ECG classification using deep neural networks: Investigating architecture optimization and transfer learning

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Abstract — Electrocardiogram (ECG) interpretation is an important medical diagnostic tool for diagnosing conditions of the heart. The use of neural networks aiming to assist cardiologists in the interpretation of ECG signals show great potential and could lead to improved patient outcomes. Here, I compared and developed several neural network architectures for classifying ECG signals from widely available datasets. Additionally, I compared modelling results when using augmented data and evaluated how well the models trained on one dataset classified data from another dataset. I found that the presented neural network architectures accurately classified pathological ECG signals, comparable to that of several published articles. I also found that data augmentation diminished overall performance, suggesting the approach taken was too simple. Finally, I found that transfer learning is possible but, classification performance was poor on pathological ECG.

Keywords —ECG, deep neural networks, architecture optimization, transfer learning, classification problems, error matrix

1.0 Introduction

An electrocardiogram (ECG) is a measurement of the electrical activity of the heart muscle. Each time the heart pumps blood it produces a characteristic time series waveform with distinct and identifiable features (right, Figure 1). In medicine, the ECG signal is used as a diagnostic tool to determine if the heart is functioning as expected. When someone has a pathology, such as heart valve leaks or tears, there can be subtle but detectable changes in the ECG signal (left, Figure 1). The ability to detect the presence of a pathology early could lead to quick diagnosis and improved outcomes for patients, not to mention reductions in long term healthcare costs [1]–[4].

Computer-aided interpretation of ECG signals has been an increasingly important tool in the clinical workflow, with first attempts dating back to the late 1950s [3]. Although sophisticated computer algorithms have shown promise, high rates of misdiagnosis are common (~50%). The presence of noise, variability in wave morphology between patients, and the mixture of both subjective and objective characteristics makes ECG classification a difficult problem, where even experienced cardiologists can disagree on prognosis [5]. The use of deep neural networks has led to major advances in image classification, speech recognition, and many medical diagnostics [1], [6], [7]. The ability of deep neural networks to recognize patterns and learn features of time varying signals without extensive preprocessing makes them particularly well suited for ECG classification.

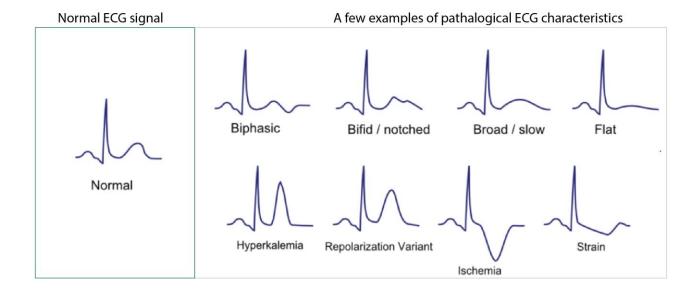


Figure 1: Example characteristics of normal and pathological ECG signals.

There are many possible pathologies that can be observed in ECG waveforms. It is common for researchers to study only certain more common pathologies, or group pathologies into categories due to limitations in the amount of data on pathological ECG signals [1], [4], [8]. In general, many research groups have shown great success with ECG classification using neural networks, achieving low misclassification errors, and accuracy's nearing 99% [2], [7], [9]–[13]. Although the results are profound and have led to startup companies and major advances, the caveats are small representative test sets, with limited patients, and limited variations in the ECG pathologies themselves. This increases the chances of missing abnormalities that may exist warranting expert cardiologist option (who can also miss these suitable features). Recently, a monumental effort led by Andrew Ng's group at Stanford university, proposed a neural network architecture for evaluating up to 12 different pathologies (more then has ever been proposed before) [3], [7]. This effort was only possible due to the extensive amount of data the group was able to collect (over 50,000 patients with all ECG annotations performed by cardiologists) which made it possible to train and evaluate their model.

This project here aims to use deep neural networks to classify normal and pathological ECG waveforms from publically available datasets. Specifically, there are several objectives. The first objective is build, develop, and train a convolutional neural network using a previously established architecture and compare its ability to classify normal and 4 pathological ECG signals to the ability of two author developed architectures. The second objective is characterize the performance variations when a simple data augmentation is performed on the dataset. Finally, the third objective is to quantify the performance of the models established in aim 1 and 2 on a novel ECG dataset via transfer learning.

2.0 Methods

2.1 Data Curation

ECG data was taken from the MIT-BIH arrhythmia (48 subjects, f = 360 Hz) and the PTB Diagnostic ECG (290 subjects, f = 1000 Hz) datasets as made available through Kaggle courtesy of [1]. Both of these ECG datasets have been labelled by trained cardiologists. The dataset has some preprocessing steps that were taken which modify it from its raw time-series form. In brief, the continuous ECG voltage signal was divided into 10s windows, voltage normalized to be between 0-1, peaks were identified using the first derivative, and the median time interval between peaks was determined ($T_{interval}$). Using the $T_{interval}$ determined for each 10 second window, the signal was then divided up such that each frame starts from a peak and continues for $1.2*T_{interval}$. Lastly, to ensure the divisions of each new frame have a fixed length, each frame is padded with zeros to make its length equal to an array of 187 in length. Each interval is then given the corresponding label which the cardiologist has previously provided. This label is added to each array as the 188^{th} value. These labels range are either normal or pathological as expressed in Table 1. This type of beat extraction does not filter the signal or make any assumption about the morphology, which has and been shown as an effective method for ECG processing. A further and more detailed overview of these preprocessing steps is provided elsewhere [1].

Table 1: The two datasets explored in this project MIT-BIH and PTB Diagnostic

| MIT-BIH | MIT-BIH ECG | | | | | | | | | |
|-----------------------------------|-------------|-----------|------------|------------------|-------|-----------|------------|--|--|--|
| ECG Class | Label | # samples | % of Total | ECG Class | Label | # samples | % of Total | | | |
| Normal | 0 | 90589 | 82.8% | Normal | 0 | 10506 | 72.2 | | | |
| Fusion of Paced and Normal | 1 | 2779 | 2.5% | Abnormal | 1 | 4046 | 27.8 | | | |
| Premature Ventricular Contraction | 2 | 7236 | 6.6% | | | | | | | |
| Atrial Premature | 3 | 803 | 0.7% | | | | | | | |
| Fusion of Ventricular and Normal | 4 | 8039 | 7.3% | | | | | | | |

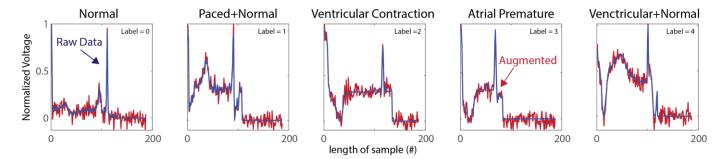
2.2 Data Augmentation

The number of representative samples for each class shows a large class imbalance (Table 1), where the majority of all data is of normal non-pathological ECG. To address this class imbalance I decided to perform a data augmentation on the minority classes [14]. It is generally suggested that augmenting just the training data increases reliability and generalization of the model [13]. With this in mind, the sample data was first split into 80% training, 20% testing. A StratifedShuffleSplit approach was taken such that the data was split equally amongst the classes. Next, the training data was further split such that 16% went into a separate validation set. The validation and testing sets were then set aside well a very simple data augmentation was performed on the training set for the MIT-BIH dataset (the main dataset explored in the bulk of this project). The majority class (normal ECG) was first reduced down to 10,000 samples and the minority classes were resampled such that each class now contained the same amount of observations. Random Gaussian noise was injected to each sample of the resampled data to further increase generalizability. The amplitude of this noise was small (0.03-0.05) to help ensure each label kept its class (it is possible though that this could change the class, this is why a small amount of noise was used). A figurative representation of the data is shown Figure 2 and Table 2 shows the summary of the data after splitting and augmenting.

Table 2: The split MIT-BIH dataset showing the number of samples in each class as well the new formed augmented data set which down sampled the majority class and up sampled the minority.

| | | | MIT-BIH ECO | G Dataset | |
|---------------|--------------|------------------------|----------------|-----------|--------------------|
| | | Training Set 1 | | | Training Set 2 |
| | | Non-Augmented Training | Validation Set | Test Set | Augmented Training |
| | % samples | 64% | 16% | 20% | 64% |
| | Label number | # samples | # samples | # samples | # samples |
| Normal | 0 | 57976 | 14495 | 18118 | 10000 |
| | 1 | 1778 | 445 | 556 | 10000 |
| Pathological | 2 | 4631 | 1158 | 1447 | 10000 |
| ratilological | 3 | 514 | 128 | 161 | 10000 |
| | 4 | 5145 | 1286 | 1608 | 10000 |

Figure 2: Raw data (MIT-BIH) from each category and data with Gaussian noise (augmented data).



2.3 Network Architectures

All networks were trained using the following set parameters: batch size = 64, patience = 20, epochs = 500. Model weights were set to be balanced meaning that for the augmented data these weights were just 1 but for the unbalanced data, the weights were balanced following the default approach proposed in Keras using callback functions. Learning rate was programed to have an exponential decay such that it started at 0.001, had 10,000 decay steps at a decay rate of 0.75. The Adam optimizer was used with beta_1 = 0.9 and beta_2, 0.999 set. The optimizer optimized for *spare categorical cross entropy* which is the recommended loss for this type of data [1].

Model 1 is a convolutional neural network with 55,013 parameters based on the architecture proposed elsewhere [1]. Model 2 is a recurrent neural network (RNN) with long short term memory (LSTM) having 54,661 parameters. This model was developed following recommendations for building RNN's and was hand tuned by trial and error during pilot experiments to include dropout [12]. RNN's are well suited for time series data of this type and should work well in classifying the ECG signals [12], [15]. Model 3 is a deep neural network with variable layers that I developed. Since developing neural network architecture is not my expertise, I wanted to use numerical optimization to help develop a simple deep neural network. This process is further explained in section **2.4 Optimization**. The model that trained on the non-augmented data converged on an architecture having 65,669 parameters. The model that trained on the augmented data converged on an architecture having 75,333 parameters (specific models details are provided in the Appendix).

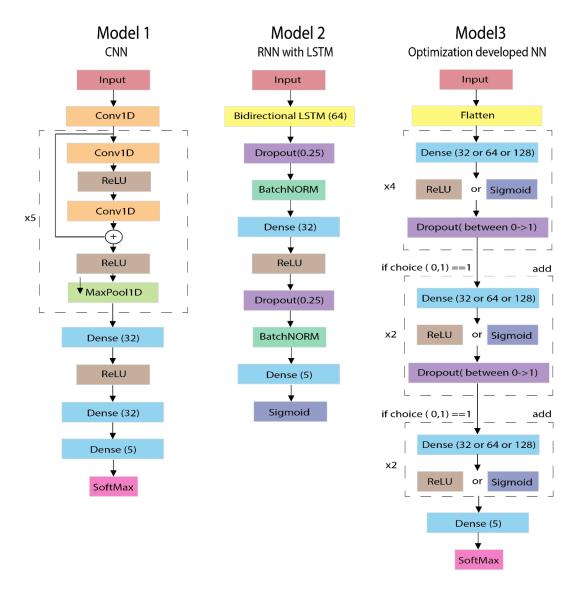


Figure 3: **Model 1**: Convolutional neural network based on [1], **Model 2**: recurrent neural network (RNN) with long short term memory (LSTM), and **Model 3**: an optimization developed neural network with variables parameter and number of layers developed using package *hyperopt* [16].

2.4 Optimization

An architecture optimization approach was implemented for Model 3 (Figure 3). A numerical optimization was setup to minimize negative test accuracy as depicted by equation 1. The optimization routine first evaluated a possible model architecture based on possible options, then model parameters were generated and optimized by minimizing the spare *categorical cross entropy*, same as for the other models. The optimization model was improved over 40 epochs of optimization (i.e. 40 different possible architectures were explored and optimized). The best combination of model architecture and model parameters resulting in the lowest objective function (Equation 1) was chosen for further evaluation (resulting model specifics shown in Appendix). This optimization routine was setup up using *hyperopt* [16].

Equation 1. Objective Function =
$$min(-(test_{set} Accuracy))$$

2.5 Transfer Learning

After training and evaluating the models on MIT-BIH dataset, I wanted to see how well the trained model would perform on a novel dataset namely the PTB Diagnostic, which has only two classes: normal and pathological (all pathological signals grouped into one) (Table 1). To do this, I used the best trained model from each architecture and added two dense layers and an output node such that the models now went from the 5 output nodes (5 classes in the MIT-BIH dataset) through two dense layers and outputted either 0 or 1 (normal or pathological) as depicted in Figure 4. This bottlenecking approach was based on methodology proposed in literature [1]. The two added dense layers where then trained on the new data while the other model weights were left untouched.

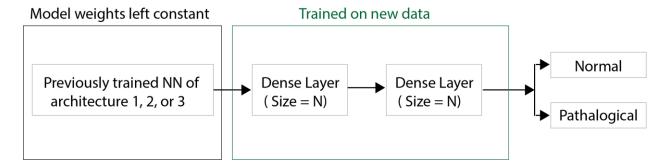


Figure 4: Approach used to add two additional dense layers to previously evaluated models. After some pilot experiments, the added dense layer was set to be a size of N = 32 as is also recommend elsewhere [1].

In addition to this approach, I also evaluated how training deep layers after adding the two dense layers would affect the transfer learning. To evaluate this in a systematic way I also evaluated the effects of training 5 layers deep and training 8 layers deep and present this in my results.

2.6 Metrics

The following metrics were utilized in this study. Recall which reflects the accurate positive identification of the ECG label as defined by equation 2.

Equation 2.
$$\mathbf{Recall} = \frac{\mathbf{True\ Positive}}{\mathbf{True\ Positive + False\ Negative}}$$

Precision which is the accurate positive identification—when the model predicted positive was it correct? This is defined by equation 3.

Equation 3. Precision =
$$\frac{\text{True Positive}}{\text{True Positive+False Positive}}$$

The F1- score is a metric for comparing the performance as a ratio of the mean of the recall and precision. The higher the F1-score the better the model did is the best way to look at it. F1-score is convenient to look at as it can be calculated for each label. In the context of our problem here this is a great metric to use to compare model performance as expressed in equation 4.

Equation 4. F1 Score
$$=\frac{2*Recall*Precision}{Recall*Precision}$$

The accuracy is the ratio between correctly predicted outcomes and the sum of all predictions. A relatively easy metric to understand for looking at global performance of the model as depicted by equation 5.

Equation 5. Accuracy
$$=\frac{\text{True Positive+True Negative}}{\text{True Positive+True Negative+False Positive+False Negative}}$$

Finally, the error matrix also known as a confusion matrix is a convenient table that shows the percentage of classified or misclassified labels in a matrix (it is the recall measure expressed for every label). Figure 5 provides an example of how this table will be presented in the results of this research. The diagonals of the matrix express the accurate identification of the correct label and the closer this accuracy is to 100% the better the model did. Any numbers shown off the diagonal can be thought of as unfavourable as this expresses misclassification of the model.

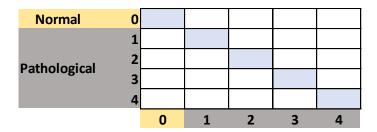


Figure 5: Example of an error matrix used for evaluating performance of a classification model. High accuracy scores across the diagonal indicates good classification.

3.0 Results

3.1 Network architecture influences performance

Network architecture has a noticeable effect on the F1-score of each model, as can be seen in Table 3. We can see that Model A has the highest overall F1-score when the data that the network is trained on is augmented (Model A, F1-Score = 0.954) but when the data the network is trained on is not augmented Model B performs best (Model B, F1-Score = 0.993). We can further see that the models all have high accuracies meaning that their ability to perform this classification task is good. The one caveat here is that the testing data is heavily skewed towards normal ECG signals. If the model correctly classifies the normal ECG it can still achieve a high accuracy without correctly classifying pathological ECG.

To better visualize the errors and misclassification occurring in the models, Figure 6 shows the error matrixes. Here we can see that all the models had a difficult time with the pathological label 1(fusion of paced and normal) as indicated by the higher misclassifications (10-35%) of this label. The models performed exceptionally well at classifying the normal ECG signal which was especially noticeable when the data was trained on the non-augmented training set (achieving almost 100% for all three architectures). The ability to well classify the larger normal set also explains why we see such large model accuracies. In general, all the architectures where good at classifying the data regardless of the training dataset howevr, Model 1 one performed noticeably

better than the other two models. Model 3 had misclassifications errors higher than other models when it came to classifying pathological ECG signals vs. normal.

Table 3: F1-scores for the 3 models trained on augmented and non-augmented datasets MIT BIT. Green shading indicates best performance. The closer the F1-score is to 1 the better the model performed at classifying.

| | F1-Score | | | |
|---------------|--|----------------|----------------|----------------|
| | Augmented Training | Model 1 | Model 2 | Model 3 |
| Normal | Normal | 0.974 | 0.960 | 0.966 |
| | Fusion of Paced and Normal | 0.680 | 0.588 | 0.679 |
| Dath alasiaal | Premature Ventricular Contraction | 0.934 | 0.891 | 0.847 |
| Pathological | Atrial Premature | 0.474 | 0.393 | 0.484 |
| | Fusion of Ventricular and Normal | 0.975 | 0.972 | 0.910 |
| | Weighted Average | 0.960 | 0.944 | 0.943 |
| | Macro Average | 0.807 | 0.761 | 0.777 |
| | | | | |
| | Non-Augmented Training | Model 1 | Model 2 | Model 3 |
| Normal | Normal | 0.992 | 0.997 | 0.989 |
| | Fusion of Paced and Normal | 0.843 | 0.925 | 0.770 |
| Dath alasted | Premature Ventricular Contraction | 0.961 | 0.984 | 0.936 |
| Pathological | | | | |
| Pathological | Atrial Premature | 0.799 | 0.895 | 0.757 |
| Patnological | Atrial Premature Fusion of Ventricular and Normal | 0.799 0.989 | 0.895 0.997 | 0.757 0.982 |
| Pathological | | | | |

Table 4: Global accuracy metrics for the 3 models trained on augmented and non-augmented datasets. Green shading indicates best performance. If the model correctly classifies the normal ECG it can still achieve a high accuracy without correctly classifying pathological ECG.

| Accuracy | Model 1 | Model 2 | Model 3 |
|------------------------|---------|---------|---------|
| Augmented Training | 95.4% | 93.2% | 97.9% |
| Non-Augmented Training | 98.5% | 99.3% | 94.0% |

| Training on Augmented Data | | | | | | | | | | | | | | | | | | |
|--------------------------------|--------------|---|-------------|-----|---------|-------------|-------|-----|-------|----------------------|--------|-----|-----|------|-------|-------|--------|-----|
| | | | | | | | ng on | Aug | mente | | | | | | | | | |
| | # of samples | | Model 1 CNN | | | Model 2 RNN | | | | Model 3 Optimization | | | | on | | | | |
| Normal | 18118 | 0 | 95% | 2% | 1% | 2% | 0% | | 93% | 3% | 1% | 2% | 0% | 96% | 1% | 1% | 1% | 1% |
| | 556 | 1 | 10% | 89% | 1% | 0% | 0% | | 10% | 88% | 2% | 0% | 0% | 29% | 68% | 3% | 0% | 0% |
| Pathological | 1447 | 2 | 1% | 0% | 96% | 2% | 0% | | 1% | 1% | 94% | 3% | 0% | 11% | 0% | 86% | 2% | 0% |
| ratifological | 161 | 3 | 2% | 1% | 6% | 92% | 0% | | 4% | 0% | 1% | 94% | 0% | 12% | 0% | 4% | 84% | 0% |
| | 1608 | 4 | 2% | 0% | 1% | 0% | 97% | | 1% | 0% | 0% | 0% | 99% | 4% | 0% | 2% | 0% | 94% |
| | | | | | | | | - | | | | | | | | | | |
| Training on Non-Augmented Data | | | | | | | | | | | | | | | | | | |
| | | | | Mod | del 1 (| CNN | | | - | Mod | el 2 R | NN | | Mo | del 3 | Optir | nizati | on |
| Normal | 18118 | 0 | 99% | 0% | 0% | 0% | 0% | | 100% | 0% | 0% | 0% | 0% | 100% | 0% | 0% | 0% | 0% |
| | 556 | 1 | 13% | 87% | 0% | 0% | 0% | | 11% | 88% | 1% | 0% | 0% | 35% | 64% | 1% | 0% | 0% |
| Pathological | 1447 | 2 | 3% | 1% | 94% | 1% | 0% | | 1% | 0% | 98% | 1% | 0% | 6% | 0% | 91% | 3% | 0% |
| ratifological | 161 | 3 | 12% | 2% | 6% | 80% | 0% | | 7% | 0% | 6% | 88% | 0% | 20% | 0% | 3% | 76% | 0% |
| | 1608 | 4 | 1% | 0% | 0% | 0% | 99% | | 0% | 0% | 0% | 0% | 99% | 3% | 0% | 0% | 0% | 97% |
| | | | 0 | 1 | 2 | 3 | 4 | | 0 | 1 | 2 | 3 | 4 | 0 | 1 | 2 | 3 | 4 |

Figure 6: Error matrixes for the 3 models explored showing the results for models trained on augmented and non-augmented training sets. Blue shading indicates diagonal squares (100% is perfect classification), red shaded color indicates noticeable misclassifications, grey shading indicates low error (<2%).

3.2 Data augmentation decreased performance

Data augmentation had a determinable effect on overall model performance as can be seen by looking at accuracy and loss performance for the three evaluated models shown in Figure 7. The model accuracy was lower and validation loss was higher for models that were trained on the augmented data set resulting in poorer F1-scores (Table 3), lower test accuracy (Table 4) and higher misclassification rates (Figure 6).

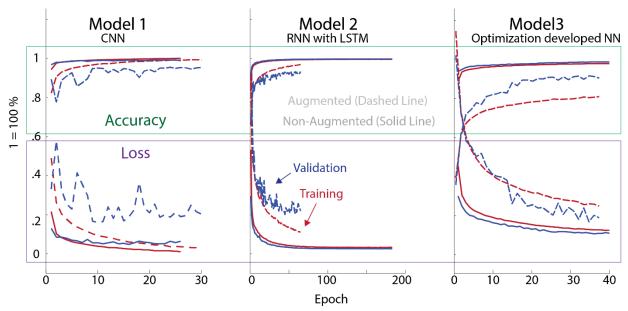


Figure 7: Accuracy (top section) and objective function loss (lower section) for validation and training sets using augmented (dashed lines) and non-augmented (solid line) training sets.

The augmented dataset required down sampling the majority class and up sampling the minority class. Perhaps the approach taken using *resample* from the Keras toolbox and adding Gaussian noise was too simple with too many similar cases for the network to train on. It is important to note that there were some improvements on classifying the pathological classes when the models trained on the augmented data (Figure 6) but in general the models performed more poorly. More sophisticated augmentation techniques appear to be beneficial for ECG data classification and should be explored in future work [11].

3.3 Transfer learning classification is strong on normal ECG but not on pathological

Figure 8 shows the F1-scores for the transfer learning results comparing: training the 2 added layers, training 5 layers deep, and training up to 8 layers deep. Applying the trained MIT-BIH models on a novel dataset resulted in a strong ability to classify the normal ECG signals but the ability to classify the pathological signal varied widely between models. Classifying the pathological data was particularly poor for the models trained on the augmented dataset. In terms of the F1-score, we can see in Figure 8 that model 3 had the best generalizability to this new dataset, resulting in the highest overall F1-score. It is interesting to note that model 3 had the poorest (although was still good at) classification performance for the MIT-BIH dataset.

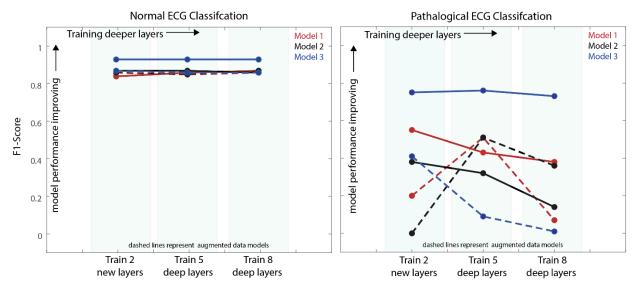


Figure 8: F1-scores for classification on PTB dataset after transfer learning from MIT_BIT trained models. PTB has two labels datasets either Normal (right) or Pathological (left). Results for model trained on augmented (dashed lines) and non-augmented (solid lines) training set are shown.

Looking further at the resulting error matrix just for the models trained on the 2 new added layers, we can see that model 3 shows a strong ability to correctly classifying the data, with Model 2 performing poorly (Figure 9). Training more layers did not appear to improve Model 3's performance and in most cases it deteriorated all models ability to classify the pathological data. In general though these results are encouraging and with further tuning improvements could further be made [1], [17].

| Training on Augmented Data | | | | | | | | | | | | |
|----------------------------|--------------------------|---|-------|------------|----------|-------------|------------|--------------|-----|--------|--|--|
| # | # of samples Model 1 CNN | | | | | Model | _ | Model 3 Opt. | | | | |
| Normal | 2102 | 0 | 98% | 2% | | 100% | 0% | | 93% | 7% | | |
| Pathological | 809 | 1 | 12% | 88% | | 100% | 0% | | 69% | 31% | | |
| | | | | | • | | <u>-</u> ' | • | | - | | |
| | | | Tra | ining on N | on-Augme | nted Data | | | | | | |
| | | | Model | 1 CNN | _ | Model 2 RNN | | | | 3 Opt. | | |
| Normal | 2102 | 0 | 82% | 18% | | 94% | 6% | | 92% | 8% | | |
| Pathological | 809 | 1 | 45% | 55% | | 73% | 27% | | 29% | 71% | | |
| | | | 0 | 1 | | 0 | 1 | • | 0 | 1 | | |

Figure 9: Error matrixes for the 3 models explored showing the transfer learning results for models trained on augmented and non-augmented training sets. Blue shading indicates diagonal squares, red shaded color indicates noticeable misclassifications, grey shading indicates low error (<2%). The PTB dataset has two classes normal and pathological. This requires the MIT-BIH trained models to bottle neck down to 2 outputs. Note: these error matrixes are only representative of results from training 2 new added dense layers.

3.4 Hyper parameters effect individual model performance

Although all of the results presented in this project used one set of hyper parameters as described in the methods, various other hyper parameter combinations were evaluated during the pilot testing of developing this systematic experiment. It warrants to present the effects of hyper parameter tuning to increase the transparency and further possibility of improving the presented results. Changes in parameters such as batch size, augmentation sample set or patience each resulted in

differences in the final outcomes, as presented by the error matrix of an arbitrary model architecture with changes made to these parameters in Figure 10. Although further tuning and exploring the best combinations of these parameters is possible, there were limitations in the hardware the network was running on and the time constraints possible to evaluate long duration computations (Anaconda using a single-core 3.6 GHz Dell OptiPlex 7020 computer with 16 Gb of RAM). I am acknowledging that it is possible that the results presented her could be further improved upon, but I feel that the conclusions and end outcomes would likely not vary drastically from what was presented here. Nevertheless, further work could lead to improvements in performance and should be explored.

| | Evaluation of hyper-paramter tuning on outcomes | | | | | | | | | | | | | | | | | | |
|--------------|---|---|------------------|-----|-----|-----|-----------------------|---|-----|-----|-----|---------------|-----|---|-----|-----|-----|-----|-----|
| | | | Batch size = 256 | | | | Augmentation = 20,000 | | | | | Patience = 10 | | | | | | | |
| Normal | 18118 | 0 | 82% | 14% | 1% | 2% | 1% | | 98% | 2% | 1% | 0% | 0% | | 93% | 5% | 1% | 0% | 0% |
| | 556 | 1 | 5% | 94% | 1% | 0% | 0% | | 9% | 90% | 1% | 0% | 0% | | 3% | 96% | 1% | 0% | 0% |
| 5 | 1447 | 2 | 2% | 3% | 91% | 4% | 1% | | 2% | 1% | 96% | 1% | 0% | | 1% | 1% | 97% | 1% | 0% |
| Pathological | 161 | 3 | 0% | 4% | 2% | 94% | 0% | | 9% | 1% | 7% | 83% | 0% | | 5% | 2% | 11% | 82% | 0% |
| | 1608 | 4 | 0% | 0% | 0% | 0% | 99% | | 1% | 0% | 0% | 0% | 99% | | 0% | 0% | 0% | 0% | 99% |
| | | | 0 | 1 | 2 | 3 | 4 | • | 0 | 1 | 2 | 3 | 4 | , | 0 | 1 | 2 | 3 | 4 |

Figure 10: Error matrixes for 1 example model using variations in hyper parameters. Results show sensitivity to user selected hyper parameters, further work is warranted.

3.5 Comparison of results to literature

The results presented in this project were compared to results from other sources. Specially, the accuracy and F1-score was compared as shown in Table 5. The performance metrics achieved in this work are strong and compare well against what is found in recent literature. Although the classification ability of the proposed work is certainly good, caution must be taken when making direct comparisons of this sort. The test set used in this project was unbalanced, small, and only had several pathological signals (for example [3][7] evaluated 12 classes on over 50,000 patients). Making comparison to other models which may achieve lower F1-score and lower accuracy for this reason is deceptive and most be taken in to account. For this specific problem misclassification of pathological ECG has larger consequences then misclassifying a healthy normal ECG. Although this was not specifically taken into account in this project it is important to consider when evaluating the models ability to classify ECG signals.

Table 4: Comparing the accuracy and F1-scores obtained in this project to that of literature

| Authors | Citation | accuracy % | Architecture | Authors | Citation | F1-score | F1-score (cardiologist) |
|--------------------------|----------|------------|------------------------|------------------------|-----------|-------------|----------------------------|
| 2019 Mousavi et al. | [13] | 99.92* | RNN+CNN | 2020 Steenkiste et al. | [17] | 0.80-0.98 | |
| 2017 Achaya et al. | [11] | 97.40 | CNN | 2019 Hannun et al. | [3][7] | 0.57-0.94** | 0.53-0.91** |
| 2016 Kiranyaz et al. | [6] | 99.10* | CNN 2D | 2019 Alfaras et al. | [10 | 0.78-0.98 | |
| 2010 Ye et al. | [8] | 99.91* | Support Vector Machine | | | | |
| | | | | | | | |
| Presented here 93.2-99.3 | | | Presented her | e | 0.39-0.99 | | |

^{*} more ECG classes explored then in this dataset and large sample size with augmentation

** Over 12 classes and dataset composed of over 50,000+ patients

4.0 Conclusions

In summary, the work presented here successfully met the objectives of the project. First, I was able to implement 3 neural network architectures for classifying ECG signals. All the proposed architectures had strong performance metrics and were able to classify the data to a high degree of accuracy. Second, I augmented the training set data and evaluated the performance effects. I found that the simple data augmentation approach led to an overall decrease in prediction performance. Lastly, I evaluated how well the models would perform when tested on a new data set. I found that the classification of normal ECG was strong but the classifying the pathological data proved to be more difficult. Further work is warranted in optimizing hyper parameters which would lead to improvements in the outcomes.

References

- [1] M. Kachuee, S. Fazeli, and M. Sarrafzadeh, "ECG Heartbeat Classification: A Deep Transferable Representation," 2018 IEEE Int. Conf. Healthc. Inform. ICHI, pp. 443–444, Jun. 2018, doi: 10.1109/ICHI.2018.00092.
- [2] S. Kiranyaz, T. Ince, and M. Gabbouj, "Real-Time Patient-Specific ECG Classification by 1-D Convolutional Neural Networks," *IEEE Trans. Biomed. Eng.*, vol. 63, no. 3, pp. 664–675, Mar. 2016, doi: 10.1109/TBME.2015.2468589.
- [3] A. Y. Hannun *et al.*, "Cardiologist-level arrhythmia detection and classification in ambulatory electrocardiograms using a deep neural network," *Nat. Med.*, vol. 25, no. 1, pp. 65–69, Jan. 2019, doi: 10.1038/s41591-018-0268-3.
- [4] S. M. Jadhav, S. L. Nalbalwar, and A. A. Ghatol, "ECG arrhythmia classification using modular neural network model," in *2010 IEEE EMBS Conference on Biomedical Engineering and Sciences (IECBES)*, Nov. 2010, pp. 62–66, doi: 10.1109/IECBES.2010.5742200.
- [5] J. Schläpfer and H. J. Wellens, "Computer-Interpreted Electrocardiograms," *J. Am. Coll. Cardiol.*, vol. 70, no. 9, pp. 1183–1192, Aug. 2017, doi: 10.1016/j.jacc.2017.07.723.
- [6] S. Kiranyaz, T. Ince, R. Hamila, and M. Gabbouj, "Convolutional Neural Networks for patient-specific ECG classification," in 2015 37th Annual International Conference of the IEEE Engineering in Medicine and Biology Society (EMBC), Milan, Aug. 2015, pp. 2608–2611, doi: 10.1109/EMBC.2015.7318926.
- [7] P. Rajpurkar, A. Y. Hannun, M. Haghpanahi, C. Bourn, and A. Y. Ng, "Cardiologist-Level Arrhythmia Detection with Convolutional Neural Networks," *ArXiv170701836 Cs*, Jul. 2017, Accessed: Mar. 18, 2020. [Online]. Available: http://arxiv.org/abs/1707.01836.
- [8] Can Ye, M. T. Coimbra, and B. V. K. Vijaya Kumar, "Arrhythmia detection and classification using morphological and dynamic features of ECG signals," in 2010 Annual International Conference of the IEEE Engineering in Medicine and Biology, Buenos Aires, Aug. 2010, pp. 1918–1921, doi: 10.1109/IEMBS.2010.5627645.
- [9] T. Li and M. Zhou, "ECG Classification Using Wavelet Packet Entropy and Random Forests," *Entropy*, vol. 18, no. 8, p. 285, Aug. 2016, doi: 10.3390/e18080285.
- [10] M. Alfaras, M. C. Soriano, and S. Ortín, "A Fast Machine Learning Model for ECG-Based Heartbeat Classification and Arrhythmia Detection," *Front. Phys.*, vol. 7, p. 103, Jul. 2019, doi: 10.3389/fphy.2019.00103.
- [11] U. R. Acharya *et al.*, "A deep convolutional neural network model to classify heartbeats," *Comput. Biol. Med.*, vol. 89, pp. 389–396, Oct. 2017, doi: 10.1016/j.compbiomed.2017.08.022.
- [12] S. Saadatnejad, M. Oveisi, and M. Hashemi, "LSTM-Based ECG Classification for Continuous Monitoring on Personal Wearable Devices," *IEEE J. Biomed. Health Inform.*, vol. 24, no. 2, pp. 515–523, Feb. 2020, doi: 10.1109/JBHI.2019.2911367.

- [13] S. Mousavi, F. Afghah, and U. R. Acharya, "Inter- and intra- patient ECG heartbeat classification for arrhythmia detection: a sequence to sequence deep learning approach," *ArXiv181207421 Phys. Q-Bio*, Mar. 2019, Accessed: Mar. 18, 2020. [Online]. Available: http://arxiv.org/abs/1812.07421.
- [14] "Resampling strategies for imbalanced datasets." https://kaggle.com/rafjaa/resampling-strategies-for-imbalanced-datasets (accessed Apr. 12, 2020).
- [15] M. Zihlmann, D. Perekrestenko, and M. Tschannen, "Convolutional Recurrent Neural Networks for Electrocardiogram Classification," in *Computing in Cardiology*, Sep. 2017, doi: 10.22489/CinC.2017.070-060.
- [16] J. Bergstra, D. Yamins, and D. D. Cox, "Making a Science of Model Search: Hyperparameter Optimization in Hundreds of Dimensions for Vision Architectures," p. 9.
- [17] G. Van Steenkiste, G. van Loon, and G. Crevecoeur, "Transfer Learning in ECG Classification from Human to Horse Using a Novel Parallel Neural Network Architecture," *Sci. Rep.*, vol. 10, no. 1, p. 186, Dec. 2020, doi: 10.1038/s41598-019-57025-2.

4.0 Appendix

Model 3: Trained on non-augmented data set

Options resulting from optimzation see Figure 3 for model

characteristics {'Activation': 0, 'Activation_1': 0, 'Activation_2': 0, 'Activation_3': 0, 'Activation_4': 0, 'Activation_5': 1, 'Activation_6': 1, 'Activation_7': 0, 'Activation_8': 0, 'Dense': 2, 'Dense_1': 2, 'Dense_2': 2, 'Dense_3': 1, 'Dense_4': 1, 'Dense_5': 1, 'Dense_6': 1, 'Dense_7': 0, 'Dense_8': 1, 'Dense_9': 3, 'Dropout': 0.3265904623792442, 'Dropout_1': 0.3431590875721917, 'Dropout_2': 0.25572580783477367, 'Dropout_3': 0.6975060560039738, 'Dropout_4': 0.3423233980953824, 'Dropout_5': 0.32317538374749766, 'a': 0, 'if': 0, 'if_1': 0, 'if_2': 0, 'if_3': 0}

Model: "sequential_103"

| Layer (type) Ou | tput Shape | Param # | |
|--------------------------|------------------|---------|--|
| flatten_107 (Flatten) | (None, 187) | 0 | |
| dense_677 (Dense) | (None, 128) | 24064 | |
| activation_636 (Activati | ion) (None, 128) | 0 | |
| dropout_456 (Dropout) | (None, 128) | 0 | |
| dense_678 (Dense) | (None, 128) | 16512 | |
| activation_637 (Activati | ion) (None, 128) | 0 | |
| dropout_457 (Dropout) | (None, 128) | 0 | |
| dense_679 (Dense) | (None, 128) | 16512 | |
| activation_638 (Activati | ion) (None, 128) | 0 | |
| dropout_458 (Dropout) | (None, 128) | 0 | |
| dense_680 (Dense) | (None, 64) | 8256 | |
| activation_639 (Activati | ion) (None, 64) | 0 | |
| dropout_459 (Dropout) | (None, 64) | 0 | |
| dense_681 (Dense) | (None, 5) | 325 | |

Total params: 65,669

Model 3: Trained on augmented data set

Options resulting from optimzation see Figure 3

| for model characteristics | Layer (type) | Output Shape | Param # | |
|-----------------------------------|----------------------|----------------------|---------|-----------|
| {'Activation': 0, | ========== | ========== | | ========= |
| 'Activation_1': 1, | flatten_135 (Flatter | n) (None, 187) | 0 | |
| 'Activation_2': 0, | | | | |
| 'Activation_3': 0, | dense_917 (Dense) | (None, 128) | 24064 | |
| 'Activation_4': 0, | | | | |
| 'Activation_5': 0, | activation_815 (Acti | ivation) (None, 128) | 0 | |
| 'Activation_6': 0, | | | | |
| 'Activation_7': 0, | dropout_582 (Dropo | out) (None, 128) | 0 | |
| 'Activation_8': 1, | | | | |
| 'Dense': 2, | dense_918 (Dense) | (None, 32) | 4128 | |
| 'Dense_1': 0, | | | | |
| 'Dense_2': 2, | activation_816 (Acti | ivation) (None, 32) | 0 | |
| 'Dense_3': 1, | | | | |
| 'Dense_4': 2, | dropout_583 (Dropo | out) (None, 32) | 0 | |
| 'Dense_5': 1, | | | | |
| 'Dense_6': 0, | dense 919 (Dense) | (None, 128) | 4224 | |
| 'Dense_7': 0, | | | | |
| 'Dense_8': 0, | activation_817 (Acti | ivation) (None, 128) | 0 | |
| 'Dense_9': 1, | | | | |
| 'Dropout': 0.4959326042904314, | dropout_584 (Dropo | out) (None, 128) | 0 | |
| 'Dropout_1': 0.5162300963476361, | | | | |
| 'Dropout_2': 0.4169915806152422, | dense_920 (Dense) | (None, 64) | 8256 | |
| 'Dropout_3': 0.25301754583882874, | | | | |
| 'Dropout_4': 0.2515810749300903, | activation_818 (Acti | ivation) (None, 64) | 0 | |
| 'Dropout_5': 0.516131118961639, | | | | |
| 'a': 1, | dropout_585 (Dropo | out) (None, 64) | 0 | |
| 'if': 0, | | | | |
| 'if_1': 0, | dense_921 (Dense) | (None, 32) | 2080 | |
| 'if_2': 1, | | | | |
| 'if_3': 0} | activation_819 (Acti | ivation) (None, 32) | 0 | |
| | dense_922 (Dense) | (None, 5) | 165 | |
| | ========== | | | |

Total params: 75,333

```
# Written by Pawel Kudzia
     import pandas as pd
     import numpy as np
     import tensorflow as tf
     import matplotlib.pyplot as plt
     from sklearn.utils import resample
     from sklearn.utils import class_weight
     from sklearn.model_selection import (TimeSeriesSplit, KFold, ShuffleSplit,
                            StratifiedKFold, GroupShuffleSplit,
                            GroupKFold, StratifiedShuffleSplit)
     from sklearn.metrics import classification_report, roc_auc_score, accuracy_score, precision_score, recall_score
     from tensorflow.keras.layers import Input, Conv1D, MaxPool1D, Activation, Add, Dense, Flatten
     from tensorflow.keras.models import Model,load_model
     from tensorflow.keras.optimizers import Adam
     from tensorflow.keras.utils import plot_model
18
     from tensorflow.keras.utils import to_categorical
     from tensorflow.keras.metrics import Precision
20
     from tensorflow.keras.layers import Input, Dropout, LSTM, Bidirectional, BatchNormalization, Dense, Activation
     from tensorflow.keras.models import Sequential, Model
23
24
     {\bf from\ tensor flow. keras. callbacks\ import\ Early Stopping, Learning Rate Scheduler,\ Model Checkpoint}
25
     from sklearn.metrics import roc_curve, auc,roc_auc_score
     from sklearn.metrics import confusion_matrix
      # CUSTOM Functions imported from folder
     from plot_Conf_Matrix import plot_Conf_Matrix
     from make_model_1 import make_model_1 # This is the biorix Architeture Model 1 in submitted report
     from make_model_2_RNN import make_model_2_RNN# this is a simple RNN model
     from hyperopt import Trials, STATUS_OK, tpe
36
     from hyperas import optim
     from hyperas.distributions import choice, uniform
     import warnings
41
42
     warnings.filterwarnings('ignore')
     warnings.simplefilter(action='ignore', category=FutureWarning)
45
     # %% Modelling Parameters and Variables CHANGE PARAMETERS HERE AS NEEDED
     saveData = 0
48
     epochs = 1
     batch = 64
     patience = 10
     verbose = 1
     ModelSaveName = 'Model1CNN UsingNotAugentedData.h5' # Change this to what you want to save the modoel as
53
     Model1 CNN =1
     \textbf{Model3\_OptimizationModel} = 0
      # %% Class labels
     classesMIT={0:"Normal",
           1 "Artial Premature"
          2: "Premature Ventricular Contraction".
           3:"Fusion of Centricular and Normal",
           4:"Fusion of paced and Normal"}
     classePTB={0:"Normal", 1:"Abnormal (MI)"}
     # %% Let define all our smaller nested functions here
66
68
     def plot_learning(history):
       plt.subplot(211)
70
71
        plt.plot(history.history['accuracy'])
        plt.legend(["accuracy"])
        plt.subplot(212)
73
74
75
        plt.plot(history.history['loss'])
        plt.plot(history.history['val_loss'], label = "val_loss")
        plt.legend(["loss", "val_loss"])
        plt.show()
77
78
     def outputMetrics(X_test,Y_Test,ModelX, batch) :
          X_{test} = X_{test.reshape(len(X_{test}), X_{test.shape[1],1)}
          X_test =X_test
       Y_Test=to_categorical(Y_Test)
       PredictionModel=ModelX.predict(X_test, batch_size=batch, verbose=1)
        TrueValue=np.argmax(Y_Test, axis=1)
        Prediction=np.argmax(PredictionModel,axis=1)
       print(classification_report(TrueValue, Prediction))
        return (Prediction, TrueValue)
     def roc_auc_score_multiclass(actual_class, pred_class, average = "macro"):
98
      #creating a set of all the unique classes using the actual class list
100
      unique_class = set(actual_class)
```

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89

90

91 92 93

94

96 97

101

```
roc_auc_aict = {}
102
       for per class in unique class:
103
         #creating a list of all the classes except the current class
104
        other_class = [x \text{ for } x \text{ in unique\_class if } x != per\_class]
105
106
         #marking the current class as 1 and all other classes as 0
107
        new_actual_class = [0 if x in other_class else 1 for x in actual_class]
108
        new pred class = [0 \text{ if } x \text{ in other class else } 1 \text{ for } x \text{ in pred class}]
109
110
         #using the sklearn metrics method to calculate the roc_auc_score
111
        roc_auc = roc_auc_score(new_actual_class, new_pred_class, average = average)
112
        roc_auc_dict[per_class] = roc_auc
113
114
        return roc auc dict
115
      def classification_report_csv(report):
117
        report_data = []
118
        lines = report.split('\n')
119
        for line in lines[2:-3]:
120
121
           row = \{\}
           row data = ''.ioin(line.split())
122
           row_data = row_data.split(' ')
123
           row['class'] = row_data[0]
124
125
           row['precision'] = float(row_data[1])
           row['recall'] = float(row data[2])
126
           row['f1_score'] = float(row_data[3])
127
128
           row['support'] = float(row_data[4])
           report_data.append(row)
130
        dataframe = pd.DataFrame.from_dict(report_data)
131
        dataframe.to_csv('classification_report.csv', index = False)
132
133
        return()
134
135
      def NetworkModel(ModelInput, TrainingDataX, TrainingDataY, ValidationX, ValidationY, TestSetX, TestSetY, epochs, batch, patience, verbose, ModelSaveName, ComputeWeight, RNN, CNN, Optimzation):
136
137
         # he learning hyper paramters are just set to what is reccomnded adjusting these paramters will not be the goal of this project.
138
         # these specifc paramaters were taken from the paper "ECG Heartbeat Classification: A Deep Transferable Representation"
139
        Ir schedule = tf.keras.optimizers.schedules.ExponentialDecay(0.001, decay_steps=10000, decay_rate=0.75)
140
        adam = Adam(learning_rate=Ir_schedule, beta_1=0.9, beta_2=0.999, amsgrad=False)
141
142
143
        if ComputeWeight:
144
           class\_weights = class\_weight.compute\_class\_weight(\begin{tabular}{c} balanced', \\ \end{tabular}
145
                                     np.unique(TrainingDataY),
146
                                      TrainingDataY)
147
148
           class_weights =[]
149
150
         # We will use the sparse C C and the optimzation function and look at accruacy for now
151
        ModelInput.compile(optimizer=adam, loss='sparse_categorical_crossentropy', metrics=['accuracy'])
152
153
         # Define some early stopping criteria here
154
        callb1= EarlyStopping(monitor='val_loss', mode='min', verbose=verbose,patience=patience) # lets monitor the loss in the opt function
155
        callb2 = ModelCheckpoint(filepath=ModelSaveName, monitor='val_loss', save_best_only=True) # Lets save it everytime it imrpoves as well
156
157
        if RNN: # fit setup for RNN network
158
159
                history = ModelInput.fit (TrainingDataX,\\
160
                            TrainingDataY.
161
                            epochs=epochs,
162
                            batch_size=batch
163
                            validation_data=(ValidationX, ValidationY),
164
                            verbose=verbose
165
                            class_weight = class_weights,
166
                            callbacks=[callb1,callb2])
168
        elif CNN: # fit setup for CNN network
169
170
                history = ModelInput.fit(np.expand_dims(TrainingDataX, axis=2),
171
                           TrainingDataY,
172
173
                           batch_size=batch,
174
                           validation\_data = (np.\ expand\_dims (Validation X,\ axis = 2),\ Validation Y),
175
                           verbose=verbose,
                           class_weight = class_weights,
177
                           callbacks=[callb1,callb2])
178
179
        plot learning(history)
180
         ModelInput.save(ModelSaveName)
181
        print('The model has been saved into your directory')
182
183
184
        return(ModelInput,history)
185
186
      \textbf{def optimization\_model} (Training Data X, \ Training Data Y, \ Validation X, \ Validation Y, X\_Test\_MIT, Y\_Test\_MIT) :
187
188
         ModelFor Training = Sequential() \\
189
        ModelForTraining.add(Flatten(input_shape=(187,1)))
190
191
         ModelForTraining.add(Dense({{choice([32,64,128])}}, input_shape=(187,1)))
192
         ModelForTraining.add(Activation({{choice(['relu', 'sigmoid'])}}))
193
        ModelForTraining.add(Dropout({{uniform(0.25, 0.75)}}))
194
195
        ModelForTraining.add(Dense({{choice([32,64,128])}}, input_shape=(187,1)))
196
197
        Model For Training. add (Activation (\{\{choice(['relu', 'sigmoid'])\}\}))
198
        ModelForTraining.add(Dropout({{uniform(0.25, 0.75)}}))
199
200
         ModelForTraining.add(Dense({{choice([32,64,128])}}, input_shape=(187,1)))
201
         ModelFor Training. add (Activation (\{\{choice([\cite{"relu"},\cite{"sigmoid"}])\}\}))
202
        ModelFor Training. add (Dropout (\{\{uniform (0.25,\ 0.75)\}\}))
203
         ModelForTraining.add(Dense({{choice([32,64,128])}}, input_shape=(187,1)))
204
205
         ModelForTraining.add(Activation({{choice(['relu', 'sigmoid'])}}))
206
        ModelFor Training. add (Dropout (\{\{uniform (0.25,\ 0.75)\}\}))
```

```
if ({\{choice(['two', 'three'])\}\}}) == 'three':
  ModelForTraining.add(Dense({{choice([32,64,128])}}))
ModelForTraining.add(Activation({{choice(['relu', 'sigmoid'])}}))
   ModelForTraining.add(Dropout({{uniform(0.25, 0.75)}}))
if ({{choice(['two', 'three'])}}) == 'three':
   ModelForTraining.add(Dense({{choice([32,64,128])}}))
   ModelForTraining.add(Activation({{choice(['relu', 'sigmoid'])}}))
   ModelForTraining.add(Dropout({{uniform(0.25, 0.75)}}))
if ({{choice(['two', 'three'])}}) == 'three':
   ModelForTraining.add(Dense({{choice([32,64,128])}}))
   ModelForTraining.add(Activation({{choice(['relu', 'sigmoid'])}}))
if ({{choice(['two', 'three'])}}) == 'three':
   ModelForTraining.add(Dense({{choice([32,64,128])}}))
   ModelForTraining.add(Activation({{choice(['relu', 'sigmoid'])}}))
ModelForTraining.add(Dense(5.activation='softmax'))
# in pilot testing i was experimenting with optimizing Model 1 to understand if the authors in the paper used the best paramters i.e if i could achieve better results
# although this was a good effort the processing was simply too slow on my hardware to make this a
# input_shape = (187, 1)
 # I = Input(input_shape)
# C = Conv1D(filters=32, kernel_size=5, strides=1)(I)
# a = {{choice(['relu', 'sigmoid', 'softmax'])}}
# C11 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(C)
# R11 = Activation(activation=a)(C11)
# C12 = Conv1D(filters=32, kernel size=5, strides=1, padding='same')(R11)
#A11 = Add()([C12, C])
# R12 = Activation(activation=a)(A11)
# M11 = MaxPool1D(pool_size=5, strides=2)(R12)
# C21 = Conv1D(filters=32, kernel_size=5, strides=1, padding='same')(M11)
# R21 = Activation(activation= a)(C21)
# C22 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(R21)
# A21 = Add()([C22, M11])
# R22 = Activation(activation=a)(A21)
# M21 = MaxPool1D(pool_size=5, strides=2)(R22)
# C31 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(M21)
# R31 = Activation(activation=a)(C31)
# C32 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(R31)
# A31 = Add()([C32, M21])
# R32 = Activation(activation=a)(A31)
# M31 = MaxPool1D(pool_size=5, strides=2)(R32)
# C41 = Conv1D(filters=32, kernel_size=5, strides=1, padding='same')(M31)
# R41 = Activation(activation=a)(C41)
# C42 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(R41)
# A41 = Add()([C42, M31])
# R42 = Activation(activation=a)(A41)
# M41 = MaxPool1D(pool_size=5, strides=2)(R42)
# C51 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(M41)
# R51 = Activation(activation=a)(C51)
# C52 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(R51)
\# A51 = Add()([M41,C52])
# R52 = Activation(activation=a)(A51)
# M51 = MaxPool1D(pool_size=5, strides=2)(R52)
# F1 = Flatten()(M51)
# D1 = Dense({{choice([32,64,128,256])}})(F1)
# R6 = Activation(activation={{choice(['relu', 'sigmoid'])}})(D1)
# D2 = Dense({{choice([32,64,128,256])}})(R6)
# D3 = Dense(5)(D2)
# O = Activation(activation='softmax')(D3)
# ModelForTraining = Model(inputs=I, outputs=O)
Ir_schedule = tf.keras.optimizers.schedules.ExponentialDecay(0.001, decay_steps=10000, decay_rate=0.75) adam = Adam(learning_rate=lr_schedule, beta_1=0.9, beta_2=0.999, amsgrad=False)
class_weights = class_weight.compute_class_weight('balanced',
                                np.unique(TrainingDataY),
                                TrainingDataY)
# class_weights = {0: 1.,1: 1.5,2: 1.5, 3:1.5,4:1.5} Experimenting here with different weights
# We will use the sparse C C and the optimzation function and look at accruacy for now
Model For Training. compile (optimizer=adam, \ loss='sparse\_categorical\_crossentropy', \ metrics=['accuracy'])
# Define some early stopping criteria here (this was not used for optimzation as it resulted in poor results)
# callb1= EarlyStopping(monitor='val_loss', mode='min', verbose=1,patience=10) # lets monitor the loss in the opt function
# callb2 = ModelCheckpoint(filepath=ModelSaveName, monitor='val_loss', save_best_only=True) # Lets save it everytime it imrpoves as well
history = ModelForTraining.fit (np.expand\_dims (TrainingDataX,\ axis=2), \\
                   TrainingDataY,
                    epochs=30.
                   batch size=64.
                    validation_data=(np.expand_dims(ValidationX, axis=2), ValidationY),
                   class weight = class weights)
                    # callbacks=[callb1])
```

test_loss, test_acc = ModelForTraining.evaluate(np.expand_dims(X_Test_MIT,axis=2), Y_Test_MIT, batch_size=64)

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```
# validation_acc = np.amax(history.history['val_accuracy']) # Not used
   print('Best validation acc of epoch:', test acc)
   return ('loss': -test acc. 'status': STATUS OK. 'model': ModelForTraining)
\textbf{def Calculate} \textbf{ModelMetrics} (\textbf{ModelOutput}, \textbf{History}, \textbf{TestSetX}, \textbf{TestSetY}, \textbf{batch}, \ \textbf{Naming}) : \textbf{ModelOutput}, \textbf{ModelOutp
    # function to make excel sheets and plot some of the data for further inspection
             results = ModelOutput.evaluate(np.expand_dims(TestSetX, axis=2), TestSetY, batch_size=batch)
             print(f"The accuracy on the testing set is {np.round(results[1]*100,1)}%")
   [Prediction, TrueValue,] = outputMetrics(TestSetX, TestSetY, ModelOutput, batch)
   clsf_reportClassification = pd.DataFrame(classification_report(y_true = TrueValue, y_pred = Prediction, output_dict=True)).transpose()
   confusionM = confusion_matrix(TrueValue, Prediction)
   clsf_report = pd.DataFrame(confusionM)
   plt.figure()
    ReportName = '%s' %(Naming)
   plot\_Conf\_Matrix(confusionM, classes=['N', 'S', 'V', 'F', 'Q'], normalize=True, title=ReportName)
     #plot_Conf_Matrix(confusionM,classes=['N', 'S'],normalize=True,title=ReportName
   plt.show()
   print("\nLogistic Regression")
     # assuming your already have a list of actual_class and predicted_class from the logistic regression classifier
   Ir_roc_auc_multiclass = roc_auc_score_multiclass(TrueValue, Prediction)
   print(lr_roc_auc_multiclass)
   hist_df = pd.DataFrame(History.history)
   hist_csv_file = 'History_%s' %(Naming)
   with open(hist_csv_file, mode='w') as f:
       hist df.to csv(f)
   jsonName = Naming.replace('.csv', ")
hist_json_file = 'History_%s.json' %(jsonName)
    with open(hist_json_file, mode='w') as f:
       hist_df.to_json(f)
   ReportName = 'ConfusionMatrix%s' %(Naming)
   clsf_report.to_csv(ReportName, index= True)
    ReportName = 'ClassificationReport%s' %(Naming)
    clsf_reportClassification.to_csv(ReportName, index= True)
def data_curation_MIT():
    # this function imports the data, performs the augmentation etc.
   SampleCounts= 10000
   noiseLevel = 0.03
   csv_MIT_train = pd.read_csv(r'E:\1_PAWEL\ENSC_project\heart_beatdata\mitbih_train.csv',header=None)
   csv\_MIT\_test=pd.read\_csv(r'E:\l_PAWEL\ENSC\_project\heart\_beatdata\mbox{\sc mitbih\_test.csv'}, header=None)
   DataSet_MIT= pd.concat([csv_MIT_train, csv_MIT_test], axis=0) # put it all into one structure
   Values MIT = DataSet MIT.values
   X_Data_MIT = Values_MIT[:,:-1]# remove the last column which is the calssification
   Y_Data_MIT = Values_MIT[:,-1] # keep the last column which will now be used for the Class Categroization
    StratifyData = StratifiedShuffleSplit(n_splits=5, test_size=0.2, random_state=100)
        what we are trying to accomplish here is to split up the data such that the number of samples in each class is somewhat evenly distrubuted
    # Since we already have a large class imbalance , this might be an issue if it is not addressed from the start. Lets split it up such that the
    # test size get 20% of the data (but catigorigaal samples evenly distrubuted)
   StratifyData.get_n_splits(X_Data_MIT, Y_Data_MIT)
   print(StratifyData)
   for train_index, test_index in StratifyData.split(X_Data_MIT, Y_Data_MIT):
       X_temp, X_Test_MIT = X_Data_MIT[train_index], X_Data_MIT[test_index]
        y_temp, Y_Test_MIT = Y_Data_MIT[train_index], Y_Data_MIT[test_index]
    # Now lets further split up our training data such that 20% of it goes to the validation sample #[X_Train_MIT, X_Valid_MIT, Y_Train_MIT, Y_Valid_MIT] = train_test_split(X_temp, y_temp, test_size=0.2)
   Stratify Data = Stratified Shuffle Split (n\_splits = 5, \ test\_size = 0.20, \ random\_state = 100)
        what we are trying to accomplish here is to split up the data such that the number of samples in each class is somewhat evenly distrubuted
    # Since we already have a large class imbalance , this might be an issue if it is not addressed from the start. Lets split it up such that the
     # test_size get 20% of the data (but catigorigcal samples evenly distrubuted)
   StratifyData.get_n_splits(X_temp, y_temp)
   \textbf{for} \ train\_index, \ test\_index \ \textbf{in} \ StratifyData.split(X\_temp, \ y\_temp):
        X_Train_MIT, X_Valid_MIT = X_temp[train_index], X_temp[test_index]
        Y_Train_MIT, Y_Valid_MIT = y_temp[train_index], y_temp[test_index]
    # TrainingDataX = X_Train_MIT # TRAINING DATA
    # TrainingDataY = Y Train MIT# TRAINING DATA Labels
   ValidationX = X_Valid_MIT
   ValidationY = Y_Valid_MIT
   temp\_X\_Train\_MIT = pd.DataFrame(X\_Train\_MIT)
   temp_Y_Train_MIT = pd.DataFrame(Y_Train_MIT)
   tempMIT_TrainingDataSet= pd.concat([temp_X_Train_MIT], axis=1,ignore_index=True) # put it all into one structure, ignore the index and autogenerate a new one
   print(tempMIT_TrainingDataSet[187].astype(int).value_counts()) # Lets see how many there are in each category. # so we have ~50k in the majority and >5k in the minoritys
    # at this point i think it makes sense to slightly reduce the sample count on the majority and then upsample the minoristy so ...
```

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#get the highest validation accuracy of the training epochs

```
410
419
        temp1=tempMIT_TrainingDataSet[tempMIT_TrainingDataSet[187]==1]
420
        temp2=tempMIT_TrainingDataSet[tempMIT_TrainingDataSet[187]==2]
421
        temp3=tempMIT_TrainingDataSet[tempMIT_TrainingDataSet[187]==3]
422
        temp 4 = temp MIT\_Training Data Set[temp MIT\_Training Data Set[187] == 4]
424
        temp0=(tempMIT_TrainingDataSet[tempMIT_TrainingDataSet[187]==0]).sample(n=SampleCounts,random_state=1) # We can downsample this
425
        temp1_upsample=resample(temp1, replace=True, n_samples=SampleCounts, random_state=2) # and we can upsample all of the minority classes
426
        temp2_upsample=resample(temp2,replace=True,n_samples=SampleCounts,random_state=3) # one note here is the random_state =var; this will let us reproduce the results everytime we run this
427
        temp3_upsample=resample(temp3,replace=True,n_samples=SampleCounts,random_state=4)
428
        temp4\_upsample = resample (temp4, replace = True, n\_samples = Sample Counts, random\_state = 5)
429
430
        # note that this resample approach will just randomly duplicate observations from the minority class. we might want to add some noise to
431
        # this data to avoid the nerual network just learning the samples. However, we need to be careful here as to not physically change he sample and its
432
        # classication. Therefore we will just add a bit of guassian noise
433
434
        MIT_Training_Resampled=pd.concat([temp0,temp1_upsample,temp2_upsample,temp3_upsample,temp4_upsample])
435
        print(MIT_Training_Resampled[187].astype(int).value_counts())
436
437
        \label{eq:mit_ming_Resampled} \mbox{MIT\_Training\_Resampled.values}
438
        X_Data_MIT_resampled_Training = MIT_Training_Resampled[:,:-1] # remove the last column which is the calssification
439
        Y_Data_MIT_resampled_Training = MIT_Training_Resampled[:,-1] # keep the last column which will now be used for the Class Categroization
440
441
       X\_Data\_MIT\_resampled\_Noise\_Training = np.empty(X\_Data\_MIT\_resampled\_Training.shape)
442
443
       for i in range(0,len(X_Data_MIT_resampled_Training)):
444
          noise1=np.random.normal(0,noiseLevel,187)
445
          noise2=np.random.normal(0,noiseLevel+0.01,187)+noise1
446
          noise=np.random.normal(0,noiseLevel-0.01,187)+noise2
447
          X\_Data\_MIT\_resampled\_Noise\_Training[i] = X\_Data\_MIT\_resampled\_Training[i] + noise \# \ \textit{lets loop through each sample and add some random noise}
448
449
450
       TrainingDataX = X_Data_MIT_resampled_Training # choose what training Dataset you want to USE (with augmnetaion or not??)
451
        Training Data Y = Y\_Data\_MIT\_resampled\_Training
452
453
        return TrainingDataX, TrainingDataY, ValidationX, ValidationY,X_Test_MIT,Y_Test_MIT
454
455
456
                     ----Plotting of data for inspection----
457
458
      # # %% Plotting of the Initial Data for Visulization
459
      # fig = plt.figure(figsize=(15,4))
460
461
     # for i in range(0,5):
462
      # plt.subplot(2,3,i + 1)
463
          all_samples_indexes = np.where(Y_Data_MIT == i)[0]
      ## rand_samples_indexes = np.random.randint(0, len(all_samples_indexes), 1)
465
        rand_samples = X_Data_MIT[all_samples_indexes[5]] # plot the 5th of each class
         plt.plot(rand_samples.transpose())
467
         plt.ylim(-0.1, 1.3)
468
        if i == 0.
469
            plt.ylabel('Norm Amplitude')
471
472
            plt.ylabel('Norm Amplitude')
         plt.title("Sample: " + classesMIT[i], loc='left', fontdict={'fontsize':10}, x=0.01, y=0.80)
473
475
476
     # fig.savefig('MIT_Data_Methods.png', format='png', dpi=800)
477
     # %%----- Train and TEST Model on MIT DataSET -----
478
479
480
      # lets load up the data for the model now
     Training Data X, \ Training Data Y, \ Validation X, \ Validation Y, X\_Test\_MIT, Y\_Test\_MIT = data\_curation\_MIT()
481
482
483
     n classes = len(np.unique(TrainingDataY)) # how many classes are there in our data
484
485
      # now pick which model we want to evaluate
486
     if Model1_CNN:
487
        print('Using Model 1')
488
        ModelForTraining = make_model_1(n_classes) # Using the archetecture CHANGE HERE
489
490
491
492
        ModelForTraining = make_model_2_RNN(n_classes) # Using the archetecture CHANGE HERE
493
494
        # for the RNN model we need to reshape the input a bit for it to work
495
        TrainingDataX_RNN = np.reshape(TrainingDataX,(TrainingDataX.shape[0],TrainingDataX.shape[1],1))
496
        ValidationX_RNN= np.reshape(ValidationX,(ValidationX.shape[0],ValidationX.shape[1],1))
497
        TestDataX\_RNN = np.reshape(X\_Test\_MIT,(X\_Test\_MIT.shape[0],X\_Test\_MIT.shape[1],1))
498
499
       del TrainingDataX, TestSetX, ValidationX
500
501
        TrainingDataX = TrainingDataX_RNN
502
        ValidationX = ValidationX_RNN
503
       X Test MIT =TestDataX RNN
504
505
       n classes = len(np.unique(TrainingDataY))
506
507
     elif Model3_OptimizationModel:
508
        print('Using Model 3')
509
       trials = Trials()
510
512
     from os import path
513
     pathFile = (r'E:\1_PAWEL\ENSC_project\%s' % (ModelSaveName))
514
      var = path.exists(pathFile)
515
     if not var:
516
517
          if Model3 OptimizationModel:
518
             Here is the setup for the optimized model, we set max evals to 40 mainly as a hardware contraint as it takes quite a long time.
519
             best_run, ModelOutput,return_space = optim.minimize(model=optimization_model,
520
                                        data = data_curation_MIT,
521
                                        algo=tpe.suggest,
522
                                        max evals=40.
523
```

trials=trials

```
526
                           saving it in this manner as to deal with issues of loading .h5 file after the optization. this seems to work now
527
                          ModelSave = trials.results[np.argmin([r['loss'] \ for \ r \ in \ trials.results])]['model']
528
                         ModelSave.save(ModelSaveName)
529
530
531
532
                         [ModelOutput, History]= NetworkModel(ModelForTraining, TrainingDataY, TrainingDataY, ValidationY, ValidationY, X_Test_MIT, Y_Test_MIT, Y_T
533
                                                                 epochs, batch, patience, verbose, Model SaveName, ComputeWeight=False, RNN=False, CNN= True, Optimzation = False)
534
535
536
                print('Loading preexcisting model from folder .....')
                \label{local_model} ModelOutput = load\_model(r'E: \label{local_model} L_PAWEL\ense \cite{Lense} Lense \cit
537
538
539
                jsonName = ModelSaveName.replace('.h5', ")
540
                hist_json_file = 'History_%s.json' %(jsonName)
541
                with open(hist_json_file, 'r') as json_file:
542
                    History= json_file.read()
543
544
              # this will not spit out the results, csv files etc into the folder to further look at
545
            History = ModelOutput.history
546
            CalculateModelMetrics(ModelOutput,History,X_Test_MIT,Y_Test_MIT,batch, Naming='MIT_resampled_Training_Weights.csv') #
547
548
549
           plt.figure(figsize=(5, 5))
550
           plt.plot(History.history['accuracy'])
           plt.plot(History.history['val_accuracy'])
552
            plt.title('model accuracy')
553
           plt.ylabel('accuracy')
554
           plt.xlabel('epoch')
           plt.legend(['train', 'validation'], loc='upper left')
            plt.show()
557
            # summarize history for loss
558
            # Plot non-normalized confusion matrix
559
           plt.figure(figsize=(5, 5))
           plt.plot(History.history['loss'])
561
            plt.plot(History.history['val_loss'])
562
           plt.title('model loss')
563
           plt.ylabel('loss')
           plt.xlabel('epoch')
565
           plt.legend(['train', 'validation'], loc='upper left')
566
           plt.show()
567
568
            # %% Exploring Transfer Learning
570
           # lets load in the other data to explore transfer learning csv_PTB_Normal = pd.read_csv(r'E:\1_PAWEL\ENSC_project\heart_beatdata\ptbdb_normal.csv',header=None)
571
572
           csv_PTB_AbNormal = pd.read_csv(r'E:\1_PAWEL\ENSC_project\heart_beatdata\ptbdb_abnormal.csv',header=None)
            DataSet_PTB = pd.concat([csv_PTB_Normal, csv_PTB_AbNormal], axis=0)
574
           Values_PTB = DataSet_PTB.values # get the values
575
576
           X_Data_PTB = Values_PTB[:,:-1] # remove the last column which is the calssification
577
            Y_Data_PTB = Values_PTB[:,-1] # keep the last column which will now be used for the Class Categroization
578
579
           StratifyData = StratifiedShuffleSplit(n_splits=5, test_size=0.2, random_state=50)
# what we are trying to accomplish here is to split up the data such that the number of samples in each class is somewhat evenly distrubuted
580
581
                Since we already have a large class imbalance, this might be an issue if it is not addressed from the start. Lets split it up such that the
582
             # test_size get 20% of the data (but catigorigcal samples evenly distrubuted)
583
           Stratify Data\_get\_n\_splits (X\_Data\_PTB,\ Y\_Data\_PTB)
584
585
           print(StratifyData)
586
587
            for train_index, test_index in StratifyData.split(X_Data_PTB, Y_Data_PTB):
588
                X\_temp,\ X\_Test\_PTB = X\_Data\_PTB[train\_index],\ X\_Data\_PTB[test\_index]
589
                y_temp, Y_Test_PTB = Y_Data_PTB[train_index], Y_Data_PTB[test_index]
590
591
           uniqueValues, occurCountTrain = np.unique(y_temp, return_counts=True) uniqueValues, occurCountValid = np.unique(Y_Test_PTB, return_counts=True)
592
593
594
           print(occurCountTrain)
595
           print(occurCountValid)
597
           for train index, test index in StratifyData.split(X temp, y temp):
598
599
               X\_Train\_PTB,\ X\_Valid\_PTB = X\_temp[train\_index],\ X\_temp[test\_index]
600
                Y_Train_PTB, Y_Valid_PTB = y_temp[train_index], y_temp[test_index]
601
602
            # # Plotting PTB database 1 sample from each cat
603
604
            # plt.figure(figsize=(15,4))
605
            # for i in range(0.2):
606
                plt.subplot(1,2,i+1)
607
                   all_samples_indexes = np.where(Y_Data_PTB == i)[0]
608
                   rand_samples_indexes = np.random.randint(0, len(all_samples_indexes), 1) rand_samples = X_Data_PTB[all_samples_indexes[5]]
609
610
                   plt.plot(rand_samples.transpose())
                   plt.title("Sample: " + classePTB[i], loc="left", fontdict={'fontsize':10})
612
613
            # change this line to change which model we want to evaluate
614
           ModelOutputTransfer = ModelOutput # CHANGE THIS TO WHAT YOU WANT TO TEST
615
616
             # lets build the new transfer learning layers
617
           D1 = Dense(32)(ModelOutputTransfer.output)
618
619
           D2 = Dense(32)(D1)
621
           O = Dense(2, activation='softmax')(D2)
622
           modelTransfer = Model(inputs=ModelOutputTransfer.input, outputs=O) # we have our new model
623
624
           for layer in modelTransfer.layers[:-3]: # change this number to explore changing more layers in case of report i looked at -3,-5, -8
625
626
627
           for layer in modelTransfer.layers[-3:]: # change this number to explore changing more layers in case of report i looked at -3,-5, -8
628
               layer.trainable = True
```

525

return_space = True)

we can use the function developed before and run it no on the new transfer learning model.

| we can use the function developed before and run it no on the new transfer learning model.

| findelTransferOutput, History] = NetworkModel(modelTransfer,X_Train_PTB,Y_Train_PTB,X_Valid_PTB,Y_Valid_PTB,X_Test_PTB,Y_Test_PTB, epochs,batch,patience,verbose,ModelSaveName,ComputeWeight=False,RNN=False, CNN=True, Optimization = False)

lets output and look at how the transfer learning did on the novel dataset...

CalculateModelMetrics(modelTransferOutput,History,X_Test_PTB,Y_Test_PTB,batch, 'Transfer')

```
from tensorflow.keras.layers import Input, Conv1D, MaxPool1D, Activation, Add, Dense, Flatten
    from tensorflow.keras.models import Model
2
3
4
    def make model 1(final layer size):
5
      input\_shape = (187, 1)
6
      I = Input(input_shape)
7
      C = Conv1D(filters=32, kernel_size=5,strides=1)(I)
8
9
      C11 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(C)
10
      R11 = Activation(activation='relu')(C11)
11
      C12 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(R11)
12
      A11 = Add()([C12, C])
13
      R12 = Activation(activation='relu')(A11)
14
      M11 = MaxPool1D(pool_size=5, strides=2)(R12)
15
16
      C21 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(M11)
17
      R21 = Activation(activation='relu')(C21)
18
      C22 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(R21)
19
      A21 = Add()([C22, M11])
20
      R22 = Activation(activation='relu')(A21)
21
      M21 = MaxPool1D(pool_size=5, strides=2)(R22)
22
23
      C31 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(M21)
24
      R31 = Activation(activation='relu')(C31)
25
      C32 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(R31)
26
      A31 = Add()([C32, M21])
27
      R32 = Activation(activation='relu')(A31)
28
      M31 = MaxPool1D(pool_size=5, strides=2)(R32)
29
30
      C41 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(M31)
31
      R41 = Activation(activation='relu')(C41)
32
      C42 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(R41)
33
      A41 = Add()([C42, M31])
34
      R42 = Activation(activation='relu')(A41)
35
      M41 = MaxPool1D(pool_size=5, strides=2)(R42)
36
37
      C51 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(M41)
38
      R51 = Activation(activation='relu')(C51)
39
      C52 = Conv1D(filters=32, kernel_size=5,strides=1,padding='same')(R51)
40
      A51 = Add()([M41,C52])
41
      R52 = Activation(activation='relu')(A51)
42
      M51 = MaxPool1D(pool_size=5, strides=2)(R52)
43
44
      F1 = Flatten()(M51)
45
46
      D1 = Dense(32)(F1)
47
      R6 = Activation(activation='relu')(D1)
48
      D2 = Dense(32)(R6)
49
      D3 = Dense(final_layer_size)(D2)
50
      O = Activation(activation='softmax')(D3)
51
52
      return Model(inputs=I, outputs=O)
```

```
from tensorflow.keras.layers import Input, Dropout, LSTM, Bidirectional, BatchNormalization, Dense
1
2
    from tensorflow.keras.models import Sequential
3
4
5
    def make_model_2_RNN(final_layer_size):
6
7
8
      Model = Sequential()
9
      Model.add(Dense(32, input_shape=(187,1)))
10
      Model.add(Bidirectional(LSTM(64,input_shape = (187,1))))
11
      Model.add(Dropout(rate =0.25))
12
      Model.add(BatchNormalization())
13
      Model.add(Dense(32, activation='relu'))
14
      Model.add(Dropout(rate =0.25))
15
      Model.add(BatchNormalization())
16
      Model.add(Dense(final_layer_size,activation = 'sigmoid'))
17
18
19
20
      return Model
```