

# System drift and speciation

July 5, 2021

## Abstract

Even if a species' phenotype does not change over evolutionary time, the underlying mechanism may change, as distinct molecular pathways can realize identical phenotypes. Here we use linear system theory to explore the consequences of this idea, describing how a gene network underlying a conserved phenotype evolves, as the genetic drift of small changes to these molecular pathways cause a population to explore the set of mechanisms with identical phenotypes. To do this, we model an organism's internal state as a linear system of differential equations for which the environment provides input and the phenotype is the output, in which context there exists an exact characterization of the set of all mechanisms that give the same input–output relationship. This characterization implies that selectively neutral directions in genotype space should be common and that the evolutionary exploration of these distinct but equivalent mechanisms can lead to the reproductive incompatibility of independently evolving populations. This evolutionary exploration, or *system drift*, is expected to proceed at a rate proportional to the amount of intrapopulation genetic variation divided by the effective population size ( $N_e$ ). At biologically reasonable parameter values this could lead to substantial interpopulation incompatibility, and thus speciation, on a time scale of  $N_e$  generations. This model also naturally predicts Haldane's rule, thus providing another possible explanation of why heterogametic hybrids tend to be disrupted more often than homogametes during the early stages of speciation.

**Key words:** Speciation, Models/Simulations, Genetic Drift

## Introduction

It is an overarching goal of many biological subdisciplines to attain a general understanding of the function and evolution of the complex molecular machinery that translates an organism's genome into the characteristics on which natural selection acts, the phenotype. For example, there is a growing body of data on the evolutionary histories and molecular characterizations of particular gene regulatory networks [Jaeger, 2011, Davidson and Erwin, 2006, Israel et al., 2016], as well as thoughtful verbal and conceptual models [True and Haag, 2001, Weiss and Fullerton, 2000, Edelman and Gally, 2001, Pavlicev and Wagner, 2012]. Mathematical models of both particular regulatory networks and the evolution of such systems in general can provide guidance where intuition fails, and thus have the potential to discover general principles in the organization of biological systems as well as provide concrete numerical predictions [Servedio et al., 2014]. There is a substantial amount of work studying the evolution of gene regulatory networks, in frameworks both abstract [Wagner, 1994, 1996, Siegal and Bergman, 2002, Bergman and Siegal, 2003, Draghi and Whitlock, 2015] and empirically inspired [Mjolsness et al., 1991, Jaeger et al., 2004, Kozlov et al., 2015, Crombach et al., 2016, Wotton et al., 2015, Chertkova et al., 2017].

At all levels of biological organization, the problems that biological systems have evolved to solve often do not have single solutions – systems can be structurally different yet remain functionally equivalent [Edelman and Gally, 2001]. Examples can be found across nearly all levels of biological organization from the level of the genetic code itself all the way up to the convergent evolution of adaptive traits. In many cases, these functionally equivalent structures can be explored through small, local changes to the structure that leave the function unchanged. For instance, there are “neutral networks” of nucleic acid sequences that produce the same RNA secondary structure [Grüner et al., 1996], amino acid sequences that fold similarly [Babajide et al., 1997], or proteins with equivalent thermodynamic stability [Hart et al., 2014]. Further examples are found in the vast space of functionally equivalent potential regulatory sequences [Hare et al., 2008], in the logic of transcriptional [Tsong et al., 2006, Matsui et al., 2015, Dalal et al., 2016, Dalal and Johnson, 2017, Jiménez et al., 2017] and neural circuits [Trojanowski et al., 2014], and in developmental systems [von Dassow et al., 2000, True and Haag, 2001].

This capacity for isofunctional yet distinct mechanisms, sometimes called *degeneracy*, is a consequence of a many-to-one mapping between a system’s structure and function, a concept that has been explored in many fields beyond biology. For instance, in many contexts mathematical models are fundamentally *nonidentifiable* and/or *indistinguishable* – meaning that there can be uncertainty about an inferred model’s parameters or even its claims about causal structure, despite access to complete and perfect data [e.g., Bellman and Åström, 1970, Grewal and Glover, 1976, Walter et al., 1984]. Models with different parameter schemes, or even different mechanics can make equally accurate predictions, but still not actually reflect the internal dynamics of the system being modeled. In control theory, where electrical circuits and mechanical systems are often the focus, it is understood that there can be an infinite number of “realizations”, or ways to reverse engineer the dynamics of a “black box”, even if all possible input and output experiments are performed [Kalman, 1963, Anderson et al., 1966, Zadeh and Deoser, 1976]. The inherent nonidentifiability of chemical reaction networks is sometimes referred to as “the fundamental dogma of chemical kinetics” [Craciun and Pantea, 2008]. In computer science, this has been framed as the relationship among processes that *simulate* one another [Van der Schaft, 2004]. Finally, the field of *inverse problems* studies those cases in which, despite the existence of a theoretical one-to-one mapping between a model and behavior, tiny amounts of noise make inference problems nonidentifiable in practice [Petrov and Sizikov, 2005].

It has been argued that the ability to modify structure without affecting function is necessary for natural selection [Edelman and Gally, 2001], as it may function as a mechanism for biological robustness and evolvability [reviewed in de Visser et al., 2003], or manifest as *canalization* [Whitacre, 2010]. It may even contribute to the formation of new species [Gavrilets, 2014]. Redundancy of the genetic code, for instance, can make sequences more fault-tolerant to mutations [Sonneborn, 1965], and robustness to modification of genetic networks can allow adaptation without passing through a fitness valley [Wagner, 2008].

In this paper we use results on mathematical nonidentifiability from linear systems theory to study how gene regulatory networks can be modified while retaining the same function, and the possible implications for speciation. If system architectures are not functionally unique, can this open up neutral evolutionary paths, and do species explore these paths through the process termed *developmental system drift* [by True and Haag, 2001]? Is this fast enough to contribute meaningfully to speciation? To do this, we describe results on linear dynamical systems which give an analytical description of the set of all linear gene network architectures that yield identical phenotypes, and use quantitative genetics theory to estimate the speed at which system drift can lead to reproductive incompatibility and hence speciation. In this model, a population diffuses along the neutral ridges of a high-dimensional space of possible system parameters, in a similar vein as *holey landscape* models [Gavrilets, 1997, Yamaguchi and Iwasa, 2013, Pina and Schertzer, 2019].

The field of population genetics has also explored the consequences of the fact that there is often more than one way to do the same thing, and observed that speciation might be the result of changes that are themselves neutral. Indeed, Bateson [1909] first proposed that what today we call a Bateson-Dobzhansky-Muller incompatibility would likely arise through neutral changes. The potential for speciation has been analyzed in models of traits under stabilizing selection determined additively by alleles at many loci [Wright, 1935, Barton, 1986, 1989, 2001], in related fitness landscape models [Fraïsse et al., 2016], and for pairs of traits that must match but whose value is unconstrained [Sved, 1981]. It has also been shown that population structure can allow long-term stable coexistence of incompatible genotypes encoding identical phenotypes [Phillips, 1996]. However, previous simulations of system drift in regulatory sequences [Tulchinsky et al., 2014] and a regulatory cascade [Porter and Johnson, 2002] found rapid speciation under directional selection but only equivocal support for speciation under models of purely neutral drift. The rate at which hybrid incompatibility accumulates due to genetic drift creating segregation variance between isolated populations is fairly well understood [Slatkin and Lande, 1994, Rosas et al., 2010, Chevin et al., 2014], but model assumptions can strongly affect predictions, including whether variation is due to rare or common alleles [Slatkin and Lande, 1994], and the shape of the fitness landscape [Fraïsse et al., 2016]. Our main aim is to provide a concrete framework that can provide natural predictions of these model parameters across a general class of models. Furthermore, tools from system theory allow analytical predictions to be made for large populations with complex phenotypes that would be inaccessible to population simulations.

## Results

We use a model of gene regulatory networks that describes the temporal dynamics of a collection of  $n$  coregulating molecules – such as transcription factors – as well as external or environmental inputs. We write  $\kappa(t)$  for the vector of  $n$  molecular concentrations at time  $t$ . The vector of  $m$  “inputs” determined exogenously to the system is denoted  $u(t)$ , and the vector of  $\ell$  “outputs” is denoted  $\phi(t)$ . The output is merely a linear function of the internal state:  $\phi_i(t) = \sum_j C_{ij} \kappa_j(t)$  for some matrix  $C$ . Since  $\phi$  is what natural selection acts on, we refer to it as the *phenotype* (meaning the “visible” aspects of the organism), and in contrast refer to  $\kappa$  as the *kryptotype*, as it is “hidden” from direct selection. Although  $\phi$  may depend on all entries of  $\kappa$ , it is usually of lower dimension than  $\kappa$ , and we tend to think of it as the subset of molecules relevant for survival. The dynamics are determined by the matrix of regulatory coefficients,  $A$ , a time-varying vector of inputs  $u(t)$ , and a matrix  $B$  that encodes the effect of each entry of  $u$  on the elements of the kryptotype. The rate at which the  $i^{\text{th}}$  concentration changes is a weighted sum of the concentrations as well as the input:

$$\begin{aligned}\dot{\kappa}(t) &= A\kappa(t) + Bu(t) \\ \phi(t) &= C\kappa(t).\end{aligned}\tag{1}$$

Furthermore, we always assume that  $\kappa(0) = 0$ , so that the kryptotype measures deviations from initial concentrations. Here  $A$  can be any  $n \times n$  matrix,  $B$  any  $n \times m$ , and  $C$  any  $\ell \times n$  dimensional matrix, with usually  $\ell$  and  $m$  less than  $n$ . We think of the system as the triple  $(A, B, C)$ , which translates (time-varying)  $m$ -dimensional input  $u(t)$  into the  $\ell$ -dimensional output  $\phi(t)$ . Under quite general assumptions on the input (e.g.,  $|u(t)|$  is integrable) we can write the phenotype as

$$\phi(t) = \int_0^t C e^{(t-s)A} B u(s) ds,\tag{2}$$

which is a convolution of the input  $u(t)$  with the system’s *impulse response*, which we denote as  $h(t) := C e^{At} B$ .

In terms of gene regulatory networks,  $A_{ij}$  determines how the  $j^{\text{th}}$  transcription factor regulates the  $i^{\text{th}}$  transcription factor. If  $A_{ij} > 0$ , then  $\kappa_j$  upregulates  $\kappa_i$ , while if  $A_{ij} < 0$ , then  $\kappa_j$  downregulates  $\kappa_i$ . The  $i^{\text{th}}$  row of  $A$  is therefore determined by genetic features such as the strength of  $j$ -binding sites in the promoter of gene  $i$ , factors affecting chromatin accessibility near gene  $i$ , or basal transcription machinery activity. The form of  $B$  determines how the environment influences transcription factor expression levels, and  $C$  might determine the rate of production of downstream enzymes.

Wagner [1994] and others have used a similar discrete-time model (that might be written  $\phi_{t+1} = f(A\phi_t)$ , where  $f$  is a sigmoid). Our choice of continuous time does not affect the points we make here, but our restriction to *linear* systems is a stronger assumption (see the Discussion).

To demonstrate the model, we construct a simple gene network in Example 1 below.

**Example 1** (An oscillator). *For illustration, we consider an extremely simplified model of oscillating gene transcription, as for instance is found in cell cycle control or the circadian rhythm. There are two genes, whose transcript concentrations are given by  $\kappa_1(t)$  and  $\kappa_2(t)$ , and gene-2 upregulates gene-1, while gene-1 downregulates gene-2 with equal strength. Only the dynamics of gene-1 are consequential to the oscillator (perhaps the amount of gene-1 activates another downstream gene network). Lastly, both genes are equally upregulated by an exogenous signal. The dynamics of the system are described by*

$$\begin{aligned}\dot{\kappa}_1(t) &= \kappa_2(t) + u(t) \\ \dot{\kappa}_2(t) &= -\kappa_1(t) + u(t) \\ \phi(t) &= \kappa_1(t).\end{aligned}$$

In matrix form the system regulatory coefficients are given as,  $A = \begin{bmatrix} 0 & 1 \\ -1 & 0 \end{bmatrix}$ ,  $B = \begin{bmatrix} 1 \\ 1 \end{bmatrix}$ , and  $C = \begin{bmatrix} 1 & 0 \end{bmatrix}$ . If the input is an impulse at time zero (a delta function), then the phenotype is equal to the impulse response:

$$\phi(t) = h(t) = \sin t + \cos t.$$

135 The system and its dynamics are referred to in Figure 1. We return to the evolution of such a system below.



Figure 1: (Left) Diagram of the gene network in Example 1, and (right) plot of the phenotype  $\phi(t)$  against time  $t$ .

## 136 Equivalent gene networks

137 As reviewed above, some systems with identical phenotypes are known to differ, sometimes substantially, at  
 138 the molecular level; systems with identical phenotypes do not necessarily have identical kryptotypes. How  
 139 many different mechanisms perform the same function?

140 Two systems are equivalent if they produce the same phenotype given the same input, i.e., have the same  
 141 input–output relationship. We say that the systems defined by  $(A, B, C)$  and  $(\bar{A}, \bar{B}, \bar{C})$  are **phenotypically**  
 142 **equivalent** if their impulse response functions are the same:  $h(t) = \bar{h}(t)$  for all  $t \geq 0$ . This implies that for  
 143 any acceptable input  $u(t)$ , if  $(\kappa_u(t), \phi_u(t))$  and  $(\bar{\kappa}_u(t), \bar{\phi}_u(t))$  are the solutions to equation (1) of these two  
 144 systems, respectively, then

$$\phi_u(t) = \bar{\phi}_u(t) \quad \text{for all } t \geq 0.$$

145 In other words, phenotypically equivalent systems respond identically for *any* input.

146 One way to find other systems phenotypically equivalent to a given one is by change of coordinates: if  $V$   
 147 is an invertible matrix, then the systems  $(A, B, C)$  and  $(VAV^{-1}, VB, CV^{-1})$  are phenotypically equivalent  
 148 because their impulse response functions are equal:

$$\begin{aligned} h(t) &= Ce^{At}B = CV^{-1}Ve^{At}V^{-1}VB \\ &= CV^{-1}e^{VAV^{-1}t}VB = \bar{C}\bar{e}^{\bar{A}t}\bar{B} = \bar{h}(t). \end{aligned} \tag{3}$$

149 These “changes of coordinates” are not simply different ways of looking at the same system – if each  
 150 dimension of the kryptotype corresponds to the concentration of a particular transcription factor, changing  
 151  $A$  corresponds to changing the strengths of regulatory interactions. We will even see below that interactions  
 152 may change sign. However, not all phenotypically equivalent systems are of this form: systems can have  
 153 identical impulse responses without being coordinate changes of each other. In fact, systems with identical  
 154 impulse responses can involve interactions between different numbers of molecules, and thus have kryptotypes  
 155 in different dimensions altogether.

156 This implies that most systems have at least  $n^2$  degrees of freedom, where recall  $n$  is the number of  
 157 components of the kryptotype vector. This is because for an arbitrary  $n \times n$  matrix  $Z$ , taking  $V$  to be the  
 158 identity matrix plus a small perturbation in the direction of  $Z$  above implies that moving  $A$  in the direction  
 159 of  $ZA - AZ$  while also moving  $B$  in the direction of  $ZB$  and  $C$  in the direction of  $-CZ$  will leave the  
 160 phenotype unchanged to second order in the size of the perturbation. If the columns of  $B$  and the rows of  
 161  $C$  are not all eigenvectors of  $A$ , then any such  $Z$  will result in a different system.

It turns out that in general, there are more degrees of freedom, except if the system is *minimal* – meaning, informally, that it uses the smallest possible number of components to achieve the desired dynamics. Results in system theory show that any system can be realized in a particular minimal dimension (the dimension of the kryptotype,  $n_{\min}$ ), and that any two phenotypically equivalent systems of dimension  $n_{\min}$  are related by a change of coordinates. Since gene networks can grow or shrink following gene duplications and deletions, these additional degrees of freedom can apply, in principle, to any system.

Even if the system is not minimal, results from systems theory explicitly describe the set of all phenotypically equivalent systems. We refer to  $\mathcal{N}(A_0, B_0, C_0)$  as the set of all systems phenotypically equivalent to the system defined by  $(A_0, B_0, C_0)$ :

$$\mathcal{N}(A_0, B_0, C_0) = \{(A, B, C) : Ce^{At}B = C_0e^{A_0t}B_0 \text{ for } t \geq 0\}. \quad (4)$$

These systems need not have the same kryptotypic dimension  $n$ , but must have the same input and output dimensions ( $\ell$  and  $m$ , respectively).

The Kalman decomposition, which we now describe informally, elegantly characterizes this set [Kalman, 1963, Kalman et al., 1969, Anderson et al., 1966]. To motivate this, first note that the input  $u(t)$  only directly pushes the system in certain directions (those lying in the span of the columns of  $B$ ). As a result, different combinations of input can move the system in any direction that lies in what is known as the *reachable subspace*. Analogously, we can only observe motion of the system in certain directions (those lying in the span of the rows of  $C$ ), and so can only infer motion in what is known as the *observable subspace*. The Kalman decomposition then classifies each direction in kryptotype space as either reachable or unreachable, and as either observable or unobservable. Only the components that are both reachable and observable determine the system’s phenotype – that is, components that both respond to an input and produce an observable output.

Concretely, the **Kalman decomposition** of a system  $(A, B, C)$  gives a change of basis  $P$  such that the transformed system  $(PAP^{-1}, PB, CP^{-1})$  can be written in block matrix form:

$$PAP^{-1} = \begin{bmatrix} A_{r\bar{o}} & A_{r\bar{o},ro} & A_{r\bar{o},\bar{r}\bar{o}} & A_{r\bar{o},\bar{r}o} \\ 0 & A_{ro} & 0 & A_{ro,\bar{r}o} \\ 0 & 0 & A_{\bar{r}\bar{o}} & A_{\bar{r}\bar{o},\bar{r}o} \\ 0 & 0 & 0 & A_{\bar{r}o} \end{bmatrix},$$

and

$$PB = \begin{bmatrix} B_{r\bar{o}} \\ B_{ro} \\ 0 \\ 0 \end{bmatrix} \quad (CP^{-1})^T = \begin{bmatrix} 0 \\ C_{ro}^T \\ 0 \\ C_{\bar{r}o}^T \end{bmatrix}.$$

The  $n$ -dimensional system has been divided into subspaces of dimensions  $n_{r\bar{o}} + n_{ro} + n_{\bar{r}\bar{o}} + n_{\bar{r}o} = n$ , and so, for instance,  $A_{r\bar{o}}$  is the  $n_{r\bar{o}} \times n_{r\bar{o}}$  square matrix in the top left corner of  $PAP^{-1}$ . The impulse response of the system is given by

$$h(t) = C_{ro}e^{A_{ro}t}B_{ro},$$

and therefore, the system is phenotypically equivalent to the *minimal* system  $(A_{ro}, B_{ro}, C_{ro})$ .

This decomposition is unique up to a change of basis that preserves the block structure. In particular, the minimal subsystem obtained by the Kalman decomposition is unique up to a change of coordinates. This implies that there is no equivalent system with a smaller number of kryptotypic dimensions than the dimension of the minimal system. It is remarkable that the gene regulatory network architecture to achieve a given input–output map is never unique – both the change of basis used to obtain the decomposition and, once in this form, all submatrices other than  $A_{ro}$ ,  $B_{ro}$ , and  $C_{ro}$  can be changed without affecting the phenotype, and so represent degrees of freedom. (However, some of these subspaces may affect how the system deals with noise.)

*Note on implementation:* The *reachable subspace* is defined to be the closure of  $\text{span}(B)$  under applying  $A$  (or equivalently, the span of  $B, AB, A^2B, \dots, A^{n-1}B$ ), and the *unobservable subspace* is the largest  $A$ -invariant subspace contained in the null space of  $C$ . The four subspaces,  $r\bar{o}$ ,  $ro$ ,  $\bar{r}\bar{o}$ , and  $\bar{r}o$  are defined from these by intersections and orthogonal complements –  $ro$  refers to the both *reachable and observable* subspace, while  $\bar{r}\bar{o}$  refers to the *unreachable and unobservable* subspace, and similarly for  $\bar{r}o$  and  $r\bar{o}$ .

For the remainder of the paper, we interpret  $\mathcal{N}$  as the neutral set in the fitness landscape, along which a large population will drift under environmental and selective stasis. This drift need not be purely neutral – for instance, second-order selection on robustness will push the species towards “flatter” areas of genotype space [Rice, 1998, Hermisson et al., 2003]. Even if the phenotype is constrained and remains constant through evolutionary time, the molecular mechanism underpinning it is not constrained and likely will not be conserved.

Finally, note that if  $B$  and  $C$  are held constant – i.e., if the relationships between environment, kryptotype, and phenotype do not change – there are *still* usually degrees of freedom. The following example 2 gives the set of minimal systems equivalent to the oscillator of Example 1, that all share common  $B$  and  $C$  matrices. The oscillator can also be equivalently realized by a three-gene (or larger) network, and will have even more evolutionary degrees of freedom available, as in Figure 3.

**Example 2** (All equivalent rewirings of the oscillator). *The oscillator of example 1 is minimal, and so any equivalent system is a change of coordinates by an invertible matrix  $V$ . If we further require  $B$  and  $C$  to be invariant then we need  $VB = B$  and  $CV = C$ . Therefore the following one-parameter family  $(A(\tau), B, C)$  describes the set of all two-gene systems phenotypically equivalent to the oscillator:*

$$A(\tau) = \frac{-1}{\tau + 1} \begin{bmatrix} -\tau & -1 \\ 2\tau(\tau + 1) + 1 & \tau \end{bmatrix} \text{ for } \tau \neq -1.$$

The resulting set of systems are depicted in Figure 2.

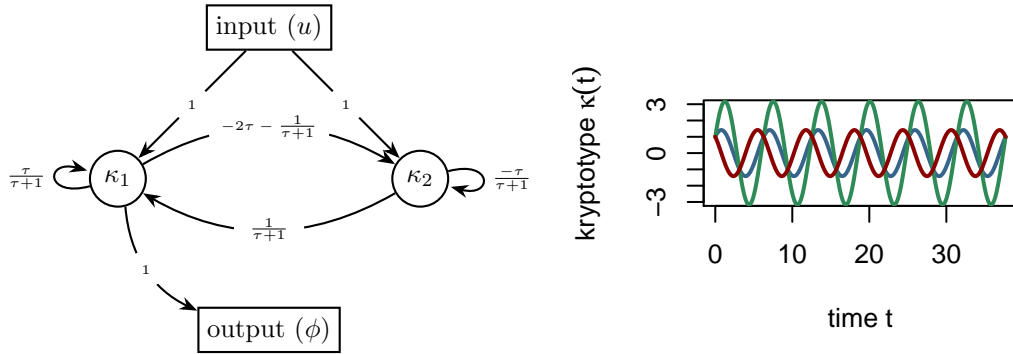


Figure 2: (Left)  $A(\tau)$ , the set of all phenotype-equivalent cell cycle control networks. (Right) Gene-1 dynamics (blue) for both systems  $A(0)$  and  $A(-2)$  are identical, however,  $A(0)$  gene-2 dynamics (red) differ from  $A(-2)$  (green).



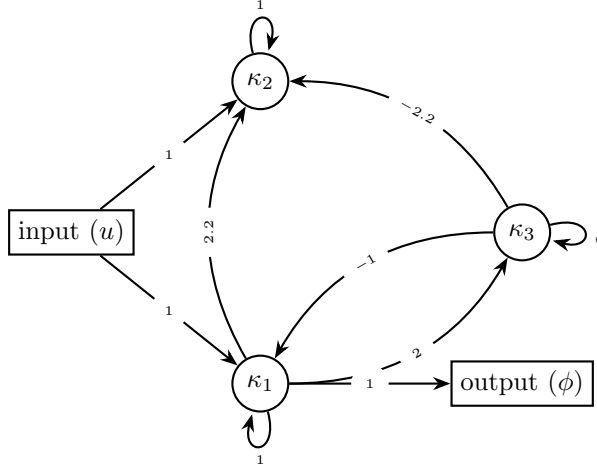


Figure 3: A possible non-minimal three-gene oscillator, phenotypically equivalent to  $A(\tau)$ , the systems in Examples 1 and 2.

**Sexual reproduction and recombination** Parents with phenotypically equivalent yet differently wired gene networks may produce offspring with dramatically different phenotypes. If the phenotypes are significantly divergent then the offspring may be inviable or otherwise dysfunctional, despite both parents being well adapted. If this is consistent for the entire population, we would consider them to be separate species, in accord with the biological species concept [Mayr, 2000].

First, we must specify how sexual reproduction acts on these systems. Suppose that each of a diploid organisms' two genomes encodes a set of system coefficients with the same kryptotype dimension. We assume that a diploid which has inherited systems  $(A', B', C')$  and  $(A'', B'', C'')$  from its two parents has phenotype determined by the system that averages these two,  $((A' + A'')/2, (B' + B'')/2, (C' + C'')/2)$ .

Each genome an organism inherits is generated by meiosis, in which both of its diploid parents recombine their two genomes, and so an  $F_1$  offspring carries one system copy from each parent, and an  $F_2$  is an offspring of two independently formed  $F_1$ s. If the parents are from distinct populations, these are simply first- and second-generation hybrids, respectively.

Exactly how the coefficients (i.e., entries of  $A$ ,  $B$  and  $C$ ) of a haploid system inherited by an offspring from a diploid parent are determined by the parent's two systems depends on the genetic basis of any variation in the coefficients. Thanks to the randomness of meiotic segregation, the result is random to the extent that each parent is heterozygous for alleles that affect the coefficients. Since the  $i^{\text{th}}$  row of  $A$  summarizes how each gene regulates gene  $i$ , and hence is determined by the promoter region of gene  $i$ , the elements of a row of  $A$  tend to be inherited together, which will create covariance between entries of the same row. It is, however, a quite general observation that the variation seen among recombinant systems is proportional to the difference between the two parental systems.

Offspring formed from two phenotypically identical systems do not necessarily exhibit the same phenotype as both of its parents – in other words  $\mathcal{N}$ , the set of all systems phenotypically equivalent to a given one, is not, in general, closed under averaging or recombination. If sexual recombination among systems drawn from  $\mathcal{N}$  yields systems with divergent phenotypes, populations containing significant diversity in  $\mathcal{N}$  can carry genetic load, and isolated populations may fail to produce hybrids with viable phenotypes.

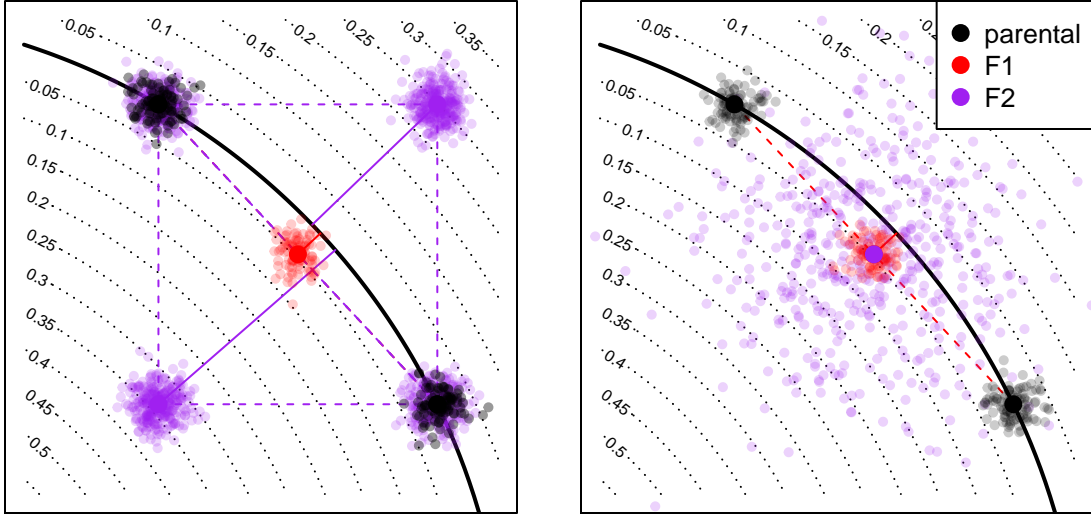


Figure 4: A conceptual figure of the fitness consequences of hybridization: axes represent system coefficients (i.e., entries of  $A$ ); the line of optimal system coefficients is down in black; solid lines give phenotypic distances to the optimum. A pair of parental populations are shown in black, along the optimum; a hypothetical population of  $F_1$ s are shown in red, and the distribution of  $F_2$ s is shown in purple. The two figures differ in the genetic basis, and hence, the distribution of  $F_2$  phenotypes: **(left)**  $F_2$ s compose all four mixed homozygotes if variation at both traits has a simple, one-locus genetic basis in both populations; and **(right)**  $F_2$  show a much wider distribution of phenotypes if the genetic basis of variation in each population is polygenic.

## Hybrid incompatibility

Two parents with the optimal phenotype can produce offspring whose phenotype is suboptimal if the parents have different underlying systems. Hybrid phenotypic break down, as a function of genetic distance between phenotypically equivalent parental oscillators (described in Example 2) is illustrated in Example 3. How quickly do hybrid phenotypes break down as genetic distance between parents increases? We will quantify how far a system's phenotype is from optimal using a weighted difference between impulse response functions. Suppose that  $\rho(t)$  is a nonnegative weighting function,  $h_0(t)$  is the *optimal* impulse response function and define the “distance to optimum” of another impulse response function to be

$$D(h) = \left( \int_0^\infty \rho(t) \|h(t) - h_0(t)\|^2 dt \right)^{1/2}. \quad (5)$$

In practice, we take  $\rho(t) = \exp(-t/4\pi)$ , so that fitness is determined by the dynamics of the system over a few multiples of  $2\pi$ , but not longer. Consider reproduction between a parent with system  $(A, B, C)$  and another displaced by distance  $\epsilon$  in the direction  $(X, Y, Z)$ , i.e., having system  $(A + \epsilon X, B + \epsilon Y, C + \epsilon Z)$ . We assume both are “perfectly adapted” systems, i.e., having impulse response function  $h_0(t)$ , and their offspring has impulse response function  $h_\epsilon(t)$ . A Taylor expansion of  $D(h_\epsilon)$  in  $\epsilon$  is explicitly worked out in Appendix A, and shows that the phenotype of an  $F_1$  hybrid between these two is at distance proportional to  $\epsilon^2$  from optimal, while  $F_2$  hybrids are at distance proportional to  $\epsilon$ . This is because an  $F_1$  hybrid has one copy of each parental system, and therefore lies directly between the parental systems (see Figure 4) – the parents both lie in  $\mathcal{N}$ , which is the valley defined by  $D$ , and so their midpoint only differs from optimal due to curvature of  $\mathcal{N}$ . In contrast, an  $F_2$  hybrid may be homozygous for one parental type in some coefficients and homozygous for the other parental type in others; this means that each coefficient of an  $F_2$  may be equal to either one of the parents, or intermediate between the two; this means that possible  $F_2$  systems may be as



far from the optimal set,  $\mathcal{N}$ , as the distance between the parents. The precise rate at which the phenotype of a hybrid diverges depends on the geometry of the optimal set  $\mathcal{N}$  relative to segregating genetic variation.

**Example 3** (Hybrid incompatibility: misregulation due to system drift). *Offspring of two equivalent systems from Example 2 can easily fail to oscillate. For instance, the  $F_1$  offspring between homozygous parents at  $\tau = 0$  and  $\tau = -2$  has phenotype  $\phi_{F_1}(t) = e^t$ , rather than  $\phi(t) = \sin t + \cos t$ . However, the coefficients of these two parental systems differ substantially, probably more than would be observed between diverging populations. In figure 5 we compare the phenotypes for  $F_1$  and  $F_2$  hybrids between more similar parents, and see increasingly divergent phenotypes as the difference between the parental systems increases. (In this example, the coefficients of  $A(\epsilon)$  differ from those of  $A(0)$  by an average factor of  $1 + \epsilon/2$ ; such small differences could plausibly be caused by changes to promoter sequences.) This divergence is quantified in Figure 6, which shows that mean distance to optimum phenotype of the  $F_1$  and  $F_2$  hybrid offspring between  $A(0)$  and  $A(\epsilon)$  increases with  $\epsilon^2$  and  $\epsilon$ , respectively.*

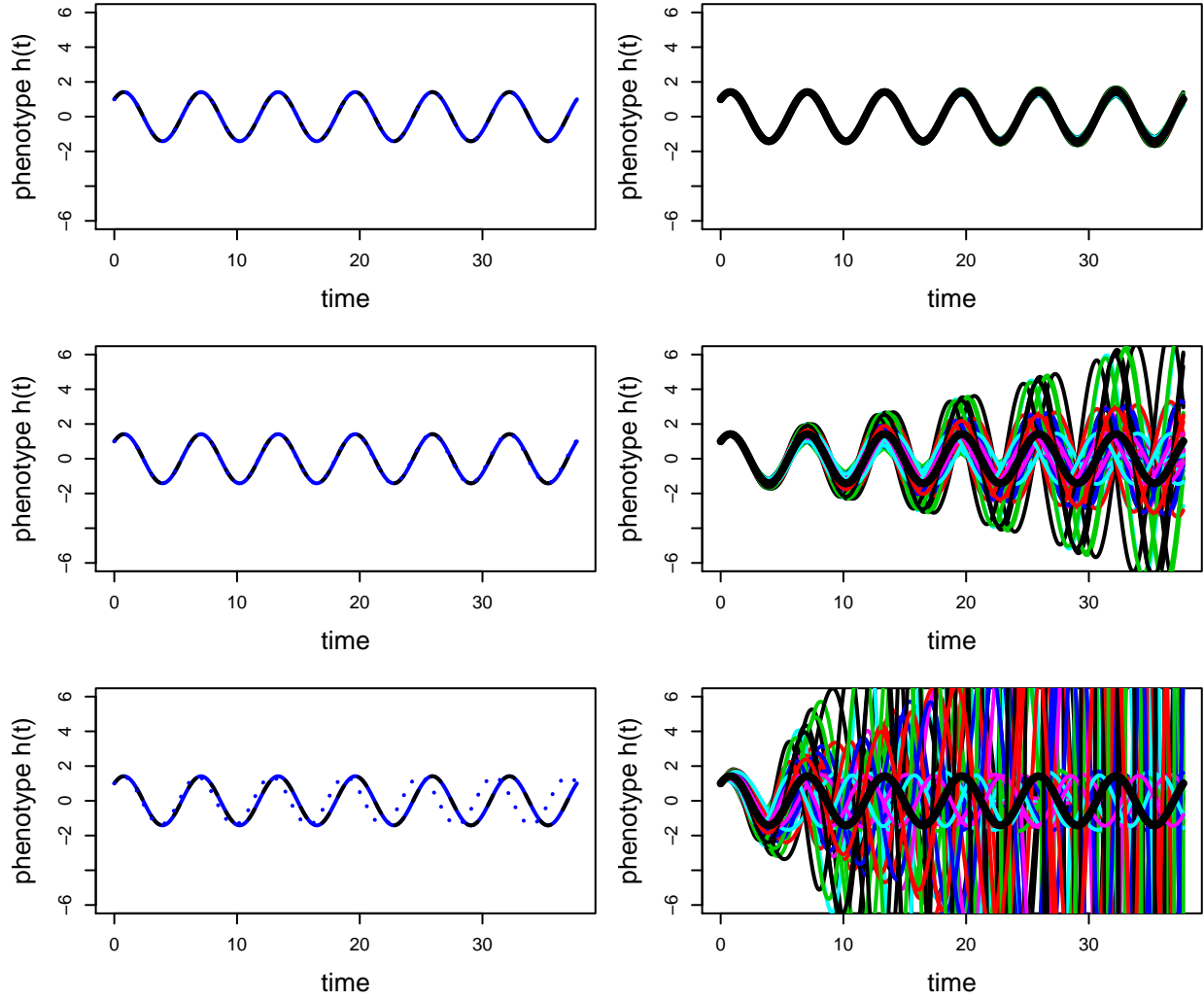


Figure 5: **(left)** Phenotypes of  $F_1$  hybrids between a homozygous  $A(0)$  parent and, top-to-bottom, homozygous  $A(1/100)$ ,  $A(1/10)$ , and  $A(1/2)$  parents, where  $A(\epsilon)$  is defined in figure 2; parental coefficients differ by around 0.5%, 5%, and 25% respectively. Parental phenotypes ( $\sin t + \cos t$ ) are shown in solid black, and hybrid phenotypes in dashed blue. **(right)** Phenotypes of all  $3^4 = 81$  possible  $F_2$  hybrids between the same set of parents, with parental phenotype again in black. Different colored lines correspond to different  $F_2$  hybrids, many of which show complete breakdown.



Figure 6: Mean hybrid phenotypic distance from optimum computed with equation (5), using  $\rho(t) = \exp(-t/4\pi)$  for  $F_1$  (black) and  $F_2$  (blue) hybrids between  $A(0)$  and  $A(\epsilon)$  parent oscillators. Genetic distance is computed as  $\left(\sum_{ij}(A_{ij}(0) - A_{ij}(\epsilon))^2\right)^{1/2}$ .

**Haldane’s rule** This model naturally predicts Haldane’s rule, the observation that if only one hybrid sex is sterile or inviable it is likely the heterogametic sex (e.g., the male in XY sex determination systems) [Haldane, 1922, Orr, 1997]. For example, consider an XY species with a two-gene network where the first gene resides on an autosome and the second gene on the X chromosome. A male whose pair of haplotypes is  $(\begin{bmatrix} A_1 & A_2 \\ X_1 & X_2 \end{bmatrix}, \begin{bmatrix} A_1 & A_2 \\ X_1 & X_2 \end{bmatrix})$  has phenotype determined by  $A = \begin{bmatrix} A_1 & A_2 \\ X_1 & X_2 \end{bmatrix}$ , if dosage compensation upregulates heterogametes by a factor of two relative to homogametes (as with *Drosophila*), while a female homozygous for the haplotype  $\begin{bmatrix} \bar{A}_1 & \bar{A}_2 \\ \bar{X}_1 & \bar{X}_2 \end{bmatrix}$ , has phenotype determined by  $A = \begin{bmatrix} \bar{A}_1 & \bar{A}_2 \\ \bar{X}_1 & \bar{X}_2 \end{bmatrix}$ . An  $F_1$  male offspring of these two will have its phenotype determined by  $\begin{bmatrix} (A_1 + \bar{A}_1)/2 & (A_2 + \bar{A}_2)/2 \\ \bar{X}_1 & \bar{X}_2 \end{bmatrix}$ . If both genes resided on the autosomes, this system would only be possible in an  $F_2$  cross. More generally, if the regulatory coefficients for a system are shared between the sex and one or more autosomal chromosomes,  $F_1$  males are effectively equivalent to purely autosomal-system  $F_2$  hybrids, and recall that  $F_2$ s are significantly less fit on average than  $F_1$ s (see Figure 6). Although many alleles will be dominant if the phenotype–fitness relationship is convex, the underlying mechanism does not depend on the *dominance theory* [Turelli and Orr, 1995] to explain Haldane’s rule: instead, it derives from the nature of segregation variance.

## The speed of speciation

We have shown that system drift can lead to speciation in principle, but is it rapid enough to be an important factor in practice? In other words, after what period of time would we expect the fitness of hybrids between two allopatric populations to be substantially lower than the parentals? Selection – on pleiotropic traits or on robustness – may actively push even a strongly constrained system along neutral directions, but even the calculations under purely neutral drift are informative. The population mean of an unconstrained quantitative trait with additive genetic variance  $V_G$  in a population with effective size  $N_e$  will move in  $t$  generations a random amount whose variance is  $tV_G/N_e$  [Lande, 1976]. The mean difference between two such populations has twice the variance. Although this mean difference is along neutral directions, we would in many cases expect the range of variation among  $F_2$ s in *all* directions to be of the same order as the differences between the populations, as depicted in Figure 4. This suggests that, naively, two such

populations that have been separated for  $t$  generations will produce  $F_2$  offspring that differ from optimal by an amount proportional to  $\sqrt{tV_G/N_e}$ . Since we assume they are at a local fitness optimum, without much loss of generality we can assume that fitness is locally quadratic, and so  $F_2$  fitness decays linearly in time: proportionally to  $tV_G/N_e$  – fastest in small, diverse populations. This predicts that we need only wait some multiple of  $N_e$  generations until substantial incompatibility has been accumulated.

It is useful to think in more detail about the assumptions in the rough argument above. The key aspect is how population differences in neutral directions (along the fitness ridge) translate to segregation variance in  $F_2$ s in selectively constrained directions. To move the system (the  $A$  matrix) a given distance generally involves moving many individual interaction coefficients (the entries  $A_{ij}$ ). The movements must be coordinated, for the population to stay near the fitness ridge. However, mixing elements between systems that have made independent sets of coordinated changes to remain on the fitness ridge is unlikely to produce a set of coordinated changes; and the resulting system could move away from the ridge in almost any direction.

## Genetic variation in empirical regulatory systems

What is known about the key quantity above, the amount of heritable variation in real regulatory networks? The coefficient  $A_{ij}$  from the system (1) measures how much the rate of net production of  $i$  changes per change in concentration of  $j$ . It is generally thought that regulatory sequence change contributes much more to inter- and intraspecific variation than does coding sequence change affecting molecular structure [Schmidt et al., 2010]. In the context of transcription factor networks this may be affected not only by the binding strength of molecule  $j$  to the promoter region of gene  $i$  but also the effects of other transcription factors (e.g., cooperativity) and local chromatin accessibility [Stefflova et al., 2013]. For this reason, the mutational target size for variation in  $A_{ij}$  may be much larger than the dozens of base pairs typically implicated in the handful of binding sites for transcription factor  $j$  of a typical promoter region, and single variants may affect many entries of  $\mathcal{N}$  simultaneously.

Variation in binding site occupancy may overestimate variation in  $A$ , since it does not capture buffering effects (if for instance only one site of many needs to be occupied for transcription to begin), and variation in expression level measures changes in steady-state concentration (our  $\kappa_i$ ) rather than the *rate* of change. Nonetheless, these measures likely give us an idea of the scale of variability. It has been shown that between human individuals, there is differential occupancy in 7.5% of binding sites of a transcription factor (p65) [Kasowski et al., 2010]. It has also been inferred that cis-regulatory variation accounts for around 2–6% of expression variation in human blood-derived primary cells [Verlaan et al., 2009], and that human population variation explained about 3% of expression variation [Lappalainen et al., 2013]. Allele-specific expression is indicative of standing genetic *cis*-regulatory variation; allele-specific expression in 7.2–8.5% of transcripts of a flycatcher species has been observed [Wang et al., 2017], as well as allele-specific expression in 23.4% of genes studied in a baboon species [Tung et al., 2015]. Taken together, this suggests that variation in the entries of  $A$  may be on the order of at least a few percent between individuals of a population – doubtless varying substantially between species and between genes.

## Discussion

In this paper, we use tools from linear system theory and quantitative genetics to study the evolution of a mechanistic model of the genotype-phenotype map, in which the phenotype is subject to stabilizing selection. In so doing, we provide an explicit model of phenogenetic drift [Weiss and Fullerton, 2000] and developmental system drift [True and Haag, 2001]. In this context, the Kalman decomposition [Kalman, 1963] gives an analytical description of all phenotypically equivalent gene networks. This description shows that the space of functionally equivalent network architectures increases with the square of a network’s size, and that this space increases further if networks grow larger than absolutely necessary – that is use more interacting components than the most efficient potential architectures. In this framework, even minimal gene network architectures – efficient architectures that contain only the requisite number of interacting parts, are not structurally unique with respect to function. Functionally equivalent architectures are often related by

continuous parameter changes, suggesting that equivalent networks might be mutationally connected, and that there exist axes of genetic variation unconstrained by natural selection. The independent movement of separated populations along these axes by genetic drift can lead to a significant reduction in hybrid viability, and thus precipitate speciation, at a speed dependent on the effective population size and the amount of genetic variation. In this model, at biologically reasonable parameter values, system drift is a significant – and possibly rapid – driver of speciation. This may be surprising because hybrid inviability appears as a consequence of recombining different, yet functionally equivalent, mechanisms, and since species are often defined by their unique adaptations or morphologies.

Consistent with empirical observation of hybrid breakdown (e.g., Plötner et al. [2017]), we see that the fitnesses of  $F_2$  hybrids drop at a much faster rate than those of  $F_1$ s. Another natural consequence of the model is Haldane’s rule, that if only one  $F_1$  hybrid sex is inviable or sterile it is likely to be the heterogametic sex. This occurs because if the genes underlying a regulatory network are distributed among both autosomes and the sex chromosome, then heterogametic  $F_1$ s show variation (and fitnesses) similar to that seen in  $F_2$  hybrids. This observation appears to be similar to the extreme hybrid phenotypes produced by transgressive segregation [Rieseberg et al., 1999], which can manifest in  $F_1$ s when only one (dominant) parental allele is expressed at heterozygous loci; this was observed in hybrid gene expression patterns, and increased as a function of parental genetic distance [Stelkens and Seehausen, 2009].

Is there evidence that this is actually occurring? System drift and network rewiring has been inferred across the tree of life [Wotton et al., 2015, Crombach et al., 2016, Dalal and Johnson, 2017, Johnson, 2017, Ali et al., 2017], and there is often significant regulatory variation segregating within populations. Transcription in hybrids between closely related species with conserved transcriptional patterns can also be divergent [Haerty and Singh, 2006, Maheshwari and Barbash, 2012, Coolon et al., 2014, Michalak and Noor, 2004, Mack and Nachman, 2016], and hybrid incompatibilities have been attributed to cryptic molecular divergence underlying conserved body plans [Gavin-Smyth and Matute, 2013]. Furthermore, in cryptic species complexes (e.g., sun skinks [Barley et al., 2013]), genetically distinct species may be nearly morphologically indistinguishable.

**The origin of species not by means of natural selection?** As classically formulated, the Dobzhansky-Muller model of hybrid incompatibility is agnostic to the relative importance of neutral versus selective genetic substitutions [Coyne and Orr, 1998], and plausible mechanisms have been proposed whereby Dobzhansky-Muller incompatibilities could originate under neutral genetic drift [Lynch and Force, 2000] or stabilizing selection [Fierst and Hansen, 2009]. The same holds for the “pathway model” [Lindtke and Buerkle, 2015], which is closer to the situation here. However, previous authors have argued that neutral processes are likely too slow to be a significant driver of speciation [Nei et al., 1983, Seehausen et al., 2014]. This has led some to conclude that hybrid incompatibility is typically a byproduct of positive selection [Orr et al., 2004, Schluter, 2009] or a consequence of genetic conflict [Presgraves, 2010, Crespi and Nosil, 2013], two processes that typically act much more rapidly than genetic drift. However, our calculations suggest that even under strictly neutral processes, hybrid fitness breaks down as a function of genetic distance rapidly enough to play a substantial role in species formation across the tree of life. This is consistent with broad patterns such as the relationship between molecular divergence and genetic isolation seen by Roux et al. [2016], and the clocklike speciation rates observed by Hedges et al. [2015].

These explanations are not mutually exclusive. All of these forces – adaptive shifts, conflict, and network drift – are plausible drivers of speciation, and may even interact. Many of our observations carry over to models of directional selection – for instance, rapid drift along the set of equivalent systems could be driven by adaptation in a different, pleiotropically coupled system. Or, reinforcement due to local adaptation might provide a selective pressure that speeds up system drift. Furthermore, while the fitness consequences of incompatibility in any one given network may be small, the cumulative impact of system drift across the many different networks an organism relies on may be substantial. It remains to be seen how the relative strengths of these forces compare.

**The dimensionality of trait space** We have focused on examples of single traits (where the phenotype is one-dimensional), but phenotypes under selection are often high-dimensional, and variation in different traits often share a genetic basis. However, we still expect many degrees of freedom as long as there are components of the system not directly and individually constrained by selection (i.e., a kryptotype). Even in networks where the phenotype and kryptotype are of the same dimension, system theory shows us that there will always be available degrees of freedom as specific system realizations are only unique up to a change of coordinates. Some phenotypes, however, require kryptotypic dimensions to be larger than that of the phenotype. For instance, many systems have minimal realizations (*e.g.*, the oscillator in Example 2) where the dimension of the kryptotype is larger than that of the phenotype, implying that for these phenotypic dynamics to be realized, the kryptotype dimension *has* to be larger than the dimension of the phenotype. Even if components of the system’s internal state are directly subject to selection and the mode of action of the environment on the internal state is constrained (so, the input and output matrices  $B$  and  $C$  are fixed) then one could still perturb  $A$  as described above by  $ZA - AZ$  if  $ZB$  and  $CZ$  are both zero, implying a number of degrees of freedom that still grows with  $n^2$  for fixed  $\ell$  and  $m$ . Generically, the number of degrees of freedom is  $n(n - \ell - m)$ , so that in a system of  $n$  components, if even one component is not directly constrained, this leads to  $n$  degrees of freedom. Whatever the true “dimensionality” of phenotype space of a typical organism, there are undoubtedly aspects of its underlying molecular machinery that are not directly constrained, suggesting large numbers of degrees of freedom. Note that pleiotropy does not directly affect this argument at all – indeed, many phenotypically equivalent changes will lead to denser  $A$  matrices and hence more pleiotropy. However, more pleiotropic genes may be more strongly constrained, making it more difficult for systems to make the required compensatory changes for system drift.

Phenotypically equivalent system evolution is probably not only driven by neutral genetic drift. For one thing, movement along the set of equivalent networks is not expected to be completely neutral, since second-order selection pushes populations towards “flatter” regions of the fitness landscape in which a population centered on the optimal set has lower genetic load [as described in different contexts by [Rice, 1998](#), [Nimwegen et al., 1999](#)]. If this bias towards more robust networks is strong enough, it may even prevent drift, but it is unclear how strong this effect would be in practice. Our results, on the other hand, do not rely on the flatness of the fitness surface around the phenotypically equivalent set, but rather on the curvature of the equivalent set itself. So long as the phenotypically equivalent set is not closed under sexual recombination, opportunities for incompatibility remain. However the speed at which system drift can generate incompatibilities might diminish if selection for robustness is strong enough to constrain a population to a small section of system space, although the strength of such effects in practice are not known. Likewise, as the speed of system drift relies on segregating genetic variation, any constraints on such variation, possibly due to epistasis, genetic architecture [[Hermisson et al., 2003](#)], adaptive inertia [[Baatz and Wagner, 1997](#), [Álvarez-Castro et al., 2009](#)] or weak gene flow could plausibly slow it down. More work on specific systems, likely coupled with simulations, will be necessary to identify the biologically relevant parameter regimes.

**Nonlinearity and model assumptions** Of course, real regulatory networks are not linear dynamical systems. Most notably, physiological limits put upper bounds on expression levels, implying saturating response curves. It remains to be seen how well our results carry over into real systems, but the fact that most nonlinear systems can be locally approximated by a linear one suggests our qualitative results may hold more generally. Furthermore, nonidentifiability (which implies the existence of neutral directions) is often found in practice in moderately complex models of biological systems [e.g., [Gutenkunst et al., 2007](#), [Piazza et al., 2008](#), [Jiménez et al., 2017](#)].

Finally, despite our model’s precise separation of phenotype and kryptotype, this relationship in nature may be far more complicated as aspects of the kryptotype may be less “hidden” than we currently assume, and the neutral network changes we describe here may only be nearly neutral. For instance, attributes excluded from the phenotype as modeled here ignore the potential energy costs associated with excessively large (non-minimal) kryptotypes, as well as the relationship between a specific network architecture and robustness to mutational, transcriptional, or environmental noise. More precise modeling will require better mechanistic understanding not only of biological systems, but also the nature of selective pressures and



genetic variation in these systems.

## References

- Sammi Ali, Sarah Signor, Konstantin Kozlov, and Sergey Nuzhdin. Quantitative variation and evolution of spatially explicit morphogen expression in *Drosophila*. *bioRxiv*, page 175711, 2017. 13
- José M Álvarez-Castro, Michael Kopp, and Joachim Hermisson. Effects of epistasis and the evolution of genetic architecture: exact results for a 2-locus model. *Theoretical population biology*, 75(2-3):109–122, 2009. 14
- BDO Anderson, RW Newcomb, RE Kalman, and DC Youla. Equivalence of linear time-invariant dynamical systems. *Journal of the Franklin Institute*, 281(5):371–378, 1966. 2, 5
- M Baatz and G.P Wagner. Adaptive inertia caused by hidden pleiotropic effects. *Theoretical Population Biology*, 51(1):49–66, February 1997. doi: 10.1006/tpbi.1997.1294. 14
- Aderonke Babajide, Ivo L Hofacker, Manfred J Sippl, and Peter F Stadler. Neutral networks in protein space: a computational study based on knowledge-based potentials of mean force. *Folding and Design*, 2(5):261–269, 1997. 1
- Claudia Bank, Reinhard Bürger, and Joachim Hermisson. The limits to parapatric speciation: Dobzhansky–Muller incompatibilities in a continent-island model. *Genetics*, 191(3):845–863, 2012. ISSN 0016-6731. doi: 10.1534/genetics.111.137513. URL <https://www.genetics.org/content/191/3/845>.
- Anthony J Barley, Jordan White, Arvin C Diesmos, and Rafe M Brown. The challenge of species delimitation at the extremes: diversification without morphological change in philippine sun skinks. *Evolution*, 67(12):3556–3572, 2013. 13
- N H Barton. The maintenance of polygenic variation through a balance between mutation and stabilizing selection. *Genet Res*, 47(3):209–216, June 1986. doi: 10.1017/S0016672300023156. URL <https://www.ncbi.nlm.nih.gov/pubmed/3744046>. 2
- Nicholas H. Barton. The divergence of a polygenic system subject to stabilizing selection, mutation and drift. *Genetics Research*, 54(1):59–78, 1989. 2
- Nicholas H. Barton. The role of hybridization in evolution. *Molecular Ecology*, 10(3):551–568, 2001. 2
- W. Bateson. *Heredity and Variation in Modern Lights*, pages 85–101–. Cambridge University Press, Cambridge, 1909. ISBN 9781108004350. doi: DOI:10.1017/CBO9780511693953.007. URL <https://www.cambridge.org/core/books/darwin-and-modern-science/heredity-and-variation-in-modern-lights/6105CC0E0388ECDCEEA76EE779E278BE>. 2
- Richard Ernest Bellman and Karl Johan Åström. On structural identifiability. *Mathematical biosciences*, 7(3-4):329–339, 1970. 2
- Aviv Bergman and Mark L Siegal. Evolutionary capacitance as a general feature of complex gene networks. *Nature*, 424(6948):549–552, 2003. 1
- François Blanquart and Thomas Bataillon. Epistasis and the structure of fitness landscapes: Are experimental fitness landscapes compatible with Fisher’s geometric model? *Genetics*, 203(2):847–862, 2016. ISSN 0016-6731. doi: 10.1534/genetics.115.182691. URL <http://www.genetics.org/content/203/2/847>.
- Aleksandra A. Chertkova, Joshua S. Schiffman, Sergey V. Nuzhdin, Konstantin N. Kozlov, Maria G. Samsonova, and Vitaly V. Gursky. In silico evolution of the *Drosophila* gap gene regulatory sequence under elevated mutational pressure. *BMC Evolutionary Biology*, 17(1):4, 2017. ISSN 1471-2148. doi: 10.1186/s12862-016-0866-y. URL <http://dx.doi.org/10.1186/s12862-016-0866-y>. 1

- Luis-Miguel Chevin, Guillaume Decorzent, and Thomas Lenormand. Niche dimensionality and the genetics of ecological speciation. *Evolution*, 68(5):1244–1256, 2014. 2
- Joseph D Coolon, C Joel McManus, Kraig R Stevenson, Brenton R Graveley, and Patricia J Wittkopp. Tempo and mode of regulatory evolution in *Drosophila*. *Genome research*, 24(5):797–808, 2014. 13
- Jerry A Coyne and H Allen Orr. The evolutionary genetics of speciation. *Philosophical Transactions of the Royal Society of London B: Biological Sciences*, 353(1366):287–305, 1998. 13
- Gheorghe Craciun and Casian Pantea. Identifiability of chemical reaction networks. *Journal of Mathematical Chemistry*, 44(1):244–259, 2008. 2
- Bernard Crespi and Patrik Nosil. Conflictual speciation: species formation via genomic conflict. *Trends in Ecology & Evolution*, 28(1):48–57, 2013. 13
- Anton Crombach, Karl R Wotton, Eva Jiménez-Guri, and Johannes Jaeger. Gap gene regulatory dynamics evolve along a genotype network. *Molecular biology and evolution*, 33(5):1293–1307, 2016. 1, 13
- Chiraj K Dalal and Alexander D Johnson. How transcription circuits explore alternative architectures while maintaining overall circuit output. *Genes & Development*, 31(14):1397–1405, 2017. 1, 13
- Chiraj K Dalal, Ignacio A Zuleta, Kaitlin F Mitchell, David R Andes, Hana El-Samad, and Alexander D Johnson. Transcriptional rewiring over evolutionary timescales changes quantitative and qualitative properties of gene expression. *eLife*, 5:e18981, 2016. 1
- Eric H Davidson and Douglas H Erwin. Gene regulatory networks and the evolution of animal body plans. *Science*, 311(5762):796–800, 2006. 1
- J. Arjan G. M. de Visser, Joachim Hermisson, Günter P. Wagner, Lauren Ancel Meyers, Homayoun Bagheri-Chaichian, Jeffrey L. Blanchard, Lin Chao, James M. Cheverud, Santiago F. Elena, Walter Fontana, Greg Gibson, Thomas F. Hansen, David Krakauer, Richard C. Lewontin, Charles Ofria, Sean H. Rice, George von Dassow, Andreas Wagner, and Michael C. Whitlock. Perspective: Evolution and detection of genetic robustness. *Evolution*, 57(9):1959–1972, 2003. doi: <https://doi.org/10.1111/j.0014-3820.2003.tb00377.x>. URL <https://onlinelibrary.wiley.com/doi/abs/10.1111/j.0014-3820.2003.tb00377.x>. 2
- Jeremy Draghi and Michael Whitlock. Robustness to noise in gene expression evolves despite epistatic constraints in a model of gene networks. *Evolution*, 69(9):2345–2358, 2015. 1
- Gerald M Edelman and Joseph A Gally. Degeneracy and complexity in biological systems. *Proceedings of the National Academy of Sciences*, 98(24):13763–13768, 2001. 1, 2
- Janna L Fierst and Thomas F Hansen. Genetic architecture and postzygotic reproductive isolation: evolution of Bateson-Dobzhansky-Muller incompatibilities in a polygenic model. *Evolution*, 2009. 13
- C Fraïsse, P A Gunnarsson, D Roze, N Bierne, and J J Welch. The genetics of speciation: Insights from Fisher’s geometric model. *Evolution*, 70(7):1450–1464, 07 2016. doi: 10.1111/evo.12968. URL <https://www.ncbi.nlm.nih.gov/pubmed/27252049>. 2
- Jackie Gavin-Smyth and Daniel R Matute. Embryonic lethality leads to hybrid male inviability in hybrids between *Drosophila melanogaster* and *D. santomea*. *Ecology and Evolution*, 3(6):1580–1589, 2013. 13
- Sergey Gavrillets. Evolution and speciation on holey adaptive landscapes. *Trends in ecology & evolution*, 12(8):307–312, 1997. 2
- Sergey Gavrillets. Models of speciation: Where are we now? *Journal of heredity*, 105(S1):743–755, 2014. 2

- M Grewal and K Glover. Identifiability of linear and nonlinear dynamical systems. *IEEE Transactions on Automatic Control*, 21(6):833–837, Dec 1976. doi: 10.1109/TAC.1976.1101375. [2](#)
- Walter Grüner, Robert Giegerich, Dirk Strothmann, Christian Reidys, Jacqueline Weber, Ivo L Hofacker, Peter F Stadler, and Peter Schuster. Analysis of rna sequence structure maps by exhaustive enumeration i. neutral networks. *Monatshefte für Chemie/Chemical Monthly*, 127(4):355–374, 1996. [1](#)
- Ryan N Gutenkunst, Joshua J Waterfall, Fergal P Casey, Kevin S Brown, Christopher R Myers, and James P Sethna. Universally sloppy parameter sensitivities in systems biology models. *PLoS Computational Biology*, 3(10):e189, 2007. [14](#)
- Wilfried Haerty and Rama S Singh. Gene regulation divergence is a major contributor to the evolution of Dobzhansky–Muller incompatibilities between species of *Drosophila*. *Molecular Biology and Evolution*, 23(9):1707–1714, 2006. [13](#)
- JBS Haldane. Sex ratio and unisexual sterility in hybrid animals. *Journal of Genetics*, 12(2):101–109, 1922. [11](#)
- Emily E Hare, Brant K Peterson, Venky N Iyer, Rudolf Meier, and Michael B Eisen. Sepsid even-skipped enhancers are functionally conserved in *Drosophila* despite lack of sequence conservation. *PLoS Genetics*, 4(6):e1000106, 2008. [1](#)
- Kathryn M Hart, Michael J Harms, Bryan H Schmidt, Carolyn Elya, Joseph W Thornton, and Susan Marqusee. Thermodynamic system drift in protein evolution. *PLoS biology*, 12(11), 2014. [1](#)
- S Blair Hedges, Julie Marin, Michael Suleski, Madeline Paymer, and Sudhir Kumar. Tree of life reveals clock-like speciation and diversification. *Molecular Biology and Evolution*, 32(4):835–845, 2015. [13](#)
- Joachim Hermisson, Thomas F Hansen, and Günter P Wagner. Epistasis in polygenic traits and the evolution of genetic architecture under stabilizing selection. *The American Naturalist*, 161(5):708–734, 2003. [6](#), [14](#)
- Jennifer W Israel, Megan L Martik, Maria Byrne, Elizabeth C Raff, Rudolf A Raff, David R McClay, and Gregory A Wray. Comparative developmental transcriptomics reveals rewiring of a highly conserved gene regulatory network during a major life history switch in the sea urchin genus *Heliocidaris*. *PLoS Biology*, 14(3):e1002391, 2016. [1](#)
- Johannes Jaeger. The gap gene network. *Cellular and Molecular Life Sciences*, 68(2):243–274, 2011. [1](#)
- Johannes Jaeger, Svetlana Surkova, Maxim Blagov, Hilde Janssens, David Kosman, Konstantin N Kozlov, Ekaterina Myasnikova, Carlos E Vanario-Alonso, Maria Samsonova, David H Sharp, et al. Dynamic control of positional information in the early *Drosophila* embryo. *Nature*, 430(6997):368–371, 2004. [1](#)
- Alba Jiménez, James Cotterell, Andreea Munteanu, and James Sharpe. A spectrum of modularity in multi-functional gene circuits. *Molecular Systems Biology*, 13(4):925, April 2017. doi: 10.15252/msb.20167347. [1](#), [14](#)
- Alexander D Johnson. The rewiring of transcription circuits in evolution. *Current Opinion in Genetics & Development*, 47:121–127, 2017. [13](#)
- Rudolf E. Kalman. Mathematical description of linear dynamical systems. *J. SIAM Control*, 1963. [2](#), [5](#), [12](#)
- Rudolf E. Kalman, Peter L. Falb, and Michael A. Arbib. *Topics in mathematical system theory*. McGraw-Hill, New York, 1969. ISBN 0754321069. [5](#)
- M. Kasowski, F. Grubert, C. Heffelfinger, M. Hariharan, A. Asabere, S. M. Waszak, L. Habegger, J. Rozowsky, M. Shi, A. E. Urban, M. Y. Hong, K. J. Karczewski, W. Huber, S. M. Weissman, M. B. Gerstein, J. O. Korbel, and M. Snyder. Variation in transcription factor binding among humans. *Science*, 328(5975):232–235, April 2010. [12](#)

571 Konstantin Kozlov, Vitaly V Gursky, Ivan V Kulakovskiy, Arina Dymova, and Maria Samsonova. Analysis  
572 of functional importance of binding sites in the *Drosophila* gap gene network model. *BMC Genomics*,  
573 2015. 1

574 Russell Lande. Natural selection and random genetic drift in phenotypic evolution. *Evolution*, 30(2):pp.  
575 314–334, 1976. ISSN 00143820. URL <http://www.jstor.org/stable/2407703>. 11

576 Tuuli Lappalainen, Michael Sammeth, Marc R Friedländer, Peter AC 't Hoen, Jean Monlong, Manuel A  
577 Rivas, Mar Gonzalez-Porta, Natalja Kurbatova, Thasso Griebel, Pedro G Ferreira, et al. Transcriptome  
578 and genome sequencing uncovers functional variation in humans. *Nature*, 501(7468):506–511, 2013. 12

579 Dorothea Lindtke and C Alex Buerkle. The genetic architecture of hybrid incompatibilities and their effect  
580 on barriers to introgression in secondary contact. *Evolution*, 69(8):1987–2004, 2015. 13

581 Michael Lynch and Allan G Force. The origin of interspecific genomic incompatibility via gene duplication.  
582 *The American Naturalist*, 156(6):590–605, 2000. 13

583 Katya L Mack and Michael W Nachman. Gene regulation and speciation. *Trends in Genetics*, 2016. 13

584 Shamoni Maheshwari and Daniel A Barbash. Cis-by-trans regulatory divergence causes the asymmetric  
585 lethal effects of an ancestral hybrid incompatibility gene. *PLoS Genetics*, 8(3):e1002597, 2012. 13

586 Takeshi Matsui, Robert Linder, Joann Phan, Fabian Seidl, and Ian M Ehrenreich. Regulatory rewiring in a  
587 cross causes extensive genetic heterogeneity. *Genetics*, 201(2):769–777, 2015. 1

588 Ernst Mayr. The biological species concept. *Species concepts and phylogenetic theory: a debate*. Columbia  
589 University Press, New York, pages 17–29, 2000. 7

590 Pawel Michalak and Mohamed AF Noor. Association of misexpression with sterility in hybrids of *Drosophila*  
591 *simulans* and *D. mauritiana*. *Journal of Molecular Evolution*, 59(2):277–282, 2004. 13

592 Eric Mjolsness, David H Sharp, and John Reinitz. A connectionist model of development. *Journal of*  
593 *Theoretical Biology*, 152(4):429–453, 1991. 1

594 Masatoshi Nei, Takeo Maruyama, and Chung-I Wu. Models of evolution of reproductive isolation. *Genetics*,  
595 103(3):557–579, 1983. 13

596 Erik Van Nimwegen, James P Crutchfield, and Martijn Huynen. Neutral evolution of mutational robustness.  
597 *PNAS*, 1999. 14

598 H Allen Orr. Haldane’s rule. *Annual Review of Ecology and Systematics*, 28(1):195–218, 1997. 11

599 H Allen Orr, John P Masly, and Daven C Presgraves. Speciation genes. *Current Opinion in Genetics &*  
600 *Development*, 14(6):675–679, 2004. 13

601 Mihaela Pavlicev and Günter P Wagner. A model of developmental evolution: selection, pleiotropy and  
602 compensation. *Trends in Ecology & Evolution*, 27(6):316–322, 2012. 1

603 Y.P. Petrov and V.S. Sizikov. *Well-posed, ill-posed, and intermediate problems with applications*, volume 49.  
604 Walter de Gruyter, 2005. 2

605 Patrick C Phillips. Maintenance of polygenic variation via a migration–selection balance under uniform  
606 selection. *Evolution*, 50(3):1334–1339, 1996. 2

607 Matthew Piazza, Xiao-Jiang Feng, Joshua D Rabinowitz, and Herschel Rabitz. Diverse metabolic model  
608 parameters generate similar methionine cycle dynamics. *Journal of Theoretical Biology*, 251(4):628–639,  
609 2008. 14

- Verónica Miró Pina and Emmanuel Schertzer. How does geographical distance translate into genetic distance? *Stochastic Processes and their Applications*, 129(10):3893–3921, 2019. 2
- Björn Plötner, Markus Nurmi, Axel Fischer, Mutsumi Watanabe, Korbinian Schneeberger, Svante Holm, Neha Vaid, Mark Aurel Schöttler, Dirk Walther, Rainer Hoefgen, et al. Chlorosis caused by two recessively interacting genes reveals a role of RNA helicase in hybrid breakdown in *Arabidopsis thaliana*. *The Plant Journal*, 2017. 13
- Adam H Porter and Norman A Johnson. Speciation despite gene flow when developmental pathways evolve. *Evolution*, 56(11):2103–2111, 2002. 2
- Daven C Presgraves. The molecular evolutionary basis of species formation. *Nature Reviews Genetics*, 11(3):175–180, 2010. 13
- Sean H. Rice. The evolution of canalization and the breaking of von Baer’s laws: Modeling the evolution of development with epistasis. *Evolution*, 52(3):647–656, 1998. doi: <https://doi.org/10.1111/j.1558-5646.1998.tb03690.x>. URL <https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1558-5646.1998.tb03690.x>. 6, 14
- Loren H Rieseberg, Margaret A Archer, and Robert K Wayne. Transgressive segregation, adaptation and speciation. *Heredity*, 83(4):363–372, 1999. 13
- Ulises Rosas, Nick H. Barton, Lucy Copsey, Pierre Barbier de Reuille, and Enrico Coen. Cryptic variation between species and the basis of hybrid performance. *PLOS Biology*, 8(7):1–12, 07 2010. doi: 10.1371/journal.pbio.1000429. URL <https://doi.org/10.1371/journal.pbio.1000429>. 2
- Camille Roux, Christelle Fraise, Jonathan Romiguier, Yoann Anciaux, Nicolas Galtier, and Nicolas Bierne. Shedding light on the grey zone of speciation along a continuum of genomic divergence. *PLoS Biology*, 14(12):e2000234, 2016. 13
- Dolph Schluter. Evidence for ecological speciation and its alternative. *Science*, 323(5915):737–741, 2009. 13
- D Schmidt, M D Wilson, B Ballester, P C Schwalie, G D Brown, A Marshall, C Kutter, S Watt, C P Martinez-Jimenez, S Mackay, I Talianidis, P Flicek, and D T Odom. Five-vertebrate ChIP-seq reveals the evolutionary dynamics of transcription factor binding. *Science*, 328(5981):1036–1040, May 2010. doi: 10.1126/science.1186176. URL <https://www.ncbi.nlm.nih.gov/pubmed/20378774>. 12
- Ole Seehausen, Roger K Butlin, Irene Keller, Catherine E Wagner, Janette W Boughman, Paul A Hohenlohe, Catherine L Peichel, Glenn-Peter Saetre, Claudia Bank, Åke Brännström, et al. Genomics and the origin of species. *Nature Reviews Genetics*, 15(3):176–192, 2014. 13
- Maria R Servedio, Yaniv Brandvain, Sumit Dhole, Courtney L Fitzpatrick, Emma E Goldberg, Caitlin A Stern, Jeremy Van Cleve, and D Justin Yeh. Not just a theory – the utility of mathematical models in evolutionary biology. *PLoS Biology*, 12(12):e1002017, 2014. 1
- Mark L Siegal and Aviv Bergman. Waddington’s canalization revisited: developmental stability and evolution. *Proceedings of the National Academy of Sciences*, 99(16):10528–10532, 2002. 1
- Montgomery Slatkin and Russell Lande. Segregation variance after hybridization of isolated populations. *Genetics Research*, 64(1):51–56, 1994. 2
- TM Sonneborn. Degeneracy of the genetic code: extent, nature, and genetic implications. In *Evolving genes and proteins*, pages 377–397. Elsevier, 1965. 2
- K. Stefflova, D. Thybert, M. D. Wilson, I. Streeter, J. Aleksic, P. Karagianni, A. Brazma, D. J. Adams, I. Talianidis, J. C. Marioni, P. Flicek, and D. T. Odom. Cooperativity and rapid evolution of cobound transcription factors in closely related mammals. *Cell*, 154(3):530–540, August 2013. 12

652 Rike Stelkens and Ole Seehausen. Genetic distance between species predicts novel trait expression in their  
653 hybrids. *Evolution: International Journal of Organic Evolution*, 63(4):884–897, 2009. 13

654 J A Sved. A two-sex polygenic model for the evolution of premating isolation. ii. computer simulation of  
655 experimental selection procedures. *Genetics*, 97(1):217–235, January 1981. URL <https://www.ncbi.nlm.nih.gov/pubmed/17249074>. 2

657 Nicholas F Trojanowski, Olivia Padovan-Merhar, David M Raizen, and Christopher Fang-Yen. Neural and  
658 genetic degeneracy underlies *caenorhabditis elegans* feeding behavior. *Journal of neurophysiology*, 112(4):  
659 951–961, 2014. 1

660 John R True and Eric S Haag. Developmental system drift and flexibility in evolutionary trajectories.  
661 *Evolution & Development*, 3(2):109–119, 2001. 1, 2, 12

662 Annie E Tsong, Brian B Tuch, Hao Li, and Alexander D Johnson. Evolution of alternative transcriptional  
663 circuits with identical logic. *Nature*, 443(7110):415–420, 2006. 1

664 Alexander Y Tulchinsky, Norman A Johnson, Ward B Watt, and Adam H Porter. Hybrid incompatibility  
665 arises in a sequence-based bioenergetic model of transcription factor binding. *Genetics*, 198(3):1155–1166,  
666 2014. 2

667 Jenny Tung, Xiang Zhou, Susan C Alberts, Matthew Stephens, and Yoav Gilad. The genetic architecture of  
668 gene expression levels in wild baboons. *eLife*, 4:e04729, 2015. 12

669 Michael Turelli and H Allen Orr. The dominance theory of Haldane’s rule. *Genetics*, 140(1):389–402, 1995.  
670 11

671 AJ Van der Schaft. Equivalence of dynamical systems by bisimulation. *IEEE transactions on automatic  
672 control*, 49(12):2160–2172, 2004. 2

673 Dominique J Verlaan, Bing Ge, Elin Grundberg, Rose Hoberman, Kevin CL Lam, Vonda Koka, Joana Dias,  
674 Scott Gurd, Nicolas W Martin, Hans Mallmin, et al. Targeted screening of cis-regulatory variation in  
675 human haplotypes. *Genome research*, 19(1):118–127, 2009. 12

676 George von Dassow, Eli Meir, Edwin M. Munro, and Garrett M. Odell. The segment polarity network is a  
677 robust developmental module. *Nature*, 406(6792):188–192, 2000. ISSN 14764687. doi: 10.1038/35018085.  
678 URL <https://doi.org/10.1038/35018085>. 1

679 Andreas Wagner. Evolution of gene networks by gene duplications: a mathematical model and its implica-  
680 tions on genome organization. *Proceedings of the National Academy of Sciences*, 91(10):4387–4391, 1994.  
681 1, 3

682 Andreas Wagner. Does evolutionary plasticity evolve? *Evolution*, pages 1008–1023, 1996. 1

683 Andreas Wagner. Robustness and evolvability: a paradox resolved. *Proceedings of the Royal Society B:  
684 Biological Sciences*, 275(1630):91–100, 2008. 2

685 Eric Walter, Yves Lecourtier, and John Happel. On the structural output distinguishability of parametric  
686 models, and its relations with structural identifiability. *IEEE Transactions on Automatic Control*, 29(1):  
687 56–57, 1984. 2

688 Mi Wang, Severin Uebbing, and Hans Ellegren. Bayesian inference of allele-specific gene expression indicates  
689 abundant cis-regulatory variation in natural flycatcher populations. *Genome Biology and Evolution*, 9(5):  
690 1266–1279, 2017. 12



- Daniel M. Weinreich and Jennifer L. Knies. Fisher’s geometric model of adaptation meets the functional synthesis: Data on pairwise epistasis for fitness yields insights into the shape and size of phenotype space. *Evolution*, 67(10):2957–2972, 2013. doi: 10.1111/evo.12156. URL <https://onlinelibrary.wiley.com/doi/abs/10.1111/evo.12156>.
- Kenneth M Weiss and Stephanie M Fullerton. Phenogenetic drift and the evolution of genotype–phenotype relationships. *Theoretical Population Biology*, 57(3):187–195, 2000. 1, 12
- James M Whitacre. Degeneracy: a link between evolvability, robustness and complexity in biological systems. *Theoretical Biology and Medical Modelling*, 7(1):6, 2010. 2
- Karl R Wotton, Eva Jiménez-Guri, Anton Crombach, Hilde Janssens, Anna Alcaine-Colet, Steffen Lemke, Urs Schmidt-Ott, and Johannes Jaeger. Quantitative system drift compensates for altered maternal inputs to the gap gene network of the scuttle fly *Megaselia abdita*. *eLife*, 4:e04785, 2015. 1, 13
- Sewall Wright. Evolution in populations in approximate equilibrium. *Journal of Genetics*, 30(2):257, 1935. URL <http://link.springer.com/content/pdf/10.1007/BF02982240.pdf>. 2
- Ryo Yamaguchi and Yoh Iwasa. First passage time to allopatric speciation. *Interface focus*, 3(6):20130026, 2013. 2
- Lotfi A Zadeh and Charles A Deoser. *Linear system theory*. Robert E. Krieger Publishing Company Huntington, 1976. 2

## A Local expansion of the fitness surface

The fitness of a system depends on the difference between the system’s impulse response and the optimal impulse response, measured as a weighted sum of the distance between the impulse response from optimal. With  $\rho(t) \geq 0$  a weighting function on  $[0, \infty)$ , and  $h_0(t) = C_0 e^{tA_0} B_0$  a representative of the optimal set, the distance is equation (5):

$$D(h) = \left( \int_0^\infty \rho(t) \|h(t) - h_0(t)\|^2 dt \right)^{1/2}. \quad (6)$$

To see how this is affected by small changes to the system, first note that if  $\|\cdot\|_\rho$  is the  $L^2(\rho)$  norm,

$$\|g\|_\rho^2 := \int_0^\infty \rho(t) \|g(t)\|^2 dt \quad (7)$$

then the distance to optimum of a perturbed system,  $h(t) = h_0(t) + \epsilon g(t)$ , is

$$D(h_0 + \epsilon g) = \epsilon \|g\|_\rho. \quad (8)$$

Now consider the perturbed systems  $(A(\epsilon), B(\epsilon), C(\epsilon)) = (A(0), B(0), C(0)) + \epsilon(U, V, W)$ , and let  $h_\epsilon(t) = C(\epsilon)e^{A(\epsilon)t}B(\epsilon)$ . If two populations with systems  $(A(0), B(0), C(0))$  and  $(A(\epsilon), B(\epsilon), C(\epsilon))$  interbreed, then the mean resulting system is  $(A(\epsilon/2), B(\epsilon/2), C(\epsilon/2))$ , but with greater variance in the  $F_2$  than the  $F_1$  offspring (see Figure 4). Indeed, if the difference between (say)  $A(0)_{ij}$  and  $A(\epsilon)_{ij}$  is due to  $n$  fixed differences with absolute additive effect size  $\epsilon/\sqrt{n}$  each, then the standard deviation of  $A_{ij}$  among the  $F_1$  offspring is still equal to that within each parent population, as each locus is heterozygous. However, if loci are unlinked then an  $F_2$  offspring is homozygous with probability  $1/2$ , and takes either homozygote with equal probability, so the variance of the contribution of each locus is  $\epsilon/4\sqrt{n}$ , and so the standard deviation across  $F_2$  offspring is  $\epsilon/4$  plus a contribution for within-population genetic variance. (This many differences of this size would be expected if the difference was due to drift, for instance.) This implies that the impulse response,  $h(t)$ , of

an  $F_2$  will also differ from optimum by something of order  $\epsilon$ . However, since the displacement between the populations lies along a ridge, the  $F_1$  offspring are closer: the Taylor expansions for  $h$  about both 0 and  $\epsilon$  are

$$h_{\epsilon/2}(t) = h_0(t) + \frac{\epsilon}{2} \partial_\epsilon h_0(t) + \frac{\epsilon^2}{8} \partial_\epsilon^2 h_0(t) + O(\epsilon^3) \quad (9)$$

$$\text{and } h_\epsilon(t) = h_0(t) - \frac{\epsilon}{2} \partial_\epsilon h_0(t) + \frac{\epsilon^2}{8} \partial_\epsilon^2 h_0(t) + O(\epsilon^3) \quad (10)$$

Since  $h_\epsilon(t) = h_0(t)$  by assumption, and  $\partial_\epsilon h_\epsilon(t) = \partial_\epsilon h_0(t) + \epsilon \partial_\epsilon^2 h_0(t) + O(\epsilon^2)$ , combining these expressions we get that

$$h_{\epsilon/2}(t) = h_0(t) - \frac{\epsilon^2}{8} \partial_\epsilon^2 h_0(t) + O(\epsilon^3). \quad (11)$$

In other words, if the parental populations are both optimal but differ by  $\epsilon$ , then the  $F_1$  offspring differ from optimal by something of order  $\epsilon^2$ , but the  $F_2$  by something of order  $\epsilon$ .

Of course, when moving away from the ridge, the fitness landscape is flatter in some directions than others, and how much so depends on the details. For completeness, we describe here how to compute the sensitivity of distance to optimum with respect to each of  $A$ ,  $B$ , and  $C$ . Changing notation slightly, let's rewrite the squared distance to optimum of a system  $(A, B, C)$ :

$$\begin{aligned} D(A, B, C)^2 &:= \int_0^\infty \rho(t) |h_A(t) - h_0(t)|^2 dt \\ &= \int_0^\infty \rho(t) |C e^{At} B - C_0 e^{A_0 t} B_0|^2 dt \\ &= \int_0^\infty \rho(t) \text{tr} \left\{ (C e^{At} B - C_0 e^{A_0 t} B_0)^T (C e^{At} B - C_0 e^{A_0 t} B_0) \right\} dt \\ &= \int_0^\infty \rho(t) \text{tr} \left\{ (C e^{At} B - C_0 e^{A_0 t} B_0) (C e^{At} B - C_0 e^{A_0 t} B_0)^T \right\} dt, \end{aligned} \quad (12)$$

where  $\text{tr } X$  denotes the trace of a square matrix  $X$ . How does this change as we perturb about  $(A_0, B_0, C_0)$ ? First we differentiate with respect to  $A$ , keeping  $B = B_0$  and  $C = C_0$  fixed. Since

$$\frac{d}{du} e^{(A+uZ)t} \Big|_{u=0} = \int_0^t e^{As} Z e^{A(t-s)} ds, \quad (13)$$

we have that

$$\begin{aligned} \frac{d}{du} D(A + uZ, B_0, C_0)^2 \Big|_{u=0} &= 2 \int_0^\infty \rho(t) \text{tr} \left\{ C_0 \left( \int_0^t e^{As} Z e^{A(t-s)} ds \right) B_0 B_0^T (e^{At} - e^{A_0 t})^T C_0^T \right\} dt \\ &= 2 \int_0^\infty \rho(t) \text{tr} \left\{ C_0 \left( \int_0^t e^{As} Z e^{A(t-s)} ds \right) B_0 (h_A(t) - h_0(t))^T \right\} dt \end{aligned} \quad (14)$$

and, by differentiating this and supposing that  $A$  is on the optimal set, i.e.,  $h_A(t) = h_0(t)$ , (so without loss of generality,  $A = A_0$ ):

$$\begin{aligned} \mathcal{H}^{A,A}(Y, Z) &:= \frac{1}{2} \frac{d}{du} \frac{d}{dv} D(A_0 + uY + vZ, B_0, C_0)^2 \Big|_{u=v=0} \\ &= \int_0^\infty \rho(t) \text{tr} \left\{ C_0 \left( \int_0^t e^{A_0 s} Y e^{A_0(t-s)} ds \right) B_0 B_0^T \left( \int_0^t e^{A_0 s} Z e^{A_0(t-s)} ds \right)^T C_0^T \right\} dt. \end{aligned} \quad (15)$$

The function  $\mathcal{H}$  will define a quadratic form. To illustrate the use of this, suppose that  $B$  and  $C$  are fixed. By defining  $\Delta_{ij}$  to be the matrix with a 1 in the  $(i, j)$ th slot and 0 elsewhere, the coefficients of the quadratic form are

$$H_{ij, k\ell}(A) := \mathcal{H}(\Delta_{ij}, \Delta_{k\ell}). \quad (16)$$

744 We could use this to get the quadratic approximation to  $D$  near the optimal set. To do so, it'd be nice  
 745 to have a way to compute the inner integral above. Suppose that we diagonalize  $A = U\Lambda U^{-1}$ . Then

$$\int_0^t e^{As} Z e^{A(t-s)} ds = \int_0^t U e^{\Lambda s} U^{-1} Z U e^{\Lambda(t-s)} U^{-1} ds \quad (17)$$

746 Now, notice that

$$\int_0^t e^{s\lambda_i} e^{(t-s)\lambda_j} ds = \begin{cases} \frac{e^{t\lambda_i} - e^{t\lambda_j}}{\lambda_i - \lambda_j} & \text{if } i \neq j \\ te^{t\lambda_i} & \text{if } i = j. \end{cases} \quad (18)$$

747 Therefore, defining

$$X_{ij}(t, Z) = \begin{cases} (U^{-1} Z U)_{ij} \frac{e^{t\lambda_i} - e^{t\lambda_j}}{\lambda_i - \lambda_j} & \text{if } i \neq j \\ (U^{-1} Z U)_{ii} te^{t\lambda_i} & \text{if } i = j, \end{cases} \quad (19)$$

748 moving the  $U$  and  $U^{-1}$  outside the integral and integrating we get that

$$\int_0^t e^{As} Z e^{A(t-s)} ds = U X(t, Z) U^{-1}. \quad (20)$$

749 This implies that

$$D(A_0 + \epsilon Z)^2 \approx \frac{1}{2} \epsilon^2 \int_0^\infty \rho(t) \operatorname{tr} \{ C U X(t, Z) U^{-1} B B^T (U^{-1})^T X(t, Z)^T U^T C^T \} dt. \quad (21)$$

750 To compute the  $n^2 \times n^2$  matrix  $H$ , we see that if  $Z = \Delta_{k\ell}$ , then

$$X_{ij}^{k\ell}(t) = \begin{cases} (U^{-1})_{\cdot k} U_{\ell} \frac{e^{t\lambda_i} - e^{t\lambda_j}}{\lambda_i - \lambda_j} & \text{if } i \neq j \\ (U^{-1})_{\cdot k} U_{\ell} te^{t\lambda_i} & \text{if } i = j, \end{cases} \quad (22)$$

751 where  $U_{k\cdot}$  is the  $k$ th row of  $U$ , and so

$$H_{ij, k\ell}(A) = \int_0^\infty \rho(t) \operatorname{tr} \{ C U X^{ij}(t) U^{-1} B B^T (U^{-1})^T X^{k\ell}(t)^T U^T C^T \} dt. \quad (23)$$

752 This implies that

$$D(A_0 + \epsilon Z)^2 \approx \frac{1}{2} \epsilon^2 \sum_{ijkl} H_{ij, k\ell}(A_0) Z_{ij} Z_{k\ell}. \quad (24)$$

753 More generally,  $B$  and  $C$  may also change. To extend this we need the remaining second derivatives of  
 754  $D^2$ . First, in  $B$ :

$$\begin{aligned} \mathcal{H}^{B,B}(Y, Z) &:= \frac{1}{2} \frac{d}{du} \frac{d}{dv} D(A_0, B_0 + uY + vZ, C_0)|_{u=v=0} \\ &= \frac{1}{2} \int_0^\infty \rho(t) \operatorname{tr} \left\{ C_0 e^{tA_0} \frac{d}{du} \frac{d}{dv} (uY + vZ)(uY + vZ)^T |_{u=v=0} e^{tA_0^T} C_0^T \right\} dt \\ &= \frac{1}{2} \int_0^\infty \rho(t) \operatorname{tr} \left\{ C_0 e^{tA_0} (Y Z^T + Z Y^T) e^{tA_0^T} C_0^T \right\} dt. \end{aligned} \quad (25)$$

755 Next, in  $C$ :

$$\begin{aligned} \mathcal{H}^{B,B}(Y, Z) &:= \frac{1}{2} \frac{d}{du} \frac{d}{dv} D(A_0, B_0, C_0 + uY + vZ)|_{u=v=0} \\ &= \frac{1}{2} \int_0^\infty \rho(t) \operatorname{tr} \left\{ B_0 e^{tA_0^T} \frac{d}{du} \frac{d}{dv} (uY + vZ)^T (uY + vZ) |_{u=v=0} e^{tA_0} B_0 \right\} dt \\ &= \frac{1}{2} \int_0^\infty \rho(t) \operatorname{tr} \left\{ B_0 e^{tA_0^T} (Y Z^T + Z Y^T) e^{tA_0} B_0 \right\} dt. \end{aligned} \quad (26)$$

756 Now, the mixed derivatives in  $B$  and  $C$ :

$$\begin{aligned}\mathcal{H}^{B,C}(Y, Z) &:= \frac{1}{2} \frac{d}{du} \frac{d}{dv} D(A_0, B_0 + uY, C_0 + vZ)|_{u=v=0} \\ &= \int_0^\infty \rho(t) \operatorname{tr} \left\{ Y e^{tA_0^T} C_0^T Z e^{tA_0} B_0 \right\} dt.\end{aligned}\tag{27}$$

757 In  $A$  and  $B$

$$\begin{aligned}\mathcal{H}^{A,B}(Y, Z) &:= \frac{1}{2} \frac{d}{du} \frac{d}{dv} D(A_0 + uY, B_0 + vZ, C_0)|_{u=v=0} \\ &= \int_0^\infty \rho(t) \operatorname{tr} \left\{ C_0 \left( \int_0^t e^{sA_0} Y e^{(t-s)A_0} ds \right) B_0 Z^T e^{tA_0} C_0 \right\} dt,\end{aligned}\tag{28}$$

758 and finally in  $A$  and  $C$ :

$$\begin{aligned}\mathcal{H}^{A,C}(Y, Z) &:= \frac{1}{2} \frac{d}{du} \frac{d}{dv} D(A_0 + uY, B_0, C_0 + vZ)|_{u=v=0} \\ &= \int_0^\infty \rho(t) \operatorname{tr} \left\{ C_0 \left( \int_0^t e^{sA_0} Y e^{(t-s)A_0} ds \right) B_0 B_0 e^{tA_0} Z \right\} dt.\end{aligned}\tag{29}$$

759 Together, numerical computation of these expressions, along with estimates of genetic covariance within  
760 a population, allow precise predictions of evolutionary dynamics of a particular system. The approximation  
761 should be good as long as the second-order Taylor approximation holds.

Resubmission Cover Letter  
*Evolution*

Joshua Schiffman  
*and* Peter Ralph  
July 5, 2021

**To the Editor(s) –**

Cover letter.  
**Sincerely,**

**Joshua Schiffman and Peter Ralph**

## Reviewer AE:

This is a very interesting theory paper that connects the modeling and evolution of regulatory networks with speciation. While quite theoretical in nature, the authors made an effort to provide examples, and I consider this paper of very high relevance for the field.

The authors have addressed most of the previous comments from the first round of reviews, and the two new reviewers only raise minor concerns and suggestions, especially regarding the paragraph on Haldane’s rule and regarding a better placement of the paper in the context of speciation research. I agree with these concerns; regarding the latter point I have three additional questions/comments, one of which was also raised in the previous round of reviews:

Thank you for the kind words.

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**(AE.1) Gene flow** *As it is described, the model only holds for allopatric populations; I would expect even a small amount of gene flow to make neutral evolution of hybrid incompatibility impossible (Gavrilets 1997, Bank et al. 2012). This is not discussed in the current version of the paper; even more interestingly, would the authors expect conditions in which emergence of incompatibility could be possible in this model also in the presence of gene flow?*

Reply:

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**(AE.2) Orr’s Snowball** *The authors state that rapid emergence of incompatibility is possible in their model. How would the accumulation of incompatibility here compare with Orr’s “snowball” model, and how is this expected to depend on the complexity of the underlying networks?*

Reply:

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**(AE.3) Genetic architecture** *The genetic architecture (e.g., interactions between cis-regulatory and coding regions) is important to determine the barrier to gene flow that an incompatibility builds. If there is a lot of short-distance interactions (physically in the genome; i.e. low recombination), they might not play a role for speciation per se because the incompatibility will often not be exposed to selection even in hybrids. In the light of our current knowledge of the genetic architecture of regulatory networks, how would this affect the conclusions from the model?*

Reply:

## Reviewer 1:

I am basically enthusiastic about this paper, which has been influential for some time on bioRxiv. The central sections, especially e.g. lines 127-219, are very clear and novel. I do think, though that the work could be better placed in context.

Thank you. We were also excited that the paper had many readers on the bioRxiv.

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**(1.1) Empirical evidence for the (minor?) role of drift in speciation:** *Lines 382-389 discuss previous discussion about the role of drift in speciation, but they focus on theory. I think that some strong arguments for the primacy of selection in speciation have been empirical (bottlenecking Drosophila populations and testing for RI, MK-tests on “speciation genes” etc.). Some of the earlier work was summarized in the last chapter of Coyne and Orr 2004. Line 396 seems to ignore a lot of this work, and so I would rephrase.*

Reply:



**(1.2) Additive quantitative trait models** The previous version of this manuscript contained a standard quantitative trait model. I think it was a good idea to remove this material, but I still think that this modelling approach (including Fisher’s geometric model) needs to be discussed a little more here. In the introduction, for example, it seems important to state clearly that “kryptotypes” (many-to-one genotype-to-phenotype maps) are also a feature of these standard models, and that previous authors have shown how phenotypically identical populations can evolve RI, in some cases via drift alone (e.g., Mani and Clarke 1990; Rosas et al. 2010; Fraitse et al. 2016 and others).

Martin 2014 argued that the results of these models and “systems-biology” type models are likely to approximate one another under some conditions, and that could be mentioned.

Reply:

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**(1.3) RI due to kryptotypes not involving drift** The section beginning on line 419 seems quite important to me, but it seems to downplay an important point. Populations might diverge genetically, or in their kryptotypes, but not in their phenotypes, if they experience fluctuating selection. This goes back the Wright-Fisher debates, and has been discussed in the context of speciation by, e.g., Mani and Clarke 1990, Barton 2001 and later authors. I think that the empirical evidence for fluctuating selection is also quite strong (e.g Bell 2010), and so it seems more important to discuss that, e.g., selection for robustness.

Reply:

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**(1.4) Interpretation of the Haldane’s Rule result** The abstract refers to “another possible explanation” of Haldane’s Rule, and the explanation given is said to be quite different from the “dominance theory”, because “it derives from the nature of segregation variance”. I wonder whether this is not misleading. My view is that the sort of model explored here (like additive quantitative trait model used by Barton 2001) naturally generates the sort of dominance relations required for the “dominance theory” to hold. Unless the authors strongly disagree, I would modify the abstract and lines 278-291, or at least provide a fuller clarification for why the explanation is different from that discussed by Barton 2001.

Reply:

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**(1.5)** 5. (very minor) Line 358: As you say, F2 hybrid breakdown has been known about for a very long time. As such, the single citation used feels a bit arbitrary.

Reply:

## Reviewer 2:

The manuscript explores how regulatory changes yielding the same phenotype may lead to hybrid incompatibilities and speciation. The authors model gene regulatory networks as sets of linear equations, where environmental inputs are integrated, giving rise to specific phenotypic outputs. Using Kalman decomposition, the authors identify the subset of such systems that produce the same phenotype and thus, characterise how a given system may be modified without affecting its phenotypic response. Resorting to a simple oscillatory network as an example, the authors show that phenotypically identical parents can generate offspring with considerably different phenotypes, resulting in hybrid incompatibilities if the set of equivalent phenotypes is not closed under the average. The phenotypic distance between parents and offspring is proportional to the square of the genetic distance between parents for F1 hybrids and directly proportional to the genetic distance between parents for F2 hybrids.

This is an interesting paper that nicely bridges core concepts of systems biology and evolution. The examples provided are very “visual”, making quite clear the idea of “equivalent systems” and will greatly assist readers not familiar with Kalman decomposition. The figures are both

informative and elegant, and the discussion aptly covers model validity and makes an effort to relate this work to standard speciation models. In addition, the authors have adequately addressed the comments of previous reviewers. Overall, this manuscript constitutes an appropriate addition to Evolution.

We recommend this paper for publication, but we have some suggestions that we consider would improve the clarity of this paper for a broader biological audience.

Thank you.

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**(2.1) Haldane's rule** *Our primary concern with the text regards Haldane's rule. The authors state that a male F1 offspring has a phenotype that could usually only be obtained in an F2 individual if all the involved genes were autosomal. Although this is true, it does not look like an arbitrary F2 individual but one with a very particular structure. This type of individual is like F1 in all autosomal genes but equal to the parental genotype in the genes present in the sex chromosome. It is not clear to us that this should necessarily generate a worse outcome than a female individual.*

*The F2 individuals only perform worse than F1 on average, and looking at figure 4 (left), we can see that some F2 individuals actually perform better than the F1 individuals.*

**Reply:** The fitness of heterogametic F1s and F2s will always depend on the genetic distance between parents and on the particular gene network. Thus it is not guaranteed that the heterogametic F1s, will always have lower fitnesses than homogametic F1s. This is clear when the genetic distance is zero between parents. The model does show, however, that the particular combination of genes found in F1 heterogametes would only be possible in F2 autosomal crosses. Our model shows that phenotypes generated by networks contained on the autosomes have lower fitnesses in F2s than in F1s, and by extension, lower fitness in the heterogametic sex when networks are contained, at least in part, on the sex chromosomes.

It is also true that some F2 individuals will perform as well as F1s and F0s – for instance it is possible to recover an F0-like genotype (and therefore phenotype) after two generations of hybridization. In this model, hybrids have lower fitness because of increased genetic variance. However, there will also be F2 individuals with genes from both parental types – meaning that there will be more low fitness F2s than low fitness F1s and F0s. *(The F2 fitness is lower on average?)*

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**(2.2)** (p. 2, l. 73): *This paragraph summarises the main body of work performed in this paper. Thus, consider moving it to the end of the Introduction (i.e. switching places with the last paragraph).* **Reply:**

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**(2.3)** (p. 2, l. 92): *“...more than one way to do the same thing...” sounds a bit too colloquial. Consider changing it to “...more than one way to accomplish the same outcome...” or some other alternative phrasing.*

**Reply:**

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**(2.4)** (p. 5, l. 192): *“The Kalman decomposition then classifies each direction in kryptotype space as either reachable or unreachable”. Is there a biological motivation for considering unreachable directions in kryptotype space as degrees of freedom of the neutral space?*

**Reply:**

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**(2.5)** (p. 6, l. 208): *“both the change of basis used to obtain the decomposition and, once in this form, all submatrices other than  $A_{ro}$ ,  $B_{ro}$ , and  $C_{ro}$  can be changed without affecting the phenotype, and so represent degrees of freedom”. A minor question on generality - does Kalman decomposition require that the coefficient matrices are time-invariant?*

**Reply:** As far as we understand, and as applied here, the Kalman decomposition is for linear time-invariant systems. Generalizations of the Kalman decomposition for time-varying systems have been discussed in... *Could cite some papers? Looks like Silverman and Meadows, EQUIVALENT REALIZATIONS OF LINEAR SYSTEMS discuss generalizations to the time-variable case.*

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**(2.6) Recombination** (p. 7) In the section “Sexual reproduction and recombination”, we think that the recombination process is not clearly explained, making it more challenging to read the text from this point. We suggest that the authors clarify how exactly recombination is implemented in the model. Perhaps an example would help the reader to have a better picture of the process. This example could be inline, like the one introduced in the section on Haldane’s rule.

**Reply:**

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**(2.7)** (p. 8, Figure 4 and p. 10, Figure 5): We suggest dividing the figure into sub-figures (e.g. A and B). Furthermore, we suggest adding “F1” and “F2” labels to the left and right plots in Figure 5, respectively, as well as the values of epsilon. This way, the reader would be immediately informed of what is changing.

**Reply:**