Session 8: Chemical Communication in **PhysiCell**



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What we learned last session

- PhysiCell full modeling workflow
- Handy C++ Tidbits for Cell Agents
- Typical form / syntax of PhysiCell functions
- The customizable functions in Cell.functions
- How to assign new functions to a cell definition
- Sampling the microenvironment at Cell locations
- Controlling initial cell placement
- Custom coloring functions

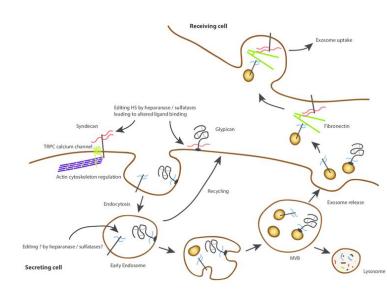
What we will learn in session 8

- 1. Secretion, uptake, export
 - How do we expect cells to communicate
 - How do we handle this process
 - ♦ Export/ Secretion/ Uptake definitions
- 2. Example: Quorum sensing (advanced)
 - Edit .cpp files to add custom functionality
 - Edit rates
 - Edit signal interactions

github repository

Chemical communication

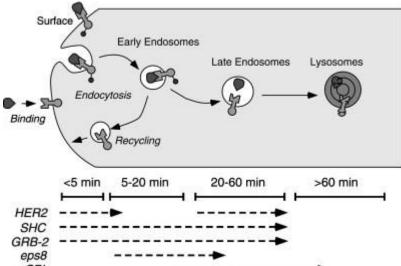
- Cells can communicate by secreting chemical factors:
 - Some stick to the extracellular matrix
 - ♦ Proteoglycans, Some forms of VEGF
 - ♦ Good for leaving "memory"
 - Many diffuse out into the surrounding tissue
 - ♦ IL6, VEGF, CXCL12, CXC/SDF1, Estrogen
 - ♦ Good for long distance communication
 - Some factors never leave the cell surface
 - ◆ Delta-like proteins (e.g., Dll4)
 - ♦ Adhesion molecules
 - ♦ Good for contact signals



Couchman, John & Multhaupt, Hinke & Sanderson, Ralph. (2016). Recent Insights into Cell Surface Heparan Sulphate Proteoglycans and Cancer. F1000Research. 5. 1541. 10.12688/f1000research.8543.1.

Chemical receptors

Chemical signaling by receptors can be rate limited by receptor trafficking



Burke, P et al. "Regulation of epidermal growth factor receptor signaling by endocytosis and intracellular trafficking." Molecular biology of the cell vol. 12,6 (2001):

• If some seeps are shown that the receptors and lose response to further signal.

Indirect chemical communication

- Single-cell processes can deplete chemical resources
 - Consume resource during metabolism
 - Consume glucose in metabolism
 - Consume growth factors during cycling
- Gradients emerge due to non-homogeneous distribution of cells
- Amounts of chemical substrates has information on local conditions
- Substrate gradients have information on the nearby environment
- Thus, even when cells don't "intend" to communicate, they send information just by altering the chemical environment. This is an indirect communication

Export mathematics

Suppose each cell exports q at a constant rate E.

$$\frac{\partial q}{\partial t} = \nabla \cdot D \nabla q - \lambda q + \sum_i \delta(x - x_i) E$$
 Now, define $Q(t) = \int_{\Omega} q \, dV$ to be the total q in the domain.

We can then find,

Q is proportional to the number of cells that secrete q.

Secretion mathematics

You could also use regular secretion:

$$\frac{\partial q}{\partial t} = \nabla \cdot D\nabla q - \lambda q + \sum_{i} \delta(\mathbf{x} - \mathbf{x}_{i}) V_{i} S_{i}(q^{*} - q)$$

What will happen:

- 1. Cells will tend secrete until the nearby q density reaches q^*
- 2. q will have higher values near larger concentrations of cells
- 3. ∇q will point towards cells.

BUT:

- 1. $\int_{\Omega} q \ dV$ will **not** be proportional to the total cell count
- 2. Even a small population can drive q towards q^* for sufficiently large S_i

Uptake mathematics

Uptake is defined as,

$$\frac{\partial q}{\partial t} = \nabla \cdot D\nabla q - \lambda q - \sum_{i} \delta (\mathbf{x} - \mathbf{x}_{i}) V_{i} U_{i} q$$

We then expect uptake dependant on the rate constant U

Accessing internalized substrates

- By default, PhysiCell keeps track of the net amount of internalized substrates.
- Each environmental substrate has a corresponding internalized substrate with the same index.

```
phenotype.molecular.internalized_total_substrates[ index ]
```

- Cells can release their contents at death. Set this (on a per-substrate basis) via phenotype.molecular.fraction_released_at_death[index]
- Similarly, if the cell is eaten, the attacking cell can acquire some or all of the contents phenotype.molecular.fraction transferred when ingested[index]

WARNING: If cells are secreting (or exporting), the internalized substrates can go to negative values unless you write code to internally generate this quantity. Use the "at death / when ingested" options with caution.

Biological example: Quorum sensing

Quorum sensing (Advanced)

- How can cells "see" or "count" how many cells are nearby?
- How can cells find nearby cells?
- How can cells build an army before attacking?

Quorum factors!

- Many cells (particularly bacteria) secrete a diffusible factor to help it communicate with others of its kind.
- Quorum factors communicate two key pieces of information:
 - They accumulate in regions of high cell density
 - The gradient points towards regions of higher cell density

Using what we know (1)

- Let's create a new project:
 - Diffusing substrate: resource
 - ♦ initial value: 0.25 (dimensionless)
 - \bullet D = 100000 μ m²/min
 - \star $\lambda = 0.1 1/min$
 - boundary value: 0.25 on x_{\min} ,
 - zero flux on all other boundaries.
 - Cells of type "bacteria":
 - ♦ Uptake resource at rate chosen to get 100 micron length scale
 - ♦ They move by chemotaxis up resource gradients.
 - Place in a circle in the center
 - Cells of type "supplier":
 - ♦ Release resource at a high rate towards saturation value of 1
 - No birth or death
 - ◆ Can't be moved
 - ♦ Place randomly in the domain

Using what we know (2) (round 1)

- Let's start with one diffusing substrate:
 - resource (Dirichlet condition 1 mmHg)
- Cell type "bacteria":
 - Proliferate proportional to resource
 - Die if resource is below a threshold
- Agent type "supplier":
 - Don't proliferate or die.
 - Can't be moved
 - Release resource

Approach

Change Dirichlet condition for resource in PhysiCell_settings.xml

- Add custom variables (to default definition):
 - R_necrosis, R_max_growth, necrosis_rate
- Create and use phenotype function for bacteria:
 - bacteria_phenotype

First, let's get a clean template project

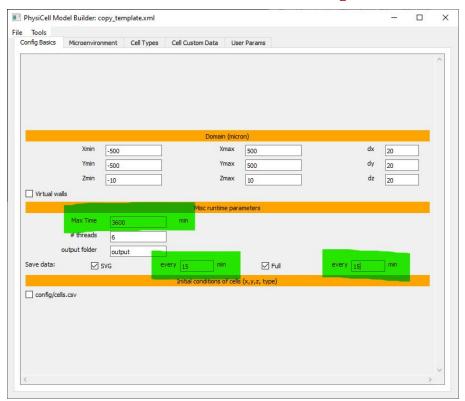
- make data-cleanup
- make reset
- make template

Simulation time and interval for outputs

From PhysiCell\config (anaconda)

python ..\..\PhysiCell-model-builder\bin\gui4xml.

Set to 3600 minutes of simulation Output SVG files every 15 minutes

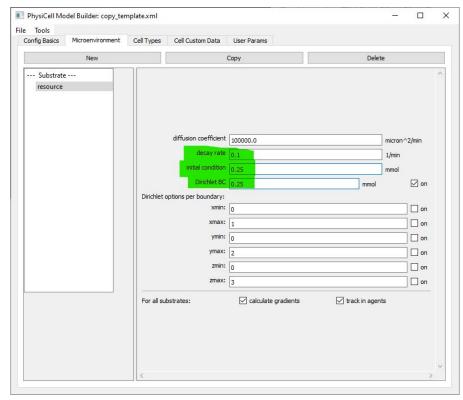


ру

Changes to PhysiCell_settings.xml (1)

Set up the substrate

- Rename substrate to resource
- Change decay rate, initial condition, and boundary condition values.



Create the new cell definitions

Create a definition to bacteria (math)

• We want a diffusion distance of 10, and D = 100000 µm²/min. We need the uptake rate U:

100
$$\mu m = L = \sqrt{\frac{D}{U}} = \sqrt{\frac{100000 \ \mu m^2 / \text{min}}{U}}$$

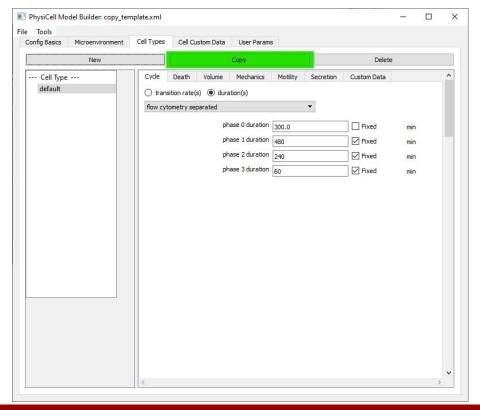
• Solving for *U*, we have:

$$U = \frac{D}{L^2} = \frac{100000 \ \mu m^2 / \text{min}}{10000 \ \mu m^2} = 10 \ \text{min}^{-1}$$

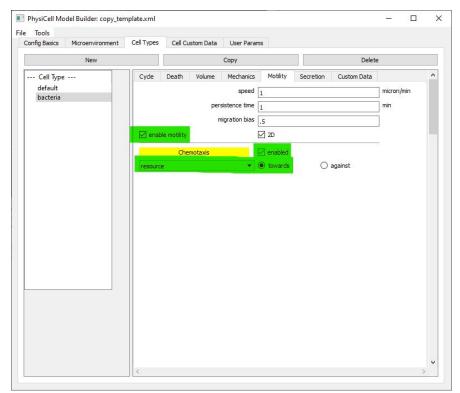
Changes to PhysiCell_settings.xml (2)

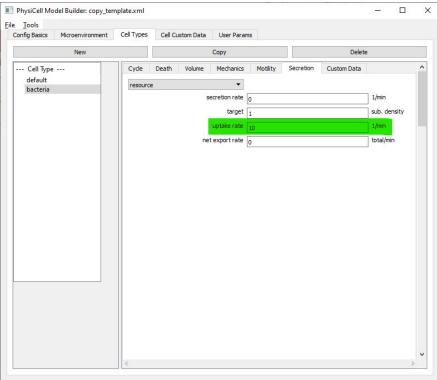
Create bacteria definition from default type

- Click on copy
- Activate chemotaxis on resource gradient direction
- Set the resource update rate



Changes to PhysiCell_settings.xml (3)

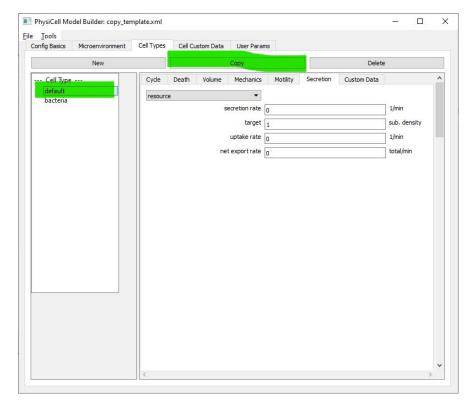




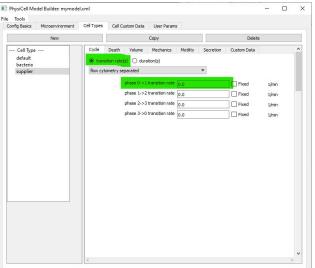
Changes to PhysiCell_settings.xml (4)

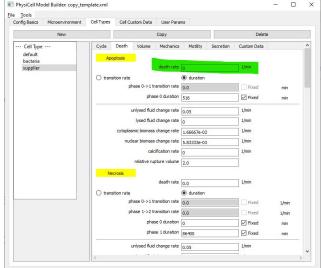
Create supplier definition from default type

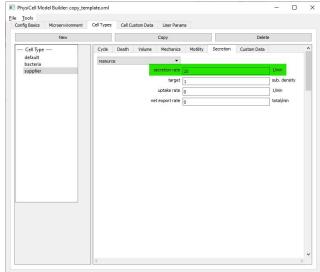
- Select default type and click on copy
- Set proliferation off
- Set dead off
- Set the resource secretion



Changes to PhysiCell_settings.xml (5)







Changes in custom.cpp

Change to create_cell_types()

```
/*
   Put any modifications to individual cell definitions here.
   This is a good place to set custom functions.
* /
cell defaults.functions.update phenotype = phenotype function;
cell defaults.functions.custom cell rule = custom function;
cell_defaults.functions.contact_function = contact_function;
Cell Definition* pBacteria = find cell definition( "bacteria" );
pBacteria->functions.update phenotype = bacteria phenotype;
   This builds the map of cell definitions and summarizes the setup.
build cell definitions maps();
display cell definitions (std::cout);
return;
```

Modify cell placement

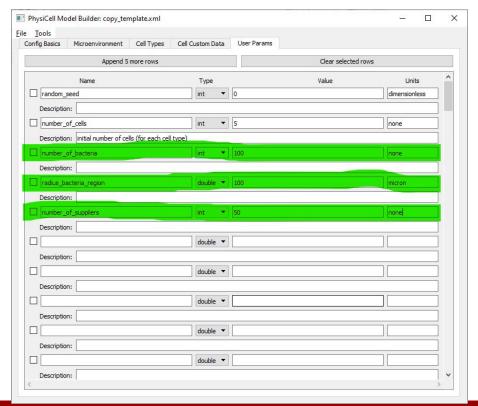
Changed to custom.cpp (setup_tissue)

```
void setup tissue ( void )
        double Xmin = microenvironment.mesh.bounding box[0];
        double Ymin = microenvironment.mesh.bounding box[1];
        double Zmin = microenvironment.mesh.bounding box[2];
        double Xmax = microenvironment.mesh.bounding box[3];
        double Ymax = microenvironment.mesh.bounding box[4];
        double Zmax = microenvironment.mesh.bounding box[5];
        if ( default microenvironment options.simulate 2D == true )
                 Zmin = 0.0;
                 Zmax = 0.0;
        double Xrange = Xmax - Xmin;
        double Yrange = Ymax - Ymin;
        double Zrange = Zmax - Zmin;
        double center x = 0.5*(Xmin+Xmax);
        double center y = 0.5*(Ymin+Ymax);
        double center z = 0.5*(Zmin+Zmax);
        // create some of each type of cell
        Cell* pC;
        // find cell definitions
        Cell Definition* pBacteria = find cell definition( "bacteria" );
```

```
// find cell definitions
Cell Definition* pBacteria = find cell definition( "bacteria" );
Cell Definition * pSupplier = find cell definition ( "supplier" );
for (int k=0; k<parameters.ints("number of bacteria"); k++ )
         std::vector<double> position = {0,0,0};
        double r = NormalRandom(0,1) *
        parameters.doubles( "radius bacteria region" );
         double theta = 6.28318530718 * UniformRandom();
        position[0] = center x + r*cos(theta);
        position[1] = center y + r*sin(theta);
        position[2] = center z;
        pC = create cell( *pBacteria );
        pC->assign position( position );
for( int k=0; kparameters.ints("number of suppliers"); k++ )
        std::vector<double> position = {0,0,0};
        position[0] = Xmin + UniformRandom()*Xrange;
        position[1] = Ymin + UniformRandom()*Yrange;
        position[2] = Zmin + UniformRandom()*Zrange;
        pC = create cell( *pSupplier );
        pC->assign position( position );
        pC->is movable = false;
return;
```

Changes to PhysiCell_settings.xml (6)

Define parameters on xml file



Bacteria phenotype (1)

```
// declare function in custom.h
void bacteria phenotype( Cell* pCell, Phenotype& phenotype , double dt );
// create it in custom.cpp
void bacteria phenotype ( Cell* pCell, Phenotype& phenotype , double dt )
       if ( phenotype.death.dead == true )
               pCell->functions.update phenotype = NULL; // don't bother doing this function again!
               return;
       // find my cell definition
       // don't use static if you plan to use this for more than one cell type
       static Cell Definition* pCD = find cell definition( pCell->type name );
       // find the index of resource
       static int nR = microenvironment.find density index( "resource" );
       // index of necrotic death model
       static int nNecrosis = 1; // PhysiCell constants::necrosis death model;
       // sample microenvironment at cell position to get resource
       double R = pCell->nearest density vector()[nR];
       // check for necrotic death
       if( R < pCell->custom data["R necrosis"]
         phenotype.death.rates[nNecrosis] = pCell->custom data["necrosis rate"]; }
       { phenotype.death.rates[nNecrosis] = 0.0; }
       // ...
```

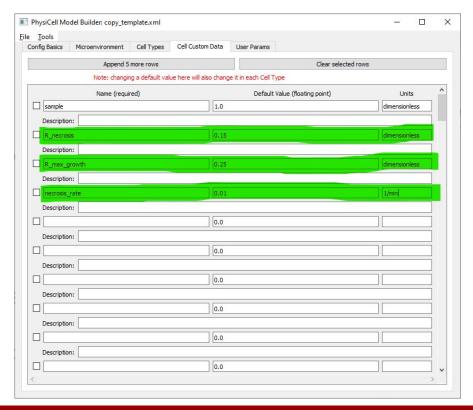
Bacteria phenotype (2)

```
// ...
// set birth rate
// set proliferation
// first, set to the cell line rate
phenotype.cycle.data.transition rate(0,1) =
    pCD->phenotype.cycle.data.transition rate(0,1);
// scale with R
double scaling factor = (R - pCell->custom data["R necrosis"])
    / (pCell->custom data["R max growth"] - pCell->custom data["R necrosis"]);
if (scaling factor > \overline{1})
{ scaling factor = 1.0; }
if ( scaling factor < 0 )
{ scaling \overline{factor} = 0.0; }
// multiply by scaling factor
phenotype.cycle.data.transition rate(0,1) *= scaling factor;
return;
```

Changes to PhysiCell_settings.xml (7)

Update the default cell definition

Include to the custom data:
 R_necrosis, R_max growth, and necrosis_rate

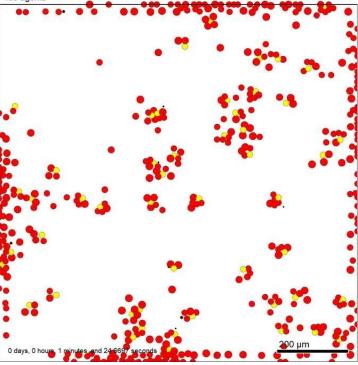


Give it a try!

```
make data-cleanup
make
.\project .\config\mymodel.xml
```

make jpeg
make movie

Current time: 2 days, 1 hours, and 0.00 minutes, z = 0.00 µm 488 agents



Link video

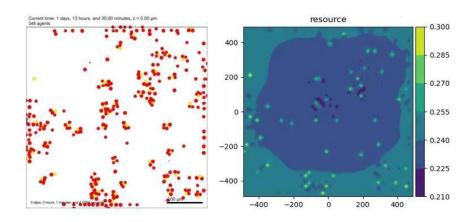
Optional visualization (advanced)

Load <u>plot_CellSubs.py</u> on \beta \\ script to plot cells + substrates using pyMCDS.py

python beta\plot_CellSubs.py 0 240 1 output

Load <u>Makefile</u> on \PhysiCell \\ generate movie cells + substrates **make movie2**

Warning: It's necessary to run 'make ipeg' before that.



Link video

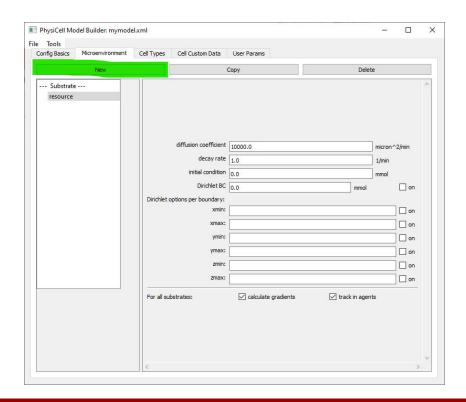
Using what we know (2) (round 2)

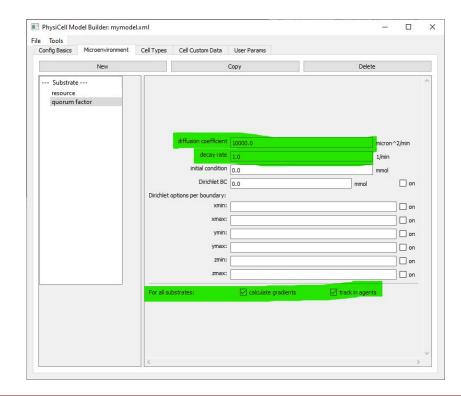
- Let's use two diffusing substrates:
 - resource (Dirichlet condition 1 mmHg)
 - quorum factor (Neumann condition)
- Cell type "bacteria":
 - Proliferate proportional to resource
 - Die if resource is below a threshold
 - Secrete q
 - Chemotax towards regions of high q
- Agent type "supplier":
 - Don't proliferate or die.
 - Can't be moved
 - Release resource

Approach

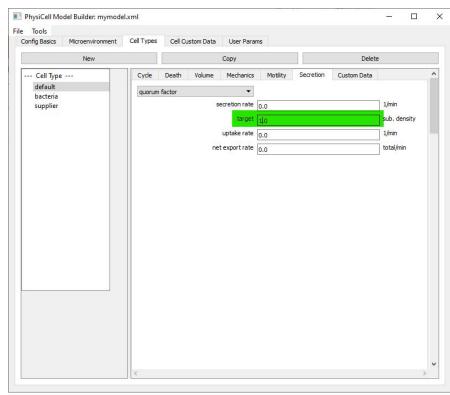
- In PhysiCell_settings.xml
 - Add "quorum factor" to microenvironment
 - ♦ Let's use a diffusion coefficient of 10000, decay rate of 1
 - ♦ Neumann conditions!
 - Add corresponding secretion / uptake to default cell definition
 - make sure bacteria export quorum factor
 - ♦ Let's use a rate of 1 for now.
 - change bacteria chemotaxis to quorum factor

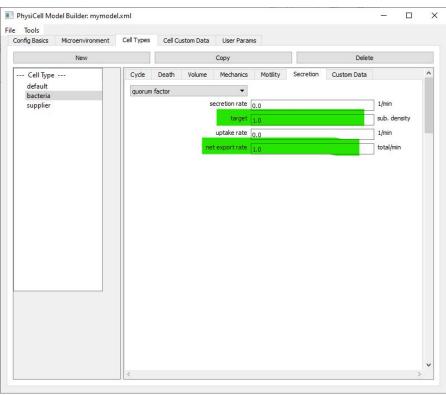
Changes to PhysiCell_settings.xml (1)



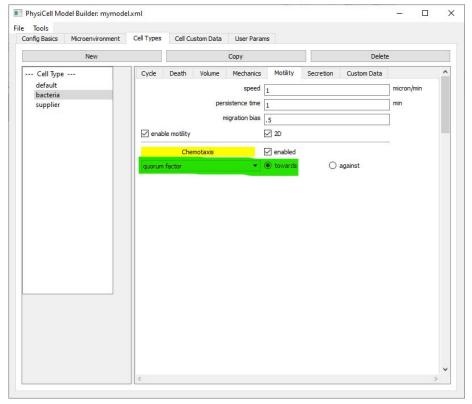


Changes to PhysiCell_settings.xml (2)





Changes to PhysiCell_settings.xml (3)

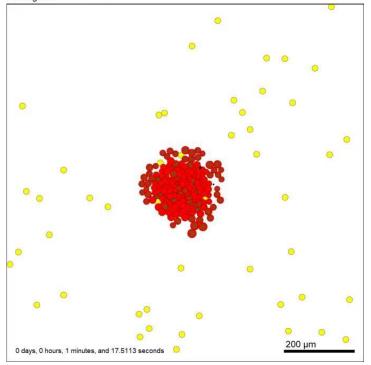


Give it a try!

make data-cleanup
make
.\project .\config\mymodel.xml

make jpeg
make movie

Current time: 1 days, 19 hours, and 15.00 minutes, z = 0.00 μm 242 agents



Link video



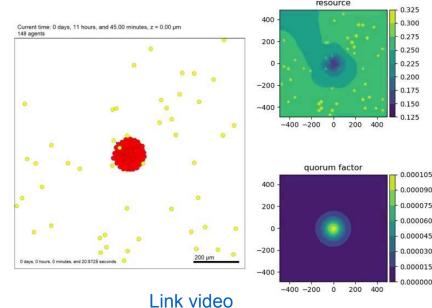
Optional visualization (advanced)

Load plot CellSubs.py on \beta \\ script to plot cells + substrates using pyMCDS.py

python beta\plot CellSubs.py 0 240 1 output

Load Makefile on \PhysiCell \\ generate movie cells + substrates make movie2

Warning: It's necessary to run 'make ipeq' before that.





Using what we know (2) (round 3)

- Let's use two diffusing substrates:
 - resource (Dirichlet condition 1 mmHg)
 - quorum factor (Neumann condition)

• Cell type "bacteria":

- Proliferate proportional to resource
- Die if resource is below a threshold
- Secrete q
- Chemotax towards regions of high q
- Slow down motility when q is high

Agent type "supplier":

- Don't proliferate or die.
- Can't be moved
- Release resource

Approach

- In PhysiCell_settings.xml
 - Add custom data to default cell definition
 - ◆ quorum_motility_slowdown (we'll default to 1e-4)
- In custom.cpp
 - In bacteria_phenotype()
 - ullet scale phenotype.motility.speed by $\max\left(0.1 \frac{q}{q_{mot}}\right)$

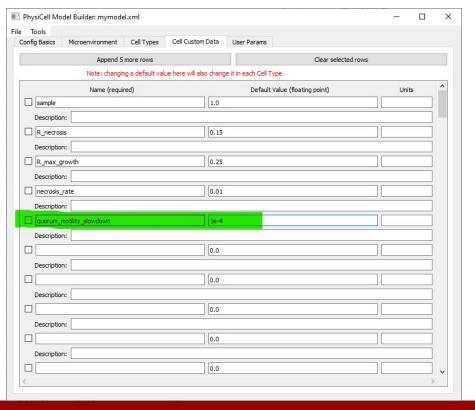
Changes to bacteria phenotype (1)

```
void bacteria phenotype (Cell* pCell, Phenotype phenotype , double dt )
      if( phenotype.death.dead == true )
            pCell->functions.update phenotype = NULL; // don't bother doing this function again!
            return;
      // find my cell definition
      // don't use static if you plan to use this for more than one cell type
      static Cell Definition* pCD = find cell definition( pCell->type name );
      // find the index of resource
      static int nR = microenvironment.find density index( "resource" );
      // find the index of quorum factor
      static int nQ = microenvironment.find density index( "quorum factor" );
      // index of necrotic death model
      static int nNecrosis = 1; // PhysiCell constants::necrosis death model;
      // sample microenvironment at cell position to get resource
      double R = pCell->nearest density vector()[nR];
      double q = pCell->nearest density vector()[nQ];
      // ...
```

Changes to bacteria phenotype (2)

```
// ...
// get the cell line's motile speed
phenotype.motility.migration speed = pCD->phenotype.motility.migration speed;
// get a scaling factor
scaling factor = 1.0 - q / pCell->custom data["quorum motility slowdown"];
if( scaling factor < 0.0 )</pre>
{ scaling factor = 0.0; }
// scale migration speed
phenotype.motility.migration speed *= scaling factor;
return;
```

Changes to PhysiCell_settings.xml

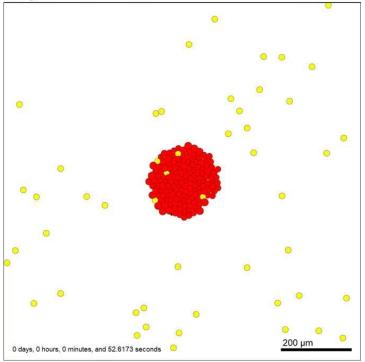


Give it a try!

```
make data-cleanup
make
.\project .\config\mymodel.xml
```

```
make jpeg
make movie
```

Current time: 1 days, 5 hours, and 0.00 minutes, $z = 0.00 \mu m$ 206 agents



Link video



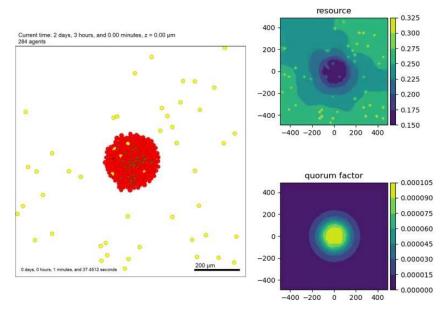
Optional visualization (advanced)

Load <u>plot_CellSubs.py</u> on \beta \\ script to plot cells + substrates using pyMCDS.py

python beta\plot_CellSubs.py 0 240 1 output

Load <u>Makefile</u> on \PhysiCell \\ generate movie cells + substrates **make movie2**

Warning: It's necessary to run 'make jpeg' before that.



Link video

Using what we know (2) (round 4)

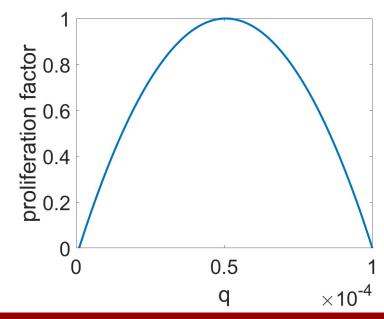
- Let's use two diffusing substrates:
 - resource (Dirichlet condition 0.25)
 - Quorum factor (Neumann condition)
- Cell type "bacteria":
 - Proliferate proportional to resource
 - Low proliferation when q is low (too far from colony) or q is too high (overcrowding)
 - Die if resource is below a threshold
 - Secrete q
 - Chemotax towards regions of high q
 - Slow down motility when q is high

Agent type "supplier":

- Don't proliferate or die.
- Can't be moved
- Release resource

Approach

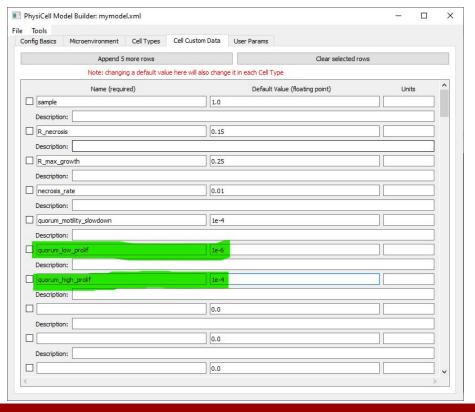
- In PhysiCell_settings.xml
 - Add custom data to default cell definition
 - ◆ quorum_low_proliferation (default this to 1e-6)
 - ♦ quorum_high_proliferation(default this to 1e-4)
- In custom.cpp
 - In bacteria_phenotype()
 - ♦ proliferation is zero if q < qL or if q > qH
 - scale proliferation by $\frac{4}{(q_H q_L)^2} (q q_L) (q_H q)$



Changes to bacteria_phenotype()

```
// ...
// scale with R
double scaling factor = (R - pCell->custom data["R necrosis"])
     / (pCell->custom data["R max growth"] - pCell->custom data["R necrosis"]);
if( scaling factor > 1 )
{ scaling factor = 1.0; }
if( scaling factor < 0 )</pre>
{ scaling factor = 0.0; }
// scale with quorum factor
double Olow = pCell->custom data["quorum low prolif"];
double Qhigh = pCell->custom data["quorum high prolif"];
double constant = 4.0 / pow( Qhigh - Qlow , 2.0 );
// no proliferation if Q is low or high
if (q < Qlow | | q > Qhigh)
{ scaling factor = 0.0; }
scaling factor *= ( constant*(q-Qlow)*(Qhiqh-q) );
// multiply by scaling factor
phenotype.cycle.data.transition rate(0,1) *= scaling factor;
```

Changes to PhysiCell_settings.xml

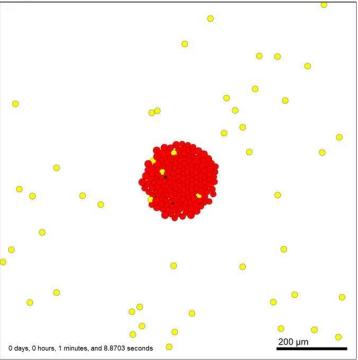


Give it a try!

```
make data-cleanup
make
.\project .\config\mymodel.xml
```

make jpeg
make movie

Current time: 1 days, 9 hours, and 45.00 minutes, z = 0.00 μ m 226 agents



Link video



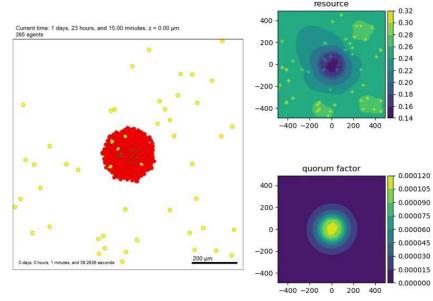
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Load <u>plot_CellSubs.py</u> on \beta \\ script to plot cells + substrates using pyMCDS.py

python beta\plot_CellSubs.py 0 240 1 output

Load <u>Makefile</u> on \PhysiCell \\ generate movie cells + substrates **make movie2**

Warning: It's necessary to run 'make ipeg' before that.



Link video

Using what we know (2) (round 5)

- Let's use two diffusing substrates:
 - resource (Dirichlet condition 1)
 - quorum factor (Neumann condition)

• Cell type "bacteria":

- Proliferate proportional to resource
- Low proliferation when q is low (too far from colony) or q is too high (overcrowding)
- Die if resource is below a threshold
- Secrete q
- Chemotax towards regions of high q
- Slow down motility when q is high
- Increase migration bias as q increases (random wandering when far from colonies)

Agent type "supplier":

- Don't proliferate or die.
- Can't be moved
- Release resource

Approach

- In custom.cpp
 - In bacteria_phenotype()
 - scale motility bias by $\frac{q}{q_m}$

Changes to bacteria_phenotype()

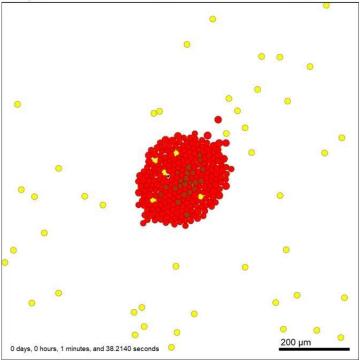
```
// get the cell line's motile speed
phenotype.motility.migration speed = pCD->phenotype.motility.migration speed;
// get a scaling factor
scaling factor = 1.0 - 1 / pCell->custom data["quorum motility slowdown"];
if (scaling factor < 0.0)
{ scaling factor = 0.0; }
// scale migration speed
phenotype.motility.migration speed *= scaling factor;
// scale migration bias by the quorum factor
scaling factor = q / pCell->custom data["quorum motility slowdown"];
if ( scaling factor > 1.0 )
{ scaling factor = 1.0; }
phenotype.motility.migration bias = pCD->phenotype.motility.migration bias;
phenotype.motility.migration bias *= scaling factor;
return;
```

Give it a try!

```
make data-cleanup
make
.\project .\config\mymodel 4.xml
```

make jpeg
make movie

Current time: 2 days, 5 hours, and 0.00 minutes, $z = 0.00 \mu m$ 298 agents



Link video



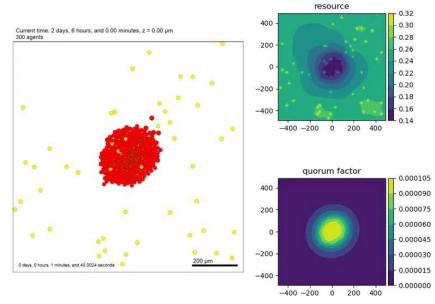
Optional visualization (advanced)

Load <u>plot_CellSubs.py</u> on \beta \\ script to plot cells + substrates using pyMCDS.py

python beta\plot_CellSubs.py 0 240 1 output

Load <u>Makefile</u> on \PhysiCell \\ generate movie cells + substrates **make movie2**

Warning: It's necessary to run 'make jpeg' before that.



Link video

Using what we know (2) (round 6)

- Let's use two diffusing substrates:
 - resource (Dirichlet condition 1)
 - quorum factor (Neumann condition)

Cell type "bacteria":

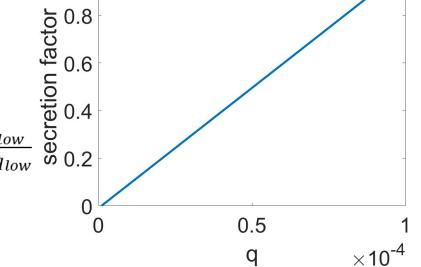
- Proliferate proportional to resource
- Low proliferation when q is low (too far from colony) or q is too high (overcrowding)
- Die if resource is below a threshold
- Secrete q
- Chemotax towards regions of high q
- Slow down motility when q is high
- Increase migration bias as q increases (random wandering when far from colonies)

Agent type "supplier":

- Don't proliferate or die.
- Can't be moved
- Release resource
- resource release increases when q is high (dynamic response to "needs" of nearby colony)

Approach

- In PhysiCell_settings.xml
 - Create new custom cell data:
 - ♦ max_R_release_rate, min_R_release_rate
 - ♦ q_max_R_release , q_min_R_release
- In custom.cpp / custom.h
 - Create supplier_phenotype()
 - Linear R release rate: $r_{low} + (r_{hi} r_{low}) \frac{q q_{low}}{q_{hi} q_{low}}$
 - Make sure supplier use this function



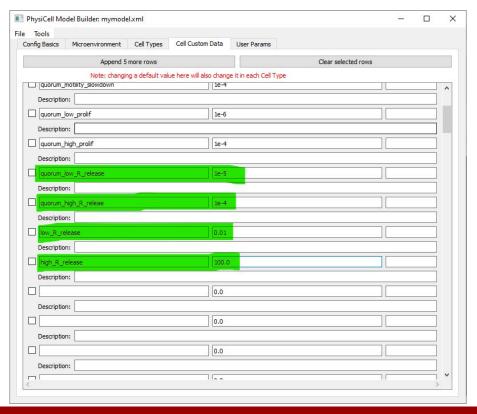
Use the function (create_cell_types())

```
// ...
cell defaults.functions.update phenotype = phenotype function;
cell defaults.functions.custom cell rule = custom_function;
Cell Definition* pBacteria = find cell definition( "bacteria" );
pBacteria->functions.update phenotype = bacteria phenotype;
Cell Definition* pSupplier = find cell definition( "supplier" );
pSupplier->functions.update phenotype = supplier phenotype;
   This builds the map of cell definitions and summarizes the setup.
build cell definitions maps();
display cell definitions( std::cout );
return;
```

Supplier phenotype

```
void supplier phenotype (Cell* pCell, Phenotype phenotype , double dt )
      // find my cell definition
      // don't use static if you plan to use this for more than one cell type
      static Cell Definition* pCD = find cell definition( pCell->type name );
      // find the index of resource
      static int nR = microenvironment.find density index( "resource" );
      // find the index of quorum factor
      static int nQ = microenvironment.find density index( "quorum factor" );
      // sample microenvironment at cell position to get resource and quorum factor
      double q = pCell->nearest density vector()[nQ];
      double Qlow = pCell->custom data["quorum low R release"];
      double Qhigh = pCell->custom data["quorum high R release"];
      double Rlow = pCell->custom data["low R release"];
      double Rhigh = pCell->custom data["high R release"];
      if(q < Qlow)
      { phenotype.secretion.secretion rates[nR] = Rlow; return; }
      if ( a > Ohigh )
      { phenotype.secretion.secretion rates[nR] = Rhigh; return; }
      double scaling factor = (q-Qlow)/(Qhigh-Qlow);
      phenotype.secretion.secretion rates[nR] = Rlow + (Rhigh-Rlow)*scaling factor;
      return;
Add void supplier phenotype (Cell* pCell, Phenotype& phenotype , double dt ) to custom.h
```

Changes in PhysiCell_settings.xml



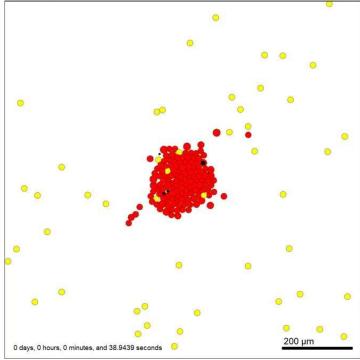
Give it a try!

```
cp ..\PhysiCell-model-builder\mymodel.xml
.\config\mymodel 6.xml
```

```
make data-cleanup
make
.\project .\config\mymodel 6.xml
```

make jpeg
make movie

Current time: 0 days, 21 hours, and 15.00 minutes, $z = 0.00 \mu m$ 186 agents



Link video



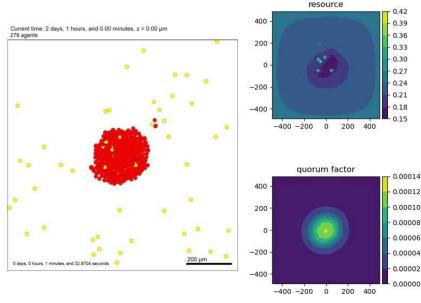
Optional visualization (advanced)

Load <u>plot_CellSubs.py</u> on \beta \\ script to plot cells + substrates using pyMCDS.py

python beta\plot_CellSubs.py 0 240 1 output

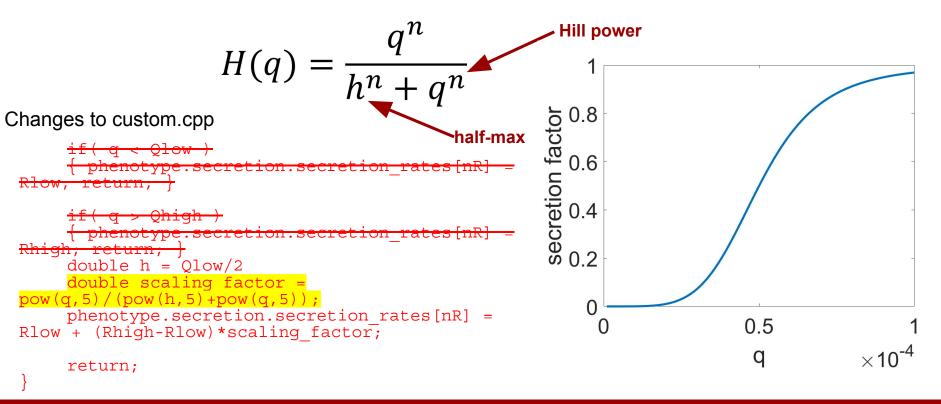
Load <u>Makefile</u> on \PhysiCell \\ generate movie cells + substrates **make movie2**

Warning: It's necessary to run 'make ipeg' before that.



Link video

Mathematics: Hill Functions





3-Types model (Intermediate homework • In physics, the **3-body problem** shows how 3 objects with very simple interactions (gravitation) can

- demonstrate chaotic behavior.
- Let's build a similar system for biology!
- 3 cell types (A,B,C) each secrete their own chemical factor
 - visualization: assume each cell fluoresces proportionally to its signal
- Each cell type can:
 - divide and die in response to resource (R), A, B, C, and pressure
 - move in response to A, B, C, and R
 - secrete (or not secrete) in response to A, B, C, and R
- What can happen in this general system?



Try this model yourself!

https://nanohub.org/tools/pc3types

Mathematics: Hill Functions

• Let's define a *Hill function* H(s):

$$H(s):$$

$$H(s) = \frac{s^n}{s^n + h^n}$$
half-max

Let's write:

$$U = \text{sum of promoter signals}$$

 $D = \text{sum of inhibitor signals}$

We can use these signals to control a parameter p via a Hill function:

$$p = \left[p_0 + \underbrace{(p_{\text{max}} - p_0)H(U)}^{p \to p_{\text{max}}} \underbrace{\text{as } U \to \infty}_{1 - H(D)} \right]$$

Exercise: tumor competition

Let's look at 2 tumor sub-clones:

- Population A:
 - Regular proliferation and death

- Population B:
 - Increased proliferation
 - Cost: more sensitive to resource depletion

Build the model

- Cell Type A:
 - Use default parameters

- Cell Type B:
 - double B_base_cycle to 0.00144
 - double B_max_cycle to 0.0144
 - increase B_necrosis_threshold to 0.6
- set to no Type C cells
 - number of C = 0

Run the model

 How do pink (Type A) and green (Type B) cells compete in high-resource regions?

 How do pink (Type A) and green (Type B) cells compete in low-resource regions?

Exercise: tumor co-option of stromal cells

- Let's look at two interacting cell populations:
 - Tumor cells attract stromal cells and "convince" them to secrete a growth factor.
- Population A (tumor):
 - Secretes signal A
 - Signal B promotes proliferation
 - No proliferation without signal B
- Population B (stromal):
 - No proliferation without signal A
 - Chemotaxis towards Signal A (and stops in regions of high signal A)
 - Signal A promotes secretion of Signal B

Build the model (1)

- Cell Type A:
 - Cycling
 - ◆ A base cycle = 0 (no proliferation without signal B)
 - ♦ A_max_cycle = 0.00072 (slow the kinetics down a bit)
 - ♦ A_cycle_B = promote (signal B enables proliferation)

4

- Cell Type B:
 - Cycling
 - ♦ B_base_cycle = 0
 - ♦ B_max_cycle = 0.000072 (less proliferation than tumor cells)
 - ♦ B_cycle_A (signal A promotes proliferation)
 - Secretion
 - ♦ B_base_secretion = 0 (no secretion without signal A)
 - ♦ B_signal_A = promote (signal A stimulates secretion)
 - Motility
 - ♦ B_speed_base = 0.5 (increase the base speed a bit)
 - ◆ B_speed_A = inhibit (slow down when you reach the tumor)

Build the model (2)

- Now, switch to the cell types tab, and select Type B
- Let's turn on chemotaxis

- Cell Type B:
 - phenotype:motility
 - enabled = on (turn on motility)
 - ♦ chemotaxis:
 - » enabled = on (use chemotaxis to guide migration)
 - » substrate = signal A (chemotaxis towards tumor cells)
 - » direction = 1 (move up the gradient towards stronger signal A)

Run the model

- Set 25 initial type A cells, and 1 initial type B cell. 0 Type C cells.
 - Where do green (type B) cells end up?
 - Where do you see the most pink cells (Type A)?
- Increase to 5 initial type B cells.
 - Where do green (type B) cells end up?
 - Where do you see the most pink cells (Type A)?
- Increase B_cycle_max to 0.00018. What happens?
- Set 0 initial Type B cells. What happens?

Exercise: a population with a quorum factor

 Cells can use quorum factors to find each other and sense population size.

- Population A (bacteria):
 - Secretes signal A
 - Signal A promotes proliferation
 - Movement towards signal A (help for aggregation)
 - Signal A inhibits migration (they stop moving when they find their home)

Build the model

Cell Type A:

- Cycling
 - ◆ A_base_cycle = 0.0 (no proliferation without signal B)
 - ◆ A_max_cycle = 0.0072 (fast birth once aggregated)
 - ♦ A_cycle_A = promote (signal B enables proliferation)
- secretion
 - ♦ A_signal_A = promote
- motility
 - ♦ A base speed = 1
 - ♦ A_max_speed = 1
 - ♦ A_speed_A = inhibit
- Turn on chemotaxis towards signal A as before in the "cell types" tab

Run the model

- Set 25 initial type A cells, and 0 Type B cells. 0 Type C cells.
 - What happens?
- Set 5 initial type A cells, and 0 Type B cells. 0 Type C cells.
 - What happens?

- Set 1 initial type A cells, and 0 Type B cells. 0 Type C cells.
 - What happens?

Exercise: add attackers

- Cells can use *quorum factors* to find each other and sense population size.
- Population A (bacteria):
 - Secretes signal A
 - Signal A promotes proliferation
 - Signal B promotes death (it's a poison)
 - Movement towards signal A (help for aggregation)
 - Signal A inhibits migration (they stop moving when they find their home)
- Population B (attackers):
 - Migration towards signal A
 - No proliferation or death
 - Signal A promotes secretion of signal B
 - Signal A **inhibits** migration (they stop moving when they find their target)

Build the model

- Cell Type A:
 - death
 - ◆ A_death_B = promote (Type B signal kills it)
 - ♦ A max death 0.005 (B signal increases death)
- · Cell Type B:
 - Cycling
 - ♦ B base cycle = 0.0
 - ♦ B max cycle = 0
 - death
 - ♦ B_base_death = 0
 - ♦ B_max_death = 0
 - secretion
 - ♦ B_signal_A = promote
 - motility
 - ♦ B_base_speed = 1
 - ♦ B_max_speed = 1
 - ♦ B_speed_A = inhibit
- Turn on chemotaxis towards signal A as before in the "cell types" tab

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