Phenological models on microbial

growth – which model is better and

\mathbf{why} ?

- PokMan HO
- Department of Life Sciences, Faculty of Natural Sciences,
- Imperial College London



Approximate Word Count: 921

⁸ Phenological models on microbial growth – which model is better

and why?

PokMan HO (CID: 01786076)

$_{\scriptscriptstyle 11}$ Abstract

9

10

12 Introduction

- Phenological models are expected to fit data trends within its biological field. Yet due to different
- 14 reasons, models developed and published from one sample may not fit the others. These reasons
- 15 may be due to data variabilities, confounding factors, inaccurate assumptions or models being
- too-specific. This project is aimed at compare and contrast published phenological models on
- 17 microbial population size data, highlighting which is a better model under what conditions.
- 18 The hypotheses are:
- published phenological models are better than polynomials in describing microbial population size;
- appropriate phenological model(s) can be identified through distinguishable shapes of
 microbial population size; and
- parameters of data under each phenological model is clustered, similar with dataset bestdescribed by the same model but different from those described by other models.

$_{25}$ Methods

- 26 Experimental microbial population growth data library were divided into individual data sub-
- sets through six filters ("Temperature (in °C)", "Microbial clade", "growth substrate materi-
- 28 als", "experimental replicate number", "population data recording unit" and "data source").
- 29 Records with data unit "OD_595" were scaled into optical density percentages (i.e. data*100)

to facilitate general analyses workflow. Independent (or explanatory) variable was "Time (hr)" and dependent (or response) variable was "population size".

Some raw data were recorded in minutes (instead of hour). This record artifact was not corrected because of two reasons: 1. shape of curves were the main concern instead of independent variable's scale; and 2. the unit was consistent within each data subset.

Six candidate models were assessed, four phenological and two polynomial equations. They were

5 Model assessment

55

56

"Verhulst (classical)"¹, "modified Gompertz"², "Baranyi"³, "Buchanan"⁴, "quadratic" and "cubic". NLLS was used only on the four phenological models and linear model-fitting was done 38 on the two polynomials. Starting values selection (for phenological models only) was described 39 below: Initial (N0) and final (K) population sizes were selected to be the minimum and maximum values of each data subset respectively. Maximum growth rate (r.max) was selected by linear 42 model through a recursive manner. For every iteration, population size data from the top 5% independent variable values were excluded from the linear model calculation. The data and 44 slope would only be recorded if it was positive, higher adjusted R² value and larger slope than 45 the recorded "best slope" value. After scanning from the maximum side, the best slope and its respective data were taken out and screened from the minimum side. Final best slope and x-intercept were regarded as the r.max and relative time lag (t.lag) of the population (in the source experiment) respectively. Time which this linear model intersected with K was regarded as the time achieving carrying capacity (t.K). Population data was then classified into three 50 groups (gx) according to the time: $g1 \le t.lag < g2 < t.K \le g3$. 5% was chosen as the scanning 51 threshold because I assumed this resolution was fine enough for achieving good starting values for NLLS fitting. Inputs for phenological modelswere listed below (popn & time were the dependent and independent variables respectively):

```
Verhulst (classical): popn = f(N0, K, r.max, time)

modified Gompertz: popn = f(N0, K, r.max, time, t.lag)

Baranyi: popn = f(N0, K, r.max, time, t.lag)

Buchanan: popn = f(N0, K, r.max, time, t.lag, gx)
```

All test starting values were than sampled from normal distribution with mean as the estimated value and standard deviation (sd) of 1. The sd value was chosen because of different reasons for each parameters. No and K were directly extracted from the raw experimental data, which could be assumed being an accurate estimate for that data subset (hence a small sd was logical). r.max was a guesstimated value from fitting linear models. This process could potentially be affected by extreme values in the data and hence a large sd should be preferred.

100 trials were done as a optimal value under a trade-off between efficiency and accuracy.

64

Only AIC⁵⁻⁷ was used to select for optimal parameter values within each phenological model and best model between the six candidates for a data subset. Reasons would be listed in Discussion section. For models with more than one parameter sets as sharing the lowest AIC value, the first set of values from the random sampling trials were used for downstream analyses. AIC tolerance threshold was expanded to min(AIC)+2⁸ to incorporate more accepted models for analyses.

71 Statistical analysis

Kruskal test was used for identify the best-fit model among all included model because the count was categorical and not assumed being normally-distributed. Pairwise Nemenyi comparisons would be carried out to identify the best test if p-value of the above test was significant.

75 Main Assumptions

- there was no negative population growth (i.e. starting population was always lower than carrying capacity), so negative population growth data were set to zeros;
- estimated parameter estimates would always result in a global optimal status in parameter space through the non-linear least squares method (NLLS)

80 Computing tools

R (ver 3.6.0)⁹ was used with "minpack.lm"¹⁰ for computing non-linear least square statistics for model comparisons. "PMCMR"¹¹ was used for carrying out statistical analyses.

3 Results

- From Fig.1, large fluctuations between each model to be described as "best-fit" were observed.
- However the occurrence difference was not statistical significant. Among the counts, there were

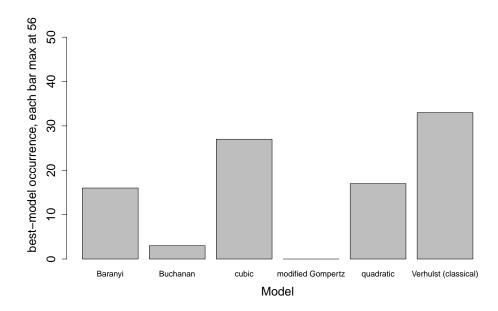


Figure 1: Barplot showing the number of "best model" identification under AIC model-selection methods with "Kruskal-Wallis rank sum test " statistic $X^2=5$, df = 5, p = 0.42

40 datasets with more than one "best-fit" models. Verhulst (classical) and cubic were the top
two models selected as "best-fit" for the 56 datasets (33 for Verhulst (classical) and 27 for
cubic). There are 11 datasets calling both "best-fit" at the same trial. Between Baranyi and
quadratic, the counts were 16 and 17 respectively with 6 datasets calling both models "best-fit".

The only outstanding performance was from modified Gompertz, which 0 datasets were called
it as "best-fit".

Discussion

AIC is considered the most suitable model-selection approach within model and between models in this project. Unlike BIC, AIC are more accurate with small sample sizes^{12,13} and sparse data¹³. AIC did not assume a "true model" was under examination^{14–16}. Since candidate models were not "nested model", BIC is not a better choice than AIC¹⁷. Hence the use of only AIC as model-selection criterion should be justified.

99 k

98

100 Conclusion

Published phenological models were data-specific, which none of them were found significantly performing better than the others in general.

103 Code and Data Availability

All scripts and data used for this report were publicity available at GitHub.

105 References

- 1. McKendrick, A. & Pai, M. K. XLV.—the rate of multiplication of micro-organisms: a mathematical study. *Proceedings of the Royal Society of Edinburgh* **31**, 649–653 (1912).
- 2. Gil, M. M., Brandão, T. R. & Silva, C. L. A modified Gompertz model to predict microbial inactivation under time-varying temperature conditions. *Journal of Food Engineering* **76.**Bugdeath, 89 –94. ISSN: 0260-8774. http://www.sciencedirect.com/science/article/pii/S0260877405003389 (2006).
- 3. Baranyi, J, McClure, P., Sutherland, J. & Roberts, T. Modeling bacterial growth responses.

 Journal of industrial microbiology 12, 190–194 (1993).
- 4. Buchanan, R., Golden, M. & Whiting, R. Differentiation of the effects of pH and lactic or
 acetic acid concentration on the kinetics of Listeria monocytogenes inactivation. *Journal* of Food Protection 56, 474–478 (1993).
- 5. Johnson, J. B. & Omland, K. S. Model selection in ecology and evolution. *Trends in ecology* evolution **19**, 101–108 (2004).
- 6. Akaike, H. in Selected papers of hirotugu akaike 199–213 (Springer, 1998).
- 7. Burnham, K. & Anderson, D. Model selection and multimodel inference: a practical information-theoretic approach. *Ecological Modelling*.
- 8. Burnham, K. P. & Anderson, D. R. Multimodel inference: understanding AIC and BIC in model selection. Sociological methods & research 33, 261–304 (2004).
- 9. R Core Team. R: A Language and Environment for Statistical Computing R Foundation for Statistical Computing (Vienna, Austria, 2019). https://www.R-project.org/.

- 10. Elzhov, T. V., Mullen, K. M., Spiess, A.-N. & Bolker, B. minpack.lm: R Interface to the Levenberg-Marquardt Nonlinear Least-Squares Algorithm Found in MINPACK, Plus
- Support for Bounds R package version 1.2-1 (2016). https://CRAN.R-project.org/
- package=minpack.lm.
- 130 11. Pohlert, T. The Pairwise Multiple Comparison of Mean Ranks Package (PMCMR) R
 package (2014). https://CRAN.R-project.org/package=PMCMR.
- 132 12. Acquah, H. D.-G. Comparison of Akaike information criterion (AIC) and Bayesian in-133 formation criterion (BIC) in selection of an asymmetric price relationship. *Journal of* 134 *Development and Agricultural Economics* 2, 001–006 (2010).
- 13. Kuha, J. AIC and BIC: Comparisons of assumptions and performance. Sociological methods

 8. Example 13. Wuha, J. AIC and BIC: Comparisons of assumptions and performance. Sociological methods

 13. Example 13. Wuha, J. AIC and BIC: Comparisons of assumptions and performance. Sociological methods

 13. Example 13. Exa
- 14. Aho, K., Derryberry, D. & Peterson, T. Model selection for ecologists: the worldviews of
 AIC and BIC. Ecology 95, 631–636 (2014).
- 139 15. Vrieze, S. I. Model selection and psychological theory: a discussion of the differences 140 between the Akaike information criterion (AIC) and the Bayesian information criterion 141 (BIC). Psychological methods 17, 228 (2012).
- 142 16. Yang, Y. Can the strengths of AIC and BIC be shared? A conflict between model inden-143 tification and regression estimation. *Biometrika* **92**, 937–950 (2005).
- 144 17. Wang, Y. & Liu, Q. Comparison of Akaike information criterion (AIC) and Bayesian information criterion (BIC) in selection of stock—recruitment relationships. *Fisheries Research*146 77, 220–225 (2006).
- 147 18. Schwarz, G. Estimating the dimension of a model. Ann. Stat. 6, 461–464 (1978).
- 148 19. Kelley, C. T. Iterative methods for optimization (SIAM, 1999).
- 149 20. Turchin, P. Complex population dynamics: a theoretical/empirical synthesis (Princeton university press, 2003).