras and Alan Lowe Lab, UCL, UK [2018]; 8) [Jennifer Francis](#) - Raphaël Levy Lab, Univ. Liverpool, UK [2018]; 9) [Teodor Viktorov Boyadzhiev](#) - Simon Ameer-Beg, KCL, UK [2019]; 10) [Marco Fantham](#) - Clemens Kaminsky Lab, University of Cambridge, UK [2019]; 11) [Sohaib Abdul Rehman](#) - Kevin O'Holleran Lab, University of Cambridge, UK [2019]; 12) [Dimitrios Kiagias](#) - Miguel Juarez Lab, University of Sheffield, UK [2019]; 13) [Yiangos Psaras](#) - Matthew Daniels Lab, University of Oxford, UK [2020]; 14) [Krystian Ubych](#) - Robert Neely Lab, University of Birmingham, UK [2020]; 15) [Maria Arista Romero](#) - Lorenzo Albertazzi Lab, Institute for Bioengineering of Catalonia, Spain [2021];

PUBLIC ENGAGEMENT AND OUTREACH

Public engagement and outreach is a major focus of our research laboratory. We particularly engage projects tackling gender equality and helping students from disadvantaged backgrounds. We are also extremely present in social media, using platforms such as [Twitter](#) (~12K followers) and Public Press [[1](#)][[2](#)][[3](#)] to promote our scientific research and engage a global audience.

Projects and Actions:

- 2013: Co-founder of [AGRAFr - Association des Diplômés Portugais en France](#) - created by a group of Portuguese researchers in Paris, AGRAFr aims to develop multidisciplinary synergy covering all areas of knowledge and to foster exchange of experiences and contacts between Portugal and France.

- 2013: Joined the [MRC-LMCB public engagement programme](#): *School visits* - annual programme where students are given background information on cell biology research, exposed to a range of lab-based activities and provided with a careers Q&A; *Back to school* - visit schools to promote knowledge in our research and science as a career; *Labathon* - open activities showcasing essential manual skills required to carry out science (e.g. pipetting, cell counting, measuring solutions by eye), which highlights the fun element of producing science to young members of the public; *Science Festivals* – science open days that include activities and workshops focused around disseminating knowledge of cell biology.
- 2015: Recurring Speaker in [Pint of Science](#) [1][2] - a science festival that brings researchers to local pubs to present their scientific discoveries.
- 2017: Joined [MRC-LMCB Athena SWAN committee](#) (Gold Award) - an initiative to foster gender equality, role models, career events, skills exchange and staff well-being.
- 2017: Joined [In2ScienceUK](#) as host lab (3 students) - an award winning initiative which empowers students from disadvantaged backgrounds to achieve their potential and progress to STEM and research careers through high quality work placements and careers guidance.
- 2022: Joined [Science4Ukraine](#) initiative as host lab (1 student) for researchers moving from Ukraine during its invasion.

TEACHING [Since 2013]

Beyond local teaching at UCL, our group participates in some of the most highly recognise international courses in advanced and super-resolution microscopy. We particularly target to train multidisciplinary researchers in quantitative advanced imaging and critical thinking in microscopy, including its limitations.

Selected UCL teaching:

- [Advanced Molecular Cell Biology](#) (previously CELL3050, now CELL0016);
- [Analysis of Biological Complexity](#) (CoMPLEX PhD Programme)
- [Mammalian Physiology](#) (PHOL1001);
- [MRes Modelling Biological Complexity](#);
- [MSci in Biological Physics](#);
- [MSci Cell Biology](#) (CELLM102);
- Personal Tutor BioMedical Sciences (5 students per year);
- [Principles of Biology](#) (BBSRC LiDo PhD Programme);
- [Super-Resolution Microscopy and Image Analysis](#) (IPLS PhD Programme);
- [Super-Resolution Microscopy and Image Analysis](#) (MRC-LMCB PhD Programme);
- [SysMIC course](#) (BBSRC LiDO PhD Programme);

Selected international teaching:

- [Edinburgh Super-Resolution Imaging Consortium Summer School](#), UK (Week-long Course) [2017, 2018, 2019];

- EMBO 3D Developmental Imaging, Portugal (Week-long Course) [2018];
- EMBL Advanced Fluorescence Imaging Techniques (Week-long Course) [2013, 2014, 2015, 2016, 2017, 2018, 2019];
- PhD Programme Lecture - Instituto de Medicina Molecular, Portugal [2015];
- PhD Programme Lecture - Instituto Gulbenkian de Ciência, Portugal [2017];
- SRRF Workshop MPI-CBG, Germany (Two-day Course) [2018];
- SRRF Workshop University of Bern, Switzerland (Two-day Course) [2018];
- Focus on Microscopy Tutorial, Singapore (Invited Lecture) [2018];

SOFTWARE DEVELOPMENT

7. [ZeroCostDL4Mic](#) - GNU GPL (PI - 2021): [von Chamier et al., N. Comm., 2021](#) - *Democratizing deep learning for microscopy with ZeroCostDL4Mic*.
6. [NanoJ](#) - GNU GPL (PI - 2018): [Laine et al., J. Phys. D, 2019](#) - *High-performance open-source super-resolution microscopy toolbox, capable of GPU acceleration*.
5. [NanoJ-Fluidics](#) - MIT License (PI - 2018): [Almada et al., Nat. Comm., 2019](#) - *Automating multimodal microscopy through inexpensive LEGO based syringe pumps*.
4. [NanoJ-SQUIRREL](#) - GNU GPL (PI - 2018): [Culley et al., Nat. Meth., 2018](#) - *Quantitative mapping and minimization of super-resolution artifacts. Commercially adapted by [Abbelight](#)*.
3. [NanoJ-SRRF](#) - GNU GPL (PI - 2016): [Gustafsson et al., Nat. Comm., 2016](#) - *New analytical super-resolution approach, led to the [first super-resolution cameras](#) by [Andor Technology](#)*.
2. [NanoJ-VirusMapper](#) - GNU GPL (PI - 2016): [Gray et al., Sci. Rep., 2016](#) - *First open-source algorithm for Single-Particle Analysis in super-resolution microscopy*.
1. [QuickPALM](#) - GNU GPL (PI - 2010): [Henriques et al., Nat. Meth., 2010](#) - *First open-source software for super-resolution analysis (PALM and STORM), one of the most used analytical packages in the Super-Resolution field*.

PUBLICATIONS [[Google Scholar](#)]

* co-corresponding author; \pm equal contribution; this list is often outdated, please visit our [laboratory website](#) for recent publications

60. Lena Harker-Kirschneck, Anne E Hafner, Tina Yao, Christian Vanhille-Campos, Xiuyun Jiang, Andre Pulschen, Fredrik Hurtig, Dawid Hryniuk, Siân Culley, [Ricardo Henriques](#), Buzz Baum, Anđela Šarić, “[Physical mechanisms of ESCRT-III-driven cell division](#)”, *PNAS* (2022).
59. Christoph Spahn*, Romain F Laine, Pedro M Pereira, Estibaliz Gómez-de-Mariscal, Lucas von Chamier, Mia Conduit, Mariana G Pinho, Séamus Holden, Guillaume Jacquemet, Mike Heilemann*, [Ricardo Henriques*](#), “[DeepBacs: Bacterial image analysis using open-source deep learning approaches](#)”, *bioRxiv*, in review (2021).
58. Mario Del Rosario, Hannah S Heil, Afonso Mendes, Vittorio Saggiomo, [Ricardo Henriques*](#), “[The Field Guide to 3D Printing in Optical Microscopy for Life Sciences](#)”, *Advanced Biology* (2021).

57. Romain F Laine, Ignacio Arganda-Carreras, Ricardo Henriques, Guillaume Jacquemet, “[Avoiding a replication crisis in deep-learning-based bioimage analysis](#)”, *Nat. Methods* (2021).
56. Bruno M Saraiva, Ludwig Krippahl, Sérgio R Filipe, Ricardo Henriques, Mariana G Pinho, “[eHooke: a tool for automated image analysis of spherical bacteria based on cell cycle progression](#)”, *Biological Imaging* (2021).
55. Bruno M Saraiva, Ludwig Krippahl, Sérgio R Filipe, Ricardo Henriques, Mariana G Pinho, “[eHooke: a tool for automated image analysis of spherical bacteria based on cell cycle progression](#)”, *Biological Imaging* (2021).
54. Kevin D Whitley, Calum Jukes, Nicholas Tregidgo, Eleni Karinou, Pedro Almada, Yann Cesbron, Ricardo Henriques, Cees Dekker, Séamus Holden, “[FtsZ treadmilling is essential for Z-ring condensation and septal constriction initiation in Bacillus subtilis cell division](#)”, *Nat. Communications* (2021).
53. Lucas von Chamier, Romain F Laine, Johanna Jukkala, Christoph Spahn, Daniel Krentzel, Elias Nehme, Martina Lerche, Sara Hernández-Pérez, Pieta K Mattila, Eleni Karinou, Séamus Holden, Ahmet Can Solak, Alexander Krull, Tim-Oliver Buchholz, Martin L Jones, Loïc A Royer, Christophe Leterrier, Yoav Shechtman, Florian Jug, Mike Heilemann, Guillaume Jacquemet*, Ricardo Henriques*, “[Democratising deep learning for microscopy with ZeroCostDL4Mic](#)”, *Nat. Communications* (2021). **Key publication.**
52. Emma Touizer*, Christian Sieben*, Ricardo Henriques*, Mark Marsh*, Romain F Laine*, “[Application of super-resolution and advanced quantitative microscopy to the spatio-temporal analysis of influenza virus replication](#)”, *Viruses* (2021).
51. Yue Yuan, Cardon A Jacobs, Isabel Llorente Garcia, Pedro M Pereira, Scott P Lawrence, Romain F Laine, Mark Marsh, Ricardo Henriques*, “[Single-Molecule Super-Resolution Imaging of T-Cell Plasma Membrane CD4 Redistribution upon HIV-1 Binding](#)”, *Viruses* (2021).
50. Alexander Spark, Alexandre Kitching, Daniel Esteban-Ferrer, Anoushka Handa, Alexander R Carr, Lisa-Maria Needham, Aleks Ponjavic, Ana Mafalda Santos, James McColl, Christophe Leterrier, Simon J Davis, Ricardo Henriques, Steven F Lee, “[vLUME: 3D virtual reality for single-molecule localization microscopy](#)”, *Nat. Methods* (2020).
49. Gautam Dey, Siân Culley, Scott Curran, Uwe Schmidt, Ricardo Henriques, Wanda Kukulski, Buzz Baum, “[Closed mitosis requires local disassembly of the nuclear envelope](#)”, *Nature* (2020).
48. Gabriel Tarrason Risa, Fredrik Hurtig, Sian Bray, Anne E Hafner, Lena Harker-Kirschneck, Peter Faull, Colin Davis, Dimitra Papatziadou, Delyan R Mutavchiev, Catherine Fan, Leticia Meneguello, Andre Arashiro Pulschen, Gautam Dey, Siân Culley, Mairi Kilkenny, Dorge P Souza, Luca Pellegrini, Robertus AM de Bruin, Ricardo Henriques, Ambrosius P Snijders, Anđela Šarić, Ann-Christin Lindås, Nicholas P Robinson, Buzz Baum, “[The proteasome controls ESCRT-III-mediated cell division in an archaeon](#)”, *Science* (2020).
47. Andre Arashiro Pulschen, Delyan R Mutavchiev, Siân Culley, Kim Nadine Sebastian, Jacques Roubinet, Marc Roubinet, Gabriel Tarrason Risa, Marleen van Wolferen, Chantal Roubinet, Uwe Schmidt, Gautam Dey, Sonja-Verena Albers, Ricardo Henriques, Buzz Baum, “[Live imaging of a hyperthermophilic archaeon reveals distinct roles for two ESCRT-III homologs in ensuring a robust and symmetric division](#)”, *Current Biology* (2020).

46. Guillaume Jacquemet*, Alexandre F. Carisey*, Hellyeh Hamidi, Ricardo Henriques*, Christophe Leterrier*, “[The cell biologist’s guide to super-resolution microscopy](#)”, *Journal of Cell Science* (2020).
45. Andre Arashiro Pulschen, Delyan R Mutavchiev, Siân Culley, Kim Nadine Sebastian, Jacques Roubinet, Marc Roubinet, Gabriel Tarrason Risa, Marleen van Wolferen, Chantal Roubinet, Uwe Schmidt, Gautam Dey, Sonja-Verena Albers, Ricardo Henriques, Buzz Baum, “[Live Imaging of a Hyperthermophilic Archaeon Reveals Distinct Roles for Two ESCRT-III Homologs in Ensuring a Robust and Symmetric Division](#)”, *Current Biology* (2020).
44. Pedro M. Pereira, Nils Gustafsson, Mark Marsh, Musa M. Mhlana, Ricardo Henriques*, “[Super-Beacons: Open-Source Probes With Spontaneous Tuneable Blinking Compatible With Live-Cell Super-Resolution Microscopy](#)”, *Traffic* (2020). **Key publication.**
43. Aki Stubb, Romain F Laine, Camilo Guzmán, Ricardo Henriques, Guillaume Jacquemet, Johanna Ivaska, “[Fluctuation-Based Super-Resolution Traction Force Microscopy](#)”, *Nano Letters* (2020).
42. Kalina L Tosheva, Yue Yuan, Pedro M Pereira, Siân Culley*, Ricardo Henriques*, “[Between Life and Death: strategies to reduce phototoxicity in super-resolution microscopy](#)”, *J. Phys. D* (2020).
41. Robert Gray, David Albrecht, Corina Beerli, Gary Cohen, Ricardo Henriques*, Jason Mercer*, “[Nanoscale Polarization of the Vaccinia Virus Entry Fusion Complex Drives Efficient Fusion](#)”, *Nat. Microbiology* (2019). **Key publication.**
40. Pedro Almada, Pedro Pereira, Siân Culley, Ghislaine Caillol, Fanny Boroni-Rueda, Christina L Dix, Romain F Laine, Guillaume Charras, Buzz Baum, Christophe Leterrier*, Ricardo Henriques*, “[Automating multimodal microscopy with NanoJ-Fluidics](#)”, *Nat. Communications* (2019). **Key publication.**
39. Jervis Vermal Thevathasan, Maurice Kahnwald, Konstanty Cieslinski, Philipp Hoess, Sudheer Kumar Peneti, Manuel Reitberger, Daniel Heid, Krishna Chaitanya Kasuba, Sarah Janice Hoerner, Yiming Li, Yu-Le Wu, Markus Mund, Ulf Matti, Pedro Matos Pereira, Ricardo Henriques, Bianca Nijmeijer-Winter, Moritz Kueblbeck, Vilma Jimenez Sabinina, Jan Ellenberg, Jonas Ries, “[Nuclear pores as versatile reference standards for quantitative superresolution microscopy](#)”, *Nat. Methods* (2019).
38. Daniel Sage, Thanh-An Pham, Hazen Babcock, Tomas Lukes, Thomas Pengo, Ramraj Velmurugan, Alex Herbert, Anurag Agrawal, Silvia Colabrese, Ann Wheeler, Anna Archetti, Bernd Rieger, Raimund Ober, Guy M. Hagen, Jean-Baptiste Sibarita, Jonas Ries, Ricardo Henriques, Michael Unser, Seamus Holden, “[Super-resolution fight club: A broad assessment of 2D & 3D single-molecule localization microscopy software](#)”, *Nat. Methods* (2019).
37. Romain F. Laine, Kalina L. Tosheva, Nils Gustafsson, Robert D. M. Gray, Pedro Almada, David Albrecht, Gabriel T. Risa, Fredrik Hurtig, Ann-Christin Lindås, Buzz Baum, Jason Mercer, Christophe Leterrier, Pedro M. Pereira*, Siân Culley*, Ricardo Henriques*, “[NanoJ: a high-performance open-source super-resolution microscopy toolbox](#)”, *J. Phys. D* (2019). **Key publication.**
36. Pedro M. Pereira*, David Albrecht*, Siân Culley, Caron Jacobs, Mark Marsh, Jason Mercer, Ricardo Henriques*, “[Fix your membrane receptor imaging: Actin cytoskeleton and CD4 membrane organization disruption by chemical fixation](#)”, *Frontiers in Immunology* (2019). **Key publication.**

35. Lucas von Chamier, Romain F Laine, Ricardo Henriques*, “[Artificial Intelligence for Microscopy: What You Should Know](#)”, *Biochem. Soc. Transactions* (2019). **Key publication.**
34. Lekha Patel, Nils Gustafsson, Yu Lin, Raimund Ober, Ricardo Henriques, Edward Cohen, “[A hidden Markov model approach to characterizing the photo-switching behavior of fluorophores](#)”, *Annals of Applied Statistics* (2019).
33. Christopher Bricogne, Michael Fine, Pedro M Pereira, Youxue Wang, Julia Sung, Maha Tijani, Ricardo Henriques, Mary K Collins, Donald Hilgemann, “[TMEM16F activation by Ca²⁺ triggers plasmalemma expansion and directs PD-1 trafficking](#)”, *Sci. Rep.* (2019).
32. Justin Cooper, Mark Browne, Hugh Gribben, Martin Catney, Colin Coates, Alan Mullan, Geraint Wilde, Ricardo Henriques, “[Real time multi-modal super-resolution microscopy through Super-Resolution Radial Fluctuations \(SRRF-Stream\)](#)”, *Single Molecule Spectroscopy and Superresolution Imaging XII* (2019).
31. Siân Culley, David Albrecht, Caron Jacobs, Pedro Matos Pereira, Christophe Leterrier*, Jason Mercer*, Ricardo Henriques*, “[Quantitative mapping and minimization of super-resolution optical imaging artifacts](#)”, *Nat. Methods* (2018). **Key publication.**
30. Martin Weigert, Uwe Schmidt, Tobias Boothe, M Andreas, Alexander Dibrov, Akanksha Jain, Benjamin Wilhelm, Deborah Schmidt, Coleman Broadbush, Siân Culley, Maurício Rocha-Martins, Fabián Segovia-Miranda, Caren Norden, Ricardo Henriques, Marino Zerial, Michele Solimena, Jochen Rink, Pavel Tomancak, Loic Royer, Florian Jug, Eugene W Myers, “[Content-Aware Image Restoration: Pushing the Limits of Fluorescence Microscopy](#)”, *Nat. Methods* (2018).
29. Sina Krokowski, Damián Lobato-Márquez, Arnaud Chastanet, Pedro Matos Pereira, Dimitrios Angelis, Dieter Galea¹, Gerald Larrouy-Maumus, Ricardo Henriques, Elias T. Spiliotis, Rut Carballido-López, Serge Mostowy, “[Septins Recognise Bacterial Cell Division for Host Defence](#)”, *Cell Host & Microbe* (2018).
28. Christina L Dix, Helen K Matthews, Marina Uroz, Susannah McLaren, Lucie Wolf, Nicholas Heatley, Zaw Win, Pedro Almada, Ricardo Henriques, Michael Boutros, Xavier Trepast, Buzz Baum, “[The Role of Mitotic Cell-Substrate Adhesion Re-modeling in Animal Cell Division](#)”, *Dev. Cell* (2018).
27. Felix Weihs, Katarzyna Wacnik, Robert D Turner, Siân Culley, Ricardo Henriques, Simon J Foster, “[Heterogeneous localisation of membrane proteins in Staphylococcus aureus](#)”, *Sci. Rep.* (2018).
26. Siân Culley, Kalina L Tosheva, Pedro Matos Pereira, Ricardo Henriques*, “[SRRF: Universal live-cell super-resolution microscopy](#)”, *Int. J. Biochem. Cell Biol.* (2018).
25. David Tomaz, Pedro Matos Pereira, Nadia Guerra, Julian Dyson, Ricardo Henriques*, Keith Gould*, “[Nanoscale colocalization of NK cell activating and inhibitory receptors controls signal integration](#)”, *bioRxiv, in revision* (2018).
24. Robert DM Gray, Jason Mercer*, Ricardo Henriques*, “[Open-source Single-particle Analysis for Super-resolution Microscopy with VirusMapper](#)”, *J. Vis. Exp.* (2017).
23. Andrea Sirianni, Sina Krokowski, Damián Lobato-Márquez, Stephen Buranyi, Julia Pfanzelter, Dieter Galea, Alexandra Willis, Siân Culley, Ricardo Henriques, Gerald Larrouy-Maumus, Michael Hollinshead, Vanessa Sancho-Shimizu, Michael Way, Serge Mostowy, “[Mitochondria mediate septin cage assembly to promote autophagy of Shigella](#)”, *EMBO Rep.* (2016).

22. Janine Scholefield[±], Ricardo Henriques[±], Anca F Savulescu, Elisabeth Fontan, Alix Boucharlat, Emmanuel Laplantine, Asma Smahi, Alain Israël, Fabrice Agou, Musa M Mhlanga, “[Super-resolution microscopy reveals a preformed NEMO lattice structure that is collapsed in incontinentia pigmenti](#)”, *Nat. Communications* (2016). **Key publication.**
21. Siân Culley, Greg J Towers, David L Selwood, Ricardo Henriques, Joe Grove, “[Infection Counter: Automated Quantification of in Vitro Virus Replication by Fluorescence Microscopy](#)”, *Viruses* (2016).
20. Barbara Clough, Joseph D Wright, Pedro M Pereira, Elizabeth M Hirst, Ashleigh C Johnston, Ricardo Henriques, Eva-Maria Frickel, “[K63-Linked Ubiquitination Targets Toxoplasma gondii for Endo-lysosomal Destruction in IFN-Stimulated Human Cells](#)”, *PLOS Pathog.* (2016).
19. Robert DM Gray, Corina Beerli, Pedro Matos Pereira, Kathrin Maria Scherer, Jerzy Samolej, Christopher Karl Ernst Bleck, Jason Mercer*, Ricardo Henriques*, “[VirusMapper: open-source nanoscale mapping of viral architecture through super-resolution microscopy](#)”, *Sci. Rep.* (2016). **Key publication.**
18. Nils Gustafsson, Siân Culley, George Ashdown, Dylan M Owen, Pedro Matos Pereira, Ricardo Henriques*, “[Fast live-cell conventional fluorophore nanoscopy with ImageJ through super-resolution radial fluctuations](#)”, *Nat. Communications* (2016). **Key publication.**
17. Joana G Silva, Nuno P Martins, Ricardo Henriques, Helena Soares, “[HIV-1 Nef Impairs the Formation of Calcium Membrane Territories Controlling the Signaling Nanoarchitecture at the Immunological Synapse.](#)”, *J. Immunol.* (2016).
16. Pedro Almada, Siân Culley, Ricardo Henriques*, “[PALM and STORM: Into large fields and high-throughput microscopy with sCMOS detectors](#)”, *Methods* (2015).
15. Pedro M Pereira, Pedro Almada, Ricardo Henriques*, “[High-content 3D multicolor super-resolution localization microscopy](#)”, *Methods Cell Biol.* (2015).
14. Nadine Tarantino, Jean-Yves Tinevez, Elizabeth Faris Crowell, Bertrand Boisson, Ricardo Henriques, Musa Mhlanga, Fabrice Agou, Alain Israël, Emmanuel Laplantine, “[TNF and IL-1 exhibit distinct ubiquitin requirements for inducing NEMO-IKK supramolecular structures](#)”, *J Cell Biol* (2014).
13. Helena Soares, Ricardo Henriques, Martin Sachse, Leandro Ventimiglia, Miguel A Alonso, Christophe Zimmer, Maria-Isabel Thoulouze, Andrés Alcover, “[Regulated vesicle fusion generates signaling nanoterritories that control T cell activation at the immunological synapse](#)”, *J. Exp. Med.* (2013).
12. Mickaël Lelek, Francesca Di Nunzio, Ricardo Henriques, Pierre Charneau, Nathalie Arhel, and Christophe Zimmer, “[Superresolution imaging of HIV in infected cells with FLAsH-PALM](#)”, *PNAS* (2012).
11. Sébastien Herbert, Helena Soares, Christophe Zimmer, Ricardo Henriques*, “[Single-Molecule Localization Super-Resolution Microscopy: Deeper and Faster](#)”, *Microscopy and Microanalysis* (2012).
10. Sébastien Herbert, Ricardo Henriques*, “[Enhanced epifluorescence microscopy by uniform and intensity optimized illumination](#)”, *Cytometry A.* (2012).

9. Soudeh Ehsani, José Carlos Santos, Cristina D. Rodrigues, Ricardo Henriques, Laurent Audry, Christophe Zimmer, Philippe Sansonetti, Guy Tran Van Nhieu, Jost Enninga, “[Hierarchies of host factor dynamics at the entry site of Shigella flexneri during host cell invasion](#)”, *Infection and immunity* (2012).
8. Carina S. S. Gomes-Santos, Maurice A. Itoe, Cristina Afonso, Ricardo Henriques, Rui Gardner, Nuno Sepúlveda, Pedro D. Simões, Helena Raquel, António Paulo Almeida, Luis F. Moita, Friedrich Frischknecht, Maria M. Mota, “[Highly Dynamic Host Actin Reorganization around Developing Plasmodium Inside Hepatocytes](#)”, *PLoS One* (2012).
7. Caron Griffiths (now Caron Jacobs), Ricardo Henriques, Musa M Mhlanga, “[PALM and STORM: a super-resolution molecular view into living cells](#)”, *Biotechnology International* (2011).
6. Ricardo Henriques, Caron Griffiths (now Caron Jacobs), Esper H Rego, Musa M Mhlanga, “[PALM and STORM: Unlocking live-cell super-resolution](#)”, *Biopolymers* (2011).
5. Ricardo Henriques*, Mickael Lelek, Eugenio F Fornasiero, Flavia Valtorta, Christophe Zimmer, Musa M Mhlanga, “[QuickPALM: 3D real-time photoactivation nanoscopy image processing in ImageJ](#)”, *Nat. Methods* (2010). **Key publication.**
4. Ricardo Henriques, Musa M Mhlanga, “[PALM and STORM: What hides beyond the Rayleigh limit?](#)”, *Biotech. Journal* (2009).
3. Luis B Barreiro, Ricardo Henriques, Musa M Mhlanga, “[High-throughput SNP genotyping: combining tag SNPs and molecular beacons](#)”, *Meth. Molecular Biology* (2009).
2. José Rino, José Braga, Ricardo Henriques, Maria Carmo-Fonseca, “[Frontiers in fluorescence microscopy](#)”, *International J. of Dev. Bio.* (2009).
1. J. Caetano-Lopes, A. Nery, R. Henriques, H. Canhão, J. Duarte, P. Amaral, M. Vale, R. Moura, P. Pereira, P. Weinmann, S. Abdulghani, M. Souto-Carneiro, P. Rego, J. Monteiro, S. Sakagushi, M. Viana Queiroz, Y. Konttinen, L. Graça, M. Vaz, J. Fonseca, “[Chronic arthritis directly induces quantitative and qualitative bone disturbances leading to compromised biomechanical properties](#)”, *Clin Exp Rheumatol.* (2009).