R E S C I E N C E C

[Re] Modeling Insect Phenology Using Ordinal Regression and Continuation Ratio Models

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1 Introduction

Phenology, the timing of seasonal biological phenomena, is a key aspect of plant and animal life. It defines the timing and duration of growth and reproduction and thereby determines the ability to capture seasonally variable resources [1]. Phenological analyses often focus on the timing of particular events, such as the dates of peak caterpillar abundance [2]. However, for many biological phenomena exact dates of particular events are more difficult to observe than the state of the system itself. For example, repeated but sparse survey visits may record whether a plant is in bud, flowering, or setting fruit, but not the exact dates when each of those stages was reached. Such observations can be used to categorize an organism's state into discrete classes which usually follow natural ordering, e.g. from least to most developed. The resulting data can be described using ordinal regression models [3, 4] which then allow inferences about phenological progression.

I here replicate a number of ordinal regression models that were employed by Dennis, Kemp, and Beckwith[5] and Candy[6] to describe insect phenology.

2 Data

The models replicated in this study are fitted to a data set on the phenology of the western spruce budworm *Choristoneura freemani* (Lepidoptera: Tortricidae), a defoliating moth that is widespread in western North America [7]. This data set was originally published in [5] and is a subset of a larger budworm survey data set analysed in [8]. The data consist of 12 sampling occasions at which counts of individual budworms in each of seven development stages (five larval instars, pupae, and adults) were recorded. The only available covariate is a measure of seasonal progression, the accumulated degree days calculated using a threshold of 5.5°C. Candy[6] noted an inconsistency in these data, namely that the reported total number of individuals did not correspond to the sum across the seven development stages for two of the sampling occasions. I therefore use the data set as it was republished in [9], where numbers in each stage have been assumed correct and the totals for each sampling occasion were adjusted accordingly.

Table 1. The data used in this replication are counts of western spruce budworm *Choristoneura freemani* across seven developmental stages on 12 sampling occasions, as published in [9].

Degree Days	Stage 1	Stage 2	Stage 3	Stage 4	Stage 5	Stage 6	Stage 7
58	16	0	0	0	0	0	0
82	10	0	0	0	0	0	0
107	23	7	0	0	0	0	0
155	3	44	0	0	0	0	0
237	0	6	45	13	0	0	0
307	0	2	9	48	15	0	0
342	0	0	1	34	37	0	0
388	0	0	1	10	87	5	0
442	0	0	0	7	53	21	0
518	0	0	0	0	10	65	1
609	0	0	0	0	0	14	26
685	0	0	0	0	0	0	42

3 Methods

The statistical models replicated here are different types of ordinal regression models [4], all with the aim of predicting the proportion of an insect population in a particular development stage at any given time. In particular, they represent three different parameterisations of the so-called cumulative model and one version of the so-called sequential model. A summary of the theory underlying these models and their derivation is provided in [10].

Both model types assume that the development of an insect follows an unobservable stochastic process $S(t)$ made up of accumulated increments of development over time t. As $S(t)$ increases, the insect passes through successive stages $j = 1, \ldots, r$, delimited by $r - 1$ moults, with the jth moult occurring when the development threshold a_j is reached:

$$
\begin{aligned}
\text{stage } 1 : &\qquad S(t) \leq a_1 \\
\text{stage } 2 : &\quad a_1 < S(t) \leq a_2 \\
&\qquad \vdots \\
\text{stage } r-1 : &\quad a_{r-2} < S(t) \leq a_{r-1} \\
\text{stage } r : &\quad a_{r-1} < S(t)
\end{aligned}
$$

The a_j values are typically unknown and their estimation from observed data was the goal of the orginal studies [5, 6].

3.1 Cumulative model with constant variance

The ordinal regression model of [6] is specified in terms of the cumulative number of individuals $m_{ij} = \sum_{k=1}^{j} n_{ik}$ observed in stages 1 to j on a sampling occasion i.

$$
\mathbf{E}(m_{ij}) = N_i Pr(S(t) < \alpha_j), \qquad j = 1, \ldots, r \tag{1}
$$
$$
= N_i G(\alpha_j + \beta z_i) \tag{2}
$$

where G is the cumulative probability density function of $S(t)$, α_j are ordered thresholds or cut-point parameters, β is a vector of regression parameters and z_i is a vector of predictor variables. If the probability of an individual being in stage j or earlier at time t_i is

$$\mu_{ij} = \mathbf{E}(m_{ij})/N_i$$

48 one can interpret G^{-1} as the link function of a generalised linear model (GLM) with the
49 linear predictor

$$\eta_{ij} = \alpha + \beta z_i$$

50 This ordinal regression model is commonly known as the cumulative model [10], and is
51 applied to the budworm data in [6] using the logit and complementary log-log (cloglog)
52 link functions. In both cases the parameterisation results in a constant variance for
53 $S(t)$. For the purpose of parameter estimation Candy[6] re-expressed the model in terms
54 of stage-specific counts n_{ij}

$$\mathbf{E}(n_{ij}) = N_i\{G(\alpha_j + \beta z_i) - G(\alpha_{j-1} + \beta z_i)\} \tag{3}$$

55 and fits it using a Poisson likelihood [11]. No code or initial values for the likelihood
56 optimisation are provided for this estimation procedure in the original paper. I there-
57 fore created an R version of the estimation procedure which directly optimizes a Poisson
58 log-likelihood for Equation 3 using the BFGS method in the `optim` function and a pa-
59 rameter scaling value of 0.01 for β relative to the cut-point parameters.. Initial values for
60 the optimisation were determined using a random search across plausible start values
61 (see Appendix A.1). The cumulative model is implemented in various R packages, in-
62 cluding in the `vglm` function in VGAM [12] and the `clm` function in `ordinal` [13] and for
63 comparison I also attempted to fit the models using these functions with their default
64 settings.

3.2 Cumulative model with proportional variance

66 Dennis, Kemp, and Beckwith[5] used a different parameterisation of the ordinal model
67 with a logit link, i.e. assuming a logistic distribution for $S(t)$, such that the probability
68 that an insect's development at time t has not exceeded s amounts to

$$Pr(S(t) \le s) = \left\{1 + \exp\left[-\left(\frac{s-t}{\sqrt{b^2 t}}\right)\right]\right\}^{-1} \tag{4}$$

69 where b^2 is a positive constant. The cumulative distribution function in Eqn. 4 corre-
70 sponds to a logistic distribution with a mean of t and a variance which increases pro-
71 portional to the mean as $(\pi^2/3)b^2 t$. At any fixed time t the thresholds a_j segment the
72 probability distribution function into r parts and the area under the curve between a_{j-1}
73 and a_j gives the probability that the insect will be in stage j at time t.
74 This modelling approach is applied to data consisting of samples i that record the num-
75 ber of insects x_{ij} in stage j at times $t_1, t_2, \ldots, t_q$ and the x_{ij} are assumed to be random
76 samples from a multinomial distribution with corresponding multinomial probabili-
77 ties p_{ij}

$$p_{ij} = Pr(a_{j-1} < S(t_i) \le a_j) \tag{5}$$

$$= \left\{1 + \exp\left[-\left(\frac{a_j - t_i}{\sqrt{b^2 t_i}}\right)\right]\right\}^{-1} - \left\{1 + \exp\left[-\left(\frac{a_{j-1} - t_i}{\sqrt{b^2 t_i}}\right)\right]\right\}^{-1} \tag{6}$$

78 To fulfill the constraint that $\sum_{j=1}^r p_{ij} = 1$, it is further assumed that $a_0 = -\infty$ and
79 $a_r = +\infty$. The model has r unknown parameters $a_1, \ldots, a_{r-1}$ and b^2 which can be
80 found by maximising the corresponding log-likelihood function

$$\ell = \log C + \sum_{j=1}^r \sum_{i=1}^q x_{ij} \log p_{ij} \tag{7}$$

81 where C is a combinatorial constant that is independent of the parameter values.
82 Dennis, Kemp, and Beckwith[5] provided SAS code and initial values to estimate the pa-
83 rameters under this likelihood using an iteratively reweighted non-linear least squares
84 approach based on `PROC NLIN`. This was updated to run in a contemporary version of
85 SAS (SAS 9.4) and is provided in the article code repository. However, since SAS is a
86 proprietary software package, I implemented two R versions of the estimation proce-
87 dure which both directly optimize the log-likelihood (7) using the L-BFGS-B optimisa-
88 tion routine [14] in the R `optim` function. The first implementation is a direct trans-
89 lation of the SAS code which uses a literal implementation of the logistic cumulative
90 distribution function and employs a lower parameter bound of 0 and initial values pro-
91 vided in [5] for the estimation. The second implementation makes use of the R function
92 `stats::plogis` to implement the logistic CDF and employs a lower parameter bound
93 of the machine precision (i.e. `.Machine$double.xmin`) and the same initial values
94 as above. Both routines used a parameter scaling value of 0.01 for b^2 relative to the cut-
95 point parameters
96 Candy[6] re-expressed the proportional variance model (Eqn. 6) to match the form of
97 Eqn. 3, which results in the following reparameterisation $\alpha_j = a_j/b$, $\beta = -1/b$, and
98 $z_i = \sqrt{t_i}$, and uses the Poisson likelihood approach described in section 3.1 for parame-
99 ter estimation. A set of example macros for the software package `GLIM` [15] is provided
100 in an earlier manuscript by the same author [9]. GLIM is no longer actively developed
101 or distributed, but initial values from the GLIM code were used in the estimation with
102 the BFGS method of R `optim` and a parameter scaling value of 0.01 for β relative to the
103 cut-point parameters.

3.3 Sequential model
104

105 A different class of ordinal regression model can be derived by treating the observations
106 as the result of a strictly ordered counting process, i.e. to achieve a stage j, all lower
107 stages $1, \ldots, j - 1$ have to be achieved first. The general form of this model is known
108 as the sequential model, and rather than assuming a single latent process $S(t)$ as in
109 the cumulative model there is a latent continuous variable S_j for each category j [10].
110 Analogous to the cumulative model it can be framed as a GLM

$$S_j = \eta + \epsilon_j \tag{8}$$

111 with a linear predictor η and an error term ϵ_j which has mean zero and is distributed
112 following some distribution G. This leads to a model of the form

$$Pr(S = a_j | S \geq a_j, \eta) = G(a_j - \eta) \tag{9}$$

113 When G is the logistic distribution this model is also known as the continuation ratio
114 model [16, 10]. Confusingly, there are two common versions of the model in the liter-
115 ature both using this name. The one outlined above describing the probability of the
116 sequential process *stopping* at stage j, and the other describing the probability of the
117 process *continuing* beyond stage j, i.e.
118 $Pr(S > a_j | S \geq a_j, \eta)$ [10, 12].
    The paper replicated here [6] used the stopping parameterisation. In their notation the
    expected value for the stage-specific counts n_{ij} is

$$\begin{aligned} \mathrm{E}(n_{ij}) &= N_i G(\beta_{01} + \beta_{11} t_i), & j = 1 \\ &= N_{ij}^* G(\beta_{0j} + \beta_{1j} t_i), & j = 2, \ldots, r - 1 \end{aligned} \tag{10}$$

119 with

$$N_{ij}^* = \left(N_i - \sum_{k=1}^{j-1} n_{ik} \right) \tag{11}$$

Table 2. Parameter estimates for the cumulative model with constant variance (Eqn. 3). This table replicates results presented in the first two rows of Table 2 of [6]. Note that `ordinal::clm` uses a parameterisation $\alpha_j - \beta z_i$ for the linear predictor yielding a parameter estimate for β with the opposite sign than the other methods. The cloglog link model failed to fit using `VGAM::vglm`.

Method	α_1	α_2	α_3	α_4	α_5	α_6	β	Link
Original [6]	5.49	9.39	12.23	15.70	21.26	27.25	-0.0457	logit
R optim	5.47	9.36	12.21	15.67	21.22	27.19	-0.0456	logit
R clm	5.47	9.36	12.21	15.67	21.22	27.19	0.0456	logit
R vglm	5.47	9.36	12.21	15.67	21.22	27.19	-0.0456	logit
Original [6]	3.32	5.85	7.72	10.03	13.46	17.52	-0.0307	cloglog
R optim	3.32	5.85	7.71	10.02	13.46	17.52	-0.0307	cloglog
R clm	3.32	5.85	7.71	10.02	13.46	17.52	0.0307	cloglog
R vglm	NA	NA	NA	NA	NA	NA	NA	cloglog

Table 3. Parameter estimates for the cumulative logit model with proportional variance. This table replicates results presented in the first row of Table 1 of [8] and the last row of Table 2 of [6].

Method	a_1	a_2	a_3	a_4	a_5	a_6	b^2	Eqn.
Original [8]	121.080	204.360	264.410	342.473	465.620	599.570	1.559	6
SAS NLIN	120.000	204.700	264.600	341.300	464.500	595.700	1.412	6
R optim	120.039	204.665	264.590	341.291	464.477	595.707	1.412	6
R optim/plogis	120.039	204.665	264.590	341.291	464.477	595.707	1.412	6
Method	α_1	α_2	α_3	α_4	α_5	α_6	β	Eqn.
Original [6]	101.000	172.200	222.700	287.200	390.900	501.300	-0.842	3
R optim	100.923	172.054	222.449	286.947	390.522	500.852	-0.841	3

and conditional probabilities

$$p_{ij}^* = G(\beta_{0j} + \beta_{1j}t_i), \qquad j = 1, \ldots, r-1 \tag{12}$$

Candy[6] uses GLM estimation routines in `GLIM` to fit the $r-1$ models defined in Eqn. 10 assuming the n_{ij} are binomially distributed conditional on N_i for stage 1 and conditional on N_{ij}^* for stages $2, \ldots, r-1$. No code is provided for this estimation procedure in the original paper, but the model is straightforward to implement using the `glm` function in R with a model formula of the form

```
cbind(count, total - N_star) ~ stage + stage:time - 1
```

where the variable `count` represents the n_{ij}, `N_star` represents N_i for $j = 1$ and N_{ij}^* for all other observations, `stage` is a factor variable encoding j and `time` are the t_i. The sequential model with stopping parameterisation is also implemented in `VGAM::vglm` [12] and for comparison I attempted to fit the model using this function with its default settings.

4 Results

4.1 Cumulative model with constant variance

Parameters were estimated using a direct optimisation of the Poisson likelihood for Eqn. 3, as well as with the R functions `VGAM::vglm` and `ordinal::clm`. The cloglog link model failed due to numerical errors when using the `vglm` function. Parameter

131 estimates were close to those of the original study for all three methods employed (Ta-
132 ble 2). Parameter estimates were generally insensitive to the choice of starting values
133 once the optimisation function was provided with appropriate parameter scaling values
134 for the optimisation routine (see Appendix).

4.2 Cumulative model with proportional variance

136 The original SAS code provided in [5] required minimal updates to run in a contemporary
137 version of SAS (SAS 9.4). Translating the model code to R was straightforward once I
138 took the decision to implement a direct minimisation of the negative log likelihood with
139 `optim`.
140 Parameter estimates from SAS `NLIN` and both R `optim` implementations (Table 3) were
141 virtually identical, but differed slightly from the parameter estimates presented in [8],
142 which was assumed to be the original source for the parameter estimates, as no param-
143 eter estimates were presented in [5]. The observed differences in estimates were $< 1\%$
144 for the cut-point parameters α_i, but c. 9% for β.
145 Based on these three sets of parameter estimates it was also possible to redraw two fig-
146 ures from [5]. Figure 1 and 2, respectively, show that despite the observed parameter
147 differences there is an overall good agreement between the original results and the repli-
148 cation.

Table 4. Parameter estimates for the sequential model with stopping ratios (Equation 10). This table replicates results presented in Table 3 of [6]. The cloglog link model failed to fit using `VGAM::vglm`.

Method	Parameter	$\beta_{\_1}$	$\beta_{\_2}$	$\beta_{\_3}$	$\beta_{\_4}$	$\beta_{\_5}$	$\beta_{\_6}$	Link
Original [6]	β_{0j}	10.410	12.960	12.020	11.160	17.700	33.730	logit
R glm	β_{0j}	10.410	12.959	12.020	11.165	17.698	33.726	logit
R vglm	β_{0j}	10.410	12.959	12.020	11.165	17.698	33.726	logit
Original [6]	β_{1j}	-0.085	-0.062	-0.046	-0.033	-0.038	-0.056	logit
R glm	β_{1j}	-0.085	-0.062	-0.046	-0.033	-0.038	-0.056	logit
R vglm	β_{1j}	-0.085	-0.062	-0.046	-0.033	-0.038	-0.056	logit
Original [6]	β_{0j}	7.350	8.530	9.120	8.440	10.090	16.300	cloglog
R glm	β_{0j}	7.347	8.537	9.124	8.442	10.087	16.298	cloglog
R vglm	β_{0j}	NA	NA	NA	NA	NA	NA	cloglog
Original [6]	β_{1j}	-0.065	-0.044	-0.037	-0.026	-0.023	-0.029	cloglog
R glm	β_{1j}	-0.065	-0.044	-0.037	-0.026	-0.023	-0.029	cloglog
R vglm	β_{1j}	NA	NA	NA	NA	NA	NA	cloglog

4.3 Sequential model

150 Parameters were estimated using the R `glm` function and `VGAM::vglm`. The GLM for-
151 mulation of the model fitted with R `glm` produced a warning that fitted probabilities
152 numerically 0 or 1 occurred. Likely due to related reasons, the cloglog link model failed
153 due to numerical errors when using the `vglm` function. Parameter estimates, where
154 obtained, were identical to the original study (Table 4) within the precision reported in
155 [6].

5 Discussion

157 Overall the results from both [5] and [6] could be replicated closely. The SAS code pro-
158 vided in [5] required only minimal updates to run in a contemporary version of SAS (SAS
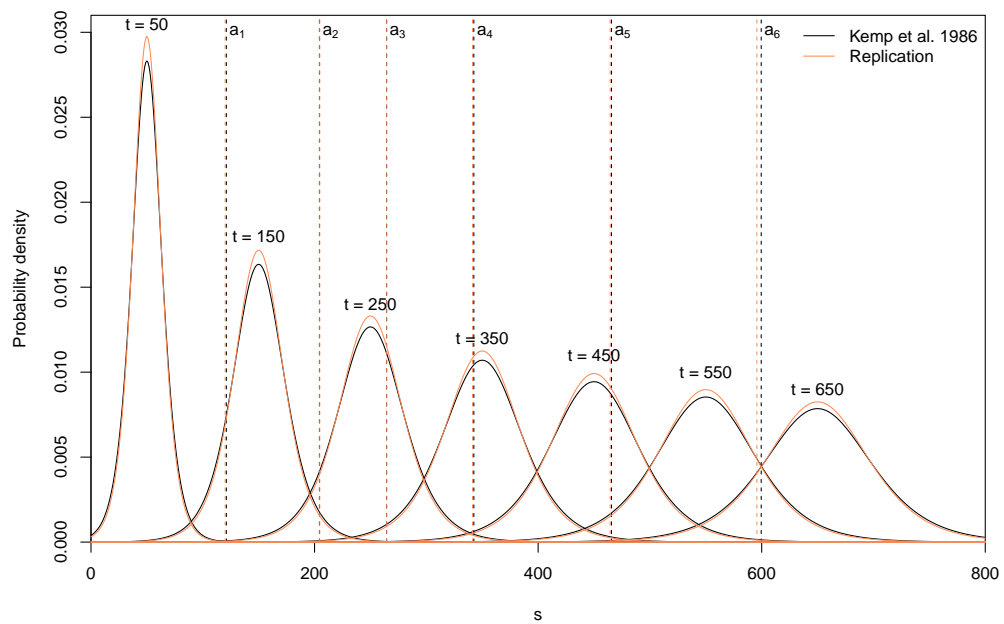
Figure 1. Logistic PDF of the cumulative model with proportional variance (Equation 6) plotted for fixed values of t. Area under the PDF between a_{j-1} and a_j gives the expected proportion of insects in stage j at time t. The graph is based on the estimates in Table 1 of [8] (black lines) and the estimates from the replication using the R `optim` implementation (red lines). This figure replicates Figure 2 in [5].

9.4) and produced virtually identical estimates to the R re-implementation. These estimates, however, differed slightly from the parameter estimates reported in [8]. Given that the same initial values were used in all implementations, I believe that this disagreement is most likely caused by the inconsistencies in the published data set described in Section 2. The corrections applied to the data by [6] result in a data set that is internally consistent but likely different to that on which the estimates in [8] are based.

No code was provided in [6]. However, the mathematical and verbal descriptions of the models were detailed enough to re-implement the estimation procedures in R. GLIM code of the cumulative model with proportional variance was available from an earlier manuscript [9]. This allowed me to use the same initial values as the original study for this model. Initial values for the model with constant variance had to be guessed. The direct optimisation of the likelihood was reported to be sensitive to the choice of initial values in [5]. Additional simulations (Appendix A.1) showed that convergence failures did occur for some initial values, particularly in the case for the cloglog-link model. However, the coefficient of variation for estimates derived from a wide range of initial values was very small (<0.01%) for both the logit and cloglog models, whenenver the model converged succesfully (Table S1).

Literal implementations of both the inverse logit and inverse complementary log-log function can suffer from numerical underflow and/or overflow for probabilities close to zero and one, and this may be the key reason for convergence failures observed in the direct optimisation, as well as in the `vglm` function. The case study presented here is based on observations where the population variation in development speed (i.e. the transition from class to class) is low compared to the overall speed of seasonal progression. As a result the early and late development stages of the observed species never occur together, and the observed data are sparse in the sense that many cell counts in Table 1 are zero. As a consequence the model estimates large effects, which although estimable on the link scale, are indistinguishable from one on the probability scale given the limitations of floating point representations. This is the underlying reason for the warnings produced by the `glm` function when fitting the sequential model. The GLIM code for the cumulative model from [9] uses multiple thresholding steps during the calculation of the linear predictor to mitigate against this, whereas the R implementation of the corresponding model makes use of a single thresholding step in the inverse link function (`gtools::inv.logit` [17] or `stats::plogis` and `VGAM::clogloglink`, respectively). Differences in parameter estimates between a literal implementation of the logit link and `stats::plogis` for the cumulative model with proportional variance (Table 4) were less than 0.01%, however, in the case of the constant variance model the complementary log-log link model proved to be numerically less stable than the corresponding logit-link model (Figure S1). This is perhaps unsurprising as the cloglog function has a steeper slope as it approaches one compared to the logit function, so values numerically close to one are attained earlier. Given the sparsity of the data set and the resulting large estimated effect sizes this likely contributed to the failure to fit either of the cloglog-link models using the `VGAM::vglm` function. The behaviour was reported to the maintainer of the `VGAM` package, and initial troubleshooting of the `VGAM` code resolved some instabilities for the fitting of the cumulative model. These updates are now available in the pre-release version of `VGAM`[1]. However, other instabilities remain. The cumulative model using the updated `VGAM` code converges far from the optimum identified by the other methods and a fix for the error affecting the sequential model are yet to be implemented.

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[1]https://www.stat.auckland.ac.nz/ yee/VGAM/prerelease/index.html

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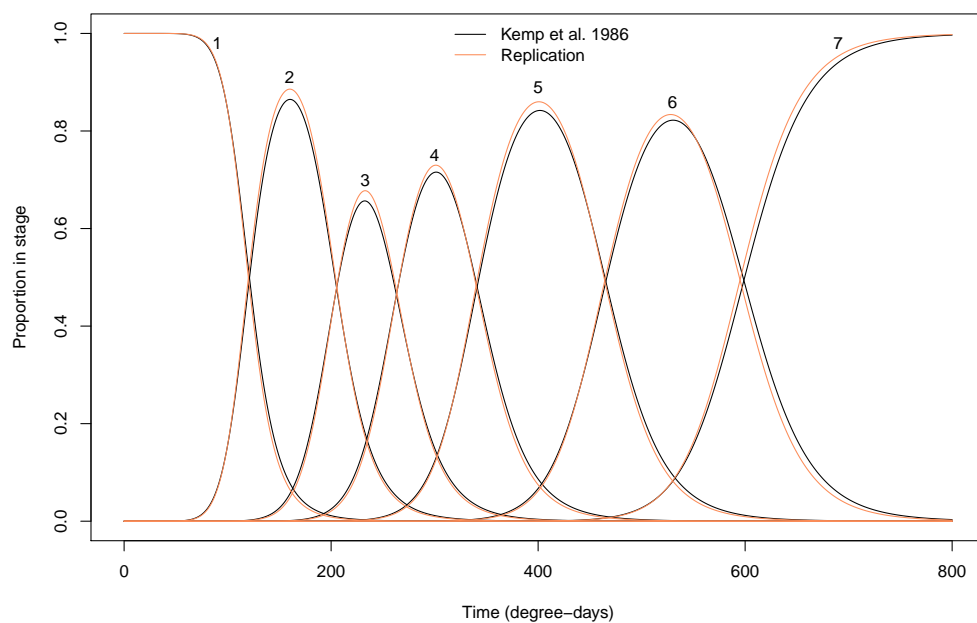
Figure 2. Proportion of insects expected in stages 1-7 under the cumulative logit model with proportional variance (Equation 6) plotted as functions of time t. Values of a_j and b^2 used in the graph are the estimates given in Table 1 of [8] (black lines) and the estimates from the replication using the R `optim` implementation (red lines). This figure replicates Figure 3 in [5].

A Appendix: Sensitivity of the optimisation procedure to initial values

A.1 Approach

The sensitivity of parameter estimates for the cumulative model with constant variance (Eqn. 3) obtained using the direct likelihood optimisation with `optim` was assessed by simulation. Simulations consisted of drawing a set of random initial values. To preserve the ordering of the cut-point parameters α_i, their initial values α_i° were assembled as the cumulative sum of six independent draws

$$x_j \sim \text{Uniform}(1, 20), \qquad j = 1, \ldots, 6$$

i.e.

$$\alpha_i^\circ = \sum_{j=1}^{i} x_j.$$

The initial value for β was drawn as

$$\beta^\circ \sim \text{Uniform}(-1, 1).$$

Optimisation proceeded using the likelihood outlined in Section 3.1 with a numerical threshold at the machine's precision to prevent the Poisson likelihood from approaching negative infinity. The optimisation routine used the BFGS method and parameter scaling value for β of 0.01 relative to the cut-point parameters α_i. This improved overall convergence rates but allowed convergence at an infinite likelihood, the likelihood was therefore evaluated one final time without thresholding at the converged parameter values and parameters were only retained when the unthresholded likelihood was finite. Parameters were further filtered to remove parameter sets where convergence was achieved at a finite likelihood, but with a substantially larger overall log-likelihood than the best model fit. Thresholds for this were determined empirically and set at 93 for the logit link and 100 for the cloglog model.

A.2 Results

The optimisation was reasonably robust for the logit link model. 73% of simulations achieved convergence at a finite likelihood and of these over 99% met the filtering threshold (Figure S1). Among the filtered parameter sets variability in the final estimates was very small (parameter-wise coefficients of variation were all smaller than 0.01%) and there was no correlation between initial values and parameter estimates when taking into account that multiple comparisons were made (Table S1). Convergence failed for initial values $\beta^\circ > 0.7$, and convergence failures were likely, but not guaranteed for $\beta^\circ \approx 0$ (Figure S2).

The optimisation of the cloglog model was less stable. Only 47% of simulations resulted in convergence, and of those 92% met the filtering threshold (Figure S1). Variability in the filtered parameter estimates was larger than for the logit model, but still small in absolute terms (parameter-wise coefficients of variation were all below 0.01%) and there was no correlation between initial values and parameter estimates (Table S1). Convergence failed for many $0 < \beta^\circ < 0.2$, and always for $\beta^\circ > 0.2$ (Figure S3).

Table S1. Summary statistics of parameter sensitivities to starting values for the cumulative model with constant variance (Eqn. 3). The coefficients of variation (CV), correlation coefficients between initial and converged values ρ^o, and their corresponding p-values P_ρ were calculated for the filtered parameter estimates only and adjusted for multiple comparisons using the Bonferroni correction.

| Par.	Link	Sim. range	Conv. range	Filtered range	CV (%)	ρ^o	P_ρ
α_1	logit	1.25 - 19.97	1.3 - 20	1.3 - 20	0.003	-0.037	1.000
α_2	logit	3.11 - 37.8	3.1 - 37.6	3.1 - 37.6	0.003	-0.050	1.000
α_3	logit	5.45 - 54.08	5.4 - 54.1	5.4 - 54.1	0.003	-0.111	0.706
α_4	logit	13.04 - 69.89	15.9 - 69.9	15.9 - 69.9	0.003	-0.135	0.322
α_5	logit	22.87 - 84.96	22.9 - 85	22.9 - 85	0.003	-0.101	0.956
α_6	logit	31.59 - 100.68	32.7 - 100.7	32.7 - 100.7	0.003	-0.087	1.000
β	logit	-0.99 - 0.99	-1 - 0.7	-1 - 0.7	0.003	-0.089	1.000
α_1	cloglog	1.09 - 19.95	1.1 - 19.9	1.1 - 19.9	0.006	0.031	1.000
α_2	cloglog	3.28 - 38.92	4 - 37.3	4 - 37.3	0.004	-0.069	1.000
α_3	cloglog	7.88 - 57.02	9.7 - 52.4	9.7 - 52.4	0.005	-0.085	1.000
α_4	cloglog	15.01 - 66.97	16.3 - 66.9	16.3 - 66.9	0.004	-0.109	1.000
α_5	cloglog	18.42 - 77.81	18.4 - 76.3	18.4 - 76.3	0.004	-0.123	1.000
α_6	cloglog	22.78 - 91.58	30.7 - 91.6	30.7 - 91.6	0.004	-0.115	1.000
β	cloglog	-1 - 1	-1 - 0.2	-1 - 0.2	0.004	-0.038	1.000
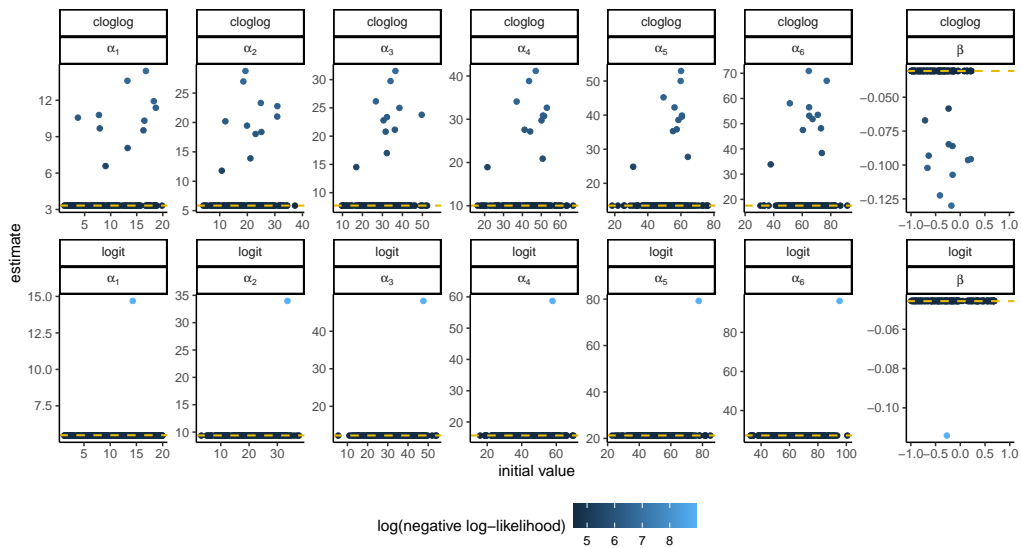
Figure S1. Optimisation results for randomly drawn initial values for the cumulative model with constant variance. Parameter estimates are coloured by the log of the negative log-likelihood of the corresponding model fit. Yellow dashed lines correspond to parameter values reported in [6]. The majority of initial values resulted in estimates close to the original publication for the logit model (bottom). Convergence to local optima far from the published parameters was more common in the cloglog link model (top).
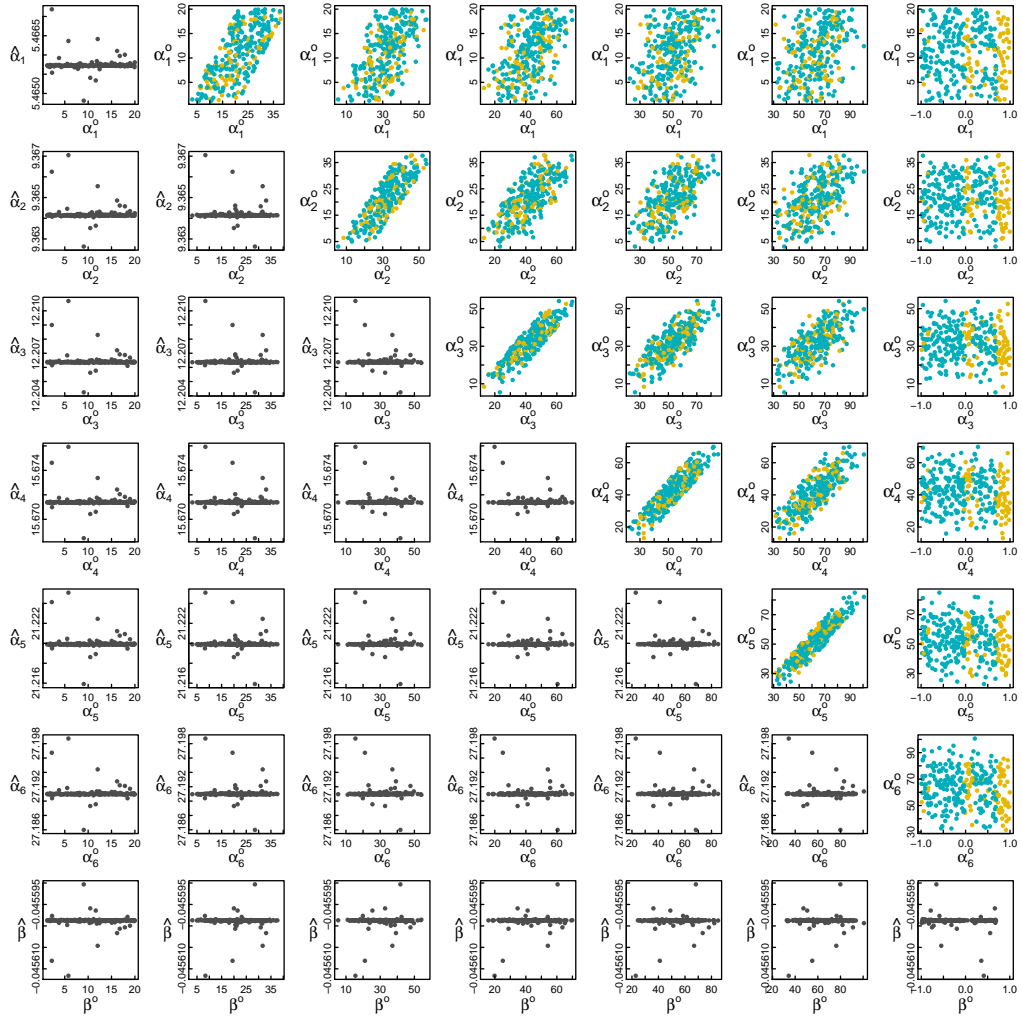
Figure S2. Sensitivity of optimisation results to initial values for the logit-link cumulative model with constant variance. The lower triangle and diagonal elements of the scatterplot matrix (grey symbols) show pairwise plots of filtered parameter estimates $\hat{\alpha}_i, \hat{\beta}$ against corresponding initial values $\alpha_i^\circ, \beta^\circ$. The upper triangle shows pairwise plots of the initial values for the same model parameters against each other. Green dots indicate successful convergence of the optimisation, yellow dots indicate convergence failures. Optimisation failed whenever $\beta^\circ > 0.7$, furthermore convergence failures were more likely when $\beta^\circ \approx 0$.
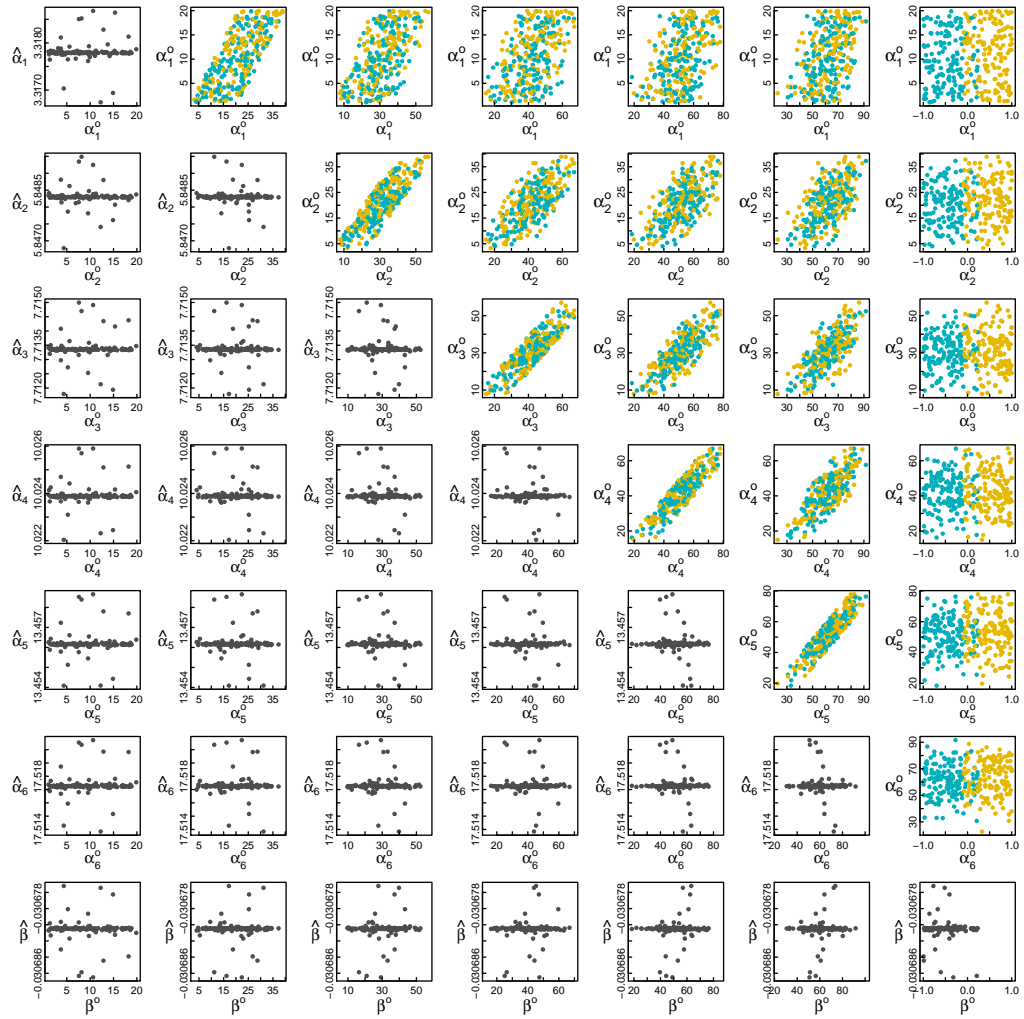
Figure S3. Sensitivity of optimisation results to initial values for the cloglog-link cumulative model with constant variance. The lower triangle and diagonal elements of the scatterplot matrix (grey symbols) show pairwise plots of filtered parameter estimates $\hat{\alpha}_i, \hat{\beta}$ against corresponding initial values $\alpha_i^\circ, \beta^\circ$. The upper triangle shows pairwise plots of the initial values for the same model parameters against each other. Green dots indicate successful convergence of the optimisation, yellow dots indicate convergence failures. Optimisation failed for most $\beta^\circ > 0$.