

# Fast quantification of splice junctions by *sjcount*

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## 1 Synopsis

The purpose of *sjcount* is to provide a fast method for quantification of splice junctions from BAM files. It is an annotation-agnostic version of bam2ssj.

## 2 Installation and usage

See README.md file for installation instructions. *sjcount* is used with the following keys

**NOTE** that you need to install samtools. Samtools package Version: 0.1.18-dev (r982:313) is compatible (and likely so are older versions).

```
sjcount -bam bam_file [-ssj junctions_output] [-ssc boundary_output]
        [-maxlen max_intron_length] [-minlen min_intron_length] [-margin length]
        [-read1 0|1] [-read2 0|1] [-nbins number_of_bins] [-binsize bin_size]
        [-lim number_of_lines] [-quiet]
```

where

- **bam\_file** is a sorted input BAM file with a header
- **junctions\_output** is the output file with junction counts
- **boundary\_output** is the output file with boundary counts
- **maxlen** upper limit on intron length, 0 = no limit (default=0)
- **minlen** lower limit on intron length, 0 = no limit (default=0)
- **margin** length, see below, (default=0)
- **read1** 0/1, reverse complement read1 no/yes (default=no)
- **read2** 0/1, reverse complement read2 no/yes (default=no)
- **binsize** size of the overhang bin, (default= $\infty$ )
- **nbins** number of overhang bins, (default=1)

- **lim** nreads stop after nreads, (default=no limit)
- **quiet** – suppress verbose output

The output consists of two parts. First, a tab-delimited file containing splice junction counts is produced. Its format is as follows

```
chr1    100 200 -1 10 25
chr1    100    200 -1    11    12
... ..
```

where the first column contains chromosome id, the second and the third columns contain positions of terminal exonic nucleotides which define the splice junction, the fourth column contains strand (1 or -1), the fifth column is the overhang (see definitions below), and the last column is the respective number of reads with these properties.

The second output is a tab-delimited file which contains counts of continuous (non-split reads) which *overlap* splice sites of splice junctions tabulated in the previous step. This second file is optional and is used to compute the completeness of splicing index [2, 3].

### 3 Definitions

By definition, we say that we observe a splice junction each we see an 'N' symbol in the CIGAR attribute of some SAM alignment. For instance, the alignment shown in Figure 1 below gives rise to two splice junctions, denoted by SJ<sub>1</sub> and SJ<sub>2</sub>. We keep the convention that coordinates of splice junctions always refer

	10	20	30	40	50	60	70	80
	12345678	9012345678901234567890123456789012345678901234567890123456789012						
Ref	AGTCTAGG*GACGGCATAGGAGGTGAGCATTGTGTACGCAGATCTACAAAACATGTGTACGGATAGGATCG							
Query	CTAGGAGACGG**TAGGAG.....ATCTA*AAAACAT.....GATa							
			<-----	SJ1	----->		<--- SJ2 --->	

The corresponding SAM line is:

```
Query    123    Ref    14    255    5M1I5M2D6M20N5M1D7M13N3M1S 1234
```

Figure 1: An example alignment and its CIGAR attribute

to terminal exonic nucleotides, i.e., SJ<sub>1</sub> is Ref<sub>31</sub>52 and SJ<sub>2</sub> is Ref<sub>64</sub>78. We also denote the length of the intron by  $l(\text{SJ})$ , i.e.  $l(\text{SJ}_1) = 52 - 31 - 1 = 20$  and  $l(\text{SJ}_2) = 78 - 64 - 1 = 13$ . Intron length is always equal to the corresponding 'N' number in the CIGAR attribute.

Each splice junctions is associated with four numbers:  $m_u$  ( $m_d$ ) — the number of matching nucleotides immediately upstream (downstream) of the junction, and  $v_u$  ( $v_d$ ) — the length in the reference of the aligned region, also called



Note that when aggregated by the fifth column, the number of counts coincides with the collapsed counts.

## References

- [1] B. Kakaradov, H. Y. Xiong, L. J. Lee, N. Jovic, and B. J. Frey. Challenges in estimating percent inclusion of alternatively spliced junctions from RNA-seq data. *BMC Bioinformatics*, 13 Suppl 6:S11, 2012.
- [2] D. D. Pervouchine, D. G. Knowles, and R. Guigo. Intron-centric estimation of alternative splicing from RNA-seq data. *Bioinformatics*, 29(2):273–274, Jan 2013.
- [3] H. Tilgner, D. G. Knowles, R. Johnson, C. A. Davis, S. Chakraborty, S. Djebali, J. Curado, M. Snyder, T. R. Gingeras, and R. Guigo. Deep sequencing of subcellular RNA fractions shows splicing to be predominantly co-transcriptional in the human genome but inefficient for lncRNAs. *Genome Res.*, 22(9):1616–1625, Sep 2012.