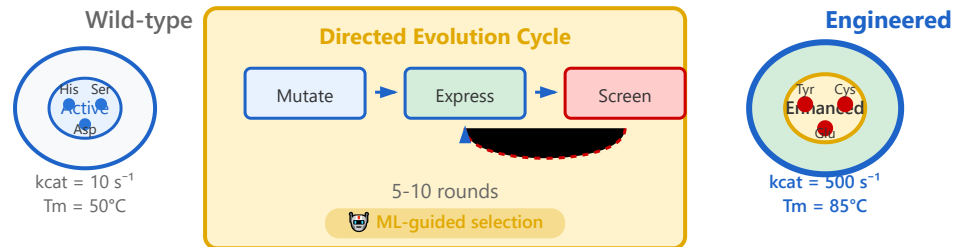


# Enzyme Engineering

## Directed Evolution & Rational Design



### Catalytic Reaction



### Industrial Applications



### Activity improvement

$k_{cat}/K_m$  optimization

### Substrate specificity

Promiscuity engineering

### Thermostability

High temperature operation

### Solvent tolerance

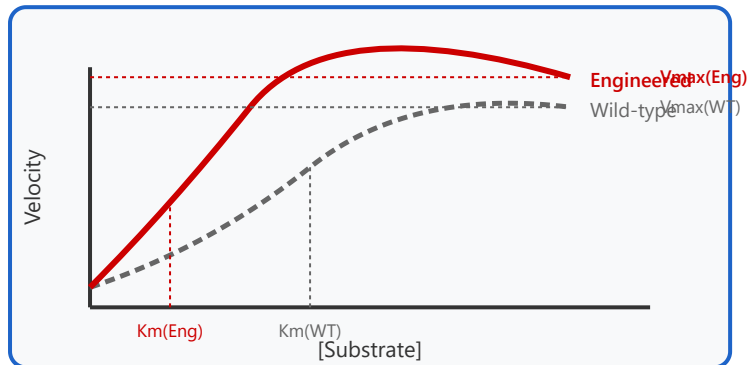
Organic solvent resistance

### Directed evolution

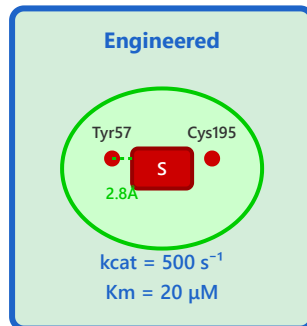
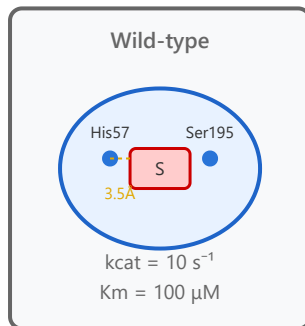
Iterative improvement cycles

# 1. Activity Improvement (kcat/Km Optimization)

## Catalytic Efficiency Enhancement



## Active Site Optimization



## Objective

Enhance the catalytic efficiency ( $k_{\text{cat}}/K_m$ ) of enzymes to increase reaction rates and substrate binding affinity. This is crucial for industrial processes requiring high throughput.

## Key Strategies

- **Transition state stabilization:** Modify active site residues to better stabilize the transition state
- **Substrate binding optimization:** Engineer binding pocket geometry for improved substrate fit
- **Product release enhancement:** Reduce product inhibition by facilitating product dissociation
- **Catalytic triad engineering:** Optimize spatial arrangement and pKa of catalytic residues

## Case Study: Subtilisin Protease

**Wild-type:**  $k_{\text{cat}}/K_m = 1.0 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$

**Engineered (N62D/G166D):**  $k_{\text{cat}}/K_m = 5.2 \times 10^6 \text{ M}^{-1}\text{s}^{-1}$

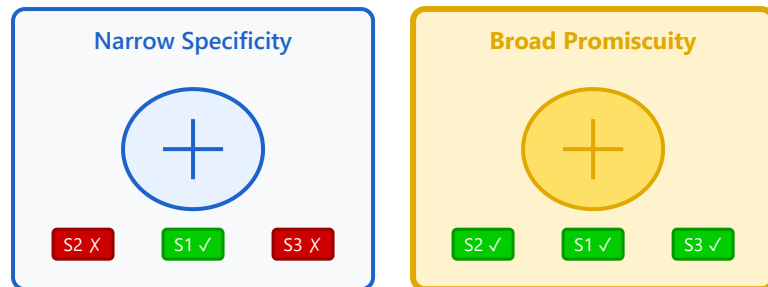
**Improvement:** 52-fold increase in catalytic efficiency through rational design of the oxyanion hole and substrate binding pocket.

**Impact:** Activity improvements of 10-1000 fold are achievable through directed evolution combined with computational design, enabling reduced enzyme loading and lower production costs.

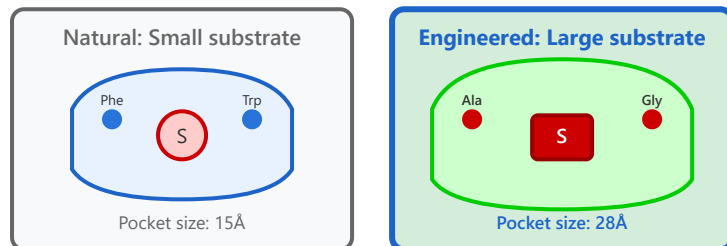
## 2. Substrate Specificity & Promiscuity Engineering

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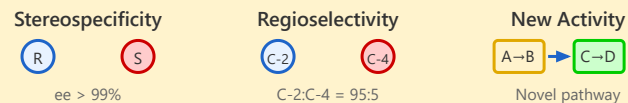
## Substrate Recognition Engineering



### Binding Pocket Reshaping



### Engineering Applications



## Objective

Modify substrate binding specificity to either narrow selectivity for a single substrate or broaden promiscuity to accept multiple substrates. This enables enzymes to process non-natural substrates or improve stereoselectivity.

## Engineering Approaches

- **Binding pocket reshaping:** Alter size and geometry through mutations (e.g., Phe→Ala for pocket enlargement)
- **Electrostatic tuning:** Change charge distribution to favor specific substrate classes
- **Hydrophobic interactions:** Engineer aromatic residues for  $\pi$ -stacking with substrates
- **Gatekeeper residue modification:** Control substrate entry and selectivity

### Case Study: P450 BM3 Hydroxylase

**Wild-type:** Hydroxylates C12-C16 fatty acids

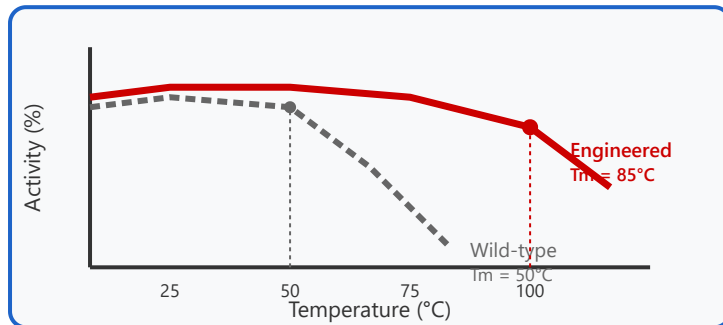
**Engineered (9 mutations):** Accepts propane and alkanes (C2-C8)

**Achievement:** Complete substrate scope inversion - from long-chain to short-chain hydrocarbons, enabling production of valuable chemical intermediates.

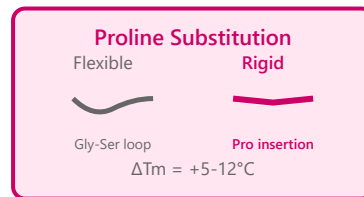
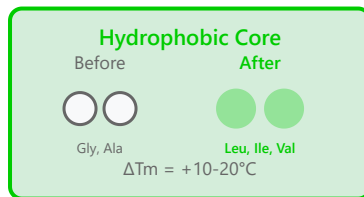
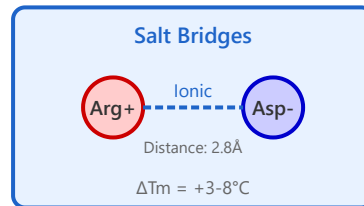
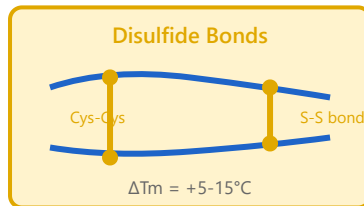
**Applications:** Pharmaceutical synthesis (>99% ee), biocatalytic cascades, plastic degradation, and production of non-natural amino acids and chemicals.

# 3. Thermostability Engineering

## Thermal Stability Enhancement



## Molecular Stabilization Mechanisms



## Objective

Increase enzyme thermal stability ( $T_m$ ) to enable operation at elevated temperatures, which improves reaction rates, reduces contamination risks, and extends enzyme shelf life in industrial processes.

## Stabilization Strategies

- **Disulfide bonds:** Introduce Cys-Cys bridges to constrain structure (5-15°C increase)
- **Salt bridges:** Engineer ionic interactions between charged residues (3-8°C increase)
- **Hydrophobic core packing:** Replace small residues (Gly, Ala) with bulky hydrophobic ones (Leu, Ile, Val) for tighter packing (10-20°C increase)
- **Proline substitution:** Reduce loop flexibility by inserting proline (5-12°C increase)
- **N/C-terminal modifications:** Add stabilizing residues or tags
- **Glycosylation:** Attach sugar moieties for protection

## Case Study: Bacillus $\alpha$ -Amylase

**Wild-type  $T_m$ :** 55°C (half-life: 15 min at 90°C)

**Engineered (15 mutations):**  $T_m = 95^\circ\text{C}$  (half-life: 7 hours at 90°C)

**Key mutations:** Introduction of 3 disulfide bonds, 5 salt bridges, and 7 core packing improvements. Now used in high-temperature starch processing.

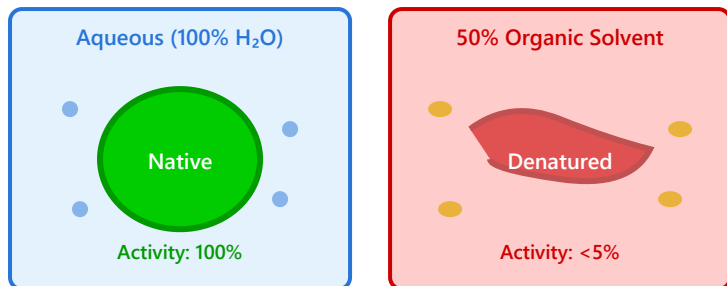
**Industrial Impact:** Thermostable enzymes enable higher process temperatures (70-100°C), reducing viscosity, increasing mass

transfer, and preventing microbial contamination without sterilization.

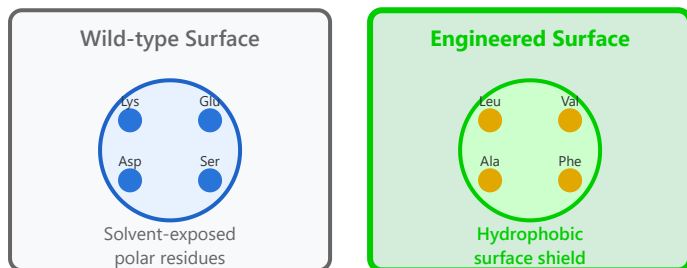
## 4. Solvent Tolerance Engineering

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## Organic Solvent Resistance



## Surface Engineering for Solvent Tolerance



## Solvent Compatibility Enhancement

Methanol	DMSO	Acetonitrile	Toluene
WT: 2% Eng: 65%	WT: <1% Eng: 40%	WT: 5% Eng: 70%	WT: 0% Eng: 35%

## Objective

Engineer enzymes to maintain activity and stability in organic solvents, enabling reactions with hydrophobic substrates and products that are poorly soluble in water.

## Engineering Strategies

- **Surface hydrophobicity:** Replace charged surface residues (Lys, Glu, Asp) with hydrophobic ones (Leu, Val, Ala, Phe)
- **Core stabilization:** Strengthen hydrophobic core to resist solvent penetration
- **Removal of water-binding sites:** Eliminate surface pockets that trap destabilizing water molecules
- **Increased rigidity:** Reduce conformational flexibility through proline substitutions and disulfide bonds
- **Active site protection:** Shield catalytic residues from solvent deactivation

## Case Study: *Candida antarctica* Lipase B (CALB)

**Wild-type:** 30% activity in 30% methanol

**Engineered (K26L/D223A/E226V):** 85% activity in 70% methanol

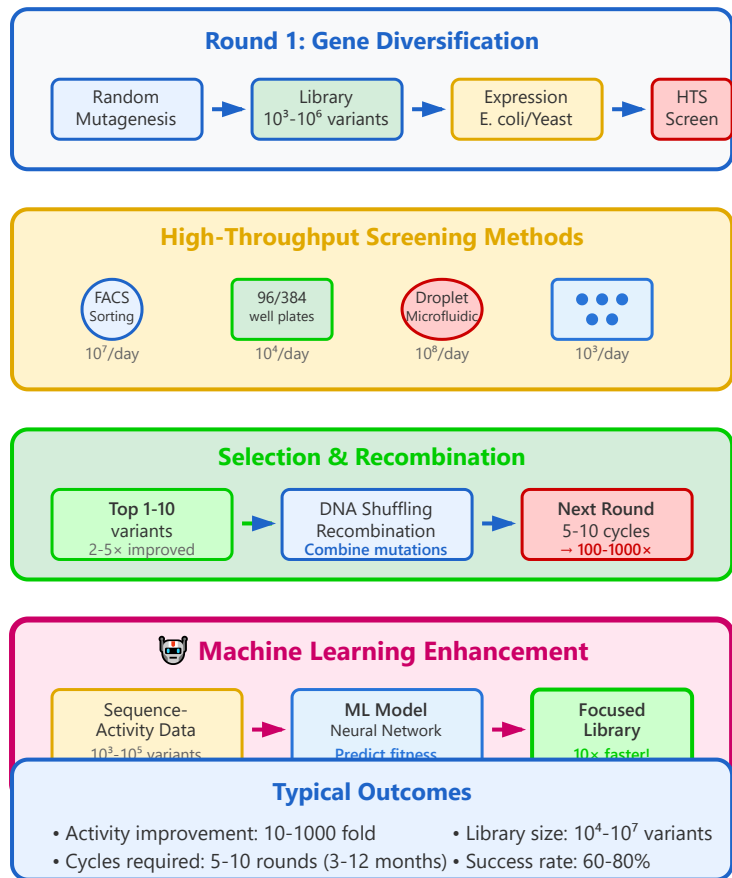
**Application:** Used in biodiesel production and synthesis of pharmaceutical esters in high solvent concentrations, dramatically improving product yields.

**Industrial Benefits:** Enables biphasic reactions, increases substrate/product solubility, reduces water activity for reversing hydrolysis, and facilitates downstream product recovery.



## 5. Directed Evolution Strategy

### Iterative Improvement Cycles



### Objective

Use iterative rounds of random mutagenesis, recombination, and selection to evolve enzymes with desired properties without requiring detailed structural knowledge. This approach mimics natural evolution but accelerated 1000-fold.

### Key Components

- **Mutagenesis methods:** Error-prone PCR (0.1-1% mutation rate), DNA shuffling, saturation mutagenesis
- **Library construction:** Generate  $10^3$ - $10^7$  variants with diverse mutations
- **High-throughput screening:** FACS ( $10^7$ /day), microfluidics ( $10^8$ /day), or plate-based assays ( $10^4$ /day)
- **Selection criteria:** Activity, stability, specificity, or multiple properties simultaneously
- **Recombination:** DNA shuffling to combine beneficial mutations from different variants
- **ML-guidance:** Machine learning models predict promising variants, reducing screening by 10-fold

### Nobel Prize Example: Frances Arnold's P450 Evolution

**Goal:** Evolve P450 for propane hydroxylation (non-natural activity)

**Starting:** 0% activity on propane

**After 5 rounds:** 300,000 turnovers with 98% selectivity

**Impact:** Enabled sustainable biocatalytic production of valuable chemicals from cheap alkanes, awarded 2018 Nobel Prize in Chemistry.

**Advantages:** No structural knowledge required, can optimize multiple properties simultaneously, discovers unexpected beneficial mutations, and integrates easily with computational design and ML prediction.