

Barriers to replicating preclinical cancer biology research challenges or opportunities?

Tim Errington

Center for Open Science

<https://cos.io/>



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Ventures

Robert Wood Johnson
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Religion Trust

TEMPLETON
World Charity Foundation



Mission: Improve
openness, integrity, and
reproducibility of
scientific research

Norms

Communality

Open sharing

Universalism

Evaluate research on own merit

Disinterestedness

Motivated by knowledge and discovery

Organized skepticism

Consider all new evidence, even against one's prior work

Quality

Counternorms

Secrecy

Closed

Particularism

Evaluate research by reputation

Self-interestedness

Treat science as a competition

Organized dogmatism

Invest career promoting one's own theories, findings

Quantity

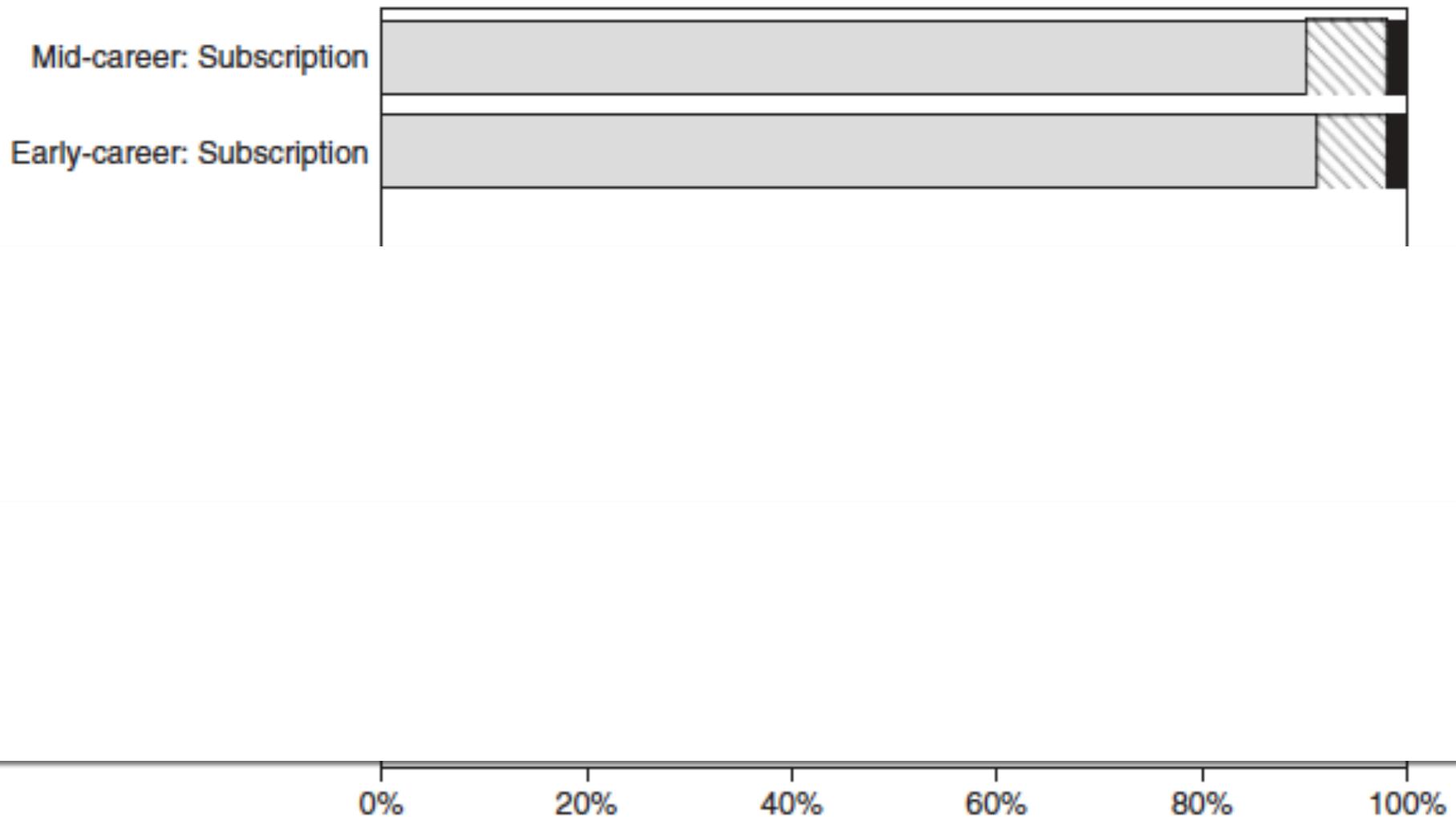


FIG. 3. Norm versus Counternorm Scores: Percent with Norm > Counternorm (dotted), Norm = Counternorm (striped), Norm < Counternorm (solid).

Barriers

1. Perceived norms (Anderson, Martinson, & DeVries, 2007)
2. Motivated reasoning (Kunda, 1990)
3. Minimal accountability (Lerner & Tetlock, 1999)
4. Concrete rewards beat abstract principles (Trope & Liberman, 2010)
5. I am busy (Me & You, 2023)
6. Incentives for individual success are focused on getting it published, not getting it right (Nosek, Spies, & Motyl, 2012)

Motivation

Believe it or not: how much can we rely on published data on potential drug targets?

Florian Prinz, Thomas Schlange and Khusru Asadullah

Raise standards for preclinical cancer research

C. Glenn Begley and Lee M. Ellis propose how methods, publications and incentives must change if patients are to benefit.

Motivation

Believe it or not: how much can we rely on published data on potential drug targets?

Florian Prinz, Thomas Schlange and Khusru Asadullah

We received input from 23 scientists (heads of laboratories) and collected data from 67 projects, most of them (47) from the field of oncology. This analysis revealed that only in ~20–25% of the projects were the relevant published data completely in line with our in-house findings (FIG. 1c). In almost two-thirds

Raise standards for preclinical cancer research

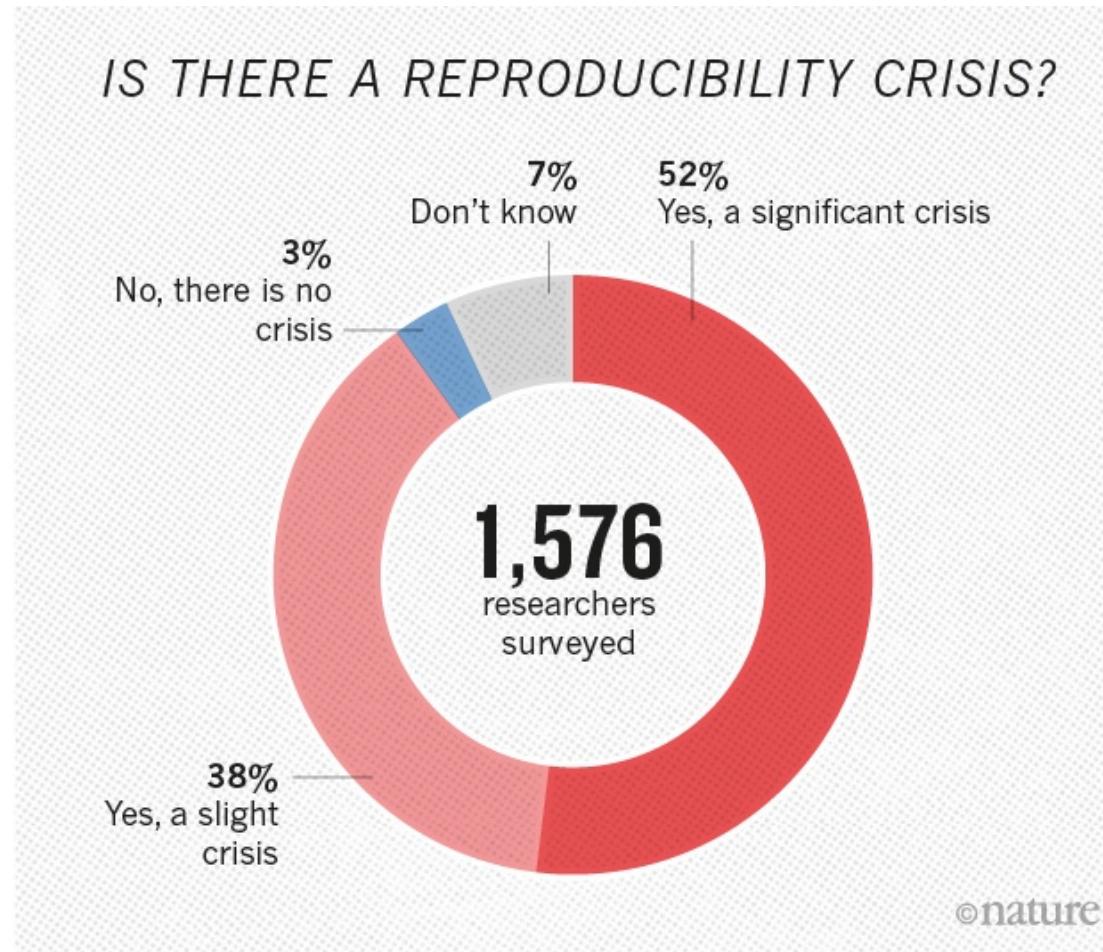
C. Glenn Begley and Lee M. Ellis propose how methods, publications and incentives must change if patients are to benefit.

tive clinical uses for existing therapeutics. Nevertheless, scientific findings were confirmed in only 6 (11%) cases. Even knowing the limitations of preclinical research, this was a shocking result.

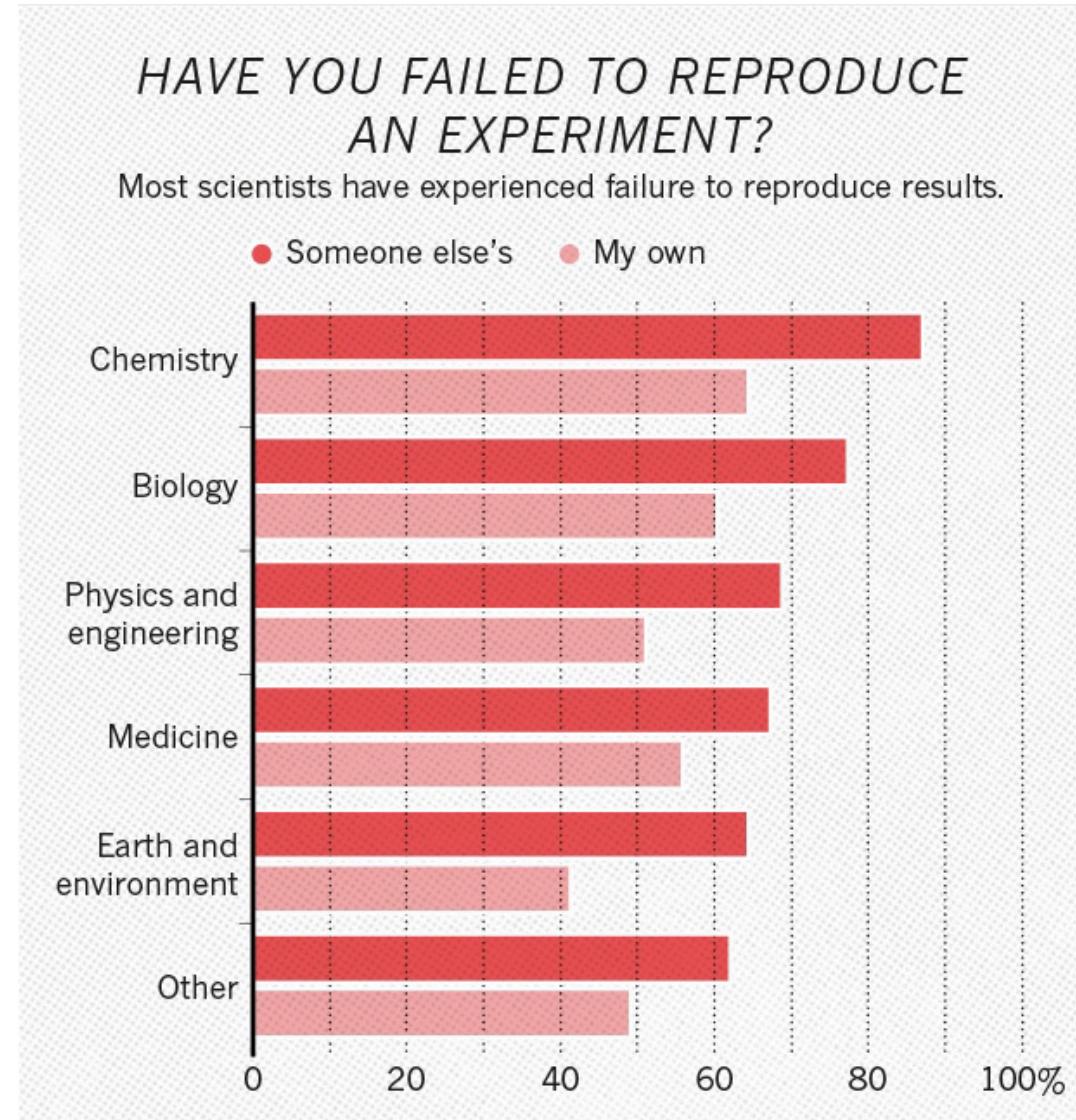
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Motivation

- Nature survey of 1,576 researchers



Motivation



Approach



eLIFE

About Careers



REPRODUCIBILITY PROJECT
Cancer Biology

The Reproducibility Project: Cancer Biology is a collaboration between the [Center for Open Science](#) and [Science Exchange](#) to independently replicate selected results from a substantial number of [high-profile papers](#) in the field of cancer biology. For each paper a Registered Report detailing the proposed experimental designs and protocols for the replications is peer reviewed and published prior to data collection. The results of these experiments will be published in a Replication Study. [The project](#) will provide evidence about reproducibility in cancer biology, and an opportunity to identify factors that influence reproducibility more generally.

Approach

Systematic sampling of 53 high-impact preclinical cancer biology papers

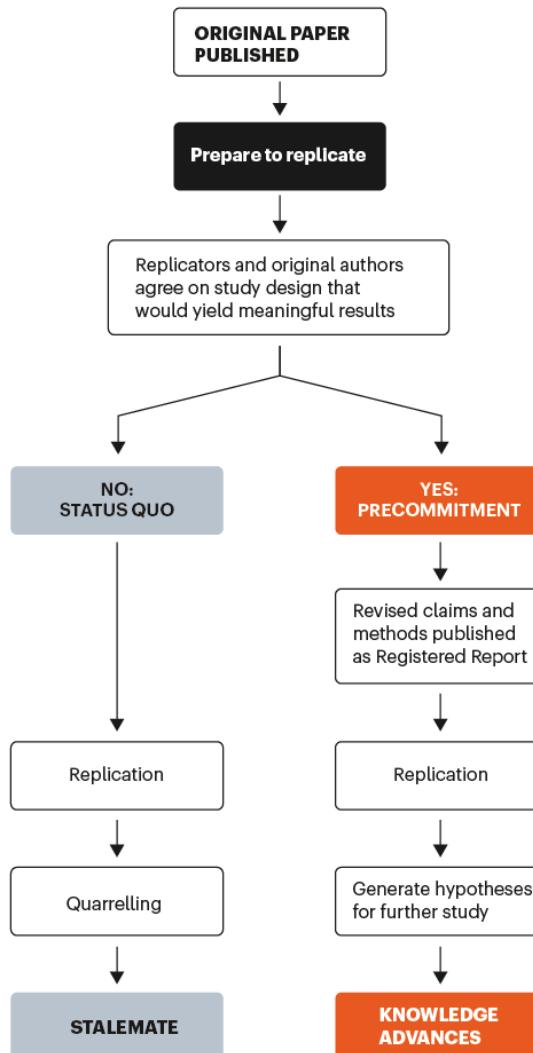
- Papers from 2010-2012
- Excluded genomics, proteomics, high-throughput assays

Primary Outcomes

- Summary of process challenges for testing replicability
- Meta-analytic summary of statistical outcomes

ROUTES TO REPLICATION

Precommitment rewards authors for providing clear, testable claims and helps to advance knowledge.

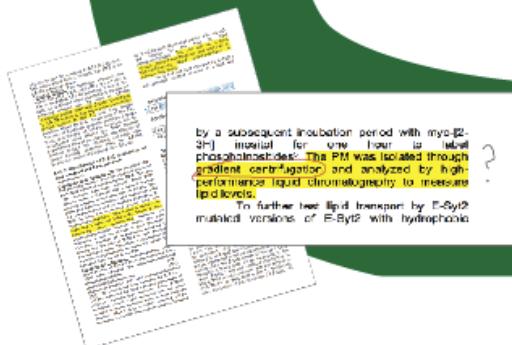


Approach

1. Identify key conclusions from the paper and a subset of data from the paper to be replicated



2. Obtain the materials and methods from the key figures of the paper and annotate them to determine the holes in the protocols that would influence the success of replication



4. Compile obtained information into organized registered report that is published at eLife and undergoes a review process, open to commentary from study authors.



5. Perform experiments, *in vitro* and *in vivo*, according to protocol from registered report.



6. Publish findings in an open access Replication Study



3. Contact laboratories to get them to fill in the holes. This often involves searching through old research notebooks, connecting with study authors that have since moved on, and interfacing with collaborators.



Challenges & Outcomes



FEATURE ARTICLE



RESEARCH ARTICLE



REPRODUCIBILITY IN CANCER BIOLOGY

Challenges for assessing replicability in preclinical cancer biology

Abstract We conducted the [Reproducibility Project: Cancer Biology](#) to investigate the replicability of preclinical research in cancer biology. The initial aim of the project was to repeat 193 experiments from 53 high-impact papers, using an approach in which the experimental protocols and plans for data analysis had to be peer reviewed and accepted for publication before experimental work could begin. However, the various barriers and challenges we encountered while designing and conducting the experiments meant that we were only able to repeat 50 experiments from 23 papers. Here we report these barriers and challenges. First, many original papers failed to report key descriptive and inferential statistics: the data needed to compute effect sizes and conduct power analyses was publicly accessible for just 4 of 193 experiments. Moreover, despite contacting the authors of the original papers, we were unable to obtain these data for 68% of the experiments. Second, none of the 193 experiments were described in sufficient detail in the original paper to enable us to design protocols to repeat the experiments, so we had to seek clarifications from the original authors. While authors were *extremely* or *very helpful* for 41% of experiments, they were *minimally helpful* for 9% of experiments, and *not at all helpful* (or did not respond to us) for 32% of experiments. Third, once experimental work started, 67% of the peer-reviewed protocols required modifications to complete the research and just 41% of those modifications could be implemented. Cumulatively, these three factors limited the number of experiments that could be repeated. This experience draws attention to a basic and fundamental concern about replication – it is hard to assess whether reported findings are credible.

**TIMOTHY M ERRINGTON*, ALEXANDRIA DENIS†, NICOLE PERFITO‡,
ELIZABETH IORNS AND BRIAN A NOSEK**

Investigating the replicability of preclinical cancer biology

Timothy M Errington^{1*}, Maya Mathur², Courtney K Soderberg¹, Alexandria Denis^{1†}, Nicole Perfito^{1‡}, Elizabeth Iorns³, Brian A Nosek^{1,4}

¹Center for Open Science, Charlottesville, United States; ²Quantitative Sciences Unit, Stanford University, Stanford, United States; ³Science Exchange, Palo Alto, United States; ⁴University of Virginia, Charlottesville, United States

Abstract Replicability is an important feature of scientific research, but aspects of contemporary research culture, such as an emphasis on novelty, can make replicability seem less important than it should be. The [Reproducibility Project: Cancer Biology](#) was set up to provide evidence about the replicability of preclinical research in cancer biology by repeating selected experiments from high-impact papers. A total of 50 experiments from 23 papers were repeated, generating data about the replicability of a total of 158 effects. Most of the original effects were positive effects (136), with the rest being null effects (22). A majority of the original effect sizes were reported as numerical values (117), with the rest being reported as representative images (41). We employed seven methods to assess replicability, and some of these methods were not suitable for all the effects in our sample. One method compared effect sizes: for positive effects, the median effect size in the replications was 85% smaller than the median effect size in the original experiments, and 92% of replication effect sizes were smaller than the original. The other methods were binary – the replication was either a success or a failure – and five of these methods could be used to assess both positive and null effects when effect sizes were reported as numerical values. For positive effects, 40% of replications (39/97) succeeded according to three or more of these five methods, and for null effects 80% of replications (12/15) were successful on this basis; combining positive and null effects, the success rate was 46% (51/112). A successful replication does not definitively confirm an original finding or its theoretical interpretation. Equally, a failure to replicate does not disconfirm a finding, but it does suggest that additional investigation is needed to establish its reliability.

Challenges



FEATURE ARTICLE



RESEARCH ARTICLE



REPRODUCIBILITY IN CANCER BIOLOGY

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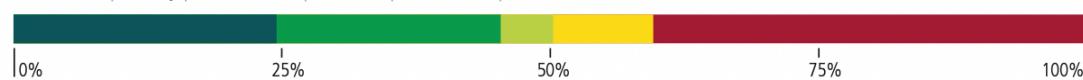
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Challenges

DESIGNED
193 experiments

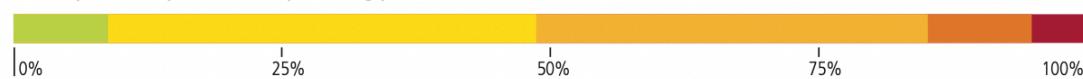
Authors helped

■ Extreme | ■ Very | ■ Moderate | ■ Some | ■ Minimal | ■ No



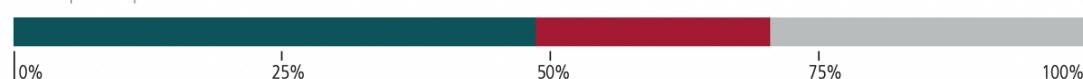
Protocol clarifications needed

■ Few | ■ Some | ■ Moderate | ■ Strong | ■ Extreme



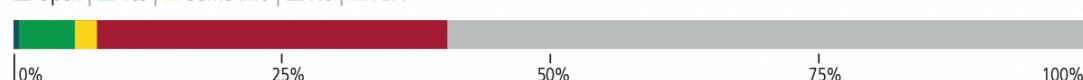
Reagents offered

■ Yes | ■ No | ■ N/A



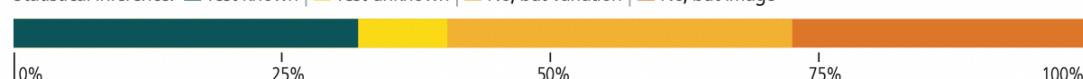
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■ Open | ■ Yes | ■ Some info | ■ No | ■ N/A



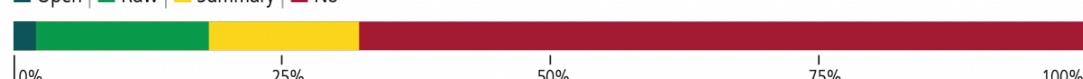
Analysis reported

Statistical inference: ■ Test known | ■ Test unknown | ■ No, but variation | ■ No, but image



Data shared

■ Open | ■ Raw | ■ Summary | ■ No



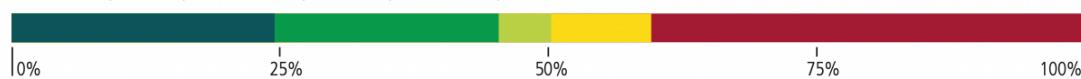
Challenges

DESIGNED
193 experiments

2% had open data; after requests 16%
shared raw data

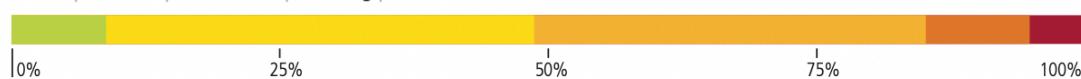
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■ Extreme | ■ Very | ■ Moderate | ■ Some | ■ Minimal | ■ No



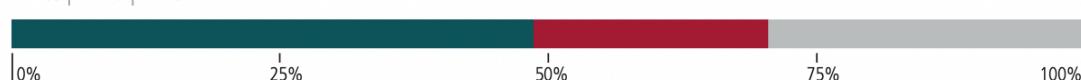
Protocol clarifications needed

■ Few | ■ Some | ■ Moderate | ■ Strong | ■ Extreme



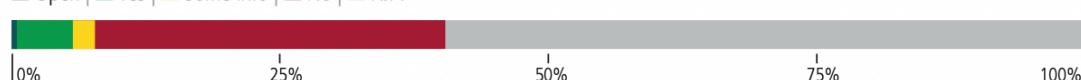
Reagents offered

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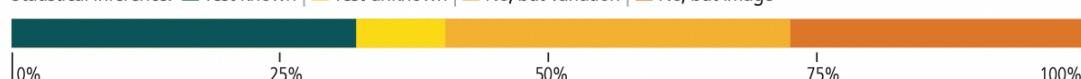
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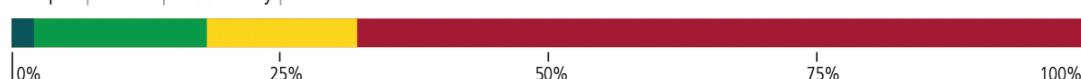
Analysis reported

Statistical inference: ■ Test known | ■ Test unknown | ■ No, but variation | ■ No, but image



Data shared

■ Open | ■ Raw | ■ Summary | ■ No



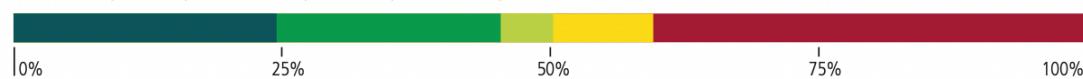
Challenges

DESIGNED
193 experiments

21% had unclear statistical tests

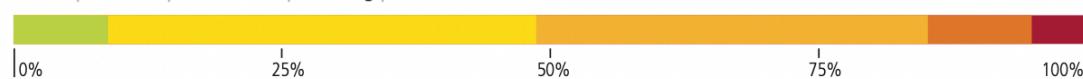
Authors helped

■ Extreme | ■ Very | ■ Moderate | ■ Some | ■ Minimal | ■ No



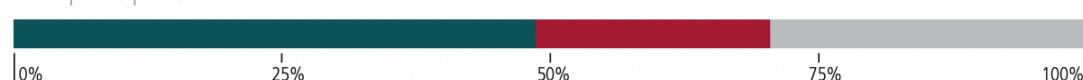
Protocol clarifications needed

■ Few | ■ Some | ■ Moderate | ■ Strong | ■ Extreme



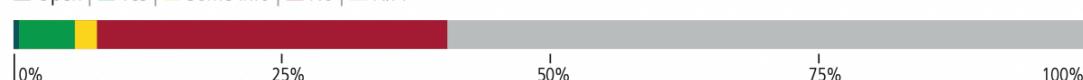
Reagents offered

■ Yes | ■ No | ■ N/A



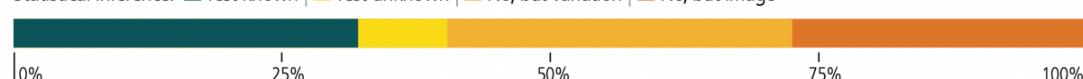
Code shared

■ Open | ■ Yes | ■ Some info | ■ No | ■ N/A



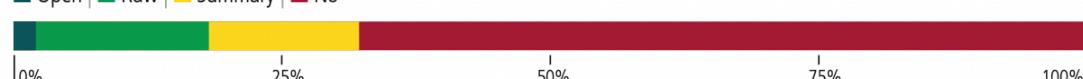
Analysis reported

Statistical inference: ■ Test known | ■ Test unknown | ■ No, but variation | ■ No, but image



Data shared

■ Open | ■ Raw | ■ Summary | ■ No



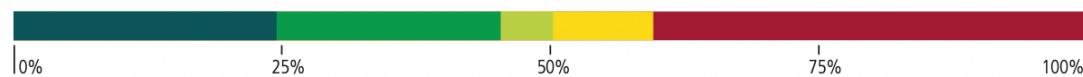
Challenges

DESIGNED
193 experiments

1% had open code; after requests 13%
shared code

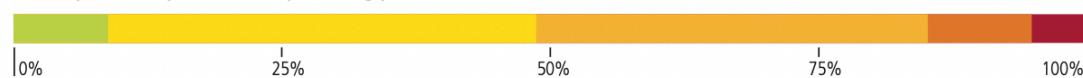
Authors helped

■ Extreme | ■ Very | ■ Moderate | ■ Some | ■ Minimal | ■ No



Protocol clarifications needed

■ Few | ■ Some | ■ Moderate | ■ Strong | ■ Extreme



Reagents offered

■ Yes | ■ No | ■ N/A



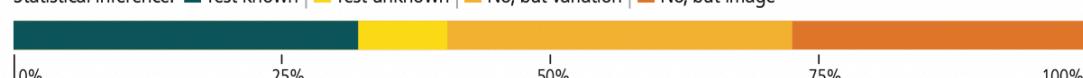
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■ Open | ■ Yes | ■ Some info | ■ No | ■ N/A



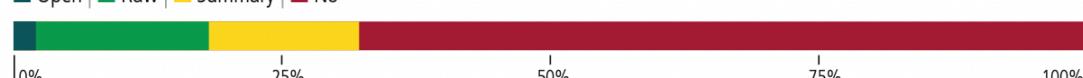
Analysis reported

Statistical inference: ■ Test known | ■ Test unknown | ■ No, but variation | ■ No, but image



Data shared

■ Open | ■ Raw | ■ Summary | ■ No



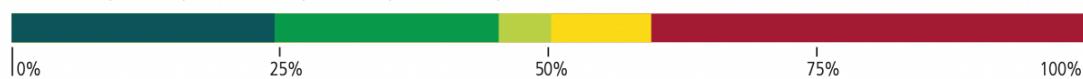
Challenges

DESIGNED
193 experiments

70% required asking for key reagents;
69% willing to share

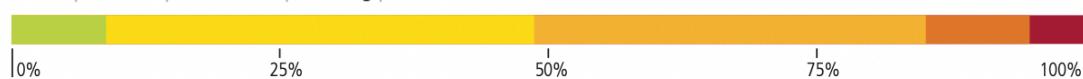
Authors helped

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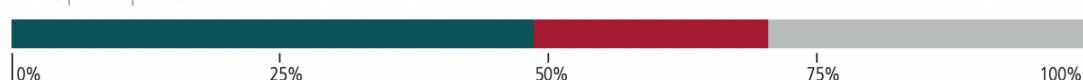
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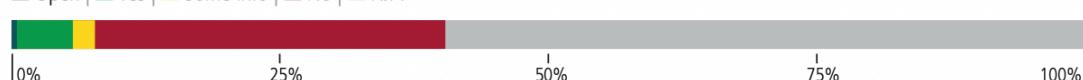
Reagents offered

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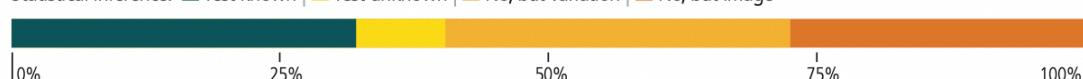
Code shared

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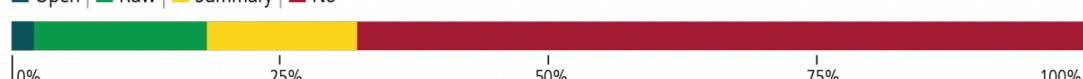
Analysis reported

Statistical inference: ■ Test known | ■ Test unknown | ■ No, but variation | ■ No, but image



Data shared

■ Open | ■ Raw | ■ Summary | ■ No



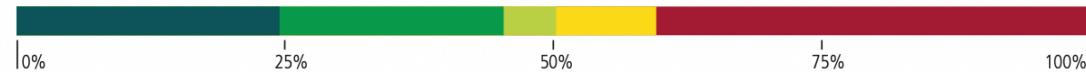
Challenges

DESIGNED
193 experiments

All needed clarifications with 49%
few/some; 20% strong/extreme

Authors helped

■ Extreme | ■ Very | ■ Moderate | ■ Some | ■ Minimal | ■ No



Protocol clarifications needed

■ Few | ■ Some | ■ Moderate | ■ Strong | ■ Extreme



Reagents offered

■ Yes | ■ No | ■ N/A



Code shared

■ Open | ■ Yes | ■ Some info | ■ No | ■ N/A



Analysis reported

Statistical inference: ■ Test known | ■ Test unknown | ■ No, but variation | ■ No, but image



Data shared

■ Open | ■ Raw | ■ Summary | ■ No



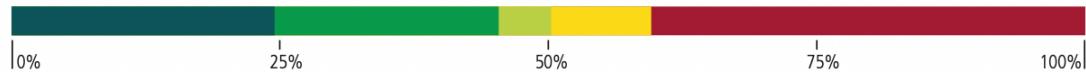
Challenges

DESIGNED
193 experiments

41% extremely/very helpful, 32% not at all helpful/no response

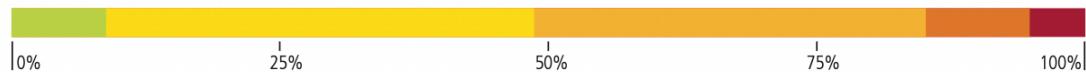
Authors helped

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Protocol clarifications needed

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Reagents offered

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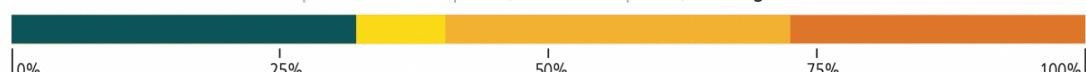
Code shared

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Analysis reported

Statistical inference: ■ Test known | ■ Test unknown | ■ No, but variation | ■ No, but image



Data shared

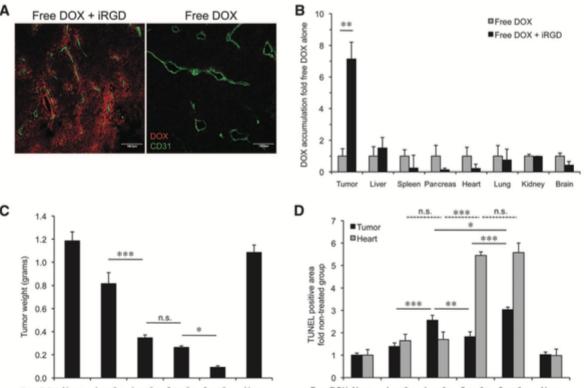
■ Open | ■ Raw | ■ Summary | ■ No



Challenges

From the publication

Fig. 2. Enhanced antitumor effect of free DOX co-injected with iRGD. (A and B) Mice bearing orthotopic 22Rv1 human prostate tumors were intra-venously injected with a mixture of DOX (10 mg/kg) and 4 μ mol/kg of iRGD or PBS. Tumors and tissues were collected 1 hour later, $n = 3$ per group. In (A), the tumors were sectioned and stained for blood vessels with anti-CD31, and the native fluorescence was used to detect DOX. Scale bars, 100 μ m. In (B), DOX in the tissues was quantified. (C) Mice bearing orthotopic 22Rv1 tumors implanted 2 weeks earlier received intravenous injections of DOX (1 or 3 mg/kg) or PBS, combined with 4 μ mol/kg of iRGD or PBS, every other day. The tumors were harvested and weighed after 24 days of treatment, $n = 10$ per group. (D) TUNEL staining was performed on tumors and hearts from the treatment study and was quantified for positive staining. Statistical analyses were performed with Student's t test (B) and ANOVA (C and D). Error bars denote mean \pm SEM. n.s., not significant; * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.



Materials and Methods

In vivo skin permeability assay was performed as described elsewhere (6).

Immunofluorescence. Tissue preparation and staining of the cryosections were performed as described (3). The primary antibodies were rat anti-mouse CD31 (BD Biosciences), FITC-labeled HLA-A,B,C (BD Biosciences), and FITC-labeled H-2kd (BD Biosciences) monoclonal, and rabbit anti-T⁺ phage polyclonal (6) antibodies. The secondary antibodies, Alexa Fluor 594 goat anti-rat, 647 goat anti-rat, and 488 donkey anti-rabbit antibodies were from Molecular Probes.

Immunohistochemistry. Tissue preparation and staining of the cryosections were performed as described (3). The primary antibodies used were biotinylated rabbit anti-FITC/Oregon green polyclonal (Molecular Probes), and biotinylated mouse anti-dextran monoclonal (Stemcell Technologies, Vancouver, BC, Canada) and biotinylated rat anti-mouse CD31 monoclonal (BD Biosciences) antibodies. Biotinylated secondary polyclonal antibodies were goat anti-rabbit and rabbit anti-human (both from Pierce Biotechnology, Rockford, IL). In some experiments, tissue sections were stained with a TUNEL assay kit (In Situ Cell Death Detection Kit, POD; Roche Applied Science, Indianapolis, IN), and quantified for the positive areas with a scanner as described elsewhere in this manuscript.

Ex vivo tumor penetration assay. PPC-1 subcutaneous tumors (about 1 cm in diameter) were excised and placed in DMEM containing 1% BSA. The tumors were first incubated with the inhibitors or peptides for 20 min at 4°C. G₇ or iRGD phage were then added to the solution and the tumors were further incubated for 90 min at 37°C or 4°C. The tumors were then washed with cold DMEM containing 1% BSA, fixed in 4% paraformaldehyde, sectioned, immunofluorescently stained, and viewed under a confocal microscope.

Quantification of ABX in tumors and tissues was performed as described elsewhere (3).

Test conditions:

- Orthotopic tumor bearing mice treated with Dox in combination with iRGD

Steps:

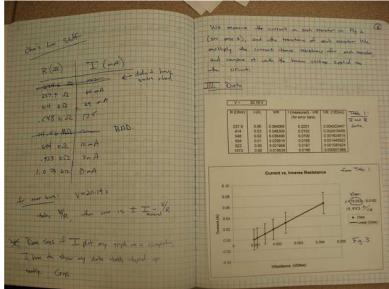
- Generate tumor bearing mice as per Protocol 1.
- Allow tumors to grow for two weeks.
- Inject mice with drug combination:
 - Negative control: Inject mice intravenously with 10 mg/kg Doxorubicin (Dox) (Doxorubicin hydrochloride, Sigma-Aldrich Cat #????) (suspended in 100 μ L PBS) and with 100 μ L PBS.
 - Can the authors please provide the catalog number of the Dox purchased from Sigma?
 - Negative control: Inject mice intravenously with 10 mg/kg Dox (suspended in 100 μ L PBS) and (7) RDSfK (suspended in 100 μ L PBS).
 - Experimental: Inject mice intravenously with 10 mg/kg Dox (suspended in 100 μ L PBS) and 4 μ mol/kg iRGD (suspended in 100 μ L PBS).
- One hour later, sacrifice mice and excise tumors:
 - Euthanize mice
 - How were mice euthanized? CO₂ and cervical dislocation?
 - Perfuse with PBS + 1% BSA.
 - Was the PBS + 1% BSA cold? Perfusion was accomplished how – through the left ventricle of the heart?
 - Excise prostate tumor tissue.
- Process, embed and section tumor tissue:
 - Were tumors fixed? If so, in what fixative and for how long? Given that Dox native fluorescence is used later in the protocol, fixation method can affect the signal from the free Dox.
 - Embed tumor tissue in Optimal Cutting Temperature medium (OCT) (purchased from where?) and freeze.
 - How thick were the sections cut from the tumors?
 - Materials and Methods section "Immunofluorescence" and "Immunohistochemistry" state: "Tissue preparation and staining of the cryosections were performed as described (3)." There is no reference 3 in the Supplemental Materials (only References 35-37), while Reference 3 in the main article is a listing of amino acid abbreviations. Can the authors please provide the details of tissue processing and staining?
- Immunostain tumor tissue with rat anti-CD31 (BD Biosciences, ?). Use native fluorescence to image Dox.
 - Can the authors please provide the Catalog number of the anti-CD31 antibody?

From the publication

- What antibodies were used?
- What sex were the mice?
- How long was the treatment period for?
- What instrument/software was used?
- What was the staining protocol used?
- How was positive area defined and measured?
- Can you share the data with us?

Challenges

From the authors



"I think that nuance was lost trying to edit the text down to size."

"I do not have those data with me and will need to dig a bit in my back-ups"



"we have been working with the protocol document. Problem was that different experiments were done by different authors"



"I do not have the raw data anymore"

"some of the details you are seeking are not readily accessible"

Challenges

Barriers to Conducting Replications in Experiments

By research stage

COMPLETED

50 experiments

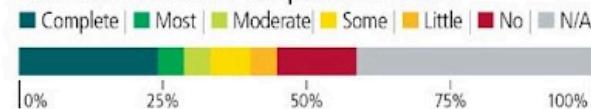
CONDUCTED

87 experiments

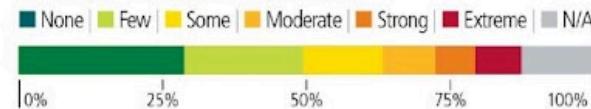


BARRIERS

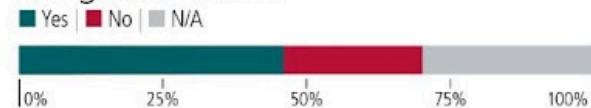
Modifications implemented



Modifications needed



Reagents shared



Challenges

Barriers to Conducting Replications in Experiments

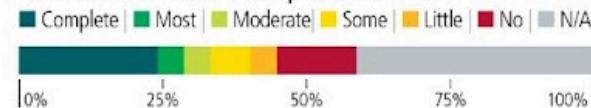
By research stage

COMPLETED

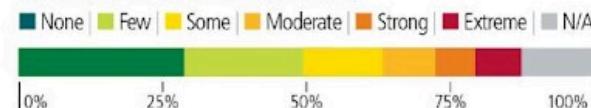
50 experiments

BARRIERS

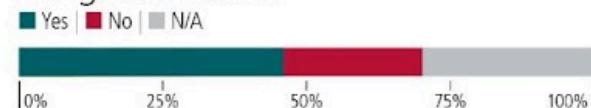
Modifications implemented



Modifications needed



Reagents shared



66% actually shared reagents

CONDUCTED

87 experiments

Barriers to Conducting Replications in Experiments

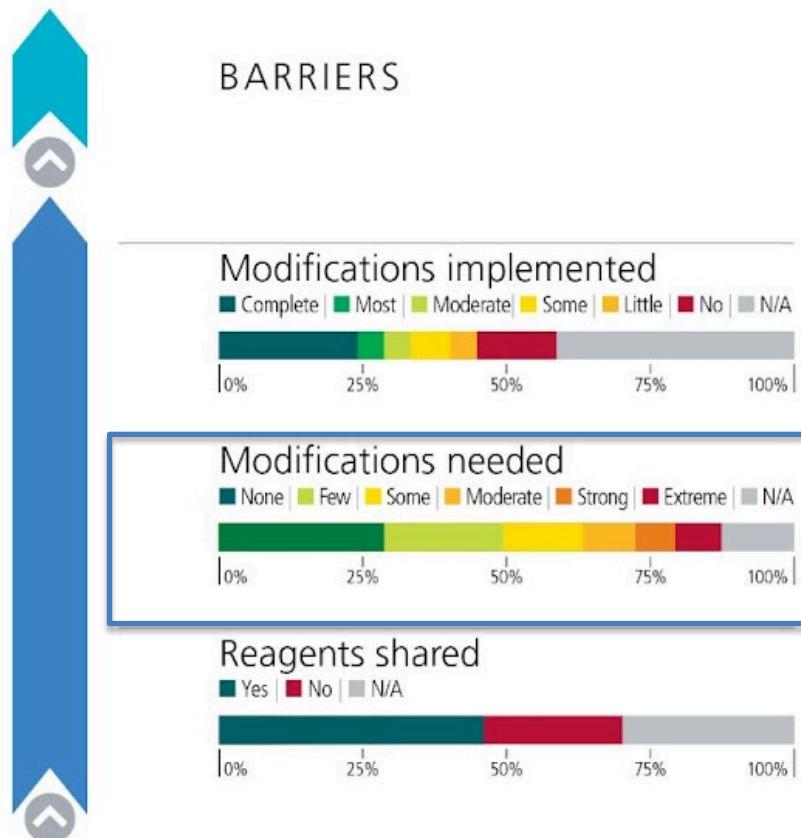
By research stage

COMPLETED

50 experiments

CONDUCTED

87 experiments



67% required modifications with 39% few/some; 17% strong/extreme

Challenges

Barriers to Conducting Replications in Experiments

By research stage

COMPLETED

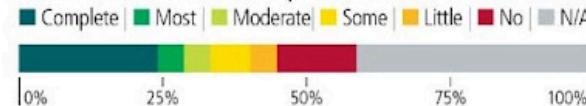
50 experiments

BARRIERS

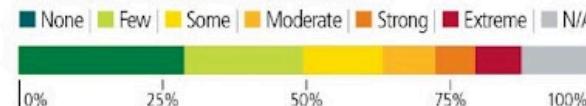
CONDUCTED

87 experiments

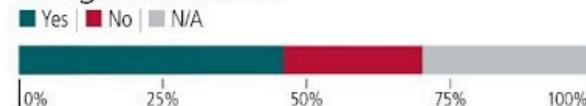
Modifications implemented



Modifications needed



Reagents shared



41% completely implemented

Testing replicability impeded by...

- Lack of transparency and accessibility of methodology
- Lack of sharing of original data and reagents
- Lack of communication for obtaining needed information
- Unexpected challenges with protocols during experimentation
- Resource challenges (cost, time, uncertainty) inflated by all the above

Outcomes



FEATURE ARTICLE



RESEARCH ARTICLE



REPRODUCIBILITY IN CANCER BIOLOGY

Challenges for assessing replicability in preclinical cancer biology

Abstract We conducted the [Reproducibility Project: Cancer Biology](#) to investigate the replicability of preclinical research in cancer biology. The initial aim of the project was to repeat 193 experiments from 53 high-impact papers, using an approach in which the experimental protocols and plans for data analysis had to be peer reviewed and accepted for publication before experimental work could begin. However, the various barriers and challenges we encountered while designing and conducting the experiments meant that we were only able to repeat 50 experiments from 23 papers. Here we report these barriers and challenges. First, many original papers failed to report key descriptive and inferential statistics: the data needed to compute effect sizes and conduct power analyses was publicly accessible for just 4 of 193 experiments. Moreover, despite contacting the authors of the original papers, we were unable to obtain these data for 68% of the experiments. Second, none of the 193 experiments were described in sufficient detail in the original paper to enable us to design protocols to repeat the experiments, so we had to seek clarifications from the original authors. While authors were *extremely* or *very helpful* for 41% of experiments, they were *minimally helpful* for 9% of experiments, and *not at all helpful* (or did not respond to us) for 32% of experiments. Third, once experimental work started, 67% of the peer-reviewed protocols required modifications to complete the research and just 41% of those modifications could be implemented. Cumulatively, these three factors limited the number of experiments that could be repeated. This experience draws attention to a basic and fundamental concern about replication – it is hard to assess whether reported findings are credible.

TIMOTHY M ERRINGTON*, ALEXANDRIA DENIS†, NICOLE PERFITO‡,
ELIZABETH IORNS AND BRIAN A NOSEK

Investigating the replicability of preclinical cancer biology

Timothy M Errington^{1*}, Maya Mathur², Courtney K Soderberg¹,
Alexandria Denis^{1†}, Nicole Perfito^{1‡}, Elizabeth Iorns³, Brian A Nosek^{1,4}

¹Center for Open Science, Charlottesville, United States; ²Quantitative Sciences Unit, Stanford University, Stanford, United States; ³Science Exchange, Palo Alto, United States; ⁴University of Virginia, Charlottesville, United States

Abstract Replicability is an important feature of scientific research, but aspects of contemporary research culture, such as an emphasis on novelty, can make replicability seem less important than it should be. The [Reproducibility Project: Cancer Biology](#) was set up to provide evidence about the replicability of preclinical research in cancer biology by repeating selected experiments from high-impact papers. A total of 50 experiments from 23 papers were repeated, generating data about the replicability of a total of 158 effects. Most of the original effects were positive effects (136), with the rest being null effects (22). A majority of the original effect sizes were reported as numerical values (117), with the rest being reported as representative images (41). We employed seven methods to assess replicability, and some of these methods were not suitable for all the effects in our sample. One method compared effect sizes: for positive effects, the median effect size in the replications was 85% smaller than the median effect size in the original experiments, and 92% of replication effect sizes were smaller than the original. The other methods were binary – the replication was either a success or a failure – and five of these methods could be used to assess both positive and null effects when effect sizes were reported as numerical values. For positive effects, 40% of replications (39/97) succeeded according to three or more of these five methods, and for null effects 80% of replications (12/15) were successful on this basis; combining positive and null effects, the success rate was 46% (51/112). A successful replication does not definitively confirm an original finding or its theoretical interpretation. Equally, a failure to replicate does not disconfirm a finding, but it does suggest that additional investigation is needed to establish its reliability.

How to assess replicability?

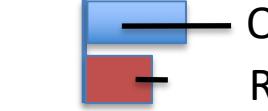
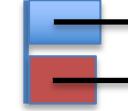
- Same direction



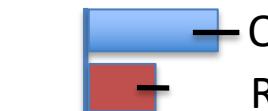
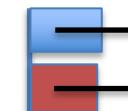
- Same direction and statistically significant



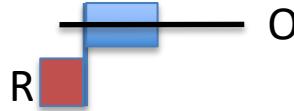
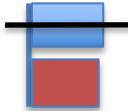
- Original effect size in replication confidence interval



- Replication effect size in original confidence interval



- Replication effect size in prediction interval



- Replication effect size compared to original effect size



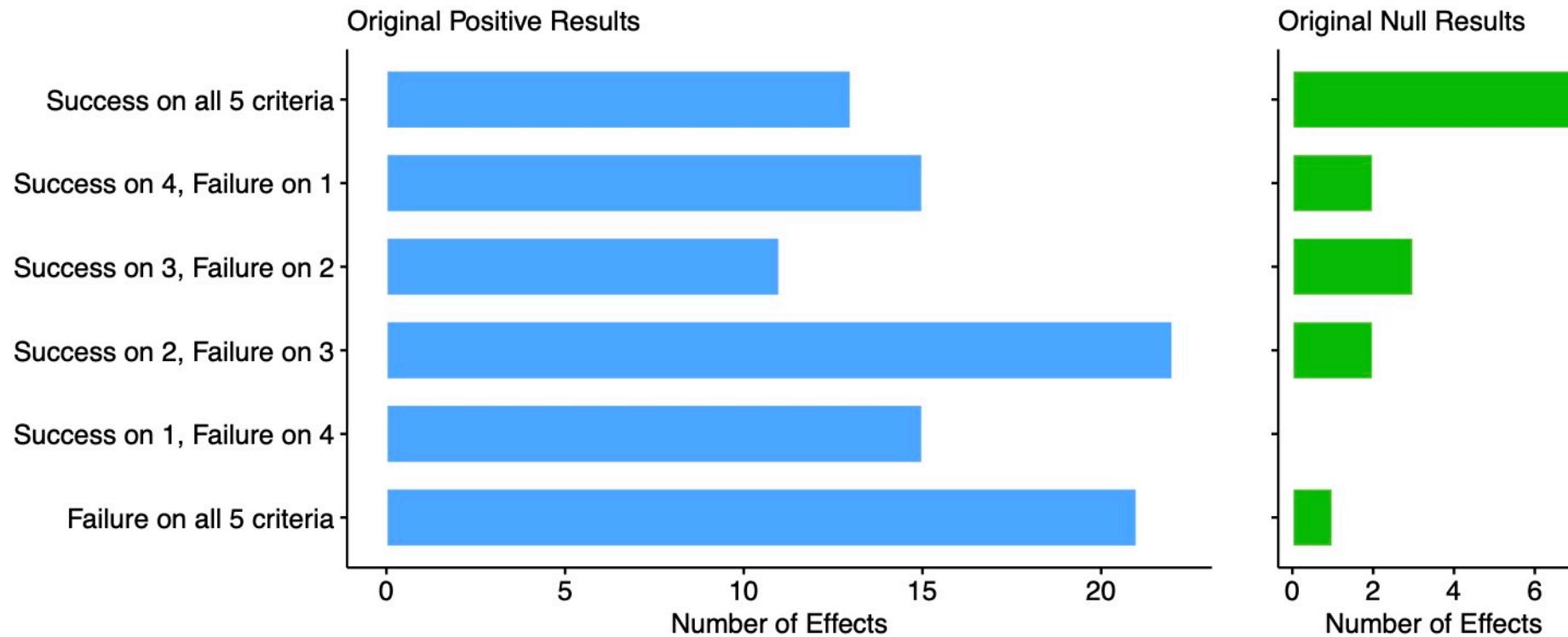
- Direction and statistical significance of meta-analysis



Summarizing across five dichotomous replication success criteria

- 47%: Same direction and statistically significant
- 25%: Original effect size in the replication 95% confidence interval
- 48%: Replication effect size in the original 95% confidence interval
- 61%: Replication effect size in the 95% prediction interval
- 63%: Meta-analysis of original and replication

Outcomes

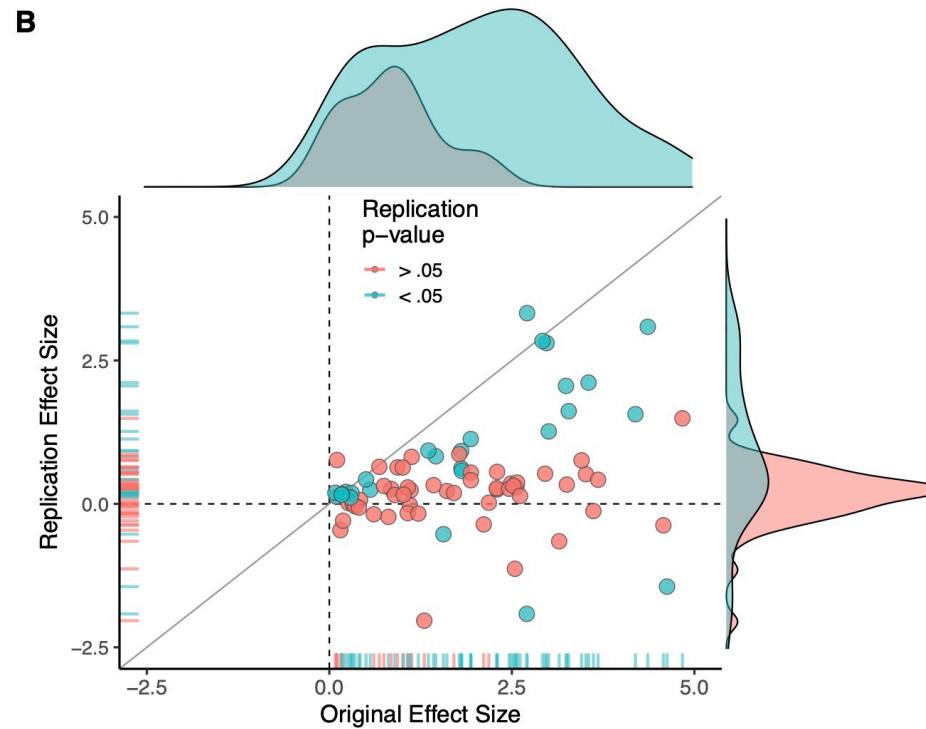
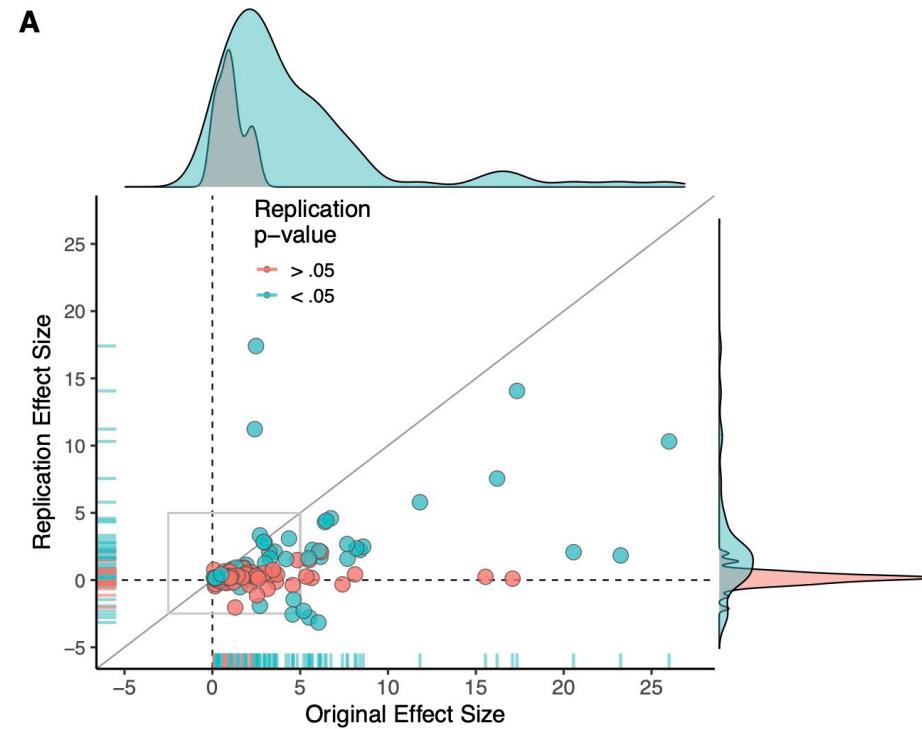


46% succeeded on most criteria, 54% failed on most criteria

80% of original null effects succeeded on most criteria

40% of original positive effects succeeded on most criteria

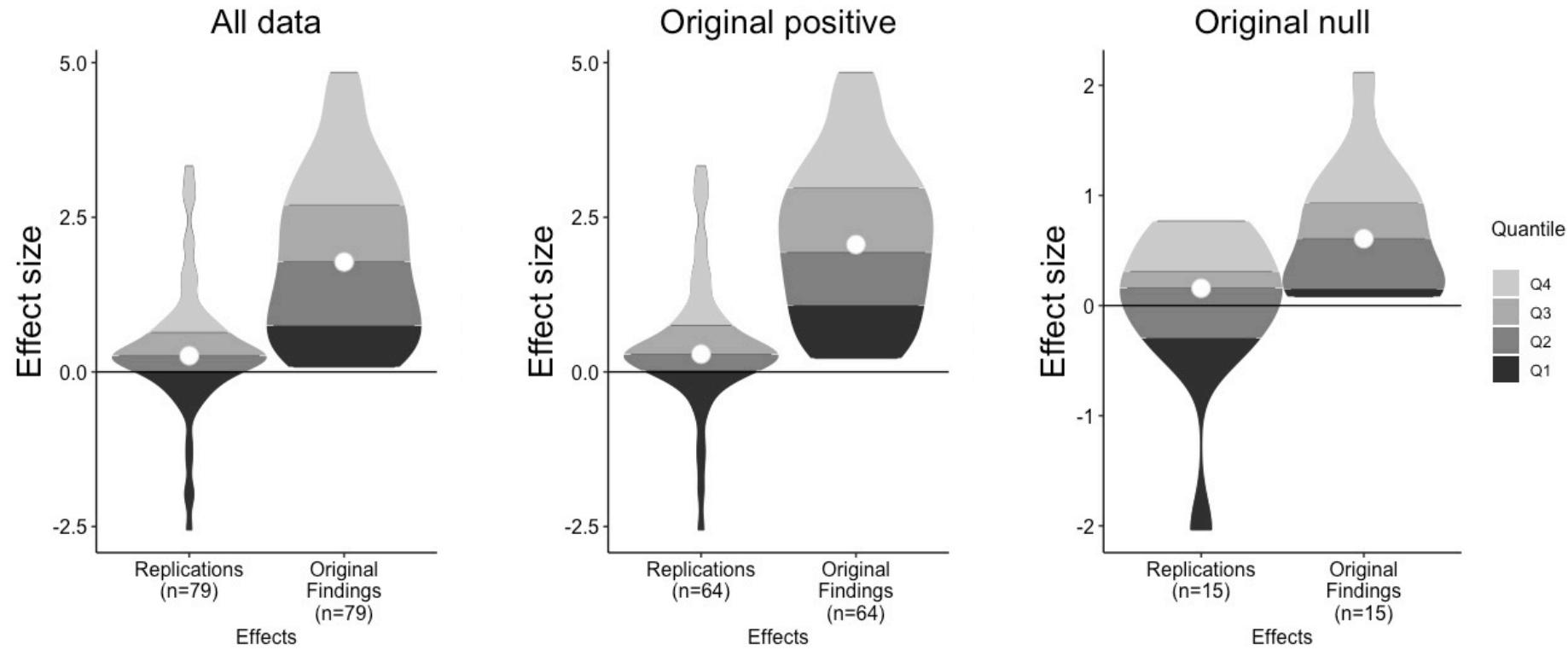
Replication effects compared with original effects



Replications 85% smaller on average

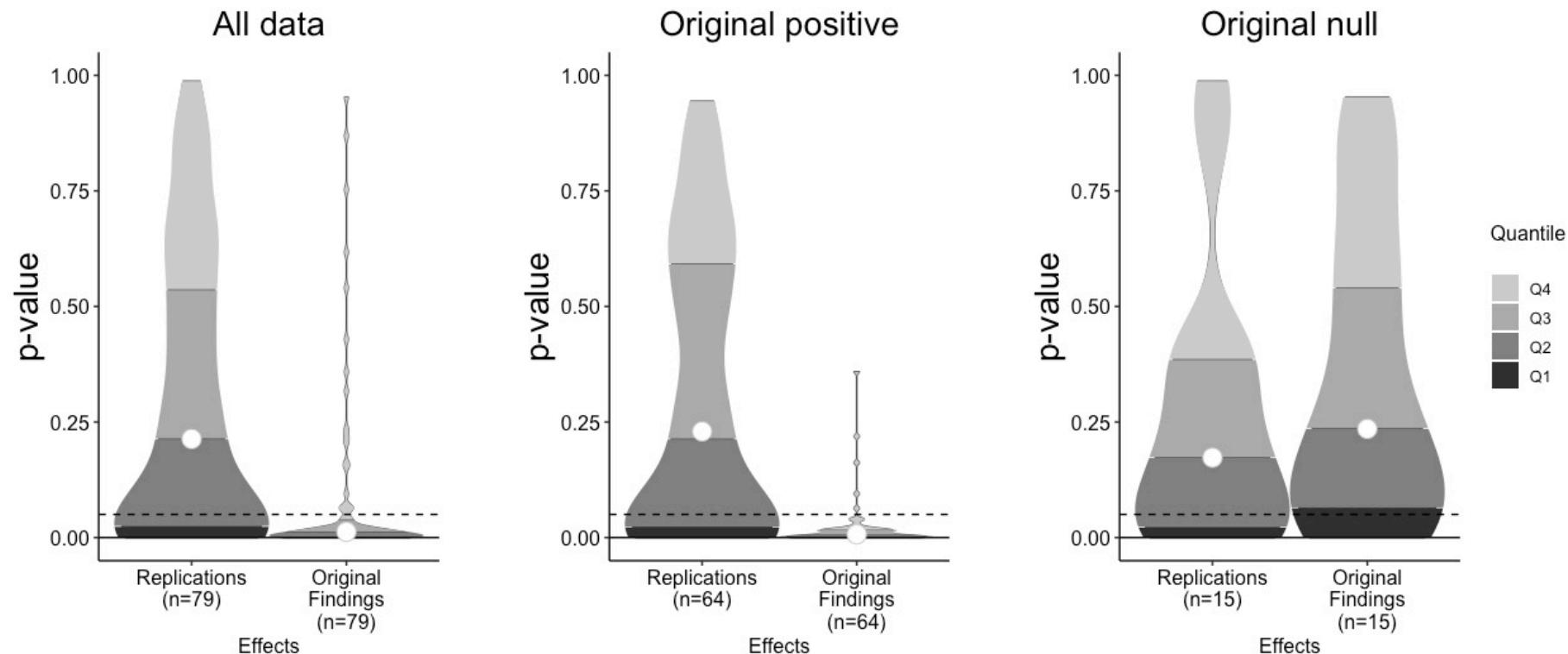
Zoomed in on effect sizes < 5

Replication effects compared with original effects



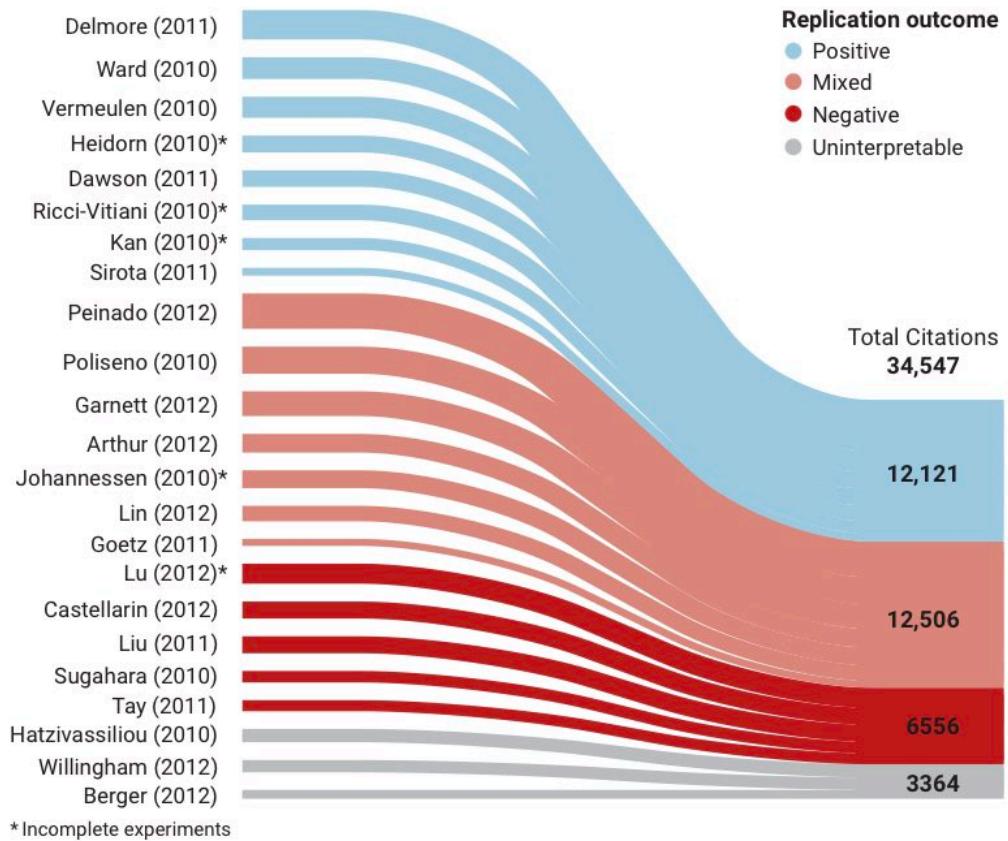
effect sizes <5

Replication p-values compared with original p-values



effect sizes <5

Subjective assessment



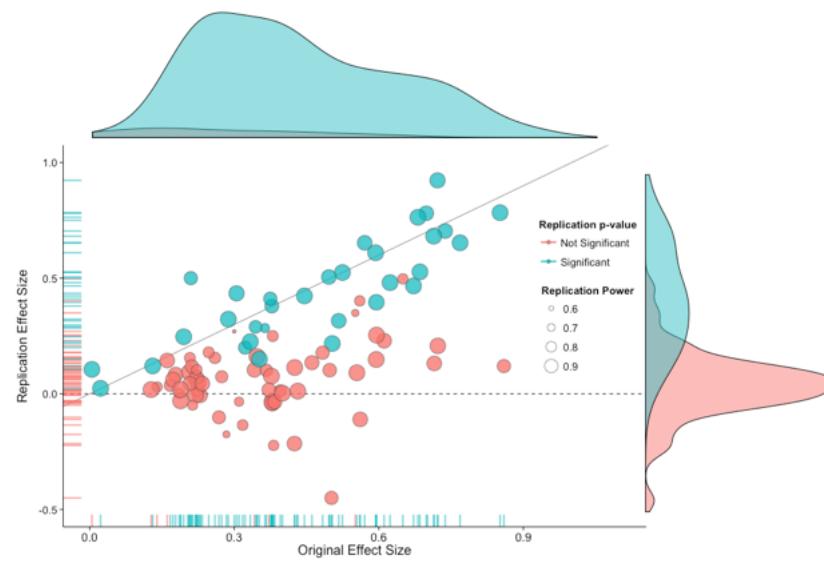
GRAPHIC: K. FRANKLIN/SCIENCE; DATA: REPRODUCIBILITY PROJECT: CANCER
BIOLOGY

Meta-analysis conclusions

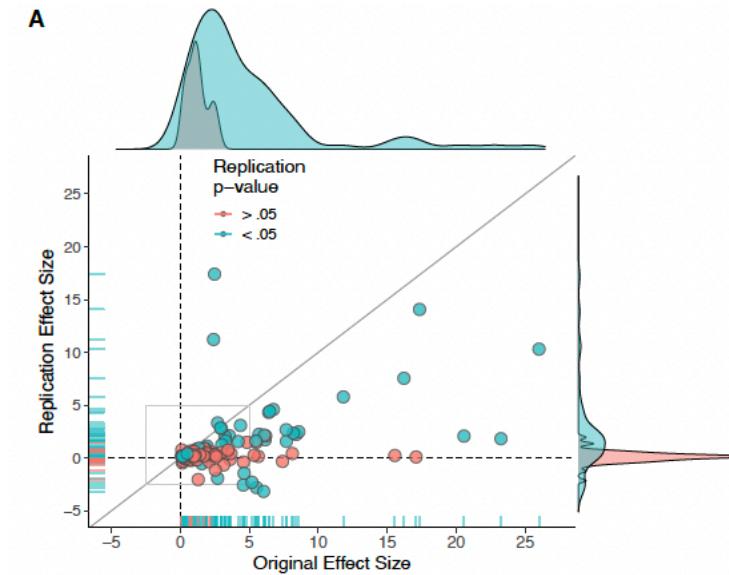
- Replication effects were much weaker than originals
- “Success” was low across replication criteria with variability due, in part to liberalness of the test
- Positive results were half as likely to replicate as null results
- Animal and non-animal declines similar magnitudes - animal effects lower success rate because small original effect sizes
- There is room for improvement

Replication in...

Psychology



Cancer Biology

Open Science Collaboration, 2015 *Science*Replications

36% significant, same direction as original
50% smaller than original on average

Errington et al., 2021 *eLife*Replications

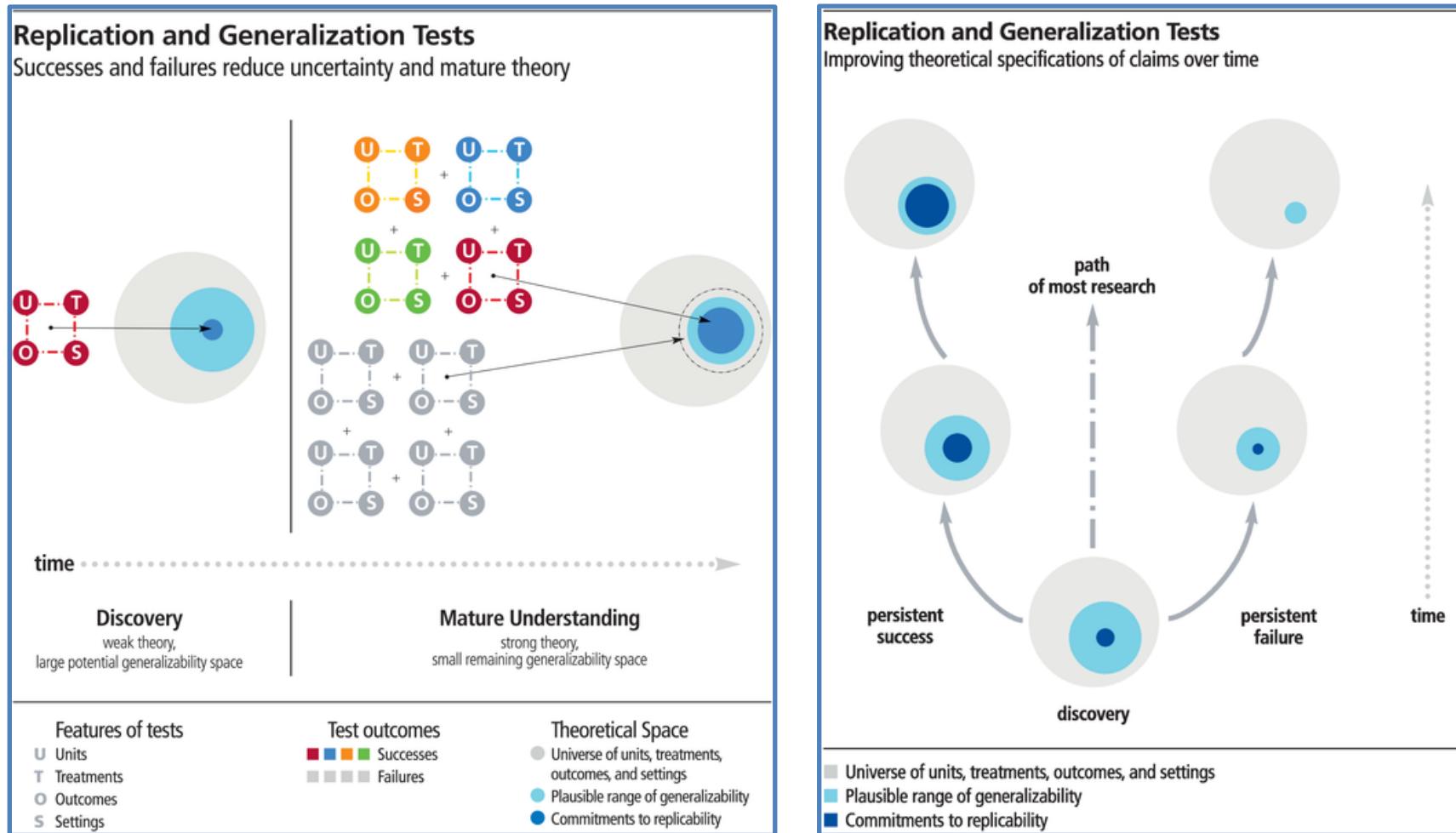
43% significant, same direction as original
85% smaller than original on average

Interpreting failures to replicate

- A failure to replicate could mean:
 - The original finding was a false positive
 - The replication was a false negative
 - Both are “true” and key conditions in the experimental design differ

Conclusions

Do we know the conditions necessary to observe a finding?

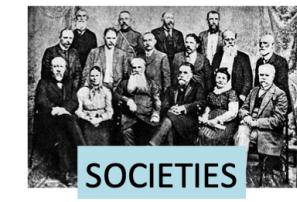


What can we do?

- Incentivize open science practices in your community
 - Aligning institutional policies with open science practices (e.g., [NASEM Roundtable](#))
 - Journal policies that incentivize open practices (e.g., [TOP Guidelines](#))
 - Assessment of researchers and scholarly research (e.g., [DORA](#))
 - Training on reproducible and open science practices



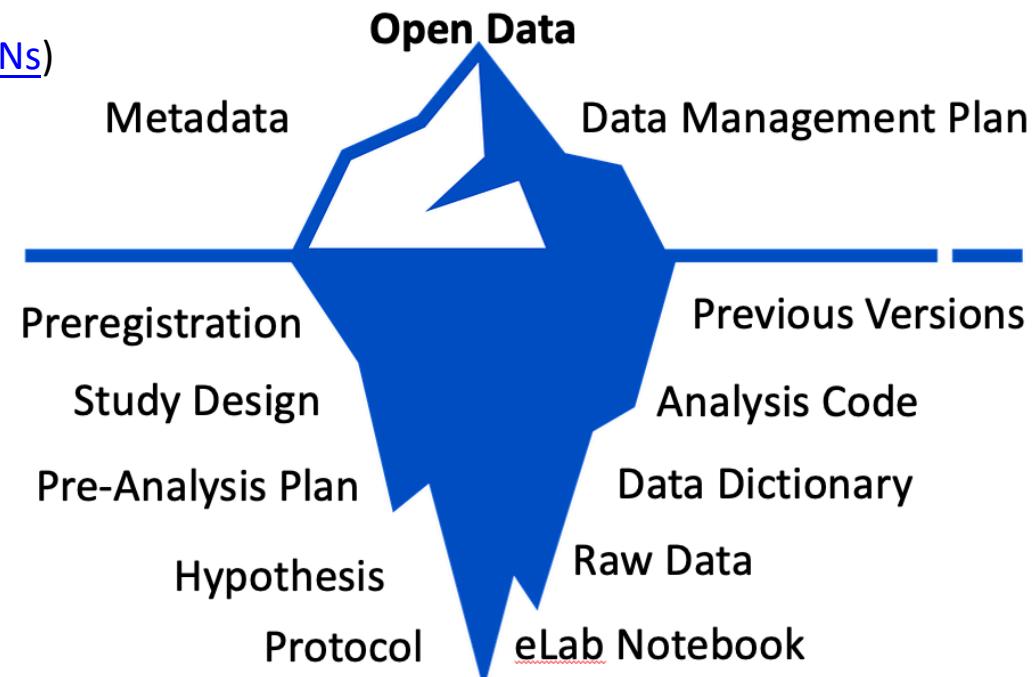
social system



What can we do?

- Incorporate open science practices in your research

- Share data/code/etc using repositories (e.g., [NIH GREI Repositories](#))
- Deposit reagents in repositories (e.g., [addgene](#))
- Make detailed protocols open (e.g., [ELNs](#))
- Try [preregistration/Registered Reports](#)



Conclusions

Study 15: Replication of Sugahara et al., 2010 (Science)

Contributors: Christine Mantis, Irawati Kandela, Fraser Aird, Elizabeth Iorns, Alexandria Denis, Stephen Williams, Nicole Perfito, Tim Errington

Affiliated institutions: Center For Open Science, Laura and John Arnold Foundation

Date created: 2013-10-22 04:11 PM | Last Updated: 2017-01-18 02:44 PM

Identifiers: DOI 10.17605/OSF.IO/XU1G2 | ARK c7605/osf.io/xu1g2

Category: Project

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Wiki

Replication Study: Coadministration of a Tumor-Penetrating Peptide Enhances the Efficacy of Cancer Drugs

Abstract

In 2015, as part of the Reproducibility Project: Cancer Biology, we published a Registered Report (Kandela et al., 2015), that described how we intended to replicate selected experiments from the paper "Coadministration of a tumor-penetrating peptide enhances the efficacy of cancer d..."

[Read More](#)

Files

Name	Modified
Study 15: Replication of Sugahara et al., 2010 (Science)	
- OSF Storage	
Replication_Study_15.docx	2016-10-25 05:39 PM
Replication_Study_15.pdf	2016-10-25 05:39 PM
Replication_Study_15.Rmd	2017-01-11 01:42 PM

Citation

osf.io/xu1g2

Components

- Quantifying the amount of Dox present in tumor tissue and major organs in mice treated with Dox with or without iRGD
- Mantis, Kandela, Aird & 5 more
39 contributions
- Effect of Dox alone or Dox in combination with iRGD on tumor growth and total body weight
- Mantis, Kandela, Aird & 5 more
47 contributions
- Assessment of TUNEL staining of tumor and heart tissue after treatment
- Mantis, Kandela, Aird & 5 more
39 contributions
- Meta-analyses
- Iorns, Denis, Williams & 2 more
117 contributions
- iRGD peptide synthesis
- Chou
63 contributions

Conclusions

Dose administration

Mice were intravenously injected (IV) by tail vein, based on body weight on injection day. For tumor and organ penetrance analysis, [Figure 1](#), mice were anesthetized, perfused, and sacrificed 1 hr after drug administration. For multi-day experiments mice were injected by IV every other day for 24 days based on body weight on injection day. On the last day, mice were anesthetized, perfused, and sacrificed 1 hr after drug administration. Further details of these methods are available at (<https://osf.io/bkhnp/>).

Conclusions

Dose administration

Mice were intravenously injected (IV) by tail vein, based on body weight on injection day. For tumor and organ penetrance analysis, Figure 1, mice were anesthetized, perfused, and sacrificed 1 hr after drug administration. For multi-day experiments mice were injected by IV every other day for 24 days based on body weight on injection day. On the last day, mice were anesthetized, perfused, and sacrificed 1 hr after drug administration. Further details of these methods are available at [\(https://osf.io/bkhnp/\)](https://osf.io/bkhnp/)

Conclusions

Dose administration

Mice were intravenously injected (IV) by tail vein, based on body weight on injection day. For tumor and organ penetrance analysis, Figure 1, mice were anesthetized with isoflurane and injected with 100 µg/kg of doxorubicin.

TUNEL analysis

TUNEL (EMD Millipore, cat # S7100) stained slides were evaluated by Dr. Gennadiy Bondarenko and Dr. Andrey Ugolkov. Images were captured using a Carl Zeiss Axial Lab A1 microscope and a 40x objective, by Dr. Ugolkov, or a Olympus BX45 microscope and a 40x objective, by Dr. Bondarenko (images available at: <https://osf.io/3fs27/>). Drs. Bondarenko and Ugolkov were blinded to group allocation, only receiving the animal ID with H or P designation to indicate heart or prostate tumor. The frequency of apoptosis was calculated as an apoptotic index, in which the proportion of cells undergoing apoptosis was expressed as a percentage of all cells observed. The apoptotic index of each tissue sample was calculated as the number of TUNEL-positive cells and bodies per 500 cells/microscopic view or 2500 cells/slide (5 slides/tissue), counted in five randomly selected microscopic fields in each tissue sample. Percent apoptotic index was calculated with the following formulation: $(i/500) \times 100\%$. i = cell undergoing apoptosis. An average was taken of the apoptotic index from all five fields of all five slices (25 fields total) which is considered one biological replicate. Negative and positive control sections were stained in parallel to the tumor and heart samples and are available at (<https://osf.io/gmcyt/>). Original counts are available at <https://osf.io/pbg7x/>.

Conclusions

Dose administration

Mice were intravenously injected (IV) by tail vein, based on body weight on injection day. For tumor analysis

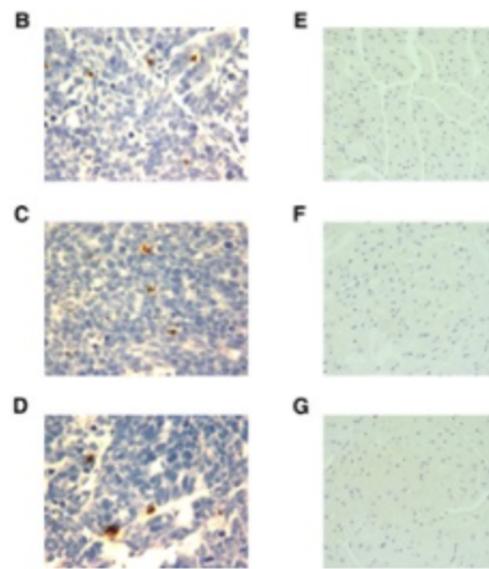
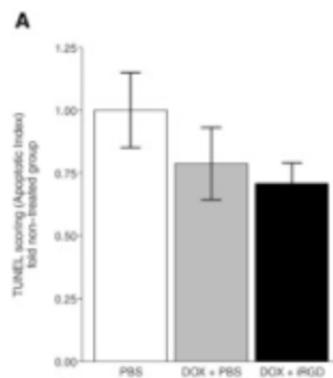
TUNEL analysis

TUNEL (EMD Millipore, catalog number S7114) was performed on tissue sections from Dr. Bondarenko and Dr. Andriantsiferova. Tissue sections were imaged on a Leica A1 microscope and a 40x objective, by Dr. Bondarenko and Ugolkova. Tissue samples were taken with H or P designation to indicate whether they were taken from the tumor or normal tissue. Apoptosis was calculated as an apoptotic index (AI). Apoptosis was expressed as the percentage of apoptotic cells per field. The total number of tissue sample was calculated as the product of the number of cells/microscopic view or selected microscopic fields multiplied by the number of fields with the following formula: AI = (number of apoptotic cells/number of total cells) × 100. The AI was taken of the apoptotic index. The AI is considered one biologic endpoint. The AI was measured in parallel to the tumor analysis. Original counts are available upon request.

Statistical analysis

Statistical analysis was performed with R software (RRID: SCR_001905), version 3.2.3 (R Core Team, 2016). All data csv files and analysis scripts are available at (<https://osf.io/xu1g2/>). Confirmatory statistical analysis was pre-registered (<https://osf.io/9hr2d/>) before the experimental work began as outlined in the Registered Report (Kandela et al., 2015). Additional exploratory analysis (area under the curve) was performed using the weights of the mice over the treatment period. Data were checked to ensure assumptions of statistical tests were met. A meta-analysis of a common original and replication effect size was performed using a random effects model and the *metafor* R package (Viechtbauer, 2010). (available at <https://osf.io/ymxaz/>). The original study data were extracted *a priori* from the published figures by determining the mean and upper/lower error values for each data point. The extracted data were published in the Registered Report (Kandela et al., 2015) and were used in the power calculations to determine the sample sizes for this study. In the meta-analyses where Glass' Δ was used, because of unequal variance between the two conditions being compared, the standard deviation of DOX + PBS was used in the calculations.

Conclusions



Download figure | Open

Figure 4.

TUNEL staining of mouse tissues.

Mice bearing orthotopic 22Rv1 human prostate tumors were intravenously injected with alone (PBS), 1 mg/kg DOX and PBS (DOX + PBS), or 1 mg/kg DOX and 4 μ mol/kg of iRGD (DOX + iRGD). TUNEL staining was performed on tumor and heart sections of each animal. (A) Boxplot of mean apoptotic index calculated from TUNEL stained tumor sections. TUNEL scores were normalized to the average score of tumors from PBS treated mice. Means reported and error bars represent s.e.m. Number of mice per condition ($n=6$; $n=18$ mice total). One-way ANOVA on apoptotic index of all groups: $F(2, 15) = 1.378, p=0.282$. Planned contrast between DOX + PBS and DOX + iRGD: $t(15) = 0.435, p=0.670$ with *a priori* alpha level = 0.05. Representative images of TUNEL staining of tumor sections from PBS (B), DOX + PBS (C), or DOX + iRGD (D) treated mice. Representative images of TUNEL staining of heart sections from PBS (E), DOX + PBS (F), or DOX + iRGD (G) treated mice. Additional details for this experiment can be found at <https://osf.io/7eynw/>.

All RP:CB papers point to the OSF components which host not only the selected representative images, but all images collected, methods, analyzes, and figure generation.

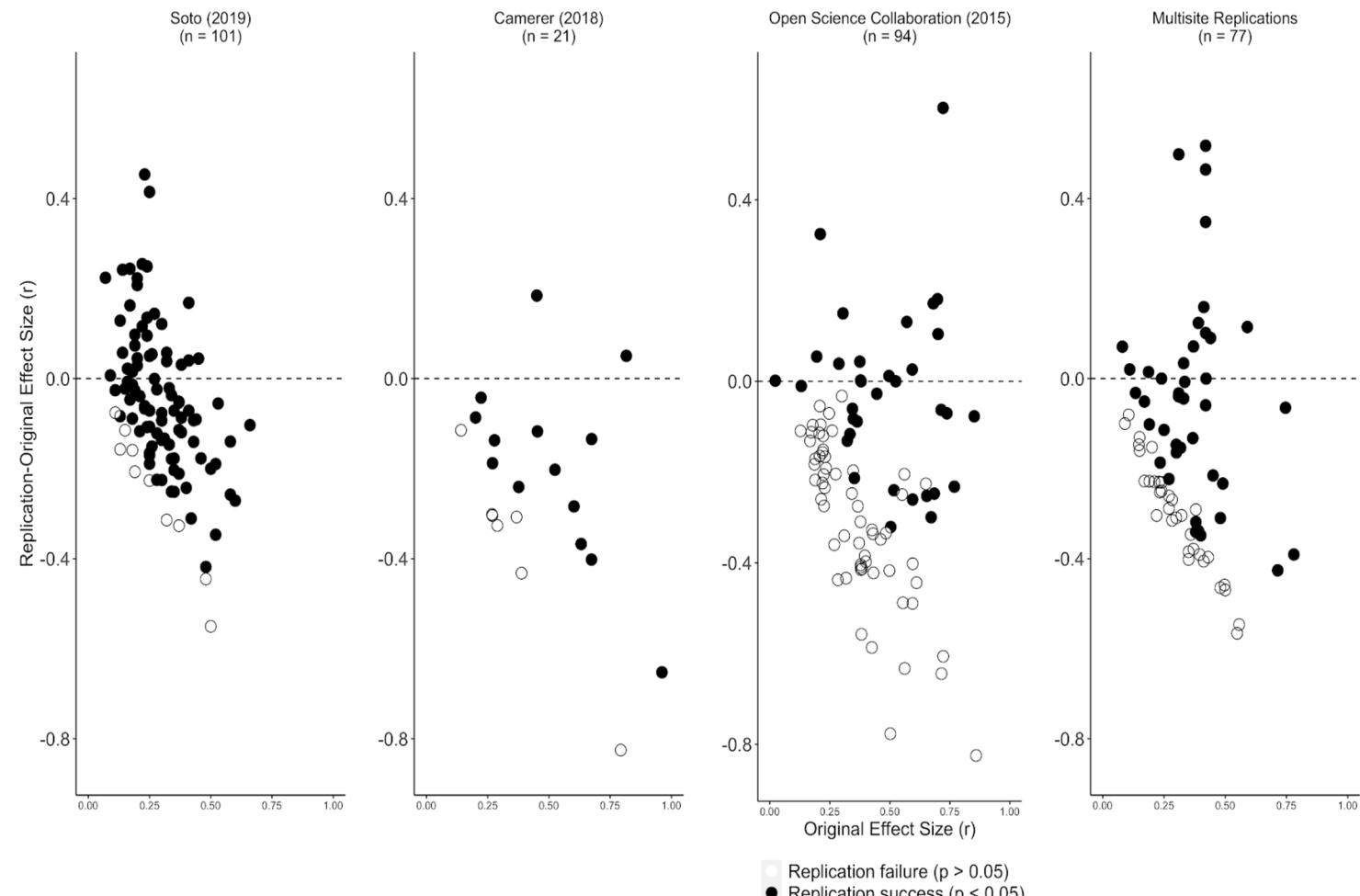
“ An article about **computational** science in a scientific publication is not the scholarship itself, it is merely advertising of the scholarship. The actual scholarship is the complete **software development environment**, and the complete set of instructions which generated the figures. ”

Buckheit & Donoho, 1995

Wavelab and Reproducible Research

Conclusions

Do adopting open science practices increase replicability?



Weaker
evidence than
original study



More information

{Take a picture}

- These slides: <https://osf.io/yf259>
- Center for Open Science: <https://cos.io/>
- Reproducibility Project: Cancer Biology: <https://cos.io/rpcb>
- NASEM Roundtable: <https://www.nationalacademies.org/our-work/roundtable-on-aligning-incentives-for-open-science>
- TOP Guidelines: <https://www.cos.io/top/>
- DORA: <https://sfdora.org/>
- NIH GREI: <https://datascience.nih.gov/news/nih-office-of-data-science-strategy-announces-new-initiative-to-improve-data-access>
- OSF: <https://osf.io/>
- Preregistration: <https://cos.io/prereg/>
- Registered Reports: <https://cos.io/rr/>
- Addgene: <https://www.addgene.org/>
- Resources on ELNs: <https://datamanagement.hms.harvard.edu/analyze/electronic-lab-notebooks>

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