

DNA (Cell Number) and Protein in Rat Brain

Second Generation (F_2) Alteration by Maternal (F_0) Dietary Protein Restriction¹

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Abstract. Female rats (F_0) were maintained on a normal (C) or a protein-restricted (8%) diet (R) 1 month prior to mating and throughout pregnancy. Their offspring (F_1) were nursed by their natural or foster mothers on normal diet or protein-restricted diet. All offspring were maintained on normal diet after weaning; the females were mated with normal males, and one group of R males with normal females. The second generation (F_2) was maintained on normal diet from birth. Body weight and cerebral weight, DNA and protein were measured in F_1 and F_2 , at birth and at 30 and 90 days. Previously-reported developmental deficiencies were observed in F_1 in all restricted groups at birth and at 30 days, but only in DNA of some groups at 90 days. The F_2 offspring from all experimental females had at birth significantly lower cerebral DNA (cell number); in most groups this deficiency in F_2 did not persist at 30 days. Cerebral deficiencies in newborn F_2 resulting from protein restriction in F_0 or F_1 were transmitted to the next (F_2) generation through females but not through males. Several explanations of this effect are offered.

Key Words

Rat
Dietary protein restriction
 F_2 generation
Brain
DNA
Protein
Cerebral deficiency
Transmission through females

In our previous work [28] we have reported that when female rats were maintained on a low-protein (8%) diet 1 month prior to mating and throughout pregnancy, their offspring at birth had significantly smaller body weights, cerebral weights, cerebral DNA (cell number) and cerebral protein. Similar results were obtained by WINICK [16] and by ZEMAN and STANBROUGH [31].

The present work was designed to investigate whether a malnutrition-induced injury in one generation can be transmitted to the next generation,

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with particular reference to brain underdevelopment. The possibility of a transfer of maternal constraint (when she was a fetus), affecting body weight and behavior of her offspring, has been indicated in the past [5, 11, 15]; it has also been suggested that such constraint has a survival value at times of food shortage [11].

Experimental

Sprague-Dawley-derived albino rats bred as a closed colony in our laboratory for 22 generations were used in the previous and present work [14, 22–28]. 60-day-old virgin females (F_0 generation) were randomly divided into an experimental and control group. The experimental group (R) was fed a protein-restricted diet (8%) and the control group (C) was fed a standard control pellet diet, as described previously [28]. Food intakes were calculated for both groups by feeding known amounts and weighing the remainder. At 90 days of age, all females were mated and maintained on their respective diets throughout gestation. The food intakes during the month prior to mating averaged 12.6 g/24 h for group R and 13.4 g/24 h for group C. During pregnancy the intake was 13.1 g/24 h for group R and 13.2 g/24 h for group C.

In the F_1 generation all litters at birth (within 6 h after delivery) were standardized to 8 newborns per female; the excess newborns were used for biochemical analysis (table I). After decapitation, the brains (cerebral hemispheres, without cerebellum and olfactory lobes) were immediately removed and weighed; they were then frozen and subsequently used for analysis. DNA was determined by a modification of the diphenylamine colorimetric method [10, 19, 21]; protein was determined by a modification of the Lowry colorimetric method [9].

For the F_1 generation groups C and R were further subdivided depending on the diet fed during weaning. The designations for these groups are as follows: CC, CR, RR, RC, C(R)R, and R(C)C. The first letter indicates the *pre-partum* diet of the biological mother, and the second letter indicates the type of maternal diet during weaning. Two groups were cross-fostered and the letter in parenthesis indicates the *pre-partum* diet of the foster mother.

During the weaning period the food intakes were calculated as 16.8 g/24 h for females on the control diet and 16.0 g/24 h for those on the protein-restricted diet. After weaning all F_1 animals were placed on the control diet for the remaining part of the experiment.

At maturity the F_1 females were mated with normal males. The males used for mating were fed control diet except that in one experiment, to test the nontransmission to F_2 of male deficiencies in F_1 , adult males of group RC were mated with control females.

Biochemical analyses were performed on F_1 and F_2 newborns (table I), 30-days F_1 and F_2 males (table II), and adult F_1 and F_2 females (table III).

Results

Tables I–III represent the effects of maternal protein restriction on neonatal, 30- and 90-day-old offspring in F_1 and F_2 . Maternal (F_0) protein restriction to 8% 1 month prior to mating and throughout gestation resulted

Table I. The effect of maternal protein restriction on neonatal offspring in F₁ and F₂

Group ¹	Genera- tion	Number	Body weight ² g	Cerebrum ²		
				weight, mg	DNA, μ g	protein, mg
CC	F ₁	123	5.7 \pm 0.6	172 \pm 13	572 \pm 44	9.7 \pm 1.6
	F ₂	74	6.1 \pm 0.5	170 \pm 13	598 \pm 33	8.8 \pm 0.8
RR	F ₁	136	4.9 \pm 0.9*	149 \pm 21*	532 \pm 46*	8.1 \pm 1.2*
	F ₂	59	6.1 \pm 0.8	168 \pm 12	562 \pm 43*	8.8 \pm 0.9
RC	F ₁	same as group RR				
	F ₂	63	5.7 \pm 0.5*	158 \pm 15*	565 \pm 35*	8.3 \pm 0.8**
R(C)C	F ₁	same as group RR				
	F ₂	59	5.9 \pm 0.5	159 \pm 16*	555 \pm 32*	8.2 \pm 1.1**
C(R)R	F ₁	same as CC				
	F ₂	50	6.3 \pm 0.8	169 \pm 17	557 \pm 38*	8.7 \pm 0.9
CR	F ₁	same as CC				
	F ₂	48	5.9 \pm 0.7	160 \pm 18**	569 \pm 41*	8.3 \pm 1.2

¹ See text (Experimental) for explanation of groups.

² Each value represents the mean \pm SD.

* Significant at $p < 0.001$ level. ** Significant at $0.01 > p > 0.001$ level. Standard t-test. All comparisons made to the control (CC).

in significant decreases in all measured parameters (body weight, cerebral weight, DNA and protein) in their offspring (F₁) at birth (table I, groups RR, RC, R[C]C). F₂ newborns in groups RC and R(C)C had significant deficiencies in neonatal cerebral DNA and practically all other neonatal parameters. In these 2 groups (RC and R[C]C), at the age of 30 days, most of these deficiencies remained significant for F₁ but not for F₂, in which even a transient increase in DNA was observed for RC. Practically all deficiencies became nonsignificant at maturity (90 days) for both F₁ and F₂.

When group RC F₁ adult males were mated with control females, no significant deficiency of any parameter in F₂ could be demonstrated even for neonatal animals (body weight 6.7 g; cerebrum: weight 175 mg; DNA 587 μ g, protein 8.02 mg).

Groups CR and C(R)R in the F₁ generation were significantly lower in all parameters at 30 days and in cerebral weight and cerebral DNA – at 90 days.

Table II. The effect of maternal protein restriction on 30-day-old male offspring in F₁ and F₂

Group ¹	Generation	Number	Body weight ² , g	Cerebrum ²		
				weight, mg	DNA, μ g	protein, mg
CC	F ₁	27	73 \pm 9	1,031 \pm 40	913 \pm 52	80.1 \pm 6.8
	F ₂	27	73 \pm 9	1,031 \pm 40	913 \pm 52	80.1 \pm 6.8
RR	F ₁	10	36 \pm 11*	904 \pm 75*	848 \pm 45**	71.6 \pm 6.2**
	F ₂	15	77 \pm 14	1,031 \pm 48	1,034 \pm 81*	79.2 \pm 15.0
RC	F ₁	15	72 \pm 18	968 \pm 56**	836 \pm 48*	71.7 \pm 5.4*
	F ₂	28	67 \pm 15	999 \pm 85	1,013 \pm 62*	78.6 \pm 11.7
R(C)C	F ₁	16	60 \pm 14**	1,008 \pm 50	846 \pm 51*	70.4 \pm 4.2*
	F ₂	12	77 \pm 23	1,002 \pm 79	916 \pm 61	70.5 \pm 6.6**
C(R)R	F ₁	11	38 \pm 19*	879 \pm 91*	803 \pm 58*	63.0 \pm 6.1*
	F ₂	11	65 \pm 12	975 \pm 44**	985 \pm 62**	74.1 \pm 9.4
CR	F ₁	20	28 \pm 6*	882 \pm 62*	865 \pm 58**	69.7 \pm 7.4*
	F ₂	10	73 \pm 15	1,012 \pm 79	912 \pm 42	70.6 \pm 5.9**

¹ Groups as in table I.

² As in table I.

* and **, see table I.

Table III. The effect of maternal protein restriction on 90-day-old female offspring in F₁ and F₂

Group ¹	Generation	Number	Body weight ² , g	Cerebrum ²		
				weight, mg	DNA, μ g	protein, mg
CC	F ₁	77	269 \pm 41	1,197 \pm 52	1,040 \pm 105	93.8 \pm 9.0
	F ₂	49	262 \pm 43	1,206 \pm 57	1,103 \pm 71	98.6 \pm 6.4
RR	F ₁	15	243 \pm 35	1,084 \pm 91*	972 \pm 72**	87.9 \pm 16.8
	F ₂	24	254 \pm 33	1,167 \pm 54**	1,025 \pm 57*	97.3 \pm 7.0
RC	F ₁	20	247 \pm 45	1,129 \pm 69*	997 \pm 77	100.2 \pm 11.8
	F ₂	28	271 \pm 32	1,210 \pm 60	1,087 \pm 47	98.9 \pm 7.7
R(C)C	F ₁	15	280 \pm 32	1,167 \pm 70	1,001 \pm 102	102.1 \pm 14.0
	F ₂	6	276 \pm 30	1,129 \pm 86	1,004 \pm 91	94.8 \pm 7.5
C(R)R	F ₁	12	249 \pm 23	1,097 \pm 59*	953 \pm 77**	92.6 \pm 10.8
	F ₂	4	244 \pm 15	1,115 \pm 19**	996 \pm 47	89.2 \pm 7.9
CR	F ₁	18	260 \pm 36	1,127 \pm 57*	916 \pm 67*	92.5 \pm 13.6
	F ₂	6	249 \pm 15	1,197 \pm 66	1,002 \pm 40**	102.7 \pm 10.3

¹ See table I.

² See table I.

* and **, see table I.

In F_2 decreases were significant for cerebral DNA and/or cerebral weight at birth and 90 days, but not at 30 days when a transient increase in DNA in group C(R)R was observed.

Discussion

With reference to neonatal parameters in F_1 the present work confirms the results previously reported by us [28] and confirmed by others [16, 31]: Maternal prenatal (F_0) protein restriction (8%) results in significant decreases in neonatal body weight, cerebral weight, cerebral DNA and cerebral protein. It is of interest that these decreases persist postnatally till at least 30 days even in groups RC and R(C)C where the lactating female was receiving control diet. As discussed in previous publications, the determination of brain DNA is a convenient and objective quantitative method for determination of total brain cell numbers [22, 23, 25]; neuron (neuroblast) proliferation in the rat ceases at or before birth [review in ref. 23 and 25], and normal glia (glioblast) proliferation – at weaning [review in ref. 16]. It has been reported that at birth the neuron/glia ratio reaches 5, and at 30 days 2 or 3 [2]; this suggests that the DNA (cell number) decrease observed at birth reflects the deficiency of neuroblasts themselves. The fact that the percentage decrease of DNA (cell number) remains essentially the same at 30 days (despite a 60-percent increase in total DNA) suggests that the number of glia (glioblasts), proliferating after birth, also suffered a deficiency, and that this deficiency essentially followed the deficiency in number of neurons (neuroblasts).

In addition to quantitative changes in number of cells, the cerebrum in F_1 also undergoes qualitative changes: At birth and at 30 days the deficiency in total cerebral protein is higher than the deficiency in the number of cerebral cells which signifies less protein per cell.

Thus, prenatal nutritional protein restriction alone (RC and R[C]C) is sufficient to sustain cerebral deficiencies past the age (21 days) at which the rat cerebrum is considered essentially mature [19].

Additional postnatal deprivation (group RR) adds considerably to the decreases in body weight and cerebral weight at 30 days, but does not alter decreases in cerebral DNA (neuron and glia number) and cerebral protein; thus, it appears that for these 2 cerebral parameters the postnatal nutritional insult did not augment the effects of the prenatal one. It is of interest that WINICK [17] did observe augmentation in a different type of pre-weaning malnutrition.

Between 30 and 90 days, cerebral DNA (controls) increases by an additional 14%. This increase might be due to further proliferation of non-neuronal cells² or further conversion of diploid cells into polyploid [7]. It is of interest that most of the deficiencies present at 30 days in groups RC and R(C)C, disappear at the age of 90 days [compare also ref. 4 and 12]. Conceivably, in these experimental animals, there is more proliferation of non-neuronal cells, or even glial cells, to fill the deficient volume; the latter is known to occur after brain injuries. Thus, disappearance of deficiencies at 90 days does not mean that the animals are rehabilitated: The deficiency of neurons (final at birth) may persist even though total DNA becomes normal. In group RR, the added postnatal restriction of F_1 suffices to carry their DNA deficiency to at least 90 days.

Entirely unexpected results were obtained when the females that appeared essentially normal in F_1 at 90 days (groups RC and R(C)C) were mated with normal males: Their offspring at birth still had significant decreases in almost all parameters. This effect was transmitted only through females: The F_1 males of group RC, mated with normal females, produced normal offspring. Thus, the effect does not follow Mendelian genetics. Chromosome abnormalities, produced by malnutrition, have recently been reported [1]; in the present study, however, the transmission appears to be due not to genetic but to environmental effects on the organism of the F_1 mothers.

The nature of this effect is, at present, not clear. One possible explanation could be that the effect on F_2 neonatal cerebra was due to poor lactation of F_0 -nursing mothers that were protein-restricted before and during pregnancy (group RC). That this was not the case is apparent from group R(C)C, in which the effects on the neonatal cerebral parameters were essentially the same as in RC, even though the nursing mothers were never protein-restricted. Also, when the nursing F_0 mothers were protein-restricted before and during pregnancy as well as during the lactation period (group RR), the effect on cerebral DNA (cell number) in F_2 was not greater than in the case of protein restriction before and during pregnancy alone (groups RC and R(C)C).

Another explanation is based on the possibility that due to protein restriction of F_0 mothers before and during pregnancy, the F_1 animals were born handicapped, not only with regard to the brain [28] but also in other respects. ZEMAN [30] and HALL and ZEMAN [6] have reported that the off-

² The estimated proportion of nonneuronal cells in cerebral cortex of young adult rats is 30–35% [2].

spring of rats similarly protein-restricted during pregnancy suffer from retardation of kidney development and altered kidney function. They may also lack the vigor to suckle [29]. LEE and CHOW [8] have reported that the restricted progeny showed reduced feed efficiency and low nitrogen balance; they excreted more amino acids than the controls. Thus, such progeny (F_1) may indeed suffer from *cryptic* malnutrition, even when postnatally given full access to normal food (groups RC and R[C]C), and consequently their F_2 progeny would have a deficiency in all parameters.

Another possibility is that the handicap of F_1 animals involved their endocrine glands, as demonstrated recently by STEPHAN *et al.* [13]; in this case their deficiency would be not of the direct nutritional nature as postulated above, but would rather involve hormonal regulation. The glands affected (underdeveloped?) could be the hypothalamus, known to be sensitive to amino acid levels, and the pituitary [13], known to affect placental development [for a recent discussion, see ref. 25]; the latter might be correlated with brain development [20]. Thus, cerebral development in F_2 offspring would be limited by maternal (but not paternal) endocrine gland development in F_1 .

Since the above deficiencies are demonstrated in neonatal (F_2) cerebra, they are likely to reflect primarily the deficiency in the number of neurons, as previously discussed. After the birth of F_2 , placental F_1 underdevelopment becomes immaterial, and evidently the injury is of a different nature or different extent than in the previous generation, because the deficiencies cannot be demonstrated even at 30 days (groups RC and R[C]C). At 30 days (RC) there is even an unexplainable transient increase in DNA [compare also ref. 4], possibly due to proliferation of nonneuronal cells. Again, this does not imply that the animals are now rehabilitated because the neuron deficiency is likely to persist even if postnatal total DNA becomes normal or elevated.

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References

- 1 ARMENDARES, S.; SALAMANCA, F., and FRENK, F.: Chromosome abnormalities in severe protein calorie malnutrition. *Nature, Lond.* 232: 271 (1971).
- 2 BRIZZEE, K. R.; VOGT, J., and KHARETCHKO, X.: Postnatal changes in glia/neuron index with a comparison of methods of cell enumeration in the white rat. *Progr. Brain Res.* 4: 136 (1964).

- 3 CALEY, D.W. and MAXWELL, D.S.: An electron microscopic study of neurons during postnatal development of rat cerebral cortex. *J. comp. Neurol.* **133**: 17 (1968).
- 4 CHOW, B.F. and STEPHAN, J.K.: Fetal undernourishment and growth potential. *Nutr. Rep. Int.* **4**: 245 (1971).
- 5 COWLEY, J.J. and GRIESEL, R.D.: The effect on growth and behaviour of rehabilitating first and second generation low protein rats. *Anim. Behav.* **14**: 506 (1966).
- 6 HALL, S.M. and ZEMAN, F.J.: Kidney function of the progeny of rats fed a low protein diet. *J. Nutr.* **95**: 49 (1968).
- 7 HERMAN, C.J. and LAPHAM, L.W.: Neuronal polyploidy and nuclear volumes in cat central nervous system. *Brain Res.*, **15**: 35 (1969).
- 8 LEE, C.J. and CHOW, B.F.: Metabolism of proteins by progeny of underfed mother rats. *J. Nutr.* **94**: 20 (1968).
- 9 LOWRY, O.M.; ROSEBROUGH, N.J.; FARR, A.L., and RANDALL, R.J.: Protein measurement with the Folin phenol reagent. *J. biol. Chem.* **193**: 265 (1951).
- 10 MARGOLIS, F.L.: DNA and DNA-polymerase activity in chicken brain regions during
- 11 OUNSTED, M.: Familial factors affecting fetal growth; in *Prenatal factors affecting human development*. Pan-Amer. Health Organization, Scientific Publication No. 185, p. 60 (World Health Organization, Washington, D.C. 1969).
- 12 STEPHAN, J.K.: The permanent effect of prenatal dietary restriction on the brain of the progeny. *Nutr. Rep. Int.* **4**: 257 (1971).
- 13 STEPHAN, J.K.; CHOW, B.; FROHMAN, L.A., and CHOW, B.F.: Relationship of growth hormone to the growth retardation associated with maternal dietary restriction. *J. Nutr.* **101**: 1453 (1971).
- 14 VAN MARTHENS, E. and ZAMENHOF, S.: Deoxyribonucleic acid of neonatal brain cerebrum increased by operative restriction of litter size. *Exp. Neurol.* **23**: 214 (1969).
- 15 WEHMER, F.; PORTER, R.H., and SCALES, B.: Pre-mating and pregnancy stress in rats affects behaviour of grand pups. *Nature, Lond.* **227**: 622 (1970).
- 16 WINICK, M.: Malnutrition and brain development. *J. Pediat.* **74**: 667 (1969).
- 17 WINICK, M.: 1970 Cellular growth in intrauterine malnutrition. *Pediat. Clin. N. Amer.* **17**: 69 (1970).
- 18 WINICK, M. and NOBLE, A.: Cellular response in rats during malnutrition at various ages. *J. Nutr.* **89**: 300 (1966).
- 19 ZAMENHOF, S.; BURSHTYN, H.; RICH, K., and ZAMENHOF, P. J.: The determination of deoxyribonucleic acid and of cell number in brain. *J. Neurochem.* **11**: 505 (1964).
- 20 ZAMENHOF, S.; GRAUEL, L., and VAN MARTHENS, E.: Study of possible correlations between prenatal brain development and placental weight. *Biol. Neonat.* **18**: 140 (1971).
- 21 ZAMENHOF, S.; GRAUEL, L.; VAN MARTHENS, E., and STILLINGER, R.A.: Quantitative determination of DNA in preserved brains and brain sections. *J. Neurochem.* **19**: 61 (1971).
- 22 ZAMENHOF, S. and VAN MARTHENS, E.: Hormonal and nutritional aspects of prenatal brain development; in *PEASE Cellular aspects of neural growth and differentiation*. UCLA Forum in Medical Sciences No. 14, p. 329 (University of California Press, Berkeley 1971).
- 23 ZAMENHOF, S.; VAN MARTHENS, E., and BURSHTYN, H.: The effect of hormones on DNA synthesis and cell number in the developing chick and rat brain; in *HAMBURGH and BARRINGTON Hormones in development*, p. 101 (Appleton-Century-Crofts, New York 1971).

- 24 ZAMENHOF, S.; VAN MARTHENS, E., and GRAUEL, L.: DNA (cell number) in neonatal brain. Second generation (F_2) alteration by maternal (F_0) dietary protein restriction. *Science* 172: 850 (1971).
- 25 ZAMENHOF, S.; VAN MARTHENS, E., and GRAUEL, L.: DNA (cell number) and protein in neonatal rat brain. Alteration by timing of maternal dietary protein restriction. *J. Nutr.* 101: 1265 (1971).
- 26 ZAMENHOF, S.; VAN MARTHENS, E., and GRAUEL, L.: DNA (cell number) in neonatal brain. Alteration by maternal dietary caloric restriction. *Nutr. Rep. Int.* 4: 269 (1971).
- 27 ZAMENHOF, S.; VAN MARTHENS, E., and GRAUEL, L.: Prenatal cerebral development. Effect of restricted diet, reversal by growth hormone. *Science* 174: 954 (1971).
- 28 ZAMENHOF, S.; VAN MARTHENS, E., and MARGOLIS, F.L.: DNA (cell number) and protein in neonatal brain. Alteration by maternal dietary protein restriction. *Science* 160: 322 (1968).
- 29 ZEMAN, F.J.: Effect on the young rat of maternal protein restriction. *J. Nutr.* 93: 167 (1967).
- 30 ZEMAN, F.J.: Effect of maternal protein restriction on the kidney of the newborn young of rats. *J. Nutr.* 94: 111 (1968).
- 31 ZEMAN, F.J. and STANBROUGH, E.C.: Effect of maternal protein deficiency on cellular development in the fetal rat. *J. Nutr.* 99: 274 (1969).

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