Prediction of WHO Life Expectancy Data Using Multiple Linear Regression And Artificial Neural Network

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KEYWORDS

Life Expectancy
Multiple Linear Models
Artificial Neural Network
Machine Learning
R
Coefficient of Determination
Prediction
MSE
MAPE
Accuracy

ABSTRACT

Life expectancy is the key for assessing the population health. A lot of researches have already been done based on the factors of life expectancy data considering demographic variables, income composition and mortality rates. Researches on multiple linear regression based on a data set of one year for all the countries are available.

Now we can build a predictive model based on this data using multiple linear regression from the data set of 2000-2015 of all countries by dividing the whole dataset into two parts namely training and testing set in ratio 80:20. After fitting the model in training data and checking the model performance on both dataset we found that MLR is a very good fit for prediction, but it violates normality and homoscedasticity assumptions. To overcome this problem we fitted this data on ANN model in the same way like before which does not assume those assumptions. We found that ANN model performed even better than MLR. At last we compared their performances and tried to find pros and cons of both models.

I. INTRODUCTION

Life Expectancy is a statistical measure of the average time an organism is expected to live, based on the year of its birth, its current age and other demographic factors including gender. In other words, it is the statistical age that a person is expected to live until, based on actuarial data. Life expectancy at birth (LEB) is the most commonly used measure worldwide.

Assuming that a person maintains the minimum standard of hygiene, life expectancy may be the outcome of multiple factors such as Diet, Lifestyle, Heredity/genetic, Marital status, Local standard of healthcare and disease prevention, Access to clean water and fresh food supply, Community and social support, Absence of adverse environmental factors like pollution

To compute life expectancy it's a bit difficult & uses lot of different data sources for each country. Life expectancy is calculated through life table or life table calculator. The life table incorporates data with age

specific death rates for the population in question which requires enumeration of data for the number of people & number of death at each age for that population. Most currently available life table or life expectancy calculators are based on a person's age, gender & race. According to actuarial science, life expectancy takes into account several individual-level as well as population-level factors to arrive at a figure.

The population of many rich developed countries has life expectancy around 80 years whereas the life expectancy of poor developing country has life expectancy around 50 to 60 years.

There are many uses for it in the financial world, including life insurance, pension planning, and Social security benefits. The most important use of life expectancy is in clinical researches. Life expectancy is used in pricing and underwriting life insurance and insurance products like annuities, as well as in retirement and pension planning.

II. DATA DESCRIPTION

The dataset we used for our project was provided by the Kaggle.com website. It is a free open source platform to all. The data was collected from WHO and United Nations website with the help of Deeksha Russell and Duan Wang. This dataset has usability rating 8.2. The data consists of 2938 rows and 22 columns. As the dataset is from WHO, we assume there is no error.

Dataset Link: <u>https://www.kaggle.com/kumarajarshi/life-expectancy-who</u>

Response Variable:

Y: Life expectancy – Life Expectancy in age

Regressors:

Country - Country

Year - Year

X1: Status – Developed or developing status

X2: Adult Mortality – Adult Mortality Rates of both sexes (probability of dying between 15 and 60 years per 1000 population)

X3: Infant deaths – Number of Infant Deaths per 1000 population

X4: Alcohol – Alcohol, recorded per capita (15+) consumption (in litres of pure alcohol)

X5: Percentage expenditure – Expenditure on health as a percentage of Gross Domestic Product per capita (%)

X6: Hepatitis B – Hepatitis B (HepB) immunization coverage among 1-year-olds (%)

X7: Measles – Number of reported Measles cases per 1000 population

X8: BMI – Average Body Mass Index of entire population

X9: Under-five deaths – Number of under-five deaths per 1000 population

X10: Polio – Polio (Pol3) immunization coverage among 1-year-olds (%)

X11: Total expenditure – General government expenditure on health as a percentage of total government expenditure (%)

X12: Diphtheria – Diphtheria tetanus toxoid and pertussis (DTP3) immunization coverage among 1-year-olds (%)

X13: HIV/AIDS – Deaths per 1 000 live births HIV/AIDS (0-4 years)

X14: GDP – Gross Domestic Product per capita (in USD)

X15: Population – Population of the country

X16: Thinness 1-19 years – Prevalence of thinness among children and adolescents for Age 10 to 19 (%)

X17: Thinness 5-9 years – Prevalence of thinness among children for Age 5 to 9(%)

X18: Income composition of resources – Human Development Index in terms of income composition of resources (index ranging from 0 to 1)

X19: Schooling – Number of years of Schooling (years)

```
> str(data)
              2938 obs. of 22 variables:
'data.frame':
                               : chr "Afghanistan" "Afghanistan" "Afghanistan" ...
$ Country
                               : int 2015 2014 2013 2012 2011 2010 2009 2008 2007 2006 ...
 $ Year
                               : chr "Developing" "Developing" "Developing" "Developing" ...
$ Status
                               : num 65 59.9 59.9 59.5 59.2 58.8 58.6 58.1 57.5 57.3
 $ Life expectancy
                               : int 263 271 268 272 275 279 281 287 295 295 ...
$ Adult Mortality
$ infant deaths
                               : int 62 64 66 69 71 74 77 80 82 84 ...
$ Alcohol
                               71.3 73.5 73.2 78.2 7.1 ...
$ percentage expenditure
                               : num
$ Hepatitis B
                                      65 62 64 67 68 66 63 64 63 64 ...
                               : int
$ Measles
                                      1154 492 430 2787 3013 1989 2861 1599 1141 1990 ...
                               : int
$ BMI
                               : num 19.1 18.6 18.1 17.6 17.2 16.7 16.2 15.7 15.2 14.7 ...
                               : int 83 86 89 93 97 102 106 110 113 116 ...
$ under.five_deaths
                                      6 58 62 67 68 66 63 64 63 58 ...
 $ Polio
                               : int
                               : num 8.16 8.18 8.13 8.52 7.87 9.2 9.42 8.33 6.73 7.43 ...
$ Total expenditure
$ Diphtheria
                               : int 65 62 64 67 68 66 63 64 63 58 ...
$ HIV_AIDS
                               : num 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 ...
$ GDP
                               : num 584.3 612.7 631.7 670 63.5 ...
                               : num 33736494 327582 31731688 3696958 2978599 ...
$ Population
$ thinness_.1.19_years
                               : num 17.2 17.5 17.7 17.9 18.2 18.4 18.6 18.8 19 19.2 ...
$ thinness_5.9_years
                               : num 17.3 17.5 17.7 18 18.2 18.4 18.7 18.9 19.1 19.3 ...
$ Income composition of resources: num 0.479 0.476 0.47 0.463 0.454 0.448 0.434 0.433 0.415 0.405 ...
$ Schooling
                               : num 10.1 10 9.9 9.8 9.5 9.2 8.9 8.7 8.4 8.1 ...
>
```

III. OBJECTIVE

The World Health Organization (WHO) keeps track of the health status and many others related factors for all countries, as well. This dataset for 193 countries has been collected from the WHO data. In this project we consider data from year 2000-2015 for 193 countries for analysis. As the dataset is from WHO there is no error in the data.

In the respective project we are going to find out the following factors:

- Whether the initially chosen predicting factors really affects the life expectancy or not & those predicting variables which actually affects the life expectancy.
- The type of correlation between life expectancy and other factors like smoking, exercise, lifestyles, alcohol consumptions etc.
- The impact of immunization coverage on life expectancy.
- Whether we can build a reliable predictive model to predict life expectancy in future or for any external dataset.

For doing the above we at first will build up a multiple linear regression model which is a supervised ML Model, based on Training data by dividing the whole data set in training set and test set randomly in ratio 80:20. Then we will check the multicollinearity, outliers, normality, and homoscedasticity. We will find out how well the model fits to the training data and how it predicts the test data by coefficient of determination along with MSE, MAPE and Accuracy. Then we will apply a Artificial Neural Network on the train data and check the model adequacy just like before. At last we will compare both models and conclude our findings accordingly.

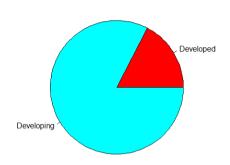
IV. THE R ENVIRONMENT

Here we are using R language for our analysis. R is a language and environment for statistical computing and graphics. R provides a wide variety of statistical (linear and nonlinear modeling, classical statistical tests, timeseries analysis, classification, clustering ...) and graphical techniques, and is highly extensible.

R is one of the major languages for data science. It provides excellent visualization features, which is essential to explore the data before submitting it to any automated learning, as well as assessing the results of the learning algorithm. Many R packages for machine learning are available off the shelf and many modern methods in statistical learning are implemented in R as part of their development.

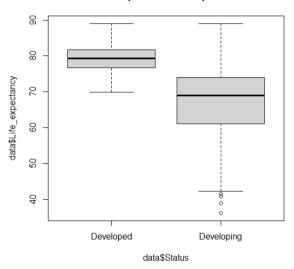
V. DATA INSIGHTS

Status wise data Share



82.57318 % of total dataset comes from developing countries and 17.42682 % data comes from developed countries.

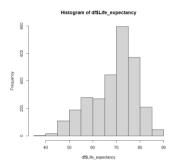
Comparison of Boxplots

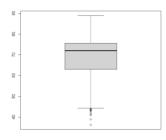


Note that avg. life expectancy in developed countries is around 80 years and spread is less compared to developing countries whereas in developing countries avg. life expectancy is around 70 years. There are some outliers in developing data points.

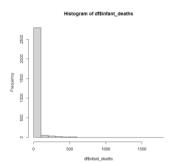
Now let us take a look into the data visualizations. Below we have histogram and boxplot of each variable

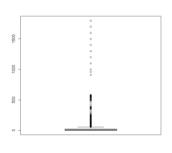
Y: Life expectancy



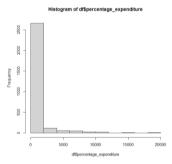


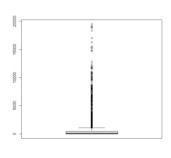
X3: Infant deaths



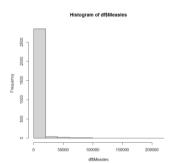


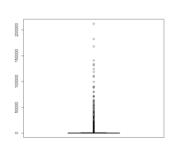
X5: Percentage expenditure



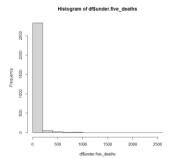


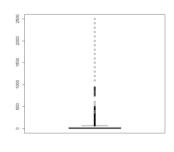
X7: Measles



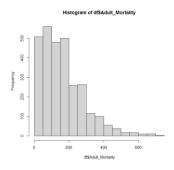


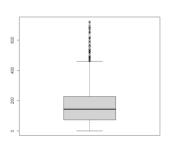
X9: Under-five deaths





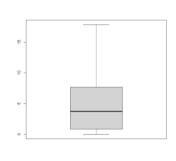
X2: Adult Mortality



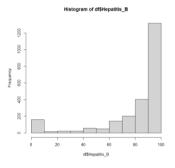


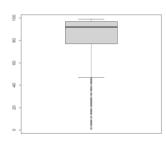
X4: Alcohol

Histogram of dfSAlcohol

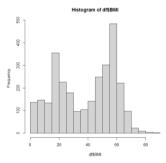


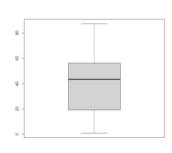
X6: Hepatitis B



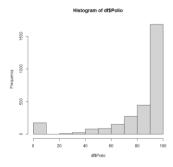


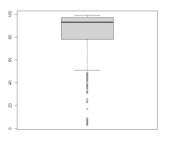
X8: BMI



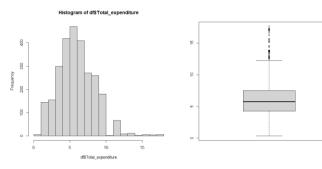


X10: Polio

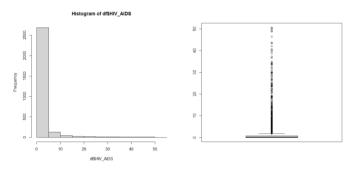




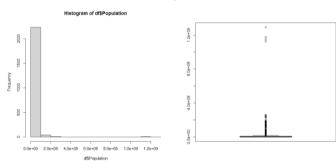
X11: Total expenditure



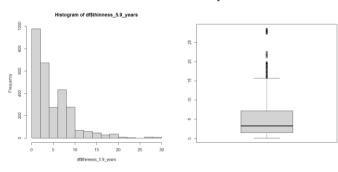
X13: HIV/AIDS



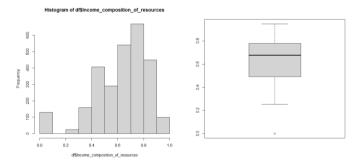
X15: Population



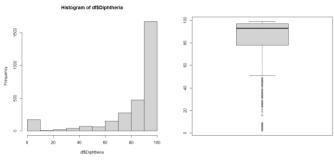
X17: Thinness 5-9 years



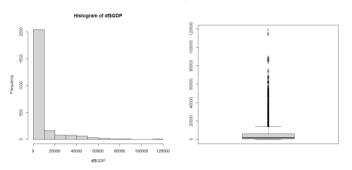
X19: Schooling



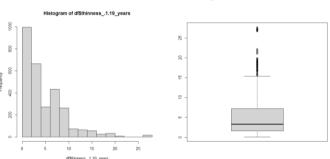
X12: Diphtheria



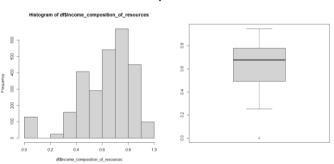
X14: GDP



X16: Thinness 1-19 years

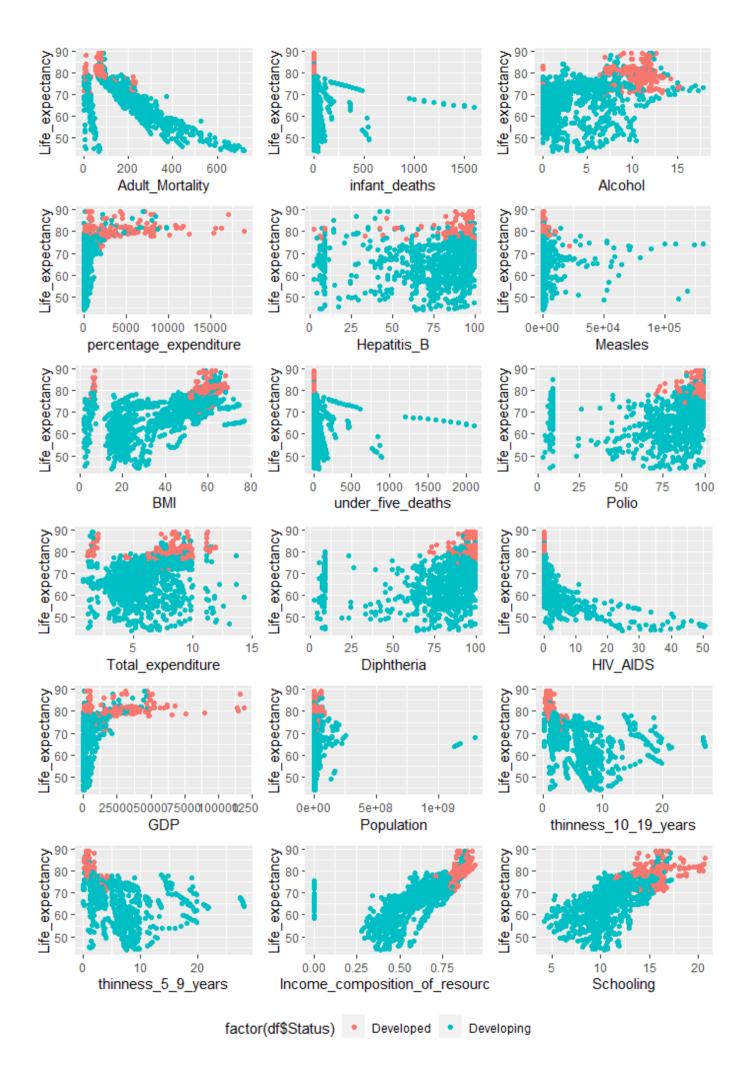


X18: Income composition of resources



We can see most of the variables are highly skewed and contain too much outlier.

Now we will draw individual scatterplot to visualize the relationship of each regressor with the response variable. The blue points correspond to developing country and red points belong to developed country.



VI. DATA PREPROCESSING

For machine learning algorithms to work it is necessary to convert the raw data into a clean data set and dataset must be converted to numeric data. You have to encode all the categorical labels to column vectors with binary values. Missing values or NaNs in the dataset is an annoying problem. You have to either drop the missing rows or fill them up with mean or interpolated values. Both the training data and test data must have same dimensions for the model.

Dropping Columns which are not useful

At first step we will drop some of the columns which may not contribute much to our machine learning model such as Country, Year.

Renaming Columns

For ease of our study and modeling we have renamed a few columns such as: "Life.expectancy" to "Life_expectancy"; or "thinness..1.19.years" to "thinness 10 19 years" etc.

Dealing with Missing values

```
> sapply(data, function(x) sum(is.na(x))/length(x)*100)
                                                            Status
                Life expectancy
                       0.3403676
                                                        0.0000000
                Adult Mortality
                                                    infant deaths
                       0.3403676
                                                        0.0000000
                         Alcohol
                                           percentage expenditure
                       6.6031314
                                                        0.0000000
                                                          Measles
                     Hepatitis B
                      18.8223281
                                                        0.0000000
                             BMI
                                                under_five_deaths
                       1.1572498
                                                        0.0000000
                           Polio
                                                Total_expenditure
                       0.6466984
                                                         7.6923077
                                                         HIV AIDS
                      Diphtheria
                                                        0.0000000
                       0.6466984
                             GDP
                                                       Population
                      15.2484683
                                                       22.1919673
           thinness_10_19_years
                                               thinness_5_9_years
                       1.1572498
                                                         1.1572498
Income composition of resources
                                                        Schooling
                       5.6841389
```

We can see that a lot of values are missing in this data. If we delete those rows completely we might lose a large amount of data. So instead of removing the null values completely, we have used data imputation technique. We have replaced the missing cells by interpolation technique. At last we have removed remaining null values.

VII. METHOD OF ANALYSIS

[A] MULTIPLE LINEAR REGRESSION

Here our multiple linear model is Y=Xβ+ε
Where,
Y=vector of responses
X=Design matrix (Vector of Regressors)

β=vector of unknown parameters

 $\varepsilon = \text{error of linear model}$

Assumptions:

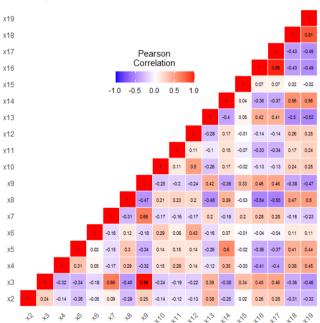
- Errors are i.i.d. distributed with $E(\epsilon)=0$ & $var(\epsilon)=\sigma 2In$.
- Explanatory variables are independent i.e. X is of full column rank.

Data Transformation

From the Histogram and boxplot of the Regressors it is evident that most of them are highly skewed. Skewed data is cumbersome and common. Different features in the data set may have values in different ranges. Data transformation comes to our aid in such situations. Here we have used log transformation and cube root transformation for skewed data.

Heatmap & correlation Matrix

a good way to quickly check correlations among columns is by visualizing the correlation matrix as a heatmap.



We see there is high correlation between x18-x19, x16-x17& x3-x9. So we have removed x19, x16 & x9 from the data frame.

Detection and Removal of Outlier

In statistics, an outlier is defined as an observation which stands far away from the most of other observations. Often an outlier is present due to the measurements error. Therefore, one of the most important tasks in data analysis is to identify and (if is necessary) to remove the outliers. Outliers can occur by chance in any distribution, but they often indicate either measurement error or that the population has a heavy-tailed distribution. In the former case one

wishes to discard them or use statistics that are robust to outliers, while in the latter case they indicate that the distribution has high skewness and that one should be very cautious in using tools or intuitions that assume a normal distribution. There is no rigid mathematical definition of what constitutes an outlier; determining whether or not an observation is an outlier is ultimately a subjective exercise. There are various methods of outlier detection.

Cook's Distance: Di is used in Regression Analysis to find the influential outliers in a set of predictor variables. In other words, it's a way to identify points that negatively affect your regression model. The measurement is a combination of each observation's leverage and residual values; the higher the leverage and residuals, the higher the Cook's distance.

Technically, Cook's Di is calculated by removing the ith data point from the model and recalculating the regression. It summarizes how much all the values in the regression model change when the ith observation is removed. The formula for Cook's distance is:

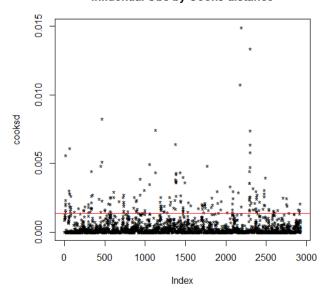
$$D_{i} = \frac{\sum_{j=1}^{n} (\hat{Y}_{j} - \hat{Y}_{j(i)})^{2}}{(p+1)\hat{\sigma}^{2}}$$

Where, Yj(i) is the fitted response value obtained when excluding i.

Decision Rule:

If (cook's distance) Di > 4/n
Then we conclude that point is outlier.
Where, n= no of rows in dataset.

Influential Obs by Cooks distance



The points above the red line are considered to be outliers.

After this we have divided the whole dataset in training and test set randomly in the ratio 80:20. Then we have deployed MLR on the training dataset.

Multicollinearity

Variance Inflation Factor: Consider the following linear model with k independent variables:

 $Y = \beta 0 + \beta 1 X1 + \beta 2 X2 + ... + \beta p Xp + \epsilon$.

VIF = 1/(1-Rj2), for all j = 1(1) p

Where, Rj2 is the coefficient of determination when Xj is regressed on the remaining regressor. High value of Rj2 suggests that Xj is well explained by the remaining set of regressor.

VIF > 2 is considered large & it suggests the presence of multicollinearity.

```
> vif(fit)
x1 x2 x3 x4 x5 x6 x7 x8
1.758423 1.283388 2.653781 1.560637 1.820716 1.350836 1.773769 1.777512
x10 x11 x12 x13 x14 x15 x17 x18
1.348084 1.258930 1.579042 1.859377 2.112175 1.205035 1.982151 2.234704
```

Variable Selection

Forward Backward & Stepwise Regression: The stepwise regression (or stepwise selection) consists of iteratively adding and removing predictors, in the predictive model, in order to find the subset of variables in the data set resulting in the best performing model that is a model that lowers prediction error.

There are three strategies of stepwise regression (James et al. 2014,P. Bruce and Bruce (2017)):

Forward selection, which starts with no predictors in the model, iteratively adds the most contributive predictors, and stops when the improvement is no longer statistically significant.

Backward selection (or **backward elimination**), which starts with all predictors in the model (full model), iteratively removes the least contributive predictors, and stops when you have a model where all predictors are statistically significant.

Stepwise selection (or sequential replacement), which is a combination of forward and backward selections. You start with no predictors, then sequentially add the most contributive predictors (like forward selection). After adding each new variable, remove any variables that no longer provide an improvement in the model fit (like backward selection). After stepwise selection, we will apply

Mellow Cp Statistic, AIC and adjusted R2 criterion for variable selection.

Mellow Cp statistic: In Mellow Cp statistic, we check the value of E(p) for different p(no. of regressor). A common method is to perform all possible regressions, then use Mallow's Cp to compare the results. The smaller Cp value is better as it indicate smaller amounts of unexplained error. We select the model of those p regressor for which **E** (p) \cong p.

AIC: The Akaike Information Criterion, or AIC for short, is a method for scoring and selecting a model. To use AIC for model selection, we simply choose the model giving smallest AIC over the set of models considered.

Output in Forward selection

```
> step_model_forward = stepAIC(fit ,direction="forward",trace=1)
Start: AIC=5708.14

y ~ x1 + x2 + x4 + x5 + x6 + x7 + x8 + x10 + x11 + x12 + x13 + x14 + x15 + x17
> step model forward
 lm(formula = y \sim x1 + x2 + x4 + x5 + x6 + x7 + x8 + x10 + x11 + x12 + x13 + x14 + x15 + x17, data = mlr_train) 
Coefficients:
 (Intercept) x1Developing
             -1.51952
   57.85164
                              -0.93520
                                           1.06096
                                                         0.19285
                             0.02849
                                           0.66452
                                                        0.13929
                -0.07476
   -0.02288
                x13
-3.43951
                                           -0.01035
                                                         -0.73650
                              0.35661
    1.00246
> best_subset_forward <-ols_step_best_subset(step_model_forward)
> best_subset_forward
                   Best Subsets Regression
Model Index Predictors
               x13
               x5 x13
               x4 x5 x13
               x4 x5 x13 x17
     5
               x2 x4 x5 x13 x17
x2 x4 x5 x12 x13 x17
     6
               x2 x4 x5 x7 x12 x13 x17
     8
               x2 x4 x5 x7 x12 x13 x14 x17
               x1 x2 x4 x5 x7 x12 x13 x14 x17
               x1 x2 x4 x5 x7 x8 x12 x13 x14 x17
               x1 x2 x4 x5 x7 x8 x10 x12 x13 x14 x17
               x1 x2 x4 x5 x7 x8 x10 x11 x12 x13 x14 x17
    12
               x1 x2 x4 x5 x7 x8 x10 x11 x12 x13 x14 x15 x17
               x1 x2 x4 x5 x6 x7 x8 x10 x11 x12 x13 x14 x15 x17
```

					Subsets Regr
Model	R-Square	Adj. R-Square	Pred R-Square	C (p)	AIC
1	0.6948	0.6947	0.6943	1978.1067	13300.3883
2	0.7628	0.7626	0.7623	1053.4563	12751.8446
3	0.7930	0.7927	0.7923	644.3261	12456.6826
4	0.8063	0.8059	0.8054	465.3964	12313.8917
5	0.8170	0.8166	0.8159	321.5698	12191.7727
6	0.8248	0.8243	0.8236	217.5346	12098.8736
7	0.8296	0.8291	0.8283	153.6494	12039.7598
8	0.8333	0.8327	0.8318	106.0126	11994.5607
9	0.8361	0.8354	0.8346	69.6374	11959.3575
10	0.8385	0.8378	0.8369	38.2563	11928.4721
11	0.8398	0.8390	0.838	22.7116	11912.9872
12	0.8408	0.8399	0.8389	11.1269	11901.3520
13	0.8408	0.8399	0.8388	13.0223	11903.2467
14	0.8408	0.8398	0.8386	15.0000	11905.2242

SBIC: Sawa's Bayesian Information Criteria SBC: Schwarz Bayesian Criteria MSEP: Estimated error of prediction, assuming multivariate normality FPE: Final Prediction Error HSP: Hocking's Sp

APC: Amemiva Prediction Criteria

Here, the value of E(Cp) =13 corresponding the p(13), So, we select 13th subset model. Also, the value of adj. R2 correspond to 14th subset model is 0.8399 which is highest among all the subset models & AIC is minimum.

Output in Backward selection

```
> step_model_backward = stepAIC(fit ,direction="backward",trace=1)
   Start: AIC=5708.14
y ~ x1 + x2 + x4 + x5 + x6 + x7 + x8 + x10 + x11 + x12 + x13 +
       x14 + x15 + x17
          Df Sum of Sq RSS AIC
1 0 29423 5706.2
   - x6
   <none>
                          29423 5708.1
  <none>
- x11   1
- x10   1
- x8   1
- x1   1
- x12   1
- x14   1
- x7   1
- x17   1
- x17   1
                    183 29606 5719.7
                      226 29649 5722.9
                     400 29822 5735.6
                      449 29872 5739 2
                     465 29888 5740.4
                     563 29986 5747.5
                     652 30075 5754.0
   - x5
                    1176 30599 5791.7
1605 31027 5822.1
   - x2
   - x13 1
                   39445 68868 7562.6
  Step: AIC=5706.16
  y ~ x1 + x2 + x4 + x5 + x7 + x8 + x10 + x11 + x12 + x13 + x14 + x15 + x17
  Df Sum of Sq RSS AIC - x15 1 1 29425 5704.3 
  - x11 1
- x10 1
- x8 1
                      183 29606 5717.7
226 29649 5720.9
  y ~ x1 + x2 + x4 + x5 + x7 + x8 + x10 + x11 + x12 + x13 + x14 + x17
   Step: AIC=5704.27
   y \sim x1 + x2 + x4 + x5 + x7 + x8 + x10 + x11 + x12 + x13 + x14 +
       x17
           Df Sum of Sq
                             RSS
   <none>
                           29425 5704.3
                      184 29609 5715.9
   - x11
   - x10
                       226 29651 5719.0
 - x10 1
- x8 1
- x1 1
- x14 1
- x12 1
- x7 1
- x17 1
- x4 1
- x5 1
- x2 1
                      398 29823 5731.6
                      436 29860 5734.4
                      464 29888 5736.4
                      523 29947 5740.7
                      598 30023 5746.2
                     660 30085 5750.7
                       692 30116 5753.0
                     1190 30614 5788.8
  - x2 1 1606 31031 5818.3
- x13 1 39693 69117 7566.5
   > step model backward
   Call:
   lm(formula = y \sim x1 + x2 + x4 + x5 + x7 + x8 + x10 + x11 + x12 + x13 + x14 + x17, data = mlr_train)
```

Coefficients:				
(Intercept)	x1Developing	x 2	x4	x 5
57.68485	-1.51624	-0.93552	1.06293	0.19341
x 7	x8	x10	x11	x12
-0.07536	0.02837	0.66265	0.13963	0.99013
x13	x14	x17		
-3.44054	0.35469	-0.73823		

> best_subset_backward <-ols_step_best_subset(step_model_backward) > best_subset_backward

Best Subsets Regression

Model Index	Predictors
1	x13
2	x5 x13
3	x4 x5 x13
4	x4 x5 x13 x17
5	x2 x4 x5 x13 x17
6	x2 x4 x5 x12 x13 x17
7	x2 x4 x5 x7 x12 x13 x17
8	x2 x4 x5 x7 x12 x13 x14 x17
9	x1 x2 x4 x5 x7 x12 x13 x14 x17
10	x1 x2 x4 x5 x7 x8 x12 x13 x14 x17
11	x1 x2 x4 x5 x7 x8 x10 x12 x13 x14 x17
12	x1 x2 x4 x5 x7 x8 x10 x11 x12 x13 x14 x17

Subsets Regre:

		Adj.	Pred		
Model	R-Square	R-Square	R-Square	C(p)	AIC
1	0.6948	0.6947	0.6943	1981.6981	13300.3883
2	0.7628	0.7626	0.7623	1056.2471	12751.8446
3	0.7930	0.7927	0.7923	646.7618	12456.6826
4	0.8063	0.8059	0.8054	467.6757	12313.8917
5	0.8170	0.8166	0.8159	323.7232	12191.7727
6	0.8248	0.8243	0.8236	219.5964	12098.8736
7	0.8296	0.8291	0.8283	155.6542	12039.7598
8	0.8333	0.8327	0.8318	107.9746	11994.5607
9	0.8361	0.8354	0.8346	71.5662	11959.3575
10	0.8385	0.8378	0.8369	40.1563	11928.4721
11	0.8398	0.8390	0.838	24.5964	11912.9872
12	0.8408	0.8399	0.8389	13.0000	11901.3520

```
AIC: Akaike Information Criteria
SBIC: Sawa's Bayesian Information Criteria
SBC: Schwarz Bayesian Criteria
MSEP: Estimated error of prediction, assuming multivariate normality
FFE: Final Prediction Error
HSP: Hocking's Sp
APC: Amemiya Prediction Criteria
```

Here, the value of E(Cp) =13 corresponding the p(12), So, we select 12^{th} subset model. Also, the value of adj. R2 correspond to 14^{th} subset model is 0.8399 which is highest among all the subset models & AIC is minimum.

Output in Stepwise selection

```
step_model_both = stepAIC(fit ,direction="both",trace=1)
Start: AIC=5708.14 y \sim x1 + x2 + x4 + x5 + x6 + x7 + x8 + x10 + x11 + x12 + x13 + x14 + x15 + 
             x14 + x15 + x17
                       Df Sum of Sq RSS
                                                                                              ATC
                                                           0 29423 5706.2
- x6
 - x15 1
                                                         1 29424 5706.2
                                                                  29423 5708.1
<none>
                                                   183 29606 5719.7
- x11
- x10
                                                   226 29649 5722.9
                                                    400 29822 5735.6
- x8
- x1
                                                   437 29860 5738.3
- x12
                                                    449 29872 5739.2
                                                    465 29888 5740.4
- x14
                                                    563 29986 5747.5
- x17
                                                    652 30075 5754.0
- x4
                                                    688 30111 5756.6
                                                1176 30599 5791.7
 - x5
                                                  1605 31027 5822.1
- x13
                       1
                                            39445 68868 7562.6
Step: AIC=5706.16
y \sim x1 + x2 + x4 + x5 + x7 + x8 + x10 + x11 + x12 + x13 + x14 + x15 + x17
                      Df Sum of Sq RSS AIC
1 1 29425 5704.3
- x15
                                                                 29423 5706.2
<none>
                                                         0 29423 5708.1
- x11
                                                  183 29606 5717.7
                                                   226 29649 5720.9
- x10
                                                   399 29823 5733.6
- x1
                                                    437 29860 5736.3
- x14
                                                    465 29888 5738.4
                                                    524 29947 5742.7
- x12
                                                    564 29987 5745.6
- x17
                                                  659 30083 5752.5
                                                     688 30111 5754.6
                                                 1180 30603 5790.0
- x2
                                                 1606 31029 5820.2
```

39487 68910 7561.9

- x13

```
Step: AIC=5704.27
y ~ x1 + x2 + x4 + x5 + x7 + x8 + x10 + x11 + x12 + x13 + x14 +
    x17
       Df Sum of Sq RSS AIC 29425 5704.3
<none>
+ x15
                  1 29423 5706.2
+ x6
                  0 29424 5706.2
                184 29609 5715.9
- x11
                226 29651 5719.0
- x8
                398 29823 5731.6
- x1
               436 29860 5734.4
- x14
               464 29888 5736.4
- x12
               523 29947 5740.7
- x7
               598 30023 5746.2
- x17
               660 30085 5750.7
                692 30116 5753.0
- x5
              1190 30614 5788.8
- x2
               1606 31031 5818.3
- x13 1
             39693 69117 7566.5
> step model both
Call:
```

```
lm(formula = y \sim x1 + x2 + x4 + x5 + x7 + x8 + x10 + x11 + x12 +
   x13 + x14 + x17, data = mlr train)
Coefficients:
 (Intercept) x1Developing
               -1.51624
                                -0.93552
   57.68485
                                               1.06293
                                                             0.19341
                                 x10
0.66265
    -0.07536
                   0.02837
                                               0.13963
                                                             0.99013
                                -0.73823
   -3.44054
                  0.35469
```

> best_subset_both <-ols_step_best_subset(step_model_both) > best_subset_both

Best Subsets Regression

Model Index	Predictors
1	x13
2	x5 x13
3	x4 x5 x13
4	x4 x5 x13 x17
5	x2 x4 x5 x13 x17
6	x2 x4 x5 x12 x13 x17
7	x2 x4 x5 x7 x12 x13 x17
8	x2 x4 x5 x7 x12 x13 x14 x17
9	x1 x2 x4 x5 x7 x12 x13 x14 x17
10	x1 x2 x4 x5 x7 x8 x12 x13 x14 x17
11	x1 x2 x4 x5 x7 x8 x10 x12 x13 x14 x17
12	x1 x2 x4 x5 x7 x8 x10 x11 x12 x13 x14 x17

Subsets Regre

Model	R-Square	Adj. R-Square	Pred R-Square	C (p)	AIC
				- 12-7	
1	0.6948	0.6947	0.6943	1981.6981	13300.3883
2	0.7628	0.7626	0.7623	1056.2471	12751.8446
3	0.7930	0.7927	0.7923	646.7618	12456.6826
4	0.8063	0.8059	0.8054	467.6757	12313.8917
5	0.8170	0.8166	0.8159	323.7232	12191.7727
6	0.8248	0.8243	0.8236	219.5964	12098.8736
7	0.8296	0.8291	0.8283	155.6542	12039.7598
8	0.8333	0.8327	0.8318	107.9746	11994.5607
9	0.8361	0.8354	0.8346	71.5662	11959.3575
10	0.8385	0.8378	0.8369	40.1563	11928.4721
11	0.8398	0.8390	0.838	24.5964	11912.9872
12	0.8408	0.8399	0.8389	13.0000	11901.3520

```
AIC: Akaike Information Criteria
SBIC: Sawa's Bayesian Information Criteria
SBC: Schwarz Bayesian Criteria
MSEP: Estimated error of prediction, assuming multivariate normality
FFE: Final Prediction Error
HSP: Hocking's Sp
APC: Amemiya Prediction Criteria
```

Here, the value of E(Cp) = 13 corresponding the p(12), So, we select 12^{th} subset model.

Also, the value of adj. R2 correspond to 14th subset model is 0.8399 which is highest among all the subset models & AIC is minimum.

Final Model

After, variable selection we lead to the final model which is:

```
y = (\beta 1 \times 1 + \beta 2 \times 2 + \beta 4 \times 4 + \beta 5 \times 5 + \beta 7 \times 7 + \beta 8 \times 8 + \beta 10 \times 10 + \beta 11 \times 11 + \beta 12 \times 12 + \beta 13 \times 13 + \beta 14 \times 14 + \beta 17 \times 17)
```

```
> fit1 <- lm(y ~x1 + x2 + x4 + x5 + x7 + x8 + x10 +
              x11 + x12 + x13 + x14 + x17, data=mlr_train)
> summary(fit1)
Call:
lm(formula = y \sim x1 + x2 + x4 + x5 + x7 + x8 + x10 + x11 + x12 +
    x13 + x14 + x17, data = mlr train)
   Min
             1Q Median
                             30
                                    Max
min 10 median 30 max
-12.986 -2.226 -0.027 2.326 11.484
Coefficients:
              Estimate Std. Error t value Pr(>|t|)
(Intercept) 57.684851
                         1.063049
                                   54.264
                         0.267458 -5.669 1.63e-08 ***
x1Developing -1.516242
             -0.935517
                         0.085957 -10.884
x2
              1.062930
                         0.148813
                                    7.143 1.24e-12 ***
                                           < 2e-16 ***
×5
              0.193408
                         0.020648
                                    9.367
             -0.075356
x7
                         0.011346
                                   -6.642 3.91e-11 ***
              0.028366
                         0.005236
                                    5.418 6.70e-08 ***
                                    4.085 4.56e-05 ***
x10
              0.662648
                         0.162202
                                    3.687 0.000232 ***
x11
              0.139625
                         0.037866
              0.990132
                         0.159480
                                     6.209 6.39e-10 ***
x12
                                           < 2e-16 ***
                         0.063591 -54.104
x13
             -3.440537
              0.354689
                         0.060643
                                   5.849 5.70e-09
x14
x17
             -0.738233
                         0.105801 -6.978 3.97e-12 ***
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 3.682 on 2170 degrees of freedom
Multiple R-squared: 0.8408,
                                 Adjusted R-squared: 0.8399
F-statistic: 955.2 on 12 and 2170 DF, p-value: < 2.2e-16
```

Test for Normality

Shapiro - Wilk Test

The Shapiro – Wilk test is a test of normality of a random sample.

HO: Sample comes from normally distributed population

HA: Sample does not come from normal population Test Statistic:

$$W = \frac{\left(\sum_{i=1}^{n} a_{i} x_{(i)}\right)^{2}}{\sum_{i=1}^{n} (x_{i} - \overline{x})^{2}}$$

Where, xi's are the ordered random sample values. ai's are the constant generated from the covariance, variances and means of the sample (size n) from a normally distributed sample.

Decision Rule:

We reject Ho if W < W α ,at α % level of significance otherwise accept Ho

Normal Q-Q Plot Segretary of the second of

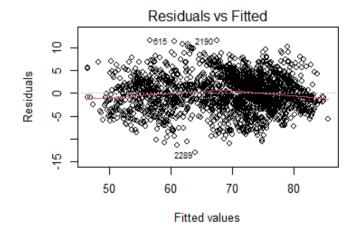
Since, p-value < 0.05 thus, we reject the null hypothesis at 5% level of significance. Hence, data is non-normal.

W = 0.94986, p-value < 2.2e-16

Homoscedasticity

H0: the residual is homoscedastic H1: the residual is heteroscedastic **Residual Plot:**

data: mlr train\$y



- > #Breusch-Pagan test
- > bptest(fit1)

studentized Breusch-Pagan test

```
data: fit1
BP = 59.712, df = 12, p-value = 2.548e-08
```

Since, p-value < 0.01 thus, we reject the null hypothesis at 1% level of significance. Hence, Training data is **Heteroskedastic.**

Auto correlation

H0: there is no serial correlation H1: there is serial correlation

```
> dwt(fit1)
lag Autocorrelation D-W Statistic p-value
1  -0.01266898  2.021858  0.552
Alternative hypothesis: rho != 0
```

Since, p-value> 0.01 thus, we fail to reject the null hypothesis at 1% level of significance. Hence, there is no serial autocorrelation.

Model Performance

It is important to appropriately estimate the prediction error of a model, since (a) it provides insight into its accuracy; (b) it allows comparison of different models, and (c) it is used to define warning thresholds. In order to facilitate the analysis of final calculation results, different performance evaluation functions are adopted in this paper, that is, mean square error (MSE), maximum absolute percentage error (MAPE).

The mean absolute percentage error (MAPE) is a statistical measure of how accurate a forecast system is. It measures this accuracy as a percentage,

and can be calculated as the average absolute percent error for each time period minus actual values divided by actual values. Where A_t is the actual value and F_t is the forecast value, this is given by:

$$\mathrm{M} = rac{1}{n} \sum_{t=1}^n \left| rac{A_t - F_t}{A_t}
ight|$$

The mean squared error (MSE) tells you how close a regression line is to a set of points. It does this by taking the distances from the points to the regression line (these distances are the "errors") and squaring them. The squaring is necessary to remove any negative signs. It also gives more weight to larger differences. It's called the mean squared error as you're finding the average of a set of errors.

```
> MSE_train
[1] 13.47896
> MSE_test
[1] 14.57727
> train_Mape
[1] 4.304494
> test_Mape
[1] 4.458779
> accuracy_train
[1] 99.68379
> accuracy_test
[1] 99.94273
```

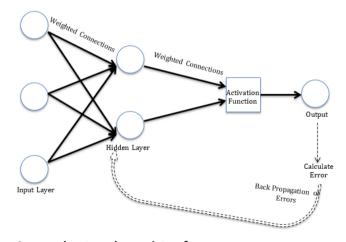
Multiple Linear regression Equation

```
 (\text{Life\_expectancy}) = 57.684851 + (-1.516242)*(\text{Status}) + (-0.935517)*(\text{Adult\_Mortality}) + (1.062930)*(\text{Alcohol}) + (0.193408)*(\text{percentage\_expenditure}) + (-0.075356)*(\text{Measles}) + (0.028366)*(\text{BMI}) + (0.662648)*(\text{Polio}) + (0.139625)*(\text{Total\_expenditure}) + (0.990132)*(\text{Diphtheria}) + (-3.440537)*(\text{HIV\_AIDS}) + (0.354689)*(\text{GDP}) + (-0.738233) *(\text{thinness\_5\_9\_years})
```

Even though MLR performs extremely well in this data for model Building and Prediction, but it violates homoscadasticity and normality assumptions. This encourages us to go for another algorithm that does not take into consideration normality.

[B] ARTIFICIAL NEURAL NETWORK

It has the ability to learn by examples. ANN is an information processing model inspired by the biological neuron system. It is composed of a large number of highly interconnected processing elements known as the neuron to solve problems. It follows the non-linear path and process information in parallel throughout the nodes. A neural network is a complex adaptive system. Adaptive means it has the ability to change its internal structure by adjusting weights of inputs.



A neural network consists of:

- 1. *Input layers:* Layers that take inputs based on existing data. Here we have 7 input variables.
- 2. *Hidden layers:* Layers that use back propagation to optimize the weights of the input variables in order to improve the predictive power of the model.

3. *Output layers:* Output of predictions based on the data from the input and hidden layers.

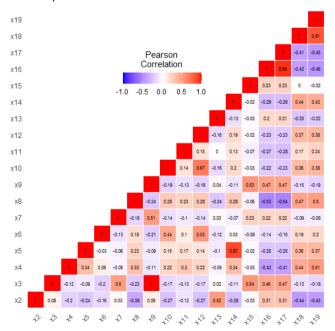
Before fitting a neural network, some preparation needs to be done. Neural networks are not that easy to train and tune. As a *first step*, we are going to address data preprocessing. In this case also we removed unnecessary columns, renamed them, and imputed missing values by interpolation.

Data Normalization

One of the most important procedures when forming a neural network is data normalization. This involves adjusting the data to a common scale so as to accurately compare predicted and actual values. Failure to normalize the data will typically result in the prediction value remaining the same across all observations, regardless of the input values. Most of the times the algorithm will not converge before the number of maximum iterations allowed. You can choose different methods to scale the data (z-normalization, min-max scale, etc...). I chose to use the min-max method and scale the data in the interval [0, 1]

Heatmap & correlation Matrix

A good way to quickly check correlations among columns is by visualizing the correlation matrix as a heatmap.



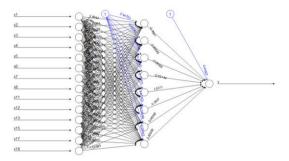
We see there is high correlation between x18-x19, x16-x17,x5-x14, x10-x12 & x3-x9. So we have removed x19, x16, x10, x14 & x9 from the data frame to avoid over fitting.

After this we have divided the whole dataset in training and test set randomly in the ratio 80:20. Then we have deployed MLR on the training dataset.

Parameter Setting

There is no fixed rule as to how many layers and neurons to use although there are several more or less accepted rules of thumb. Usually, if at all necessary, one hidden layer is enough for a vast numbers of applications. As far as the number of neurons is concerned, it should be between the input layer size and the output layer size, usually 2/3 of the input size. Here we have 1 response and 14 regressors so we used 8 neurons in the only hidden layer. The configuration of our neural network is (14:8:1)

This is the graphical representation of the model with the weights on each connection:



Predicting using the neural network

Now we can try to predict the values for the both train & test set and calculate the MSE, MAPE & Accuracy. Remember that the net will output a normalized prediction, so we need to scale it back in order to make a meaningful comparison (or just a simple prediction).

```
> MSE.nn1_train
[1] 4.684468
> MSE.nn1_test
[1] 7.257011
> train_Mape
[1] 2.323643
> test_Mape
[1] 2.882731
> accuracy_train
[1] 99.88873
> accuracy_test
[1] 99.78785
```

VIII. PERFORMACE OF THE ALGORITHMS

To compare the performance of these two algorithms we will use MSE, MAPE & accuracy of the both models on both training data and test data.

Statistical performance of the MLR and ANN

	М	LR	ANN	
	Training Testing		Training	Testing
MSE	13.47896	14.57727	4.684468	7.257011
MAPE	4.304494	4.458779	2.323643	2.882731
Accuracy	99.68379	99.94273	99.88873	99.78785

IX. CONCLUSION

In both the models we can see that all the Regressors are not contributing in predicting Life Expectancy. We had to drop few Regressors due to multicollinearity.

MLR model indicates that Alcohol, BMI, Polio, Total_expenditure, Diphtheria, GDP and percentage_expenditure have positive impact on Life Expectancy. On the other hand Measles, HIV_AIDS and thinness_5_9_years have negative impact on Life Expectancy.It is evident that immunization coverage improves life expectancy.

Both the models have performed exceptionally in predicting Life Expectancy. But neural networks usually outperform linear regression as they deal with non-linearity automatically with large number of inputs, whereas in linear regression you need to mention explicitly. Here also MAPE and MSE suggest that neural network has performed better than MLR in Prediction Life Expectancy.

FUTURE WORK

- There are many scopes in future with this data.
 We can run cross validation models to check if we can improve results.
- We can include the year column to do time series analysis, but for that we need more data.
- We can deploy other algorithms such as k means or decision tree according to our requirements.
- We can divide the whole dataset into two parts.
 One for developed country, another for developing country, and then we can again run MLR on that.

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#Pie Chart

```
setwd("C:\\Users\\user\\Desktop\\GD project")
data<- read.csv("Life Expectancy Data.csv")
data \leftarrow data[c(3,4)]
data$Status <- as.factor(data$Status)</pre>
data1 <- data[data$Status == "Developed", ]</pre>
data2 <- data[data$Status == "Developing",]</pre>
a<-nrow(data1)/(nrow(data1)+nrow(data2))*100
b<-nrow(data2)/(nrow(data1)+nrow(data2))*100
Developed LE <- data1[ 2 ]
Developing LE <- data2[ 2 ]</pre>
# Create data for the graph.
x < -c(a, b)
labels <- c("Developed", "Developing")</pre>
# Plot the chart with title and rainbow color pallet.
pie(x, labels, main = "Status wise data Share",
    col = rainbow(length(x)))
#Use new R-script
#Histogram & Boxplot
library(boot)
library(car)
library(lmtest)
library (MASS)
library(ggplot2)
library(questionr)
library(e1071)
library(reshape2)
#load data
setwd("C:\\Users\\user\\Desktop\\GD project")
data<- read.csv("Life Expectancy Data.csv")</pre>
df < - data[ -c(1,2) ]
df \leftarrow df[,c(2,1,3:19,20)]
df$Status<-as.factor(df$Status)</pre>
df<-rename.variable(df,"Life.expectancy","Life expectancy")</pre>
df<-rename.variable(df,"Adult.Mortality","Adult Mortality")
df<-rename.variable(df,"infant.deaths","infant deaths")</pre>
df<-rename.variable(df,"percentage.expenditure","percentage expenditure")</pre>
```

df<-rename.variable(df,"Hepatitis.B","Hepatitis_B")</pre>

```
df<-rename.variable(df, "under.five.deaths", "under five deaths")
df<-rename.variable(df,"Total.expenditure","Total expenditure")</pre>
df<-rename.variable(df,"HIV.AIDS","HIV AIDS")</pre>
df<-rename.variable(df,"thinness.5.9.years","thinness 5 9 years")
df<-rename.variable(df,"thinness..1.19.years","thinness 10 19 years")</pre>
rename.variable(df, "Income.composition.of.resources", "Income composition of res
ources")
#comparison of boxplots of life expectancy due to status
plot(df$Life expectancy,, main, xlab, ylab, xlim, ylim, axes)
boxplot(df$Life expectancy ~ df$Status, data = df, main = "Comparison of
Boxplots")
#Histogram and boxplot of regressors
par(mfrow=c(1,2))
hist(df$Life expectancy)
boxplot(df$Life expectancy)
hist(df$Life expectancy)
boxplot(df$Adult Mortality)
hist(df$infant deaths)
boxplot(df$infant deaths)
hist(df$Alcohol)
boxplot(df$Alcohol)
hist(df$percentage expenditure)
boxplot(df$percentage expenditure)
hist(df$Hepatitis B)
boxplot(df$Hepatitis B)
hist(df$Measles)
boxplot(df$Measles)
hist(df$BMI)
boxplot(df$BMI)
hist(df$under five deaths)
boxplot(df$under_five_deaths)
hist (df$Polio)
boxplot(df$Polio)
```

hist(df\$Total_expenditure)
boxplot(df\$Total expenditure)

hist(df\$Diphtheria)
boxplot(df\$Diphtheria)

```
hist(df$HIV AIDS )
boxplot(df$HIV AIDS )
hist (df$GDP)
boxplot (df$GDP)
hist (df$Population)
boxplot(df$Population)
hist (df$thinness 10 19 years)
boxplot(df$thinness_10_19_years)
hist(df$thinness 5 9 years)
boxplot(df$thinness 5 9 years)
hist(df$Income composition of resources)
boxplot(df$Income composition of resources)
hist (df$Schooling)
boxplot (df$Schooling)
#Use R studio
#Scatterplot
install.packages("gridExtra", dependencies=T)
library(gridExtra)
library(ggplot2)
library(questionr)
setwd("C:\\Users\\user\\Desktop\\GD project")
data<- read.csv("Life Expectancy Data.csv")
df < - data[ -c(1,2) ]
data \leftarrow df[,c(2,1,3:19,20)]
data$Status<-as.factor(df$Status)</pre>
data<-na.omit(data)</pre>
str(data)
data <- rename.variable (data, "Life.expectancy", "Life expectancy")
data<-rename.variable(data, "Adult.Mortality", "Adult Mortality")
data <- rename.variable (data, "infant.deaths", "infant deaths")
data <- rename.variable (data, "percentage.expenditure", "percentage expenditure")
data<-rename.variable(data, "Hepatitis.B", "Hepatitis B")
data <- rename.variable (data, "under.five.deaths", "under five deaths")
data<-rename.variable(data, "Total.expenditure", "Total expenditure")
data<-rename.variable(data,"HIV.AIDS","HIV AIDS")</pre>
data<-rename.variable(data, "thinness.5.9.years", "thinness 5 9 years")
data<-rename.variable(data, "thinness..1.19.years", "thinness 10 19 years")
rename.variable(data, "Income.composition.of.resources", "Income_composition_of_r
esources")
```

```
g1<-ggplot(data, aes(x = Adult Mortality, y = Life_expectancy)) +
  geom_point(aes(color = factor(Status ))) + theme(legend.position="none")
g2 <- ggplot(data, aes(x = infant_deaths, y = Life_expectancy)) +</pre>
 geom point(aes(color = factor(Status ))) + theme(legend.position="none")
g3 < -ggplot(data, aes(x = Alcohol, y = Life expectancy)) +
  geom_point(aes(color = factor(Status ))) + theme(legend.position="none")
g4 <- ggplot(data, aes(x = percentage_expenditure, y = Life_expectancy)) +
 geom_point(aes(color = factor( Status ))) + theme(legend.position="none")
g5 \leftarrow ggplot(data, aes(x = Hepatitis B, y = Life expectancy)) +
 geom_point(aes(color = factor( Status ))) + theme(legend.position="none")
g6 <- ggplot(data, aes(x = Measles, y = Life_expectancy)) +</pre>
 geom point(aes(color = factor(Status ))) + theme(legend.position="none")
g7 \leftarrow ggplot(data, aes(x = BMI, y = Life_expectancy)) +
 geom_point(aes(color = factor( Status ))) + theme(legend.position="none")
g8<-ggplot(data, aes(x = under_five_deaths, y = Life_expectancy)) +
  geom_point(aes(color = factor(Status ))) + theme(legend.position="none")
g9 < -ggplot(data, aes(x = Polio, y = Life_expectancy)) +
  geom_point(aes(color = factor( Status ))) + theme(legend.position="none")
g10 < -ggplot(data, aes(x = Total_expenditure, y = Life_expectancy)) +
  geom point(aes(color = factor( Status ))) + theme(legend.position="none")
g11 < -ggplot(data, aes(x = Diphtheria, y = Life_expectancy)) +
  geom_point(aes(color = factor( Status ))) + theme(legend.position="none")
g12 < -ggplot(data, aes(x = HIV_AIDS, y = Life_expectancy)) +
  geom_point(aes(color = factor( Status ))) + theme(legend.position="none")
g13 < -ggplot(data, aes(x = GDP, y = Life_expectancy)) +
```

```
geom_point(aes(color = factor( Status ))) + theme(legend.position="none")
g14<-ggplot(data, aes(x = Population, y = Life_expectancy)) +
    geom_point(aes(color = factor( Status ))) + theme(legend.position="none")
g15<-ggplot(data, aes(x = thinness_10_19_years, y = Life_expectancy)) +
    geom_point(aes(color = factor( Status ))) + theme(legend.position="none")
g16<-ggplot(data, aes(x = thinness_5_9_years, y = Life_expectancy)) +
    geom_point(aes(color = factor( Status ))) + theme(legend.position="none")
g17<-ggplot(data, aes(x = Income_composition_of_resources, y =
    Life_expectancy)) +
    geom_point(aes(color = factor( Status ))) + theme(legend.position="none")
g18<-ggplot(data, aes(x = Schooling, y = Life_expectancy)) +
    geom_point(aes(color = factor(Status ))) + theme(legend.position="none")
grid.arrange(g1, g2,g3,g4,g5,g6,g7,g8,g9, nrow = 3)
grid.arrange(g1,g12,g13,g14,g15,g16,g17,g18, nrow = 3)</pre>
```

R CODE FOR MULTIPLE LINEAR REGRESSION #### #### USE A NEW R-SCRIPT

#Installing Required packages

```
install.packages("questionr", dependencies=TRUE)
install.packages("dplyr", dependencies=TRUE)
install.packages("car", dependencies=TRUE)
install.packages("boot", dependencies=TRUE)
install.packages("QuantPsyc", dependencies=TRUE)
install.packages("lmtest", dependencies=TRUE)
install.packages("sandwich", dependencies=TRUE)
install.packages("vars", dependencies=TRUE)
install.packages("MASS", dependencies=TRUE)
install.packages("nortest", dependencies=TRUE)
install.packages("ggplot2", dependencies=TRUE)
install.packages("imputeTS", dependencies=TRUE)
install.packages("perturb", dependencies=TRUE)
install.packages("leaps", dependencies=TRUE)
install.packages("olsrr", dependencies=TRUE)
install.packages("e1071", dependencies=TRUE)
install.packages("reshape2", dependencies=TRUE)
```

Calling the liabraries

```
library(boot)
library(car)
library(QuantPsyc)
library(lmtest)
library(sandwich)
library(vars)
library(nortest)
library (MASS)
library(ggplot2)
library(imputeTS)
library(perturb)
library(leaps)
library(olsrr)
library(e1071)
library(reshape2)
library(questionr)
```

#load data

```
setwd("C:\\Users\\user\\Desktop\\GD project")
data<- read.csv("Life_Expectancy_Data.csv")
str(data)</pre>
```

Dropping year and country column

```
data \leftarrow data [-c(1,2)]
```

```
data <- data[,c(2,1,3:19,20)]
data$Status<-as.factor(data$Status)</pre>
```

structure and summary of dataset

```
str(data)
summary(data)
```

#Renaming the columns

```
colnames(data)
data<-rename.variable(data,"Life.expectancy","Life_expectancy")
data<-rename.variable(data,"Adult.Mortality","Adult_Mortality")
data<-rename.variable(data,"infant.deaths","infant_deaths")
data<-rename.variable(data,"percentage.expenditure","percentage_expenditure")
data<-rename.variable(data,"Hepatitis.B","Hepatitis_B")
data<-rename.variable(data,"under.five.deaths","under_five_deaths")
data<-rename.variable(data,"Total.expenditure","Total_expenditure")
data<-rename.variable(data,"HIV.AIDS","HIV_AIDS")
data<-rename.variable(data,"thinness.5.9.years","thinness_5_9_years")
data<-rename.variable(data,"thinness.1.19.years","thinness_10_19_years")
data<-
rename.variable(data,"Income.composition.of.resources","Income_composition_of_resources")</pre>
```

#Checking for missing values and data imputation by interpolation

sapply(data, function(x) sum(is.na(x))/length(x)*100)

```
data$Adult_Mortality <- na_interpolation(data$Adult_Mortality)
data$Alcohol<-na_interpolation(data$Alcohol)
data$Hepatitis_B<-na_interpolation(data$Hepatitis_B)
data$BMI<-na_interpolation(data$BMI)
data$Polio<-na_interpolation(data$Polio)
data$Total_expenditure<-na_interpolation(data$Total_expenditure)
data$Diphtheria<-na_interpolation(data$Diphtheria)
data$GDP<-na_interpolation(data$GDP)
data$Population<-na_interpolation(data$Population)
data$thinness_10_19_years<-na_interpolation(data$thinness_10_19_years)
data$thinness_5_9_years<-na_interpolation(data$thinness_5_9_years)
data$Income_composition_of_resources<-
na_interpolation(data$Income_composition_of_resources)
data$Schooling<-na_interpolation(data$Schooling)
```

#omitting Null rows

```
data <- na.omit(data)
str(data)
summary(data)</pre>
```

#Checking Skewness

```
skewness(data$Life_expectancy)
skewness(data$thinness_5_9_years)
skewness(data$thinness_10_19_years)
skewness(data$Population)
```

```
skewness(data$Income_composition_of_resources)
skewness(data$GDP)
skewness(data$HIV_AIDS)
skewness(data$Diphtheria)
skewness(data$Total_expenditure)
skewness(data$Polio)
skewness(data$Polio)
skewness(data$MEI)
skewness(data$MEI)
skewness(data$Measles)
skewness(data$Hepatitis_B)
skewness(data$Polio)
skewness(data$Polio)
skewness(data$Measles)
skewness(data$Measles)
skewness(data$Polio)
skewness(data$Polio)
skewness(data$Polio)
skewness(data$Polio)
skewness(data$Polio)
skewness(data$Polio)
```

#Response Variable

y <- data\$Life expectancy

#Regressor Variables

- x1 <- data\$Status
- x2 <- data\$Adult Mortality
- x3 <- data\$infant deaths
- x4 <- data\$Alcohol
- x5 <- data\$percentage expenditure
- x6 <- data\$Hepatitis_B
- x7 <- data\$Measles
- x8 <- data\$BMI
- x9 <- data\$under five deaths
- x10 <- data\$Polio
- x11 <- data\$Total expenditure
- x12 <- data\$Diphtheria
- x13 <- data\$HIV AIDS
- x14 <- data\$GDP
- x15 <- data\$Population
- x16 <- data\$thinness_10_19_years</pre>
- x17 <- data\$thinness 5 9 years
- x18 <- data\$Income composition of resources</pre>
- x19 <- data\$Schooling

Data Normalization/Standardization

```
y <- data$Life_expectancy
x1 <- data$Status
x2 <- log(data$Adult_Mortality)
x3 <- (data$infant_deaths)^(1/3)
x4 <- data$Alcohol^(1/3)
x5 <- (data$percentage_expenditure)^(1/3)
x6 <- log(data$Hepatitis_B)
x7 <- (data$Measles)^(1/3)
x8 <- data$BMI
x9 <- (data$under_five_deaths)^(1/3)
x10 <- log(data$Polio)
x11 <- data$Total expenditure
```

```
x12 <- log(data$Diphtheria)
x13 <- log(data$HIV AIDS)
x14 <- log(data$GDP)</pre>
x15 <- log(data$Population)</pre>
x16 <- log(data$thinness 10 19 years)
x17 <- log(data$thinness 5 9 years)</pre>
x18 <- data$Income composition of resources
x19 <- data$Schooling
# Creating new dataframe
data1 <- data.frame(y, x1, x2, x3, x4, x5, x6, x7, x8, x9, x10,
x11, x12, x13, x14, x15, x16, x17, x18, x19
head (data1)
nrow(data1)
str(data1)
summary(data1)
#Heatmap & correlation Matrix to visualize Multicolinearity
df2 < - data1[ -c(1,2) ]
cormat <- round(cor(df2),2)</pre>
melted cormat <- melt(cormat)</pre>
head(melted cormat)
# Get upper triangle of the correlation matrix
get upper tri <- function(cormat) {</pre>
    cormat[lower.tri(cormat)]<- NA</pre>
    return (cormat)
  }
upper tri <- get upper tri(cormat)</pre>
# Melt the correlation matrix
library(reshape2)
melted cormat <- melt(upper_tri, na.rm = TRUE)</pre>
# Heatmap
library(ggplot2)
ggheatmap <- ggplot(data = melted_cormat, aes(Var2, Var1, fill = value))+</pre>
geom tile(color = "white")+
scale fill gradient2(low = "blue", high = "red", mid = "white",
   midpoint = 0, limit = c(-1,1), space = "Lab",
   name="Pearson\nCorrelation") +
  theme minimal()+
 theme (axis.text.x = element text(angle = 45, vjust = 1,
    size = 8, hjust = 1)) +
 coord fixed()
ggheatmap +
geom text(aes(Var2, Var1, label = value), color = "black", size = 2) +
theme (
  axis.title.x = element blank(),
  axis.title.y = element blank(),
```

```
panel.grid.major = element blank(),
  panel.border = element blank(),
  panel.background = element blank(),
  axis.ticks = element blank(),
  legend.justification = c(1, 0),
  legend.position = c(0.6, 0.7),
  legend.direction = "horizontal") +
  guides(fill = guide colorbar(barwidth = 7, barheight = 1,
                title.position = "top", title.hjust = 0.5))
#removing unnecessery variables
data1 <- data.frame(y, x1, x2, x3, x4, x5, x6, x7, x8, x10,
x11, x12, x13, x14, x15, x17, x18
#Outlier Detection (Cook's Distance)
sapply (data1, function (x) sum(is.na(x))/length(x)*100)
model < -lm(y \sim x1 + x2 + x3 + x4 + x5 + x6 + x7 + x8 + x10 +
x11 + x12 + x13 + x14 + x15 + x17 + x18, data=data1)
cooksd <- cooks.distance(model)</pre>
# Plot the Cook's Distance using the traditional 4/(n=sample zize) criterion
sample size <- nrow(data1)</pre>
# plot cook's distance
plot(cooksd, pch="*", cex=1, main="Influential Obs by Cooks distance")
# add cutoff line
abline(h = (4/(sample size)), col="red")
# Removing Outliers
# influential row numbers
influential <- as.numeric(names(cooksd)[(cooksd > (4/(sample size-19-1)))])
data <- data1[-influential, ]</pre>
# Training and Test Data
index <- sample(1:nrow(data), round(0.8*nrow(data)))</pre>
mlr train <- data [index,]</pre>
mlr test <- data [-index,]</pre>
head(mlr train)
nrow(mlr train)
# Fitting Multiple Linear Regression Model on training data
fit < - lm(y \sim x1 + x2 + x3 + x4 + x5 + x6 + x7 + x8 + x10 +
            x11 + x12 + x13 + x14 + x15 + x17 + x18, data=mlr train)
fit <-lm(y ~x1 + x2 + x4 + x5 + x6 + x7 + x8 + x10 +
            x11 + x12 + x13 + x14 + x15 + x17, data=mlr train)
```

```
# Summary of Model
#P value of independent variable (p value should be less than .05)
summary(fit)
#Again Checking Multicolinearity vif>2 means presence of multicollinearity
vif(fit)
#Stepwise regression
fit <-lm(y \sim x1 + x2 + x4 + x5 + x6 + x7 + x8 + x10 +
            x11 + x12 + x13 + x14 + x15 + x17, data=mlr train)
summary(fit )
#Forward Selection
step model forward = stepAIC(fit ,direction="forward",trace=1)
step model forward
best subset forward <-ols step best subset(step model forward)
best subset forward
#Backward Elimination
step model backward = stepAIC(fit ,direction="backward",trace=1)
step model backward
best subset backward <-ols step best subset(step model backward)
best subset backward
#Stepwise Regression
step model both = stepAIC(fit ,direction="both",trace=1)
step model both
best subset both <-ols step best subset(step model both)</pre>
best subset both
#Final MOdel
fit1 <- lm(y \sim x1 + x2 + x4 + x5 + x7 + x8 + x10 +
            x11 + x12 + x13 + x14 + x17, data=mlr train)
summary(fit1)
# Test for Normality
qqnorm(mlr train$y)
qqline(mlr_train$y,col="red")
shapiro.test(mlr train$y)
ad.test(mlr train$y)
#Breusch-Pagan test
par(mfrow = c(2,2))
plot(fit1)
bptest(fit1)
#Autocorrelation
dwt(fit1)
```

Get the predicted or fitted values on training data

train pred<-fitted(fit1)</pre>

#Validation on train set

attach(mlr train)

train_Mape<-(sum((abs(mlr_train\$ytrain_pred))/mlr_train\$y))/nrow(mlr_train)*100
MSE_train <- sum((train_pred - mlr_train\$y)^2)/nrow(mlr_train)
deviation_train=((mlr_train\$y-train_pred)/mlr_train\$y)
accuracy train=(1-abs(mean(deviation train)))*100</pre>

Get the predicted or fitted values on training data

test pred <- predict(fit1,mlr test)</pre>

#Validation on test set

attach(mlr test)

test_Mape<-(sum((abs(mlr_test\$y-test_pred))/mlr_test\$y))/nrow(mlr_test)*100
MSE_test <- sum((test_pred - mlr_test\$y)^2)/nrow(mlr_test)
deviation_test=((mlr_test\$y-test_pred)/mlr_test\$y)
accuracy_test=(1-abs(mean(deviation_test)))*100</pre>

MSE_train
MSE_test
train_Mape
test_Mape
accuracy_train
accuracy_test

R CODE FOR ARTIFICIAL NEURAL NETWORK #### #### USE A NEW R-SCRIPT

```
install.packages("car",dependencies=TRUE)
install.packages("boot",dependencies=TRUE)
install.packages("QuantPsyc",dependencies=TRUE)
install.packages("lmtest",dependencies=TRUE)
install.packages("sandwich",dependencies=TRUE)
install.packages("vars",dependencies=TRUE)
install.packages("MASS",dependencies=TRUE)
install.packages("nortest",dependencies=TRUE)
install.packages("ggplot2",dependencies=TRUE)
install.packages("imputeTS",dependencies=TRUE)
install.packages("perturb",dependencies=TRUE)
install.packages("leaps",dependencies=TRUE)
install.packages("olsrr",dependencies=TRUE)
install.packages("el071",dependencies=TRUE)
install.packages("reshape2",dependencies=TRUE)
```

#Required Libraries

library(boot)
library(car)
library(QuantPsyc)
library(lmtest)
library(sandwich)
library(vars)
library(mortest)
library(MASS)
library(ggplot2)
library(imputeTS)
library(perturb)
library(leaps)
library(olsrr)
library(e1071)
library(reshape2)

#load data

```
setwd("C:\\Users\\user\\Desktop\\GD project")
data<- read.csv("Life_Expectancy_Data.csv")
str(data)

summary(data)

data <- data[ -c(1,2) ]
data <- data[,c(2,1,3:19,20)]
data$Status<-as.factor(data$Status)
data$Status <- ifelse(data$Status == "Developing",0,1)</pre>
```

#Renaming the columns

colnames (data)

```
data <- rename.variable (data, "Life.expectancy", "Life expectancy")
data<-rename.variable(data,"Adult.Mortality","Adult Mortality")
data<-rename.variable(data,"infant.deaths","infant deaths")</pre>
data <- rename.variable (data, "percentage.expenditure", "percentage expenditure")
data<-rename.variable(data, "Hepatitis.B", "Hepatitis B")
data<-rename.variable(data, "under.five.deaths", "under five deaths")
data<-rename.variable(data, "Total.expenditure", "Total expenditure")
data<-rename.variable(data,"HIV.AIDS","HIV AIDS")</pre>
data<-rename.variable(data, "thinness.5.9.years", "thinness 5 9 years")
data <- rename.variable (data, "thinness..1.19.years", "thinness 10 19 years")
data<-
rename.variable(data, "Income.composition.of.resources", "Income composition of r
esources")
# Checking for missing values and data imputation by interpolation
sapply(data, function(x) sum(is.na(x))/length(x)*100)
data$Adult Mortality <- na interpolation(data$Adult Mortality )</pre>
data$Alcohol<-na interpolation(data$Alcohol)</pre>
data$Hepatitis B<-na interpolation(data$Hepatitis B)</pre>
data$BMI<-na interpolation(data$BMI)</pre>
data$Polio<-na interpolation(data$Polio)</pre>
data$Total expenditure<-na interpolation(data$Total expenditure)
data$Diphtheria<-na interpolation(data$Diphtheria)</pre>
data$GDP<-na interpolation(data$GDP)</pre>
data$Population<-na interpolation(data$Population)
data$thinness 10 19 years<-na interpolation(data$thinness 10 19 years)
data$thinness 5 9 years<-na interpolation(data$thinness 5 9 years)
data$Income composition of resources<-
na interpolation(data$Income composition of resources)
data$Schooling<-na interpolation(data$Schooling)</pre>
# omitting Null rows
data <- na.omit(data)</pre>
sapply (data, function (x) sum (is.na(x)) /length (x) *100)
str(data)
summary (data)
#Response Variable
y <- data$Life expectancy
#Regressor Variables
x1 <- data$Status
x2 <- data$Adult Mortality
x3 <- data$infant deaths
x4 <- data$Alcohol
x5 <- data$percentage expenditure
x6 <- data$Hepatitis B
x7 <- data$Measles
x8 <- data$BMI
x9 <- data$under five deaths
```

```
x10 <- data$Polio
x11 <- data$Total expenditure
x12 <- data$Diphtheria
x13 <- data$HIV AIDS
x14 <- data$GDP
x15 <- data$Population
x16 <- data$thinness 10 19 years
x17 <- data$thinness 5 9 years
x18 <- data$Income composition of resources
x19 <- data$Schooling
data \leftarrow data.frame(y,x1,x2,x3,x4,x5,x6,x7,x8,x9,x10,
x11, x12, x13, x14, x15, x16, x17, x18, x19
#Scaling dataset
maxs <- apply(data, 2, max)</pre>
mins <- apply(data, 2, min)
scaled <- as.data.frame(scale(data, center = mins, scale = maxs - mins))</pre>
summary(scaled )
str(scaled)
scaled$x19 <- NULL
scaled$x16 <- NULL
scaled$x10 <- NULL
scaled$x9 <- NULL
scaled$x14 <- NULL
#Heatmap & correlation Matrix
df2 < - scaled [ -c(1,2) ]
cormat <- round(cor(df2),2)</pre>
melted cormat <- melt(cormat)</pre>
head(melted cormat)
# Get upper triangle of the correlation matrix
 get upper tri <- function(cormat) {</pre>
    cormat[lower.tri(cormat)]<- NA</pre>
    return (cormat)
upper tri <- get upper tri(cormat)</pre>
# Melt the correlation matrix
library(reshape2)
melted cormat <- melt(upper tri, na.rm = TRUE)</pre>
# Heatmap
library(ggplot2)
ggheatmap <- ggplot(data = melted_cormat, aes(Var2, Var1, fill = value))+</pre>
geom tile(color = "white")+
 scale fill gradient2(low = "blue", high = "red", mid = "white",
   midpoint = 0, limit = c(-1,1), space = "Lab",
```

```
name="Pearson\nCorrelation") +
  theme minimal()+
 theme(axis.text.x = element text(angle = 45, vjust = 1,
    size = 8, hjust = 1))+
 coord fixed()
qqheatmap +
geom text(aes(Var2, Var1, label = value), color = "black", size = 2) +
  axis.title.x = element blank(),
  axis.title.y = element blank(),
  panel.grid.major = element blank(),
  panel.border = element blank(),
  panel.background = element blank(),
  axis.ticks = element blank(),
  legend.justification = c(1, 0),
  legend.position = c(0.6, 0.7),
  legend.direction = "horizontal") +
  guides(fill = guide colorbar(barwidth = 7, barheight = 1,
                title.position = "top", title.hjust = 0.5))
sapply (scaled, function (x) sum (is.na(x)) /length (x)*100)
# Training and Test Data
index <- sample(1:nrow(scaled), round(0.8*nrow(scaled)))</pre>
ann train <- scaled[index,]</pre>
ann test <- scaled [-index,]</pre>
summary(ann train)
summary(ann test)
install.packages("neuralnet", dependencies=T)
library(neuralnet)
n <- names(ann train)</pre>
f \leftarrow as.formula(paste("y \sim", paste(n[!n %in% "y"], collapse = " + ")))
## model1
set.seed(123)
nn1 <- neuralnet(f,data=ann train,hidden=8,linear.output=T)</pre>
plot(nn1)
nn1$result.matrix
summary(nn1)
nrow(ann train)
b <- ann train[,2:15]</pre>
head(b)
pr.nn1 test <- compute(nn1,ann test[,2:15])</pre>
pr.nn1 test <- pr.nn1 test$net.result*(max(data$y)-min(data$y))+min(data$y)</pre>
```

```
test.r <- (ann_test$y) * (max(data$y) -min(data$y)) +min(data$y)
MSE.nn1_test <- sum((test.r - pr.nn1_test)^2) /nrow(ann_test)
test_Mape<- (sum((abs(test.r-pr.nn1_test)) / test.r)) /nrow(ann_test) *100

pr.nn1_train <- compute(nn1,b)
pr.nn1_train <- pr.nn1_train$net.result*(max(data$y) -min(data$y)) +min(data$y)
train.r <- (ann_train$y) * (max(data$y) -min(data$y)) +min(data$y)
MSE.nn1_train <- sum((train.r - pr.nn1_train)^2) /nrow(ann_train)
train Mape<- (sum((abs(train.r-pr.nn1_train))) / train.r)) /nrow(ann_train) *100</pre>
```

Accuracy

```
predicted_train=results_train$prediction
actual_train=results_train$actual
deviation_train=((actual_train-predicted_train)/actual_train)
comparison_train=data.frame(predicted_train,actual_train,deviation_train)
accuracy_train=(1-abs(mean(deviation_train)))*100

results_test <- data.frame(actual = test.r, prediction = pr.nn1_test)
results_test

predicted_test=results_test$prediction
actual_test=results_test$actual
deviation_test=((actual_test-predicted_test)/actual_test)
comparison_test=data.frame(predicted_test,actual_test,deviation_test)
accuracy_test=(1-abs(mean(deviation_test)))*100</pre>
```

MSE.nn1_train MSE.nn1_test train_Mape test_Mape accuracy_train accuracy_test