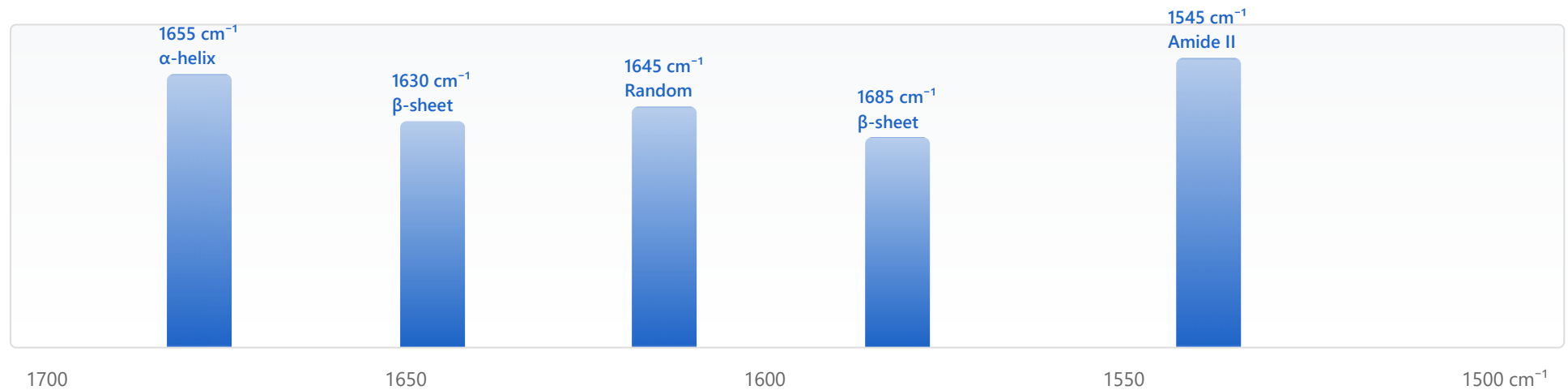


# FTIR for Biomolecules

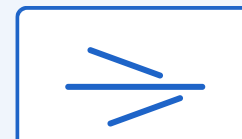
## Amide Bands

- Amide I ( $1600\text{--}1700\text{ cm}^{-1}$ ): C=O stretch, secondary structure sensitive
- Amide II ( $1510\text{--}1580\text{ cm}^{-1}$ ): N-H bend, C-N stretch
- $\alpha$ -helix:  $1650\text{--}1658$  |  $\beta$ -sheet:  $1620\text{--}1640$ ,  $1680\text{--}1690$  | Random:  $1640\text{--}1650$

Representative Protein FTIR Spectrum



## Secondary Structure Characteristics



### $\alpha$ -helix

1650-1658  $\text{cm}^{-1}$

### $\beta$ -sheet

1620-1640, 1680-1690  $\text{cm}^{-1}$

### Random coil

1640-1650  $\text{cm}^{-1}$

**$\alpha$ -helix:** Forms regular hydrogen bonding patterns due to helical structure, showing strong absorption at 1650-1658  $\text{cm}^{-1}$  from C=O stretching vibration.

**$\beta$ -sheet:** Shows two characteristic peaks depending on parallel or antiparallel structure. Absorbed at 1620-1640  $\text{cm}^{-1}$  (strong peak) and 1680-1690  $\text{cm}^{-1}$  (weak peak).

**Random coil:** Exhibits broad absorption band at 1640-1650  $\text{cm}^{-1}$  due to irregular structure.

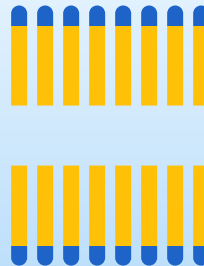
## Lipid Analysis

C-H stretch 2800-3000  $\text{cm}^{-1}$ . Membrane fluidity studies.

## Spectral Deconvolution

Fourier self-deconvolution resolves overlapping bands.

## Lipid Analysis

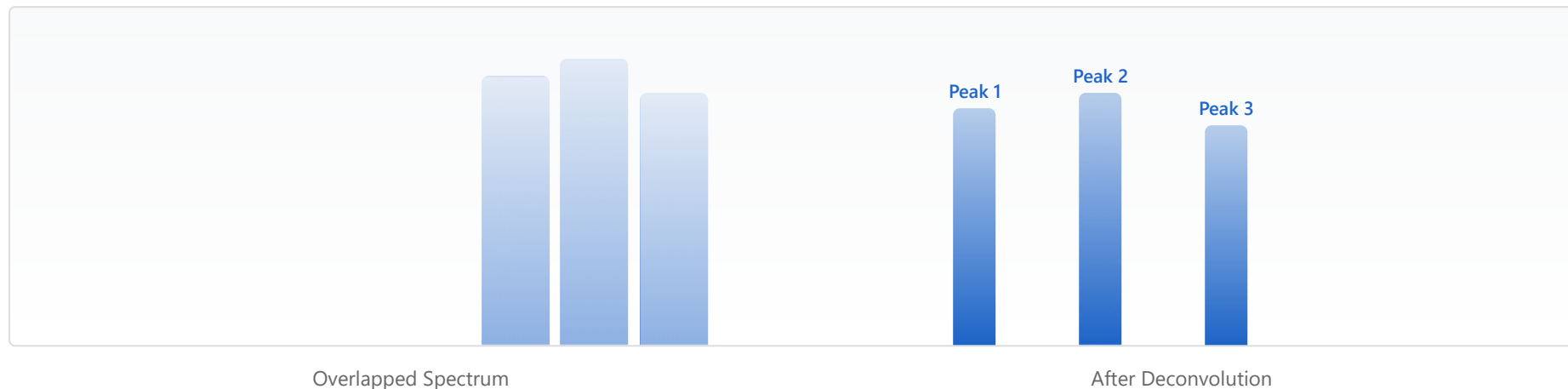


### C-H Stretching Vibration (2800-3000 $\text{cm}^{-1}$ ):

- 2920  $\text{cm}^{-1}$ : Asymmetric  $\text{CH}_2$  stretch
- 2850  $\text{cm}^{-1}$ : Symmetric  $\text{CH}_2$  stretch
- Changes in position and intensity of these peaks can be used to analyze fluidity and phase transitions of lipid bilayers.
- Peak shift to higher wavenumbers with increasing temperature indicates increased membrane fluidity.
- Applied in biological membrane studies, drug-membrane interactions, and lipid oxidation research.

## Spectral Deconvolution

### Separation of Overlapped Peaks



#### Fourier Self-Deconvolution:

- Separates overlapped peaks into individual components using mathematical algorithms.
- Enables quantification of various secondary structure components in the complex Amide I region spectrum.
- Used in conjunction with second derivative to enhance resolution.
- Essential technique for protein aggregation, structural changes, and ligand binding studies.
- Caution: Excessive deconvolution can generate artificial peaks, so appropriate parameter selection is important.

## Major Applications of Biomolecular FTIR



### Protein Structure Analysis

Monitoring protein folding, denaturation, and aggregation states. Studying protein-ligand interactions in drug development.



### Pharmaceutical Research

Drug crystalline forms, polymer formulations, stability assessment and quality control of protein therapeutics.



### Cell Membrane Research

Phase transitions of lipid bilayers, membrane fluidity, and drug-membrane interaction analysis.



### Biosensors

Label-free sensing for biomolecular recognition, immunoassays, and disease marker detection.

## Experimental Tips & Precautions

### Sample Preparation:

- Use D<sub>2</sub>O solution or measure in dry film form to minimize water interference
- Appropriate concentration: Protein 10-50 mg/mL, Lipid 5-20 mg/mL

### Spectrum Acquisition:

- Resolution: 2-4 cm<sup>-1</sup> (general), 1 cm<sup>-1</sup> (high-resolution analysis)
- Number of scans: Minimum 64 or more (improves signal-to-noise ratio)
- Acquire fresh background spectrum before each measurement

### Data Processing:

- Baseline correction
- Atmospheric water vapor/CO<sub>2</sub> correction
- Appropriate smoothing (Savitzky-Golay filter recommended)