Genetic studies through the lens of gene networks

1 When citing this paper, please use the following:

Subirana-Granés M, Hoffman J, Zhang H, Nandi S, Akirtava C, Fotso K, Pividori M. Genetic studies through the lens of gene networks. 2025. Annu. Rev. Biomed. Data Sci. 8. https://doi.org/10.1146/annurev-biodatasci-103123-095355

This manuscript (<u>permalink</u>) was automatically generated from <u>pivlab/annual review of biomedical data science@c8cdd37</u> on October 8, 2024.

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Abstract

This is a very cool paper about something. We are doing this and that. And this are the conclusions. And this is in **boldface**.

Introduction

Understanding the genetics of complex traits remains a formidable challenge, primarily addressed through Genome-Wide Association Studies (GWAS) that identify variant-trait associations without providing a mechanistic understanding of how these variants influence phenotypes. A significant limitation of GWAS is that most associated variants reside in non-coding regions of the genome, underscoring the critical role of gene regulation in generating diverse phenotypes from identical DNA sequences. To bridge this gap, Transcriptome-Wide Association Studies (TWAS) have been developed to link genetic variants to gene expression (eQTLs) or alternative splicing (sQTLs) and subsequently to specific traits or phenotypes. However, TWAS and similar approaches typically focus on single variants or individual genes, thereby overlooking the intricate gene-gene interactions that are pivotal according to modern theories such as the omnigenic model of genetic architecture for complex traits. Capturing these interactions necessitates more sophisticated methodologies. Traditional approaches involve pathways or gene sets, which rely on predefined groups of functionally related genes but are constrained by existing knowledge. Alternatively, QTL analyses like trans-eQTLs and sQTLs explore distal genetic relationships and splicing variations, respectively. Another promising avenue is the construction of gene regulatory networks based on co-expression patterns, introducing the concept of gene modules. These gene modules are often identified using unsupervised learning techniques applied to diverse and heterogeneous datasets, enabling the modeling of both linear and nonlinear relationships within the data. Such methods transcend basic differential expression analyses by capturing higher-order connections between genes. Tools like MAGMA and the more recent PhenoPLIER exemplify this integrative approach by combining variant-trait associations from GWAS or variant-gene expression-trait associations from TWAS with groups of functionally related genes, thereby enhancing the reliability of gene modules by mitigating false positives through additional layers of evidence. Furthermore, these gene modules can be refined using quantitative omnigenic models that incorporate transcription factors (TFs) to better elucidate gene-gene interactions and potentially include aspects of alternative splicing. Ongoing research continues to explore and develop novel methods that more effectively link expression patterns with genetic variation, aiming to provide a more comprehensive mechanistic understanding of the genetic basis of complex traits.

Single-variant / single-gene approaches

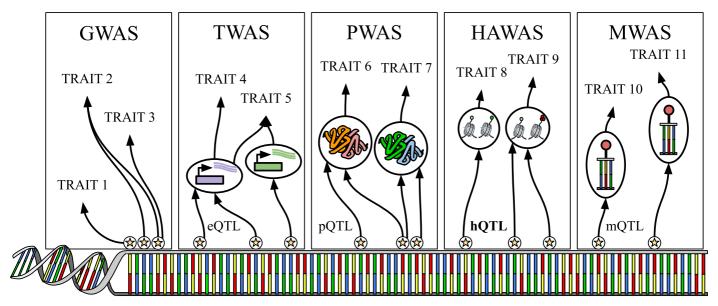


Figure 1: Major approaches based on single-variant/single-gene strategies to understand the genetic basis of complex traits. From left to right GWAS, TWAS, PWAS, HAWAS and MWAS.

Understanding the genetic underpinnings of complex traits and diseases has been a pivotal pursuit in the field of human genetics. Early endeavors predominantly utilized genome-wide association studies (GWAS), which have been instrumental in identifying a foundational map of genomic regions linked to traits, offering valuable insights into the heritable components of complex diseases.

Building upon GWAS findings, efforts have been made to link these associated variants to specific genes, operating under the premise that variation in a single gene can elucidate disease-relevant biology. Techniques such as Transcriptome-Wide Association Studies (TWAS) have emerged, integrating GWAS data with gene expression profiles to pinpoint genes whose expression levels correlate with phenotypic variation. These approaches have advanced our understanding by moving from variant-level associations to functional gene-level insights.

From genetic variants to traits: genome-wide association study (GWAS)

In a simple trait, a single gene can be responsible for the disease, for example in the case of Sickle Cell Anemia and Huntington's disease. However, most traits are not so simplistic and are the result of many mutations across the genome. Even these monogenic traits are impacted by the polygenic background [1]. Genome-wide association studies (GWAS) examine the relationship between genetic variants and specific phenotypes by comparing allele frequencies in individuals of similar ancestry with distinct phenotypic traits (**Figure 1**). The statistical associations are typically assessed using linear regression for continuous traits and logistic regression for binary traits. It is crucial to control for covariates such as age, sex, population structure, and ancestry to minimize confounding effects [2,3]. As many variants are found to be significant and due to the phenomena of linkage disequilibrium between causal and non-causal variants, the results of GWAS are often grouped into risk loci. These loci represent clusters of variants that demonstrate significant associations, although not every variant may be causal. Since the first GWAS or age-related macular degeneration (AMD) in 2005, the GWAS catalog rapidly expanded, containing ~400,000 curated SNP-trait associations from >45,000 published GWAS across >5,000 human traits [4]. These studies have successfully identified risk loci for a variety of traits and diseases such as type 2 diabetes, auto-immune disease, schizophrenia, major depressive disorder, and more [5,6]. As GWAS participants increase, more loci are able to be identified. For example, a study investigating insomnia with a sample size of >1M identified 202 risk loci [7], compared to earlier studies with a sample size of ~110,00, which were only able to identify 3 risk loci [8]. The increased sample sizes allows for identification of more common variants as well as

rare and low-frequency variants [5]. Beyond the identification of risk loci, GWAS has also led to the discovery of novel biological mechanisms. In Crohn's disease, a novel SNP was identified through GWAS in the gene *ATG16L1* (rs2241880) [9]. This SNP causes a missense mutation that enhances caspase-3-mediated cleavage of *ATG16L1* and diminished autophagy. As a result, intracellular bacterial clearance is impaired causing a chronic inflammatory state [10]. A potential clinical application of GWAS involves quantifying a patient's risk for a specific disease based on relevant SNPs represented by a polygenic risk score (PRS) [11]. PRS's can eventually be combined with current risk prediction models that include clinical, biochemical, and lifestyle features. However, due to the majority of GWAS participants being from European ancestry, PRS has shown to not be generalizable to other groups.

Although GWAS have identified numerous genetic variants associated with complex traits, translating these findings into biological insights remains challenging [5]. Many significant GWAS variants are located in non-coding regions, making it difficult to identify the specific genes that drive trait associations [12]. A simple strategy to link GWAS-associated variants with a gene is by physical proximity, which typically selects the closest gene to the most significant SNP at each locus. However, SNPs can have distal effects on the expression of the target gene [13]. Genes can also have epistatic interactions, a secondary loci affecting a primary loci. The peripheral effect of the secondary loci are often too small to be picked up through GWAS alone [14]

Recent approaches combine multi-omics data from various cell types and tissues with GWAS to identify potential mechanisms of SNPs and the associated gene through molecular quantitative trait loci (moQTLs) [15] (**Figure 1**). An expression quantitative trait loci (eQTL) is a genetic region where variants are associated with the expression levels of a nearby or distant gene [16]. eQTL studies estimate that most causal genes linked to GWAS loci are not the proximal ones [17]. They have also allowed for insight on epistatic interactions. A large eQTL study was able to detect epistatic interactions, showing that many primary loci affecting gene expression were also modified by secondary eQTLs [14]. Splicing quantitative trait loci (sQTLs) are another major causal mechanism in GWAS loci, but are complex to interpret due to the production of unknown isoforms [18]. To overcome this, LeafCutter [19] maps variable splicing events, alleviating the need for transcript annotations. how this mogtls other mechanisms

Both GWAS and QTL studies have been penalized by their large multiple testing burden, causing the need to adopt a high level of significance. This results in GWAS being underpowered to detect all heritability explained by SNPs [5]. A strategy to combat this limitation is reducing the number of tests performed by focusing on gene co-expression modules or other prioritized genes [ttps://doi.org/10.1038/s41576-019-0127-1].

There are many strategies for leveraging GWAS data to identify gene sets related to a trait such as MAGMA [20] and FLAGS [21]. FLAGS uses predefined gene sets from prior knowledge while MAGMA is able to infer its own gene sets by first correlating genes with phenotypes and other genes. Both methods then statistically associate these modules with specific phenotypes. These tools are able to include trans-interactions, which TWAS misses, however by looking at the effect of SNPs on a gene set, specific SNP gene connections are lost [22].

From GWAS to gene: transcriptome-wide association studies (TWAS)

Transcriptome-wide association studies (TWAS) address this gap by integrating GWAS data with gene expression data (**Figure 1**), typically derived from eQTL studies, to prioritize genes whose expression across different tissues is influenced by GWAS variants [23,24]. By leveraging predicted gene expression levels, TWAS provides a mechanistic link between genetic variants and traits, allowing researchers to move beyond associations with individual SNPs to identify putatively causal genes [24]. Since TWAS models the genetic regulation of gene expression, the approach allows researchers to

impute expression levels in GWAS cohorts where expression data may not be available. A key advantage of TWAS over GWAS lies in its ability to increase interpretability by providing a gene-trait association: TWAS connects trait-associated SNPs (which are mostly non-coding) to genes, which are biologically functional units.

Several TWAS approaches have been introduced [25], including Predixcan [26], FUSION [27] and TIGAR [28]. However, all of them implement a similar framework that consists in three steps: 1) model training, 2) gene expression imputation, and 3) gene-trait association. For example, during 1), PrediXcan, builds one expression prediction model per gene and tissue using penalized linear regression with ElasticNet to model sparse genetic architectures. These models contain weights for each SNP used as predictor for a gene expression in a tissue. Given genotype data in a cohort without measured gene expression, during 2), the SNPs weights from the models can be used to impute tissue-specific gene expression for individuals. During 3), a gene-tissue-trait association is computed by correlating the tissue-specific imputed gene expression with the trait of interest. Most methods (such as Summary-PrediXcan [29] or S-PrediXcan), however, offer a shortcut by computing a genetissue-trait association directly from GWAS summary statistics without the need of individual-level data. This process, however, requires the user to select a tissue of interest, which might not be straightforward [24]. To address this limitation, approaches such as MultiXcan [30] or UTMOST [31] combine information across tissues to generate a gene-trait association. These multi-tissue approaches are generally more powerful than single-tissue ones, although they do not provide a direction of effect (i.e., whether a higher or lower predicted expression is associated with a higher or lower disease risk).

Going beyond TWAS

The flexibility of this 3-step framework can also be used to test whether other molecular phenotypes might mediate the association between GWAS variants and a trait of interest. In addition to integrating eQTLs, the framework has been implemented also with splicing QTLs (sQTLs), protein QTLs (pQTLs), histone acetylation QTLs (haQTLs) and methylation QTLs (mQTLs).

Mapping protein function to disease: protein-wide association studies (PWAS)

PWAS complements GWAS and TWAS by aggregating genetic variations in protein-coding regions to assess their combined impact on protein function and phenotypes (**Figure 1**) [32]. While GWAS focuses on individual variant associations, PWAS emphasizes the broader functional consequences of coding variants, providing a more comprehensive view of potential links to disease. By aggregating multiple variants of a gene, PWAS reduces the multiple testing burden and reveals complex inheritance patterns, such as dominant and recessive traits. PWAS shares a methodology with GWAS, using genotype, phenotype, and covariate data, but extends this approach by evaluating the cumulative impact of multiple variants on protein function.

To evaluate how genetic variants—such as missense, nonsense, and frameshift mutations—affect protein function, PWAS uses the FIRM machine learning model to assign impairment scores [32]. Genes are then evaluated for dominant and recessive inheritance patterns, followed by statistical tests to determine significant associations between gene-level scores and phenotypes. For example, using PWAS with the UK Biobank confirmed the association of *MUTYH* with colorectal cancer, even though no individual variant reached genome-wide significance in GWAS [32]. This demonstrates PWAS's ability to uncover functional associations missed by other methods. Similarly, PWAS identified associations in genes like *HLA-DQA2* and *PSMB9* in asthma, highlighting its utility in exploring the genetic components of complex diseases. [33]. However, PWAS relies heavily on high-quality proteomic data and may overlook associations driven by non-coding variants. It is also

computationally intensive, and future efforts may focus on incorporating rare variants and more diverse data.

Enhancing genetic studies with epigenetic data: Epigenome-wide association studies (EWAS)

Epigenome-wide association studies (EWAS) offer another valuable approach to understanding complex traits, specifically by exploring single-variant and single-gene relationships within the epigenetic context. Unlike GWAS, which primarily investigate genetic variants, EWAS focuses on epigenetic markers such as DNA methylation and histone modifications to uncover their roles in gene regulation and disease etiology [34]. Within the EWAS framework, methylome-wide association studies (MWAS) (Figure 1) and histone acetylome-wide association studies (HAWAS) (Figure 1), based on haQTLs and mQLTs respectability, represent specialized methodologies targeting specific types of epigenetic modifications. MWAS concentrates on DNA methylation patterns, enabling the identification of methylation changes at individual loci associated with particular traits or diseases. For instance, a MWAS conducted by Shen et al. provided evidence for a causal relationship between DNAm and depression, suggesting that epigenetic modifications may mediate the genetic risk for depression, exemplifying the effectiveness of EWAS in uncovering the biological mechanisms that link genetic predisposition to psychiatric disorders [35]. HAWAS focuses on histone acetylation modifications, which are crucial for regulating chromatin structure and gene expression. Sun et al. conducted a HAWAS on autism spectrum disorder (ASD) and identified thousands of differential acetylation loci in the brains of individuals with ASD, indicating that histone acetylation plays a significant role in ASD pathogenesis [36].

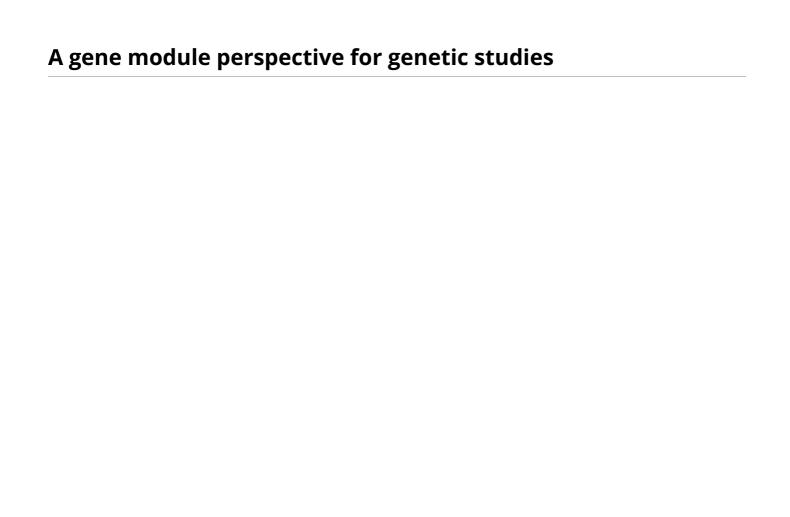
The tissue-specific nature of epigenetic modifications, alongside the capacity to capture cellular plasticity and environmental influences, enhances our insight into the effects of genetic variants across distinct tissues, temporal cellular states, and gene-environment interactions. This understanding contributes to a more detailed knowledge of the molecular mechanisms and regulatory dynamics underlying complex diseases, thereby supplementing the findings from GWAS [37,38].

Single-gene approaches are not enough

Despite the advancements facilitated by single-variant and single-gene methodologies, a common thread persists: the focus remains on one gene at a time. The underlying expectation is that identifying a gene linked to a trait will directly unveil the biological mechanisms driving disease processes. While this has been successful in certain monogenic disorders, complex traits often involve intricate interactions among multiple genes and environmental factors. Consequently, the one-gene-at-a-time paradigm may oversimplify the multifaceted nature of these traits. In light of these considerations, it becomes imperative to reevaluate our approaches to dissecting the genetic architecture of complex traits. This necessitates a shift towards methodologies that can capture the polygenic and network-based interactions inherent in complex diseases.

From single genes to gene networks: the omnigenic model for complex traits

From gene networks to machine learning derived gene modules: hands-on strategies for inferring gene-gene interactions



Future perspectives

Conclusions

References

1. Polygenic background modifies penetrance of monogenic variants for tier 1 genomic

Akl C Fahed, Minxian Wang, Julian R Homburger, Aniruddh P Patel, Alexander G Bick, Cynthia L Neben, Carmen Lai, Deanna Brockman, Anthony Philippakis, Patrick T Ellinor, ... Amit V Khera *Nature Communications* (2020-08-20) https://doi.org/gg9hqv

DOI: <u>10.1038/s41467-020-17374-3</u> · PMID: <u>32820175</u> · PMCID: <u>PMC7441381</u>

2. Genome-wide association studies

Emil Uffelmann, Qin Qin Huang, Nchangwi Syntia Munung, Jantina de Vries, Yukinori Okada, Alicia R Martin, Hilary C Martin, Tuuli Lappalainen, Danielle Posthuma *Nature Reviews Methods Primers* (2021-08-26) https://doi.org/gmk7p2

DOI: 10.1038/s43586-021-00056-9

3. **Open problems in human trait genetics**

Nadav Brandes, Omer Weissbrod, Michal Linial *Genome Biology* (2022-06-20) https://doi.org/gqc962

DOI: 10.1186/s13059-022-02697-9 · PMID: 35725481 · PMCID: PMC9208223

4. The NHGRI-EBI GWAS Catalog: knowledgebase and deposition resource

Elliot Sollis, Abayomi Mosaku, Ala Abid, Annalisa Buniello, Maria Cerezo, Laurent Gil, Tudor Groza, Osman Güneş, Peggy Hall, James Hayhurst, ... Laura W Harris *Nucleic Acids Research* (2022-11-09) https://doi.org/gq73r5

DOI: <u>10.1093/nar/gkac1010</u> · PMID: <u>36350656</u> · PMCID: <u>PMC9825413</u>

5. Benefits and limitations of genome-wide association studies

Vivian Tam, Nikunj Patel, Michelle Turcotte, Yohan Bossé, Guillaume Paré, David Meyre *Nature Reviews Genetics* (2019-05-08) https://doi.org/ggcxxb
DOI: 10.1038/s41576-019-0127-1 · PMID: 31068683

6. 10 Years of GWAS Discovery: Biology, Function, and Translation

Peter M Visscher, Naomi R Wray, Qian Zhang, Pamela Sklar, Mark I McCarthy, Matthew A Brown, Jian Yang

The American Journal of Human Genetics (2017-07) https://doi.org/gcsmnm
DOI: 10.1016/j.ajhg.2017.06.005 • PMID: 28686856 • PMCID: PMC5501872

7. Genome-wide analysis of insomnia in 1,331,010 individuals identifies new risk loci and functional pathways

, Philip R Jansen, Kyoko Watanabe, Sven Stringer, Nathan Skene, Julien Bryois, Anke R Hammerschlag, Christiaan A de Leeuw, Jeroen S Benjamins, Ana B Muñoz-Manchado, ... Danielle Posthuma

Nature Genetics (2019-02-25) https://doi.org/gfv4p2
DOI: 10.1038/s41588-018-0333-3 · PMID: 30804565

8. Genome-wide association analysis of insomnia complaints identifies risk genes and genetic overlap with psychiatric and metabolic traits

Anke R Hammerschlag, Sven Stringer, Christiaan A de Leeuw, Suzanne Sniekers, Erdogan Taskesen, Kyoko Watanabe, Tessa F Blanken, Kim Dekker, Bart HW te Lindert, Rick Wassing, ... Danielle Posthuma

Nature Genetics (2017-06-12) https://doi.org/gbhzs5

DOI: 10.1038/ng.3888 · PMID: 28604731 · PMCID: PMC5600256

9. A genome-wide association scan of nonsynonymous SNPs identifies a susceptibility variant for Crohn disease in ATG16L1

Jochen Hampe, Andre Franke, Philip Rosenstiel, Andreas Till, Markus Teuber, Klaus Huse, Mario Albrecht, Gabriele Mayr, Francisco M De La Vega, Jason Briggs, ... Stefan Schreiber *Nature Genetics* (2006-12-31) https://doi.org/b9fr2w

DOI: <u>10.1038/ng1954</u> · PMID: <u>17200669</u>

10. A Crohn's disease variant in Atg16l1 enhances its degradation by caspase 3

Aditya Murthy, Yun Li, Ivan Peng, Mike Reichelt, Anand Kumar Katakam, Rajkumar Noubade, Merone Roose-Girma, Jason DeVoss, Lauri Diehl, Robert R Graham, Menno van Lookeren Campagne

Nature (2014-02) https://doi.org/f5r7wh

DOI: <u>10.1038/nature13044</u> · PMID: <u>24553140</u>

11. Polygenic risk scores: from research tools to clinical instruments

Cathryn M Lewis, Evangelos Vassos

Genome Medicine (2020-05-18) https://doi.org/ghdnpx

DOI: <u>10.1186/s13073-020-00742-5</u> · PMID: <u>32423490</u> · PMCID: <u>PMC7236300</u>

12. Demystifying non-coding GWAS variants: an overview of computational tools and methods

Marijn Schipper, Danielle Posthuma

Human Molecular Genetics (2022-08-16) https://doi.org/grqdp7

DOI: 10.1093/hmg/ddac198 · PMID: 35972862 · PMCID: PMC9585674

13. The GTEx Consortium atlas of genetic regulatory effects across human tissues

, François Aguet, Shankara Anand, Kristin G Ardlie, Stacey Gabriel, Gad A Getz, Aaron Graubert, Kane Hadley, Robert E Handsaker, Katherine H Huang, ... Simona Volpi

Science (2020-09-11) https://doi.org/ghbnhr

DOI: 10.1126/science.aaz1776 · PMID: 32913098 · PMCID: PMC7737656

14. Molecular mechanisms of epistasis within and between genes

Ben Lehner

Trends in Genetics (2011-08) https://doi.org/fk3d5b

DOI: 10.1016/j.tig.2011.05.007 · PMID: 21684621

15. Strengthening Causal Inference for Complex Disease Using Molecular Quantitative Trait

Sonja Neumeyer, Gibran Hemani, Eleftheria Zeggini

Trends in Molecular Medicine (2020-02) https://doi.org/ggmq94

DOI: <u>10.1016/j.molmed.2019.10.004</u> · PMID: <u>31718940</u>

16. eQTL studies: from bulk tissues to single cells

Jingfei Zhang, Hongyu Zhao

Journal of Genetics and Genomics (2023-12) https://doi.org/g7n6d9

DOI: 10.1016/j.jgg.2023.05.003 · PMID: 37207929 · PMCID: PMC10656365

17. The omnigenic model and polygenic prediction of complex traits

lain Mathieson

The American Journal of Human Genetics (2021-09) https://doi.org/gmv9s5

DOI: <u>10.1016/j.ajhg.2021.07.003</u> · PMID: <u>34331855</u> · PMCID: <u>PMC8456163</u>

18. Splicing QTL analysis focusing on coding sequences reveals mechanisms for disease susceptibility loci

Kensuke Yamaguchi, Kazuyoshi Ishigaki, Akari Suzuki, Yumi Tsuchida, Haruka Tsuchiya, Shuji Sumitomo, Yasuo Nagafuchi, Fuyuki Miya, Tatsuhiko Tsunoda, Hirofumi Shoda, ... Yuta Kochi *Nature Communications* (2022-08-24) https://doi.org/ggr76g

DOI: 10.1038/s41467-022-32358-1 · PMID: 36002455 · PMCID: PMC9402578

19. Annotation-free quantification of RNA splicing using LeafCutter

Yang I Li, David A Knowles, Jack Humphrey, Alvaro N Barbeira, Scott P Dickinson, Hae Kyung Im, Jonathan K Pritchard

Nature Genetics (2017-12-11) https://doi.org/gcwk77

DOI: <u>10.1038/s41588-017-0004-9</u> · PMID: <u>29229983</u> · PMCID: <u>PMC5742080</u>

20. MAGMA: Generalized Gene-Set Analysis of GWAS Data

Christiaan A de Leeuw, Joris M Mooij, Tom Heskes, Danielle Posthuma *PLOS Computational Biology* (2015-04-17) https://doi.org/gf92gp

DOI: <u>10.1371/journal.pcbi.1004219</u> · PMID: <u>25885710</u> · PMCID: <u>PMC4401657</u>

21. FLAGS: A Flexible and Adaptive Association Test for Gene Sets Using Summary Statistics

Jianfei Huang, Kai Wang, Peng Wei, Xiangtao Liu, Xiaoming Liu, Kai Tan, Eric Boerwinkle, James B Potash, Shizhong Han

Genetics (2016-01-14) https://doi.org/f8db5f

DOI: <u>10.1534/genetics.115.185009</u> · PMID: <u>26773050</u> · PMCID: <u>PMC4788129</u>

22. A transcriptome-wide association study implicates specific pre- and post-synaptic abnormalities in schizophrenia

Lynsey S Hall, Christopher W Medway, Oliver Pain, Antonio F Pardiñas, Elliott G Rees, Valentina Escott-Price, Andrew Pocklington, Nicholas J Bray, Peter A Holmans, James TR Walters, ... Michael C O'Donovan

Human Molecular Genetics (2019-11-06) https://doi.org/gst8k8

DOI: <u>10.1093/hmg/ddz253</u> · PMID: <u>31691811</u> · PMCID: <u>PMC7416679</u>

23. From GWAS to Gene: Transcriptome-Wide Association Studies and Other Methods to Functionally Understand GWAS Discoveries

Binglan Li, Marylyn D Ritchie

Frontiers in Genetics (2021-09-30) https://doi.org/g7n6kb

DOI: 10.3389/fgene.2021.713230 · PMID: 34659337 · PMCID: PMC8515949

24. Opportunities and challenges for transcriptome-wide association studies

Michael Wainberg, Nasa Sinnott-Armstrong, Nicholas Mancuso, Alvaro N Barbeira, David A Knowles, David Golan, Raili Ermel, Arno Ruusalepp, Thomas Quertermous, Ke Hao, ... Anshul Kundaje

Nature Genetics (2019-03-29) https://doi.org/gf3hmr

DOI: <u>10.1038/s41588-019-0385-z</u> · PMID: <u>30926968</u> · PMCID: <u>PMC6777347</u>

25. Transcriptome-wide association studies: recent advances in methods, applications and available databases

Jialin Mai, Mingming Lu, Qianwen Gao, Jingyao Zeng, Jingfa Xiao *Communications Biology* (2023-09-01) https://doi.org/gs9f8r

DOI: 10.1038/s42003-023-05279-y · PMID: 37658226 · PMCID: PMC10474133

26. A gene-based association method for mapping traits using reference transcriptome data

Eric R Gamazon, Heather E Wheeler, Kaanan P Shah, Sahar V Mozaffari, Keston Aquino-Michaels, Robert J Carroll, Anne E Eyler, Joshua C Denny, Dan L Nicolae, ... Hae Kyung Im *Nature Genetics* (2015-08-10) https://doi.org/f7p9zv

DOI: 10.1038/ng.3367 · PMID: 26258848 · PMCID: PMC4552594

27. Integrative approaches for large-scale transcriptome-wide association studies

Alexander Gusev, Arthur Ko, Huwenbo Shi, Gaurav Bhatia, Wonil Chung, Brenda WJH Penninx, Rick Jansen, Eco JC de Geus, Dorret I Boomsma, Fred A Wright, ... Bogdan Pasaniuc *Nature Genetics* (2016-02-08) https://doi.org/f3vf4p

DOI: <u>10.1038/ng.3506</u> · PMID: <u>26854917</u> · PMCID: <u>PMC4767558</u>

28. TIGAR: An Improved Bayesian Tool for Transcriptomic Data Imputation Enhances Gene Mapping of Complex Traits

Sini Nagpal, Xiaoran Meng, Michael P Epstein, Lam C Tsoi, Matthew Patrick, Greg Gibson, Philip L De Jager, David A Bennett, Aliza P Wingo, Thomas S Wingo, Jingjing Yang *The American Journal of Human Genetics* (2019-08) https://doi.org/g7n6dv

DOI: 10.1016/j.ajhg.2019.05.018 · PMID: 31230719 · PMCID: PMC6698804

29. Exploring the phenotypic consequences of tissue specific gene expression variation inferred from GWAS summary statistics

Alvaro N Barbeira, Scott P Dickinson, Rodrigo Bonazzola, Jiamao Zheng, Heather E Wheeler, Jason M Torres, Eric S Torstenson, Kaanan P Shah, Tzintzuni Garcia, Todd L Edwards, ... Hae Kyung Im

Nature Communications (2018-05-08) https://doi.org/gdjvp5

DOI: 10.1038/s41467-018-03621-1 · PMID: 29739930 · PMCID: PMC5940825

30. Integrating predicted transcriptome from multiple tissues improves association detection

Alvaro N Barbeira, Milton Pividori, Jiamao Zheng, Heather E Wheeler, Dan L Nicolae, Hae Kyung Im

PLOS Genetics (2019-01-22) https://doi.org/ghs8vx

DOI: <u>10.1371/journal.pgen.1007889</u> · PMID: <u>30668570</u> · PMCID: <u>PMC6358100</u>

31. A statistical framework for cross-tissue transcriptome-wide association analysis

, Yiming Hu, Mo Li, Qiongshi Lu, Haoyi Weng, Jiawei Wang, Seyedeh M Zekavat, Zhaolong Yu, Boyang Li, Jianlei Gu, ... Hongyu Zhao

Nature Genetics (2019-02-25) https://doi.org/gijt5f

DOI: <u>10.1038/s41588-019-0345-7</u> · PMID: <u>30804563</u> · PMCID: <u>PMC6788740</u>

32. PWAS: proteome-wide association study—linking genes and phenotypes by functional variation in proteins

Nadav Brandes, Nathan Linial, Michal Linial

Genome Biology (2020-07-14) https://doi.org/gnvtr9

DOI: 10.1186/s13059-020-02089-x · PMID: 32665031 · PMCID: PMC7386203

33. PWAS Hub: Exploring Gene-Based Associations of Common Complex Diseases

Guy Kelman, Roei Zucker, Nadav Brandes, Michal Linial

Cold Spring Harbor Laboratory (2024-01-22) https://doi.org/g7n6hx

DOI: <u>10.1101/2024.01.20.23300645</u>

34. Epigenome-wide association studies: current knowledge, strategies and recommendations

Maria Pia Campagna, Alexandre Xavier, Jeannette Lechner-Scott, Vicky Maltby, Rodney J Scott, Helmut Butzkueven, Vilija G Jokubaitis, Rodney A Lea

Clinical Epigenetics (2021-12) https://doi.org/gn3kds

DOI: <u>10.1186/s13148-021-01200-8</u> · PMID: <u>34863305</u> · PMCID: <u>PMC8645110</u>

35. DNA methylome-wide association study of genetic risk for depression implicates antigen processing and immune responses

Xueyi Shen, Doretta Caramaschi, Mark J Adams, Rosie M Walker, Josine L Min, Alex Kwong, Gibran Hemani, Miruna C Barbu, Heather C Whalley, ... Andrew M McIntosh *Genome Medicine* (2022-03-31) https://doi.org/g7n6jn

DOI: <u>10.1186/s13073-022-01039-5</u> · PMID: <u>35354486</u> · PMCID: <u>PMC8969265</u>

36. Histone Acetylome-wide Association Study of Autism Spectrum Disorder

Wenjie Sun, Jeremie Poschmann, Ricardo Cruz-Herrera del Rosario, Neelroop N Parikshak, Hajira Shreen Hajan, Vibhor Kumar, Ramalakshmi Ramasamy, TGrant Belgard, Bavani Elanggovan, Chloe Chung Yi Wong, ... Shyam Prabhakar

Cell (2016-11) https://doi.org/f9dpzf

DOI: <u>10.1016/j.cell.2016.10.031</u> · PMID: <u>27863250</u>

37. Histone Variants: Guardians of Genome Integrity

Juliette Ferrand, Beatrice Rondinelli, Sophie E Polo *Cells* (2020-11-05) https://doi.org/g6vwfw

DOI: <u>10.3390/cells9112424</u> · PMID: <u>33167489</u> · PMCID: <u>PMC7694513</u>

38. Epigenetics: The DNA Methylation Profile of Tissue-Dependent and Differentially Methylated Regions in Cells

J Ohgane, S Yagi, K Shiota

Placenta (2008-03) https://doi.org/dfph4p

DOI: <u>10.1016/j.placenta.2007.09.011</u> · PMID: <u>18031808</u>