



ORIGINAL RESEARCH



Removal of sodium diclofenac from aqueous solutions by rice hull biochar

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Received: 27 June 2020 / Accepted: 24 November 2020
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Abstract

Diclofenac is an ecotoxic pharmaceutical compound affecting both aquatic and terrestrial ecosystems even at low concentrations. In this study, batch adsorption experiments were conducted to remove sodium diclofenac (SD) from aqueous solutions using rice hull biochar (RHB) adsorbents. Higher SD removals were obtained with increasing initial SD concentration, RHB dosage, contact time, and decreasing initial pH. Isotherm studies revealed that Langmuir isotherm best fitted the experimental data with the highest coefficient of determination for both pH 2 ($R^2 = 0.9827$) and pH 7 ($R^2 = 0.9460$). For kinetic studies, the pseudo second-order model gave $R^2 = 0.9999$ for both pH 2 and pH 7. SD removals reached up to 97% at pH 2 and up to 80% at pH 7. Higher SD removals were achieved at solution pH lower than the adsorbent pH_{pzc} since electrostatic repulsion was eliminated. Fourier-transform infrared analysis showed the major involvement of C=O in the adsorption process. This study demonstrated the potential of using agricultural residues such as rice hulls for the treatment of wastewater contaminated with pharmaceutical compounds.

Graphic abstract



Keywords Adsorption · Biochar · Isotherm · Kinetics · Rice hull · Sodium diclofenac

Nomenclature

C_0 Initial SD concentration, mg L^{-1}

C Final SD concentration, mg L^{-1}

Q Equilibrium adsorption capacity, mg g^{-1}

q_m Maximum adsorption capacity, mg g^{-1}

b Energy of adsorption, L mg^{-1}

C_e Equilibrium concentration of the adsorbate, mg L^{-1}

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R_L	Separation factor or equilibrium parameter
K_f	Freundlich capacity factor, ($\text{mg}^{1-1/n} \text{L}^{1/n} \text{g}^{-1}$)
$1/n$	Intensity parameter
β	Constant related to heat of sorption, J mol^{-1}
α	Temkin isotherm equilibrium binding constant, L g^{-1}
q_t	Amount of adsorbate at contact time t , mg g^{-1}
k_1	Pseudo-first-order rate constant, min^{-1}
k_2	Pseudo-second-order rate constant, $\text{g mg}^{-1} \text{min}^{-1}$
k_{id}	Intraparticle diffusion rate constant, $\text{mg g}^{-1} \text{min}^{-0.5}$

1 Introduction

Pharmaceuticals and personal care products (PPCPs) are emerging contaminants that have been a cause for concern in recent years (Hu et al. 2011). Extensive use of PPCPs, such as analgesics, antibiotics, contraceptives, lipid regulators, insect repellents, sunscreens, lotions, soaps, toothpaste, detergents, fragrances, and skin, hair, and dental care products, both in medical and veterinary practice has resulted in their presence in unaltered form in surface waters (Martínez et al. 2011; Liu and Wong 2013), since these persistent compounds are not removed completely by sewage treatment plants (STPs) and other conventional treatment methods (Gotostos et al. 2014). When these compounds are released into recipient waters, they can be bioaccumulated and, therefore, may present potential risks to human health and aquatic organisms (Sotelo et al. 2012).

Among the pharmaceutical substances that reach the aquatic environments via industrial effluents and human use, sodium diclofenac (SD) is the most widely detected (Wu et al. 2019). Chemically known as sodium 2-[(2,6-dichlorophenyl) amino] phenyl] acetate, SD is among the most extensively-used nonsteroidal anti-inflammatory drugs (NSAIDs) (Ensano et al. 2017). In fact, this drug is widely used to treat livestock especially across the Indian sub-continent (Green et al. 2004). The global consumption of diclofenac is estimated to be 940 tons per year, with detection concentration in surface waters, groundwaters, and even in drinking waters in the range of ng L^{-1} to $\mu\text{g L}^{-1}$ in the last years (Malefane et al. 2019). Extensive studies on SD metabolism in humans and animals have been conducted (Fei et al. 2006; Tiwari et al. 2015), and it was discovered that after ingestion, unchanged diclofenac, together with its hydroxylated metabolites, are excreted by the body through urine and feces (Stülten et al. 2008). These compounds enter wastewater treatment plants (WWTP) where the removal percentage of SD typically ranges from 21% to 40% during the treatment processes (de Luna et al. 2017).

The adverse effects of these pharmaceutical compounds, such as SD, which has been proven to be ecotoxic and affects both aquatic and terrestrial ecosystems (Oaks et al. 2004),

have led to strict regulations that require the elimination of pharmaceutical residues from industrial wastewaters. PPCPs are commonly removed from wastewaters by membrane filtration (de Luna et al. 2019), advanced oxidation processes (Lin et al. 2016), photodegradation (Paragas et al. 2018), and adsorption (Abo El Naga et al. 2019). However, the large-scale application of these technologies is hindered by the high cost of installation. Among the various treatment systems for pharmaceutical wastewaters, adsorption is by far one of the most promising. In particular, the use of biochar as sorbent material addresses the high cost of commercially available activated carbon.

Biochar is a carbonaceous product of biomass thermal conversion under low-oxygen environment (Kim et al. 2020). It is the prime product of torrefaction at 200–300 °C, and a pyrolysis by-product at 300–600 °C (Genuino et al. 2018; Arriola et al. 2020). It is an organic-rich material that is effective in the adsorption of pharmaceuticals (Genuino et al. 2017). Availability, abundance, low cost, and the renewable nature of the adsorbent material, along with the simple preparation method and ease of process operations, are among the advantages of this technology (Antunes et al. 2012; Ifthikar et al. 2017). Several agricultural wastes and other materials like municipal solid wastes have been used as biochar precursors for decontaminating industrial/domestic wastewater (Sumalinog et al. 2018). A particularly abundant agricultural waste that can be used for biochar production is rice hull, which is the residue obtained from the rice milling process. While rice hulls have been used as fuel in boiler furnaces of various industries to produce steam, these agricultural residues have limited or no practical use in the Philippines. The annual generation of rice hulls exceeds 140 million tons worldwide, 96% of which comes from developing countries (Kalderis et al. 2008). Since the rice industry is expected to continue for a long time, with predicted total rice consumption at 450 million tons by 2020 (milled basis) (Lim et al. 2012), the availability of rice by-products will remain high. Thus, utilization of rice hull as adsorbent can add value to this waste material and significantly contribute to effective waste management. Several studies have reported on the application of rice hull biochar (RHB) for dye (Khan et al. 2020) and heavy metals (Han et al. 2020) removal. However, the application of RHB for the removal of SD from water matrices has not yet been fully explored.

Biochar adsorbents derived from rice hulls were previously prepared in various researches at temperatures ranging from 270 °C to 700 °C (Leksungnoen et al. 2019; Shi et al. 2019), with some studies utilizing a variety of modifications, such as microwave-assisted pyrolysis, steam activation, acid activation, alkaline activation, CO_2 activation and electrochemical modification (Wang et al. 2020b). However, rice hull adsorbents may also be prepared through torrefaction at lower temperatures (200–300 °C) (Arpia et al. 2021) and

without modifications or use of chemical activating agents (de Luna et al. 2013). This process conforms with the essential advantages of using biochar as adsorbents which are low cost and compatibility with green manufacturing (Wang et al. 2018). Compared with biochar modification processes that require high activation temperatures and chemical activating agents, the process used in this study is more economical, requires low energy consumption, and is environment friendly.

The study aims to determine the applicability of RHB in removing SD from aqueous solutions. Parametric studies were done to determine the effects of operating parameters such as initial SD concentration, RHB dosage, initial solution pH, and contact time on SD removal. Box–Behnken design (BBD) was used to determine the optimum conditions of these parameters. Isotherm and kinetic studies were employed to identify the best-fitting models, as well as the dominant mechanism of the adsorption process.

2 Materials and methods

2.1 Preparation of SD aqueous solutions and rice hull adsorbents

All chemicals used in this study were of analytical grade. A synthetic SD stock solution (500 mg L^{-1}) was prepared by dissolving diclofenac sodium salt ($\text{C}_{14}\text{H}_{10}\text{Cl}_2\text{NNaO}_2$, 99%, Sigma-Aldrich) in distilled water. Working SD solutions were prepared by diluting the stock solution with distilled water.

Rice hulls were obtained from a rice mill in Muñoz, Nueva Ecija, Philippines. The rice hulls were washed thoroughly with distilled water to remove particles and water-soluble impurities. The washed hulls were dried in an oven at 100°C for 30 min and pulverized in a Thomas-Wiley laboratory mill. The powdered hulls were sieved to obtain particles that were $300\text{--}500 \mu\text{m}$ in size and severely torrefied in an oven at 350°C for 1 h. The RHB powder was then stored in clean and dry containers.

2.2 Analytical methods

The elemental compositions of raw rice hulls (RRH) and RHB were examined by energy dispersive X-ray analysis (EDX, TM-1000, Hitachi). The functional groups involved in SD adsorption were analyzed by Fourier-transform infrared (FTIR) spectroscopy (Nicolet 6700, Thermo Scientific). The point-of-zero charge (pH_{pzc}) was determined by mass titration method that used increasing amounts of RHB added to water to obtain suspensions of 0.01, 0.1, 1, 5, 10, and 20% by weight. These samples were agitated for 24 h and then filtered. The final pH of each suspension was measured

after 24 h of contact time for which the value of pH equilibrium was reached. This value was approximately equal to the adsorbent pH_{pzc} .

Sample SD concentrations were determined following the study by Matin et al. (2005). SD-containing samples (2 mL) were combined with 1 mL concentrated nitric acid (HNO_3 63% w/v, Merck) to produce a yellowish compound diluted to 10 mL with distilled water. The absorbance of the resulting solution was read at maximum wavelength (λ_{max}) of 365 nm using a UV–Vis spectrophotometer (Model UV 1601, Shimadzu). Diclofenac removal was computed using the following equation:

$$\text{SD Removal (\%)} = \frac{C_0 - C}{C_0} \times 100. \quad (1)$$

2.3 Batch adsorption experiments: optimization, isotherm, and kinetic studies

Erlenmeyer flasks containing 20 mL samples of varying initial SD concentrations, RHB dosage, initial pH, and contact time were agitated using a magnetic stirrer at 300 rpm and 25°C . The pH of the solution was adjusted with 0.1 M sodium hydroxide (NaOH 99%, Merck) and 0.1 M hydrochloric acid (HCl 99%, Merck). Box–Behnken design with three levels was used to optimize process parameters, namely initial SD concentration (30, 65, 100 mg L^{-1}), RHB dosage (0.1, 0.3, 0.5 g), initial pH (2, 5.5, 9), and contact time (10, 35, 60 min). Adsorption experiments were carried out at optimum conditions. For the isotherm study, SD solutions of different initial SD concentrations (20, 40, 60, 80, and 100 mg L^{-1}) were prepared at pH 2 and pH 7. Kinetic studies were conducted at varying durations (10, 20, 30, 40, 50, and 60 min). All the experiments were performed in triplicates. The adsorption capacity (Q) was computed using the following equation:

$$Q = \frac{(C_0 - C) \text{ volume of solution}}{\text{mass of adsorbent}}. \quad (2)$$

3 Results and discussion

3.1 Adsorbent characterization

Raw rice hulls contain cellulose, hemicellulose, lignin, extractives, water and mineral ash. The mineral content include oxides of silicon, potassium, magnesium, iron, aluminum, and calcium (Nakbanpote et al. 2000). Figure 1 shows that RRH was composed mainly of carbon (16.53 wt%), oxygen (40.51 wt%), and silicon (42.96 wt%). Similar findings were previously reported (Souza et al. 2002). After severe torrefaction,

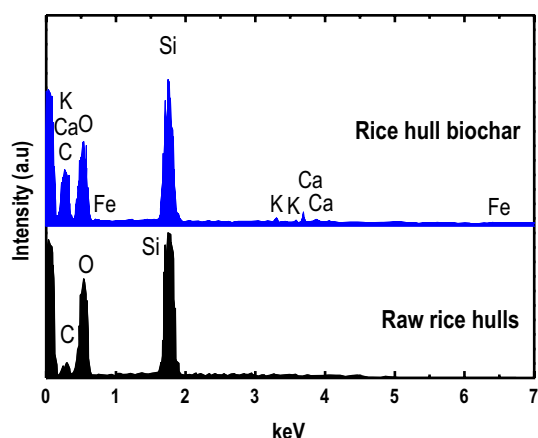


Fig. 1 EDX atomic composition of raw rice hulls (RRH) and rice hull biochar (RHB)

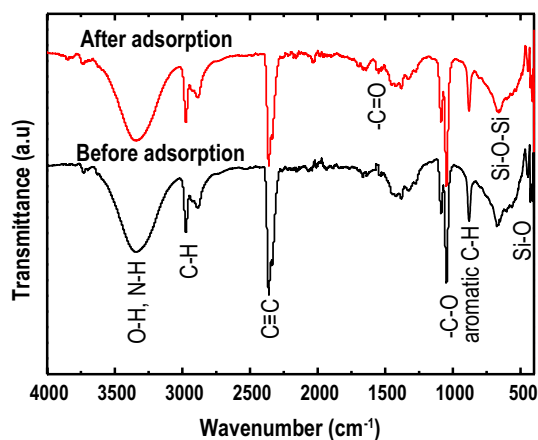


Fig. 2 FTIR spectra of RHB before and after adsorption

other elements from the mineral ash, such as potassium, calcium, and iron, were liberated and exposed. In addition to this, the atomic composition of carbon significantly increased from 25.3% to 40.48% after torrefaction, thus making RHB a more effective adsorbent compared to RRH (Nakbanpote et al. 2000; Wang et al. 2001; Souza et al. 2002).

The results of the FTIR analysis of RHB before and after adsorption are shown in the FTIR spectra presented in Fig. 2. The broad peak at 3300–3400 cm^{-1} corresponds to the stretching of OH groups due to SiOH and adsorbed water (de Luna et al. 2013). The peak at 3000 cm^{-1} and those at 2100–2400 cm^{-1} are attributed to C-H stretching and $\text{C}\equiv\text{C}$ stretching, respectively. Indications of aromatic C-H and C-O stretching are seen in the peaks at 800–1000 cm^{-1} and 1000–1100 cm^{-1} , respectively (de Luna et al. 2013). Si-O bond stretching vibration is also observed in the peak at 700 cm^{-1} (Chen et al. 2012). In addition, the peak at

1700 cm^{-1} , which corresponds to C=O stretching vibrations, resulted in the largest shift to 1900 cm^{-1} after SD adsorption (Coates 2000; Jeon 2011). This indicates that the stretching of the C=O in the carbonyl or carboxyl groups participated the most in the SD adsorption process (Yu et al. 2013).

3.2 Optimization of the adsorption parameters

In this study, Box–Behnken design (BBD) was used to investigate the effects of important operating parameters in the removal of SD using RHB adsorbent, and to determine the optimum condition for SD adsorption. BBD is a type of response surface method (RSM) which is based on a three-level incomplete factorial design, and requires fewer runs compared to other RSM designs (de Luna et al. 2012). From Box–Behnken design, the highest SD removal was 97% using the following experimental conditions: initial SD concentration = 100 mg L^{-1} , solution $\text{pH}=2$, RHB dosage = 0.3 g and contact time = 35 min (Table 1). In contrast, the lowest SD removal was 41% at initial SD concentration = 65 mg L^{-1} , $\text{pH}=9$, RHB dosage = 0.1 g and contact time = 35 min. This is consistent with the findings on the effect of varying initial solution pH on SD removal.

The results of the analysis of variance (ANOVA) presented in Table 2 show that the response surface quadratic model obtained was significant, with model F -value at 9.93. This indicates that the model can explain most of the variations in the response (de Luna et al. 2015). The high coefficient of determination (R^2) of 0.9145 and an adjusted R^2 value of 0.8224 indicate that the quadratic model is reliable in predicting responses. Likewise, all main parameters (initial SD concentration, RHB dose, initial pH , and contact time) and the interactions between initial SD concentration and contact time were significant, with p values of less than 0.05. Three-dimensional (3D) response surface plot (Fig. 3) was generated using RSM to show the significant interaction effects between contact time and initial SD concentration. As observed, longer contact time and higher initial SD concentration resulted in higher SD removal. This is attributed to the increased interaction between SD molecules and the available binding sites on the RHB surface.

The quadratic model equation in terms of coded factors is expressed by the following equation:

$$Y = 76.797 + 13.262X_1 + 6.999X_2 - 14.921X_3 + 6.899X_3 + 5.655X_1X_2 + 5.037X_1X_3 + 9.311X_1X_4 + 2.816X_2X_3 - 4.000X_2X_4 + 1.379X_3X_4 - 6.914X_1^2 - 4.964X_2^2 - 0.452X_3^2 - 5.911X_4^2, \quad (3)$$

where Y is the predicted response variable for SD removal and X_1 , X_2 , X_3 and X_4 correspond to the independent

Table 1 BBD for SD removal using RHB

Run	X_1 : Initial SD conc (mg L ⁻¹)	X_2 : RHB Dosage (mg)	X_3 : Initial pH	X_4 : Contact Time (min)	Actual SD removal (%)	Predicted SD removal (%)
1	30	100	5.50	35	50	48
2	100	100	5.50	35	71	63
3	30	500	5.50	35	42	51
4	100	500	5.50	35	86	89
5	65	300	2.00	10	80	79
6	65	300	9.00	10	41	46
7	65	300	2.00	60	93	90
8	65	300	9.00	60	60	62
9	65	300	5.50	35	83	75
10	65	300	5.50	35	70	75
11	30	300	5.50	10	55	51
12	100	300	5.50	10	57	59
13	30	300	5.50	60	45	46
14	100	300	5.50	60	85	91
15	65	100	2.00	35	74	81
16	65	500	2.00	35	91	89
17	65	100	9.00	35	41	45
18	65	500	9.00	35	70	65
19	65	300	5.50	35	77	75
20	65	300	5.50	35	83	75
21	30	300	2.00	35	82	81
22	100	300	2.00	35	97	98
23	30	300	9.00	35	45	41
24	100	300	9.00	35	81	78
25	65	100	5.50	10	52	52
26	65	500	5.50	10	76	74
27	65	100	5.50	60	76	74
28	65	500	5.50	60	84	80
29	65	300	5.50	35	72	81
30	65	300	5.50	35	76	81

variables: initial SD concentration, RHB dosage, initial pH and contact time. From Eq. (3), higher initial SD concentration, RHB dosage, and contact time and lower initial pH will result in better SD removals. Table 1 also shows the proximity of the predicted and actual percent SD removals, confirming the reliability of the quadratic model.

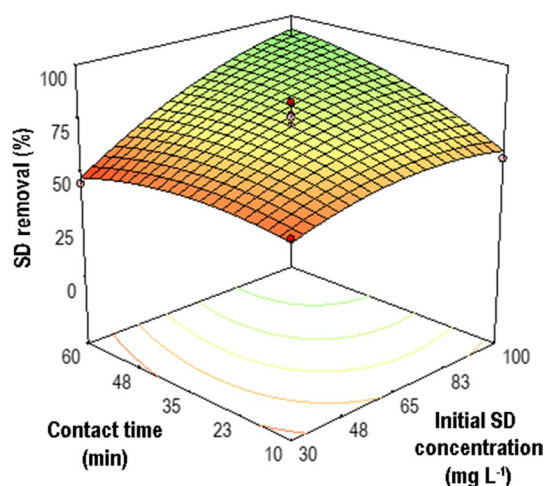
The numerical optimization of process parameters achieved a 96% SD removal (predicted) at 100 mg L⁻¹ initial SD concentration, 0.5 g of RHB dose and pH 2. At this condition, the residual SD concentration was found at 4 mg L⁻¹ and the adsorption capacity was at 3.3 mg g⁻¹. Experimental validation of the model at optimum conditions resulted in 95.5% SD removal, 3.6 mg L⁻¹ residual SD concentration and adsorption capacity of 3.4 mg g⁻¹. Based on the percent error (0.52% for SD removal), the model generated is robust and reliable (de Luna et al. 2015).

3.3 Factors affecting SD removal

The effect of initial pH of the water matrix on SD removal is shown in Fig. 4a. The highest SD removals were obtained at pH 2 for both 0.1 and 0.5 g RHB dosages. It is evident from Fig. 4a that SD removal decreased as the solution pH was raised, with the lowest removal observed at pH 7. The pH point-of-zero charge (pH_{pzc}) of RHB is at pH 6.21, as determined by mass titration. At pH 2 ($pH < pH_{pzc}$), the surface of the RHB is positively charged, whereas at pH 7 ($pH > pH_{pzc}$), it is negatively charged (de Luna et al. 2017). Furthermore, SD is an acid and has a dissociation constant (pK_a) value of 4.15. At $pH < pK_a$, the SD molecules are positively charged, while at the $pH > pK_a$, these molecules are negatively charged (de Luna et al. 2017). Thus, at pH 7 where the solution pH was greater than both the adsorbent pH_{pzc} and

Table 2 ANOVA for the response surface model for SD removal using RHB

Source	Sum of squares	df	Mean square	F value	p value	Prob > F
Model	7237.57	14	516.97	9.93	< 0.0001	Significant
X_1 -initial SD conc. (mg L^{-1})	2107.25	1	2107.25	40.48	< 0.0001	Significant
X_2 -RHB dosage (mg)	588.00	1	588.00	11.30	0.0051	Significant
X_3 -initial pH	2671.49	1	2671.49	51.32	< 0.0001	Significant
X_4 -contact time (min)	571.11	1	571.11	10.97	0.0056	Significant
X_1X_2	127.91	1	127.91	2.46	0.1410	Not significant
X_1X_3	101.48	1	101.48	1.95	0.1860	Not significant
X_1X_4	346.74	1	346.74	6.66	0.0228	Significant
X_2X_3	31.72	1	31.72	0.61	0.4490	Not significant
X_2X_4	64.00	1	64.00	1.23	0.2876	Not significant
X_3X_4	7.61	1	7.61	0.15	0.7084	Not significant
X_1^2	327.81	1	327.81	6.30	0.0261	Significant
X_2^2	168.95	1	168.95	3.25	0.0948	Not significant
X_3^2	1.40	1	1.40	0.027	0.8723	Not significant
X_4^2	239.62	1	239.62	4.60	0.0514	Not significant
Residual	676.69	13	52.05			
Lack of fit	559.67	10	55.97	1.43	0.4252	Not significant
Pure error	117.02	3	39.01			
Cor total	8192.21	29				

**Fig. 3** 3D response surface plot of the significant interaction effects of contact time and initial SD concentration on SD removal

the adsorbate pK_a , both RHB and SD had negatively charged surfaces. This explains the low SD removal at higher solution pH, as the increasing electrostatic repulsion reduced the adsorption of SD onto the RHB. When pH is around pK_a of 4.15, the increase of negative species of SD may cause a decrease of removal efficiency. At $\text{pH}=2$, most of the SD are neutral whereas the adsorbent is positively charged, thus the hydrogen bonding between SD (carbonyl oxygen) and adsorbent (hydroxyl hydrogen) may exist. Also, at lower solution

pH, the carboxyl groups ($-\text{COOH}$; $\text{pK}_a=1.7\text{--}4.7$) formed negatively charged carboxylate groups ($-\text{COO}^-$) (Tran et al. 2017). These adsorbent binding sites were mainly responsible for the high SD removal through interaction with the positively charged SD molecules, even at pH levels below pH_{pzc} . This can be seen from the FTIR results that show large shifts of $\text{C}=\text{O}$ of carbonyl/carboxyl groups before and after adsorption. For comparison, pH 2 and pH 7 were used in the succeeding experiments.

Figure 4b shows the relationship between SD removal and initial SD concentration (mg L^{-1}) at two different solution pH. As the initial SD concentration was raised from 30 to 100 mg L^{-1} at solution pH of 2, SD removal improved from 88% to 97%. At higher solution pH of 7, SD removals also improved from 61% to 77% for 30 and 100 mg L^{-1} SD solutions, respectively. These indicate that the SD removals are dependent on initial SD concentration. The observed high SD removal at the low initial SD concentration of 30 mg L^{-1} , especially at pH 2, can be attributed to the lower amount of SD molecules competing for sorption onto the RHB surface (Ong et al. 2018). The improved SD removal as the initial SD concentration was raised was due to the increased interaction through collision between the SD molecules and the available binding sites on the RHB, thereby resulting in higher probability of these molecules to attach to the adsorbent (Mane et al. 2007).

Figure 4c shows the relationship between SD removal and RHB dosage. At solution pH of 2 and as RHB

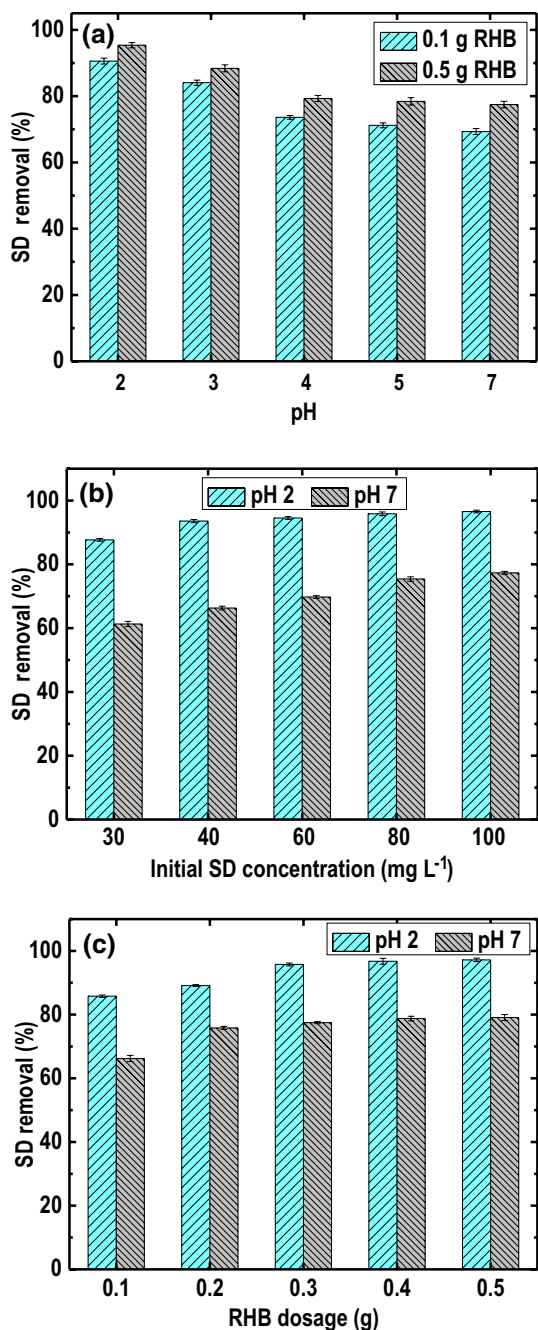


Fig. 4 SD removal at varying **a** initial pH, **b** initial SD concentration, and **c** RHB dosage

dosage increased from 0.1 to 0.5 g, SD removal significantly improved from 86% to 97%. A similar trend was observed at pH 7 where a noticeable rise in SD removal (66–79%) occurred as RHB dosage was increased from 0.1 to 0.5 g. The improvement in SD removal at higher RHB dosage is attributed to the greater number of available binding sites for interaction with SD molecules (Overah 2011).

3.4 Adsorption isotherms

One of the most important criteria in selecting a suitable adsorbent is by studying the adsorption isotherm which provides fundamental physiochemical data for evaluating the adsorption capacities of an adsorbent. The data obtained at the two pH levels were fitted using Langmuir, Freundlich, and Temkin adsorption equations.

The Langmuir isotherm model represents the equilibrium distribution of pollutant molecules between the solid and liquid phases. It is valid for monolayer adsorption onto a surface containing a finite number of identical sites (Langmuir 1918). The model assumes uniform energies of adsorption onto the surface and no transmigration of adsorbate occurs in the plane of the surface. Formation of a monolayer adsorbate on the outer surface of the adsorbent eventually reaches a saturation point where the maximum adsorption of the surface is achieved and no further adsorption takes place (Boparai et al. 2011). Langmuir isotherm model is expressed mathematically in Eq. (4). The values of Q_m and b are computed from the slope and y-intercept of the plot of $1/Q$ and $1/C_e$.

$$\frac{1}{Q} = \left[\frac{1}{bQ_m} \right] \frac{1}{C_e} + \frac{1}{Q_m} \quad (4)$$

The separation factor, R_L , also called the equilibrium parameter, is a dimensionless constant used to validate if experimental data conform to the Langmuir isotherm. The criteria for applicability to the Langmuir isotherm depends on the R_L value as computed using Eq. (5) where: ($R_L = 1$) follows a linear isotherm, ($R_L > 1$) means an unfavorable isotherm, ($R_L < 1$) corresponds to a favorable isotherm, and ($R_L = 0$) is an irreversible isotherm.

$$R_L = \frac{1}{1 + bC_o} \quad (5)$$

Meanwhile, the Freundlich isotherm assumes heterogeneous surface and adsorption sites with varying energy of adsorption (Freundlich 1906). The model assumes that the stronger binding sites on the heterogeneous surface are initially occupied, and a decrease in binding strength occurs as the degree of site occupation is increased. The Freundlich isotherm can be expressed as follows:

$$\ln Q = \ln K_f + \frac{1}{n} \ln C_e \quad (6)$$

The slope of $1/n$ ranging between 0 and 1 is a measure of adsorption intensity or surface heterogeneity, becoming more heterogeneous as its value approaches zero. A value for $1/n$ below one indicates normal Langmuir isotherm.

The Temkin isotherm equation is characterized by a uniform distribution of binding energies (Temkin and Pyzhev 1940). The slope and intercepts are determined by plotting the quantity adsorbed Q against $\ln C_e$ and the constants and take into account the presence of indirect interactions between the adsorbent and adsorbate as shown in Eq. (7). The model assumes that heat of adsorption (function of temperature) of all molecules in the layer decreases with coverage linearly rather than logarithmically.

$$Q = \beta \ln \alpha + \beta \ln C_e \quad (7)$$

Shown in Fig. 5 are the plots of the equilibrium adsorption isotherm of diclofenac onto the RHB surface, Langmuir, Freundlich, and Temkin isotherms for initial pH 2 and pH 7, respectively, while Table 3 shows the values for the isotherm parameters. In this study, the Langmuir isotherm gave the highest coefficient of determination for both pH 2 ($R^2 = 0.9827$) and pH 7 ($R^2 = 0.9460$). This indicates that monolayer coverage of SD onto the RHB surface, which contained homogenous and energetically equivalent sites for adsorption, is favored (Ifthikar et al. 2018). Likewise,

Table 3 Isotherm parameters for SD removal using RHB

Parameter	Value	
	pH 2	pH 7
Langmuir		
q_m (mg g ⁻¹)	14.7929	11.8483
b (L mg ⁻¹)	0.1703	0.0052
R^2	0.9827	0.9460
Freundlich		
K_f (mg ^{1-1/n} L ^{1/n} g ⁻¹)	2.0943	0.0618
n	1.1148	0.9831
R^2	0.9740	0.9258
Temkin		
α (L g ⁻¹)	2.3621	0.1439
β (J mol ⁻¹)	2.3458	2.3349
R^2	0.8040	0.8332

no transmigration of diclofenac happened in the layer of the RHB surface (de Luna et al. 2013). For the separation factor, R_L , the values ranged from 0.73 to 0.92 at pH 2 and 0.64

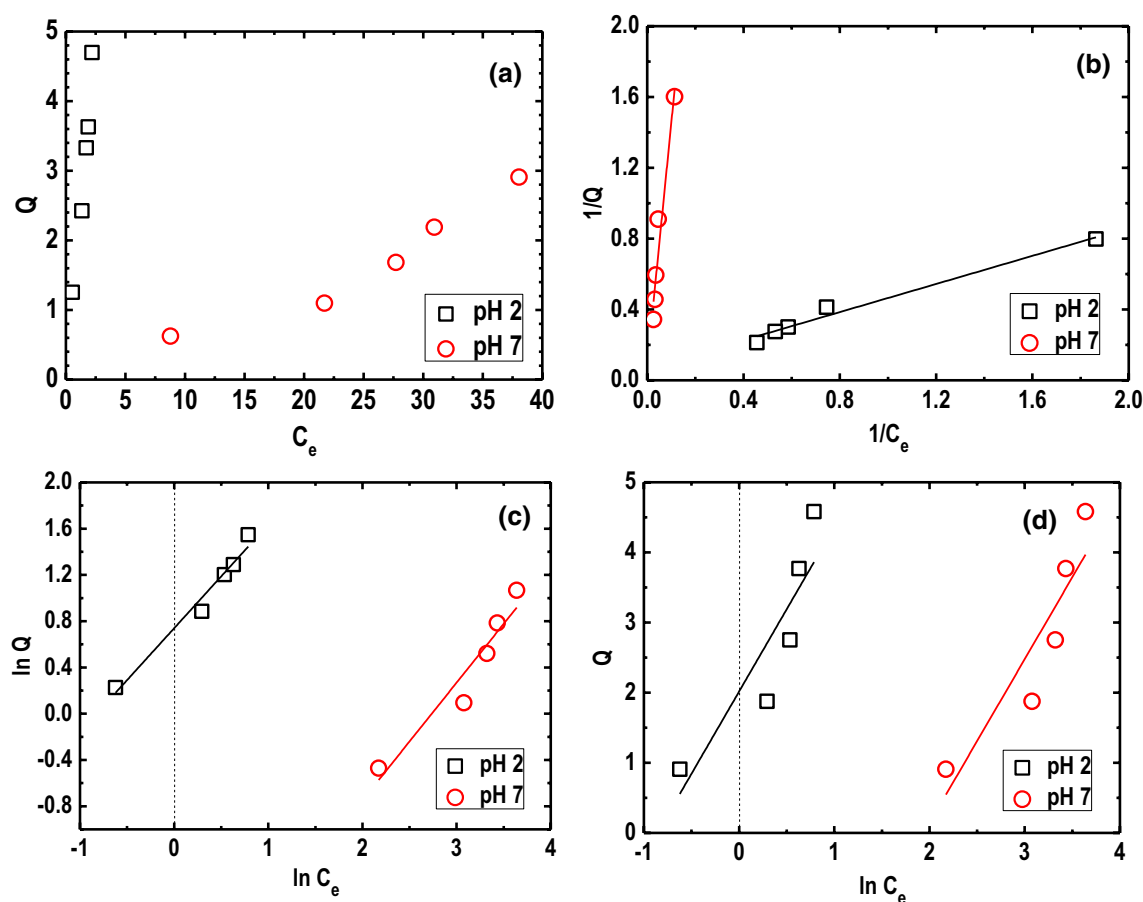


Fig. 5 a Adsorption isotherm of SD over RHB, b Langmuir, c Freundlich and d Temkin isotherm plots at pH 2 and pH 7

to 0.89 at pH 7, confirming that the isotherm is favorable at both pH levels. Thus, RHB is a suitable adsorbent for SD.

3.5 Adsorption kinetics

Kinetic modeling was done using pseudo-first-order, pseudo-second-order, and intraparticle diffusion equations for both pH 2 and pH 7 solutions to determine the dominant mechanism that controls the adsorption rate, as shown in Eqs. (8, 9, 10). Pseudo-first-order kinetic model (Lagergren 1898) describes the rate of pollutant removal as directly proportional to the pollutant concentration and assumes physisorption as the rate-controlling mechanism. The integrated linear form of the pseudo-first-order model is shown in the following equation:

$$\log(Q - q_t) = \log Q - \frac{k_1}{2.303}t. \quad (8)$$

On the other hand, the pseudo-second-order kinetic model (Ho and McKay 1998) maintains that the rate of pollutant degradation is directly proportional to the square of pollutant concentration and suggests chemisorption as the

rate-controlling step in the adsorption process. This equation indicates the fast decrease in pollutant concentration in the aqueous solution at a short time of contact suggesting an electrostatic interaction. The linearized form of the pseudo-second-order model is shown in the following equation:

$$\frac{t}{q_t} = \frac{1}{k_2 Q^2} + \frac{1}{Q}t. \quad (9)$$

The intraparticle diffusion model describes the transport of the solute from the bulk of the solution onto the adsorbent through an intraparticle process or pore diffusion. The intraparticle diffusion model demonstrating a linear relationship means that the pore diffusion also affects the rate of adsorption. The intraparticle diffusion model is given in the following equation:

$$q_t = k_{id}t^{0.5} + C. \quad (10)$$

If the rate-limiting step is intraparticle diffusion, a plot of solute adsorbed against the square root of the contact time should yield a straight line passing through the origin (Oladaja et al. 2008). However, if the y-intercept, C , is not equal to zero, the mechanism does not uniquely limit the

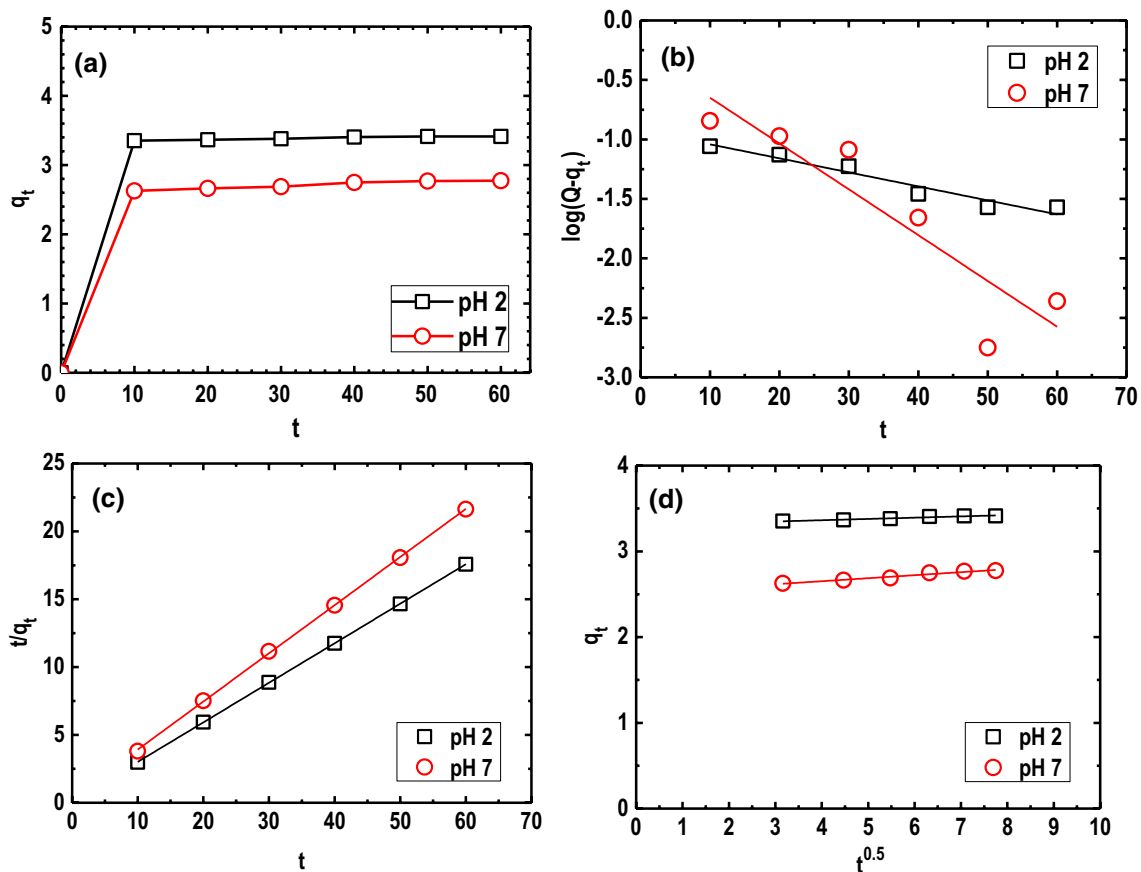


Fig. 6 a Effect of contact time, b pseudo-first-order c pseudo-second-order and d intraparticle diffusion kinetic plots at pH 2 and pH 7

Table 4 Kinetic parameters for SD removal by RHB

Parameter	Value	
	pH 2	pH 7
Pseudo-first order		
Q (mg g ⁻¹)	0.1192	0.5438
k_1 (min ⁻¹)	0.0272	0.0887
R^2	0.9426	0.8293
Pseudo-second order		
Q (mg g ⁻¹)	3.4317	2.8193
k_2 (g mg ⁻¹ min ⁻¹)	0.8647	0.3393
R^2	0.9999	0.9998
Intraparticle diffusion		
C (mg g ⁻¹)	3.3033	2.511
k_{id} (mg g ⁻¹ min ^{-1/2})	0.0149	0.0351
R^2	0.9552	0.9651

overall process, but it may be a complex combination of adsorption and intraparticle transport (Cunha et al. 2010).

The kinetic data in Fig. 6a revealed that the adsorption process was fast during the first 10 min, and leveled off immediately afterwards when the adsorption equilibrium was achieved. From Table 4, the pseudo-second-order model provided the best fit, and the plot of t/q_t against t is shown in Fig. 6. This model best described the experimental data as shown by the high coefficients of determination ($R^2=0.9999$) for both SD removals at pH 2 and pH 7. This suggests that chemisorption controlled the rate of the adsorption process (Wang et al. 2020a). This mechanism can be described as complexation of the SD molecules with the functional groups present on the RHB, with C=O in the carbonyl or carboxyl groups having the most significant participation as indicated in the large shift of peak attributed to C=O stretching vibrations observed in Fig. 2. As shown in Table 4, the adsorption capacity values ($q_{pH\ 2}=3.4$ and $q_{pH\ 7}=2.8$ mg g⁻¹) determined using regression of the concentration against time were very close to the values ($q_{pH\ 2}=3.4$ and $q_{pH\ 7}=2.3$ mg g⁻¹) obtained during optimization. This renders the pseudo-second-order model more reliable. The value of the pseudo-second-order rate constant, k_2 , was higher at pH 2. Thus, SD molecules have high probability of binding to the functional groups on the RHB at low solution pH.

4 Conclusions

The RHB prepared via torrefaction was found effective in removing SD from water matrices. EDX analysis revealed that the elemental composition of the RHB adsorbent surface before and after torrefaction showed an increase in carbon atomic percentage from 25.3% to 40.48%. FTIR spectra for

RHB before and after adsorption revealed that C=O stretching vibrations that correspond to the carbonyl or carboxyl groups had major participation in the adsorption process. In the adsorption study, the increase in SD concentration, contact time, and RHB dosage improved SD removal up to 97%, while the increase in the solution pH reduced SD removal. At optimum operating conditions (initial SD concentration = 100 mg L⁻¹, RHB dosage = 0.5 g, pH = 2), SD removal reached 96% and the calculated equilibrium adsorption capacity was 3.3 mg g⁻¹. In addition, the Langmuir isotherm and pseudo-second order kinetics best described SD adsorption on RHB, with the highest coefficients of determination at 0.9827 and 0.9999, respectively. Overall, the biochar adsorbents produced via the torrefaction of rice hulls and in the absence of activating chemicals showed excellent potential for the removal of pharmaceutical compounds such as sodium diclofenac from water matrices.

Acknowledgements The authors would like to thank the Department of Science and Technology, Philippines for providing financial support for this research undertaking.

Author contributions Conceptualization: JQF, MDGDL; methodology: JQF, KKR and SMBPO; investigation: RRMA, KKR and DCO; validation: JQF, SMBPO and RRMA; formal analysis: JQF, RRMA, DCO and SMBPO; resources: MDGDL; data curation: KKR, RRMA, DCO; writing original draft: JQF, KKR, DCO; writing—review and editing: DCO, SMBPO and MDGDL; visualization: RRMA, DCO and KKR; supervision: MDGDL; project administration: MDGDL; funding: MDGDL.

Compliance with ethical standards

Conflict of interest The authors declare no competing financial interest.

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