

Computational hyperspectral SPIM for quantitative multicolor imaging

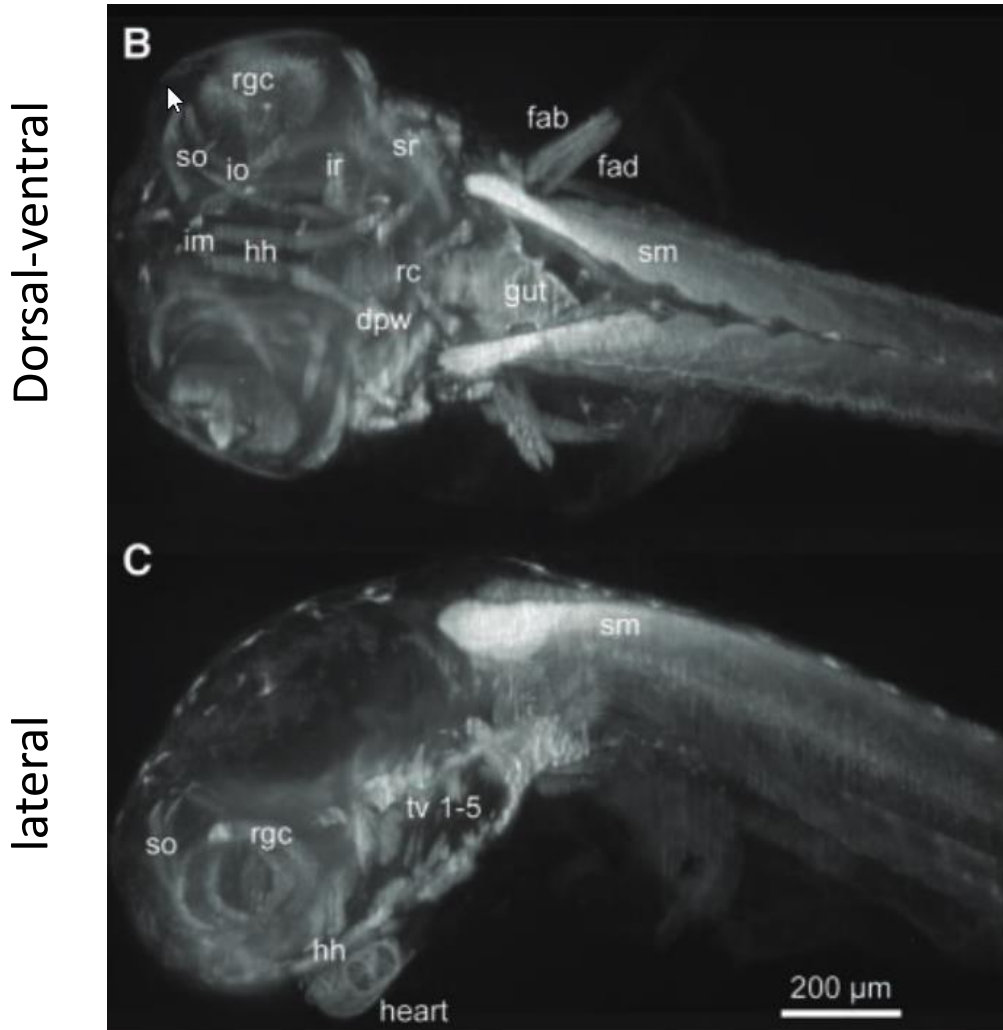
Nicolas Ducros^{1, 2}

¹CREATIS, Univ Lyon, INSA-Lyon, UCB Lyon 1, CNRS, Inserm, CREATIS UMR 5220, U1206, Lyon, France

²IUF, Institut Universitaire de France

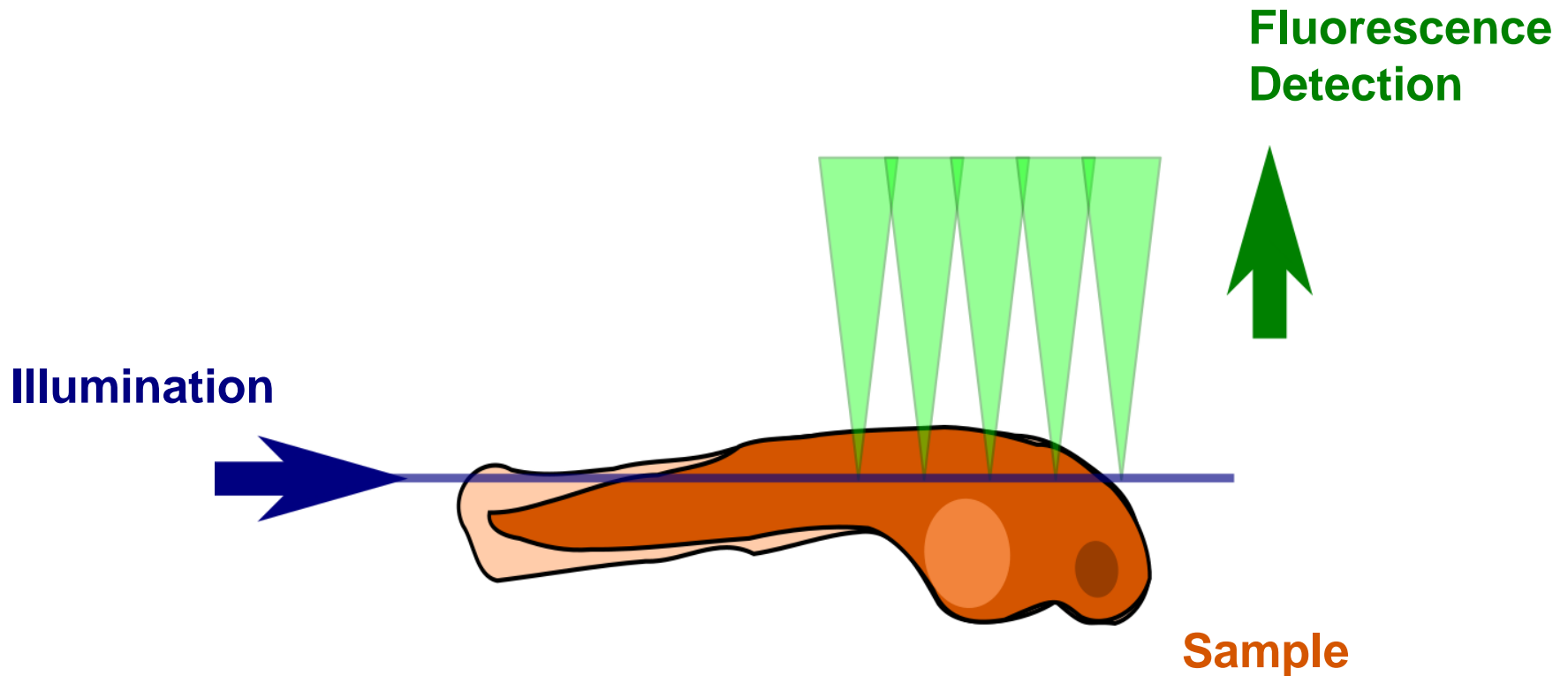
Joint work with: T Baudier, S Crombez, Chloé Exbrayat-Heritier, A Lorente-Mur, L Mahieu-Williams, C Ray, F Ruggiero

Single Plane Illumination Microscopy (SPIM)



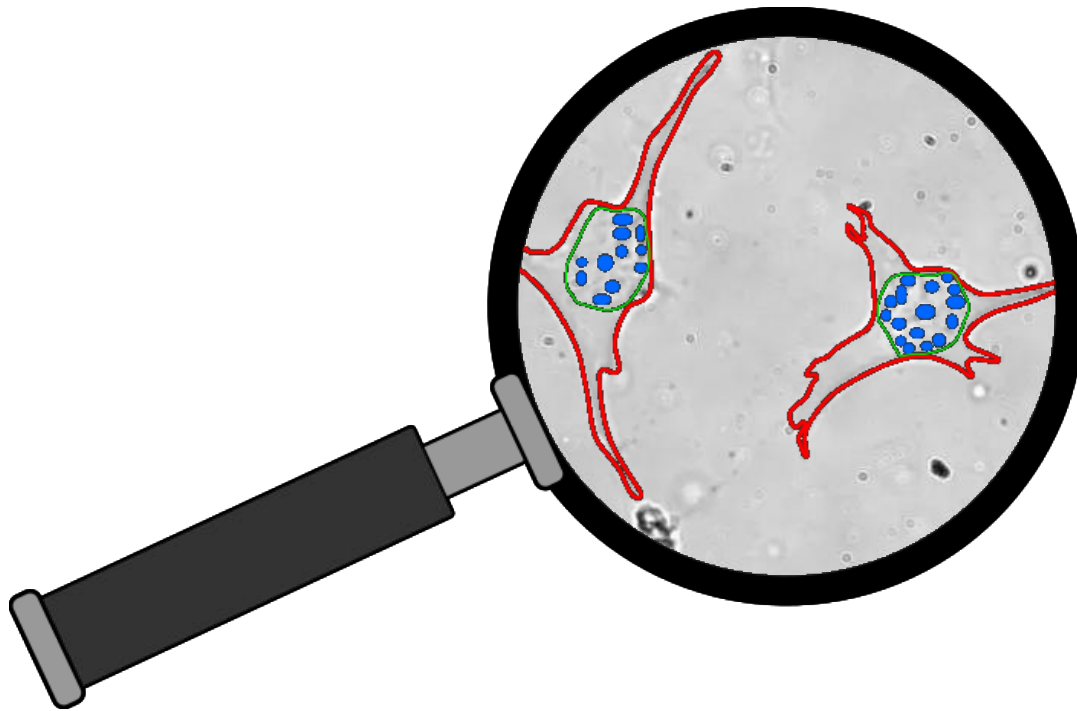
- ✓ Wide field ($\sim\text{mm}^3$),
- ✓ High res ($\sim 6\ \mu\text{m}$)
- ✓ Fast (10 fps)
- ✓ Fluorescence samples
- ✓ GFP-labeled transgenic embryos
- ✓ Developmental study

[J Huisken *et al*,
Science, **305**, (2004)]

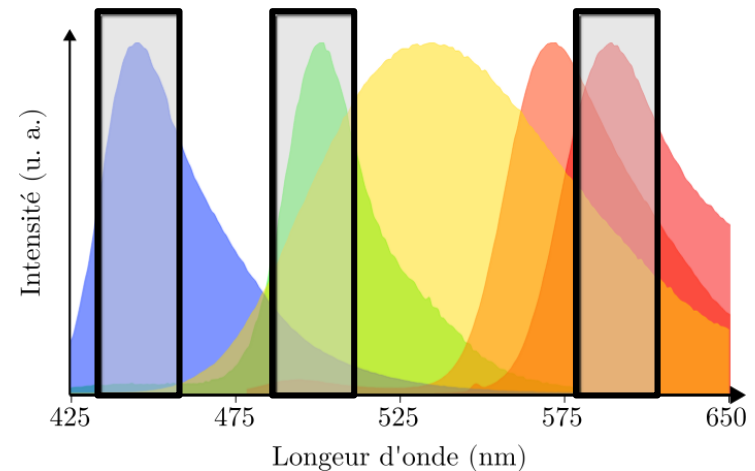


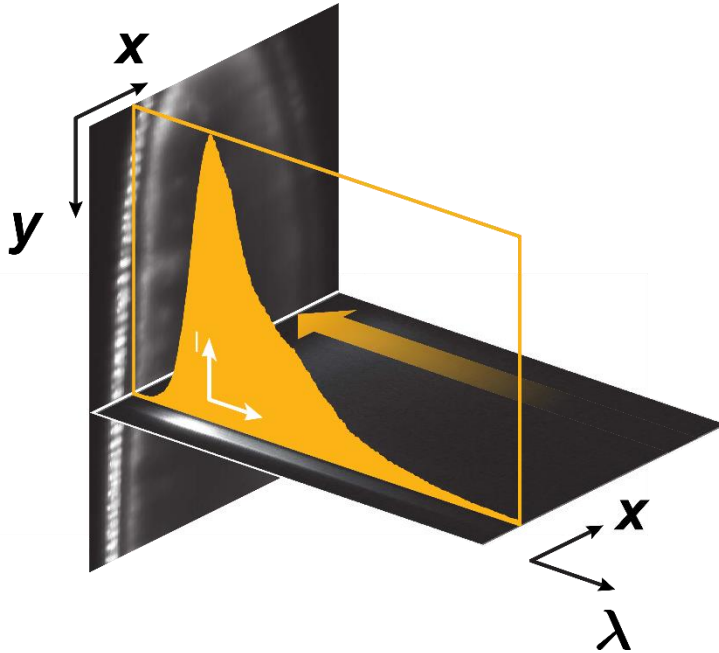
- ✓ Optical sectioning (6 μm resolution) as deep as 500 μm)
- ✓ Low photobleaching

[Image adapted from [Wikimedia Commons](#)]

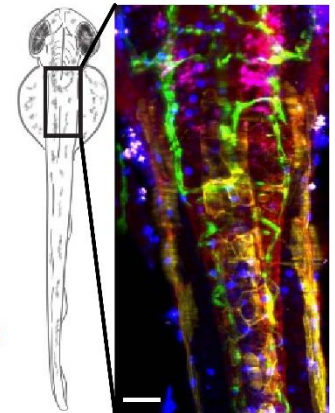
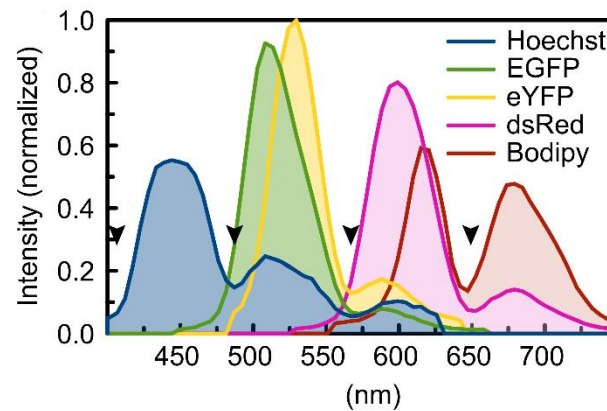


- ✓ Many photons are rejected
- ✓ Overlapping fluorophores cannot be separated
- ✓ Undesired fluorophores cannot be rejected

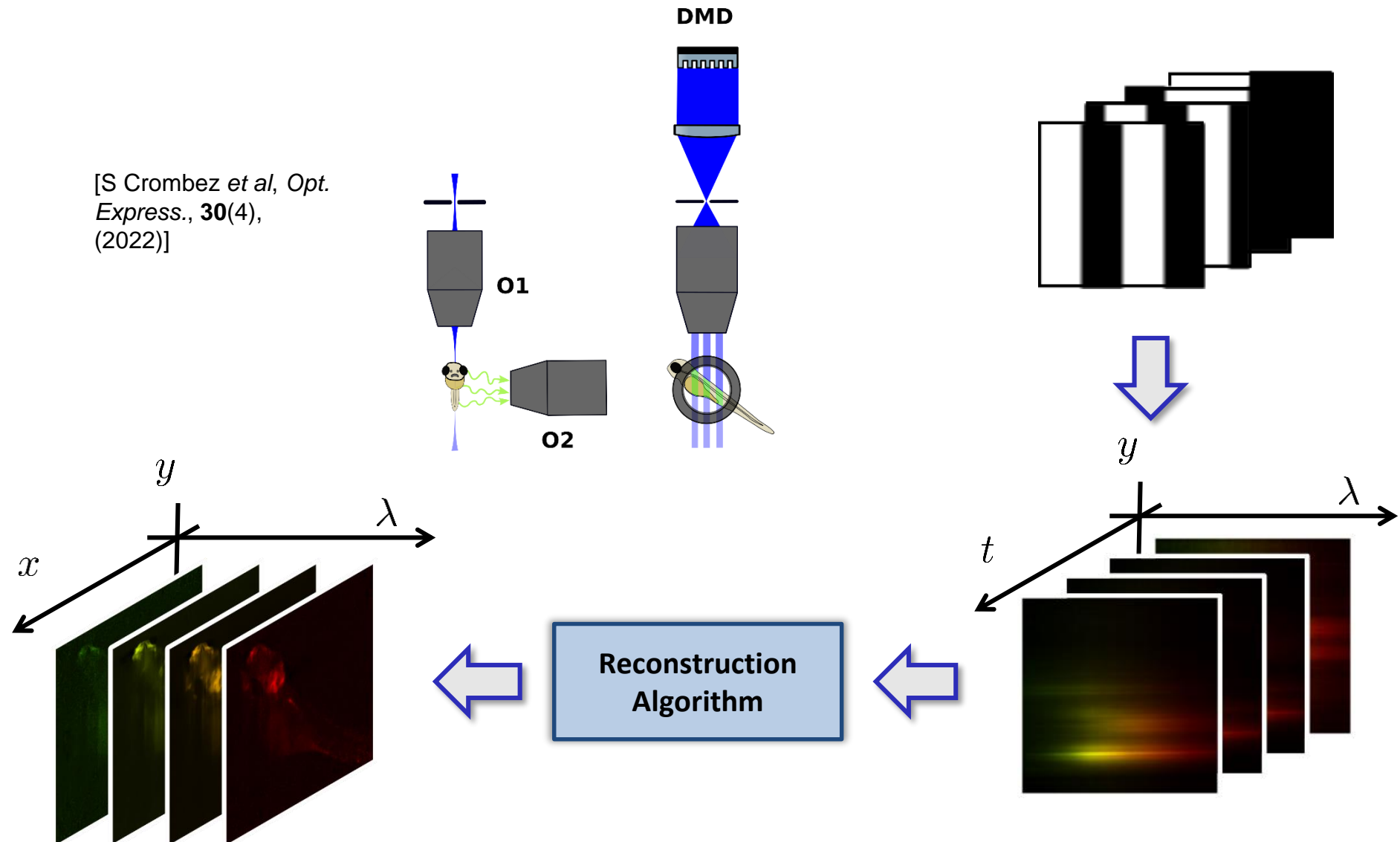


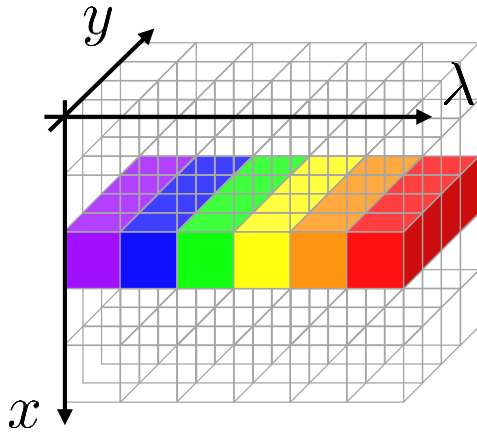


[W Jahr *et al*, *Nat. Commun.*, **6**, (2015)]

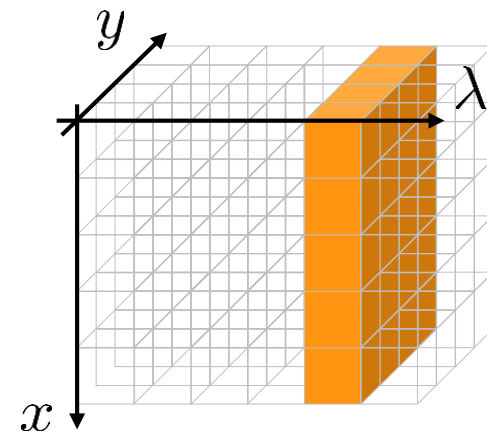


[S Crombez *et al*, *Opt. Express.*, **30**(4), (2022)]

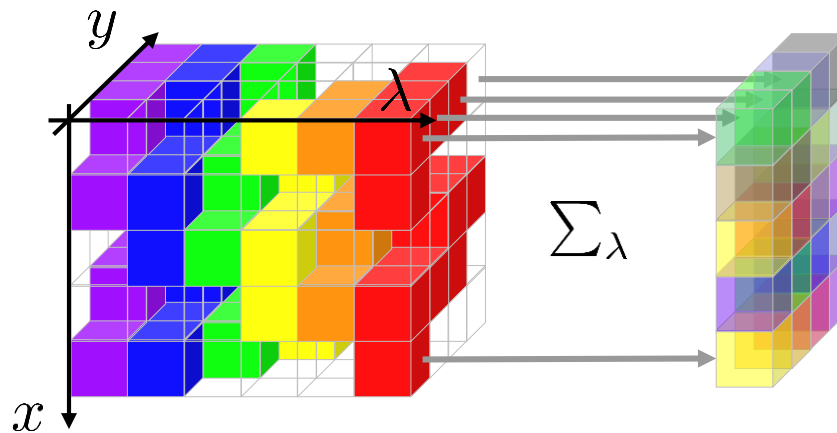




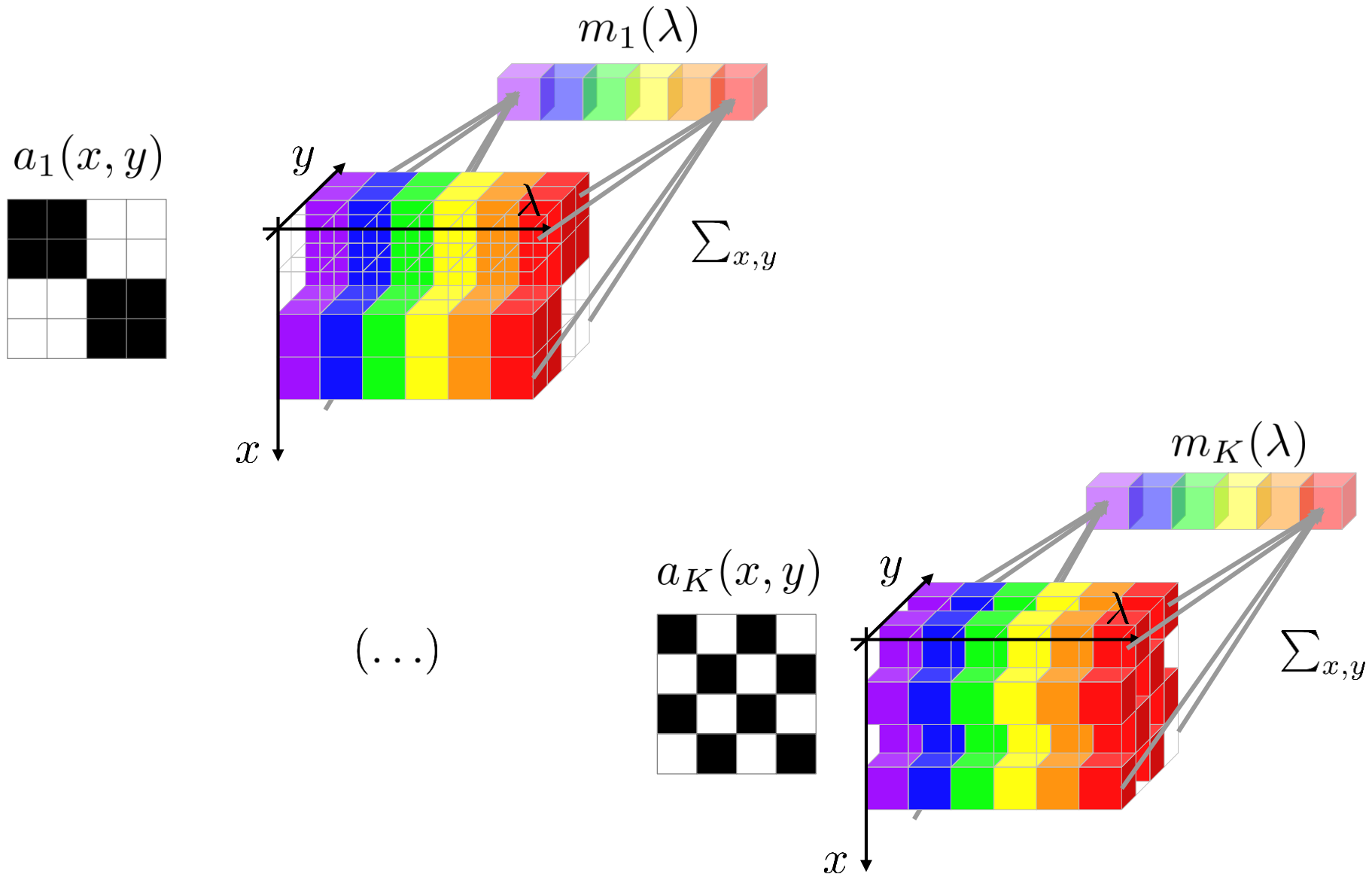
Pushbroom

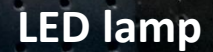


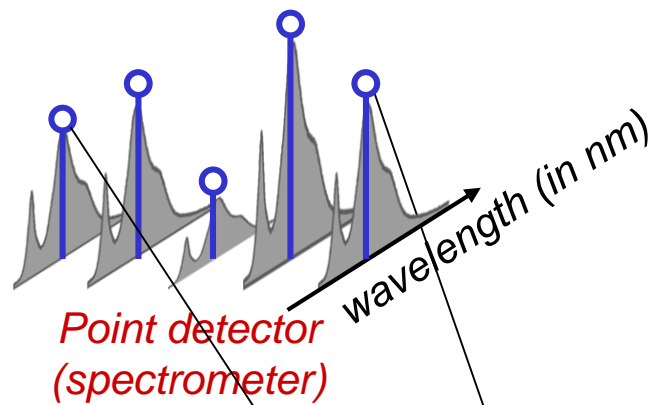
Filter-based



Computational

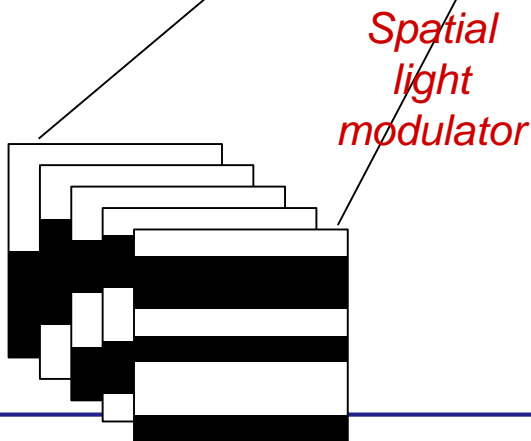






$$\mathbf{m}_\lambda = [m_{1,\lambda}, \dots, m_{K,\lambda}]^\top \in \mathbb{R}^K$$

$$\mathbf{P} = [\mathbf{p}_1^\top, \dots, \mathbf{p}_K^\top]^\top \in \mathbb{R}^{K \times N}$$



➤ Linear model

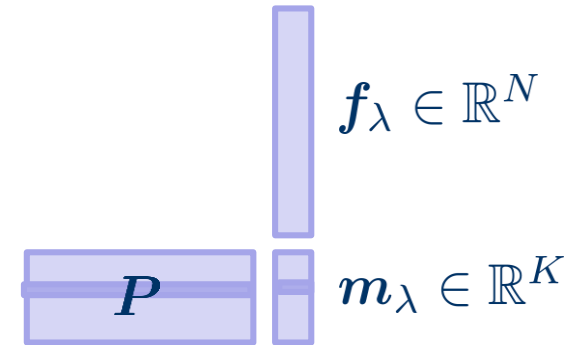
$$\mathbf{m}_\lambda = \mathbf{P} \mathbf{f}_\lambda$$

1. “Weight design”: How to choose the patterns \mathbf{P} ?
2. Reconstruction: How to recover the image \mathbf{f} ?

➤ Fast acquisitions

- ❖ Sequential measurements lead to long acquisition times
- Limit to a few patterns

$$K \ll N$$



➤ Compressed sensing

- ❖ Choose a “random” P
- ❖ Recover f from m via (constrained/LI) optimization

$$\min_f \|m - Pf\|_2^2 + \alpha \mathcal{R}(f)$$

○ Total Variation (TV)

$$\mathcal{R}_{\text{TV}}(f) = \|\nabla f\|_1$$

... which requires iterative algorithms

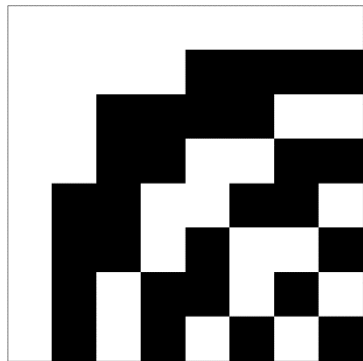
→ Fast acquisitions: long reconstructions!

256 × 256 image
 $M = 6,500$ random measurements

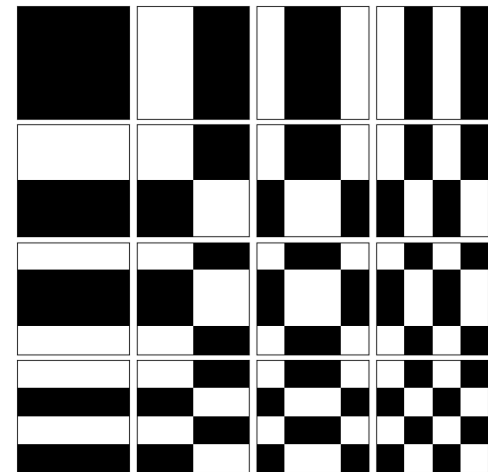


[Duarte *et. al*, IEEE SPM, 2008]

- In the case $K = N$, choose Hadamard patterns



$$K = N = 8$$



$$K = N = 4^2$$

$$P^{\top} P = NI$$

- **Hadamard patterns are optimal for additive white Gaussian noise**

$$m_i \sim \mathcal{G}(\mu = 0, \sigma^2) \quad 1 \leq k \leq N$$

❖ Raster scan

$$\text{var}(f_n^*) = \sigma^2$$

1	0	0	0
0	1	0	0
0	0	1	0
0	0	0	1

$$P^\top P = NI$$

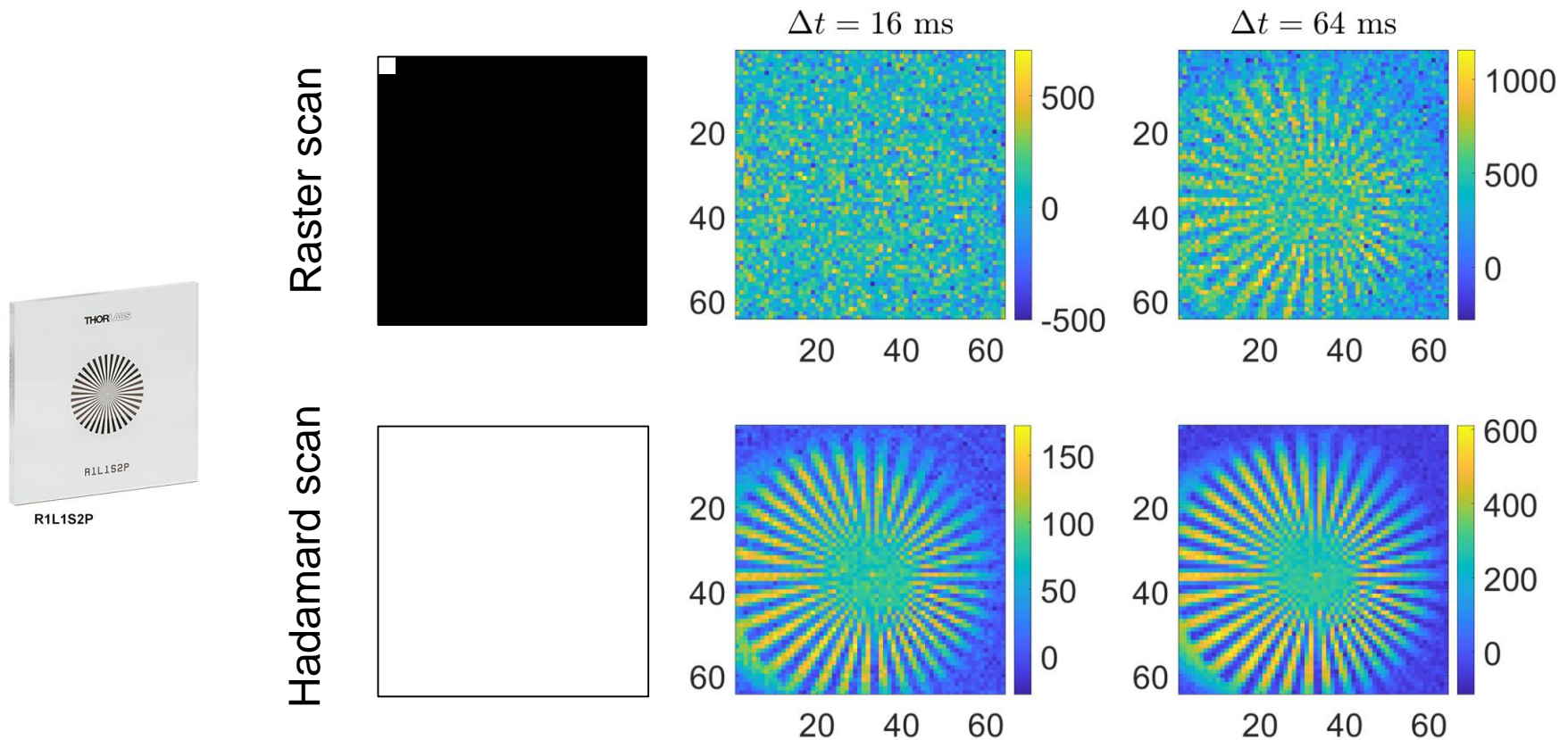
$$f = \frac{1}{N} P^\top m$$

❖ Hadamard

$$\text{var}(f_n^*) = \frac{1}{N} \sigma^2$$

+	+	+	+
+	+	-	-
+	-	-	+
+	-	+	-

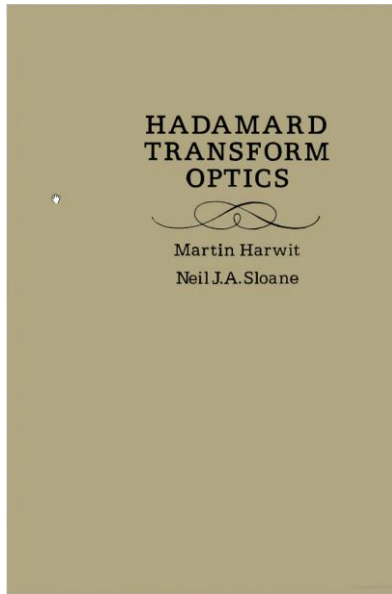
➤ Hadamard optimality, a. k. a. Felgett's advantage



[N. Ducros *et al.*,
working paper, 2021]

The Felgett's Advantage for Spectroscopy

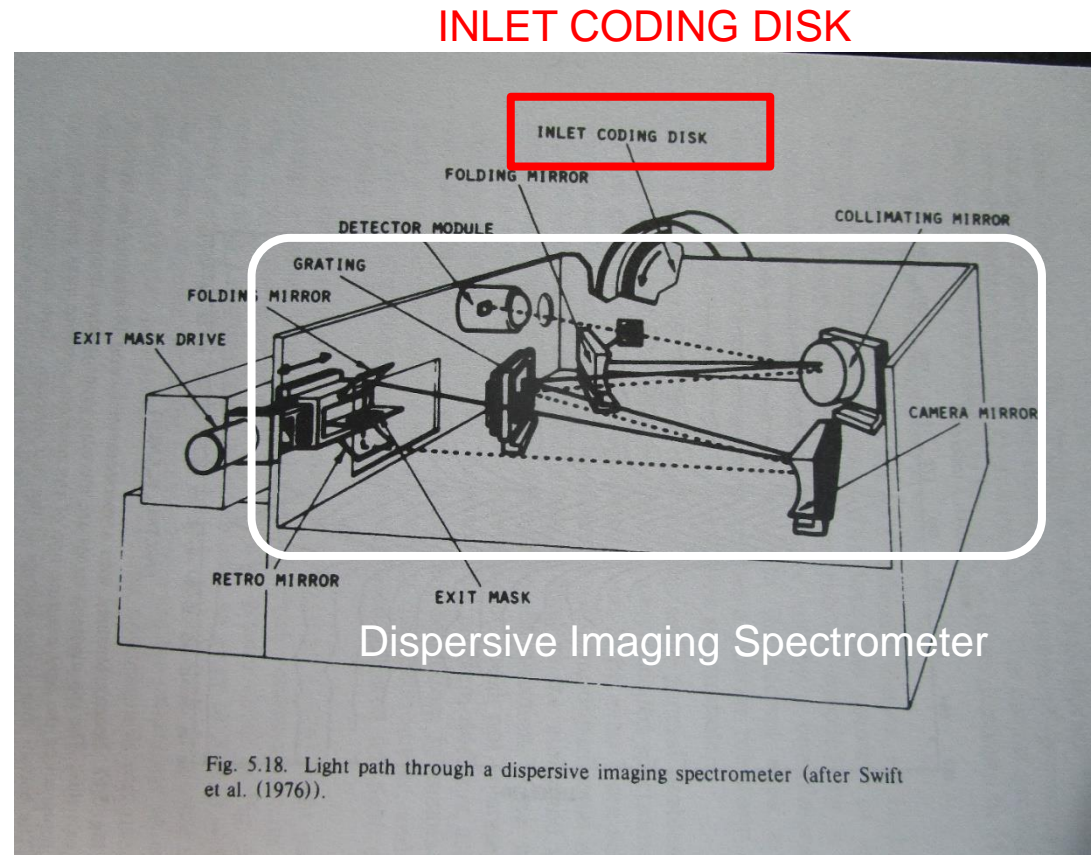
15



M. Harwit,
Cornell University,
Center for Radiophysics and
Space Research

N. Sloane,
Bell Laboratories

(1979)



- ✓ Only one spatial dimension should be compressed, not two.

$$m_k(y, \lambda) = \int p_k(x) f(x, y, \lambda) \, dx, \quad 1 \leq k \leq K$$

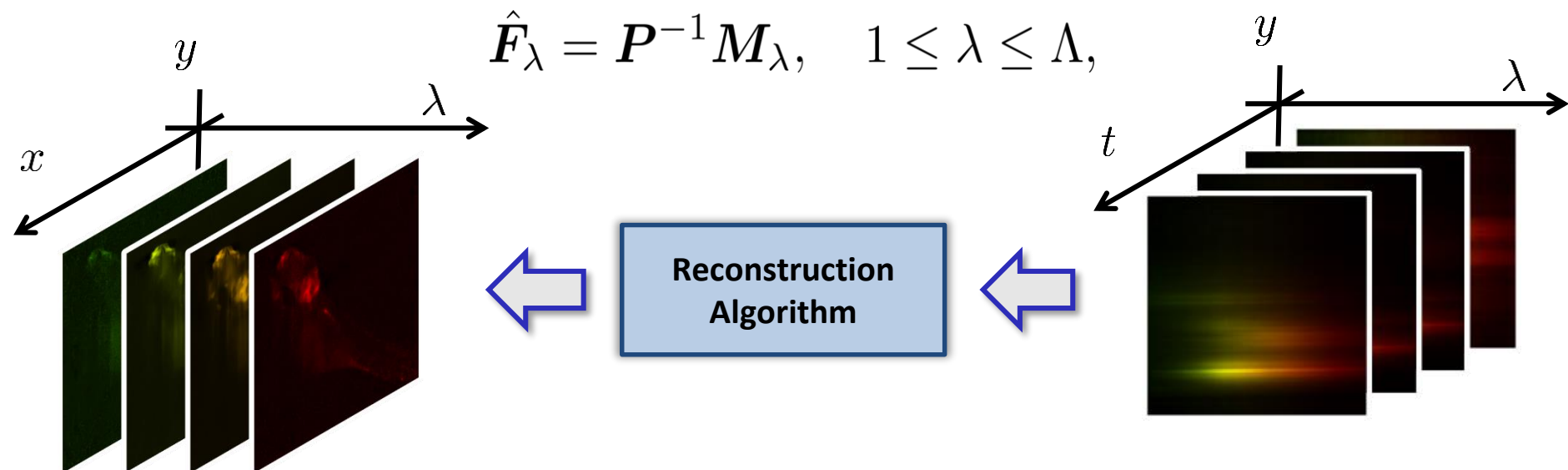
- ✓ Felgett's advantage (contrary to original hspim)
- ✓ No extra compression (as in hspi)

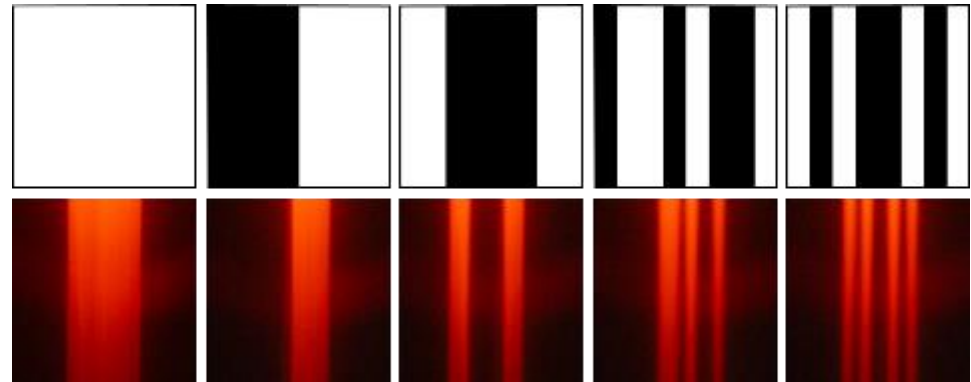
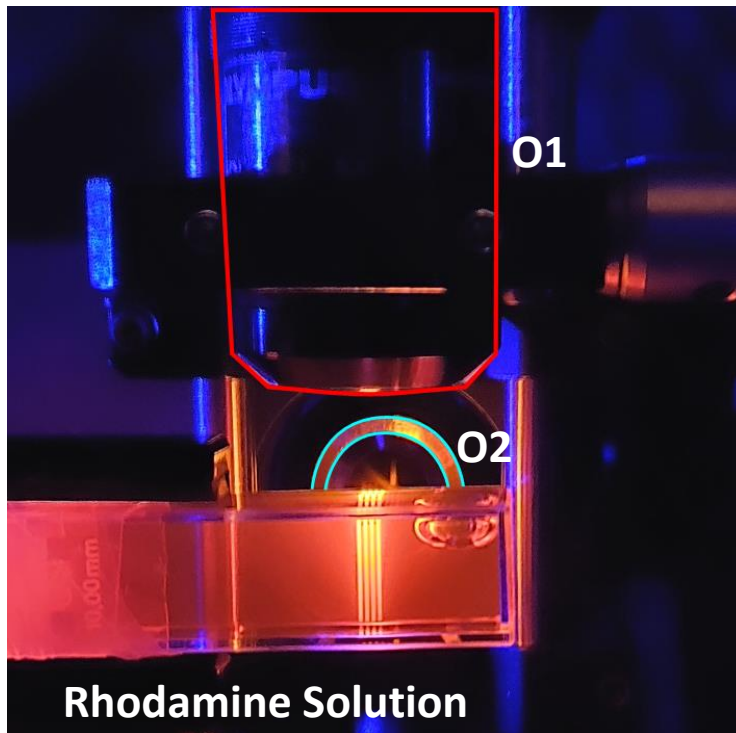
➤ Image formation model

$$M_\lambda = PF_\lambda + E, \quad 1 \leq \lambda \leq \Lambda,$$

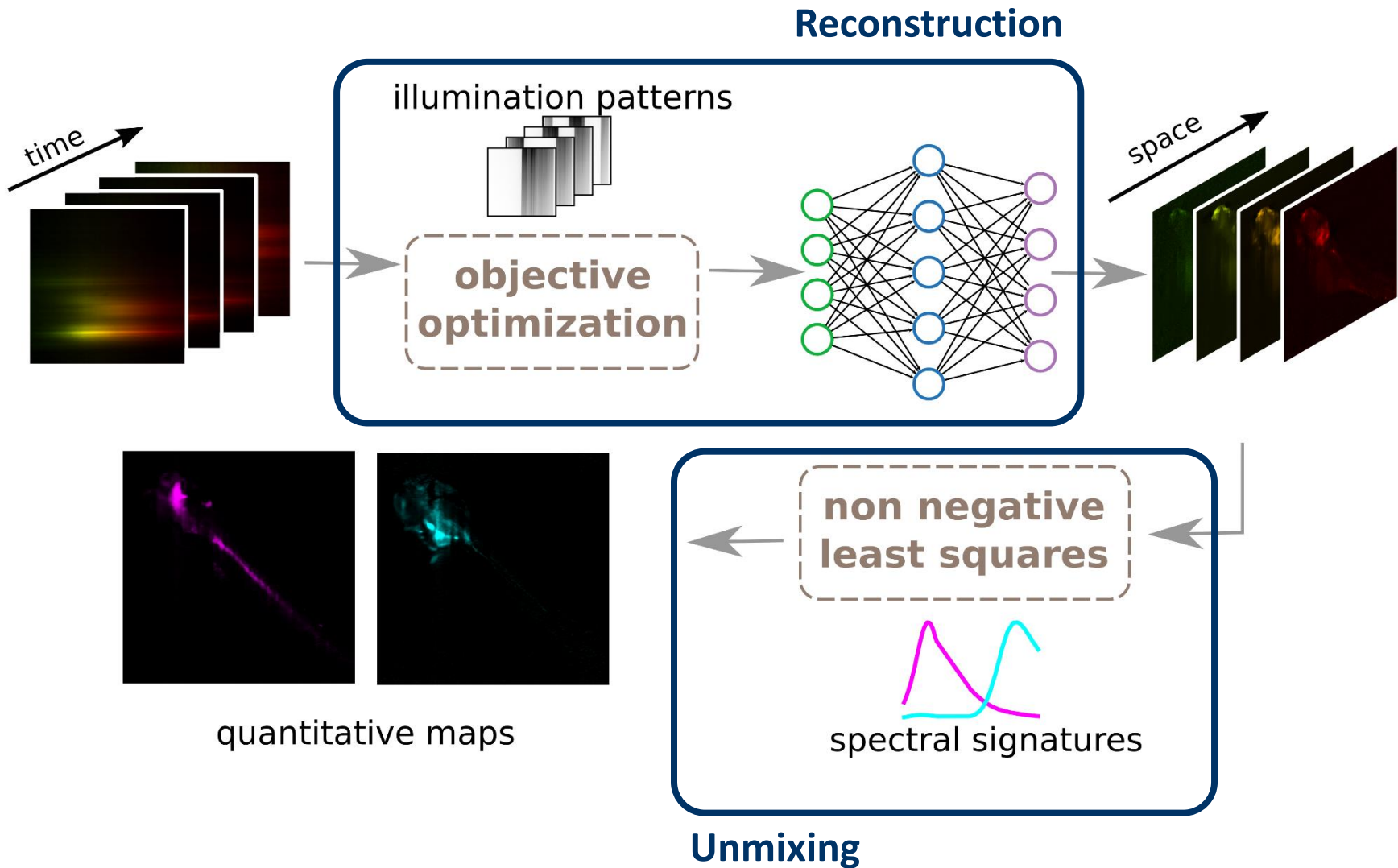
$K \times N_y$ measurement
 $K \times K$ pattern matrix
 $K \times N_y$ hypercube slice

➤ (Basic) Image reconstruction





- ✓ Light patterns need to be calibrated
- ✓ “Good” patterns (e.g., matrix with low condition number) are highly desirable



➤ **Learnt reconstruction**

❖ Reconstruction (inference)

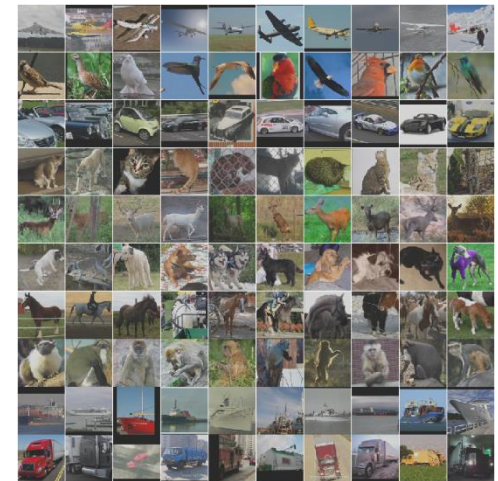
$$\mathbf{F}_\lambda^* = \mathcal{H}_{\theta^*}(\mathbf{M}_\lambda)$$

↖ *network
parameters*

❖ Learning

$$\theta^* \in \arg \min_{\theta} \frac{1}{L} \sum_{\ell} \|\mathcal{H}(\theta; \mathbf{M}^{\ell}) - \mathbf{F}^{\ell}\|_2^2$$

STL-10 dataset



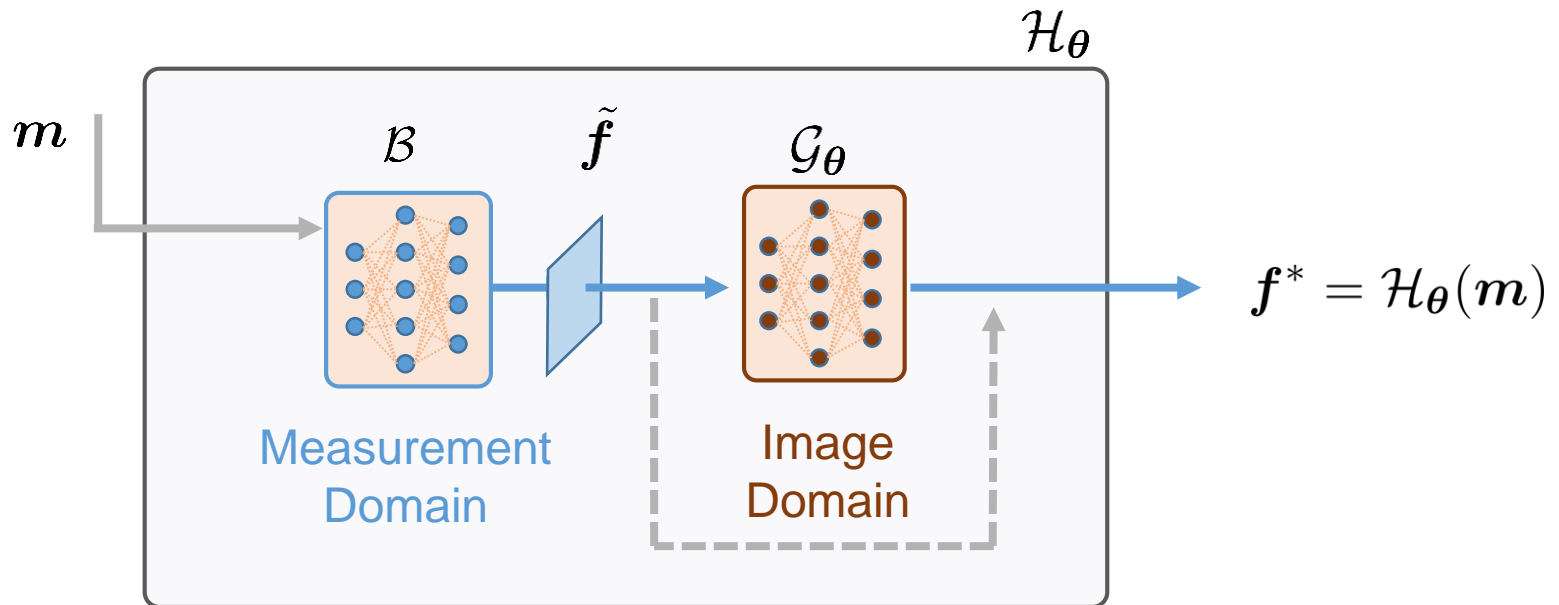
➤ **Computation times**

- ❖ Training phase is slow, i.e., several hours or days
- ❖ Inference is fast, i.e., tens or hundreds of milliseconds

➤ How to choose the ‘model’ \mathcal{H} ?

- ❖ Black box
- ❖ Two step
- ❖ Unrolled, plug & play

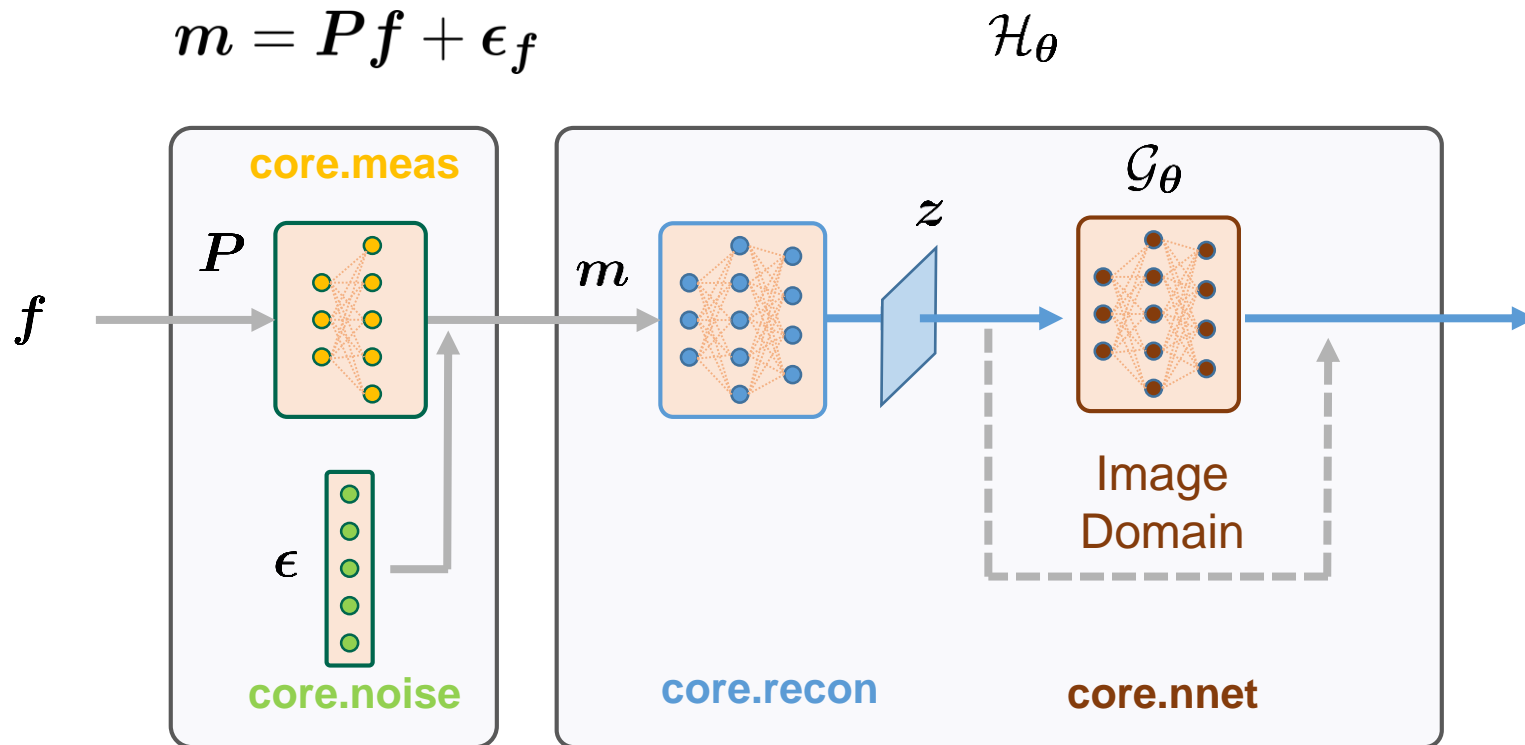
$$F_{\lambda}^* = \mathcal{H}_{\theta^*}(M_{\lambda}) = (\mathcal{G}_{\theta^*} \circ \mathcal{B})(M_{\lambda})$$



$$\mathcal{B}(M_{\lambda}) = \Sigma P^{\top} (P \Sigma P^{\top} + \Gamma)^{-1} M_{\lambda}$$

➤ Core components

<https://spyrit.readthedocs.io/en/master/>

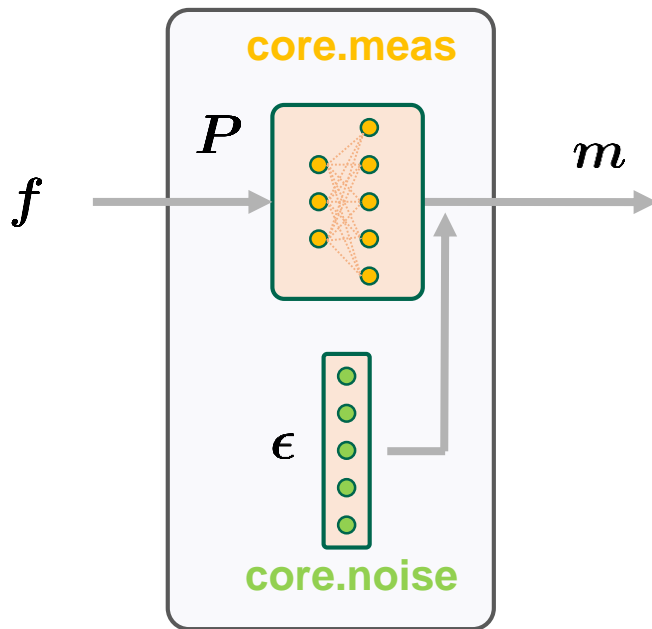


$$\theta^* \in \arg \min_{\theta} \frac{1}{L} \sum_{\ell} \|(\mathcal{G}_{\theta} \circ \mathcal{H})(f^{\ell}) - f^{\ell}\|_2^2$$

➤ Core components

<https://spyrit.readthedocs.io/en/master/>

$$m = Pf + \epsilon_f$$



Data simulation

```
from spyrit.core.meas import HadamSplit
from spyrit.core.noise import Poisson
```

```
meas_op = HadamSplit(512, 128)
noise_op = Poisson(meas_op, 100)
m = noise_op(f)
```

➤ Spectral unmixing

- ❖ Assuming C fluorophores (or combination of fluorophores, e.g., autofluorescence)

$$\min_A \|F - AS\|_2^2 + \alpha \mathcal{R}(A)$$

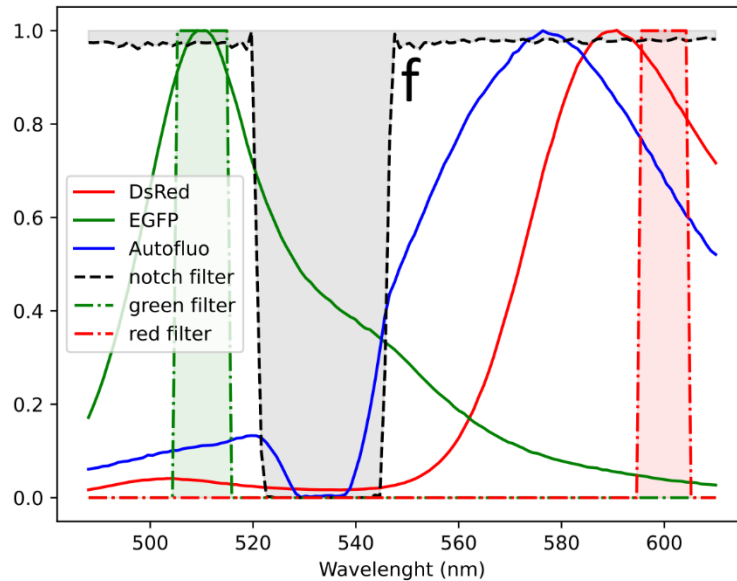
$N \times \Lambda$ hypercube

$C \times \Lambda$ spectra

$N \times C$ abundance maps

$N = K \times N_y$ pixels

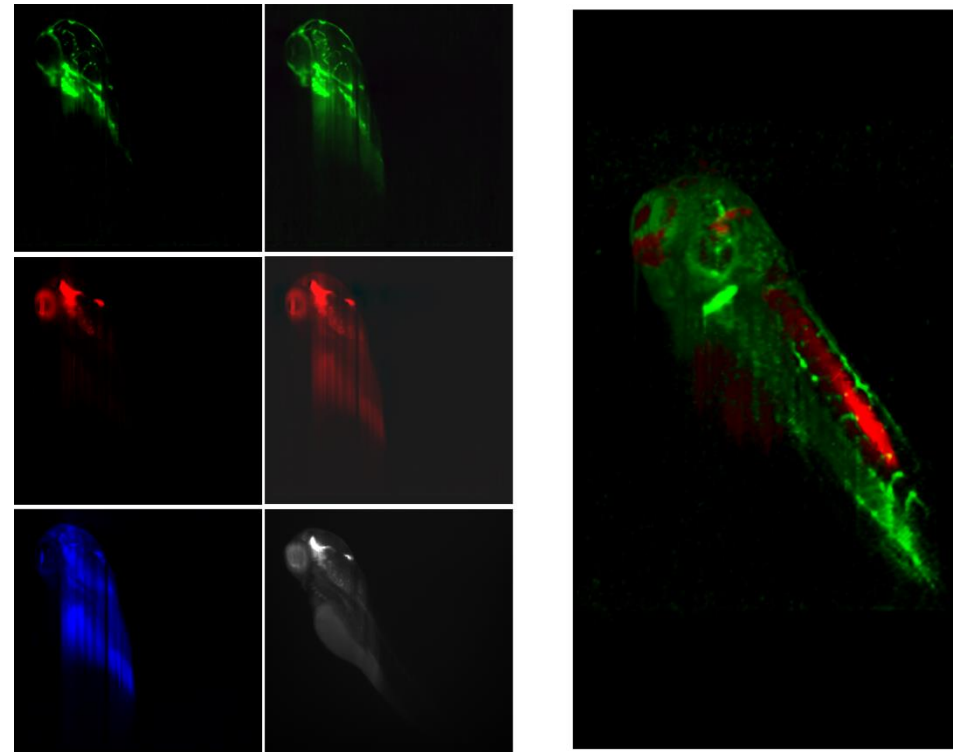
- ❖ Regularization can enforce prior knowledge about the abundance maps or be “learnt” from “examples”



Filter

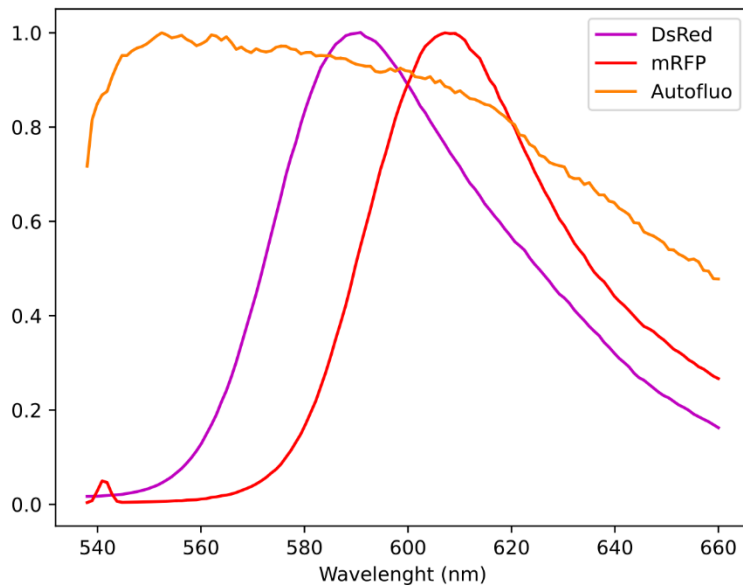
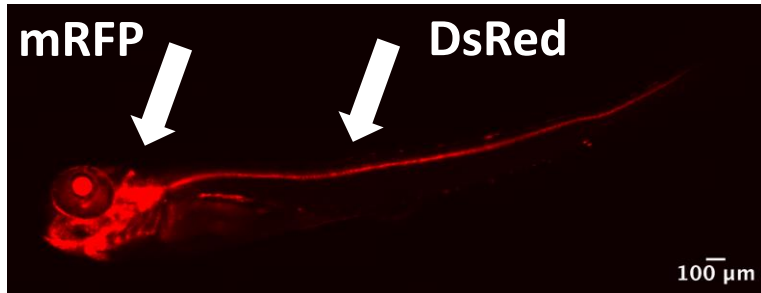
Spectral

Autofluo free (3D)

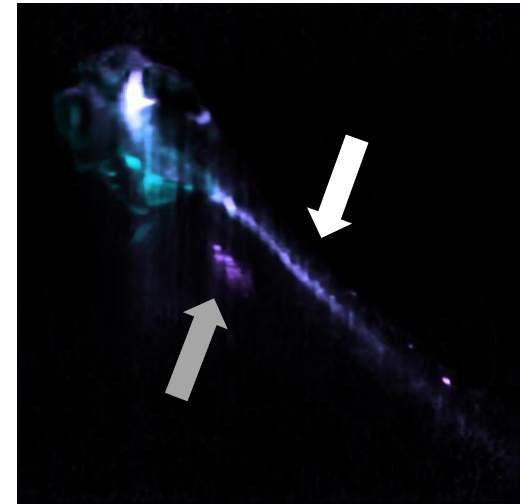


Separation of Overlapping Fluorophores

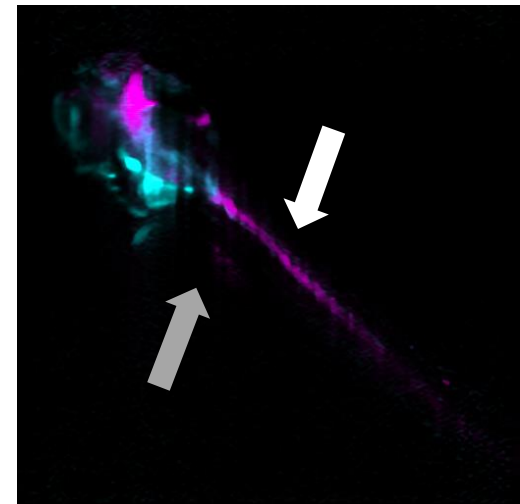
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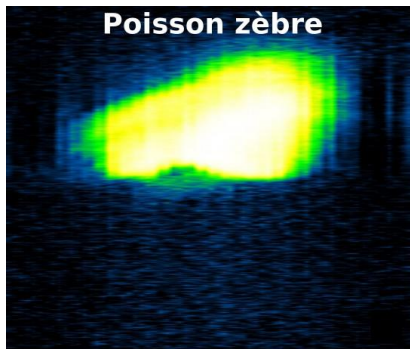


Filter

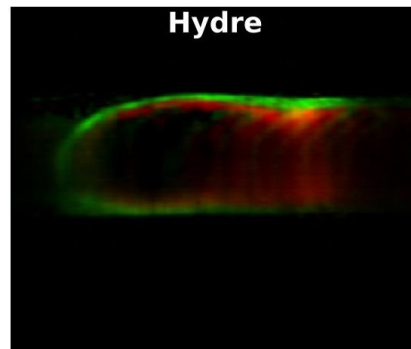


Spectral

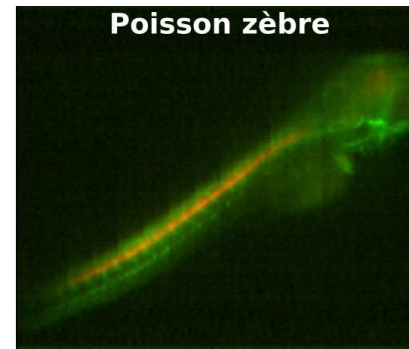




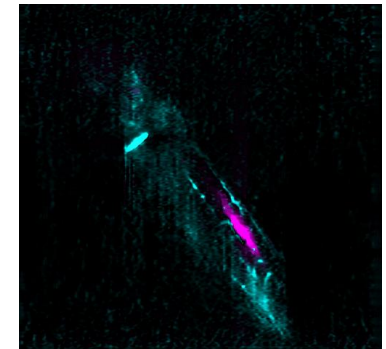
Octobre 2019



Juin 2020



Nov 2021



2023



Open questions :

- ❖ Improved reconstruction / improved unmixing
- ❖ Joint reconstruction-unmixing
- ❖ Choice of the patterns
- ❖ Two-arm reconstruction (i.e., pansharpening)
- ❖ Reconstruction hyperspectral 3D (x, y, z, λ)
- ❖ Dynamic reconstruction (x, y, z, λ, t)