Strain-space model for Sars-CoV-2

Peter C. Jentsch, PhD ^{1,4} Finlay Maguire, PhD ^{3,5} Samira Mubareka, MD, FRCPC ^{1,2}

¹Sunnybrook Research Institute, Toronto, Canada

²University of Toronto, Toronto, Canada

³Dalhousie University, Halifax, Canada

⁴Simon Fraser University, Burnaby, Canada

⁵Shared Hospital Laboratory, Toronto, Canada

June 14, 2022

- Dynamic model of Sars-CoV-2 evolution, representing antigenic diversity on a lattice (as in e.g. [Gog and Grenfell, 2002, Kryazhimskiy et al., 2007, Marchi et al., 2021])
- Antigenically distinct variants of the virus are mapped to 2D grid, distance between variants corresponds to the proportional reduction in maximum serum viral titre [Wilks et al., 2022, van der Straten et al., 2022]

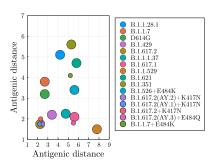


Figure: Antigenic cartography of Sars-CoV-2, reproduced from [Wilks et al., 2022], Fig. 2

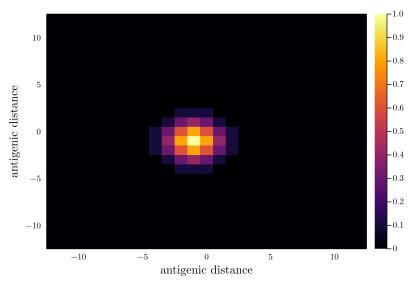
Model parameters/variables

| Symbol | Description |
|-----------------|---|
| N | Size of variant grid |
| S_{ij} | Population susceptible to variant $(i, j) \in [0, N]^2$ |
| I_{ij} | Population infected by variant $(i, j) \in [0, N]^2$ |
| R_{ij} | Recovered/Immune to variant $(i, j) \in [0, N]^2$ |
| σ_{ijkl} | Probability that exposure to variant (i, j) causes |
| | immunity |
| | to variant (k, l) |
| eta_{ij} | Transmission rate of variant (i, j) |
| ξ | Recovery rate of all strains |
| γ | Rate of immunity loss of all strains |

Table: Table of symbols for Model 2

σ matrix

In practice, we assume σ_{ijkl} is just a 2-D gaussian distribution parameterized by the distance between (i, j) and (k, l).



Model Equations

$$\frac{S_{ij}}{dt} = -\sum_{kl} \beta_{kl} \sigma_{ijkl} S_{ij} I_{kl} + \gamma R_{ij} \tag{1}$$

$$\frac{I_{ij}(t)}{dt} = \beta_{ij} S_{ij} I_{ij} - \xi I_{ij} + M \left(-4I_{ij} + I_{i-1,j} + I_{i+1,j} + I_{i,j-1} + I_{i,j+1} \right)$$
(2)

$$\frac{R_{ij}(t)}{dt} = \xi I_{ij} - \gamma R_{ij} \tag{3}$$

Boundary conditions: $I_{0,j} = 0, I_{j,0} = 0, I_{N,j} = 0, I_{j,N} = 0$ Initial conditions computed from genomic data in GISAID To incorporate more realistic mutation rates, we can go to continuous strain-space and use nonlocal reaction-diffusion dynamics as in [Rouzine and Rozhnova, 2018, Bessonov et al., 2021]

$$S_{t}(x, y, t) = \int_{-\infty}^{\infty} \int_{-\infty}^{\infty} \beta(x', y') \sigma(x, y, x', y') S(x, y, t) I(x', y', t) dx' dy' + \gamma R_{ij} - \eta(t) v(x, y) S(x, y, t)$$

$$I_{t}(x, y, t) = \beta(x, y)S(x, y, t)I(x, y, t) - \xi I(x, y, t) + M\left(I_{x}(x, y, t) + I_{y}(x, y, t)\right)$$
(5)

$$R_t(x, y, t) = \xi I(x, y, t)I(x, y, t) - \gamma R(x, y, t) + \eta(t)v(x, y)S(x, y, t)$$
(6)

where β, σ, v have been generalized to their continuous counterparts. Given a dispersion kernel $K(x,y) \in L_2 : \mathbb{R}^2 \to \mathbb{R}$ this can be generalised to non-local diffusion as follows

$$I_{t}(x, y, t) = \beta(x, y)S(x, y, t)I(x, y, t) - \xi I(x, y, t) + M\left(\int_{-\infty}^{\infty} \int_{-\infty}^{\infty} K(x - x', y - y')I(x', y', t)dx'dy'\right)$$
(7)

Developing an antigenic distance map

- We would like an approximate measure of antigenic distance for every sample genome
- Using all samples, we compute pairwise distances between each unique genome in some way that encodes antigenic response
- Many possible ways to do this, so far none of them seem to work very well
- Project to 2-d (hopefully) space with multidimensional scaling

Genome distance

Assume:

- a, b are SARS-CoV-2 genomes aligned with the reference
- \bullet a_i the *i*th nucleotide base in a and

$$\chi(a_i, b_i) = \begin{cases} 1 & \text{if } a_i = b_i \\ 0 & \text{otherwise} \end{cases}$$

- \bullet h_i is a vector containing the number of homoplasic mutations at site i in the global tree
- **\mathfrak{B}(a)** computes the polyclonal binding affinity of genome a as per [Starr et al., 2020]

One option for a distance measure is something like

$$d(a,b) = \frac{\mathfrak{B}(a) + \mathfrak{B}(b)}{2} + \sum_{i} \chi(a_i, b_i) h_i \tag{8}$$

That is, the average binding between two genomes plus the SNP distance weighted by the relative homoplasy of each mutation.

Example antigenic distance map

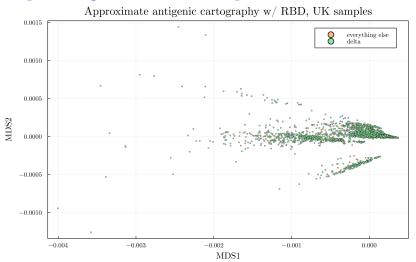


Figure: Multidimensional scaling plot using samples from the UK up to mid November

Homoplasy in global tree

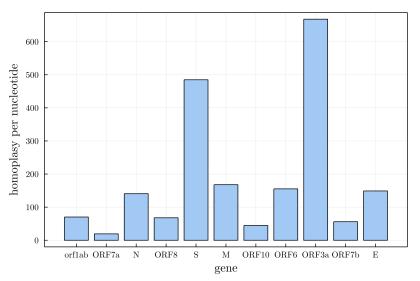
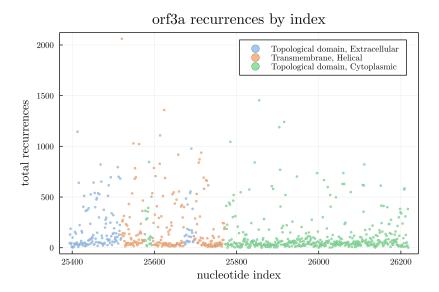


Figure: Number of recurrent (homoplasic) mutations per base by gene, (normalized by gene length)

Homoplasy in orf3a



Bessonov, N., Bocharov, G., Meyerhans, A., Popov, V., and Volpert, V. (2021).

Existence and dynamics of strains in a nonlocal reaction-diffusion model of viral evolution. SIAM Journal on Applied Mathematics, 81(1):107–128.

Gog, J. R. and Grenfell, B. T. (2002).

Dynamics and selection of many-strain pathogens.

Proceedings of the National Academy of Sciences,
99(26):17209–17214.

Kryazhimskiy, S., Dieckmann, U., Levin, S. A., and Dushoff, J. (2007).

On State-Space Reduction in Multi-Strain Pathogen Models, with an Application to Antigenic Drift in Influenza A.

PLOS Computational Biology, 3(8):e159.

Marchi, J., Lässig, M., Walczak, A. M., and Mora, T. (2021).

Antigenic waves of virus-immune coevolution. *Proceedings of the National Academy of Sciences*, 118(27):e2103398118.

Rouzine, I. M. and Rozhnova, G. (2018). Antigenic evolution of viruses in host populations. *PLoS Pathogens*, 14(9):e1007291.

Starr, T. N., Greaney, A. J., Hilton, S. K., Ellis, D., Crawford, K. H., Dingens, A. S., Navarro, M. J., Bowen, J. E., Tortorici, M. A., Walls, A. C., et al. (2020). Deep mutational scanning of sars-cov-2 receptor binding domain reveals constraints on folding and ace2 binding. Cell, 182(5):1295–1310.

van der Straten, K., Guerra, D., van Gils, M., Bontjer, I., Caniels, T. G., van Willigen, H. D., Wynberg, E., Poniman, M., Burger, J. A., Bouhuijs, J. H., et al. (2022). Mapping the antigenic diversification of sars-cov-2.

medRxiv.

Wilks, S. H., Mühlemann, B., Shen, X., Türeli, S., LeGresley, E. B., Netzl, A., Caniza, M. A., Chacaltana-Huarcaya, J. N., Daniell, X., Datto, M. B., Denny, T. N., Drosten, C., Fouchier, R. A. M., Garcia, P. J., Halfmann, P. J., Jassem, A., Jones, T. C., Kawaoka, Y., Krammer, F., McDanal, C., Pajon, R., Simon, V., Stockwell, M., Tang, H., van Bakel, H., Webby, R., Montefiori, D. C., and Smith, D. J. (2022). Mapping SARS-CoV-2 antigenic relationships and serological responses. Preprint, Immunology.