## **Supplementary material of** "OptimJV3:

an optimization tool for improving genomic data compression using genetic algorithms"

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## 1 Datasets and materials

Table 1 presents all datasets (sequences) used in the experiments, excluding samples.

## 2 Random and Local Search Results - Supplementary Material

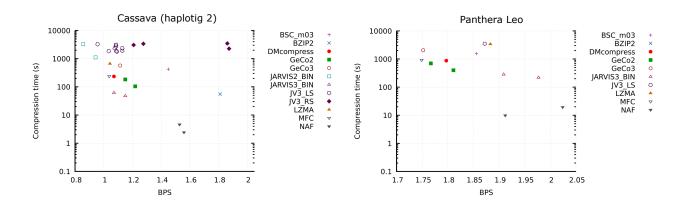


Figure S1: Results of Random Search (RS) for Cassava (haplotig2), and Local Search (LS) for Pathera Leo.

## 3 Reproducibility

## 3.1 Quick Demo

To clone the OptimJV3 project and change script permissions, execute the following instructions:

```
git clone https://github.com/cobilab/OptimJV3.git
chmod +x OptimJV3/scripts/*.sh
```

After cloning and activating the script permissions, the following demonstration, where a GA is applied to the compression the human chromosome Y (CY), can be executed in the scripts folder:

```
1 ./GetDSinfo.sh # map sequences into its DS, sorted by size; view sequences info
2 ./GA.sh -s cy -ga "demo" -lg 100 -t 10 # optimize compression of CY using canonical GA for 100
generations and 10 threads
```

The results will then be stored in a folder with prefix "DS" within the "OptimJV3" project. To identify this folder name, the following command can be executed:

```
1 ./Main.sh -v
```

For instance, assuming that there are no other sequences installed, the output folder should be "DS1".

Table S1: Enumeration of all datasets considered; RS/LS breadth experiments span this full list, whereas MGA/MOGA deep analyses are reported for CY, HG, and Cassava H1/H2. Scripts and outputs for the remaining datasets are available in the repository. ID refers to the NCBI sequence ID or an abbreviation if from

DS	Source	ID	Description	Size
DS1	NCBI	KT868810.1	Cutavirus strain BR-283 NS1 gene, partial cds; and putative VP1, hypothetical protein, VP2, and hypothetical protein genes	4.09668 KB
DS2	NCBI	CM029732.1	Pollicipes pollicipes isolate AB1234 mitochondrion, complete sequence $$	14.7363 KB
DS3	DNACorpus	BuEb	Bundibugyo ebolavirus genome	18.4961 KB
DS4	NCBI	OM812693.1	SARS-CoV-2 isolate $\rm HUN/Hun\text{-}1/2020,$ complete genome	29.0693 KB
DS5	AlcoR	alcor30KB	Synthetic sequence	29.2969 KB
DS6	DNACorpus	AgPh	Aggregatibacter phage S1249 genome	42.9395 KB
DS7	DNACorpus	YeMi	Yellowstone lake mimivirus genome	71.9619 KB
DS8	NCBI	NC_001664.4	Human betaherpesvirus 6A, variant A, isolate U1102, complete genome	155.643 KB
DS9	NCBI	NC_000898.1	Human herpesvirus 6B, complete genome	158.314 KB
DS10	NCBI	NC_000908.2	Mycoplasmoides genitalium G37, complete sequence	566.48 KB
DS11	DNACorpus	AeCa	Aeropyrum camini genome	1.51734 MB
DS12	DNACorpus	HePy	Helicobacter pylori genome	1.59056 MB
DS13	AlcoR	alcor2MB	Synthetic sequence with five low-complexity regions	2 MB
DS14	NCBI	NC_004461.1	Staphylococcus epidermidis ATCC 12228, complete sequence	2.3835 MB
DS15	DNACorpus	НаНі	Haloarcula hispanica genome	3.7098 MB
DS16	DNACorpus	EsCo	Escherichia coli genome	4.42662 MB
DS17	DNACorpus	PlFa	Plasmodium falciparum genome	8.5704 MB
DS18	DNACorpus	WaMe	Wallemia mellicola genome	8.72081 MB
DS19	DNACorpus	ScPo	Schizosaccharomyces pombe genome	10.1587 MB
DS20	NCBI	NC_000024.1	Human chromosome Y	21.6181 MB
DS21	DNACorpus	EnIn	Entamoeba invadens genome	25.1799 MB
DS22	DNACorpus	DrMe	Drosophila miranda chromosome 2	30.6906 MB
DS23	NCBI	BA000046.3	Pan troglodytes DNA, chromosome 22, complete sequence	31.1909 MB
DS24	NCBI	NC_000021.9	Human chromosome 21	38.2315 MB
DS25	DNACorpus	OrSa	Oriza sativa Japonica chromosome 1	41.2584 MB
DS26	NCBI	NC_073246.2	Gorilla gorilla isolate KB3781 chromosome 22	55.7969 MB
DS27	DNACorpus	DaRe	Danio rerio chromosome 3	59.6667 MB
DS28	NCBI	NC_072005.2	Pongo abelii isolate AG06213 chromosome 20	62.3704 MB
DS29	DNACorpus	AnCa	Anolis carolinensis genome	135.603 MB
DS30	NCBI	NC_000008.11	Human chromosome 8	138.062 MB
DS31	NCBI	NC_058373.1	Felis Catus chromosome B3	141.611 MB
DS32	DNACorpus	GaGa	Gallus gallus chromosome 2	141.651 MB
DS33	DNACorpus	HoSa	Human chromosome 4	180.962 MB
DS34	CNGB <sup>1</sup>	Cassava2	Cassava (haplotig 2)	673.607 MB
DS35	$CNGB^2$	Cassava	Cassava (haplotig 1)	727.074 MB
DS36	NCBI	GCA.008795835.1	Panthera Leo genome	2.22454 GB
DS37	3	HG	Human genome	2.9032 GB

<sup>1</sup> https://s3.ap-northeast-1.wasabisys.com/gigadb-datasets/live/pub/10.5524/102001\_103000/102193/00\_Assembly\_Fasta/haplotigs/
TME204.HiFi\_HiC.haplotig2.fa
2 https://s3.ap-northeast-1.wasabisys.com/gigadb-datasets/live/pub/10.5524/102001\_103000/102193/00\_Assembly\_Fasta/haplotigs/

https://s3.ap-northeast-1.wasabisys.com/gigadb-datasets/live/pub/10.5524/102001\_103000/102193/00\_Assembly\_Fasta/haplotigs/TME204\_HiFi\_HiC\_haplotig1\_fa

TME204.HiFi\_HiC.haplotig1.fa 

https://s3-us-west-2.amazonaws.com/human-pangenomics/T2T/CHM13/assemblies/analysis\_set/chm13v2.0.fa.gz

### 3.2 Setup

After cloning and changing script permissions, the following instruction should be executed to install all tools and sequences required to reproduce the experiments:

./Setup.sh

Alternatively, in the scripts folder, the following instructions should be executed:

```
./InstallTools.sh  # install listed compressors, GTO, and AlcoR

./DownloadFASTA.sh  # downloads FASTA files

./GetCassava.sh  # gunzip cassava files

./GetAlcoRFASTA.sh  # simulates and stores 2 synthetic FASTA sequences

./FASTA2seq.sh  # cleans FASTA files and stores raw sequence files

./DownloadDNAcorpus.sh  # download raw sequences from a balanced sequence corpus

./GetSample.sh -s cassava -sp 0.4 -mb 100 -so cassava100MB

./GetSample.sh -s cassava -sp 0.4 -mb 50 -so cassava50MB

./GetSample.sh -s cassava -sp 0.4 -mb 25 -so cassava25MB

./GetSample.sh -s cassava -sp 0.4 -mb 12.5 -so cassava12d5MB

./GetSample.sh -s human -sp 0.2 -mb 10 -so human10MB

./GetDSinfo.sh  # map sequences into their ids, sorted by size; view sequences info
```

## 3.3 Reproducing the "Non-MGA results"

To reproduce this experiment, the following commands should be executed in the scripts folder:

```
bash -x ./GA.sh -s $sequence -ga "rs" -lr 0 -lg 1 1> out 2> err &
```

### 3.4 Reproducing the "Canonical MGA experiment"

To reproduce this experiment, the following commands should be executed in the scripts folder:

```
bash -x ./GA.sh -s "escherichia_coli" -ga "e0_ga1_lr0_cmga" -lr 0 -lg 100 1> out 2> err &
bash -x ./GA.sh -s cy -ga "e0_ga1_lr0_cmga" -lr 0 -lg 100 1> out 2> err &
bash -x ./GA.sh -s cassava -ga "e0_ga1_lr0_cmga" -lr 0 -lg 20 1> out 2> err &
```

## 3.5 Reproducing the "CY experiments"

To reproduce these experiments, the following command should be executed in the scripts folder:

```
1 bash -x ./Main.sh -s cy -lg 100 -t 10 1> out 2> err &
```

## 3.6 Reproducing the "Cassava sampling experiment"

To reproduce this experiment, run:

```
./SamplingDemoCassava.sh
```

## 3.7 Reproducing the "Human genome sampling experiments"

To reproduce this experiment for human genome samples greater than 10 MB, run:

```
./SamplingDemo.sh
```

As for the 10 MB experiment, the following command should be executed to run a GA algorithm for 500 generations:

```
bash -x ./GA.sh -s human10MB -lg 500 1> out 2> err &
```

## 4 Parameters and options of OptimJV3

All implemented features are listed in the following scripts:

```
1 ./InstallTools.sh -h
2 ./GetDSinfo.sh -h
3 ./Main.sh -h
4 ./GA.sh -h
5 ./Initialization.sh -h
6 ./Run.sh -h
7 ./Evaluation.sh -h
```

```
8 ./Selection.sh -h
9 ./CrossMut.sh -h
10 ./GetSamples.sh -h
11 ./PlotGA.sh -h
12 ./PlotGAcmp.sh -h
```

#### 4.1 Install tools menu

To access the menu of the script that installs all required tools, the following command can be executed

```
./InstallTools.sh -h
```

This command will output the following content

with the options/parameters available.

## 4.2 Download FASTA sequences menu

To access the menu of the script that downloads a set of FASTA sequences, or a specific FASTA sequence given its NCBI ID, the following command can be executed

```
./DownloadFASTA.sh -h
```

This command will output the following content

with the options/parameters available.

## 4.3 Map sequences to dataset ID menu

The GetDSinfo.sh maps and displays information about sequences, their dataset id, and their size. The command to access its features is:

```
./GetDSinfo.sh -h
```

This command will output the following content

with the options/parameters available.

#### 4.4 Main menu

The main script runs a set of pre-configured GAs and applied to a set of sequences.

The command to access the main menu with its features is

```
./Main.sh -h
```

This command will output the following content

```
OptimJV3 - optimize JARVIS3 CM and RM parameters
  Program options -----
  -h|--help......Show this
  -v|--view-ds|--view-datasets...View sequence names, size
            of each in bytes, MB, and BG, and their group
  -s|--seq|--sequence......Select sequence by its name
  -sg|-sequence-grp|--seq-group..Select group of sequences
                                          by their size
12
  -ds|--dataset.....Select sequence by its dataset number
  -dr|--drange|--dsrange|--dataset-range......Select
                   sequences by range of dataset numbers
  -fg|--first-gen|--first-generation......Define first
                                     generation number
  -lg|--last-gen|--last-generation..........Define last
                                      generation number
19
  -rg|--range-gen|--range-generation.....Define generation
20
                                                 range
  -t|--nthreads....Define number of threads to run JARVIS3
23
  -sd|--seed......Define pseudo-random seed
24
  -si|--seed-increment...........Define seed increment
25
  example 1: ./Main.sh -s human
  example 2: ./Main.sh -s cassava -s human
```

with the options/parameters available.

### 4.5 GA menu

The GA script applies a GA to a set of sequences.

The command to access the GA menu with the options/parameters of OptimJV3 is

```
1 ./GA.sh -h
```

15	<u> </u>	
16	-sg seq-grp sequence-groupSelect sequence group	
17	1 7	
18	-dr drange dsrange dataset-rangeSelect	
19	sequences by range of dataset numbers	
20	-ps psize population population-sizeDefine	
21	population size	
22	-sd seedDefine pseudo-random seed	
23	-si seed-incrementDefine seed increment	
24		
25	Program options (initialization)	
26		
27	-hei heuristic-initializationActivate heuristic	
28	initialization/local initialization	
29	-hyi hybrid-initializationActivate hybrid	
30	initialization	
31	-hhp hybrid-heuristic-percentageDefine percentage	
32	of tests generated by heuristic/local search	
33	-hhp hybrid-heuristic-percentageDefine number	
34	of tests generated by heuristic/local search	
35	-mCM m-cm min-cmDefine minimum number of context	
36	models (CMs)	
37	-MCM M-cm max-cmDefine maximum number of	
38	context models (CMs)	
39	-mRM m-rm min-rmDefine minimum number of	
40	copy/repeat models (RMs)	
41	-MRM M-rm max-rmDefine maximum number of	
42	copy/repeat models (RMs)	
43	-lr learning-rateDefine learning rate	
44	-hs hidden-sizeDefine hidden size	
45	-sing seedingActivate seeding feature	
45 46	-sing seedingActivate seeding feature to populate with few hardcoded solutions. Only works for	
	-sing seedingActivate seeding feature to populate with few hardcoded solutions. Only works for human genome	
46	to populate with few hardcoded solutions. Only works for	
46 47	to populate with few hardcoded solutions. Only works for	
46 47 48 49	to populate with few hardcoded solutions. Only works for human genome	
46 47 48 49 50	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48	to populate with few hardcoded solutions. Only works for human genome  Program options (run)  -t nthreadsDefine number of threads to run JARVIS3	
46 47 48 49 50 51 52	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53	to populate with few hardcoded solutions. Only works for human genome  Program options (run)  -t nthreadsDefine number of threads to run JARVIS3	
46 47 48 49 50 51 52 53	to populate with few hardcoded solutions. Only works for human genome  Program options (run)  -t nthreadsDefine number of threads to run JARVIS3 in parallel	
46 47 48 49 50 51 52 53 54	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 59 60	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 59 60	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 60 61 62	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 60 61 62 63	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 59 60 61 62 63 64	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 60 61 62 63 64 65	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 60 61 62 63 64 65 66	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 67 60 61 62 63 64 65 66	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 60 61 62 63 64 65 66 67 68	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 60 61 62 63 64 65 66 67 68 69	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 59 60 61 62 63 64 65 66 67 68 69 70	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 60 61 62 63 64 65 66 67 68 69 70 71	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 60 61 62 63 64 65 66 67 68 69 70 71 72	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 60 61 62 63 64 65 66 67 68 69 70 71	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	
46 47 48 49 50 51 52 53 54 55 56 57 58 60 61 62 63 64 65 66 67 68 69 70 71 72	to populate with few hardcoded solutions. Only works for human genome  Program options (run)	

```
Program options (crossover) -----
  -cr|-ccr|--command-crossover-rate..Define crossover rate
                           of a selected pair of commands
  -mrc|--model-crossover-rate......Define crossover rate
                         of a selected pair of CMs or RMs
81
  -cc|--command-crossover......Choose command crossover
   operator: 'mrc' (metameric random crossover) (default),
                    (metameric canonical crossover) 'mcc'
  -c|--crossover........Choose model crossover operator:
85
     'xpoint' (default), 'uniform', 'average', 'discrete',
                                     'flat', 'heuristic'
88
   Program options (mutation) -----
89
90
  -hm|--heuristic-mutation.....Activate heuristic mutation
                             for narrower range mutations
92
  -mr|-cmr|--command-mutation-rate....Define mutation rate
93
                                            of a command
  -pmr|--parameter-mutation-rate.....Define mutation rate
                                          of a parameter
96
97
  Program options (stop criteria) ------
98
  -sc|--stop-criteria......Define stop criteria: '1' to
              halt program when no offspring is produced,
                   else (default) stop at last generation
```

## 4.6 Initialization menu

The initialization script starts a GA by initializing a population of commands that compress a chosen sequence. To access the initialization menu, the following menu can be executed

```
1 ./Initialization.sh -h
```

```
OptimJV3 - optimize JARVIS3 CM and RM parameters
Program options -----
 -h|--help......Show this
-v|--view-ds|--view-datasets..View sequence names, size
         of each in bytes, MB, and BG, and their group
-s|--seq|--sequence......Select sequence by its name
-sg|--sequence-grp|--seq-group.Select group of sequences
                                      by their size
-a|-ga|--genetic-algorithm...Define (folder) name of the
                                  genetic algorithm
-s|--seq|--sequence......Select sequence name
-sg|--seq-grp|--sequence-group.....Select sequence group
-ds|--dataset.....Select sequence by its dataset number
-dr|--drange|--dsrange|--dataset-range......Select
                 sequences by range of dataset numbers
-ps|--psize|--population|--population-size......Define
                                    population size
-sd|--seed......Define pseudo-random seed
-si|--seed-increment...........Define seed increment
```

```
Program options (initialization) -----
26
  -hei|--heuristic-initialization.....Activate heuristic
                      initialization/local initialization
  -hyi|--hybrid-initialization.....Activate hybrid
29
                                         initialization
30
  -hhp|--hybrid-heuristic-percentage.....Define percentage
31
             of tests generated by heuristic/local search
  -hhp|--hybrid-heuristic-percentage......Define number
33
             of tests generated by heuristic/local search
  -mCM|--m-cm|--min-cm....Define minimum number of context
                                           models (CMs)
  -MCM|--M-cm|--max-cm......Define maximum number of
37
                                    context models (CMs)
38
  -mRM|--m-rm|--min-rm.......Define minimum number of
                                copy/repeat models (RMs)
  -MRM|--M-rm|--max-rm.......Define maximum number of
41
                                copy/repeat models (RMs)
  -lr|--learning-rate......Define learning rate
  -hs|--hidden-size......Define hidden size
  -sing|--seeding......Activate seeding feature
  to populate with few hardcoded solutions. Only works for
                                           human genome
```

#### 4.7 Run menu

The run script is used to run the initial population or offspring from a certain generation.

To access the menu of the run script, the following command can be executed

./Run.sh -h

This command will output the following content

```
OptimJV3 - optimize JARVIS3 CM and RM parameters
  Program options -----
  -h|--help......Show this
  -v|--view-ds|--view-datasets...View sequence names, size
           of each in bytes, MB, and BG, and their group
  -s|--seq|--sequence......Select sequence by its name
  -sg|--sequence-grp|--seq-group.Select group of sequences
                                        by their size
12
  -a|-ga|--genetic-algorithm...Define (folder) name of the
13
                                    genetic algorithm
  -ds|--dataset.....Select sequence by its dataset number
  -dr|--drange|--dsrange|--dataset-range......Select
16
                  sequences by range of dataset numbers
  -ps|--psize|--population|--population-size......Define
                                      population size
  -sd|--seed......Define pseudo-random seed
20
  -si|--seed-increment..........Define seed increment
21
22
  Program options (run) ------
  -t|--nthreads....Define number of threads to run JARVIS3
                                          in parallel
  -to|--timeout......Define timeout (default: 1 hour)
```

with the options/parameters available.

#### 4.8 Evaluation menu

This script is used to evaluate and sort the current generation based on their compression, expressed in BPS, and compression time.

To access the menu of evaluation script, the following command can be executed

1 ./Evaluation.sh -h

This command will output the following content

```
_____
   OptimJV3 - optimize JARVIS3 CM and RM parameters
   Program options -----
  -h|--help......Show this
  -v|--view-ds|--view-datasets...View sequence names, size
            of each in bytes, MB, and BG, and their group
  -s|--seq|--sequence......Select sequence by its name
  -sg|--sequence-grp|--seq-group.Select group of sequences
                                        by their size
  -a|-ga|--genetic-algorithm...Define (folder) name of the
                                     genetic algorithm
  -s|--seq|--sequence......Select sequence name
  -sg|--seq-grp|--sequence-group.....Select sequence group
  -ds|--dataset.....Select sequence by its dataset number
  -dr|--drange|--dsrange|--dataset-range......Select
18
                   sequences by range of dataset numbers
  -ps|--psize|--population|--population-size......Define
                                       population size
  -sd|--seed.................Define pseudo-random seed
  -si|--seed-increment...........Define seed increment
23
24
   Program options (evaluation) -----
26
   -moga|--moga-wm|--moga-weighted-metric...Activate Multi
27
        Objective Genetic Algorithm (MOGA) using weighted
29
                                       metric function
   -moga-ws|--moga-weighted-sum...Activate Multi-Objective
30
     Genetic Algorithm (MOGA) using weighted sum function
31
  -pe|--p-exp|--p-expoent........Define expoent for MOGA
32
                              weighted metric function
  -w1|-wBPS|--w-bps|--weight-bps.....Bits Per Symbol (BPS)
34
                               weight for MOGA function
  -w2|-wCTIME|--w-ctime|--weight-ctime....Compression time
                               weight for MOGA function
```

with the options/parameters available.

#### 4.9 Selection menu

This script selects commands from the population as potential candidates for producing offspring.

To access the selection menu, the following command can be executed

```
./Selection.sh -h
```

```
OptimJV3 - optimize JARVIS3 CM and RM parameters

Program options ------
```

```
-h|--help......Show this
  -v|--view-ds|--view-datasets...View sequence names, size
           of each in bytes, MB, and BG, and their group
  -s|--seq|--sequence......Select sequence by its name
  -sg|--sequence-grp|--seq-group.Select group of sequences
                                        by their size
  -a|-ga|--genetic-algorithm...Define (folder) name of the
                                    genetic algorithm
  -s|--seq|--sequence......Select sequence name
15
  -sg|--seq-grp|--sequence-group.....Select sequence group
  -ds|--dataset......Select sequence by its dataset number
  -dr|--drange|--dsrange|--dataset-range......Select
                   sequences by range of dataset numbers
19
  -ps|--psize|--population|--population-size......Define
20
                                      population size
21
  -sd|--seed................Define pseudo-random seed
  -si|--seed-increment..........Define seed increment
23
24
  Program options (selection) -----
  27
    operator: 'elitist' (default), 'roulette', 'rank', or
28
                                         'tournament'
  -ns|--num-sel-cmds...Define number of commands to select
  -sr|--selection-rate...Define rate of commands to select
```

### 4.10 Crossover and Mutation menu

The crossover and mutation script is used to produce offspring.

To access the menu of this script, the following command can be executed

./CrossMut.sh -h

```
OptimJV3 - optimize JARVIS3 CM and RM parameters
  Program options -----
  -h|--help......Show this
  -v|--view-ds|--view-datasets...View sequence names, size
           of each in bytes, MB, and BG, and their group
  -s|--seq|--sequence......Select sequence by its name
  -sg|--sequence-grp|--seq-group.Select group of sequences
                                       by their size
  -a|-ga|--genetic-algorithm...Define (folder) name of the
                                    genetic algorithm
  -s|--seq|--sequence......Select sequence name
  -sg|--seq-grp|--sequence-group.....Select sequence group
16
  -ds|--dataset......Select sequence by its dataset number
  -dr|--drange|--dsrange|--dataset-range......Select
                   sequences by range of dataset numbers
  -ps|--psize|--population|--population-size......Define
20
                                      population size
  -sd|--seed......Define pseudo-random seed
  -si|--seed-increment.......Define seed increment
23
24
  Program options (crossover) -----
```

```
-cr|-ccr|--command-crossover-rate..Define crossover rate
                            of a selected pair of commands
28
  -mrc|--model-crossover-rate......Define crossover rate
                          of a selected pair of CMs or RMs
  -cc|--command-crossover.......Choose command crossover
   operator: 'mrc' (metameric random crossover) (default),
                     (metameric canonical crossover) 'mcc'
  -c|--crossover.........Choose model crossover operator:
     'xpoint' (default), 'uniform', 'average', 'discrete',
35
                                       'flat', 'heuristic'
   Program options (mutation) -----
38
39
  -hm|--heuristic-mutation.....Activate heuristic mutation
40
                              for narrower range mutations
  -mr|-cmr|--command-mutation-rate....Define mutation rate
43
  -pmr|--parameter-mutation-rate.....Define mutation rate
                                            of a parameter
```

### 4.11 Get Sample menu

The features of the script that outputs a sample file can be found by running the following command

```
./GetSample.sh -h
```

This command will output the following content

with the options/parameters available.

#### 4.12 Get Samples menu

The features of the script that outputs four sample files from a sequence - with sizes of 100MB, 50MB, 25MB and 12.5MB - can be found by running the following command

```
./GetSamples.sh -h
```

```
9 -----
```

#### 4.13 Plot GA menu

The features of the script that plots the evolutionary plots and histograms of a single GA can be found by running the following command

```
./PlotGA.sh -h
```

This command will output the following content

```
OptimJV3 - optimize JARVIS3 CM and RM parameters
Program options -----
-h|--help......Show this
-a|-ga|--genetic-algorithm...Define (folder) name of the
                                   genetic algorithm
-s|--seq|--sequence............Choose sequence name/file
-ds|--dataset.....Select sequence by its dataset number
-pb|--percentage-best.........Define percentage of best
                                  individuals to plot
-b|--best.....Define number of best individuals to plot
-fg|--first-generation...Specify first generation number
-lg|--last-generation.....Select last generation number
-br|--b-range......Define x-axis (BPS range)
-trs|--trange-s...Define y-axis (time range, in seconds)
-trm | -- trange-m... Define y-axis (time range, in minutes)
-trh|--trange-h....Define y-axis (time range, in hours)
-hi|--hist-interval......Define bin size for histogram
```

with the options/parameters available.

## 4.14 Plot GA comparison menu

The features of the script that plots the comparison between GAs of a specific experiment can be found by running the following command

```
1 ./PlotGAcmp.sh -h
```

```
-br|--b-range.................Define x-axis (BPS range)
-trs|--trange-s...Define y-axis (time range, in seconds)
-trm|--trange-m...Define y-axis (time range, in minutes)
-trh|--trange-h.....Define y-axis (time range, in hours)
```

## 5 Parameters and options of CompressSequences

```
./InstallTools.sh -h
./DownloadFASTA.sh -h
./GetDSinfo.sh -h
./RunTestsExample.sh
./ProcessBenchRes.sh -h
./Plot.sh -h
```

#### 5.1 Install tools menu

See "Install tools menu" subsection from "Parameters and options of OptimJV3" section.

## 5.2 Download FASTA sequences menu

See "Download FASTA sequences menu" subsection from "Parameters and options of OptimJV3" section.

## 5.3 Map sequences to dataset ID menu

See "Map sequences to dataset ID menu" subsection from "Parameters and options of OptimJV3" section.

#### 5.4 Plot menu

The features of the script that plots the results of benchmark data can be found by running

```
./Plot.sh -h
```

This command will output the following content

```
CompressSequences - benchmark
     Plot Script
     -s|--sequence......Select sequence
    --sequence|--seq|-s.....Select sequence by its name
    --sequence-group|--seq-grp|-sg.Select group of sequences
12
                                               by their size
                1. Sequences with size lower than 1MB
                2. Sequences with size between 1MB and 100\,\mathrm{MB}
14
                3. Sequences with size between 100MB and 1GB
15
16
                4. Sequences with size between 1GB and 3GB
                5. Sequences with size greater than 3GB
17
    -br|--b-range..........Define x-axis (BPS range)
18
    -trs|--trange-s...Define y-axis (time range, in seconds)
19
    -trm | --trange -m... Define y-axis (time range, in minutes)
20
21
    -trh|--trange-h.....Define y-axis (time range, in hours)
    -m|--mode....Select data to plot ('bench', 'NGA', 'all')
22
23
```

with the options/parameters available.

#### 5.5 Run menu

The features of the script that runs the generated JARVIS3 commands can be found by running

```
./Run.sh -h
```

Alternatively, to access the menu of a similar script which runs fewer tests the following instruction can be executed:

```
1 ./RunLess.sh -h
```

Both instructions will output the following content

```
______
  CompressSequences - benchmark
  Run Script
  Program options -----
  -h|--help......Show this
  -v|--view-ds|--view-datasets...View sequence names, size
9
          of each in bytes, MB, and GB, and their group
_{11} -s|--seq|--sequence.......Select sequence by its name
  -sg|--sequence-grp|--seq-group.Select group of sequences
                                   by their size
13
_{14} -a|-ga|--genetic-algorithm...Define (folder) name of the
                                genetic algorithm
16 -ds | --dataset . . . . . Select sequence by its dataset number
17 -dr | --drange | --dsrange | --dataset -range . . . . . . . . . . . . Select
               sequences by range of dataset numbers
18
-t|--nthreads....Define number of threads to run JARVIS3
21
                                   in parallel
```

with the options/parameters available.

## 5.6 Process Benchmark results menu

The features of the script that processes the benchmark data can be found by running

```
1 ./ProcessBenchRes.sh -h
```

This command will output the following content

with the options/parameters available.

#### 5.7 JARVIS3 Parameters

JARVIS3 [1] Context Models (-cm) and Repeat Models (-rm) are the following:

```
-cm [NB_C]:[NB_D]:[NB_I]:[NB_G]/[NB_S]:[NB_E]:[NB_R]:[NB_A]
        Template of a context model.
3
        Parameters:
        [NB_C]: (integer [1;14]) order size of the regular context
6
                model. Higher values use more RAM but, usually, are
                related to a better compression score.
        [NB_D]: (integer [1;5000]) denominator to build alpha, which
                is a parameter estimator. Alpha is given by 1/[NB_D].
9
                Higher values are usually used with higher [NB_C],
                and related to confident bets. When [NB_D] is one,
                the probabilities assume a Laplacian distribution.
12
        [NB_I]: (integer \{0,1,2\}) number to define if a sub-program
13
                which addresses the specific properties of DNA
14
```

```
sequences (Inverted repeats) is used or not. The
                 number 1 turns ON the sub-program using at the same
16
                 time the regular context model. The number 2 does
17
18
                 only contemple the inversions only (NO regular). The
                 number 0 does not contemple its use (Inverted repeats
19
                 {\tt OFF})\,. The use of this {\tt sub\text{-}program} increases the
20
                 necessary time to compress but it does not affect the
21
                 RAM.
22
         [NB_G]: (real [0;1)) real number to define gamma. This value
23
                 represents the decayment forgetting factor of the
24
25
                 regular context model in definition.
         [NB_S]: (integer [0;20]) maximum number of editions allowed
26
                 to use a substitutional tolerant model with the same
27
                 memory model of the regular context model with
28
                 order size equal to [NB_C]. The value 0 stands for
29
                 turning the tolerant context model off. When the
30
                 model is on, it pauses when the number of editions
31
                 is higher that [NB_C], while it is turned on when
32
                 a complete match of size [NB_C] is seen again. This
33
```

impact of this model is usually only noticed for higher [NB\_C]. [NB\_R]: (integer  $\{0,1\}$ ) number to define if a sub-program which addresses the specific properties of DNA sequences (Inverted repeats) is used or not. It is

is probabilistic-algorithmic model very useful to

sequences. When  $[NB_S] > 0$ , the compressor used more

processing time, but uses the same RAM and, usually, achieves a substantial higher compression ratio. The

handle the high substitutional nature of genomic

- similar to the [NR\_I] but for tolerant models. [NB\_E]: (integer [1;5000]) denominator to build alpha for substitutional tolerant context model. It is analogous to [NB\_D], however to be only used in the probabilistic model for computing the statistics of the substitutional tolerant context model.
- [NB\_A]: (real [0;1)) real number to define gamma. This value represents the decayment forgetting factor of the substitutional tolerant context model in definition. Its definition and use is analogus to [NB\_G].

... (you may use several context models)

-rm [NB\_R]:[NB\_C]:[NB\_B]:[NB\_L]:[NB\_G]:[NB\_I]:[NB\_W]:[NB\_Y] Template of a repeat model. Parameters:

[NB\_R]: (integer [1;10000] maximum number of repeat models

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- for the class. On very repetive sequences the RAM increases along with this value, however it also improves the compression capability.
- [NB\_C]: (integer [1;14]) order size of the repeat context model. Higher values use more RAM but, usually, are related to a better compression score.
- $[NB_B]$ : (real (0;1]) beta is a real value, which is a parameter for discarding or maintaining a certain repeat model.
- [NB\_L]: (integer (1;20]) a limit threshold to play with [NB\_B]. It accepts or not a certain repeat model.
- [NB\_G]: (real [0;1)) real number to define gamma. This value represents the decayment forgetting factor of the regular context model in definition.
- [NB\_I]: (integer {0,1,2}) number to define if a sub-program which addresses the specific properties of DNA sequences (Inverted repeats) is used or not. The number 1 turns  ${\tt ON}$  the  ${\tt sub-program}$  using at the same time the regular context model. The number 0 does not contemple its use (Inverted repeats OFF). The number 2 uses exclusively Inverted repeats. The use of this sub-program increases the necessary time to compress but it does not affect the RAM.
- [NB\_W]: (real (0;1)) initial weight for the repeat class.
- [NB\_Y]: (integer  $\{0\}$ , [1;50]) maximum cache size. This will use a table cache with the specified size. The size must be in balance with the k-mer size [NB\_C].

# References

[1] M. J. P. Sousa et al. JARVIS3: an efficient encoder for genomic data. Bioinformatics, 7(7), 2025.