

Mechanistic Modeling as an Explanatory Tool for Clinical Treatment of Chronic Catatonia

Patrick D Roberts^{1,*} and James Conour²

¹Amazon Web Services, Portland, OR, USA

²Cascadia Behavioral Healthcare, Portland, OR, USA

Correspondence*:

Patrick D Roberts

rbertsp@amazon.com

2 ABSTRACT

3 Mathematical modeling of neural systems is an effective means to integrate complex information
4 about the brain into a numerical tool that can help explain observations. However, the use of neural
5 models to inform clinical decisions has been limited. In this study, we use a simple model of brain
6 circuitry, the Wilson-Cowan model, to predict changes in a clinical measure for catatonia, the Bush-
7 Francis Catatonia Rating Scale, for use in clinical treatment of schizophrenia. This computational
8 tool can then be used to better understand mechanisms of action for pharmaceutical treatments,
9 and to fine-tune dosage in individual cases. We present the conditions of clinical care for a
10 residential patient cohort, and describe methods for synthesizing data to demonstrated the
11 functioning of the model. We then show that the model can be used to explain effect sizes
12 of treatments and estimate outcomes for combinations of medications. We conclude with a
13 demonstration of how this model could be personalized for individual patients to inform ongoing
14 treatment protocols.

15 **Keywords:** Schizophrenia, Bush-Francis, antipsychotic, benzodiazepine, lamotrigine, Wilson-Cowan

1 INTRODUCTION

16 The treatment of severe and persistent mental illness has been a central challenge for psychiatry.
17 Individuals with the most debilitating forms of schizophrenia often derive limited benefit from medications.
18 Additionally, the efficacy of pharmacologic treatments can be highly variable. A full response to a medical
19 intervention may take weeks or months to materialize. Moreover, it can be difficult to accurately assess
20 the impact of a specific medication. These challenges are compounded by the inconsistent history of care
21 for many psychiatric patients and the significant amounts of polypharmacy they have been prescribed.
22 Technical tools offer a promising augmentation to a psychiatrist's experience to design treatment plans and
23 may help reduce the inconsistencies and refine treatment for individual cases.

24 Catatonia manifests as a cluster of symptoms including rituals, repetitive movements, perseveration,
25 and withdrawal [Northoff (2002)]. There is common co-morbidity with both psychiatric and medical
26 illnesses [Bhati et al. (2007)] and catatonia is often not recognized in its chronic form because it can present
27 subtly and idiosyncratically [Penland et al. (2006)]. In individuals with treatment resistant schizophrenia,
28 chronic catatonic may be quite common, and direct treatment of catatonic symptoms improves cognition
29 [Wilcox and Reid Duffy (2015); Ungvari et al. (2005)]. For this reason, we have focussed on using the

Bush-Francis Catatonia Rating Scale (BFCRS) [Bush et al. (1996)] as a measure of symptoms and then model pharmacological mechanisms that explain how medications alleviate catatonic symptoms.

The data in this study is based on a cohort of schizophrenia patients admitted to Cascadia Behavioral Healthcare for residential care. The clinical practice in treating these patients has been to introduce a minimal set of medications with a known effect of reducing psychiatric symptoms. For patients admitted with a diagnosis of schizophrenia, antipsychotic medication was transitioned to clozapine (if possible), and augmented lamotrigine and a benzodiazepine based on functional status and safety. Lamotrigine has been previously observed to reduce symptoms in combination with clozapine [Gray and Risch (2009); Tiihonen et al. (2003)]. Benzodiazepines have shown a strong therapeutic efficacy in reducing catatonia symptoms [Rosebush and Mazurek (2010); Northoff et al. (2004)] and are considered a first-line treatment for acute or chronic catatonia [Ungvari et al. (2005)]. A significant reduction in catatonic symptoms, as measured by BFCRS, was observed in the clinic with this treatment along with a corresponding improvement in psychiatric symptoms. However, a mechanistic understanding of the action of this combination is desirable to improve treatments and seek new strategies for psychiatric disease maintenance.

1.1 Modeling as an explanatory tool

Physiological modeling of pharmacological systems can provide insight into mechanisms of therapeutic treatments by coupling molecular action to observable function. Explanatory models require a balance between biological detail and conceptual simplicity to express how specific treatments result in observed functional changes. The psychomotor abnormalities observed in catatonia can be conceptualized as a seizing of motor patterns on a time scale long enough to result in the clinical observations such as posturing and repetitive movements. Clinical and imaging studies have suggested that the physiological basis of catatonia symptoms are cortical in origin [Northoff (2002); Hirjak et al. (2019)] resulting from an over-excitation of circuitry and under-gating of movement termination. The effective treatments also support the concept of an imbalance of inhibition and excitation in cortical structures because targets of lamotrigine reduce pyramidal cell excitation [Poolos et al. (2002); Xie et al. (1995)], and benzodiazepines increase inhibition [Miller et al. (1987)].

A neural model describing interactions of excitatory and inhibitory neurons, with sufficient structure to couple medication actions, is the Wilson-Cowan model [Wilson and Cowan (1972)]. This model is interpreted as two interacting populations of cortical neurons where a single variable for each population represents the average spike rate (Figure 1a). The Wilson-Cowan model is mathematically well-understood [Cowan et al. (2016); Bressloff (2010); Benayoun et al. (2010); Buice et al. (2010); Negahbani et al. (2015)] with dynamics that can display excitatory bursts and oscillations for different choices of parameters. For the purposes of the current study, we select a parameter range so that the dynamics represent two steady states of spiking activity, a high-rate and low-rate, in two basins separated by a barrier. The height of the barrier is determined by the parameters of the model and determines the perturbation required to transition from the high-rate state to the low-rate state. The transition from the high-rate state to the low-rate state represents the termination of a cortical activity pattern. If the barrier is high then the system becomes “stuck” in a functional pattern and is interpreted to represent symptoms of catatonia such as posturing or perseveration. Parameters of the model determine the synaptic coupling between populations of neurons and internal neural excitability, and these parameters are affected by medications.

In our model, we start with baseline parameter settings with a high barrier to represent catatonia, then calculate the changes in parameters based on the doses of medications in the clinical treatment. We show that the change in the barrier can be correlated with the change in BFCRS score to explain how each

medication is impacting symptoms of catatonia. By using the model as a clinical guide to treatment, the clinician can conceptualize the physiological effects of a treatment as controlling cortical excitability to treat catatonia. This allows guidance beyond the safety and efficacy of individual medications to integrate polypharmacy into utilizing additive effects maximize positive outcomes.

2 MATERIALS AND METHODS

2.1 Data synthesis

Clinical data on BFCRS scores and daily medication dosages were collected and analyzed for clinical treatment purposes. For demonstration purposes, we synthesized surrogate data based on the statistics of the original data set (Conour (2015)). Using the *SVD* [Patki et al. (2016)] python package, we constructed a Gaussian copula model based on the daily dosages of medications, and BFCRS scores before and after changes in medication for 12 individuals. The statistical reconstruction method ensured that no personal patient data is present in the published study. The copula model generated many spurious data samples with unrealistic medication doses because there were few individuals included in training the model. To eliminate spurious data, we added rules determined by JC to be unlikely under clinical conditions (see *Data Selection Filter*, Supplementary Data). The copula model generated 700 subjects and 58 subjects remained after filtering.

A summary of the synthesized dataset is shown in Figure 2. The mean BFCRS score before treatment is 17.3 ± 3.9 (std) and after treatment is 4.1 ± 2.8 , resulting in an effect size of 2.7 for the treatment. The distribution of doses for medications upon admission (pre-treatment) and following stabilization of the treatment (post-treatment) are shown in Figure 2b and 2c. These statistics of the medication combinations and dose ranges are consistent with the clinical patient data set.

2.2 Wilson-Cowan model

The pharmaceutical treatments include 3 classes of medications: anticonvulsants, benzodiazepines, and antipsychotics. These medications operate via multiple mechanisms of action, and our approach couples their action to a model of cortical activity. In order to quantify the effects, we developed a two-state model of cortical dynamics that can predict how varying doses affect catatonic symptoms. The Wilson-Cowan equations are (1):

$$\begin{aligned}\dot{x}_0 &= -x_0 + F_0(w_{00}x_0 + w_{01}x_1) \\ \dot{x}_1 &= -x_1 + F_1(w_{10}x_0 + w_{11}x_1)\end{aligned}\quad (1)$$

with the spike probability (rate) function:

$$F_a(x_a) = \frac{1}{1 + \exp[-\mu_a(x_a - \theta_a)]}\quad (2)$$

We interpret x_0 as the average rate of inhibitory interneurons (parvalbumin positive) and x_1 as the average rate of excitatory neurons (cortical pyramidal cells). The parameters of the model were initialized to express 3 fixed points, one stable fixed point representing a low spike rate, one stable fixed point representing a high spike rate, and a saddle point that is the barrier between the two states. The initial synaptic parameters: $w_{11} = 8.65$, $w_{10} = 4$, $w_{01} = 13$, and $w_{00} = 9$. The parameters for the rate function are $\mu_1 = 1.2$, $\theta_1 = 2.8$, $\mu_0 = 1.0$, and $\theta_0 = 4.0$.

The barrier is calculated by a cumulative summation of the excitatory rate gradient along the I-nullcline (Figure 1b) from the high-rate fixed point to the barrier fixed point. This is to represent the minimal perturbation necessary to transition out of the high-rate state basin. The high-rate state is unstable under perturbations and when noise is added, the system will spontaneously transition to the low-rate state (Figure 1c). The duration of the time in the high-rate state can be interpreted as a form of working memory [Katori et al. (2011)], but here we consider the duration as a phase of activity [Bagi et al. (2022)] that can lead to perseveration when the barrier is too high and a large perturbation is required for a state transition. Medications act on parameters of the model to raise or lower the boundary and affect catatonia symptoms.

2.3 Coupling treatment doses to model parameters

Clinical doses were converted to changes in the model parameters through a series of calculations. First we approximated the pharmacokinetics of each medication (see *Pharmacokinetic Parameters*, Supplementary Data) to arrive at a concentration in the cerebrospinal fluid (CSF). Next we calculate the binding to a target, and finally approximate an effect on the model parameters [Spiros et al. (2010); Geerts et al. (2013)]. The following provides details of the pharmacokinetics and coupling for lamotrigine, lorazepam (and applies to other benzodiazepines according to their affinities), and antipsychotics.

Pharmacokinetics

To compute the average CSF concentration, C_{ave} , we apply the following function to the clinical daily dose for the synthesized data:

$$C_{ave} = K_{csf} \frac{d}{T_d} \frac{AUC}{AUC_d} \frac{10^3}{M_{wt}} \quad (3)$$

where K_{csf} is the brain/blood transport ratio, d is the dose (mg), T_d is the dose interval (hr), AUC is the area under the curve of the plasma concentration (mg/hr), AUC_d is the dose (mg) when the AUC was measured, and M_{wt} is the molecular weight to convert (g/mol) to (nM).

Lamotrigine

Na^+ -current: Lamotrigine reduces Na^+ -current in pyramidal cells Xie et al. (1995),

$$\Delta I_{Na} = I/I_{max} = 1 - \frac{C_{LTG}}{(C_{LTG} + K_C)^n} \quad (4)$$

where $K_C = 513$ uM, $n = 0.9$. To affect the rate, we will shift θ_1 by the mechanism of action, $M_{Na} = 1 - p_{Lam}(1 - \Delta I_{Na})$, where $p_{Lam} = 0.15$. The reduction in the Na current increases the threshold in excitatory neurons by, $\theta'_1 = \theta_1/M_{Na}$.

I_h -current: Lamotrigine shifts the I-V activation curve of the I_h current Poolos et al. (2002) and decreased evoked firing rate, $\Delta x_1 = 1 - 0.004 * C_{LTG}$, for $\Delta x_1 > 0$ and where C_{LTG} is the average concentration of lamotrigine in CSF. To represent this effect in our model parameters, we shift the threshold, θ_1 , in pyramidal cells. The shift is based on the spike probability function linearized near threshold $F_1(x_1) = 1/2 + (\mu_1/4)(x_1 - \theta_1)$, so that θ_1 will be shifted by the mechanism of action, $M_h = 1 - p_{Lam}(1 - \Delta x_1)$, where $p_{Lam} = 0.15$. The reduction in the Na current increases the threshold in excitatory neurons by, $\theta'_1 = \theta_1/M_h$.

Glutamate release: Lamotrigine reduces glutamate release from excitatory synapses Wang et al. (2001), $\Delta G = 1 - 0.004 * C_{LTG}$ for $\Delta G > 0$. The excitatory synaptic parameters, w_{11} and w_{10} , are affected by

the mechanism of action, $M_{glu} = 1 - p_{Lam}(1 - \Delta G)$, where $p_{Lam} = 0.15$. The reduction in the Glutamate release decreases the excitatory synaptic parameters by, $w'_{11} = w_{11}M_{glu}$ and $w'_{10} = w_{10}M_{glu}$.

Benzodiazepines

Benzodiazepines such as lorazepam increase GABA_A currents following binding at the BZD receptor site. The receptor occupation is calculated by Miller et al. (1987):

$$R_{oc} = \frac{(C_{Lor})^A}{(C_{Lor})^A + B} \quad (5)$$

where $A = 1.4328$, $B = 73.89$ (ng/gm), and C_{Lor} is the average concentration of lorazepam in CSF. The maximal effect is about doubles the GABA_A conductance. The inhibitory synaptic parameters, w_{01} and w_{00} , are affected by the mechanism of action, $M'_{gaba} = M_{gaba} + R_{oc}(1 - M_{gaba})$, to increase the inhibitory synaptic parameters by, $\Delta w'_{11} = w_{11}(1 + M_{gaba})$ and $w'_{10} = w_{10}(1 + M_{gaba})$.

Antipsychotics

These medications bind competitively with endogenous neurotransmitters to specific receptors. We use an exact form of the competitive binding formula Wang (1995):

$$\begin{aligned} a &= K_A + K_B + C_A + C_B - 1 \\ b &= K_B(C_A - 1) + K_A(C_B - 1) + K_A K_B \\ c &= -K_A K_B \\ \theta &= \arccos \left(\frac{-2a^3 + 9ab - 27c}{2\sqrt{(a^2 - 3b)^3}} \right) \\ R_{oc} &= C_A \frac{2\sqrt{a^2 - 3b} \cos(\theta/3) - a}{3K_A + (2\sqrt{a^2 - 3b} \cos(\theta/3) - a)} \end{aligned} \quad (6)$$

where K_A is the binding affinity of the endogenous neurotransmitter, C_A is the average concentration of the endogenous neurotransmitter, K_B is the binding affinity of the medication, and C_B is the average concentration of the medication. R_{oc} is the receptor occupation by the endogenous neurotransmitter and will be used to estimate the activation level of the receptor. In this study, endogenous levels of neurotransmitters were dopamine (tonic) = 37 mM, dopamine (burst) = 200 mM, serotonin = 3.9mM, and acetylcholine = 10mM (Dreyer et al. (2010); Paterson et al. (2010)).

D1 activation effect: The endogenous concentration at dopamine synapses depend on the firing patterning so that simulations estimate Dreyer et al. (2010) that tonic activity yields 37 ± 1.2 nM and bursts yield 100 - 300 nM. According to data in Lapish et al. (2007), D1 activation decreases the slope parameter (μ_1) of the rate function in excitatory neurons, $\mu'_1 = \mu_1(1 - (R_{oc} - R_{con})/R_{con})$, where R_{con} is the control level. D1 activation decreases the threshold (θ_0) in inhibitory interneurons, $\theta'_0 = \theta_0(1 - (R_{oc} - R_{con})/R_{con})$. D1 activation increases w_{11} , and w_{10} because at low concentrations (<50 uM) by acting preferentially on D1-like receptors to increase NMDA receptor-mediated transmission Lee et al. (2002), and increases w_{01} , that we represent by $w'_{ab} = w'_{ab}(1 + (R_{oc} - R_{con})/R_{con})$ where $(a, b) = (1, 1)$, $(1, 0)$, and $(0, 1)$.

D2 activation effect: At high concentrations (≥ 100 uM) DA activates D2-like receptors and suppress NMDA function Kotecha et al. (2002), that we represent by decreasing w_{11} and w_{10} , that we represent by $w'_{ab} = w'_{ab}(1 - (R_{oc} - R_{con})/R_{con})$ where $(a, b) = (1, 1)$ and $(1, 0)$. D2 also Increases the slope

parameter (μ_1) of probability function in excitatory neurons (pyramidal cells Lapish et al. (2007)), $\mu'_1 = \mu_1(1 + (R_{oc} - R_{con})/R_{con})$.

5-HT1A activation effect: Increases the threshold (θ_1) in excitatory neurons (pyramidal cells, [Foehring (1996)]), $\theta'_1 = \theta_1(1 + (R_{oc} - R_{con})/R_{con})$.

5-HT2A activation effect: Decreases the threshold (θ_1) in excitatory neurons (pyramidal cells, [Carr et al. (2002)]), $\theta'_1 = \theta_1(1 - (R_{oc} - R_{con})/R_{con})$.

M1 activation effect: Decreases the threshold (θ_1) in excitatory neurons (pyramidal cells) [Perez-Rosello et al. (2005)], $\theta'_1 = \theta_1(1 - (R_{oc} - R_{con})/R_{con})$.

When the changes in model parameters are calculated, we multiply by an overall factor of 0.35 to limit the dynamics of the system to maintain 2 stable fixed points separated by an unstable barrier fixed-point and ensure that the ground state of the system is the low-rate fixed point for all cases. For the personalization demonstration, we replaced this single parameter with a vector to individually tune the response to medications for each subject.

3 RESULTS

3.1 Dose sensitivity

To illustrate the effects of each medication in the post-treatment cases, we interpolated across the range of doses from the clinical data and tested the model for the change in the barrier for lamotrigine, benzodiazepines, and antipsychotics. Each medication reduced the barrier in a nearly linear dose response in this range (Figure 3a), but through different mechanisms of action. Lamotrigine acts to reduce excitation by both reducing the excitability of the excitatory neuron population and reducing the excitatory synaptic weights. The benzodiazepines act through increasing the inhibitory synaptic weights to reduce the boundary between states.

The antipsychotics have more complicated mechanisms of action through dopamine, serotonin, and muscarinic receptors. We model two types of dopamine receptors, D1 and D2. In our model, D1 receptor activation decrease the excitability of the excitatory neuron population and increase the excitability of the inhibitory neuron population, both contributing to increasing barrier when D1 receptors are blocked by antipsychotics. However, D1 activation also increases excitatory synaptic transmission to have the opposite effect on the barrier by antipsychotics that block D1. The D2 receptor activation reduces excitatory synaptic transmission and increases the excitability of the excitatory neuron population leading to opposite effects. Activation of the two serotonin receptors included the model (5-HT1A and 5-HT2A) have opposite effects on the excitability of the excitatory neuron population, and M1 receptor activation increases their excitability. The affect of each antipsychotic depends on the affinity of the molecule to each receptor in competition with the background level of neurotransmitter, and we find that there is a net decrease in the barrier for increasing dose of both clozapine and olanzapine.

To help untangle the competing effects of the medications, we investigated the dose response of model parameters, as shown in Figure 3. The benzodiazepine (clonazepam) has the simplest action and affects only the inhibitory synaptic weights (w_{00} and w_{01}) in the model. Lamotrigine affects both the excitatory neuron threshold and the excitatory synapses to yield an overall effect of reducing excitation. Clozapine has a more mixed effect on several parameters with the largest effect on the threshold of excitatory neuron that reduces their overall excitability.

209 3.2 BFCRS clinical scale

210 We calculated the changes in the model parameter for each synthesized subject caused by medications at
211 admission, and then after treatment was stabilized. With the modified parameters we could calculate the
212 barrier between the high-rate state and the low-rate state to observe whether the barrier was reduced. A
213 reduction in the barrier is interpreted as an improvement in catatonic symptoms. We find that the barrier
214 was reduced in all cases, as observed in clinical observations. We could then compare the BFCRS clinical
215 score with the barrier to visualize the effect of the treatment (Figure 5).

216 Although there is a clear reduction in the barrier, consistent with the reduction in BFCRS score, there is
217 an uncorrelated spread across the subjects before and after treatment ($r^2 = 0.73$). We have confirmed that
218 this is not due to lost correlations in our synthesized data, and must be attributed to individual differences
219 between subjects in both their pre-treatment disease state and their response to the medications.

220 3.3 Combination efficacy

221 The combination of medications in the treatment has been clinically observed to be additive, and this
222 observation can be explained by the parallel mechanisms of action. Lamotrigine and the benzodiazepines
223 act on different sites, excitatory neurons and inhibitory synapses. Although the antipsychotics have some
224 overlap with these parameters in the model, they act through different receptors. In the dynamic range of
225 medication effects on the barrier size, the dose response is nearly linear, and we find an additive effect of
226 the combination (Figure 6A). To relate the effect back to the clinic, we can use the correlation between the
227 BFCRS score and the boundary to map the boundary back to the BFCRS score to predict the effect of each
228 medication and their combinations on the average subject. We calculate the linear fit between the BFCRS
229 score and barrier before and after the treatment to obtain the mapping, and then plot the BFCRS score in
230 Figure 6B.

231 3.4 Personalization

232 The model is good at predicting large changes in BFCRS score for the population as a whole, but more
233 exact predictions of individuals should be possible with further parametrization. Ultimately, the model
234 could then be used as a tool for informing clinical care and refining treatments. Because the model has
235 few parameters to tune, then each subject could have a personalized model for use in the clinic. We
236 personalized the model by calibrating the initial state with model parameters, and then adjusted the dose
237 response parameter for each individual subject.

238 The first adjustment was to tune individual Wilson-Cowan model based on the the initial BFCRS score for
239 each patient. The barrier size can be adjusted in the Wilson-Cowan model so that patients with high BFCRS
240 scores will have a corresponding model with a high barrier. We have attempted to tune the w_{01} model
241 parameter to this end, but no clear result could be seen in the correlation of the outcomes to treatment.
242 Further research will be needed to determine whether different model parameters need to be tuned to be
243 more representative of the pathology underlying catatonia.

244 The second adjustment was to calibrate the individual dose response with model coupling parameters to
245 the effect on BFCRS score. As patients are admitted to the residence, they transition their medication to
246 the new regimen, and measures of the BFCRS score inform how each individual is affected by removing
247 and adding medications. These changes in BFCRS score could be used to calibrate individual mechanisms
248 and how they couple to model parameters. Such a tuning could create a model that adapts along with the
249 patient, and improves in its prediction power over time.

The results of these two modification are shown in Figure 7 where the new prediction of the barrier is compared with the BFCRS score. The higher correlation between the model barrier and the clinical score ($r^2 = 0.97$) gives confidence that the effects predicted by the model can guide further changes in medication, and aid the psychiatrist in clinical decisions.

4 DISCUSSION

The objective of this study was to demonstrate that a simple cortical model, with excitatory and inhibitory neural populations, is sufficiently descriptive to explain and predict clinical outcomes in schizophrenia patients with catatonia. The pharmaceutical coupling of the treatments to model parameters are based on known mechanisms of action in cortical neurons: pyramidal cells and parvalbumin positive inhibitory interneurons. We have demonstrated the utility of the model for explaining the observed clinical outcomes by tracing the action of medications to changes in the model dynamics by interpreting the change in the barrier between states as a change in a clinical measure, the BFCRS score. The model supports the clinical observation that the 3-medication combination, clozapine, lamotrigine, and a benzodiazepine, is additive, and explains how the pathways of action are independent on a mechanistic level. Finally, we took a first step at personalization of the model for individual subjects, with the goal of supporting individual clinical decisions with mechanistic explanations.

Augmenting psychiatric practice with a simple mechanistic model encourages a conceptual shift to a focus on reducing cortical excitability, either through reducing excitability of pyramidal neurons, or increasing inhibition. Each of the 3 medications are optimized on their own for safety and efficacy, but since they act on the excitability of the system through different mechanisms, they can have an additive effect on catatonic symptoms. Further use of this approach can suggest other means of controlling cortical excitability and inspire new treatment protocols.

Conceptualizing the action of this treatment as modifying excitability and connectivity of neuron populations also suggests mechanisms of observed clinical improvements. The clinical observation that reduced chronic catatonic features lead to meaningful improvements in social and cognitive function suggests that reducing the barrier represents a physical improvement in brain network connectivity and dynamical processing. Bursts of neural activity that control behavioral patterns become more flexible with a reduced barrier between states of excitation, and that flexibility leads to more fluid cognitive function and social behavior.

4.1 Extensions of the model

The model is based on cortical circuitry, in part because catatonia is thought to have a cortical origin. However, antipsychotics also target the striatum. Extending the model to include a cortical-striatum-thalamic loop would include additional dynamics that are presently missing. As yet, it is unknown if such an extension will add a precision that is visible in clinical usage, but this would be a rich area to explore.

One avenue to improve the model's predictions is to further personalize the model by individualizing the pharmacokinetics for each patient. When clozapine is administered, safety considerations require blood samples, and blood levels of clozapine have been recorded from many patients in this cohort. There is a wide variation in the dose response to blood serum concentration of clozapine, and these variations are not currently included in the model. We have tested the robustness of our results to ensure this observed variance does not affect the conclusions in this study, but clearly such an addition to the model will help to refine individual cases.

Further clinical variables may provide new insights into how model outputs can be interpreted. Although the BFCRS score has provided a good clinical guidance for this cohort, the addition of either cognitive or motor measures could augment the model's interpretation. Furthermore, additional clinical measures could add constraints that require a more detailed model, such as adding a striatum, that the BFCRS score alone will not capture. Although further complications may degrade the causal interpretability because of added complex dynamics, there are likely parameter regions with simpler dynamics that may broaden the applicability to other symptoms of psychiatric disease.

CONFLICT OF INTEREST STATEMENT

Author PDR is employed by Amazon Web Services, and author JC is/was employed by Cascadia Behavioral Healthcare. This research does not relate to PDR's position or activities at Amazon Web Services. The authors declare that the research was conducted in the absence of any other commercial or financial relationships that could be construed as a potential conflict of interest.

AUTHOR CONTRIBUTIONS

JC was in charge of clinical care and worked with PDR to synthesize data with statistics similar to clinical practice and observations. PDR developed the model and visualizations and wrote the initial draft of the manuscript. Both authors contributed equally on planning and finalizing the manuscript.

DATA AVAILABILITY STATEMENT

The datasets generated for this study can be found in the repo for Cascadia-Behavioral-Healthcare: <https://github.com/pdroberts/cascadia-behavioral-healthcare.git>.

REFERENCES

- Bagi, B., Brecht, M., and Sanguinetti-Scheck, J. I. (2022). Unsupervised discovery of behaviorally relevant brain states in rats playing hide-and-seek. *Current Biology*
- Benayoun, M., Cowan, J. D., van Drongelen, W., and Wallace, E. (2010). Avalanches in a stochastic model of spiking neurons. *PLoS computational biology* 6, e1000846
- Bhati, M. T., Datto, C. J., and O'Reardon, J. P. (2007). Clinical manifestations, diagnosis, and empirical treatments for catatonia. *Psychiatry (Edgmont)* 4, 46
- Bressloff, P. C. (2010). Metastable states and quasicycles in a stochastic wilson-cowan model of neuronal population dynamics. *Physical Review E* 82, 051903
- Buice, M. A., Cowan, J. D., and Chow, C. C. (2010). Systematic fluctuation expansion for neural network activity equations. *Neural computation* 22, 377–426
- Bush, G., Fink, M., Petrides, G., Dowling, F., and Francis, A. (1996). Catatonia. i. rating scale and standardized examination. *Acta Psychiatrica Scandinavica* 93, 129–136
- Carr, D. B., Cooper, D. C., Ulrich, S. L., Spruston, N., and Surmeier, D. J. (2002). Serotonin receptor activation inhibits sodium current and dendritic excitability in prefrontal cortex via a protein kinase c-dependent mechanism. *Journal of Neuroscience* 22, 6846–6855
- [Dataset] Conour, J. (2015). Compositions and methods for the treatment of catatonia. US Patent 9,066,949
- Cowan, J. D., Neuman, J., and van Drongelen, W. (2016). Wilson-cowan equations for neocortical dynamics. *The Journal of Mathematical Neuroscience* 6, 1–24

- 324 Dreyer, J. K., Herrik, K. F., Berg, R. W., and Hounsgaard, J. D. (2010). Influence of phasic and tonic
325 dopamine release on receptor activation. *The Journal of Neuroscience* 30, 14273–14283
- 326 Foehring, R. C. (1996). Serotonin modulates n- and p-type calcium currents in neocortical pyramidal
327 neurons via a membrane-delimited pathway. *Journal of Neurophysiology* 75, 648–659
- 328 Geerts, H., Roberts, P., and Spiros, A. (2013). A quantitative system pharmacology computer model for
329 cognitive deficits in schizophrenia. *CPT: pharmacometrics & systems pharmacology* 2, 1–8
- 330 Gray, J. A. and Risch, S. C. (2009). When clozapine is not enough: Augment with lamotrigine? *Current*
331 *Psychiatry* 8, 40–47
- 332 Hirjak, D., Kubera, K. M., Northoff, G., Fritze, S., Bertolino, A. L., Topor, C. E., et al. (2019). Cortical
333 contributions to distinct symptom dimensions of catatonia. *Schizophrenia bulletin* 45, 1184–1194
- 334 Katori, Y., Sakamoto, K., Saito, N., Tanji, J., Mushiaki, H., and Aihara, K. (2011). Representational
335 switching by dynamical reorganization of attractor structure in a network model of the prefrontal cortex.
336 *PLoS computational biology* 7, e1002266
- 337 Kotecha, S. A., Oak, J. N., Jackson, M. F., Perez, Y., Orser, B. A., Van Tol, H. H., et al. (2002). A D2
338 class dopamine receptor transactivates a receptor tyrosine kinase to inhibit NMDA receptor transmission.
339 *Neuron* 35, 1111–1122
- 340 Laphish, C. C., Kroener, S., Durstewitz, D., Lavin, A., and Seamans, J. K. (2007). The ability of the
341 mesocortical dopamine system to operate in distinct temporal modes. *Psychopharmacology* 191,
342 609–625
- 343 Lee, F. J., Xue, S., Pei, L., Vukusic, B., Chéry, N., Wang, Y., et al. (2002). Dual regulation of nmda receptor
344 functions by direct protein-protein interactions with the dopamine d1 receptor. *Cell* 111, 219–230
- 345 Miller, L. G., Greenblatt, D. J., Paul, S. M., and Shader, R. I. (1987). Benzodiazepine receptor occupancy
346 in vivo: correlation with brain concentrations and pharmacodynamic actions. *Journal of Pharmacology*
347 *and Experimental Therapeutics* 240, 516–522
- 348 Negahbani, E., Steyn-Ross, D. A., Steyn-Ross, M. L., Wilson, M. T., and Sleight, J. W. (2015).
349 Noise-induced precursors of state transitions in the stochastic wilson–cowan model. *The Journal*
350 *of Mathematical Neuroscience (JMN)* 5, 1–27
- 351 Northoff, G. (2002). What catatonia can tell us about “top-down modulation”: a neuropsychiatric hypothesis.
352 *Behavioral and Brain Sciences* 25, 555–577
- 353 Northoff, G., Kötter, R., Baumgart, F., Danos, P., Boeker, H., Kaulisch, T., et al. (2004). Orbitofrontal
354 cortical dysfunction in akinetic catatonia: a functional magnetic resonance imaging study during negative
355 emotional stimulation. *Schizophrenia bulletin* 30, 405–427
- 356 Paterson, L. M., Tyacke, R. J., Nutt, D. J., and Knudsen, G. M. (2010). Measuring endogenous 5-HT release
357 by emission tomography: promises and pitfalls. *Journal of Cerebral Blood Flow & Metabolism* 30,
358 1682–1706
- 359 Patki, N., Wedge, R., and Veeramachaneni, K. (2016). The synthetic data vault. In *2016 IEEE International*
360 *Conference on Data Science and Advanced Analytics (DSAA)*. 399–410
- 361 Penland, H. R., Weder, N., and Tampi, R. R. (2006). The catatonic dilemma expanded. *Annals of General*
362 *Psychiatry* 5, 14
- 363 Perez-Rosello, T., Figueroa, A., Salgado, H., Vilchis, C., Tecuapetla, F., Guzman, J. N., et al. (2005).
364 Cholinergic control of firing pattern and neurotransmission in rat neostriatal projection neurons: role of
365 cav2. 1 and cav2. 2 Ca²⁺ channels. *Journal of neurophysiology* 93, 2507–2519
- 366 Poolos, N. P., Migliore, M., and Johnston, D. (2002). Pharmacological upregulation of h-channels reduces
367 the excitability of pyramidal neuron dendrites. *Nature neuroscience* 5, 767–774
- 368 Rosebush, P. I. and Mazurek, M. F. (2010). Catatonia and its treatment. *Schizophrenia bulletin* 36, 239–242

- [Dataset] Roth, B. and Lopez, E. (2006). Psychoactive drug screening program Ki database. <https://kiddbdev.med.unc.edu/databases/downloadki.html>
- Spiros, A., Carr, R., and Geerts, H. (2010). Not all partial dopamine d2 receptor agonists are the same in treating schizophrenia. exploring the effects of bifeprunox and aripiprazole using a computer model of a primate striatal dopaminergic synapse. *Neuropsychiatric disease and treatment* 6, 589
- Tiihonen, J., Hallikainen, T., Ryyänänen, O.-P., Repo-Tiihonen, E., Kotilainen, I., Eronen, M., et al. (2003). Lamotrigine in treatment-resistant schizophrenia: a randomized placebo-controlled crossover trial. *Biological psychiatry* 54, 1241–1248
- Ungvari, G. S., Leung, S. K., Ng, F. S., Cheung, H.-K., and Leung, T. (2005). Schizophrenia with prominent catatonic features ('catatonic schizophrenia'): I. demographic and clinical correlates in the chronic phase. *Progress in Neuro-Psychopharmacology and Biological Psychiatry* 29, 27–38
- Wang, S.-J., Sihra, T. S., and Gean, P.-W. (2001). Lamotrigine inhibition of glutamate release from isolated cerebrocortical nerve terminals (synaptosomes) by suppression of voltage-activated calcium channel activity. *Neuroreport* 12, 2255–2258
- Wang, Z.-X. (1995). An exact mathematical expression for describing competitive binding of two different ligands to a protein molecule. *FEBS letters* 360, 111–114
- Wilcox, J. A. and Reid Duffy, P. (2015). The syndrome of catatonia. *Behavioral Sciences* 5, 576–588
- Wilson, H. R. and Cowan, J. D. (1972). Excitatory and inhibitory interactions in localized populations of model neurons. *Biophys. J.* 12, 1–24
- Xie, X., Lancaster, B., Peakman, T., and Garthwaite, J. (1995). Interaction of the antiepileptic drug lamotrigine with recombinant rat brain type iia na⁺ channels and with native na⁺ channels in rat hippocampal neurones. *Pflügers Archiv* 430, 437–446

SUPPLEMENTARY DATA

391 *Data Selection Filter*

- 392 • Remove subjects if pre-treatment includes clozapine.
- 393 • Remove subjects if pre-treatment includes lamotrigine.
- 394 • Remove subjects if pre-treatment has olanzapine is greater than 20mg/d.
- 395 • Remove subjects with pre-treatment includes both clozapine and olanzapine.
- 396 • Remove subjects if post-treatment of Lamotrigine is less than 400mg of greater than 500mg.
- 397 • Remove subjects if pre-treatment does not include antipsychotics.
- 398 • Remove subjects if post-treatment does not include antipsychotics.
- 399 • Remove subjects if post-treatment includes methylphenidate.
- 400 • Remove subjects if pre-treatment BFCRS score is less than or equal to post-treatment score.
- 401 • Remove subjects if their BFCRS score changes less than 9 (for consistency with clinical dataset).
- 402 • Remove subjects if the number of post-treatment medications is less than 3.
- 403 • Remove subjects if post-treatment includes both clonazepam and lorazepam.
- 404 • Remove subjects if pre-treatment has more than 1 mood stabilizer.
- 405 • Balanced the number of subjects with clozapine post-treatment to 80%.

406 *Pharmacokinetic Parameters:* Partial list of the most prominent medications in the study.

| | param | Clozapine | Olanzapine | Lamotrigine | Clonazepam | Lorazepam |
|-----|-------------------|-----------|------------|-------------|------------|-----------|
| | bioavailability | 0.65 | 0.87 | 0.950 | 0.9 | 0.85 |
| 407 | clearance (L/hr) | 31.00 | 21.80 | 2.100 | 2.1 | 4.30 |
| | blood/brain ratio | 1.00 | 1.00 | 1.900 | 1.0 | 1.53 |
| | mole wt (g/mol) | 326.82 | 312.43 | 256.091 | 315.7 | 321.20 |
| | pk_param | 1.00 | 1.00 | 1.000 | 0.3 | 10.00 |

408 *Receptor Affinities:* Partial list of the most prominent medications in the study from Roth and Lopez
409 (2006).

| | | Dopamine | 5-HT | Clozapine | Olanzapine | Lamotrigine | Clonazepam | Lorazepam |
|-----|-----------|----------|--------|-----------|------------|-------------|------------|-----------|
| | receptors | | | | | | | |
| 410 | D1 | 130.0 | 9690.0 | 89.0 | 25.00 | 10000 | 10000 | 10000 |
| | D2 | 469.43 | 10000 | 28.0 | 3.00 | 10000 | 10000 | 10000 |
| | 5-HT1A | 8248.0 | 2.789 | 104.8 | 610.00 | 10000 | 10000 | 10000 |
| | 5-HT2A | 10000 | 20.80 | 1.0 | 1.48 | 10000 | 10000 | 10000 |
| | M1 | 10000 | 10000 | 1.4 | 2.00 | 10000 | 10000 | 10000 |

FIGURE CAPTIONS

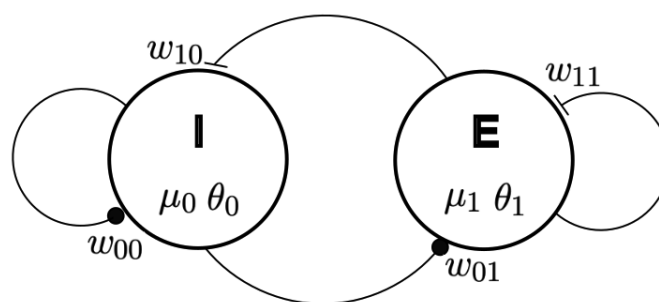


Figure 1a. Wilson-Cowan circuit with an inhibitory (I) and excitatory (E) neuron population. The model parameters associated with each circuit element are shown.

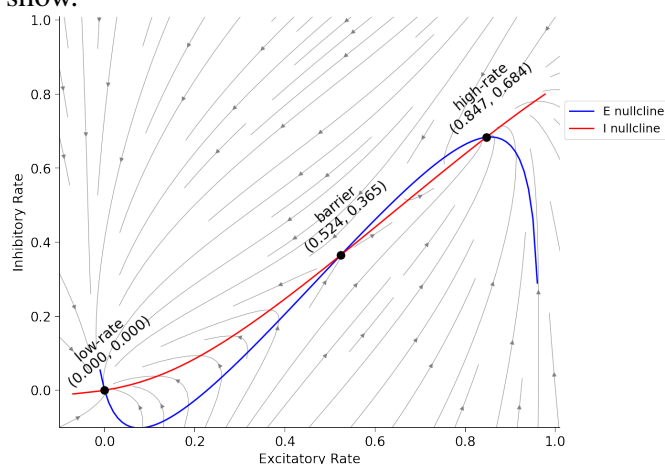


Figure 1b. Phase plane of the Wilson-Cowan model with trajectories, nullclines and fixed points labeled.

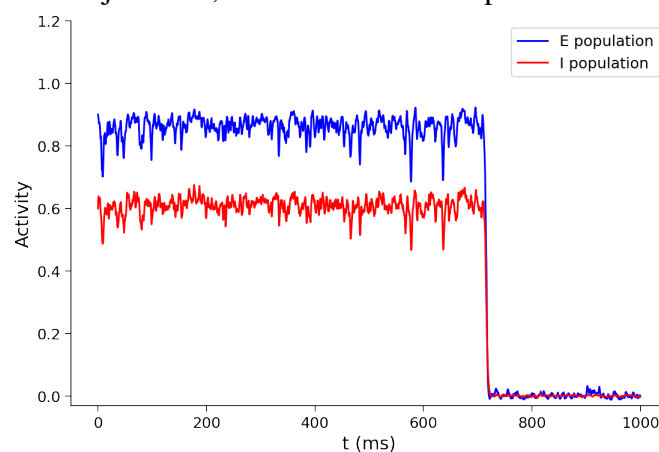


Figure 1c. Sustained activity under the influence of noise eventually decays. If the boundary is too high, then the sustained burst continues indefinitely. Treatments reduce the boundary between the states and transitions become more fluid.

Figure 1. Wilson-Cowan model and dynamics. (a) Wilson-Cowan model circuit. (b) Phase plane of the Wilson-Cowan model. (c) Sustained activity in the high-rate state.

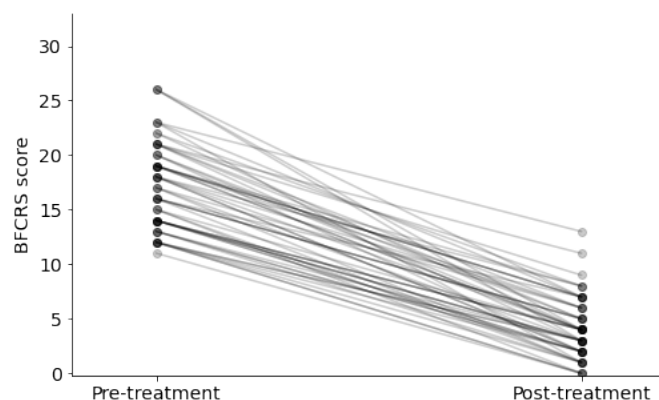


Figure 2a. BFCRS score for 59 synthesized data subjects before and after treatment.

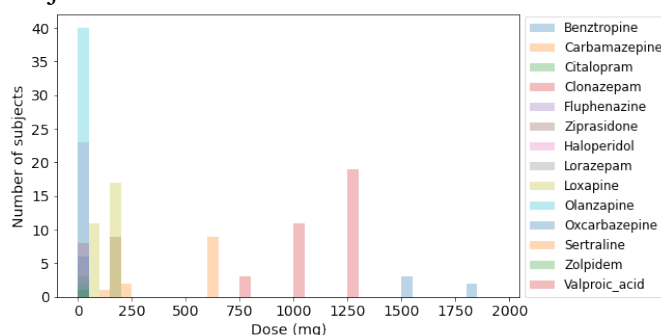


Figure 2b. Distribution of medication doses across all subjects before treatment.

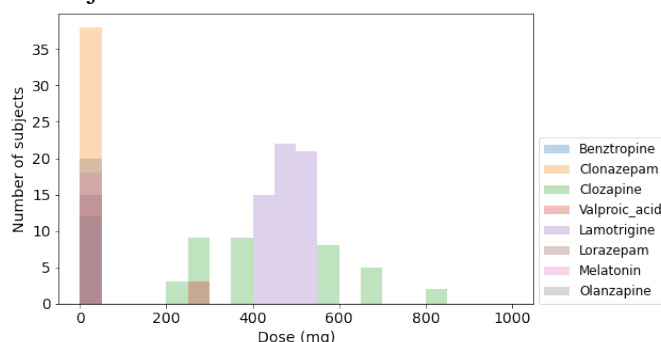


Figure 2c. Distribution of medication doses across all subjects after treatment.

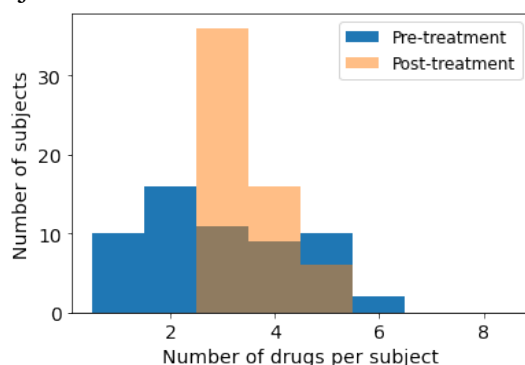


Figure 2d. Distribution of the number of medications for each subject before and after treatment.

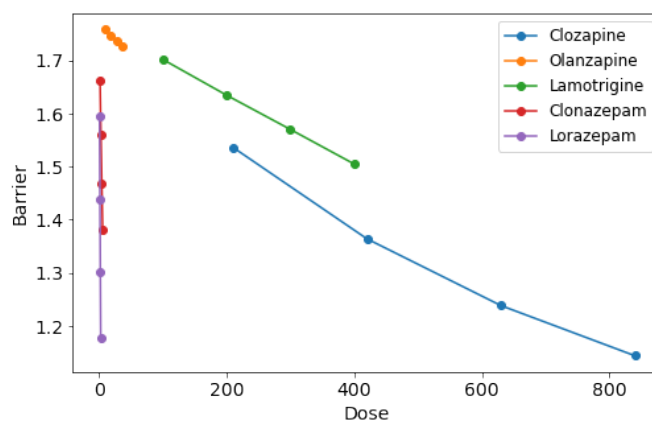


Figure 3a. Dose response of the barrier between the high-rate state and the low-rate state for medications in the treatment.

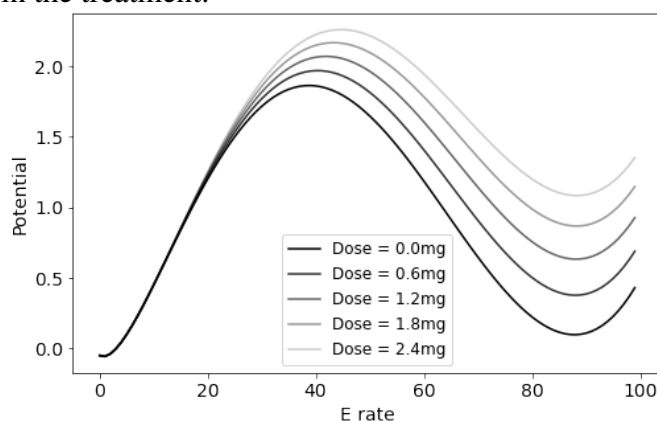


Figure 3b. Dose response of potential to show how the barrier becomes smaller with increasing doses of lorazepam. The two stable fixed points are where the excitatory rate is 0 and ~ 90 . The peak of the barrier is the unstable fixed point where the excitatory rate is ~ 40 . The vertical distance from the high-rate basin to the unstable peak is the barrier.

Figure 3. Dose response of model parameters for lamotrigine, clozapine, and clonazepam. The dose response is nearly linear in this range of the model.

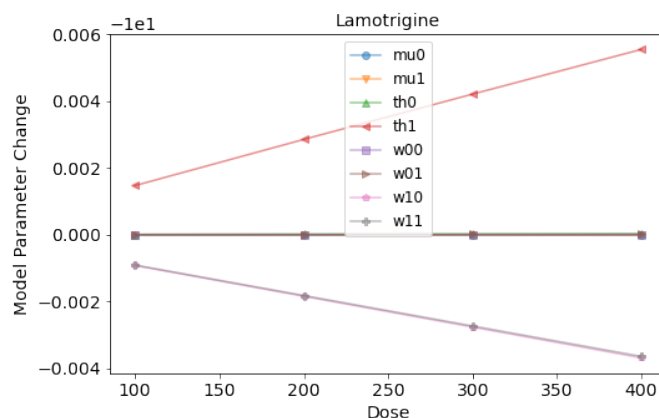


Figure 4a. Dose response of model parameters for lamotrigine. The threshold of excitatory increases with increasing dose leading to a decrease of the neuron's excitability. The excitatory synaptic parameters (w_{11} and w_{10}) decrease leading to a reduced excitation of the system.

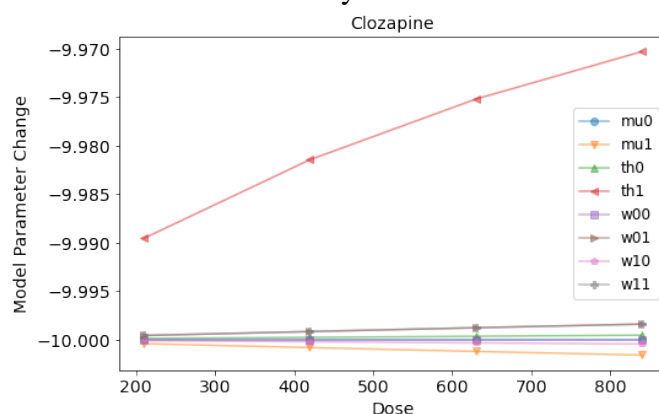


Figure 4b. Dose response of model parameters for clozapine. Several parameters are affected, but the largest effect is an increase of the threshold in excitatory neurons due to blocking M1 and 5-HT_{2A} receptors reducing the excitation of the system.

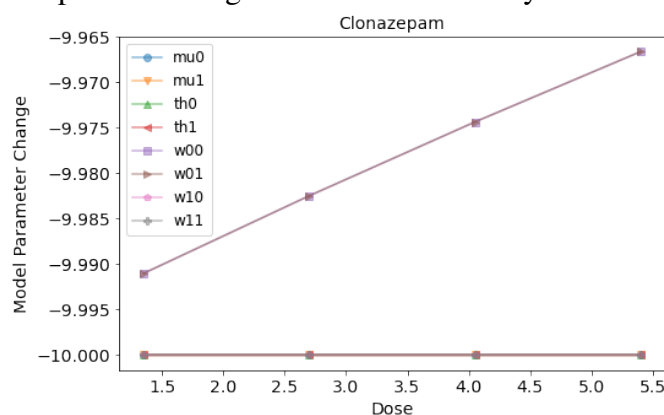


Figure 4c. Dose response of model parameters for clonazepam. Only the inhibitory synaptic parameters are affected.

Figure 4. Dose response of model parameters for lamotrigine, clozapine, and clonazepam.

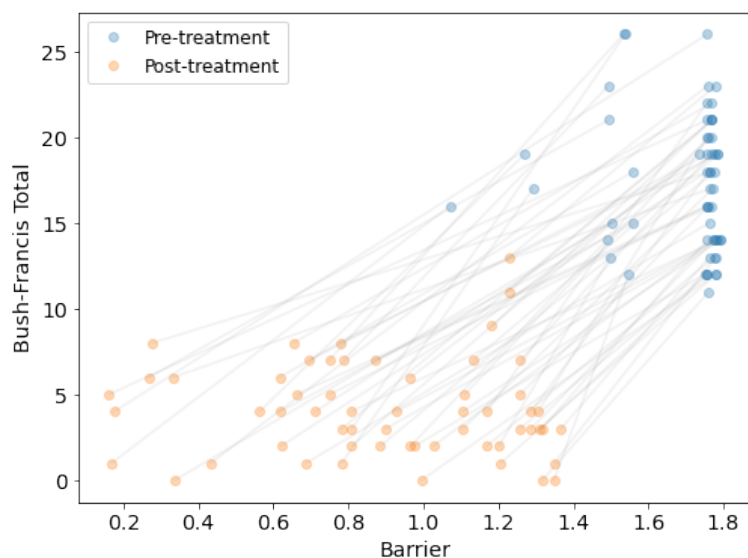


Figure 5. Model predicted barrier and the synthesized BFCRS score for subjects before and after treatment. The grey lines associate the pre- and post-treatment scores for the same subject.

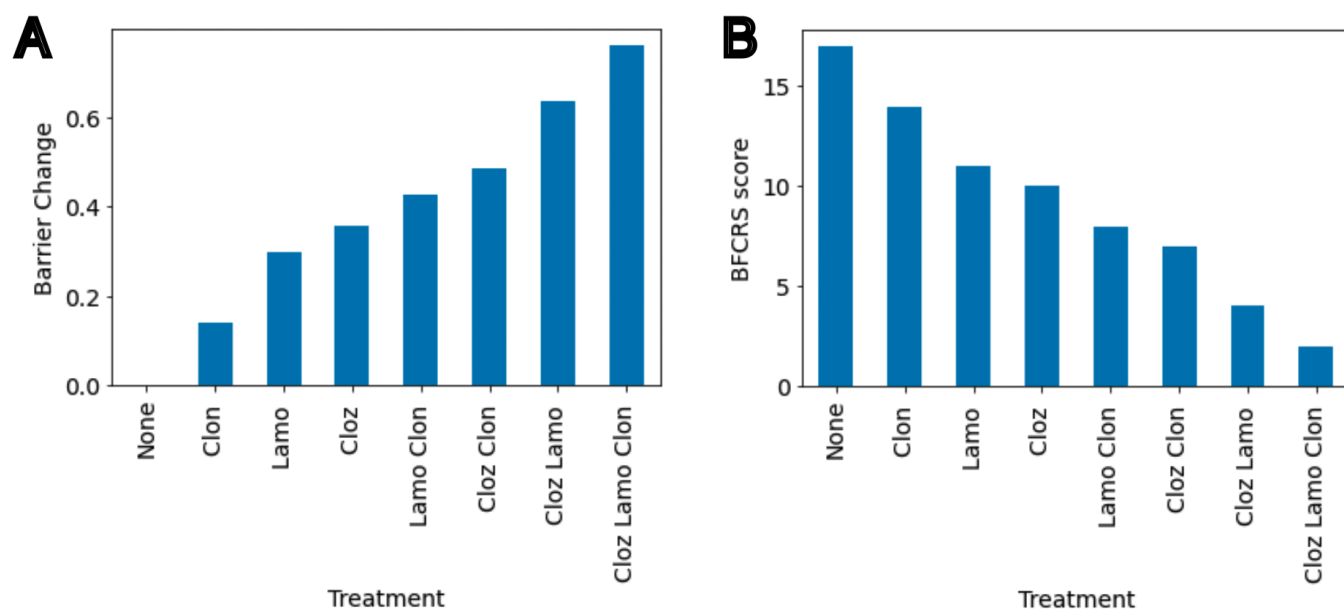


Figure 6. Model results for combinations of lamotrigine, clozapine, and clonazepam demonstrating the additive effects. A. Barrier for combinations of medications in the treatment protocol. B. Predicted BFCRS for combinations of medications in the treatment protocol.

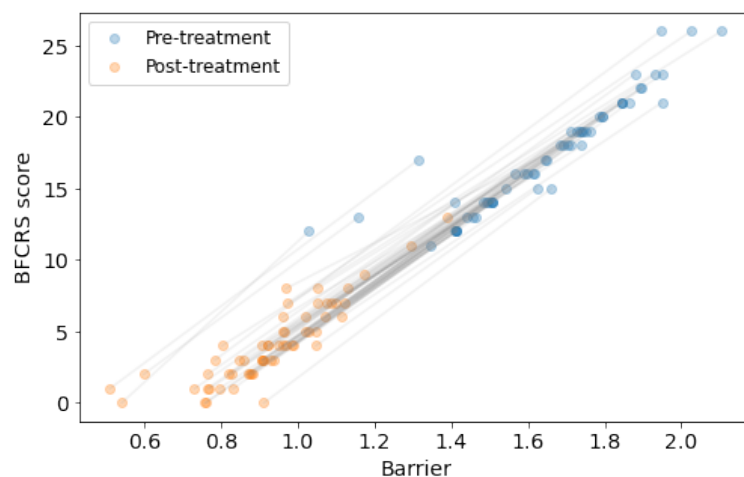


Figure 7. Personalized model prediction of barrier and the synthesized BFCRS score for subjects before and after treatment. The grey lines associate the pre- and post-treatment scores for the same subject.