

# pcwOrd: (partial) (constrained) (weighted) ordination

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# 1 Motivation

Many ordination packages exist for ecologists using R - **vegan** is excellent, for example. But none readily perform (to my knowledge) weighted, partial constrained ordinations.

The motivation for this is to analyze compositional ecological data - especially high throughput sequencing data - with methods that are robust, reproducible, and transparent. Compositional data contains only relative data (think relative abundance), and so requires a bit of extra care to analyze. The dominant approach is to perform a log-ratio transformation, then use principle components analysis - combined, this is called log-ratio analysis. For an entrance into this literature, see Greenacre and Aitchison (2002), Greenacre and Lewi (2009), and Quinn et al (2018).

In particular, Greenacre and Lewi (2009) suggest using weighted log-ratio analysis, as unweighted log-ratio analysis is susceptible to noise in low-abundance features. Raw values of low-abundance features have higher relative variance than raw values of high-abundance features; therefore, the log-ratios of low abundance features are also of lower certainty. When low-abundance features dominate a solution the distances and inferences made about samples is less robust to error and noise. Note that correspondence analysis also is susceptible to dominance by low-abundance features.

Weighted log-ratio analysis down-weights these uncertain, low-abundance features. As a result, the observed distances among samples should be more reproducible across experiments.

**vegan** can perform constrained and partialled log-ratio analysis if the community matrix is log-transformed beforehand. However, it does not provide a straightforward way to weight columns and working with ordination objects is not intuitive. **easyCODA** allows weighted and constrained log-ratio analysis, but does not allow partialing of nuisance effects and does not provide hypothesis testing. **pcwOrd** allows partialled, constrained, and weighted ordinations, and also provides utility functions for investigating ordination objects, visualization, and hypothesis testing. The code is written to be self-documenting. Maybe you'll agree.

This isn't an R package yet, so load the functions with **source()**:

```
libs = c('easyCODA', # log-ratio analysis and CLR analysis
        'vegan')     # multivariate statistics
for(i in libs) {
  if (!require(i, character.only=TRUE)) {
    install.packages(i)
    if (!require(i, character.only=TRUE)) stop(paste("Cannot load", i))
  }
}

# pcwOrd
load('../pcwOrd_0.1.Rdata')

# data
spider = readRDS('Spider.RDS')
```

This vignette will demonstrate the features of **pcwOrd**: ordination, hypothesis-testing, and visualization. I'll use the **spiders** dataset from **mvabund**. This dataset has two tables:

1. A **data.frame** of environmental variables, to which I'll add a categorical version of **soil.dry\***.
2. A **matrix** of community data.

Certain **pcwOrd** functions use the rownames of the environmental and community data to cross-reference, so we want to ensure both tables have rownames. Usually these are something meaningful, like sample IDs.

\*technically this should be ordinal, but we'll treat it as categorical.

```

# environmental data
envi = as.data.frame(spider$x)

# categorical soil wetness
cats = c('dry', 'damp', 'wet', 'soaking')
cc = ceiling(envi$soil.dry)
envi$soil.cat = factor(cats[cc],
                      levels=cats)

# community data
comm = as.matrix(spider$abund)

# rownames
nobs = nrow(comm)
row_names = c(letters[], LETTERS[])[1:nobs]
rownames(envi) = row_names
rownames(comm) = row_names

str(envi)

## 'data.frame': 28 obs. of 7 variables:
## $ soil.dry : num 2.33 3.05 2.56 2.67 3.02 ...
## $ bare.sand : num 0 0 0 0 0 ...
## $ fallen.leaves: num 0 1.79 0 0 0 ...
## $ moss : num 3.04 1.1 2.4 2.4 0 ...
## $ herb.layer : num 4.45 4.56 4.61 4.62 4.62 ...
## $ reflection : num 3.91 1.61 3.69 3 2.3 ...
## $ soil.cat : Factor w/ 4 levels "dry","damp","wet",...: 3 4 3 3 4 4 4 3 3 3 ...

```

```
comm[1:8, 1:8]
```

```

## Alopacce Alopcone Alopfabr Arctlute Arctperi Auloalbi Pardlugu Pardmont
## a      25      10       0        0        0        4        0       60
## b       0       2       0        0        0       30       1       1
## c      15      20       2        2        0        9       1      29
## d       2       6       0        1        0       24       1       7
## e       1      20       0        2        0        9       1       2
## f       0       6       0        6        0        6       0      11
## g       2       7       0       12        0       16       1      30
## h       0      11       0        0        0        7      55       2

```

## 2 Basic Functions

### 2.1 Ordination

We'll first perform a basic principle components analysis of the community with `pcwOrd()` and visualize the results with `plot_ord()`:

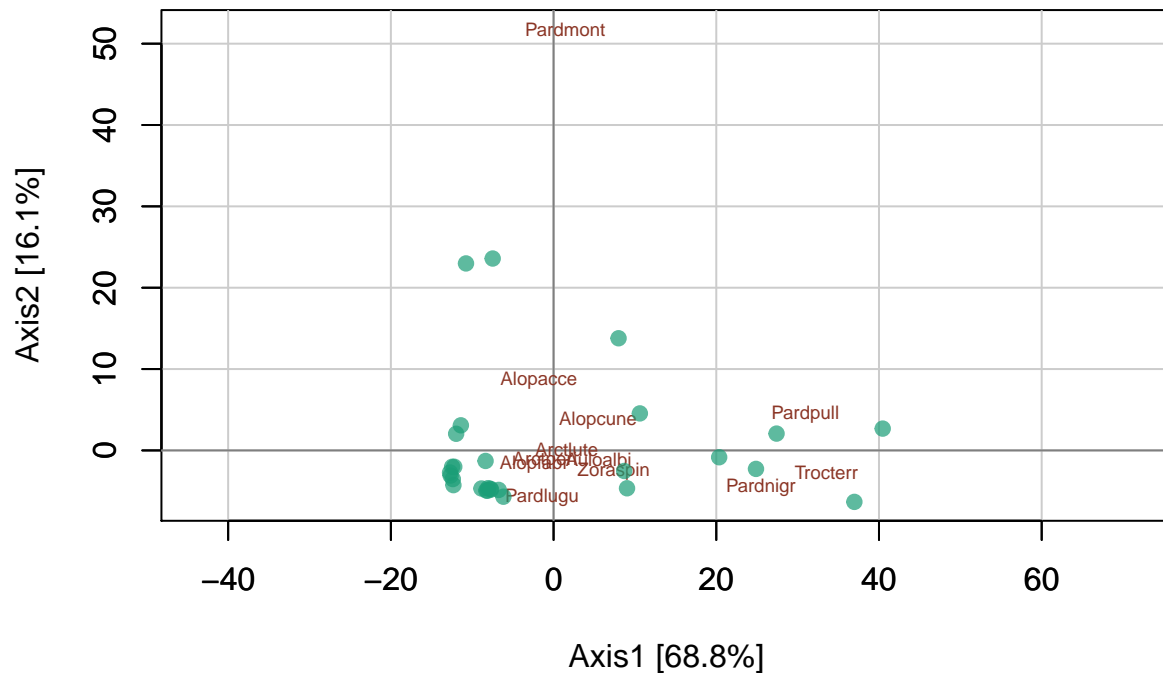
```

pca = pcwOrd(comm)
plot_ord(pca,
         main='Spiders: Basic PCA')

```

```
## Rescaling standard scores by 15.45
```

## Spiders: Basic PCA



The `pca` object is a list of class `pcwOrd` that contains a number of items. See the documentation for `pcwOrd`. Important ones are: `'Y_scaled'` - the community matrix after centering, scaling, and/or weighting but before further analysis; `'unconstrained'` - the singular value decomposition matrices (left `'u'` and right `'v'`) and values (`'d'`). All relevant information about this ordination can be calculated from these values.

### 2.1.1 Eigenvalues

For example, if we want to calculate eigenvalues of this unconstrained ordination, we need to access the singular values (vector `d`) of the unconstrained solution:

```
pca$unconstrained$d^2
```

```
## [1] 251.7432705 58.7674666 25.3873166 11.7224346 5.6704074 4.1539497
## [7] 2.7649418 2.3272743 1.6733360 0.8060037 0.6634983 0.1884040
```

### 2.1.2 Axis Variances

We can view these same data by calling `ord_variance`, which summarizes the variance explained by each axis in the solution:

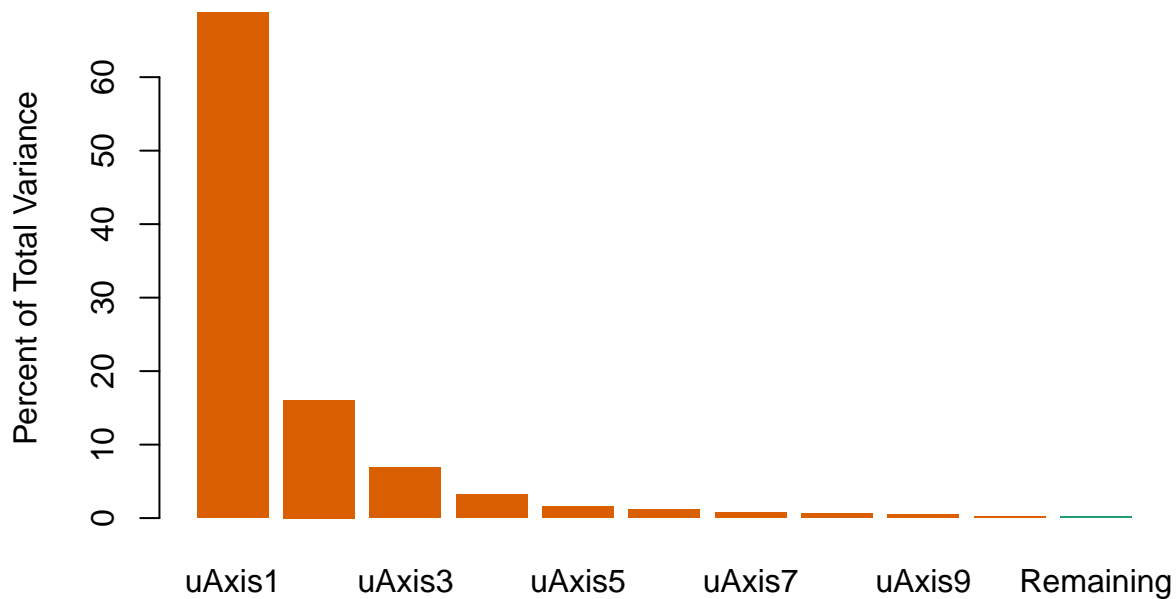
```
ord_variance(pca)
```

```
## $type
## [1] "variance"
##
## $summary
##           total partialled constrained unconstrained
## value      365.8683          0          0      365.8683
## pct_total 100.0000          0          0     100.0000
##
## $total
##           Total
## value      365.8683
## pct_group 100.0000
## pct_total 100.0000
##
## $partialled
##
## value
## pct_group
## pct_total
##
## $constrained
##
## value
## pct_group
## pct_total
##
## $unconstrained
##           uAxis1  uAxis2  uAxis3  uAxis4  uAxis5  uAxis6  uAxis7
## value      251.74327 58.76747 25.387317 11.722435 5.670407 4.153950 2.7649418
## pct_group   68.80707 16.06246  6.938922  3.204004 1.549849 1.135367 0.7557205
## pct_total   68.80707 16.06246  6.938922  3.204004 1.549849 1.135367 0.7557205
##           uAxis8  uAxis9  uAxis10  uAxis11  uAxis12
## value      2.3272743 1.6733360 0.8060037 0.6634983 0.18840395
## pct_group   0.6360962 0.4573602 0.2202989 0.1813489 0.05149502
## pct_total   0.6360962 0.4573602 0.2202989 0.1813489 0.05149502
```

### 2.1.3 Scree plots

We can also visualize variances across axes in a scree plot with `ord_scree()`:

```
ord_scree(pca)
```



#### 2.1.4 Calculate scores

If you want to make your own biplots, you can calculate row, column, centroid, and biplot scores by calling `ord_scores`. Scores can be principle, standard, or contribution - with the latter being raw singular values (either 'u' or 'v'). To make plotting easy, we can also add grouping information from the `envi` table:

```
grp = envi[, 'soil.cat', drop=FALSE] # preserve rownames
rowscores = ord_scores(pca,
  choice='row',
  scaling='principle',
  axes=c(1:3),
  add_grouping=grp)
head(rowscores)
```

```
##      Axis1      Axis2      Axis3 soil.cat
## a  7.988176 13.786265 -3.604174      wet
## b  9.030352 -4.652883 -5.718181    soaking
## c 10.606032  4.549642 -3.574303      wet
## d 24.881370 -2.272295 -8.576151      wet
## e 36.974719 -6.324696 19.094659    soaking
## f  8.683557 -2.525640 -1.763815    soaking
```

When plotting both standard and principle scores, often you'll need to rescale one of them for visualization:

```

colscores = ord_scores(pca,
                        choice='column',
                        scaling='standard',
                        axes=c(1:3))
scaled_colscores = scale_scores(colscores,
                                rowscores) # can also specify manual scaling factor

```

```
## Rescaling standard scores by 12.63
```

```

lapply(list(original = colscores,
            rescaled = scaled_colscores),
       head)

```

```

## $original
##           Axis1      Axis2      Axis3
## Alopacce -0.11660037  0.558436920  0.21907567
## Alopcone  0.35414730  0.243325952  0.17224233
## Alopfabr -0.15364830 -0.112002939  0.30171516
## Arctlute  0.10396386  0.008826392  0.04486536
## Arctperi -0.06877624 -0.081008292  0.12759310
## Auloalbi  0.35918227 -0.068503756 -0.42665830
##
## $rescaled
##           Axis1      Axis2      Axis3
## Alopacce -1.4732357  7.0558024  2.768002
## Alopcone  4.4746206  3.0744024  2.176267
## Alopfabr -1.9413330 -1.4151475  3.812145
## Arctlute  1.3135744  0.1115207  0.566870
## Arctperi -0.8689818 -1.0235328  1.612128
## Auloalbi  4.5382371 -0.8655391 -5.390791

```

### 2.1.5 Identify top features

Finally, it's useful to identify the top features. The default `scaling='contribution'` gives top contribution scores - i.e. the features that contribute most to the axes chosen. Here, we identify the spider taxa that have the five highest mean contributions across the first two axes:

```
top_scores(pca, n=5, choice='column', scaling='contribution', axes=c(1:2))
```

```

##           mean_contribution
## Pardmont      0.9681618
## Trocterr      0.6288081
## Pardpull      0.5841149
## Pardnigr      0.4835553
## Alopacce      0.1646834
## attr(,"axes")
## [1] 1 2

```

## 3 More ordinations

### 3.1 Log-ratio analysis

To perform log-ratio analysis:

1. Remove zeros from your data. For simplicity, I'll replace zeros with a pseudocount of 0.5. Note that I am *not* adding the pseudocount as  $x + 0.5$ , as this will distort the ratios between observations.
2. Close the community matrix, so that all abundances are relative abundances and  $\text{rowSums}(x) = 1$ .
3. Perform the log-ratio transformation of your choice. The centered log-ratio (CLR), where values are centered on the geometric mean, is a common choice.
4. Run principle components analysis on the log-ratio transformed matrix

```
# Pseudocounts
cc = comm
cc[cc==0] = 0.5 # replace zeros without adding to all values!

# Close the community
closed_comm = sweep(cc, 1, rowSums(comm), '/') # close to relative abundance

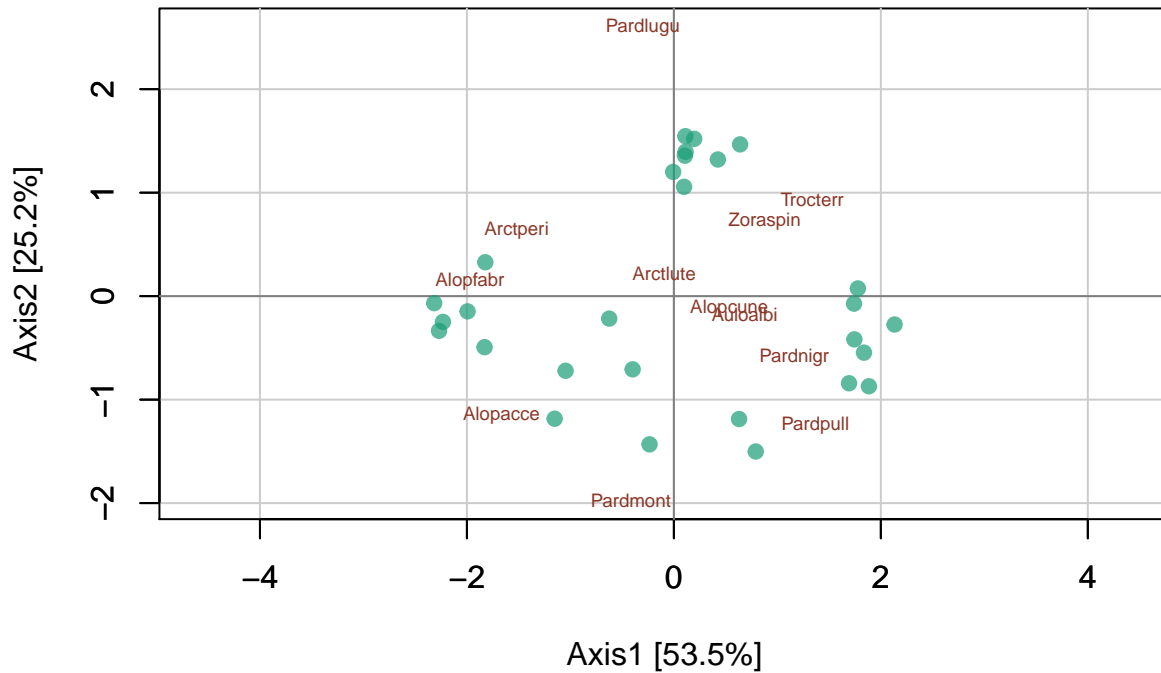
# perform centered log-ratio
geo_mean = apply(closed_comm, 1,
                 function(x) {
                   exp(sum(log(x))/length(x))
                 })
lr_comm = sweep(closed_comm, 1, geo_mean, '/')
lr_comm = log(lr_comm, 2)

lra = pcwOrd(lr_comm)
plot_ord(lra,
         main='Spiders: Log Ratio Analysis')
```

```
## Rescaling standard scores by 1.146
```



## Spiders: Log Ratio Analysis



### 3.2 Weighted ordinations

To weight either rows or columns, for example by sequencing depth or column prevalence, you have three options:

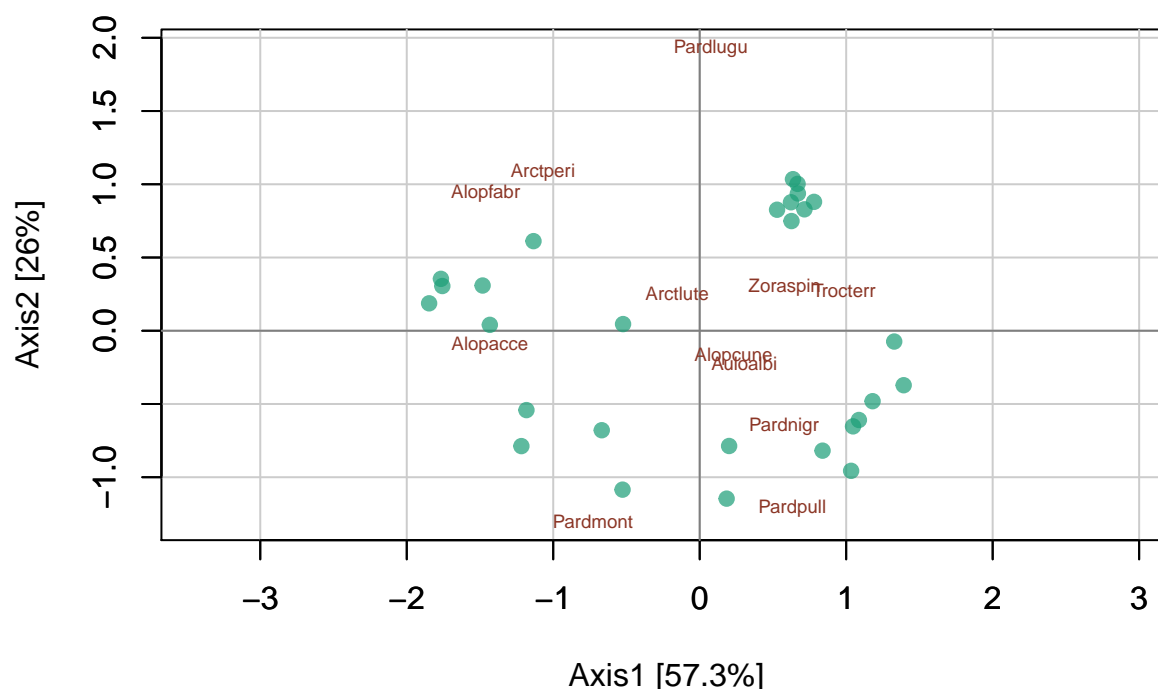
1. Tell `pcwOrd` to calculate weights automatically by setting `weight_rows=TRUE` or `weight_columns=TRUE`. This uses row or column masses as weights (via `rowSums`, for example).
2. Specify a vector of length `nrow()` or `ncol()` with the weights.
3. Input the result of `easyCODA::CLR()`, which returns a weighted CLR transformation and row-weights. This approach is demonstrated below:

Here is a weighted LRA:

```
wlr_comm = CLR(closed_comm) # CLR based on column-weighted geometric mean, plus column weights
w_lra = pcwOrd(wlr_comm)
plot_ord(w_lra,
         main='Spiders: Column-Weighted LRA')
```

```
## Rescaling standard scores by 0.9226
```

## Spiders: Column-Weighted LRA



In this situation, the results of weighted and unweighted ordinations are approximately the same. See Greenacre and Lewi (2009) for a discussion of when weighted log-ratio analysis might be advantageous over a non-weighted log ratio analysis.

### 3.3 Constrained ordination

We might be interested in how these spiders associate with environmental variables. For example, we might be interested in whether moss and reflection structures spider communities. A constrained ordination like redundancy analysis will do this: `pcwOrd` will regress the community matrix against moss and reflectivity, and then ordinate the fitted values.

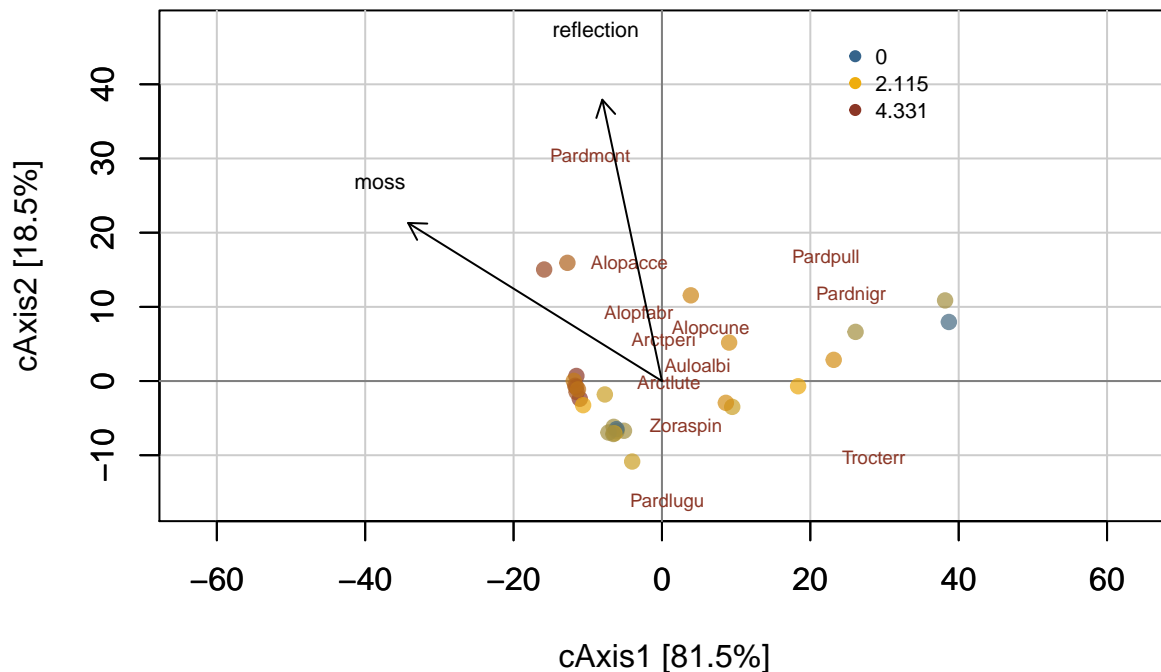
```
X = envi[, c('moss', 'reflection')] # preserve rownames
rda = pcwOrd(comm, X=X)

plot_ord(rda,
  main='Spiders by Moss Coverage',
  color_legend_position=c(0.8, 0))
```

```
## Rescaling standard scores by 13.4
```

```
## Rescaling standard scores by 48.24
```

## Spiders by Moss Coverage



In this plot, colors show moss coverage, the first column in X.

### 3.3.1 Significance testing

To test whether spiders vary with wetness class, run a PERMANOVA on the ordination with `permute_ord()`:

```
pp = permute_ord(rda)
pp
```

```
##   permute_on total_variance variance_after_partialing   fitted residuals num_df
## 1   partial      365.8683          365.8683 98.72652  267.1418      2
##   denom_df   F_stat p_val F_perm
## 1       25 4.619575 0.004   999
```

`permute_ord()` has a number of permutation models that parallel the options in `vegan::permutest()` - and `permute_ord()` will return the same results as `vegan::permutest()` with the same randomization seed. We see a p-value of 0.004 after 999 permutations.

### 3.4 Partial ordination

Say we want to look at how moss affects spider communities independently of moisture class. We can partial this out as Z:

```

Z = envi[, 'soil.cat', drop=FALSE] # preserve rownames
X = envi[, c('moss', 'reflection'), drop=FALSE]

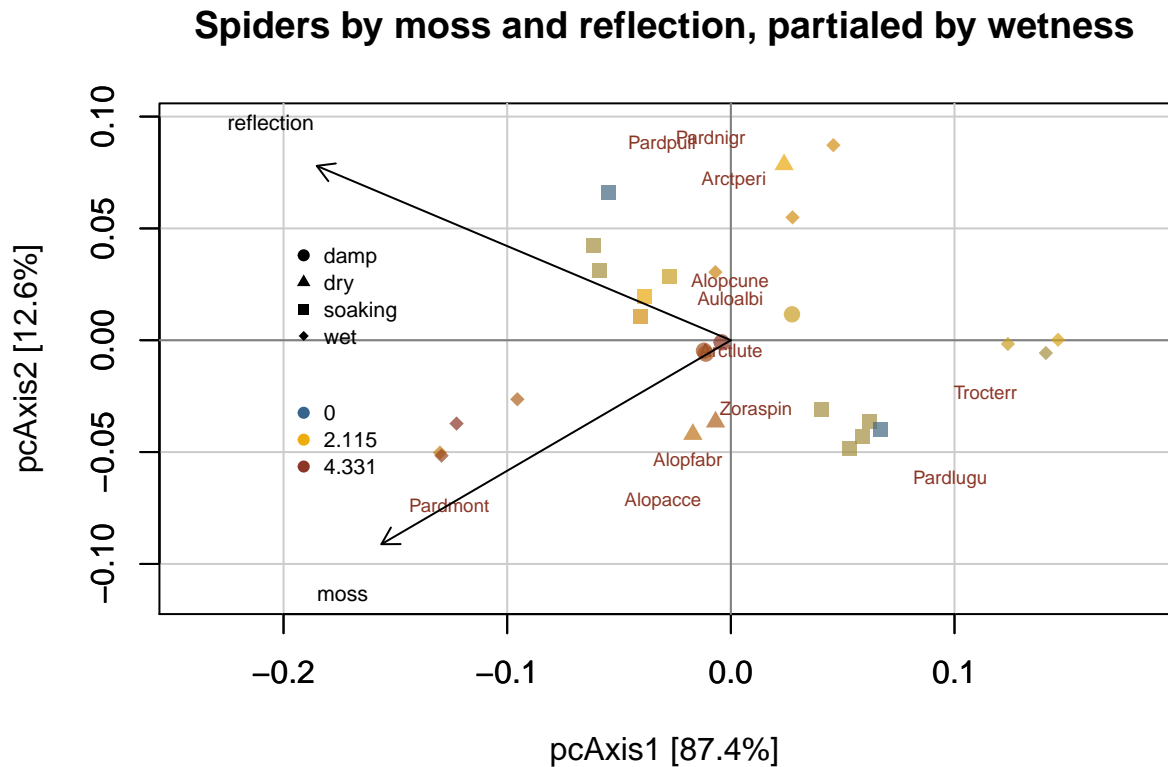
pcrda = pcwOrd(closed_comm, X, Z)

plot_ord(pcrda,
  main='Spiders by moss and reflection, partialled by wetness',
  shape_legend_position=c(-0.2, 0.05))

```

```
## Rescaling standard scores by 0.05837
```

```
## Rescaling standard scores by 0.2961
```



Each of the wetness categories is centered at zero, as expected due to partialing. Moss increases to the bottom left (darker red, and arrows). This model is highly significant:

```
permute_ord(pcrda)
```

```

##   permute_on total_variance variance_after_partialing   fitted   residuals
## 1   partial      0.01927485          0.01070139 0.004584283 0.006117103
##   num_df denom_df   F_stat p_val F_perm
## 1      2       22 8.243627 0.001   999

```

### 3.5 Partialled, constrained, weighted ordination

The reason for this package is to perform partial, constrained, weighted ordinations. Here is a weighted log-ratio analysis of the spider community, constrained by moss and reflection, with moisture category partialled out:

```
Y = CLR(closed_comm)

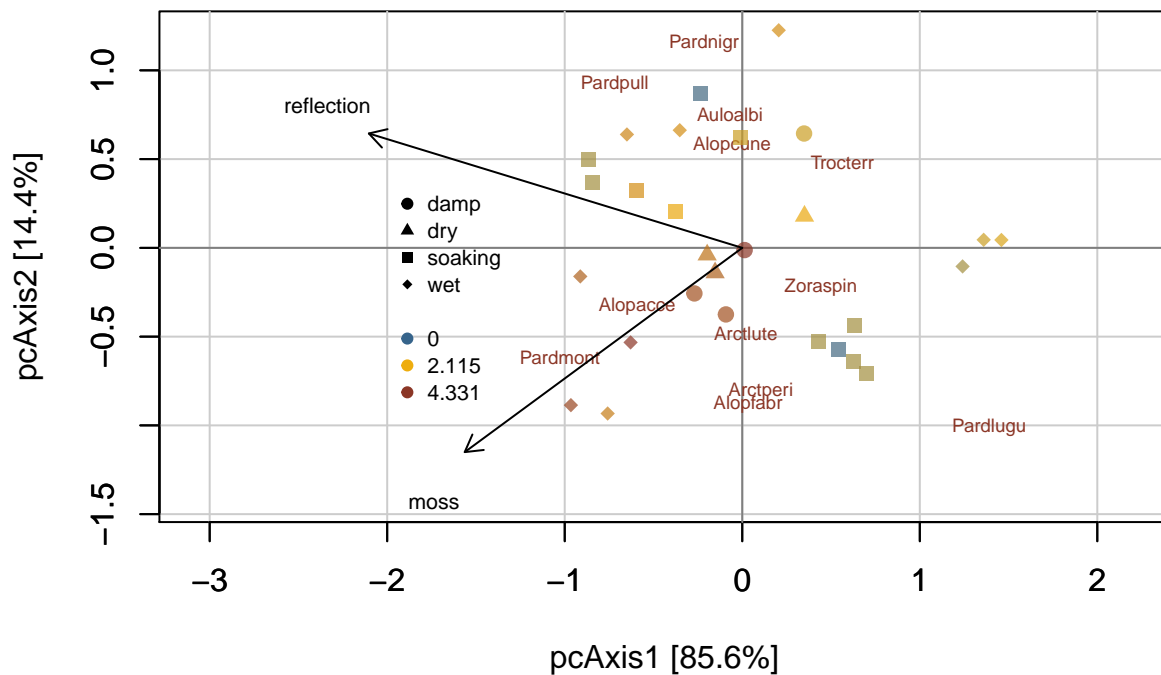
pcw_ord = pcwOrd(Y, X, Z)

plot_ord(pcw_ord,
  main='Partialled, Constrained, Weighted Log-Ratio Analysis',
  shape_legend_position=c(-2, 0.4))
```

```
## Rescaling standard scores by 0.7005
```

```
## Rescaling standard scores by 3.215
```

#### Partialled, Constrained, Weighted Log-Ratio Analysis



## 4 Comparison to vegan and easyCODA

pcwOrd gives the same results as comparable `vegan` and `easycODA` functions.

## 4.1 Eigenvalues

Returning to eigenvalues: these eigenvalues are exactly the same as calculated by easyCODA's PCA, and solutions are the same:

```
easy_pca = PCA(comm, weight=FALSE)

rbind(pcwOrd = pca$unconstrained$d^2,
      easyCODA = easy_pca$sv^2
)
```

```
##           [,1]      [,2]      [,3]      [,4]      [,5]      [,6]      [,7]      [,8]
## pcwOrd    251.7433  58.76747  25.38732  11.72243  5.670407  4.15395  2.764942  2.327274
## easyCODA  251.7433  58.76747  25.38732  11.72243  5.670407  4.15395  2.764942  2.327274
##           [,9]      [,10]     [,11]     [,12]
## pcwOrd     1.673336  0.8060037  0.6634983  0.188404
## easyCODA   1.673336  0.8060037  0.6634983  0.188404
```

Eigenvalues are not identical to `vegan` eigenvalues, because `pcwOrd` (and `easyCODA`) weights the initial community matrix rows by  $1/\sqrt{nrow}$ , while `vegan` weights rows by  $1/\sqrt{nrow-1}$ . You can access `vegan`-style weighting, which will agree with `vegan` results:

```
veganlike_pca = pcwOrd(comm, as_vegan=TRUE)
vegan_pca = rda(comm)

rbind(pcwOrd = veganlike_pca$unconstrained$d^2,
      vegan = vegan_pca$CA$eig)
```

```
##           PC1      PC2      PC3      PC4      PC5      PC6      PC7      PC8
## pcwOrd  3132.805  731.3285  315.9311  145.8792  70.56507  51.6936  34.40816  28.96164
## vegan   3132.805  731.3285  315.9311  145.8792  70.56507  51.6936  34.40816  28.96164
##           PC9      PC10     PC11     PC12
## pcwOrd   20.82374  10.03027  8.256867  2.344583
## vegan    20.82374  10.03027  8.256867  2.344583
```

These weightings don't change relative variance or relative distances, only absolute variances and distances. Generally, we're concerned with relative variances and distances, so the choice of (equal) weighting doesn't matter.

```
list(
  coda_weightings = ord_variance(pca)$unconstrained,
  vegan_weightings = ord_variance(veganlike_pca)$unconstrained
)
```

```
## $coda_weightings
##           uAxis1  uAxis2  uAxis3  uAxis4  uAxis5  uAxis6  uAxis7
## value      251.74327  58.76747  25.387317  11.722435  5.670407  4.153950  2.7649418
## pct_group   68.80707  16.06246  6.938922  3.204004  1.549849  1.135367  0.7557205
## pct_total   68.80707  16.06246  6.938922  3.204004  1.549849  1.135367  0.7557205
##           uAxis8  uAxis9  uAxis10  uAxis11  uAxis12
## value      2.3272743  1.6733360  0.8060037  0.6634983  0.18840395
## pct_group   0.6360962  0.4573602  0.2202989  0.1813489  0.05149502
```

```
## pct_total 0.6360962 0.4573602 0.2202989 0.1813489 0.05149502
##
## $vegan_weightings
##           uAxis1    uAxis2    uAxis3    uAxis4    uAxis5    uAxis6
## value      3132.80514 731.32847 315.931051 145.879186 70.565070 51.693597
## pct_group   68.80707  16.06246  6.938922   3.204004  1.549849  1.135367
## pct_total   68.80707  16.06246  6.938922   3.204004  1.549849  1.135367
##           uAxis7    uAxis8    uAxis9    uAxis10   uAxis11   uAxis12
## value      34.4081650 28.9616352 20.8237366 10.0302687 8.2568673 2.34458251
## pct_group   0.7557205 0.6360962 0.4573602 0.2202989 0.1813489 0.05149502
## pct_total   0.7557205 0.6360962 0.4573602 0.2202989 0.1813489 0.05149502
```

## 4.2 Scores

```
ord_scores(pca, 'row', 'principle')
```

```
##           Axis1      Axis2
## a    7.988176 13.7862654
## b    9.030352 -4.6528827
## c   10.606032  4.5496420
## d   24.881370 -2.2722951
## e   36.974719 -6.3246958
## f    8.683557 -2.5256396
## g   40.458746  2.6835088
## h   -6.167524 -5.6887597
## i  -11.395552  3.0854298
## j  -11.973728  2.0464630
## k  -10.765634 22.9846309
## l   -7.483546 23.5898261
## m   27.409257  2.0678738
## n   20.354007 -0.8378813
## o   -7.687032 -4.8091247
## p   -6.740881 -4.8547748
## q   -8.058080 -4.6416878
## r   -7.769638 -4.7163087
## s   -8.108993 -4.9559103
## t   -8.264546 -4.9285167
## u   -8.866147 -4.6802406
## v  -12.383293 -3.5219133
## w  -12.469082 -2.0121482
## x  -12.652253 -3.0858499
## y   -8.360181 -1.2930062
## z  -12.312196 -4.2679844
## A  -12.721757 -2.7161932
## B  -12.206151 -2.0078266
```

```
easy_pca$rowcoord[, 1:2]
```

```
##           [,1]      [,2]
## [1,]  0.5034643  1.7983663
## [2,]  0.5691487 -0.6069510
```

```
## [3,] 0.6684578 0.5934836
## [4,] 1.5681780 -0.2964123
## [5,] 2.3303757 -0.8250327
## [6,] 0.5472915 -0.3294601
## [7,] 2.5499606 0.3500536
## [8,] -0.3887156 -0.7420772
## [9,] -0.7182182 0.4024827
## [10,] -0.7546585 0.2669534
## [11,] -0.6785169 2.9982583
## [12,] -0.4716594 3.0772038
## [13,] 1.7275011 0.2697463
## [14,] 1.2828355 -0.1092984
## [15,] -0.4844843 -0.6273322
## [16,] -0.4248521 -0.6332871
## [17,] -0.5078701 -0.6054906
## [18,] -0.4896907 -0.6152247
## [19,] -0.5110789 -0.6464798
## [20,] -0.5208828 -0.6429064
## [21,] -0.5587995 -0.6105197
## [22,] -0.7804718 -0.4594203
## [23,] -0.7858788 -0.2624771
## [24,] -0.7974233 -0.4025375
## [25,] -0.5269104 -0.1686678
## [26,] -0.7759908 -0.5567424
## [27,] -0.8018039 -0.3543171
## [28,] -0.7693072 -0.2619134
```

### 4.3 Permutation Tests

`vegan::permutest()` gives the same as `pcwOrd::permute_ord()`, if you set the same seed.

```
vegan_rda = rda(comm, X)
veganlike_rda = pcwOrd(comm, X, as_vegan=TRUE)

set.seed(445)
vegan_permutes = permutest(vegan_rda,
                           permutations=999)

set.seed(445)
pcw_permutes = permute_ord(veganlike_rda)
keep = c('fitted', 'residuals', 'num_df', 'denom_df', 'F_stat', 'p_val')

list(vegan = vegan_permutes,
     pcwOrd = pcw_permutes[keep])

## $vegan
##
## Permutation test for rda under reduced model
##
## Permutation: free
## Number of permutations: 999
##
## Model: rda(X = comm, Y = X)
## Permutation test for all constrained eigenvalues
```



```
##           Df Inertia      F Pr(>F)
## Model      2  1228.6 4.6196  0.004 **
## Residual 25  3324.4
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## $pcwOrd
##      fitted residuals num_df denom_df    F_stat p_val
## 1 1228.597  3324.431      2        25 4.619575 0.004
```

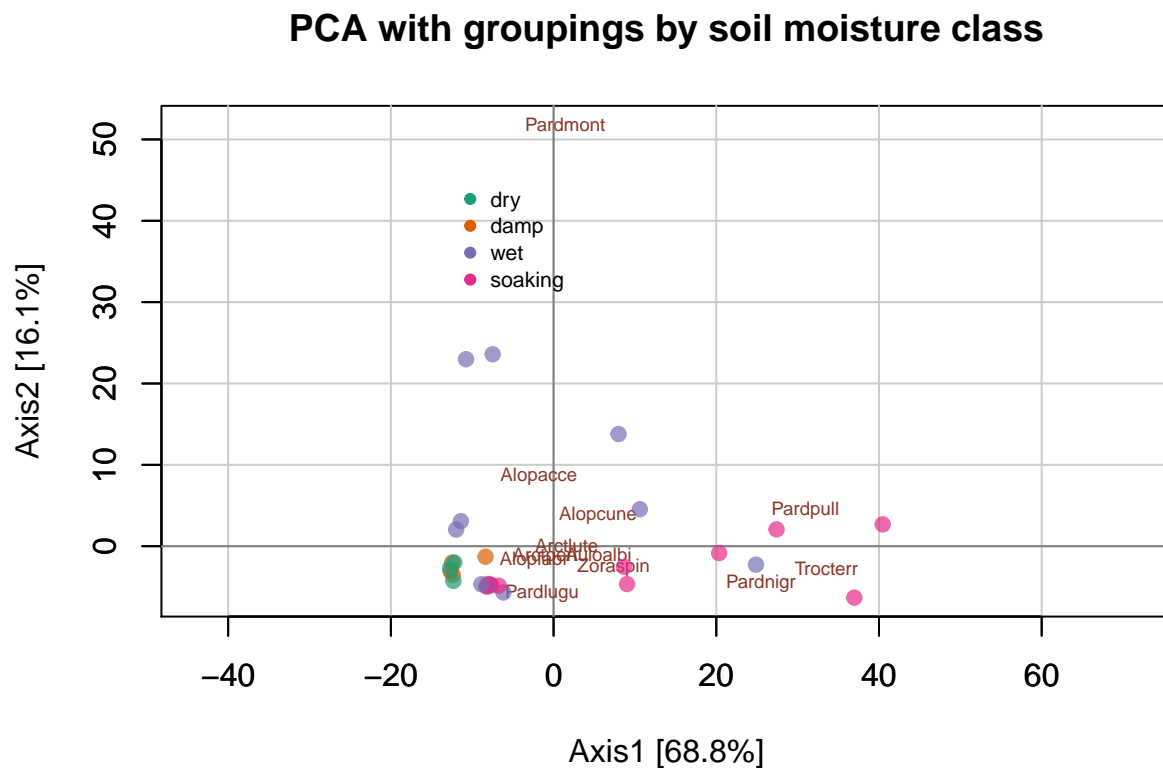
## 5 Additional plotting options

### 5.1 Added categories

Add groupings to any ordination. They can be continuous.

```
row_group = env[, 'soil.cat', drop=FALSE]
plot_ord(pca,
         row_group=row_group,
         main='PCA with groupings by soil moisture class')
```

```
## Rescaling standard scores by 15.45
```

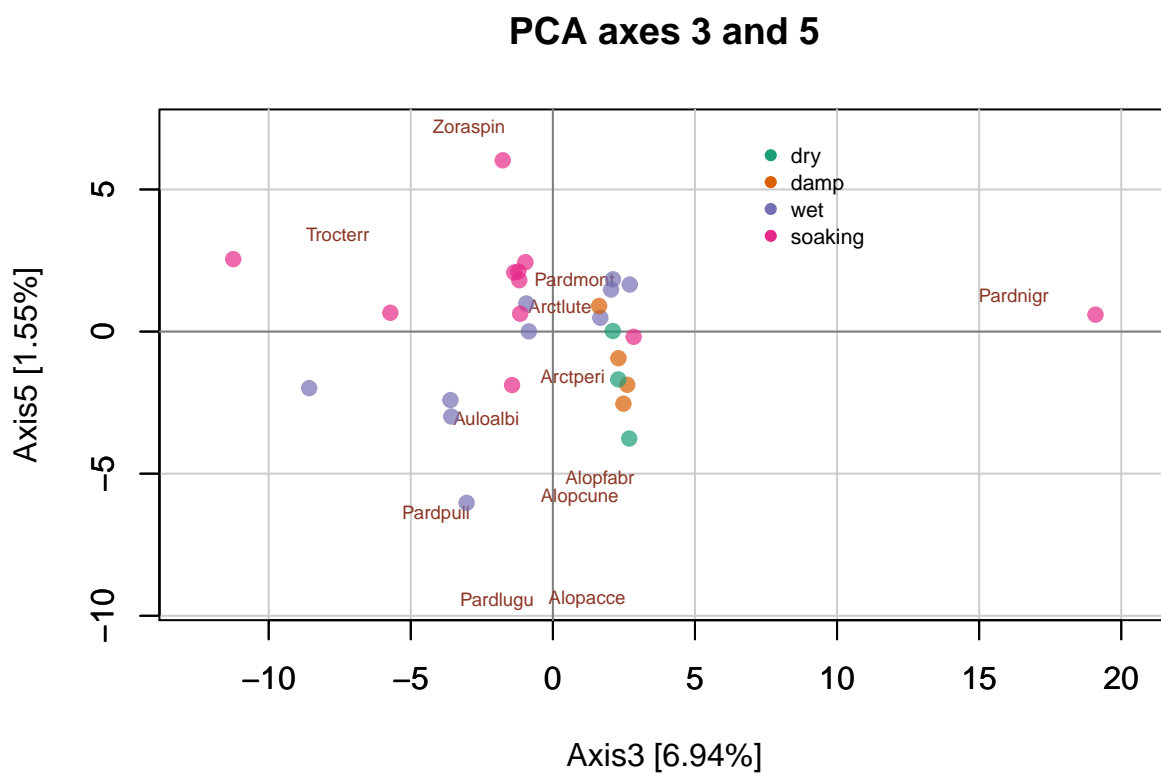


## 5.2 Different axes

Plot different axes

```
plot_ord(pca,  
         row_group=row_group,  
         axes=c(3,5),  
         main='PCA axes 3 and 5',  
         color_legend_position=c(0.6, 0))
```

```
## Rescaling standard scores by 5.487
```

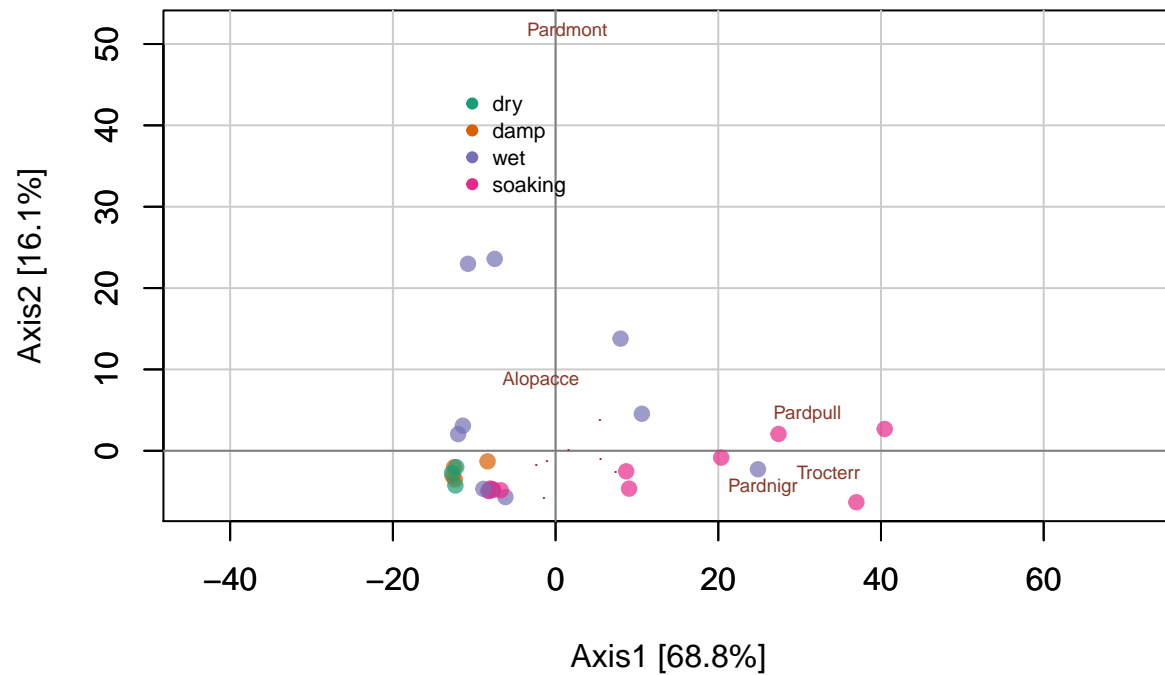


## 5.3 Limit column labels

Reduce clutter by only labeling the top columns (by plotted score)

```
plot_ord(pca,  
         row_group=row_group,  
         max_labels=5)
```

```
## Rescaling standard scores by 15.45
```



## 5.4 colors and shapes

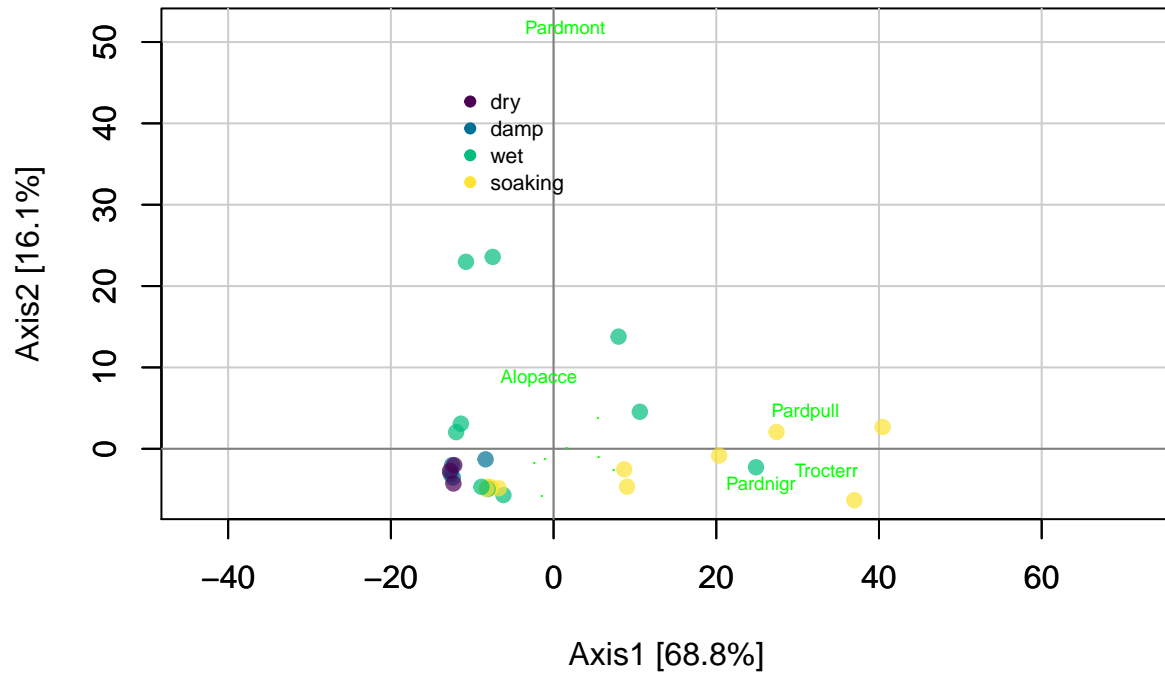
Change the colors of text, groupings, arrows, and change shapes.

```
new_colors = hcl.colors(4, palette='Viridis')

plot_ord(pca,
         row_group=row_group,
         max_labels=5,
         discrete_scale=new_colors,
         col_text='green',
         main='Fluorescent PCA')
```

```
## Rescaling standard scores by 15.45
```

## Fluorescent PCA



If you constrain an ordination with categories, the plot function will automatically recognize these and plot them as named centroids:

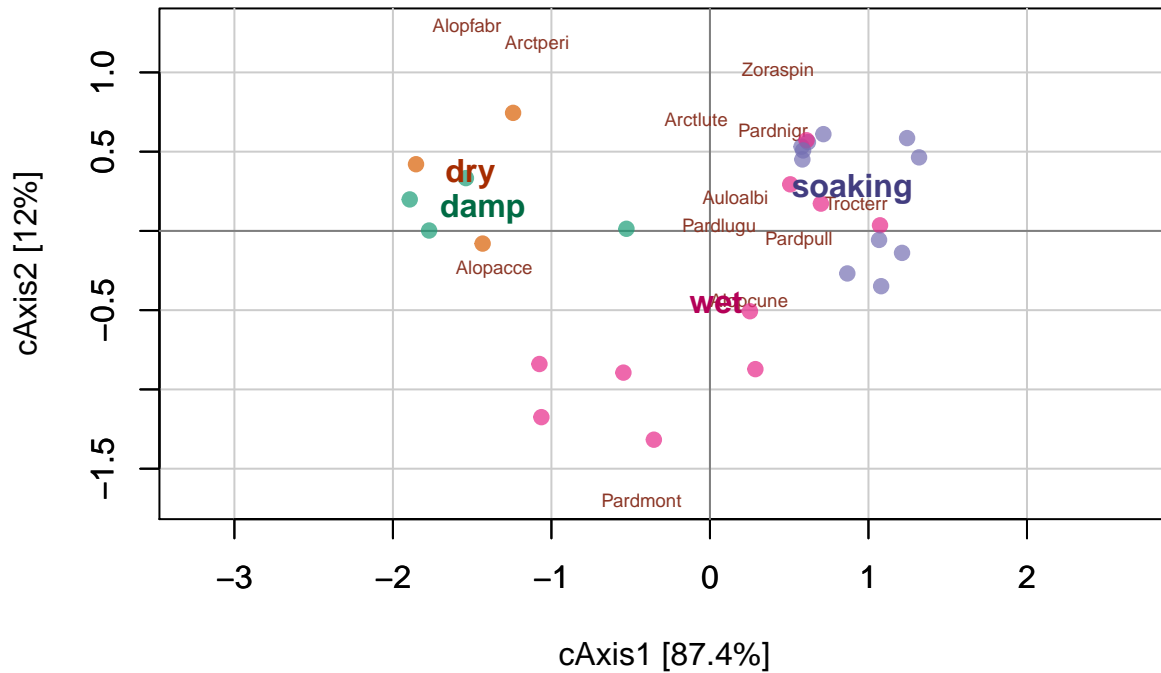
```
X = envi[, 'soil.cat', drop=FALSE]
Y = CLR(closed_comm)

cat_cwLRA = pcwOrd(Y, X)

plot_ord(cat_cwLRA,
          main='Spiders wLRA with Centroids')
```

```
## Rescaling standard scores by 0.8593
```

## Spiders wLRA with Centroids



### 5.5 Screeplots as grobs

Screeplots can be produced as grobs (via `ggplot2`) for downstream manipulation, saving as R objects, and arranging with `gridExtra`. If you want to do this with ordinations, you're on your own!

```
ord_screep(cwLRA, main='ggplot2 Scree Plot', as_grob=TRUE) +  
  theme_dark()
```

```
## Loading required package: ggplot2
```

ggplot2 Scree Plot

