

# Stats Reasoning 6

Paige and Peter

```
library(tidyverse) # For data wrangling
```

```
Warning: package 'ggplot2' was built under R version 4.4.3
```

```
Warning: package 'tibble' was built under R version 4.4.3
```

```
Warning: package 'purrr' was built under R version 4.4.3
```

```
-- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
v dplyr     1.1.4     v readr     2.1.5
v forcats   1.0.0     v stringr   1.5.1
v ggplot2   4.0.1     v tibble    3.3.1
v lubridate  1.9.4     v tidyr    1.3.1
v purrr    1.2.1
-- Conflicts ----- tidyverse_conflicts() --
x dplyr::filter() masks stats::filter()
x dplyr::lag()    masks stats::lag()
i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to becom
```

```
library(brms) # For stats
```

```
Warning: package 'brms' was built under R version 4.4.3
```

```
Loading required package: Rcpp
Loading 'brms' package (version 2.23.0). Useful instructions
can be found by typing help('brms'). A more detailed introduction
to the package is available through vignette('brms_overview').
```

```
Attaching package: 'brms'
```

```
The following object is masked from 'package:stats':
```

```
ar
```

```
library(ggeffects) # for plotting model predictions
```

```
Warning: package 'ggeffects' was built under R version 4.4.3
```

```
# Note: I needed to also install the `insight` and `see` packages to get `modelbased` to inst
# install.packages('modelbased') # if you need to install this package
library(modelbased) # for plotting model predictions. supports the link scale (ggeffects does
```

```
Warning: package 'modelbased' was built under R version 4.4.3
```

```
Attaching package: 'modelbased'
```

```
The following objects are masked from 'package:ggeffects':
```

```
pool_predictions, residualize_over_grid
```

```
# install.packages('faraway') # if you need to install this package
library(faraway) # For data on galapagos species richness
```

```
Warning: package 'faraway' was built under R version 4.4.3
```

```
Attaching package: 'faraway'
```

```
The following object is masked from 'package:brms':
```

```
epilepsy
```

## **Q1.1**

### **Q1.1a**

1. Numerical (0 to infinity, positive, whole numbers)
2. Y/N, 0/1
3. 0-1 or 0-100 , percent
4. Numerical
5. Numerical (positive)

### **Q1.1b Choose a distribution that fits each of the response variables**

1. Poisson
2. Bernoulli
3. Beta
4. Gamma
5. Gamma

### **Q1.2 Choose a distribution that fits your final project response variable**

Now, 1. Write the response variable that you are using for your final project.

#### **Beta diversity**

2. What values can your response variable take on? (e.g. real numbers, negative real numbers, positive real numbers, zeroes, integers, fractions, 0's/1's, etc.)

#### **Continuous, real numbers**

3. What distribution (or distributions) fits your response variable?

#### **Gaussian / gamma**

## 1.2

```
# Read in the pre-stored data
data("gala")
# Check out the first 6 rows
head(gala)
```

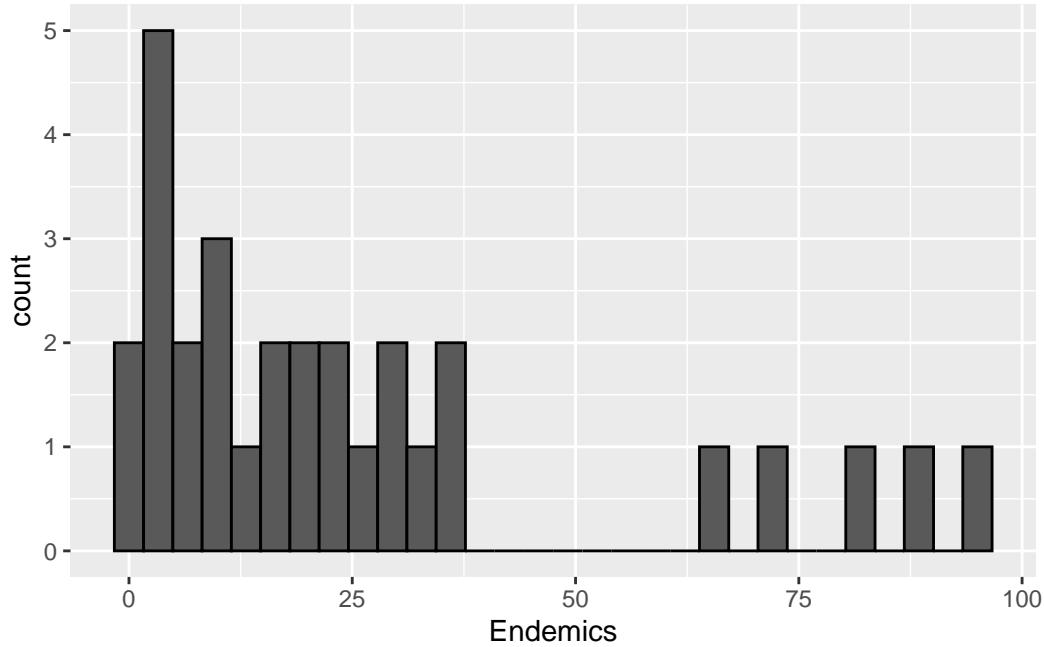
	Species	Endemics	Area	Elevation	Nearest	Scruz	Adjacent
Baltra	58	23	25.09	346	0.6	0.6	1.84
Bartolome	31	21	1.24	109	0.6	26.3	572.33
Caldwell	3	3	0.21	114	2.8	58.7	0.78
Champion	25	9	0.10	46	1.9	47.4	0.18
Coamano	2	1	0.05	77	1.9	1.9	903.82
Daphne.Major	18	11	0.34	119	8.0	8.0	1.84

```
# Look at the help page too!
```

## Q1.3

```
ggplot(gala, aes(x= Endemics))+
  geom_histogram(color="black")
```

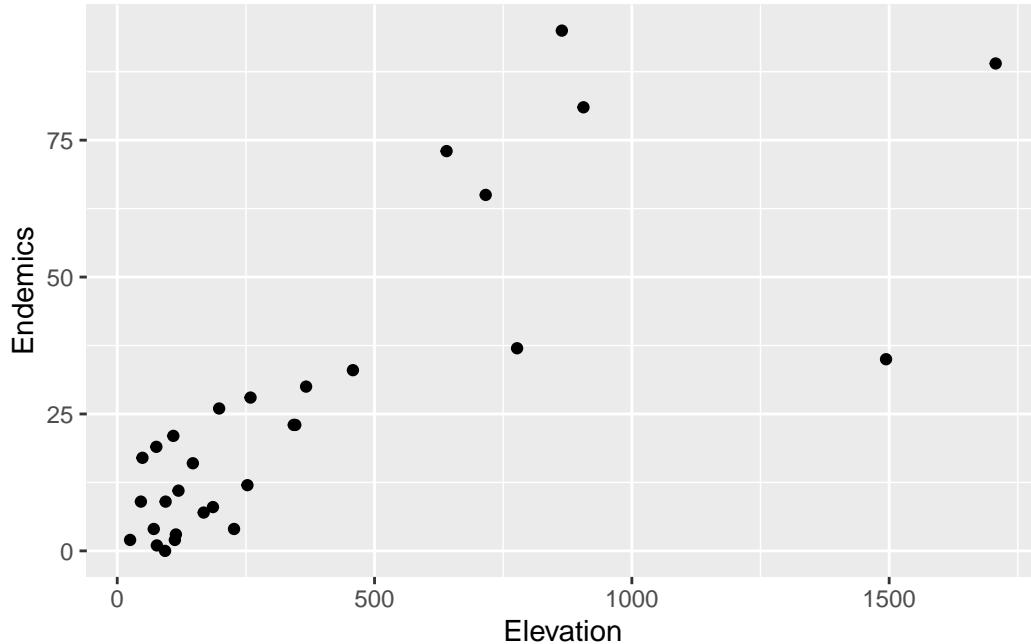
```
`stat_bin()` using `bins = 30`. Pick better value `binwidth`.
```



#### Q1.4 Plot Endemics ~ Elevation

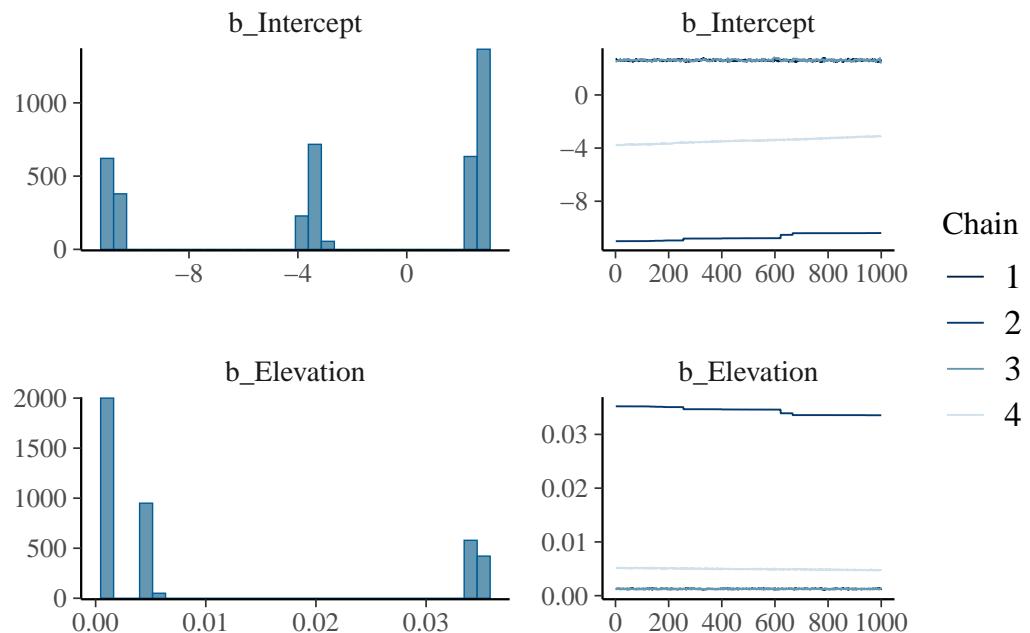
Make a plot to visualize the model we are about to run: Endemics as a function of Elevation.

```
ggplot(gala, aes(x= Elevation, y=Endemics))+  
  geom_point()
```

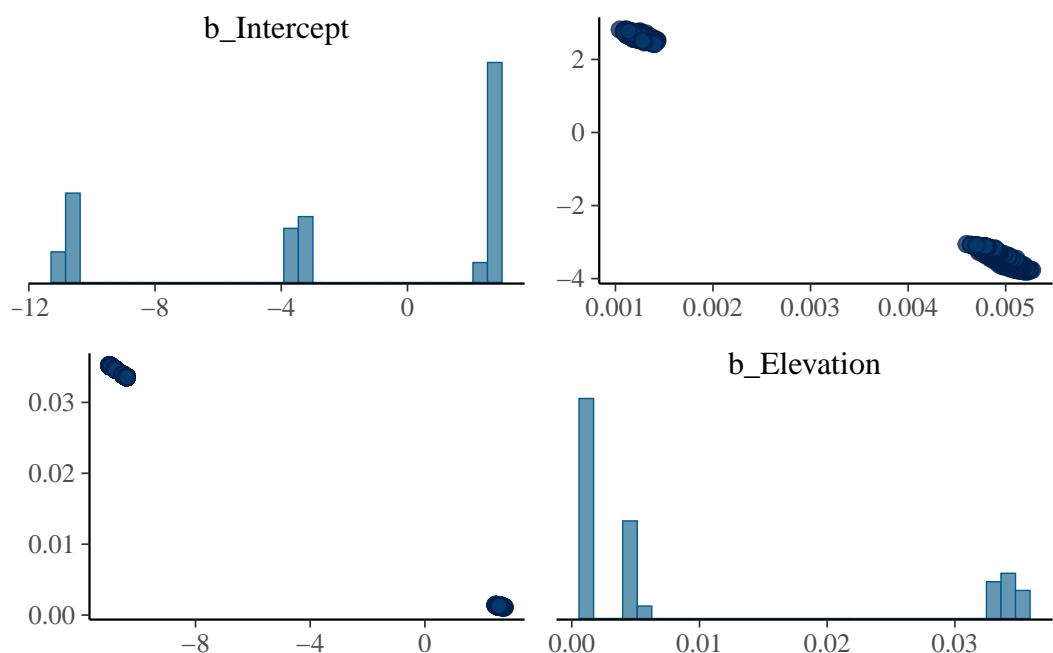


```
# Endemics ~ Elevation
m.elev <-
  brm(data = gala, # Give the model the penguins data
       # Choose a poisson distribution - THIS IS THE NEW PART!
       family = poisson(link = "log"),
       # Specify the model here.
       Endemics ~ 1 + Elevation,
       # Here's where you specify parameters for executing the Markov chains
       # We're using similar to the defaults, except we set cores to 4 so the analysis runs faster
       iter = 2000, warmup = 1000, chains = 4, cores = 4,
       # Save the fitted model object as output - helpful for reloading in the output later
       file = "output/m.elev")
```

`plot(m.elev)`



```
pairs(m.elev)
```



```
summary(m.elev)
```

Warning: Parts of the model have not converged (some Rhats are > 1.05). Be careful when analysing the results! We recommend running more iterations and/or setting stronger priors.

```
Family: poisson  
Links: mu = log  
Formula: Endemics ~ 1 + Elevation  
Data: gala (Number of observations: 30)  
Draws: 4 chains, each with iter = 2000; warmup = 1000; thin = 1;  
total post-warmup draws = 4000
```

Regression Coefficients:

	Estimate	Est.Error	1-95% CI	u-95% CI	Rhat	Bulk_ESS	Tail_ESS
Intercept	-2.23	5.48	-11.00	2.70	2.31	5	11
Elevation	0.01	0.01	0.00	0.04	2.30	5	11

Draws were sampled using sampling(NUTS). For each parameter, Bulk\_ESS and Tail\_ESS are effective sample size measures, and Rhat is the potential scale reduction factor on split chains (at convergence, Rhat = 1).

## Q 1.5

Answer: The Rhat values are ~2.3 and very different from 1. The posterior plots are not unimodal and the posterior chains are not overlapping.

Clustering at either extreme of the elevation. Not enough data at either low or high elevations to properly train the model and set the intercept.

## Q1.6 Center the predictors

```
print(m.elev2, digits=4)
```

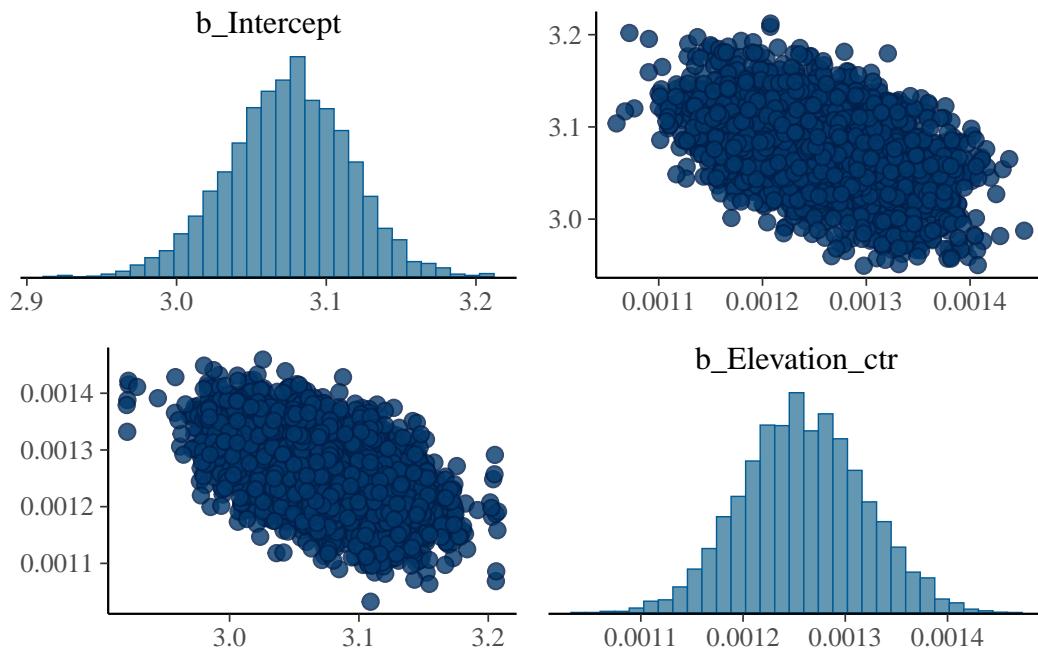
```
Family: poisson  
Links: mu = log  
Formula: Endemics ~ 1 + Elevation_ctr  
Data: gala (Number of observations: 30)  
Draws: 4 chains, each with iter = 6000; warmup = 4000; thin = 1;  
total post-warmup draws = 8000
```

Regression Coefficients:

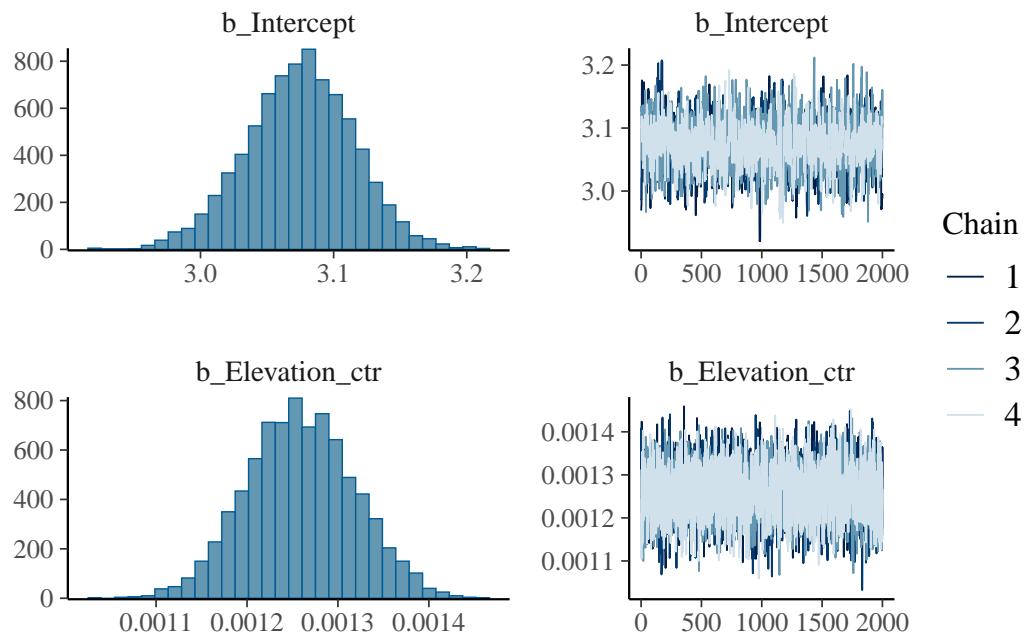
	Estimate	Est.Error	l-95% CI	u-95% CI	Rhat	Bulk_ESS	Tail_ESS
Intercept	3.0747	0.0402	2.9931	3.1514	1.0016	1397	1298
Elevation_ctr	0.0013	0.0001	0.0011	0.0014	1.0003	3749	4499

Draws were sampled using sampling(NUTS). For each parameter, Bulk\_ESS and Tail\_ESS are effective sample size measures, and Rhat is the potential scale reduction factor on split chains (at convergence, Rhat = 1).

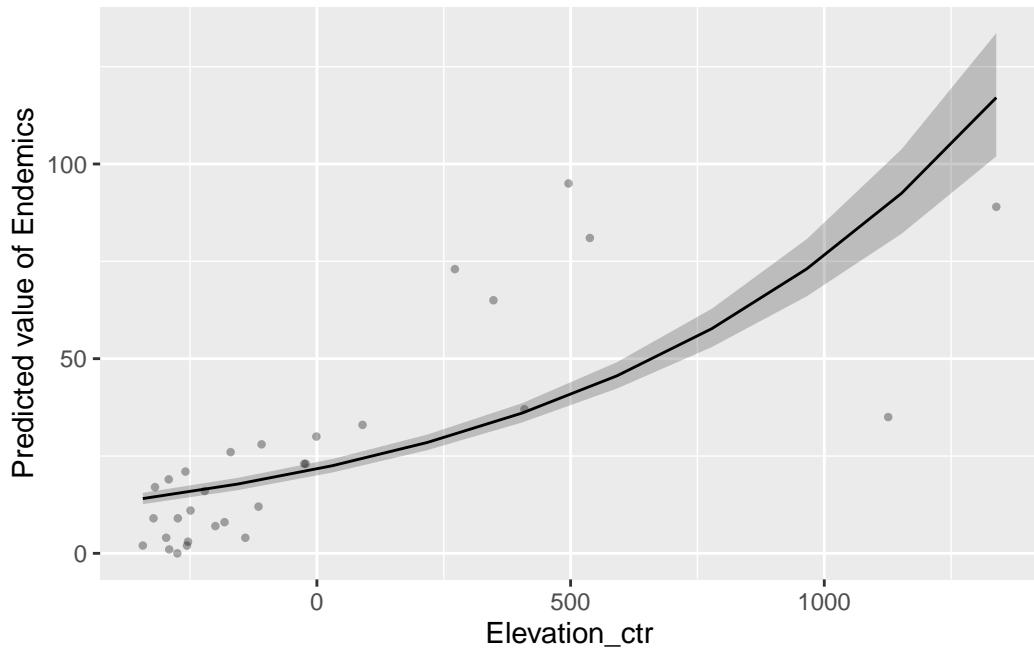
```
pairs(m.elev2)
```



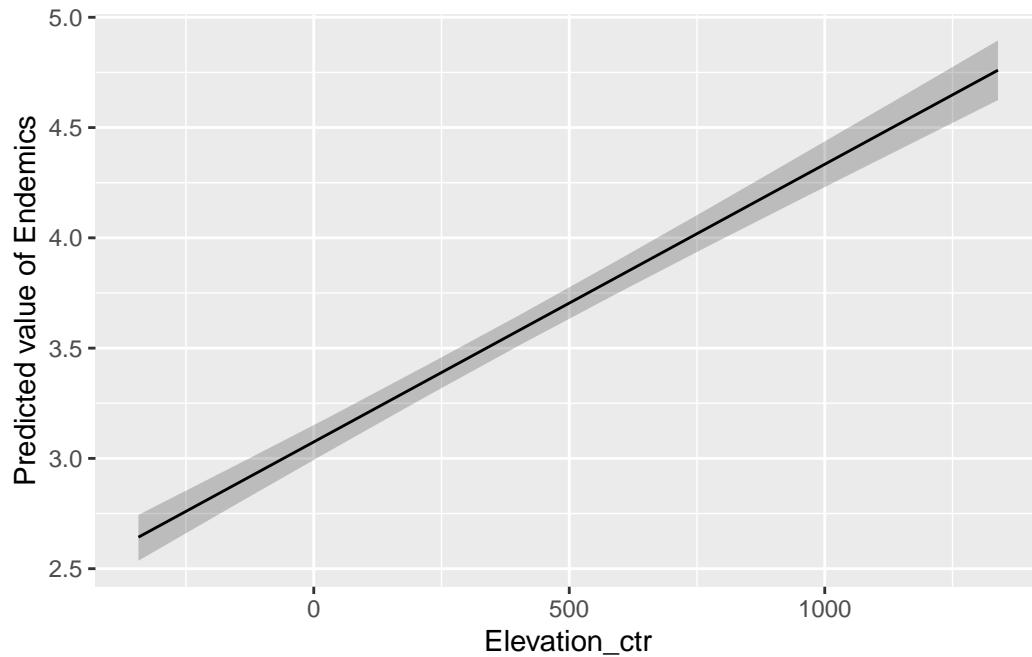
```
plot(m.elev2)
```



```
preds <- estimate_expectation(m.elev2, by = 'Elevation_ctr')
plot(preds, show_data = TRUE)
```



```
predslog <- estimate_expectation(m.elev2, by = 'Elevation_ctr', predict = 'link')
plot(predslog)
```



```
print(m.elev2, digits = 4)
```

Family: poisson  
 Links: mu = log  
 Formula: Endemics ~ 1 + Elevation\_ctr  
 Data: gala (Number of observations: 30)  
 Draws: 4 chains, each with iter = 6000; warmup = 4000; thin = 1;  
 total post-warmup draws = 8000

#### Regression Coefficients:

	Estimate	Est.Error	l-95% CI	u-95% CI	Rhat	Bulk_ESS	Tail_ESS
Intercept	3.0747	0.0402	2.9931	3.1514	1.0016	1397	1298
Elevation_ctr	0.0013	0.0001	0.0011	0.0014	1.0003	3749	4499

Draws were sampled using sampling(NUTS). For each parameter, Bulk\_ESS and Tail\_ESS are effective sample size measures, and Rhat is the potential scale reduction factor on split chains (at convergence, Rhat = 1).

1. Back transform - My slope value is 0.0013, so I will take the `exp()` of that value:

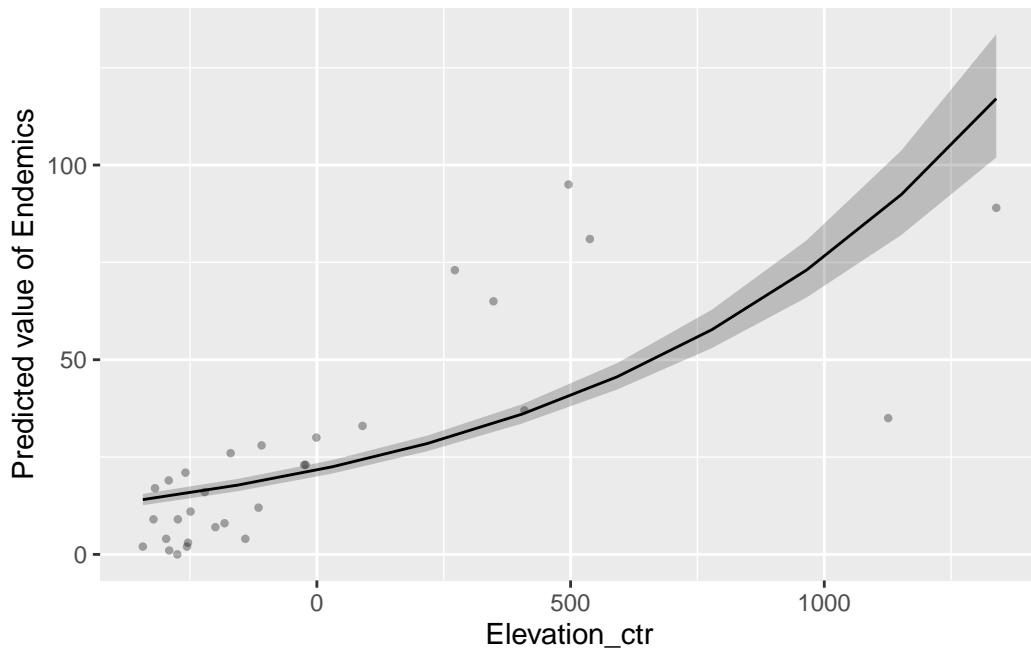
```
exp(0.0013)
```

```
[1] 1.001301
```

This back transforms to 1.0013

## 2. Interpret:

```
plot(preds, show_data = TRUE)
```



### Q1.7 What is the percent change on the response scale?

1. Number of Clarkias blooming as a function of temperature in Celsius: 1.09

```
exp(1.09)
```

```
[1] 2.974274
```

For every increase in degree C, there is a 2.9% increase in clarkias blooming.

2. Density of sea urchins per square meter in a quadrat as a function of number of sea otters: -2.5

```
exp(-2.5)
```

```
[1] 0.082085
```

For every additional sea otter, urchin density decreases by 0.08% per square meter

3. Number of tomatoes per plant as a function of kg of fertilizer: 6.24

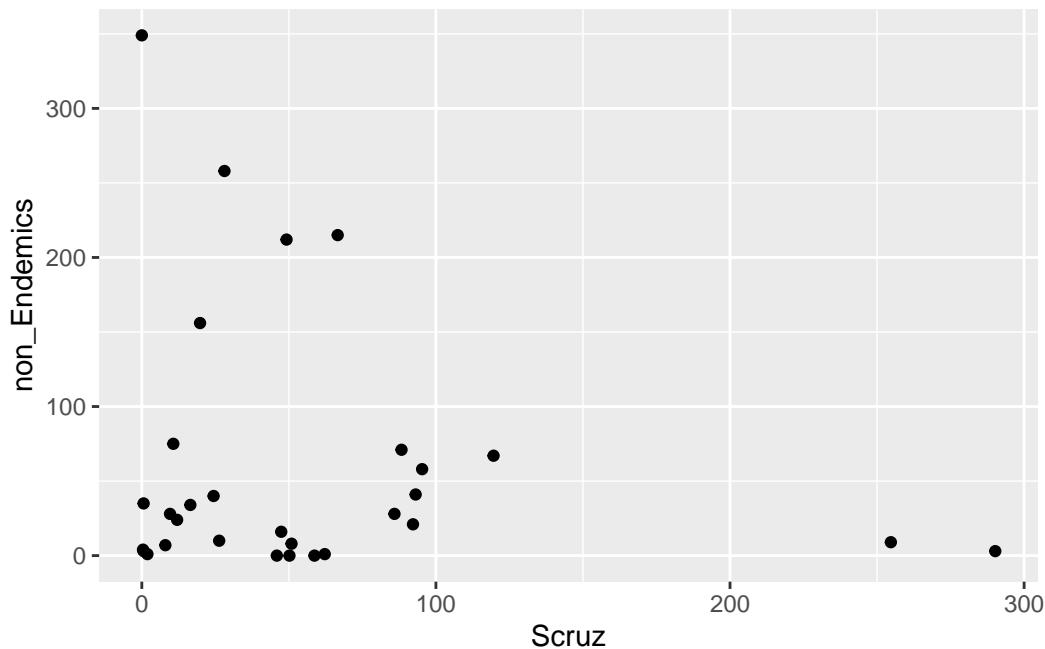
```
exp(6.24)
```

```
[1] 512.8585
```

### Q1.8 Create a non-endemic column

```
gala2 <- gala %>%
  mutate(non_Endemics = Species - Endemics)

ggplot(gala2, aes(x= Scruz, y= non_Endemics))+
  geom_point()
```



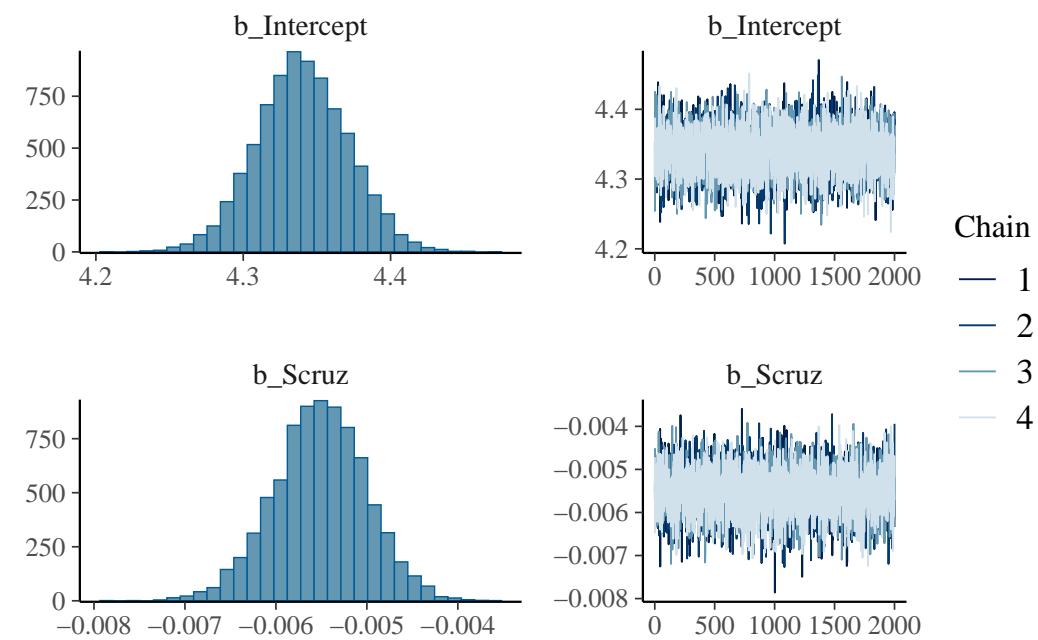
### Q1.9 Run a model of non Endemics ~ distance from Santa Cruz Island

Run a model with a Poisson distribution and log-link function that models the number of non-endemic species as a function of distance from Santa Cruz Island.

```
m.nonedemic <-  
  brm(data = gala2, # Give the model the penguins data  
        # Choose a poisson distribution - THIS IS THE NEW PART!  
        family = poisson(link = "log"),  
        # Specify the model here.  
        non_Endemics ~ 1 + Scruz,  
        # Here's where you specify parameters for executing the Markov chains  
        # We're using similar to the defaults, except we set cores to 4 so the analysis runs fa  
        iter = 6000, warmup = 4000, chains = 4, cores = 4,  
        prior = prior(normal(0, 0.1), class = b),  
        # Save the fitted model object as output - helpful for reloading in the output later  
        file = "output/m.nonenedemic")
```

### Q1.10 Evaluate the output

```
plot(m.nonedemic)
```



```
summary(m.nonedemic)
```

Family: poisson  
Links: mu = log  
Formula: non\_Endemics ~ 1 + Scruz  
Data: gala2 (Number of observations: 30)  
Draws: 4 chains, each with iter = 6000; warmup = 4000; thin = 1;  
total post-warmup draws = 8000

Regression Coefficients:

	Estimate	Est.Error	1-95% CI	u-95% CI	Rhat	Bulk_ESS	Tail_ESS
Intercept	4.34	0.03	4.28	4.40	1.00	5472	5174
Scruz	-0.01	0.00	-0.01	-0.00	1.00	5920	5002

Draws were sampled using sampling(NUTS). For each parameter, Bulk\_ESS and Tail\_ESS are effective sample size measures, and Rhat is the potential scale reduction factor on split chains (at convergence, Rhat = 1).

```
print(m.nonedemic, digits = 4)
```

Family: poisson  
Links: mu = log  
Formula: non\_Endemics ~ 1 + Scruz  
Data: gala2 (Number of observations: 30)  
Draws: 4 chains, each with iter = 6000; warmup = 4000; thin = 1;  
total post-warmup draws = 8000

Regression Coefficients:

	Estimate	Est.Error	1-95% CI	u-95% CI	Rhat	Bulk_ESS	Tail_ESS
Intercept	4.3400	0.0311	4.2787	4.4009	1.0013	5472	5174
Scruz	-0.0055	0.0005	-0.0066	-0.0045	1.0005	5920	5002

Draws were sampled using sampling(NUTS). For each parameter, Bulk\_ESS and Tail\_ESS are effective sample size measures, and Rhat is the potential scale reduction factor on split chains (at convergence, Rhat = 1).

rhat values are 1, posterior chains converge, and the posterior plots are unimodal/uniform

### **Q1.11 Interpret the output**

Interpret the model coefficients by writing a 2-3 sentence results paragraph that answers:

- a) The original value was -0.1
- b)

```
exp(-0.01)
```

```
[1] 0.9900498
```

- c) The percent change is then 0.99%

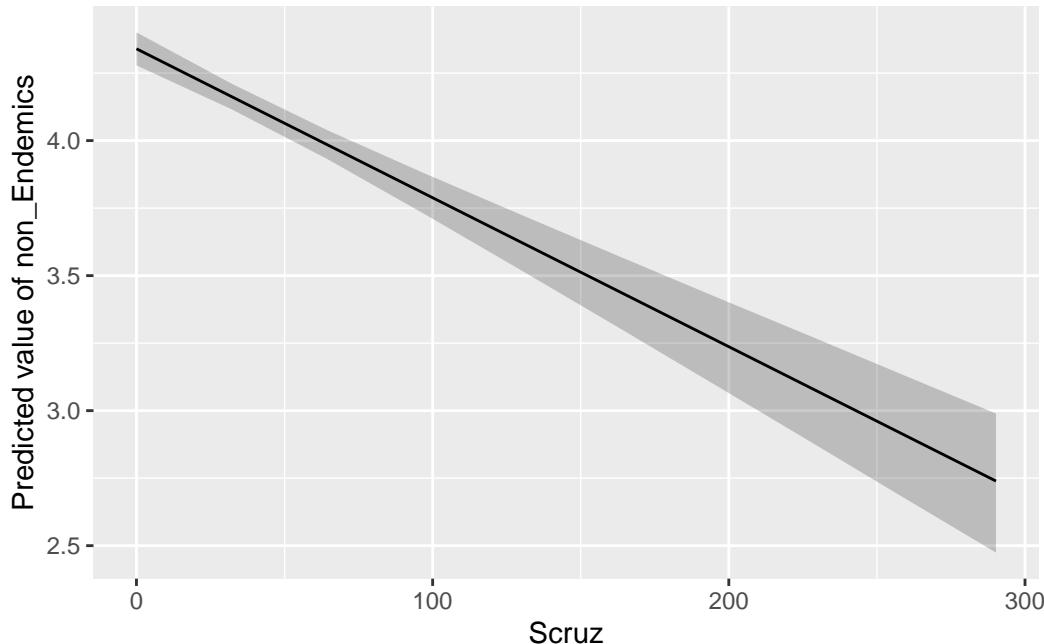
As distance from Santa Cruz Island increases, the frequency of nonendemics decreases. With every unit increase in distance from Santa Cruz island, there is a 1% decrease in non-endemic species.

2. Does it seem like the slope estimate is different from zero? Why?

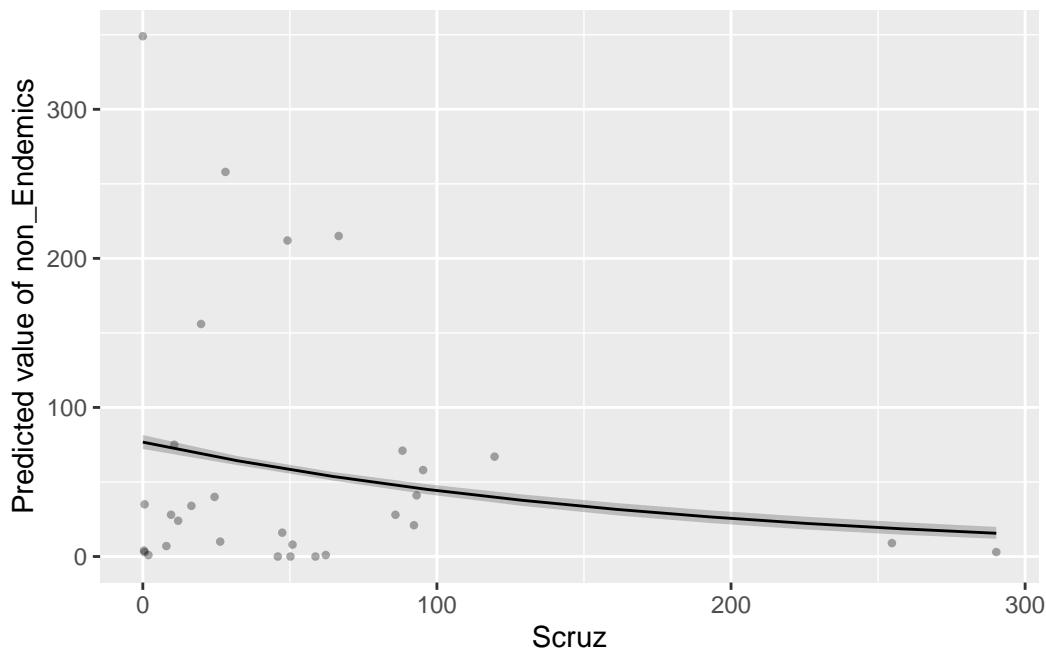
Yes, the 95% CI's exclude 0

### **Q1.12 Plot the posterior**

```
predslog <- estimate_expectation(m.nondenmic, by = 'Scruz', predict = 'link')  
plot(predslog)
```



```
preds <- estimate_expectation(m.nonedemic, by = 'Scruz')
plot(preds, show_data = TRUE)
```

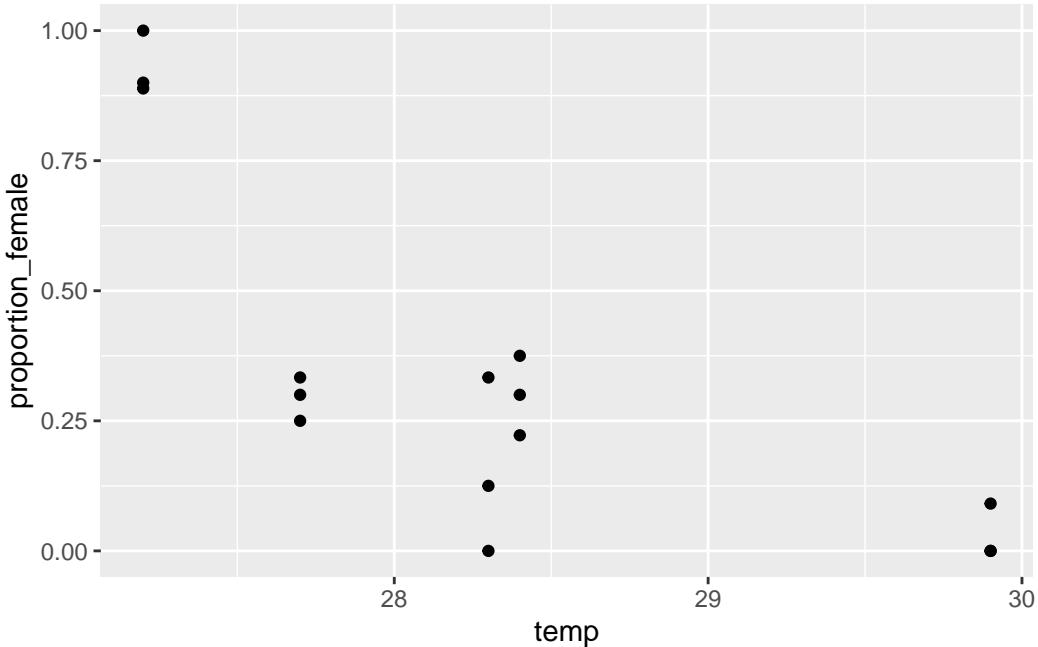


### 1.3 GLM with a logit link

Bring in turtle data

```
turtle <- faraway::turtle %>%
  mutate(total_turtles = male + female,
       proportion_female = female/total_turtles)
```

```
turtle %>%
  ggplot(aes(x = temp, y = proportion_female)) +
  geom_point()
```



```
m.turt <-
  brm(data = turtle, # Give the model the data
       # Choose a binomial distribution - THIS IS THE NEW PART!
       family = binomial(link = "logit"),
       # Specify the model here.
       female | trials(total_turtles) ~ 1 + temp,
       # Here's where you specify parameters for executing the Markov chains
       # We're using similar to the defaults, except we set cores to 4 so the analysis runs faster
       iter = 4000, warmup = 1000, chains = 4, cores = 4,
       # Save the fitted model object as output - helpful for reloading in the output later
       file = "output/m.turt")

summary(m.turt)
```

Family: binomial  
 Links: mu = logit  
 Formula: female | trials(total\_turtles) ~ 1 + temp  
 Data: turtle (Number of observations: 15)  
 Draws: 4 chains, each with iter = 4000; warmup = 1000; thin = 1;  
 total post-warmup draws = 12000

Regression Coefficients:

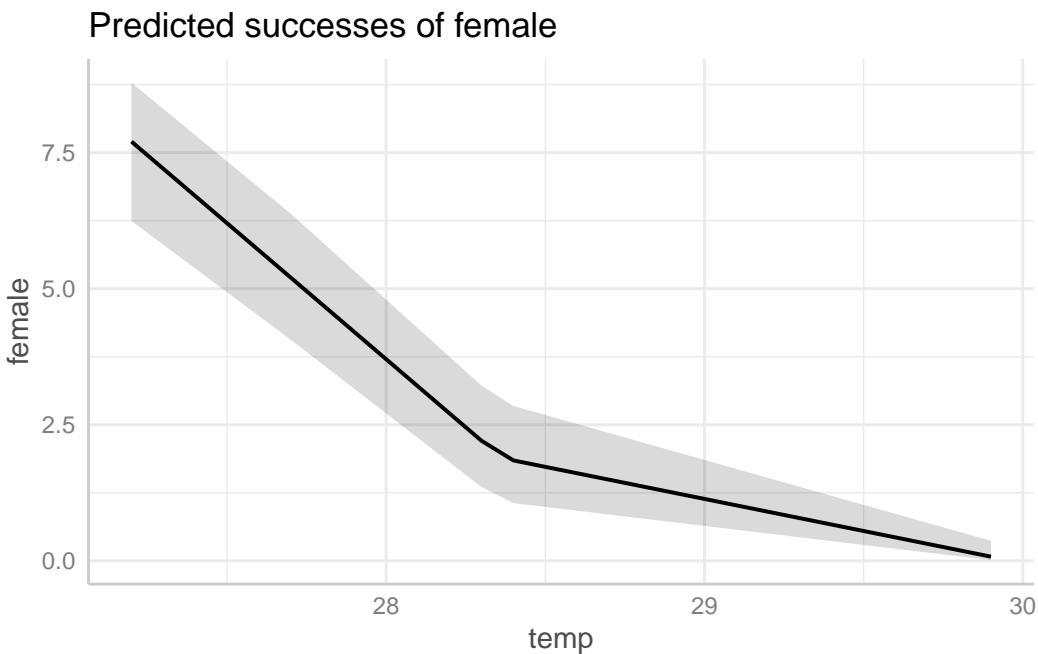
	Estimate	Est.Error	1-95% CI	u-95% CI	Rhat	Bulk_ESS	Tail_ESS
Intercept	62.75	11.92	40.58	86.97	1.00	4347	6472
temp	-2.26	0.43	-3.13	-1.47	1.00	4294	6358

Draws were sampled using sampling(NUTS). For each parameter, Bulk\_ESS and Tail\_ESS are effective sample size measures, and Rhat is the potential scale reduction factor on split chains (at convergence, Rhat = 1).

```
pred <- predict_response(m.turt, condition = c(total_turtles = 10))
```

```
Warning in check_dep_version(dep_pkg = "TMB"): package version mismatch:  
glmmTMB was built with TMB package version 1.9.18  
Current TMB package version is 1.9.16  
Please re-install glmmTMB from source or restore original 'TMB' package (see '?reinstalling'
```

```
plot(pred)
```



### Q2.1 Fixed effects vs random effects

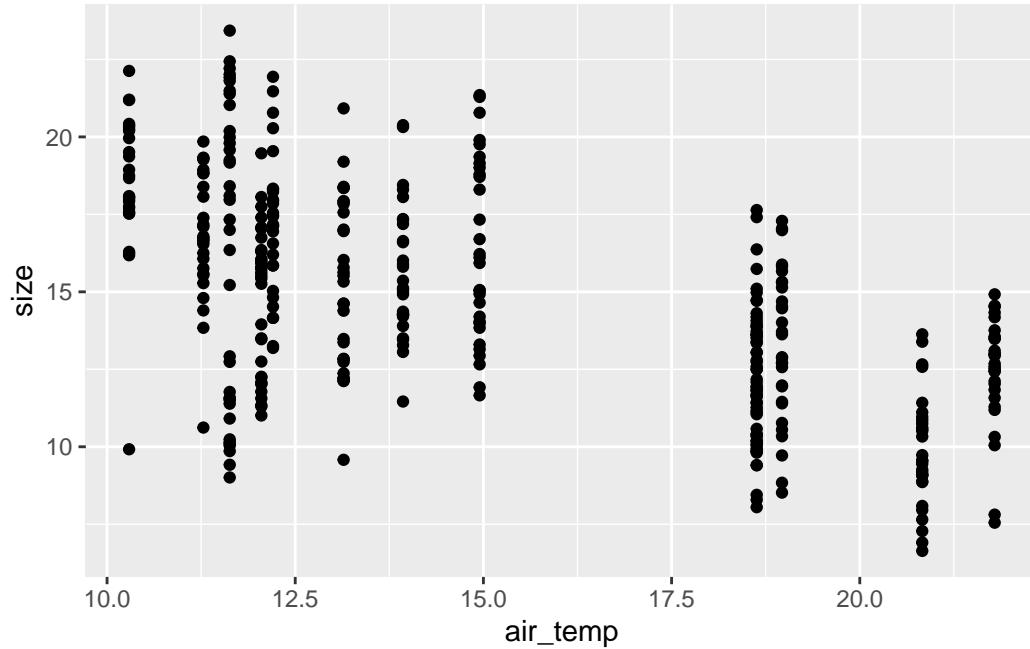
For the following variables in the model examples below, denote which variables are the fixed effects and which could be accounted for as random effects (some variables could be either, but consider then as being eligible to be random effects):

1. Student high school graduation rates as a function of: parental income, state of residence, and school district
  - Fixed: parental income
  - Random: State and district
2. Density of kelp as a function of: latitude, site, transect number, and density of sea urchins
  - Fixed: latitude, density of sea urchins
  - Random: site, transect number
3. Probability of whale giving birth as a function of: age, annual temperature, year, individual ID
  - Fixed: temperature, age
4. Random: individual Id, year

## Fiddler Crabs

```
pie_crab <- lterdatasampler::pie_crab %>%
  mutate(site = as.factor(site))

pie_crab %>%
  ggplot(aes(x = air_temp, y = size)) +
  geom_point()
```



```
m.watertemp <-
  brm(data = pie_crab, # Give the model the penguins data
       # Use a gamma distribution
       family = Gamma(link = "log"),
       # Specify the model here.
       size ~ 1 + water_temp,
       # Here's where you specify parameters for executing the Markov chains
       # We're using similar to the defaults, except we set cores to 4 so the analysis runs faster
       iter = 2000, warmup = 1000, chains = 4, cores = 4,
       # Save the fitted model object as output - helpful for reloading in the output later
       file = "output/m.watertemp")

  print(m.watertemp, digits = 3)
```

Family: gamma  
 Links: mu = log  
 Formula: size ~ 1 + water\_temp  
 Data: pie\_crab (Number of observations: 392)  
 Draws: 4 chains, each with iter = 2000; warmup = 1000; thin = 1;  
 total post-warmup draws = 4000

Regression Coefficients:

	Estimate	Est.Error	l-95%	CI	u-95%	CI	Rhat	Bulk_ESS	Tail_ESS
Intercept	3.355	0.056	3.246	3.467	1.000	4372	3105		
water_temp	-0.038	0.003	-0.045	-0.032	1.000	4570	3015		

Further Distributional Parameters:

	Estimate	Est.Error	l-95%	CI	u-95%	CI	Rhat	Bulk_ESS	Tail_ESS
shape	23.259	1.619	20.191	26.516	1.004	2931	2625		

Draws were sampled using sampling(NUTS). For each parameter, Bulk\_ESS and Tail\_ESS are effective sample size measures, and Rhat is the potential scale reduction factor on split chains (at convergence, Rhat = 1).

```
m.watertemp.site <-
  brm(data = pie_crab, # Give the model the penguins data
       # Use a gamma distribution
       family = Gamma(link = "log"),
       # Specify the model here.
       size ~ 1 + water_temp + (1|site),
       # Here's where you specify parameters for executing the Markov chains
       # We're using similar to the defaults, except we set cores to 4 so the analysis runs faster
       iter = 2000, warmup = 1000, chains = 4, cores = 4,
       # Save the fitted model object as output - helpful for reloading in the output later
       file = "output/m.watertemp.site")

print(m.watertemp.site, digits = 3)
```

Family: gamma  
 Links: mu = log  
 Formula: size ~ 1 + water\_temp + (1 | site)  
 Data: pie\_crab (Number of observations: 392)  
 Draws: 4 chains, each with iter = 2000; warmup = 1000; thin = 1;  
 total post-warmup draws = 4000

Multilevel Hyperparameters:  
 ~site (Number of levels: 13)

	Estimate	Est.Error	l-95%	CI	u-95%	CI	Rhat	Bulk_ESS	Tail_ESS
sd(Intercept)	0.123	0.033	0.075	0.208	1.002	897	1288		

Regression Coefficients:

	Estimate	Est.Error	l-95%	CI	u-95%	CI	Rhat	Bulk_ESS	Tail_ESS
Intercept	3.368	0.181	3.014	3.734	1.002	1058	1594		
water_temp	-0.039	0.010	-0.059	-0.020	1.001	1158	1744		

Further Distributional Parameters:

	Estimate	Est.Error	l-95% CI	u-95% CI	Rhat	Bulk_ESS	Tail_ESS
shape	30.075	2.167	25.932	34.506	1.002	2738	2321

Draws were sampled using sampling(NUTS). For each parameter, Bulk\_ESS and Tail\_ESS are effective sample size measures, and Rhat is the potential scale reduction factor on split chains (at convergence, Rhat = 1).

## Q2.2 What is the effect of water\_temp on crab size on the response scale?

Interpret the output by writing a 2-3 sentence results paragraph that answers:

1. What is the effect of water\_temp on creab size? Report the a) original output on the log scale, b) your backtransformed value, and c) the percent change that this translates to. Describe the effect using the proper units.
2. Does it seem like the slope estimate is different from zero? Why?

## Q2.3 Compare WAIC and PSIS of the two models

Between the two models, which has the better predictive power?

```
loo(m.watertemp)
```

Computed from 4000 by 392 log-likelihood matrix.

```
          Estimate    SE
elpd_loo   -984.7 12.1
p_loo        2.8  0.2
looic     1969.5 24.3
-----
```

MCSE of elpd\_loo is 0.0.

MCSE and ESS estimates assume MCMC draws (r\_eff in [0.7, 1.1]).

All Pareto k estimates are good (k < 0.7).

See help('pareto-k-diagnostic') for details.

```
loo(m.watertemp.site)
```

```
Computed from 4000 by 392 log-likelihood matrix.
```

	Estimate	SE
elpd_loo	-938.5	13.7
p_loo	12.0	0.8
looic	1876.9	27.3

-----  
MCSE of elpd\_loo is 0.1.  
MCSE and ESS estimates assume MCMC draws (r\_eff in [0.5, 1.6]).

All Pareto k estimates are good (k < 0.7).  
See help('pareto-k-diagnostic') for details.

```
waic(m.watertemp)
```

```
Computed from 4000 by 392 log-likelihood matrix.
```

	Estimate	SE
elpd_waic	-984.7	12.1
p_waic	2.8	0.2
waic	1969.4	24.3

```
waic(m.watertemp.site)
```

```
Computed from 4000 by 392 log-likelihood matrix.
```

	Estimate	SE
elpd_waic	-938.4	13.7
p_waic	12.0	0.8
waic	1876.8	27.3

The second model that includes site as a random effect is the better model (lower loo and waic values)

```
preds <- predict_response(m.watertemp.site,
                           interval = "prediction",
                           terms = "site",
```

```
type = "random")  
plot(preds)
```

