

Green Synthesis, Antioxidant, and Plant Growth Regulatory Activities of Novel α -Furfuryl-2-alkylaminophosphonates

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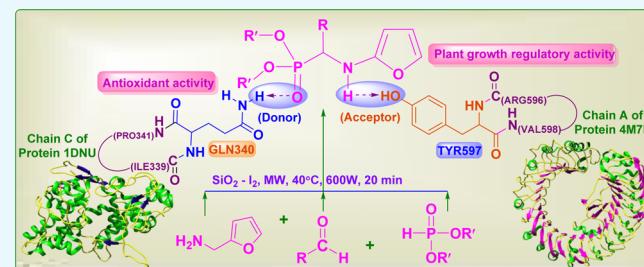
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ABSTRACT: A series of novel α -furfuryl-2-alkylaminophosphonates have been efficiently synthesized from the one-pot three-component classical Kabachnik–Fields reaction in a green chemical approach by addition of an *in situ* generated dialkylphosphite to Schiff's base of aldehydes and amines by using environmental and eco-friendly silica gel supported iodine as a catalyst by microwave irradiation. The advantage of this protocol is simplicity in experimental procedures and products were resulted in high isolated yields. The synthesized α -furfuryl-2-alkylaminophosphonates were screened to *in vitro* antioxidant and plant growth regulatory activities and some are found to be potent with antioxidant and plant growth regulatory activities. These *in vitro* studies have been further supported by ADMET (absorption, distribution, metabolism, excretion, and toxicity), quantitative structure–activity relationship, molecular docking, and bioactivity studies and identified that they were potentially bound to the GLN340 amino acid residue in chain C of 1DNU protein and TYR597 amino acid residue in chain A of 4M7E protein, causing potential exhibition of antioxidant and plant growth regulatory activities. Eventually, title compounds are identified as good blood–brain barrier (BBB)-penetrable compounds and are considered as proficient central nervous system active and neuroprotective antioxidant agents as the neuroprotective property is determined with BBB penetration thresholds.



1. INTRODUCTION

Organophosphorus chemistry research bestowed an elegant locus to α -aminophosphonates as they have exhilarated with potential antioxidant,¹ antifungal,² antibacterial,³ antidiabetic,⁴ anticancer,⁵ antiviral,⁶ antitumor,⁷ cytotoxic,⁸ antiproliferative,⁹ antibiotic,¹⁰ antithrombotic,¹¹ peptidomimetic,¹² catalytic antibodies hapten,¹³ neuroactive enzyme inhibiting, and anti-bone resorption¹⁴ activities. As they are tetrahedral transition-state mimics, they are identified as HIV protease,¹⁵ synthase,¹⁶ serine hydrolase,¹⁷ and UDP-galactopyranose mutase¹⁸ inhibitors. They have also been identified as surrogates of both natural and synthetic α -amino acids¹⁹ as constituted with significant motifs in medicine. In agriculture, phosphorus is identified as an essential element^{20,21} and is widely used in the production of crop fertilizers. They find their use as insecticides,²¹ plant growth regulators,^{22–24} herbicides,²⁵ pesticides,²⁶ fungicides,²⁷ and in photosynthesis as ADP and ATP. In the industry, they are used as additive polymers,²⁸ surfactants,²⁹ lubricants,²⁹ surface treaters on metals,³⁰ and flame retardants.³¹ In organic chemistry,³² they have been used as catalysts³³ in coupling³⁴ and complex reactions³⁵ and also as catalytic promoters.³⁶ Many aminophosphonic acids/peptides/esters from both synthetic and natural origin have displayed an extensive array of biological activities.³⁷ They act as substrates

in the synthesis and development of impending drugs against various metabolic disorders.³⁸

Owing to these, the synthesis of α -aminophosphonates had grown more inquisitiveness in medicinal chemistry researchers to study their structure–activity relationship and to design some more potential libraries to the existing repositories. Although a number of synthetic methodologies have been reported to construct bioactive α -aminophosphonates,^{39–42} the nucleophilic phosphite addition to Schiff's base is the utmost prevailing and modest method. Here, certain active catalysts such as titanium dioxide, boric acid, sulfonic acid functionalized ionic liquid, nanoporous AlSBA-15,⁴³ Magnetic BaFe₁₂O₁₉/Al₂O₃,⁴⁴ and silica sulfuric acid⁴⁵ are used with conventional heating, sonication, microwave irradiation in organic solvents,⁴⁶ and even with simple grinding. Among them, reactions stimulated with ultrasonic/microwave irradiation led to the specific attention as these energizing

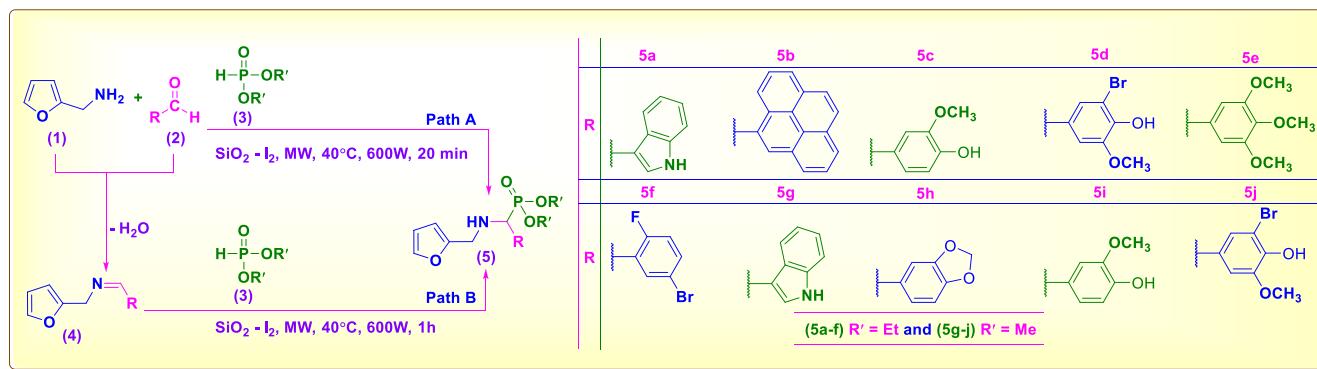
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Scheme 1. Synthesis of α -Furfuryl-2-alkylaminophosphonates (**5a–j**)

techniques advance the reaction and complete it in less time under trivial conditions and produces high yields and also more pronounced scientifically. Concerning the vitality of furfuryl and tetrahydrofurfuryl groups in pharmaceuticals, fibers, and perfumes,^{47–51} one-pot microwave Kabachnik–Fields synthesis of some novel and structurally diversified α -furfuryl-2-ethyl aminophosphonates (**5a–j**) has been designed through the reaction of aryl aldehydes, furfurylamine, and dialkyl phosphonates (Scheme 1). The synthesized α -furfuryl-2-alkylaminophosphonates were structurally confirmed by physical characterization and spectral studies, and their antioxidant and plant growth regulatory activities have been evaluated by *in silico* and *in vitro* methods. Eventually, herein we describe the microwave-supported synthesis, spectral and physical characterization, and biological activity evaluation of novel furfurylamine-based α -aminophosphonates. The additional studies such as molecular docking, quantitative structure–activity relationship (QSAR), and ADMET (absorption, distribution, metabolism, excretion, and toxicity) performed to study their bioactivity properties infer that the bioactive proficiency of **5a–j** has been administrated through a stereogenic α -carbon's configuration bonded to phosphorus.

2. RESULTS AND DISCUSSION

2.1. Chemistry. Synthesis of diethyl (((furan-2-ylmethyl)-amino) (4-hydroxy-3-methoxyphenyl)methyl) phosphonate (**5a**) has been accomplished by two different reaction pathways A and B. Path A involves a straight one-pot and three-component, $\text{SiO}_2\text{-I}_2$ -catalyzed Kabachnik–Fields reaction of vanillin (**2a**), furfurylamine (**1**), and diethylphosphite (**3a**) promoted by microwave irradiation. In path B, the reaction is carried out through the addition of diethylphosphite (**3a**) to the vanillin-derived Schiff base, vanillinidene-furfurylamine (**4**) (Scheme 1); by following the same procedure, the remaining analogues of the series (**5b–j**) have been synthesized.

2.1.1. Optimization of Reaction Conditions for the Synthesis of **5a.** Both reactions were run under microwave irradiation at different conditions and observed that best product yields were obtained in both the pathways at 40 °C and 600 W (Table 1). Among the two reaction pathways, better product yields were obtained in path A in less time because of the synergic formation of Schiff's base from vanillin, furfurylamine, and addition of phosphite to it. However, in path B, water formed during the formation of imine hydrolyzes the imine back to aldehyde that results in lower yields. We observed that the reaction goes slowly and takes 6 h to complete without any catalyst. When the reaction was

Table 1. Optimization of Synthesis of **5a**^a

temperature (°C)	microwave irradiation (W)	catalyst (10 mol %)	time (min)	isolated yield (%) ^b
25	100	—	60	10.24
25	200	—	60	12.49
25	300	—	60	15.17
25	400	—	60	20.66
25	500	—	60	25.48
25	600	—	60	32.59
25	700	—	60	32.38
25	800	—	60	20.27
30	600	—	60	32.69
40	600	—	60	35.74
50	600	—	60	35.93
60	600	—	60	25.87
40	600	Al_2O_3	45	51.79
40	600	SiO_2	42	66.30
40	600	$\text{Al}(\text{OTf})_3$	40	70.52
40	600	nano Al_2O_3	43	57.95
40	600	$\text{BF}_3\cdot\text{EtO}$	40	63.41
40	600	I_2	35	76.54
40	600	$\text{SiO}_2\text{-I}_2$	20	92.29

^aReaction conditions: diethyl phosphite (**3a**), vanillin-derived Schiff base (**4**), and $\text{SiO}_2\text{-I}_2$ as the catalyst. ^bIsolated yield.

conducted with different catalysts such as Al_2O_3 , SiO_2 , $\text{Al}(\text{OTf})_3$, and $\text{BF}_3\cdot\text{EtO}$, moderate product yields (35–45%) were obtained. When the reaction was conducted in 10 mol % of $\text{SiO}_2\text{-I}_2$, a great increase in the rate of reaction yields in lesser times takes place. At the outset, it was thought that $\text{SiO}_2\text{-I}_2$ was acquainted for catalytic support; besides, kinetic studies have revealed that it enhances the rate of reaction because of its acidic characteristic and ease of binding of silicone with several atoms of the substrates. Recently, several research groups have utilized some silica-supported catalysts such as $\text{SiO}_2\text{-CeCl}_2$ and $\text{SiO}_2\text{-ZnCl}_2$ ⁵² effectively to synthesize α -amino/carbamoyl/carbamothioyl phosphonates.

In addition, iodine was used as a catalyst owing to its abundant availability and low cost, ease of handling, stability, and use in numerous chemical transformations.^{53,54} Further, the catalyst mixture $\text{SiO}_2\text{-I}_2$ was prepared from iodine and silica gel (100–200 mesh) in a 2:1 ratio. It worked as a heterogeneous catalyst and conveniently handled and separated from the reaction mixture through filtration for reuse and is environment-friendly. To our best knowledge, the $\text{SiO}_2\text{-I}_2$ catalyst has not been reported to synthesize α -aminophosphonates, and hence, we have explored the one-pot and

three-component Kabachnik–Fields synthesis of α -furfuryl-2-alkylamino-phosphonates under heterogeneous catalytic action of $\text{SiO}_2\text{--I}_2$ in association with solvent-free, microwave irradiation conditions.

Hence, the finest conditions for reacting 2.0 mmol of aldehyde (2) and furfurylamine (1) with 2.1 mmol of dialkylphosphite (3) and 10 mol % of $\text{SiO}_2\text{--I}_2$ under microwave irradiation and neat conditions have accomplished and produced **5a–j** in 83–92% yields (Table 2). It is observed

Table 2. Reaction Time and Yield of the Products **5a–j**

entry	reaction time (min)	isolated yield (%)
5a	20	92.35
5b	22	90.42
5c	15	95.28
5d	22	89.41
5e	15	96.53
5f	25	85.17
5g	20	90.34
5h	25	83.76
5i	15	93.89
5j	22	85.06

that this reaction method is applicable to a wide range of aromatic aldehydes (electron-rich as well as electron-deficient) and diethyl and dimethyl phosphites and that the reactions proceed smoothly in 20–30 min and produced **5a–j** in excellent isolated yields. Eventually, it is noticed that the effect of electron-withdrawing groups on the carbonyl compound decreases the product yield and also increases the reaction time, while the effect of electron-releasing groups on carbonyl compound increases the product yield and decreases the reaction time.

2.1.2. Plausible Reaction Mechanism for the Synthesis of **5a–j.** Obviously, this reaction is a multistep procedure with the transition state having a quite complex mechanism (Figure 1). In fact, in the first step, the catalyst has greatly promoted the reaction by the virtue of acidic nature of silicon, is well influenced by iodine by withdrawing electrons from silica, and activates electrophilically the carbonyl carbon of the aldehyde

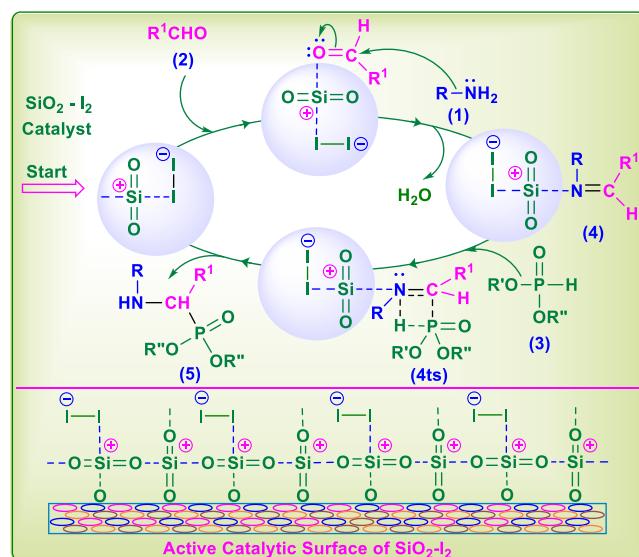


Figure 1. Mechanism for $\text{SiO}_2\text{--I}_2$ -catalyzed synthesis.

(2), which facilitates nucleophilic substitution by the amine (1) to form the imine (4).

Because its formation is a reversible process, it is trapped immediately by the catalyst as a four-membered imine–phosphite– $\text{SiO}_2\text{--I}_2$ complex transition state (4ts). Formation of 4ts not only prevents hydrolysis of the imine substrate (4) to the amine (1) and aldehyde (2) by water formed during the condensation of amine (1) and aldehyde that also force the reaction forward. The $\text{SiO}_2\text{--I}_2$ catalyst coordinates to the nitrogen atom of 4ts renders it more electron rich by attracting the electron density of imino bond and facilitates concerted break-up of 4ts in such a way as to lead to the nucleophilic addition of phosphino phosphorus to the iminocarbon atom and electrophilic addition of hydrogen to the iminonitrogen atom to form α -aminophosphonate (5) with simultaneous release of the catalyst.

2.2. Antioxidant Activity. The title compounds were screened for *in vitro* plant growth and antioxidant activities and supported by molecular docking studies. Compounds which inhibit the oxidation are known to be antioxidants. If oxidation takes place, then there is a chance of formation of free radicals, which leads to chain reactions that may damage the active cells. Hence, the main aim of the antioxidants is to inhibit generation of free radicals, thereby protecting the active cells.^{55–58} To evaluate the capacity of antioxidants, different types of methods are developed, and among them, 1,1-diphenyl-2-picrylhydrazyl (DPPH) and H_2O_2 methods are more popular and effective. Hence, the synthesized novel α -furfuryl-2-ethylaminophosphonates (**5a–j**) are screened for antioxidant activity by these methods.

2.2.1. DPPH Radical Scavenging Assay. Compounds **5a–j** were subjected to free radical scavenging *in vitro* by the DPPH radical assay;⁵⁹ the IC_{50} value denotes the concentration of the compound that inhibits 50% process of oxidation and is obtained by plotting the concentration *versus* percentage of inhibition graph and the obtained IC_{50} values shown against their compounds (Figure 2). Among the synthesized

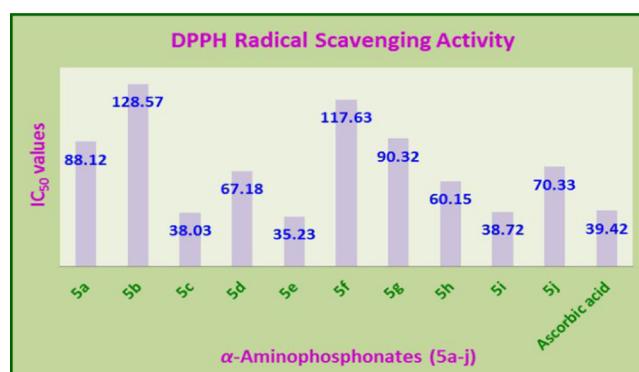


Figure 2. DPPH radical scavenging activity of **5a–j**.

compounds **5a–j**, **5e** ($\text{IC}_{50} = 35.23$) showed higher bioactivity compared to standard ascorbic acid ($\text{IC}_{50} = 39.42$). Also, **5c** (38.03) and **5i** (38.72) showed good results in comparison to ascorbic acid. Perhaps their significant activities are due to the presence of electron-donating methoxy ($-\text{OCH}_3$) and hydroxy ($-\text{OH}$) substitutions in **5c**, **5i** at meta and para position, respectively. In **5e**, the two methoxy groups at the meta position being less conjugated than the methoxy group located at the para position delivers additional electron density to

overcome oxidation. The identical process in hydroxyl group present in **5c** and **5i**, but which are less compared to methoxy group, hydroxyl group and hence they have shown less antioxidant activity than **5e**. Furthermore, the presence of electron-withdrawing groups in **5f** (fluorine at the ortho position and bromine at the meta position) decreases the availability of electrons and is responsible for the lower activity.

2.2.2. H_2O_2 Radical Scavenging Assay. In practice, the hydroxyl free radical scavenging assay of **5a–j** has been performed by deoxyribose method⁶⁰ and was referenced to ascorbic acid standard. In H_2O_2 radical scavenging assay, the compound **5e** ($IC_{50} = 35.38$) exhibited noteworthy activity than the standard ascorbic acid (39.57), whereas compounds **5c** (37.03) and **5i** (38.72) exhibit moderate activities (Figure 3). Here also, the same structure–activity has been observed

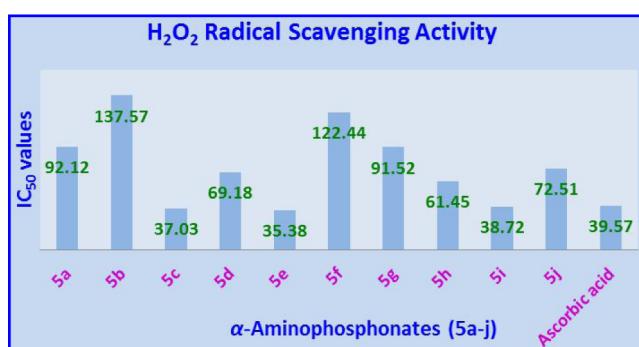


Figure 3. H_2O_2 radical scavenging activity of **5a–j**.

among the series of compounds as noticed in the DPPH radical scavenging assay. The results obtained from DPPH and H_2O_2 free radical scavenging assays followed the linear correlation and is comparable to each other. It is not necessary that the OH group alone shows antioxidant activity. The concept of electron donor and acceptor also plays a very important role in the redox reactions. Therefore, any functional group which is capable of donating electrons may act as a good antioxidant. Among the synthesized title compounds, **5c**, **5e**, and **5i** showed relatively higher antioxidant activity potentiality compared to standard ascorbic acid.

2.3. Plant Growth Activity. As organophosphorus compounds are bioactive toward both animals and plants and in such aminophosphonates are comparatively more potent than glyphosate in terms of biological activity. It is analyzed and identified that the biological activity of title

compounds intensely correlates with the lipophilic characteristic of Trakephon and hence also resembles the plant growth regulatory activity.⁶¹ Therefore, it is precisely credible that the action of title compounds is linked with distracting plant membrane's function as proposed by many researchers. Based on this notion, we have analyzed the effect of prepared α -furfuryl-2-ethylaminophosphonates (**5a–j**) differentiated by the nature of groups substituted on the phosphoryl phosphorus and carbonyl carbon atoms and hence indicating diverse polarity and hydrophobicity properties. Their plant growth regulatory activity was tested on roots and shoots of aquatic plant *sorghum bicolor*, using the technique of Smith and Fowden.⁶²

The results demonstrated that all the compounds (**5a–j**) clearly influenced the growth of the plant roots as well as shoots, but their effect on shoots is slightly less than that on the roots. Compound **5c** at 100 ppm showed highest root length (7.06 cm), and compound **5e** showed highest shoot length (1.96 cm). Further the point of structure–activity relationships, compounds substituted with electron-releasing groups in the aromatic ring (**5c**, **5f**, and **5j**) promoted growth, while those substituted with electron-withdrawing groups (**5f**) inhibit the growth of the root. However, in the case of **5d** and **5j**, the presence of both electron-withdrawing and hydroxyl groups showed moderate results. In the case of effect on shoot lengths, all the compounds showed mostly equal effect. With respect to the effect of concentration of the compound in solution, plant growth also increases with the increase of concentration of the compound. The results are represented in Figure 4.

2.4. Molecular Docking Studies. The *in vitro* DPPH and H_2O_2 radical scavenging activity and plant growth regulatory activity of **5a–j** have been supported by their binding potentialities against the considered enzymatic protein chains as revealed based on molecular docking studies. Here, the molecular docking was executed for the structures of **5a–j** against the target protein receptors in chain C of 1DNU (human myeloperoxidase–thiocyanate complex) protein and chain A of 4M7E (BL-induced activation of the BRI1–BAK1 complex) protein, respectively, for the antioxidant and plant growth regulatory activities to study their protein–ligand binding interactions.

The potential binding modes of amino acid residues against hydrogen bond acceptor and donor atoms of **5a–j** and their docking poses are presented in Tables 3–6. The binding specificity of **5a–j** with chain C (containing 466 amino acids)

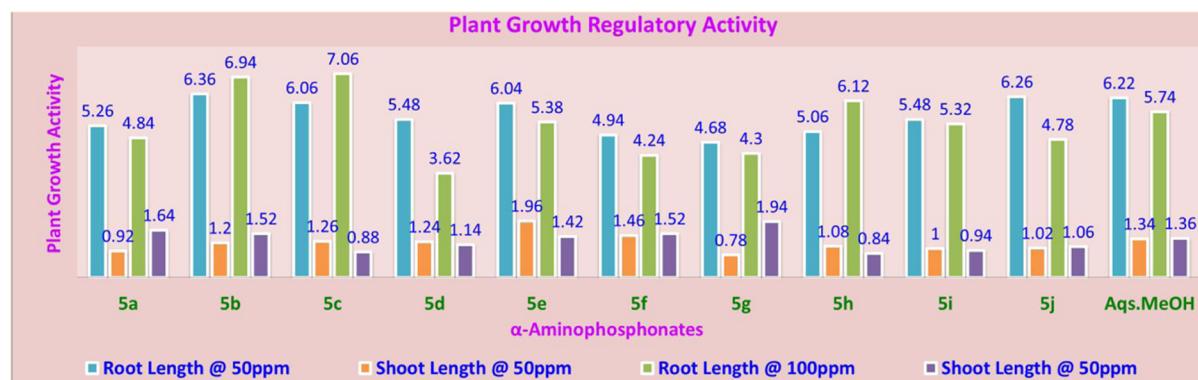
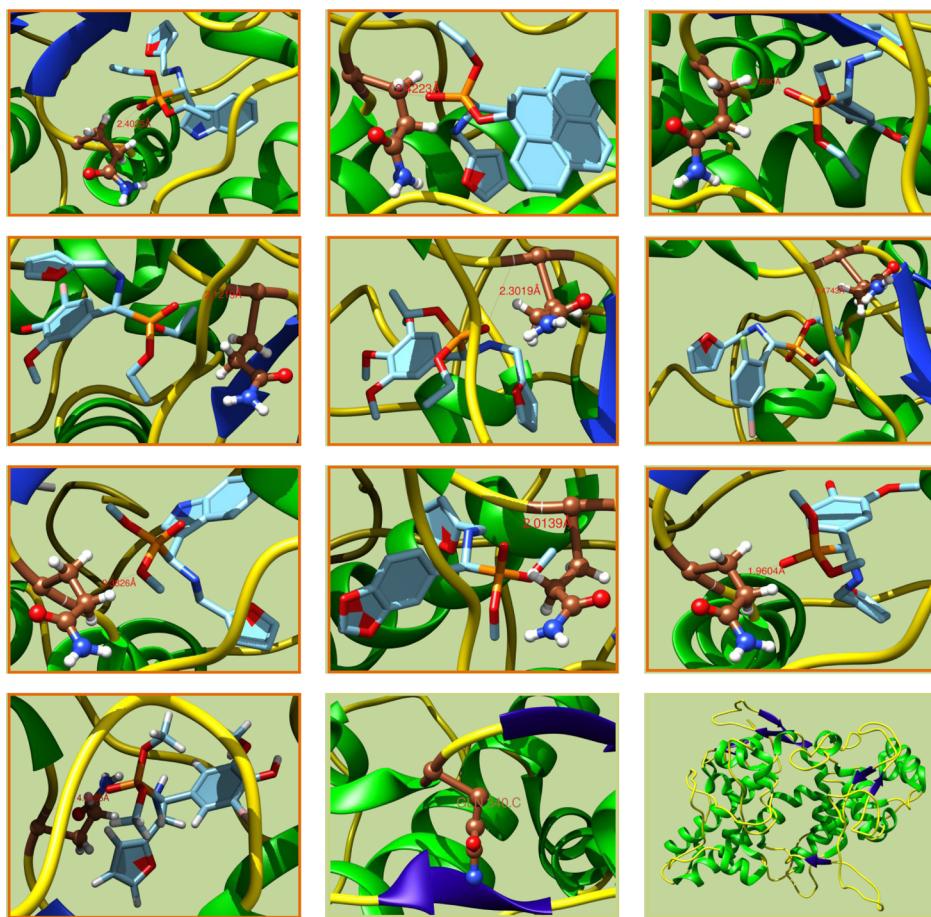


Figure 4. Plant growth regulatory activity of compounds **5a–j**.

Table 3. Potential Protein–Ligand Interactions of **5a–j** with Chain C of Protein 1DNU at GLN340 Amino Acid Residue^a

^aDocking poses of **5a–j**, GLN340 in chain C of 1DNU protein and chain C of 1DNU protein (in order—top left to bottom right).

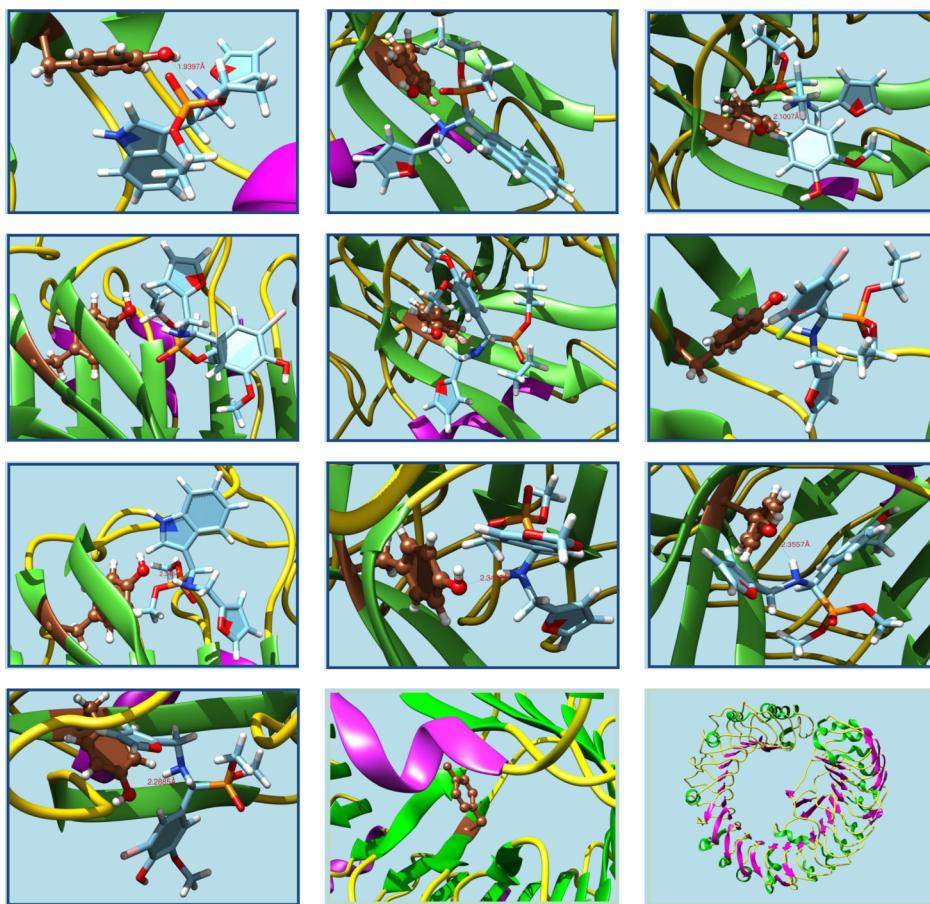
Table 4. Molecular Docking Interactions of **5a–j** with Chain C of Protein 1DNU at GLN340 Amino Acid Residue

S. no.	cluster number	cluster rank	binding energy (kcal/mol)	no. of hydrogen bonds	H-bond ligand atoms	H-bond receptor atoms	binding interaction	bond length (Å)	H-bond type
5a	4	1	-8.2531	1	1	1	ligand(P=O)...(HN) GLN340	2.4025	donor
5b	1	4	-9.2938	1	1	1	ligand(P=O)...(HN) GLN340	2.4223	donor
5c	0	0	-8.2422	1	1	1	ligand(P=O)...(HN) GLN340	2.3590	donor
5d	5	5	-8.6906	1	1	1	ligand(P=O)...(HN) GLN340	2.1219	donor
5e	4	3	-9.2858	1	1	1	ligand(P=O)...(HN) GLN340	2.3019	donor
5f	19	0	-8.0713	1	1	1	ligand[P=O]...(HN) GLN340	2.4743	donor
5g	0	0	-7.7212	1	1	1	ligand[P=O]...(HN) GLN340	2.3626	donor
5h	0	1	-8.2572	1	1	1	ligand(P=O)...(HN) GLN340	2.0139	donor
5i	0	11	-8.3254	1	1	1	ligand(P=O)...(HN) GLN340	1.9604	donor
5j	0	1	-8.4110	1	1	1	ligand(P=O)...(HN) GLN340	2.3885	donor

of 1DNU (myeloperoxidase) protein has confirmed their antioxidant potentiality as they were intensely bound with the glutamine amino acid residue (GLN340) located at the 340 position of the amino acid series in the chain. This assures the core interaction of synthesized compounds at the central

position of chain C, which is a valid consideration to support antioxidant activity *in vitro*.

Remarkably, they were bound to GLN340 with a binding energy of -9.2938 to -7.7212 kcal/mol with a rational bond length of 1.9604–2.4743 Å in forming hydrogen bonds. Profoundly, compounds **5a–j** formed hydrogen bonding with

Table 5. Potential Protein–Ligand Interactions of **5a–j** with Chain A of Protein 4M7E at TYR597 Amino Acid Residue^a

^aDocking poses of **5a–j**, TYR597 in chain A of 4M7E protein and chain A of 4M7E protein (in order—top left to bottom right).

Table 6. Molecular Docking Interactions of **5a–j** with Chain A of Protein 4M7E at TYR597 Amino Acid Residue

S. no.	cluster number	cluster rank	binding energy (kcal/mol)	no. of hydrogen bonds	H-bond ligand atoms	H-bond receptor atoms	binding interaction	bond length (Å)	H-bond type
5a	2	1	−8.0654	1	1	1	ligand(P—C(H)—NH)…(O—H)TYR597	1.9397	acceptor
5b	33	1	−8.1484	1	1	1	ligand(P—C(H)—NH)…(O—H)TYR597	1.9236	acceptor
5c	0	0	−7.9312	1	1	1	ligand(P—C(H)—NH)…(O—H)TYR597	2.1007	acceptor
5d	6	6	−7.9150	1	1	1	ligand(P—C(H)—NH)…(O—H)TYR597	2.3359	acceptor
5e	20	1	−7.8425	1	1	1	ligand(P—C(H)—NH)…(O—H)TYR597	2.7589	acceptor
5f	40	0	−7.6491	1	1	1	ligand(P—C(H)—NH)…(O—H)TYR597	2.0703	acceptor
5g	1	3	−7.6353	1	1	1	ligand(P—C(H)—NH)…(O—H)TYR597	2.3484	acceptor
5h	2	1	−7.8134	1	1	1	ligand(P—C(H)—NH)…(O—H)TYR597	2.3462	acceptor
5i	21	2	−7.9364	1	1	1	ligand(P—C(H)—NH)…(O—H)TYR597	2.3557	acceptor
5j	22	1	−7.3909	1	1	1	ligand(P—C(H)—NH)…(O—H)TYR597	2.2885	acceptor

GLN340 as an oxygen atom of the phosphoryl group has been strongly bound to an amino hydrogen atom of the amide bond present on the glutamine; hence, here the title compounds act as the electron cloud donors in forming the hydrogen bond. Similarly, the binding specificity of **5a–j** with chain A (containing 767 amino acids) of 4M7E (brassinosteroid

insensitive 1) protein has confirmed their plant growth regulatory activity as they were intensely bound with tyrosine amino acid residue (TYR597) located at the S97 position of the amino acid series in the chain. This assures the core interaction of synthesized compounds at central position of chain A, which is a valid consideration to support plant growth

Table 7. ADMET Properties of 5a–j

entry	<i>in vivo</i> BBB penetration (C.brain/C.blood) ^a	<i>in vitro</i> Caco-2 cell permeability(nm/s) ^b	human intestinal absorption(HIA, %) ^c	<i>in vitro</i> MDCK cell permeability(nm/s) ^d	<i>in vitro</i> plasma protein binding (%) ^e	toxicity ^f
5a	1.2749	21.7129	89.5313	191.0100	95.7056	negative
5b	2.5729	21.7217	97.2754	79.4515	99.2126	negative
5c	0.4368	21.7072	89.8057	122.3450	90.6733	negative
5d	0.4900	18.8444	94.9217	27.9760	91.1146	negative
5e	0.6212	21.7231	94.1342	74.8857	96.2773	negative
5f	1.2911	21.4068	96.4950	38.0802	98.2989	negative
5g	0.7318	21.6890	88.5328	186.4370	93.6175	negative
5h	0.4699	21.7074	92.7086	87.8067	93.2263	negative
5i	0.4518	21.6737	86.9710	122.9902	94.1828	negative
5j	0.4560	16.1225	94.4193	29.6239	94.1007	negative

^aBBB penetration = [brain]/[blood]. ^bCaco-2 cells derived from human colon adenocarcinoma, possessing multiple drug transport pathways through intestinal epithelium. ^cHIA (human intestinal absorption), the sum of absorption and bioavailability evaluated from ratio of excretion in urine, bile, and feces. ^dMDCK cell system is used as a tool for rapid permeability screening. ^e% of drug that binds to plasma protein. ^f*In vitro* Ames test by metabolic and nonmetabolic activated TA100 and TA1535 strains collected from rat liver homogenate.

Table 8. Molecular Descriptor Properties of 5a–j^a

entry	Lipinski parameters					Veber parameters			other parameters						
	MW	HB Don	HB Acc	log P (o/w)	MR	Lip. Vio.	TPSA	no. of RB	Veb. Vio.	no. of H	V. Vol.	ρ	S	CLP	% ABS
5a	362.37	6	2	2.08	96.64	0	86.30	9	0	25	325.07	1.246	-3.61	2.52	79.23
5b	447.47	5	1	4.24	126.18	0	60.71	9	0	32	400.65	1.275	-3.54	3.19	88.06
5c	369.35	7	2	2.23	93.67	0	99.97	10	0	25	329.66	1.228	-2.81	1.74	74.51
5d	448.25	7	2	3.02	101.3	0	99.97	10	0	26	347.54	1.414	-3.64	2.46	74.51
5e	413.41	8	1	2.01	104.91	0	98.20	12	0	28	372.73	1.180	-3.14	1.95	75.12
5f	420.21	5	1	3.70	93.36	0	70.51	9	0	24	318.91	1.407	-4.24	2.89	84.67
5g	334.31	6	2	1.77	87.14	0	76.50	7	0	23	291.47	1.297	-3.01	1.38	82.60
5h	339.28	7	1	1.51	81.79	0	89.17	7	0	23	286.42	1.323	-3.20	1.45	78.24
5i	341.30	7	7	0.96	84.18	0	99.17	8	0	23	296.05	1.275	-2.21	1.20	74.78
5j	420.20	7	2	1.92	91.80	0	99.37	8	0	24	313.94	1.480	-3.04	1.65	74.72

^aMW: molecular weight; HB Don: hydrogen bond donors (*n* ON); HB Acc: hydrogen bond acceptors (*n* OH NH); log P: log of octanol to water partition coefficient; MR: molecular refractivity (cm³/mol); Lip. Vio.: lipinski violations; TPSA: total polar surface area (Å)²; no. of RB: number of rotatable bonds; Veb. Vio.: Veber violations; no. of "H": number of hydrophobic atoms; V. Vol.: van der Waals volume; ρ : density (gm/cc); S: solubility; CLP: clog P; and % ABS: % of absorption.

regulatory activity. Remarkably, they were bound to TYR597 with a binding energy of -8.1484 to -7.3909 kcal/mol with a rational bond length of 1.9397–2.7589 Å in forming hydrogen bonds. Profoundly, compounds 5a–j formed hydrogen bonding with TYR597 as the amino hydrogen atom of the α -aminophosphoryl group has been strongly bound to the phenolic oxygen atom present on the tyrosine; hence, here the compounds 5a–j act as electron cloud acceptors in forming the hydrogen bond. This investigation unveils that GLN340 and TYR597 amino acids are strongly bound to 5a–j and are act as donors and acceptors, respectively, and responsible for the predicted *in vitro* activities.

2.5. ADMET Properties. The analysis of ADMET profiles for a library of molecules under scrutiny helps to figure out the physicochemical collaborations and evaluate their druglikeness properties. This sort of high-throughput screening assists in characterizing a lead one out of a large group in the captivated field of a target.⁶³ This critical investigation had supported in predicting the pharmacokinetic effects of 5a–j and to disclose the druglikeness properties. Among the ADMET properties, the human intestinal absorption designates transporting of the effective composites to the target cell tissues *via* blood stream and sustains their mutual interaction. During this the oral administration of a compound under study, its degree of absorption has been taken into account, which depends on its

intrinsic bioavailability characteristics. In turnout, the absorbed quantity of the analyte is distributed to the muscles and then to other organs in self-circulation mode through extracellular sites. Then, the distribution of analyte drops down its plasma concentration autonomously and then metabolized; from there, such metabolites will be dispersed to the target sites by enzymatic redox reactions. In pharmacological perspectives, potentially distributed metabolites will work competently for cellular organisms, and on the other side, the inactive metabolites deactivate the controlled entity and moderate its possible effect *in vivo*; finally, the inert metabolites are inevitably emitted by kidneys. The analysis of the acquired ADMET profiles (Table 7) of 5a–j infers us about the *in vivo* blood–brain barrier (BBB) penetration potentiality ratio which is operational in the range of 0.4518–2.5729 and authorizes their high central nervous system (CNS) significance and commends their greater permeability for its distribution *in vivo*. It is strengthened on the source that the *in vitro* Caco-2 cell permeability property is effective within 16.1225 to 21.7231 nm/s range and establishes their insistent moderate permeability to bind with plasma proteins and to pierce into the BBB system. The *in vitro* plasma protein binding (PPB) affinity ranging from 90.6733 to 99.2126% empowers their robust binding ability to plasma proteins. The *in vitro* Maden Darby Canine kidney (MDCK) cell

Table 9. Molecular Fingerprint Properties of **5a–j^a**

entry	point group	H.F. (kcal/mol)	T.E. (eV)	E.E. (eV)	C.C.R. (eV)	G.N.	dipole (Debye)	no. of F.L.	I.P. (eV)	H.E. (eV)	L.E. (eV)	C.A. (Å ²)	C.V. (Å ³)
5a	C1	-106.66	-4165.36	-31020.65	26855.30	0.98	1.89	64	8.36	-8.36	0.07	375.31	430.50
5b	C1	-105.79	-4165.32	-31142.12	26976.79	0.94	1.84	64	8.14	-8.14	0.30	372.69	431.76
5c	C1	-209.24	-4489.49	-33075.83	28586.33	0.92	2.49	66	8.79	-8.79	-0.05	370.76	440.94
5d	C1	-209.11	-4823.17	-35478.33	30655.16	0.99	2.09	69	8.91	-8.91	-0.24	386.63	463.71
5e	C1	-241.18	-5123.82	-40651.74	35527.92	0.90	1.53	75	9.07	-9.0	-0.30	402.91	497.09
5f	C1	-167.25	-4487.54	-30712.54	26224.99	0.97	1.97	63	9.17	-9.17	-0.60	365.74	429.05
5g	C1	-96.74	-3852.44	-26907.70	23055.26	0.99	1.81	58	8.38	-8.38	0.06	335.01	384.06
5h	C1	-187.95	-4147.75	-27663.14	23515.38	0.91	1.84	59	8.86	-8.86	-0.20	330.80	375.71
5i	C1	-202.23	-4176.70	-28679.19	24502.48	0.98	2.43	60	8.73	-8.73	0.05	335.26	391.00
5j	C1	-199.95	-4510.28	-30781.31	26271.03	0.95	0.82	63	9.09	-9.09	-0.38	357.86	417.55

^aH.F.: heat of formation; T.E.: total energy; E.E.: electronic energy; C.C.R.: core–core repulsion; G.N.: gradient norm; F.L.: filled levels; I.P.: ionization potential; H.E.: HOMO energy; L.E.: LUMO energy; C.A.: COSMO area; and C.V.: COSMO volume.

Table 10. Bioactivity Scores, Drug Properties, and Toxicity Risks of **5a–j^a**

entry	bioactivity						drug properties		toxicity risks			
	GPCRL	ICM	KI	NRL	PI	EI	drug-likeness	drug score	Mut	Tum	Irrit	R.E.
5a	-0.20	0.14	-0.20	-0.46	0.01	0.11	-32.76	0.42	green	green	green	green
5b	-0.24	0.07	-0.27	-0.33	0.02	0.10	-34.08	0.08	green	green	green	green
5c	-0.46	0.01	-0.47	-0.51	-0.07	0.02	-33.04	0.43	green	green	green	green
5d	-0.64	-0.12	-0.56	-0.68	-0.23	-0.11	-34.72	0.33	green	green	green	green
5e	-0.45	-0.04	-0.44	-0.59	-0.09	-0.03	-30.14	0.44	green	green	green	green
5f	-0.49	-0.04	-0.46	-0.62	-0.18	-0.08	-38.50	0.34	green	green	green	green
5g	-0.21	0.20	-0.29	-0.58	0.01	0.15	-25.33	0.16	green	green	green	green
5h	-0.47	-0.01	-0.65	-0.75	-0.06	0.10	-25.97	0.15	green	green	green	green
5i	-0.47	0.06	-0.57	-0.61	-0.07	0.06	-25.68	0.16	green	green	green	green
5j	-0.66	-0.07	-0.66	-0.79	-0.22	-0.08	-27.42	0.14	green	green	green	green

^aGPCRL: G protein-coupled receptor ligand; ICM: ion channel modulator; KI: kinase inhibitor; NRL: nuclear receptor ligand; PI: protease inhibitor; EI: enzyme inhibitor; Mut: mutagenic; Tum: tumorigenic; Irrit: irritant; R.E.: reproductive effect; green: green color indicates drug-conform behavior.

permeability readings ranging from 27.9760 to 191.0100 nm/s discloses them as worthy permeable. In the same way, the human intestinal absorption (% HIA) in the range of 86.9710–97.2754 ratio promises the interactions of **5a–j** with suitable species of the anticipating target domains. The negative sign of toxicity properties of **5a–j** assures them as safer and nontoxic drugs. In summary, this analysis has revealed the physicochemical interactions and druglikeness properties of **5a–j**.

2.6. QSAR Studies. The results obtained from *in silico* molecular descriptor properties (**Table 8**) designates that compounds **5a–j** are designed to obey the properties and their molecular weights ranging from 334.31 to 448.25 (less than 500 Da) approves the same. Similarly, they have been validated with log P values in the range of 4.24–0.96 (less than 5) suggesting their good permeability through cell membranes. Likewise in the line of Lipinski's rule of five, the number of hydrogen bond donors <5 and acceptors <10 are also validated in all molecules that were synthesized. Similarly, the molecular refractivity ranging from 81.79 to 126.18 cm³/mol has also obeyed the valid range, that is, 40–130 cm³/mol.

Hence, ascertained compounds **5a–j** are following the Lipinski rule of five, and all of them are significantly orally active drugs with worthy druglikeness properties. Furthermore, the total polar surface area is precised by the total effect of all polar atoms such as oxygen, nitrogen, and attached hydrogens;⁶⁴ for instance, compounds **5a–j** are also in the range of 60.71–99.97 Å², which are ultimately less than 140 Å², and are

also satisfying the Veber rule. Henceforth, compounds **5a–j** are assessed as they can be easily diffused, absorbed, and transported in the systems to which they are exposed. At this juncture, TPSA is highly interrelated with the hydrogen bonding of a compound and is accompanied with the transport properties of that drug across membranes and the correlation of BBB for the intestinal crossing. Compounds with TPSA ≤160 Å² have been identified as good intestinal absorption compounds with BBB penetrability ≤60 Å² as BBB-penetrable and CNS-active agents.⁶⁵ TPSA for the scrutinized sequences all the offshoots have been identified as the best intestinal absorbing ones. Similarly, the number of rotatable bonds among all the compounds is identified with a range of 7–12, which is according to the Veber rule, and eventually, they are also obeying the Veber's rule and established themselves as orally administrable drugs. Furthermore, the percentage of absorption in the range of 79.23–88.06, density ranging from 1.246 to 1.480 gm/cc, solubility in the range of -2.21 to -4.24, van der Waals volume ranging from 286.42 to 400.65 Å³, and clog P ranging from 1.20 to 3.19 establish compounds **5a–j** drug-like compounds and are significantly safer. These properties helped a lot in considering their physicochemical interactions against the target entities and eventually helped in deciding the drug potentialities that are associated by the bioactivity and toxicity risks studies.

The synthesized molecules **5a–j** are identified with the fingerprint properties (**Table 9**) as having all of them are having the C1 point group of symmetry elements in them and

heat of formation is identified within the range of -96.74 to -241.18 kcal/mol, total energy ranges from -3852.44 to -5123.82 eV, electronic energy ranges from -26907.7 to -40651.74 eV, and core–core repulsion energy ranges from 23055.26 to 35527.92 eV. Similarly, the gradient norm is identified in the ranges of values from 0.90 to 0.99 , whereas the dipole difference is identified with 0.82 – 2.49 debye. The filled levels were identified with 58 – 75 units, and the ionization potential values are identified from 8.14 to 9.17 eV. Similarly, highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO) energies are identified from -8.14 to -9.17 eV and from -0.6 to 0.3 eV, respectively. Similarly, the Cosmo area and Cosmo volumes are identified within the range of 330.80 – 402.91 \AA^2 and 375.71 – 497.09 \AA^3 , respectively. All these predicted values confirm that the synthesized molecules are correlating with the potential limits of safer drugs and pharmaceuticals, and hence, in complementary perspectives of the molecular descriptors, they are obeying the QSAR properties.

2.7. Bioactivity and Toxicity Risk Studies. The study of bioactivity and toxicity risk properties of **5a**–**j** (Table 10) inferred the bioactivity properties such as GPCRL (GPCR ligand property), ICM (ion channel modulator), KI (kinase inhibitor), NRL (nuclear receptor ligand) interactions, PI (protease inhibitor) and EI (enzyme inhibitor) interactions, and drug properties (druglikeness and drug scores), which have ascertained them as nontoxic and potential molecules. Prediction of these properties by Molinspiration software had comprehensively helped to assess the cheminformatics of **5a**–**j** by relating with *in vitro* and *in vivo* results based on functional group similarity. The results of toxicity risk study supporting **5a**–**j** as safer compounds as they are encompassed with no risks of tumorigenicity, mutagenicity, irritant and are showing no effect on the reproductive system and conformed their drug-like behavior. The positive magnitude of druglikeness represents most of the fragments that are the part of commercial drug molecules; however, many compounds signify the negative magnitudes.⁶⁶ Another important factor that infers about the association of a substance from the administering site to the blood stream is solubility, and good solubility leads to good absorption.⁶⁷ Similarly, drug score is considered as a harmonizing parameter that wholly represents clog *P*, druglikeness, log *S*, toxicity risks, and molecular weight and accounted to decide the molecule's inclusive potential to denote a drug. Eventually, it is identified that all the synthesized analogues **5a**–**j** showed higher scores (Table 10).

2.8. Structure–Activity Relationship Studies. The molecular docking studies of **5a**–**j** have well established the structure–activity relationship for them as molecules were intensely bound to glutamine (GLN340) located on chain C of 1DNU protein. Specifically, hydrogen bonding with GLN340 is identified as such the oxygen atom of the phosphoryl group has been strongly bound to the amino hydrogen atom of the amide bond present on the glutamine and is responsible for the antioxidant activity.

Similarly, molecules are strongly bound to tyrosine (TYR597) located on chain A of 4ME7 protein. Specifically, hydrogen bonding with TYR597 is identified as such the amino hydrogen atom of the α -aminophosphoryl group has been strongly bound to the phenolic oxygen atom present on the tyrosine and is responsible for the antioxidant activity (Figure 5). Moreover, the oxidative stress comprehends the pathogenesis of neurodegenerative diseases triggered by the

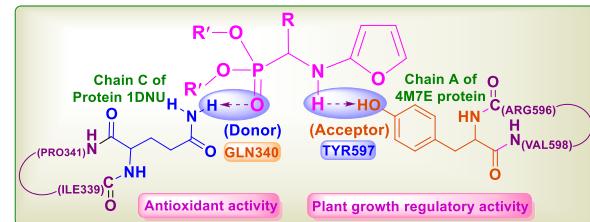


Figure 5. Structure–activity relationship of **5a**–**j**.

excess quantity of reactive oxygen species created *in vivo* by uneven biochemical disorders and which again causes damage of neuronal cells and their death.^{68–71} In relation to this, the title compounds are identified as good BBB-penetrable compounds; they can be considered as neuroprotective antioxidants with CNS-active potentiality as the neuroprotective property is determined with BBB penetration thresholds.

3. CONCLUSIONS

Novel α -furfuryl-2-alkylaminophosphonates (**5a**–**j**) were successfully synthesized with the aid of SiO_2 – I_2 , an eco-friendly catalyst. Their antioxidant properties were studied by DPPH and H_2O_2 radical scavenging assays. The plant growth regulatory activity in terms of growth of root length and shoot length was studied and observed that compounds **5e**, **5c**, and **5i** were identified with potential activity and comparable to the standards. The additional studies such as ADMET, QSAR, molecular docking, and bioactivity and toxicity risk have been well correlating with the results predicted *in vitro*. The complementary consideration of all the studies has supported the synthesized compounds as potential antioxidants and plant growth regulators with potential CNS-active neuroprotective antioxidant activity.

4. EXPERIMENTAL SECTION

4.1. General. Chemicals are acquired from Sigma-Aldrich, solvents were distilled before use, and the reactions were performed on a CATA-4R (Catalyst System) scientific microwave instrument. Thin-layer chromatography (TLC) analysis has been performed on silica gel-G-coated aluminum sheets as procured from Merck; the analyte mobilities were visualized under UV light, iodine absorption, and ninhydrin solution. ^1H and ^{13}C NMR and spectral studies were performed on a Bruker 400 MHz spectrometer, and the spectra were recorded in deuterated chloroform (CDCl_3) by using tetramethylsilane (TMS) as an internal reference standard.

Infrared (IR) spectra were recorded on a Bruker Alpa-ECO-ATR-FTIR (attenuated total reflection Fourier transform infrared) interferometer by using a single reflection sampling module equipped with a zinc selenide (ZNSe) crystal, and the readings were expressed in reciprocals of centimeter (cm^{-1}). Mass spectra were recorded by using an Agilent-MS spectrometer.

4.2. Chemistry. Synthesis of aminophosphonates (**5a**–**j**) has been achieved from the reaction of a mixture of carbonyl derivatives (1.0 mmol), furfurylamine (1 mmol), and diethyl/dimethyl hydrogen phosphite (1.0 mmol) by using SiO_2 – I_2 (10 mol %) catalyst by irradiating under microwave at 600 W and 40 °C for 15–25 min. The reaction progress was scrutinized by TLC for the completion of the reaction and

then 25 mL of ethyl acetate was added to the crude content and filtered, followed by washing with ethyl acetate (2×10 mL). Then, the solvent contents were evaporated by a rotary evaporation process. The crude compound was purified, and the pure product was collected by column chromatography and characterized by spectral and elemental analysis.

4.2.1. Diethyl (((Furan-2-ylmethyl)amino)(1*H*-indol-3-yl)methyl)phosphonate (5a). Brown liquid, 92% yield (334.7 mg); FT-IR (ZnSe): ν_{max} 3215, 2977, 2924, 1444, 1432, 1213, 1019, 953, 734; ^1H NMR (400 MHz, CDCl_3 /TMS): δ 10.08 (br s, 1H), 7.71–7.69 ($d, J_{\text{HH}} = 7.72$ Hz, 1H), 7.38–7.35 (m, 2H), 7.24 (s, 1H), 7.17–7.08 (m, 2H), 6.29 (s, 1H), 6.09 (s, 1H), 4.46 ($d, J_{\text{HH}} = 17.4$ Hz, 1H), 4.16–4.09 (m, 2H), 4.00–3.95 (m, 1H), 3.89 ($d, J_{\text{HH}} = 14.56$, 1H), 3.81–3.74 (m, 1H), 3.72–3.68 ($d, J_{\text{HH}} = 14.56$, 1H), 3.19 (br s, 1H), 1.31–1.27 (m, 3H), 1.09–1.05 (m, 3H); ^{13}C NMR (100 MHz, CDCl_3 /TMS): δ 153.05, 142.07, 136.49, 126.96, 124.99, 121.79, 119.20, 111.74, 110.20, 108.38, 107.83, 77.39, 76.91, 62.98, 52.10, 50.48, 16.17; HRMS (ESI) m/z calcd for $\text{C}_{18}\text{H}_{23}\text{N}_2\text{O}_4\text{P}^+$ (M + H), 362.1395; found, 362.1392.

4.2.2. Diethyl (((Furan-2-ylmethyl)amino)(pyren-4-yl)methyl)phosphonate (5b). Dark brown liquid, 90% yield (402.3 mg); FT-IR (ZnSe): ν_{max} 3318, 2981, 2920, 1678, 1234, 1017, 954, 730; ^1H NMR (400 MHz, CDCl_3): 8.49 (s, 1H), 8.34 (s, 1H), 8.28 ($d, J = 7.2$ Hz, 1H), 8.20–8.12 (m, 3H), 8.05 ($d, J = 9.2$ Hz, 2H), 8.02–7.93 (m, 1H), 7.37 (s, 1H), 7.29 (s, 1H), 6.28 (s, 1H), 5.99 (s, 1H), 5.33 ($d, J = 19.6$ Hz, 1H), 4.17–4.03 (m, 2H), 3.90 ($d, J = 14.0$ Hz, 2H), 3.70 ($dd, J = 15.5, 7.4$ Hz, 1H), 3.60 ($d, J = 14.7$ Hz, 1H), 3.22 (s, 1H), 1.27 ($t, J = 6.2$ Hz, 3H), 0.99 ($t, J = 6.6$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3): δ 152.80, 142.20, 131.37, 130.97, 130.67, 130.01, 128.90, 127.63, 125.51, 124.80, 124.75, 110.20, 108.12, 77.64, 77.32, 77.01, 63.78, 62.99, 43.70, 34.88, 16.50; HRMS (ESI) m/z calcd for $\text{C}_{26}\text{H}_{26}\text{NO}_4\text{P}^+$ (M + H), 447.1599; found, 447.1598.

4.2.3. Diethyl (((Furan-2-ylmethyl)amino)(4-hydroxy-3-methoxyphenyl)methyl)phosphonate (5c). Yellow liquid, 95% yield (349.8 mg), FT-IR (ZnSe): ν_{max} 3184, 2980, 2920, 1514, 1211, 1021, 953, 738; ^1H NMR (400 MHz, CDCl_3): δ 7.34 ($d, J = 1.0$ Hz, 1H), 7.03 (s, 1H), 6.86 ($d, J = 2.1$ Hz, 2H), 6.32–6.26 (m, 1H), 6.09 ($d, J = 2.5$ Hz, 1H), 4.08–4.01 (m, 2H), 4.00–3.92 (m, 2H), 3.88 ($d, J = 3.2$ Hz, 3H), 3.87–3.83 (m, 1H), 3.82–3.76 (m, 1H), 3.56 ($d, J = 14.6$ Hz, 1H), 1.27 ($t, J = 7.1$ Hz, 3H), 1.17 ($t, J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3): δ 152.84, 146.98, 145.75, 142.01, 126.34, 122.03, 114.45, 111.07, 110.09, 107.73, 77.41, 77.09, 76.77, 62.94, 59.59, 58.04, 55.95, 43.49, 16.32; HRMS (ESI) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{NO}_6\text{P}^+$ (M + H), 369.1341; found, 362.1344.

4.2.4. Diethyl ((3-Bromo-4-hydroxy-5-methoxyphenyl)((Furan-2-ylmethyl)amino)methyl) Phosphonate (5d). Orange red liquid, 89% yield (398.7 mg), FT-IR (ZnSe): ν_{max} 3125, 2980, 1495, 1424, 1278, 1231, 1156, 1026, 728; ^1H NMR (400 MHz, CDCl_3): δ 9.73 (s, 1H), 7.35 ($d, J = 1.3$ Hz, 2H), 7.12 ($d, J = 1.9$ Hz, 2H), 6.99 ($d, J = 1.8$ Hz, 2H), 6.30 ($d, J = 1.7$ Hz, 2H), 6.10 (s, 2H), 4.11–3.95 (m, 8H), 3.89 ($d, J = 2.0$ Hz, 10H), 3.80 ($d, J = 15.0$ Hz, 2H), 3.56 ($d, J = 14.7$ Hz, 2H), 3.04–2.94 (m, 2H), 1.27 ($t, J = 8.9$ Hz, 6H), 1.21 ($t, J = 8.1$ Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3): δ 155.58, 150.54, 146.09, 145.12, 113.16, 110.89, 80.44, 80.12, 79.80, 66.34, 65.89, 62.25, 60.71, 59.43, 46.47, 19.38, 11.57; HRMS (ESI) m/z calcd for $\text{C}_{17}\text{H}_{23}\text{BrNO}_6\text{P}^+$ (M + H), 447.0446; found, 447.0448.

4.2.5. Diethyl (((Furan-2-ylmethyl)amino)(3,4,5-trimethoxyphenyl)methyl)phosphonate (5e). Pale yellow liquid, 96% yield (396.6 mg); FT-IR (ZnSe): ν_{max} 3216, 2991, 2933, 1524, 1221, 1037, 956, 735; ^1H NMR (400 MHz, CDCl_3): δ 7.33 (s, 2H), 6.67 ($d, J = 2.2$ Hz, 3H), 6.28 ($d, J = 2.4$ Hz, 2H), 6.09 ($d, J = 3.3$ Hz, 2H), 4.10–3.95 (m, 8H), 3.95–3.90 (m, 3H), 3.90–3.81 (m, 20H), 3.78 (s, 2H), 1.26 ($t, J = 7.1$ Hz, 7H), 1.18 ($t, J = 7.0$ Hz, 7H); ^{13}C NMR (100 MHz, CDCl_3): δ 153.19, 152.78, 142.03, 130.59, 110.10, 107.75, 105.67, 99.99, 77.43, 77.12, 76.80, 62.92, 60.81, 60.13, 58.59, 56.13, 43.50, 16.26; HRMS (ESI) m/z calcd for $\text{C}_{19}\text{H}_{28}\text{NO}_7\text{P}^+$ (M + H), 413.1603; found, 413.1601.

4.2.6. Diethyl (((5-Bromo-2-fluorophenyl)((furan-2-ylmethyl)amino)methyl)phosphonate (5f). Orange red liquid, 85% yield (357.2 mg); FT-IR (ZnSe): ν_{max} 3306, 2982, 1477, 1237, 1021, 958, 805, 731; ^1H NMR (400 MHz, CDCl_3): δ 7.73–7.69 (m, 1H), 7.33 (ddt, $J = 8.9, 4.5, 2.3$ Hz, 1H), 7.29 ($d, J = 1.7$ Hz, 1H), 6.90 ($t, J = 9.1$ Hz, 1H), 6.22 ($dd, J = 3.1, 1.9$ Hz, 1H), 6.07 ($d, J = 3.1$ Hz, 1H), 4.38 ($d, J = 20.3$ Hz, 1H), 4.13–4.04 (m, 2H), 3.98 (ddd, $J = 12.6, 7.7, 5.8$ Hz, 2H), 3.76 ($d, J = 14.6$ Hz, 1H), 3.55 ($d, J = 14.6$ Hz, 1H), 1.25 ($t, J = 7.1$ Hz, 3H), 1.16 ($t, J = 7.1$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 161.38, 158.91, 152.21, 142.13, 132.26, 125.63, 117.00, 110.12, 107.91, 99.99, 77.50, 77.18, 76.86, 63.06, 51.97, 50.40, 43.97, 16.20; HRMS (ESI) m/z calcd for $\text{C}_{16}\text{H}_{20}\text{BrFNO}_4\text{P}^+$ (M + H), 419.0297; found, 419.0296.

4.2.7. Dimethyl (((Furan-2-ylmethyl)amino)(1*H*-indol-3-yl)methyl)phosphonate (5g). Orange yellow solid, 90% yield (301 mg); FT-IR (ZnSe): ν_{max} 3168, 2923, 2852, 1646, 1449, 1185, 1037, 731; ^1H NMR (400 MHz, CDCl_3): δ 9.49 ($d, J = 23.7$ Hz, 1H), 7.69–7.60 (m, 1H), 7.42–7.35 (m, 2H), 7.24–7.10 (m, 3H), 6.38–6.22 (m, 2H), 6.10 ($d, J = 3.2$ Hz, 1H), 4.47 ($d, J = 17.4$ Hz, 1H), 3.96 ($d, J = 10.5$ Hz, 1H), 3.76 ($d, J = 10.5$ Hz, 3H), 3.53 ($d, J = 10.4$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3): δ 152.95, 142.04, 136.33, 126.85, 122.10, 119.71, 119.13, 118.66, 111.66, 110.15, 108.69, 107.85, 104.25, 100.00, 77.40, 77.09, 76.77, 53.46, 43.61; HRMS (ESI) m/z calcd for $\text{C}_{16}\text{H}_{19}\text{N}_2\text{O}_4\text{P}^+$ (M + H), 334.1082; found, 334.1078.

4.2.8. Dimethyl (benzo[d][1,3]dioxol-5-yl((furan-2-ylmethyl)amino)methyl)phosphonate (5h). Pale yellow solid, 83% yield (281.7 mg); FT-IR (ZnSe): ν_{max} 3218, 2945, 1620, 1232, 1025, 820, 746; ^1H NMR (400 MHz, CDCl_3): δ 7.24 ($d, J = 5.9$ Hz, 2H), 6.85 ($d, J = 1.5$ Hz, 2H), 6.71 (dd, $J = 20.1, 12.2$ Hz, 4H), 6.14 ($d, J = 1.8$ Hz, 2H), 5.98 ($t, J = 9.9$ Hz, 2H), 5.79 (s, 4H), 4.86 ($d, J = 87.0$ Hz, 1H), 3.85 ($d, J = 19.0$ Hz, 2H), 3.58 (s, 6H), 3.47 (s, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 152.57, 147.92, 147.44, 141.99, 128.49, 122.28, 110.04, 108.62, 108.15, 107.68, 101.11, 59.08, 57.53, 53.42, 43.10; HRMS (ESI) m/z calcd for $\text{C}_{14}\text{H}_{18}\text{NO}_6\text{P}^+$ (M + H), 339.0872; found, 339.0874.

4.2.9. Dimethyl (((Furan-2-ylmethyl)amino)(4-hydroxy-3-methoxyphenyl)methyl)phosphonate (5i). Yellow solid, 93% yield (317.2 mg); FT-IR (ZnSe): ν_{max} 3428, 3168, 2942, 1693, 1497, 1178, 1032, 732; ^1H NMR (400 MHz, CDCl_3): δ 7.28 ($d, J = 4.6$ Hz, 2H), 7.04 (s, 1H), 6.93 (s, 1H), 6.22 ($d, J = 1.3$ Hz, 1H), 6.04 (s, 1H), 3.90 ($d, J = 18.9$ Hz, 1H), 3.79 (s, 3H), 3.74 ($d, J = 14.5$ Hz, 2H), 3.67 ($d, J = 10.6$ Hz, 3H), 3.55 ($d, J = 10.5$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 152.31, 147.92, 143.47, 142.14, 126.81, 124.91, 110.15, 108.57, 107.97, 58.69, 57.13, 56.35, 53.80, 53.65, 43.37; HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{20}\text{NO}_6\text{P}^+$ (M + H), 341.1028; found, 341.1026.

4.2.10. Dimethyl((3-bromo-4-hydroxy-5-methoxyphenyl)((furan-2-ylmethyl)amino)methyl)phosphonate (5j). Orange

red solid, 85% yield (357.1 mg); FT-IR (ZnSe): ν_{max} 3373, 3113, 2949, 1603, 1515, 1455, 1203, 1029, 743; ^1H NMR (400 MHz, CDCl_3): δ 7.24 (*d*, $J = 7.6$ Hz, 1H), 6.92 (*s*, 1H), 6.83–6.68 (*m*, 2H), 6.17 (*d*, $J = 5.1$ Hz, 1H), 6.00 (*d*, $J = 4.9$ Hz, 1H), 3.90 (dd, $J = 18.6, 7.3$ Hz, 1H), 3.76–3.33 (*m*, 11H), ^{13}C NMR (100 MHz, CDCl_3): δ 152.55, 147.49, 146.23, 142.02, 125.49, 121.76, 121.70, 115.01, 111.28, 110.07, 107.79, 77.72, 77.41, 77.05, 59.00, 57.45, 55.80, 53.66, 43.08; HRMS (ESI) *m/z* calcd for $\text{C}_{15}\text{H}_{19}\text{BrNO}_6\text{P}^+$ ($M + \text{H}$), 419.0133; found, 419.0131.

4.3. Antioxidant Activity. **4.3.1. DPPH Radical Scavenging Activity.** In practice, the electron donation capacity or hydrogen atom donation ability of a compound is confirmed from the bleaching ability of purple color of methanolic DPPH solution to colorless. In this spectrophotometric study, 1 mL of the test sample of compounds that are to be screened for antioxidant activity with different concentrations (50, 100, and 200 $\mu\text{g}/\text{mL}$) was prepared and added to 4 mL of 0.004% (w/v) methanolic DPPH solutions.^{72–76} Then, the analytical mixture was mixed thoroughly for 1 min and then incubated at room temperature for a period of 30 min. After completion of incubation process, the absorbance of all the test samples was measured at 517 nm and was referenced to ascorbic acid reference standards. Finally, DPPH radical scavenging activity was obtained from the following equation.

$$\% \text{ of inhibition} = \frac{[(A_{\text{control}} - A_{\text{test}})]}{(A_{\text{control}})} \times 100$$

4.3.2. H_2O_2 Radical Scavenging Activity. To the 1 mL test solution of different concentrations (50, 100, and 200 $\mu\text{g}/\text{mL}$) of the synthesized compounds of the study were added with 3.75 mM of deoxyribose, 1.0 mM of H_2O_2 , 20 mM of potassium phosphate buffer (7.4 pH), 0.1 mM of FeCl_3 , and 0.1 mM of EDTA and incubated for a period of 1 h at 37 °C temperature in a water bath. The degree of degradation of deoxyribose was measured by adding thiobarbituric acid (1.0%) and trichloroacetic acid (2.0%) and heating them in a water bath at 100 °C for a period of 20 min. Eventually, the absorbance of the resulting test solutions was obtained spectrophotometrically at 532 nm from the following equation.

$$\% \text{ of inhibition} = \frac{[(A_{\text{control}} - A_{\text{test}})]}{(A_{\text{control}})} \times 100$$

4.4. Plant Growth Regulatory Activity. Seeds were germinated at 25 °C for 48 h in 12 h light and 12 h darkness. Groups of 50 uniform well-sterilized seedlings were transferred to 10 cm Petri dishes that are outlined with two discs of Whatman no. 1 filter paper wetted with 2 mL of 0.001 M aqueous methanol (control) and 2 mL of solutions of the test compounds (**5a–j**) at 50 and 100 ppm concentrations. The plants were grown in simultaneous 16 h light and 8 h dark conditions at 25 ± 2 °C for 48 h and the lengths of roots and stem were measured. The results are presented as the average increase or decrease in root length and shoot length compared with the control. Each experiment was repeated three times.

4.5. Molecular Docking Studies. The mechanistic inhibition of propagating DPPH and H_2O_2 free radicals *in vivo* by title compounds **5a–j** has been determined by studying the molecular docking interactions. As such, the three-dimensional crystalline structural information of identified proteins of the study were attained from protein data bank (as pdb files) and reinvigorated them by removing bound water,

ligands, and other cofactors present in their vicinity. The .mol2 and .pdb database files of **5a–j** ligands were produced from ChemBioOffice software (ChemBio3D ultra 14.0 suite) and docked them for the selected proteins on Swiss Dock software.⁷⁷ The interactive docking poses were considered and captured at energy-minimized optimization conditions with 0.100 of minimum root-mean standard deviation (RMSD) gradient and the interacting binding postures are imagined through UCSF Chimera.⁷⁸ The potential and highest binding energies originated with proper ligand–protein interactions of **5a–j** with target protein receptors in chain C of 1DNU protein and chain A of 4M7E protein, respectively, for the antioxidant and plant growth regulatory activities and were accounted in macromolecular structural atmosphere and acknowledged that compounds **5a–j** as efficiently bound with the proteins considered for the study and presented in Tables 3–6.

4.6. ADMET Properties. The ADMET properties of synthesized compounds **5a–j** have been calculated on preADMET online server^{79,80} and which assisted us to understand their potentialities like *in vivo* BBB penetration, % HIA, *in vitro* Caco-2 cell permeability, *in vitro* Maden Darby Canine kidney (MDCK) cell permeability, and *in vitro* PPB properties. In continuance, the prediction of tumorigenic, mutagenic, reproductive, and irritant effects has assisted in ascertaining the toxicity properties. The BBB is confined with profoundly bound endothelial cells that constrain a compound's ability to be conceded into the blood stream *via* the directed route. The analysis of BBB penetration rate (in terms of concentration ration, *i.e.*, [brain]/[blood]) supported us to inspect the ability of an entity to penetrate over BBB, which is dynamic in assigning CNS activity to the libraries of pharmacophoric assets of a compound. The compounds proficient to pass *via* BBB are denoted as CNS active (where, BBB penetration rate >0.40) and unable to pass are denoted as CNS inactive (where, BBB penetration rate < 0.40). Similarly, Caco-2 (human colon adenocarcinoma based cells) cells associates by an intestinal epithelium system in multiple drug transport avenues such as transcellular, paracellular, and active efflux transports. The compounds with *in vitro* Caco-2 cell permeability value <4 are assigned as poor, the permeability in the range of 4–70 is assigned as moderate, and permeability value >70 is assigned as extremely permeable and are efficiently transported to biochemical and cellular processes. Moreover, the extent of PPB influences distribution level of unbound quantity of compound in body tissues and distributed over cellular sites of action and hence stimulates to metabolize and excrete from systems. The % PPB *in vitro* classifies the compounds as strong if % PPB > 90% and as weak if % PPB < 90%, which eventually reflects its potentiality. Moreover, the MDCK cell system is recognized as a sensible system to filter out the prompt permeability of a compound and to decide its potentiality as cellular life span is lower than the life span of Caco-2 cells and therefore the correspondence stands high. Mostly, *in vitro* MDCK permeability <25 describes compound as poorly permeable, permeability in the range of 25–500 designated as sensibly permeable, and permeability >500 designated as greatly permeable. Likewise, the % HIA is deliberated as the percentage of an orally administered drug compound into the hepatic portal vein. Generally, it is accounted for ample bioavailability and absorption measured based on the ratio of cumulative excretion in bile, urine, and feces. The % HIA ranging from 0 to 20 entitles poor, 20–70

entitles moderate, and 70–100 entitles good absorbance of a compound under study. Coming to the toxicology perspectives, it is important to consider the toxicology properties of an entity under study so as to ignore the toxicity risks during the design itself. The negative magnitude of toxicology for a compound is the outcome of its safer drug property as they are reclining to carcinogenicity, mutagenicity, and HERG (human ether-a-go-go related gene) channel inhibition and its administration *in vivo* as presented in Table 7. This type of complementary study ascertains a chemical entity as safer drugs.

4.7. QSAR Studies. Many proposed drugs do not come to the clinical stage because of their poor ADMET liabilities, but the worthy oral bioavailability can be realized them by right poise amidst its partition and solubility.⁸¹ Computationally, Lipinski's rule of five⁸² is considered for screening newer molecules to testify their potentiality as it states that (i) molecular weight ≤ 500 Da; (ii) octanol/water partition coefficient ($\log P$) ≤ 5 ; (iv) number of hydrogen bond donors ≤ 5 ; and (iii) number of hydrogen bond acceptors ≤ 10 and summarizes the Lipinski parameters (molecular weight, hydrogen bond acceptors, hydrogen bond donors, molecular refractivity, and octanol to water partition coefficient). The drug-like properties predicted by Molinspiration⁸³ are very sturdy and work on the summation of fragment contributions and handling most of the organic compounds to compute and to know the structural sensitivity in visualizing the significant molecular properties like Veber parameters (number of rotatable bonds and total polar surface area in addition to Lipinski parameters) and other parameters like number of hydrophobic atoms; van der Waals volume, density, solubility, calculated octanol to water partition coefficient and percentage of absorption) has been considered and all the quantitative structure–activity relationship descriptors were obtained and presented in Table 8.

In this QSAR analysis study, molecular descriptors and fingerprints both play an important role as they are mathematical representations of chemical entities and serves as the data analysis methodological input sources in building QSAR models.^{84–86} In detail, molecular descriptors are the experimental results of a molecule according to their theoretically derived properties, which helps in understanding the molecular structure and biological activity of the compounds under study from different facets.^{87–89} In complementary molecular descriptors and fingerprint properties, both portray a vital role in QSAR analysis and in turn in the drug discovery process.^{90–93} The fingerprint properties of 5a–j have been predicted from the ChemDes server,^{94,95} an integrated aid for the computing the results. Here, the QSAR results were predicted in MOPAC Cartesian format, a semiempirical molecular orbital package to scrutinize molecular structures and is widely accepted for the prediction of QSAR and other drug discovery studies. These computational chemical studies were performed by modified neglect of diatomic overlap, a semiempirical approach for calculating molecular electronic structural quantum, which is designed on neglect of diatomic differential overlap integral approximation process and the results are presented in Table 9.

4.8. Bioactivity and Toxicity Risk Studies. The drug-like property profiles of compounds 5a–j have been assessed on Molinspiration cheminformatics software tool,⁶⁶ viz., v2018.10 Molinspiration engine for property exploration and v2018.03 Molinspiration engine for bioactivity score evaluations. This

evaluation revealed the bioactivity and nontoxic properties such as KI (kinase inhibitor), GPCR (G protein-coupled receptor) ligand property, ICM (ion channel modulator), and NRL (nuclear receptor ligand) interactions, and EI (enzyme inhibitor) and PI (protease inhibitor) properties, drug-likeness, and drug scores and mutagenic, tumorigenic, irritant, and reproductive effect of the synthesized compounds have been evaluated and proved as safer drugs. On the other hand, drug properties mounted with the multifarious offset of a set of structural features and molecular properties, and its evaluation helps to evaluate the close approximation of a molecule under investigation with the notorious drugs. Hence, these toxicity risk profiles and drug property parameters have predicted by the Osiris online property explorer toolkit⁶⁹ and consolidated the properties under the head of bioactivity and toxicity risk studies and are presented in Table 10. This study helped in considering the physicochemical interactions of the synthesized compounds against the targeted domains and eventually facilitated for determining corresponding drug properties by comparing with the bioactivity properties and toxicity risk profile studies.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acsomegajournal.0c05302>.

Spectra of compounds 5a–j (PDF)

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Notes

The authors declare no competing financial interest.

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