

# Class 8: Breast Cancer Mini Project

Paul Brencick (A17668863)

## Table of contents

Background . . . . .	1
Importing Our Data . . . . .	1
Performing PCA! . . . . .	3
<b>Interpreting PCA results</b>	<b>5</b>
Explaining our Variance with a screen plot . . . . .	6
Screen plot . . . . .	6
<b>4. Hierarchal Clustering</b>	<b>8</b>
Combining Method . . . . .	10
Prediction . . . . .	12

## Background

In today's class we will be employing all the R techniques for data analysis that we have learned thus far - including machine learning methods of clustering and PCA - to analyze real breast cancer biopsy data.

## Importing Our Data

Before we start analyzing we need to make sure the data is uploaded into the project so we can access it.

```
fna.data <- "WisconsinCancer.csv"
wisc.df <- read.csv(fna.data, row.names=1)
head(wisc.df, 4)
```

	diagnosis	radius_mean	texture_mean	perimeter_mean	area_mean
842302	M	17.99	10.38	122.80	1001.0
842517	M	20.57	17.77	132.90	1326.0
84300903	M	19.69	21.25	130.00	1203.0
84348301	M	11.42	20.38	77.58	386.1
	smoothness_mean	compactness_mean	concavity_mean	concave.points_mean	
842302	0.11840	0.27760	0.3001		0.14710
842517	0.08474	0.07864	0.0869		0.07017
84300903	0.10960	0.15990	0.1974		0.12790
84348301	0.14250	0.28390	0.2414		0.10520
	symmetry_mean	fractal_dimension_mean	radius_se	texture_se	perimeter_se
842302	0.2419	0.07871	1.0950	0.9053	8.589
842517	0.1812	0.05667	0.5435	0.7339	3.398
84300903	0.2069	0.05999	0.7456	0.7869	4.585
84348301	0.2597	0.09744	0.4956	1.1560	3.445
	area_se	smoothness_se	compactness_se	concavity_se	concave.points_se
842302	153.40	0.006399	0.04904	0.05373	0.01587
842517	74.08	0.005225	0.01308	0.01860	0.01340
84300903	94.03	0.006150	0.04006	0.03832	0.02058
84348301	27.23	0.009110	0.07458	0.05661	0.01867
	symmetry_se	fractal_dimension_se	radius_worst	texture_worst	
842302	0.03003	0.006193	25.38	17.33	
842517	0.01389	0.003532	24.99	23.41	
84300903	0.02250	0.004571	23.57	25.53	
84348301	0.05963	0.009208	14.91	26.50	
	perimeter_worst	area_worst	smoothness_worst	compactness_worst	
842302	184.60	2019.0	0.1622	0.6656	
842517	158.80	1956.0	0.1238	0.1866	
84300903	152.50	1709.0	0.1444	0.4245	
84348301	98.87	567.7	0.2098	0.8663	
	concavity_worst	concave.points_worst	symmetry_worst		
842302	0.7119	0.2654	0.4601		
842517	0.2416	0.1860	0.2750		
84300903	0.4504	0.2430	0.3613		
84348301	0.6869	0.2575	0.6638		
	fractal_dimension_worst				
842302	0.11890				
842517	0.08902				
84300903	0.08758				
84348301	0.17300				

We don't want the diagnosis column in our data so we have to remove it and create a new

vectors `diagnosis` so we can access both if needed. We can compare our findings to the experts with it after we are done.

```
wisc.data <- wisc.df[,-1]
diagnosis <- wisc.df$diagnosis
```

Q1. How many observations are in this dataset?

```
nrow(wisc.data)
```

```
[1] 569
```

Q2. How many of the observations have a malignant diagnosis?

```
sum(diagnosis == "M")
```

```
[1] 212
```

Q3. How many variables/features in the data are suffixed with `_mean`?

```
length(grep("_mean", colnames(wisc.data)))
```

```
[1] 10
```

## Performing PCA!

The main function in base R is called `prcomp()` we will use the optional argument `scaling=T` here as the data columns/features/dimensions are on very different scales in the original data set.

```
wisc.pr <- prcomp(wisc.data, scale = T)
summary(wisc.pr)
```

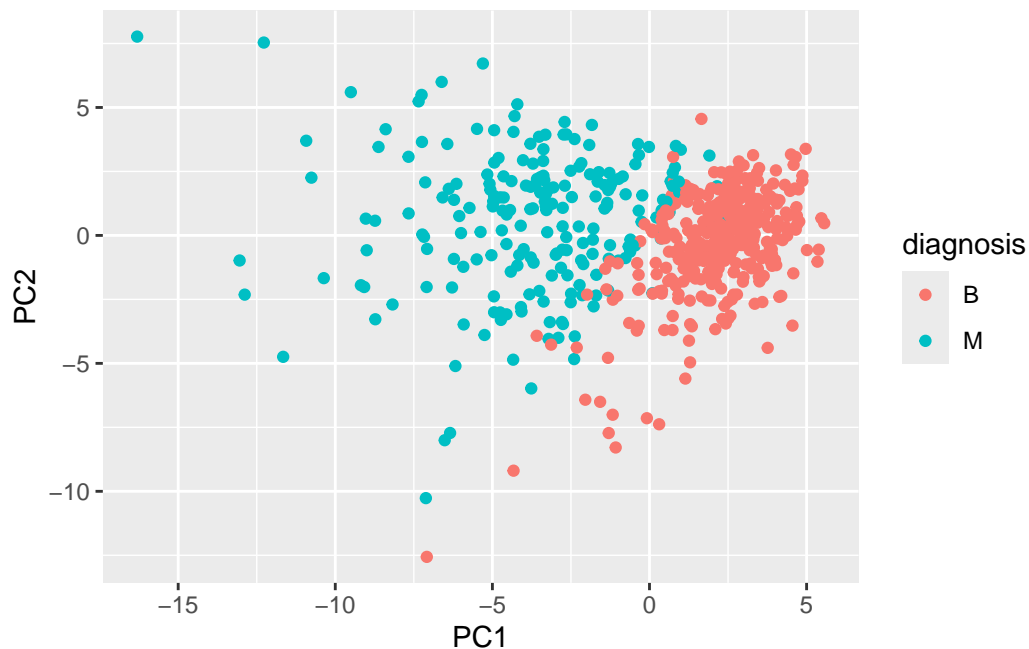
Importance of components:

	PC1	PC2	PC3	PC4	PC5	PC6	PC7
Standard deviation	3.6444	2.3857	1.67867	1.40735	1.28403	1.09880	0.82172
Proportion of Variance	0.4427	0.1897	0.09393	0.06602	0.05496	0.04025	0.02251
Cumulative Proportion	0.4427	0.6324	0.72636	0.79239	0.84734	0.88759	0.91010
	PC8	PC9	PC10	PC11	PC12	PC13	PC14
Standard deviation	0.69037	0.6457	0.59219	0.5421	0.51104	0.49128	0.39624

Proportion of Variance	0.01589	0.0139	0.01169	0.0098	0.00871	0.00805	0.00523
Cumulative Proportion	0.92598	0.9399	0.95157	0.9614	0.97007	0.97812	0.98335
	PC15	PC16	PC17	PC18	PC19	PC20	PC21
Standard deviation	0.30681	0.28260	0.24372	0.22939	0.22244	0.17652	0.1731
Proportion of Variance	0.00314	0.00266	0.00198	0.00175	0.00165	0.00104	0.0010
Cumulative Proportion	0.98649	0.98915	0.99113	0.99288	0.99453	0.99557	0.9966
	PC22	PC23	PC24	PC25	PC26	PC27	PC28
Standard deviation	0.16565	0.15602	0.1344	0.12442	0.09043	0.08307	0.03987
Proportion of Variance	0.00091	0.00081	0.0006	0.00052	0.00027	0.00023	0.00005
Cumulative Proportion	0.99749	0.99830	0.9989	0.99942	0.99969	0.99992	0.99997
	PC29	PC30					
Standard deviation	0.02736	0.01153					
Proportion of Variance	0.00002	0.00000					
Cumulative Proportion	1.00000	1.00000					

```
library(ggplot2)

ggplot(wisc.pr$x) +
  aes(PC1, PC2, col=diagnosis) +
  geom_point()
```



Q4. From your results, what proportion of the original variance is captured by the first principal component (PC1)?

0.44272 is captured within PC1

Q5. How many principal components (PCs) are required to describe at least 70% of the original variance in the data?

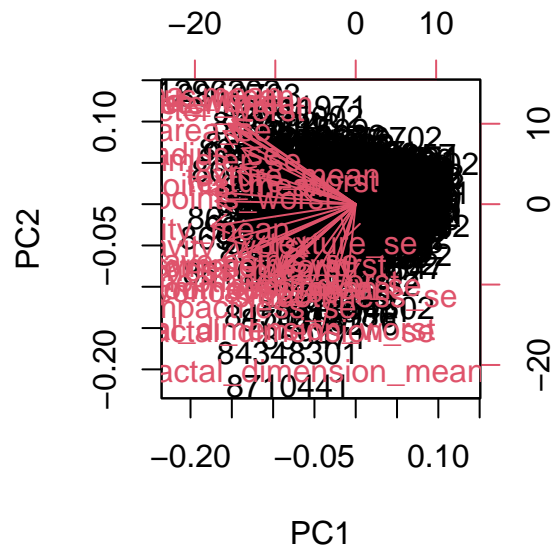
It took 3 PCs to describe at least 70% of the original variance in the data.

Q6. How many principal components (PCs) are required to describe at least 90% of the original variance in the data?

It takes 7 PCs to describe at least 90% of the original variance.

## Interpreting PCA results

```
biplot(wisc.pr)
```



Q7 What stands out to you about this plot? Is it easy or difficult to understand? Why?

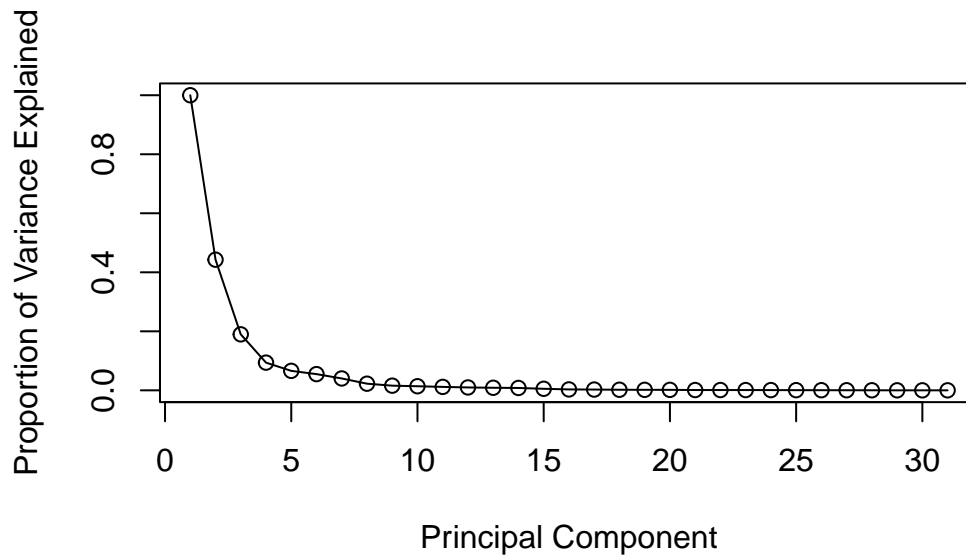
This plot is very confusing to understand and interpret as it is all overlapped. What stands out to me is the tight spread in the plot, all within  $\pm 0.20$ .

## Explaining our Variance with a screen plot

```
pr.var <- wisc.pr$sdev^2  
head(pr.var)
```

```
[1] 13.281608  5.691355  2.817949  1.980640  1.648731  1.207357
```

```
pve <- pr.var / sum(pr.var)  
  
plot(c(1,pve), xlab = "Principal Component",  
     ylab = "Proportion of Variance Explained",  
     ylim = c(0, 1), type = "o")
```



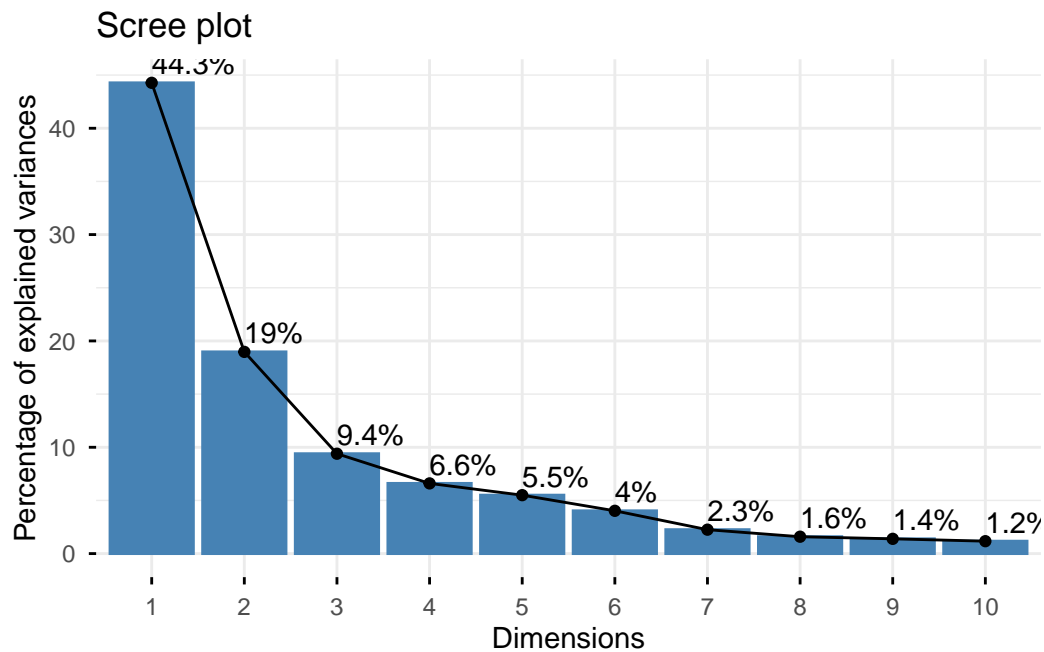
## Screen plot

```
library(factoextra)
```

Welcome! Want to learn more? See two factoextra-related books at <https://goo.gl/ve3WBa>

```
fviz_eig(wisc.pr, addlabels = TRUE)
```

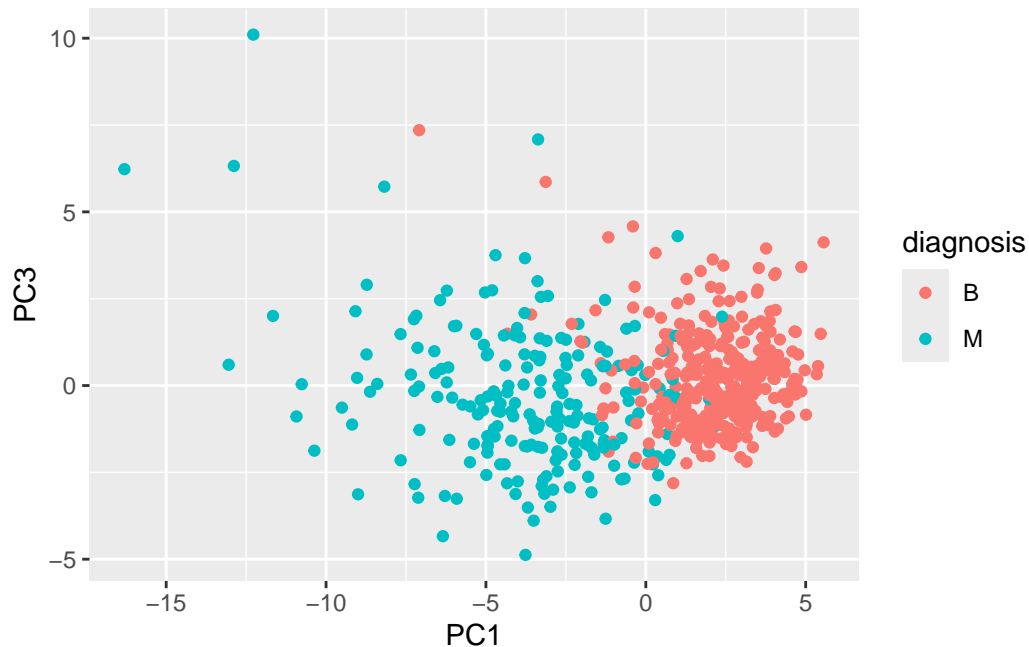
Warning in geom\_bar(stat = "identity", fill = barfill, color = barcolor, :  
Ignoring empty aesthetic: `width`.



Q8. Generate a similar plot for principal components 1 and 3. What do you notice about these plots?

```
library(ggplot2)

ggplot(wisc.pr$x) +
  aes(PC1, PC3, col=diagnosis) +
  geom_point()
```



I notice there is still a pretty strong divide between M and B and it is fairly clear where there is a difference

Q9. For the first principal component, what is the component of the loading vector (i.e. `wisc.pr$rotation[,1]`) for the feature `concave.points_mean`? This tells us how much this original feature contributes to the first PC. Are there any features with larger contributions than this one?

```
wisc.pr$rotation["concave.points_mean", "PC1"]
```

```
[1] -0.2608538
```

I could not find any feature with larger contributions.

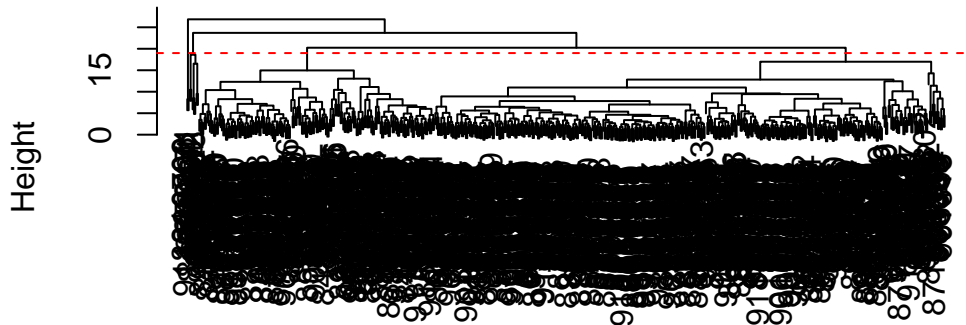
## 4. Hierarchical Clustering

The goal of this section is to do hierarchical clustering of the original data to see if there is any obvious grouping into malignant and benign clusters.

First we will scale our `wisc.data` then calculate a distance matrix then pass to `hclust()`

```
wisc.dist <- dist( scale(wisc.data))
wisc.hclust <- hclust(wisc.dist)
plot(wisc.hclust)
abline(h = 19, col = "red", lty = 2)
```

## Cluster Dendrogram



wisc.dist  
hclust (\*, "complete")

Q10. Using the plot() and abline() functions, what is the height at which the clustering model has 4 clusters?

~19

```
wisc.hclust.clusters <- cutree(wisc.hclust,k=2)
table(wisc.hclust.clusters)
```

```
wisc.hclust.clusters
 1  2
567 2
```

Q12. Which method gives your favorite results for the same data.dist dataset?  
Explain your reasoning.

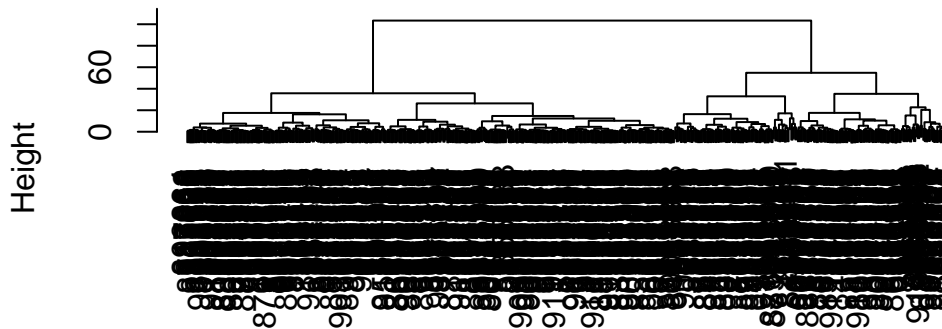
Ward.D2 hierarchical clustering on the PCA-transformed data, the separation between diagnoses is much better than when clustering the original variables.

## Combining Method

The idea here is that I can take my new variables (i.e. the scores PCs `wisc.pr$x`) that are better descriptors of the data-set than the original features (i.e. the 30 columns in `wisc.data`) and use these as a basis for clustering.

```
pc.dist<- dist(wisc.pr$x[,1:3])  
  
wisc.pr.hclust <- hclust(pc.dist, method = "ward.D2")  
  
plot(wisc.pr.hclust)
```

### Cluster Dendrogram



```
grps <- cutree(wisc.pr.hclust, k=2)  
table(grps)
```

```
grps  
  1  2  
203 366
```

```
table(diagnosis)
```

```
diagnosis
  B   M
357 212
```

Q13. How well does the newly created hclust model with two clusters separate out the two “M” and “B” diagnoses?

I can now run `table()` with both my clustering `grps` and the expert diagnoses

```
table(grps, diagnosis)
```

```
      diagnosis
grps   B    M
1    24 179
2   333  33
```

Our cluster “1” has 179 “M” diagnosis

However, cluster “2” has 333 “B” diagnosis

Group 1: 179 = True positive 24 = False positives

Group 2: 333 = True negative 33 = False negative

Sensitivity:  $TP/(TP+FN)$

```
179/(179+33)
```

```
[1] 0.8443396
```

Specificity:  $TN/(TN+FP)$

```
333/(333+24)
```

```
[1] 0.9327731
```

Q14. How well do the hierarchical clustering models you created in the previous sections (i.e. without first doing PCA) do in terms of separating the diagnoses? Again, use the `table()` function to compare the output of each model (`wisc.hclust.clusters` and `wisc.pr.hclust.clusters`) with the vector containing the actual diagnoses.

```
table(wisc.hclust.clusters, diagnosis)
```

```

              diagnosis
wisc.hclust.clusters  B   M
1 357 210
2   0   2

```

This means the clustering largely fails to distinguish between benign and malignant tumors

## Prediction

We can use our PCA model for prediction of new un-seen cases!

```

url <- "https://tinyurl.com/new-samples-CSV"
new <- read.csv(url)
npc <- predict(wisc.pr, newdata=new)
npc

```

```

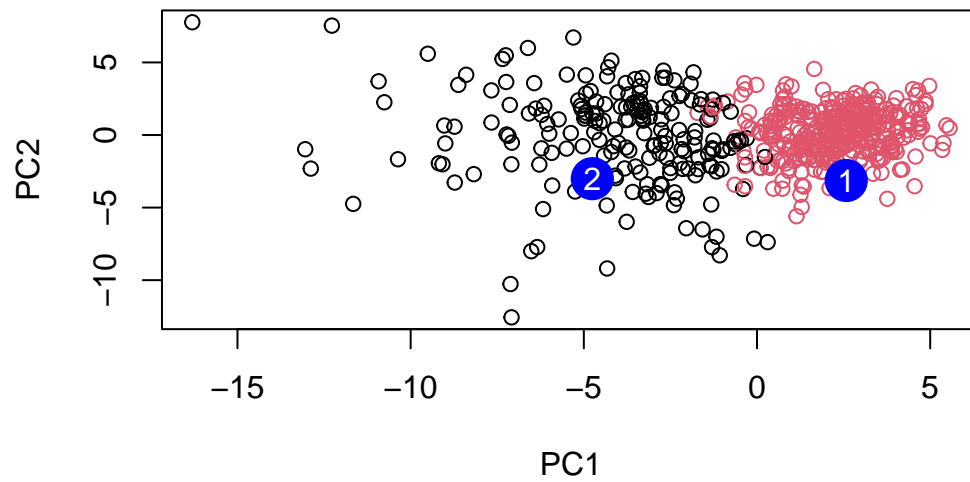
      PC1      PC2      PC3      PC4      PC5      PC6      PC7
[1,]  2.576616 -3.135913  1.3990492 -0.7631950  2.781648 -0.8150185 -0.3959098
[2,] -4.754928 -3.009033 -0.1660946 -0.6052952 -1.140698 -1.2189945  0.8193031
      PC8      PC9      PC10     PC11     PC12     PC13     PC14
[1,] -0.2307350 0.1029569 -0.9272861 0.3411457  0.375921 0.1610764 1.187882
[2,] -0.3307423 0.5281896 -0.4855301 0.7173233 -1.185917 0.5893856 0.303029
      PC15     PC16     PC17     PC18     PC19     PC20
[1,] 0.3216974 -0.1743616 -0.07875393 -0.11207028 -0.08802955 -0.2495216
[2,] 0.1299153  0.1448061 -0.40509706  0.06565549  0.25591230 -0.4289500
      PC21     PC22     PC23     PC24     PC25     PC26
[1,] 0.1228233 0.09358453 0.08347651 0.1223396 0.02124121 0.078884581
[2,] -0.1224776 0.01732146 0.06316631 -0.2338618 -0.20755948 -0.009833238
      PC27     PC28     PC29     PC30
[1,] 0.220199544 -0.02946023 -0.015620933 0.005269029
[2,] -0.001134152 0.09638361 0.002795349 -0.019015820

```

```

plot(wisc.pr$x[,1:2], col=grps)
points(npc[,1], npc[,2], col="blue", pch=16, cex=3)
text(npc[,1], npc[,2], c(1,2), col="white")

```



Q16. Which of these new patients should we prioritize for follow up based on your results?

Patient 2 is malignant and 1 is benign so we would want to follow up with patient 2 most likely first.