

## Kinetic modelling

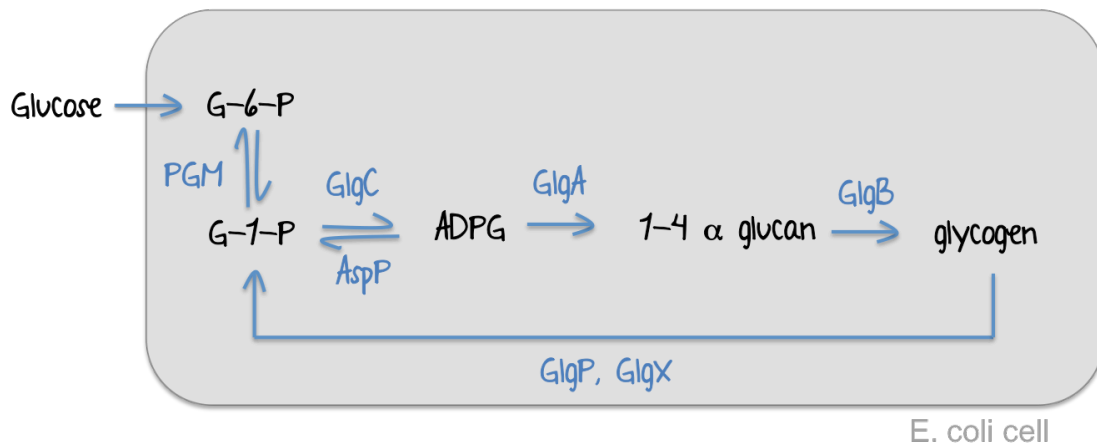
**Assumptions:** We generated a deterministic model taking into consideration the following assumptions:

- All enzyme concentrations are constant and similar. We fixed the enzyme concentrations to  $1 \times 10^{-5}$  mmol/mL
- Glucose is available in excess.
- Reactions follow simple reversible or irreversible Michaelis-Menten kinetics.
- There is no additional flux of substrates after the beginning of the simulation.
- The intracellular metabolite concentrations (ATP, AMP, ADP and PPi) are constant. They are all set to  $1 \times 10^{-5}$  mmol/mL.
- Only competitive inhibition by the product occurs. For simplicity, other external inhibitors are not considered in the model.

### Enzyme kinetic rates

Enzyme	Km (mmol/mL)	Reference
Pgm(G1P)	$2.9 \times 10^{-4}$	(1)
Pgm(G6P)	$5.6 \times 10^{-6}$	(1)
GlgC(ADPG)	$4 \times 10^{-5}$	(2)
GlgC(ATP)	$3.2 \times 10^{-4}$	(2)
GlgA	$3.5 \times 10^{-5}$	(3)
GlgB(Glucan1-4)	$1.42 \times 10^{-5}$	(4)
GlgX (Glycogen)	$1 \times 10^{-6}$	-not found-
GlgP (Glycogen)	$1 \times 10^{-6}$	-not found-

## Glycogen Pathway (5)

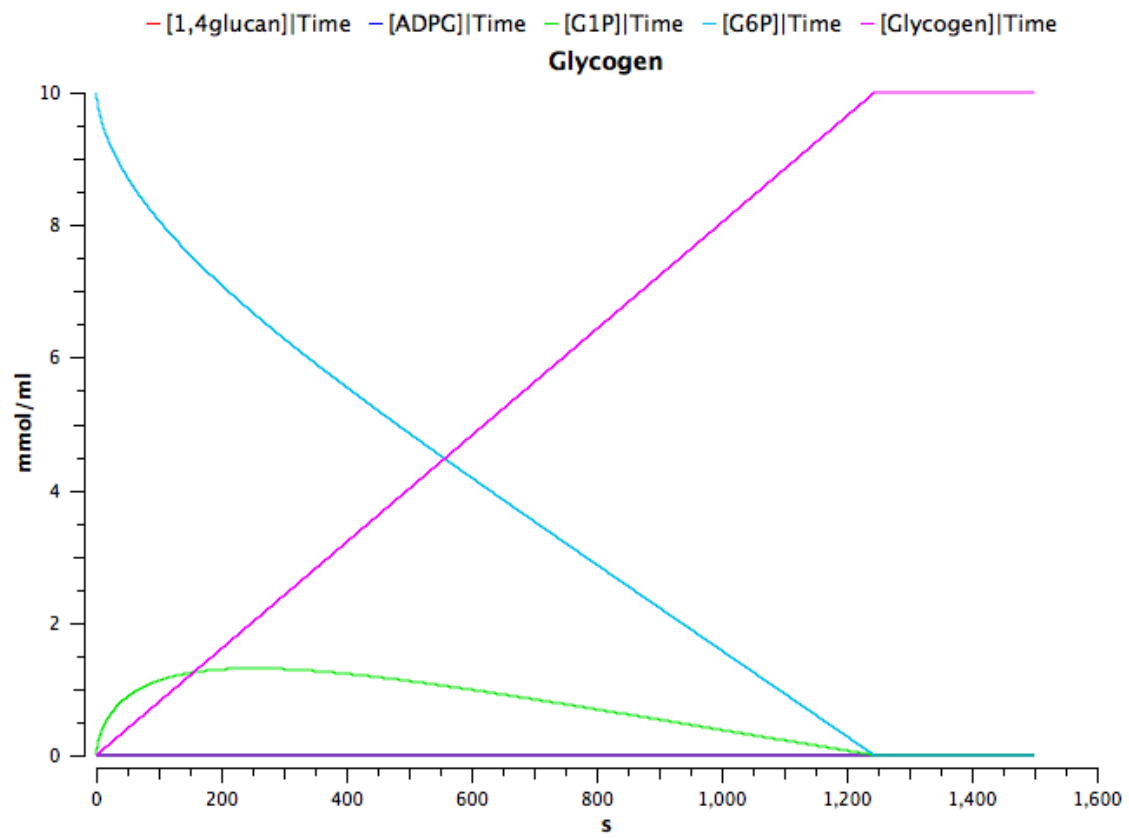


GlgB catalyzes 2 consecutive reactions. First, it cleaves an alpha 1,4 glycosidic linkage in a 1,4-alpha-D-glucan to form a non-reducing-end oligosaccharide chain that is transferred to a C-6 hydroxyl group of the same or another alpha-1,4-D-glucan. (Preiss, J. 2009. Glycogen Biosynthesis.) GlgX and GlgP were considered together as the action of both enzymes is required to get debranching of glycogen.

2 substrate irreversible reaction:  $V_{max} \text{substrateA} \text{substrateB} / (K_m \text{B} \text{substrateA} + K_m \text{A} \text{substrateB} + \text{substrateA} * \text{substrateB})$

All the reactions of the pathway are highly efficient, as all the initial glucose-6-P converted to glucose-1-P and ADP-glucose is used in the production of glycogen. Part of the glucose-1-P is recovered due to the GlgX debranching activity.

We run our model for 2500 seconds and collect the data in intervals of 0.05 seconds starting with a concentration of 10 mmol/mL of Glucose-6-P (G6P). After the time course, the majority of the starting glucose has been used to produce glycogen and only a small part of the glucose stays unbranched.



## Differential equations

$$\frac{d([G6P])}{dt} = - \left( \frac{\frac{V_f(P_{gm}) \cdot [G6P]}{K_{ms}(P_{gm})} - \frac{V_r(P_{gm}) \cdot [G1P]}{K_{mp}(P_{gm})}}{1 + \frac{[G6P]}{K_{ms}(P_{gm})} + \frac{[G1P]}{K_{mp}(P_{gm})}} \right)$$

$$\begin{aligned} \frac{d([G1P])}{dt} = & + \left( \frac{\frac{V_f(P_{gm}) \cdot [G6P]}{K_{ms}(P_{gm})} - \frac{V_r(P_{gm}) \cdot [G1P]}{K_{mp}(P_{gm})}}{1 + \frac{[G6P]}{K_{ms}(P_{gm})} + \frac{[G1P]}{K_{mp}(P_{gm})}} \right) \\ & - \left( \frac{V_{max}(GlgC) \cdot [G1P] \cdot [ATP]}{K_{mB}(GlgC) \cdot K_{mA}(GlgC) + K_{mB}(GlgC) \cdot [G1P] + K_{mA}(GlgC) \cdot [ATP] + [G1P] \cdot [ATP]} \right) \\ & + \left( \frac{V_{(GlgX-GlgP)} [Glycogen]}{K_{m(GlgX-GlgP)} + [Glycogen]} \right) \\ & + \left( \frac{V_{(AspP)} \cdot [ADPG]}{K_{m(AspP)} + [ADPG]} \right) \end{aligned}$$

$$\begin{aligned} \frac{d([ADPG])}{dt} = & - \left( \frac{V_{(GlgA)} \cdot [ADPG]}{K_{m(GlgA)} + [ADPG]} \right) \\ & + \left( \frac{V_{max}(GlgC) \cdot [G1P] \cdot [ATP]}{K_{mB}(GlgC) \cdot K_{mA}(GlgC) + K_{mB}(GlgC) \cdot [G1P] + K_{mA}(GlgC) \cdot [ATP] + [G1P] \cdot [ATP]} \right) \\ & - \left( \frac{V_{(AspP)} \cdot [ADPG]}{K_{m(AspP)} + [ADPG]} \right) \end{aligned}$$

$$\begin{aligned} \frac{d([1,4glucan])}{dt} = & + \left( \frac{V_{(GlgA)} \cdot [ADPG]}{K_{m(GlgA)} + [ADPG]} \right) \\ & - \left( \frac{V_{(GlgB)} \cdot [1,4glucan]}{K_{m(GlgB)} + [1,4glucan]} \right) \end{aligned}$$

$$\begin{aligned} \frac{d([Glycogen])}{dt} = & + \left( \frac{V_{(GlgB)} \cdot [1,4glucan]}{K_{m(GlgB)} + [1,4glucan]} \right) \\ & - \left( \frac{V_{(GlgX-GlgP)} \cdot [Glycogen]}{K_{m(GlgX-GlgP)} + [Glycogen]} \right) \end{aligned}$$

## References

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- cherichia coli. *Metab Eng* 2000, 2:104–14.
2. Figueroa CM, Esper MC, Bertolo A, Demonte AM, Aleanzi M, Iglesias AA, Ballicora MA: Understanding the allosteric trigger for the fructose-1,6-bisphosphate regulation of the aDP-glucose pyrophosphorylase from *escherichia coli*. *Biochimie* 2011, 93:1816–23.
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  5. Wilson WA, Roach PJ, Montero M, Baroja-Fernández E, Muñoz FJ, Eydallin G, Viale AM, Pozueta-Romero J: Regulation of glycogen metabolism in yeast and bacteria. *FEMS Microbiol Rev* 2010, 34:952–85.