

NGS DATA ANALYSIS

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```
setwd("D:/R/Pan_NGS_data/Rna_seq/Results")
# STEP 2 Define the samples with their read IDs (SRA run accessions) and calculate RPKM values...
samples_name <- c("AMB + LF rep 1", "AMB + LF rep 2", "AMB + LF rep 3",
"control 1", "control 2", "control 3")
names(samples_name) <- c("SRR3396388", "SRR3396389", "SRR3396390", "SRR3396391", "SRR3396392", "SRR3396392")
```

```
# Construct the file path -
files <- c()
```

```
for (s in samples_name) {
  fp <- paste0(s, ".bam")
  if (file.exists(fp)) files <- c(files, fp)
}
```

```
# Load in the BAM files
bams_files <- BamFileList(files, yieldSize = 100000, asMates=TRUE)
# Overlap BAM reads and genes
overlap <- summarizeOverlaps(exons, bams_files, mode="Union", singleEnd=FALSE, ignore.strand=TRUE, fragments=TRUE)
overlap
```

```
## class: RangedSummarizedExperiment
## dim: 7127 6
## metadata(0):
## assays(1): counts
## rownames(7127): ETS1-1 ETS1-2 ... tY(GUA)O tY(GUA)Q
## rowData names(0):
## colnames(6): AMB + LF rep 1.bam AMB + LF rep 2.bam ... control 2.bam
## control 3.bam
## colData names(0):
```

```
# Extract the raw-reads per gene
geneRawReads <- assay(overlap)
geneRawReads[1:10,]
```

```
##          AMB + LF rep 1.bam AMB + LF rep 2.bam AMB + LF rep 3.bam control 1.bam
## ETS1-1              306              262              159
##      286
## ETS1-2              311              237              161
```

```

243
## ETS2-1          96          43          37
22
## ETS2-2          83          45          34
31
## HRA1            1           1           1
1
## ICR1           104          95          78
132
## IRT1           144          103         94
129
## ITS1-1          21          25          18
15
## ITS1-2          41          52          31
36
## ITS2-1          58          69          47
63
## control 2.bam control 3.bam
## ETS1-1          447          363
## ETS1-2          487          407
## ETS2-1          28           17
## ETS2-2          27           19
## HRA1            0            1
## ICR1           104          174
## IRT1           116          152
## ITS1-1          19           32
## ITS1-2          50           45
## ITS2-1          62           64

colnames(geneRawReads) <- gsub(".bam", "", colnames(geneRawReads))
write.table(geneRawReads, "gene_raw_reads.txt", sep = "\t", quote = FALSE)

# Calculate the RPKM values per gene
# RPKM = (10^9 * C)/(N * L)
# C = Number of reads mapped to a gene
# N = Total mapped reads in the sample
# L = gene length in base-pairs for a gene

# Get the total number of reads per samples
totalMappedReads <- apply(geneRawReads, 2, sum)

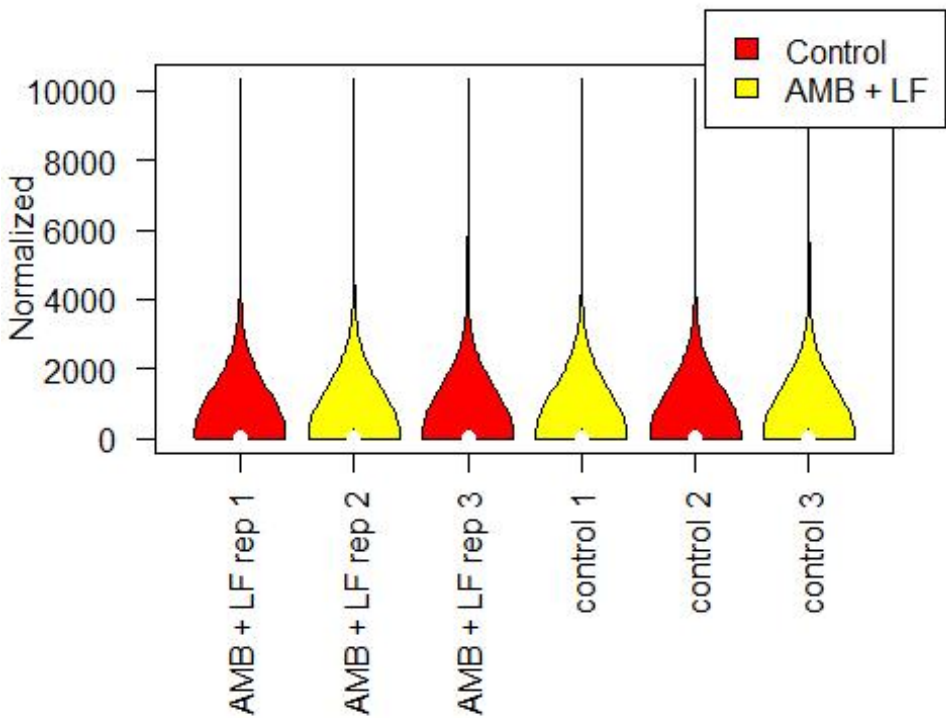
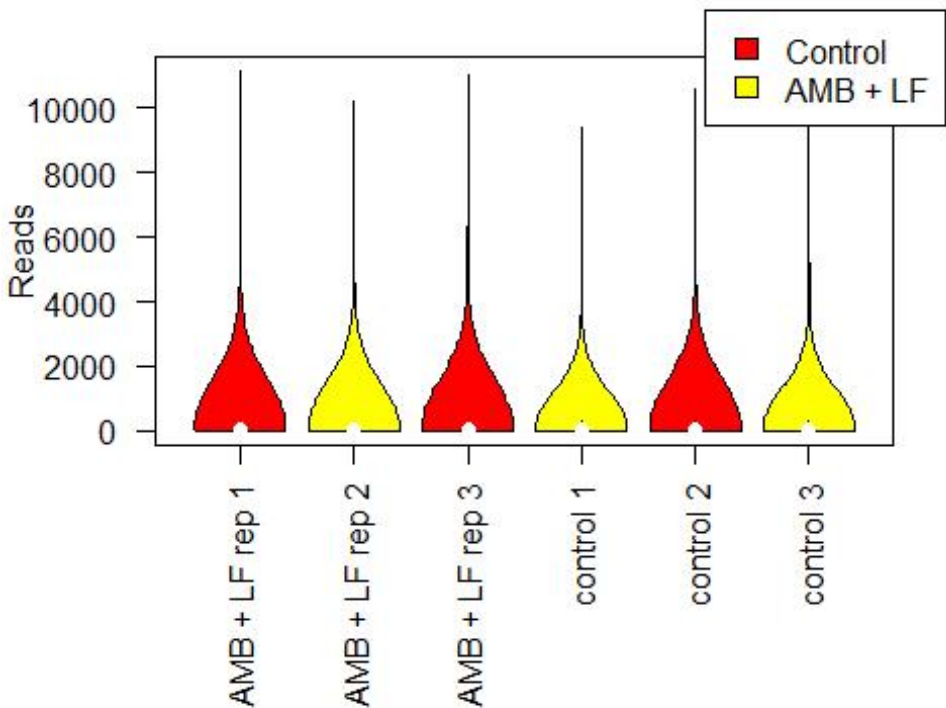
# Loop through all genes, compute RPKM
geneIdx <- 1
geneRPKM <- t(apply(geneRawReads, 1, function(C){
  geneLength <- as.numeric(gene.lengths[geneIdx])
  RPKM <- (10 ^ 9 * C) / (totalMappedReads * geneLength)
  geneIdx <- geneIdx + 1
  return(round(RPKM, d = 1))
})))

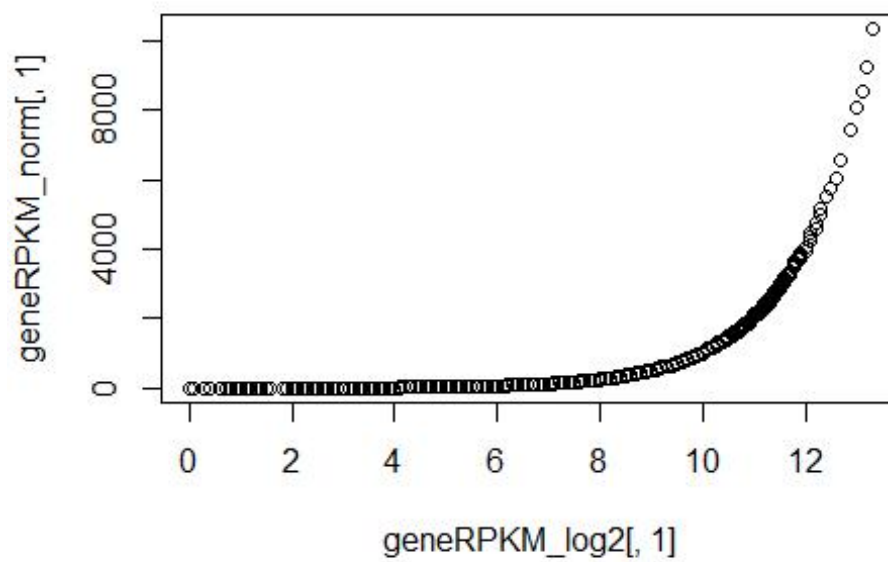
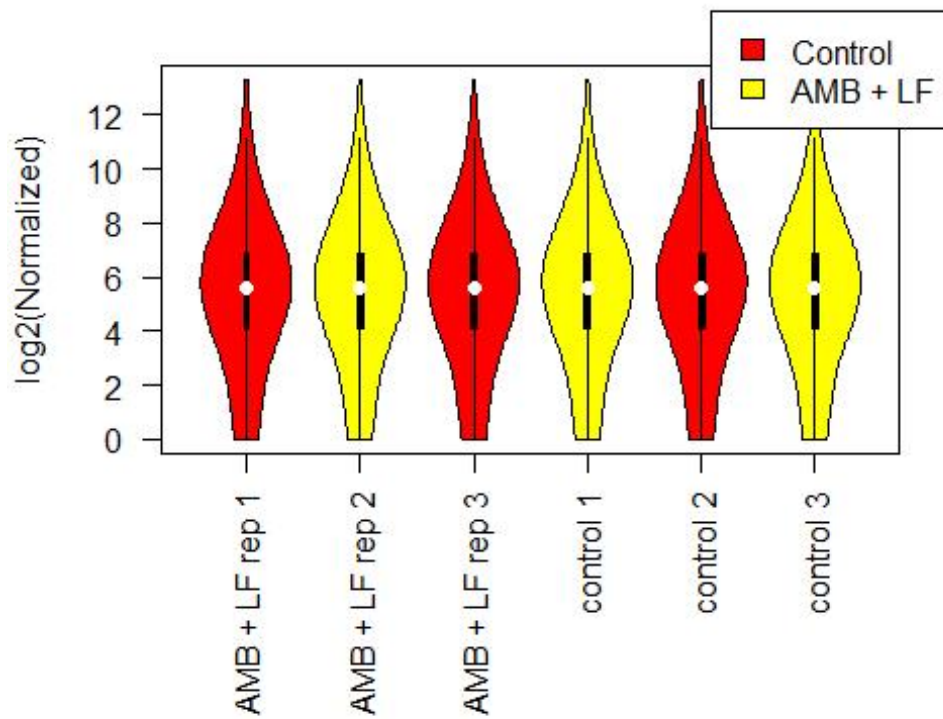
```

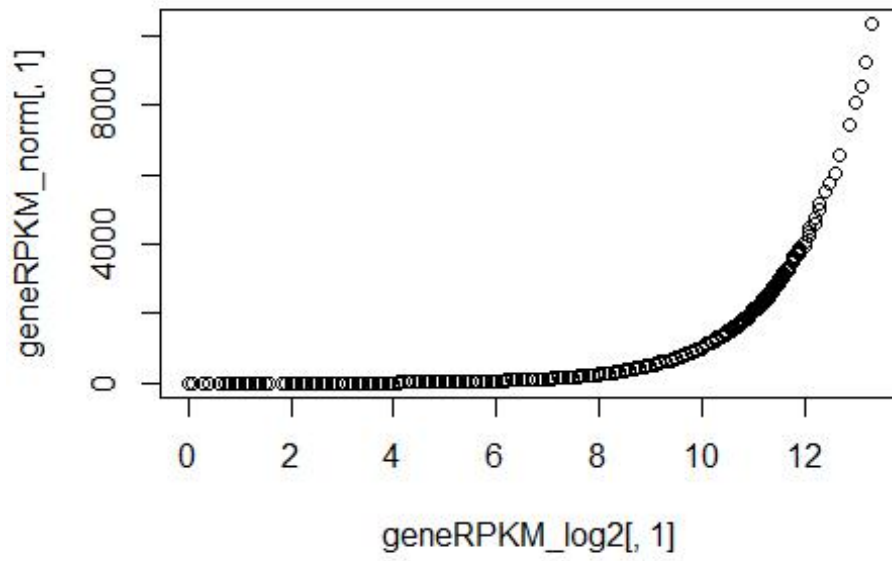
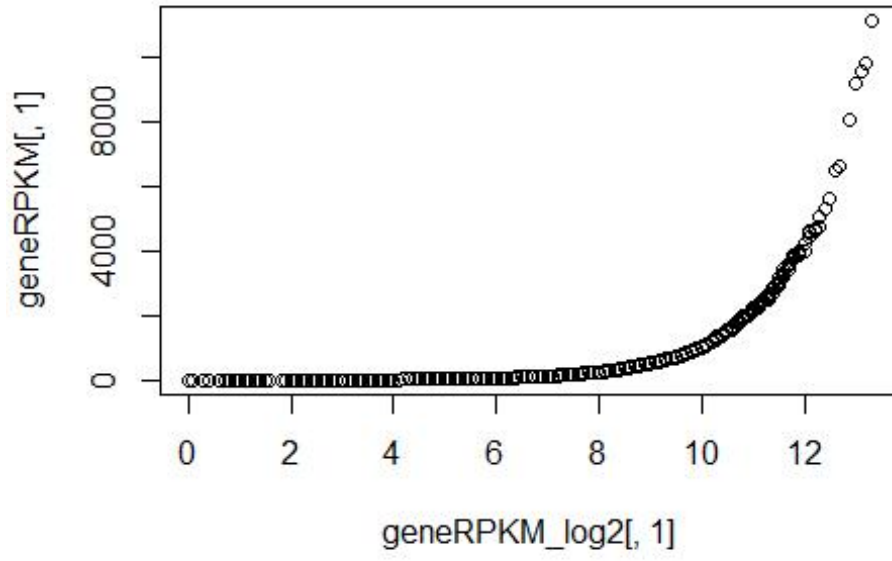
geneRPKM[1:10,]

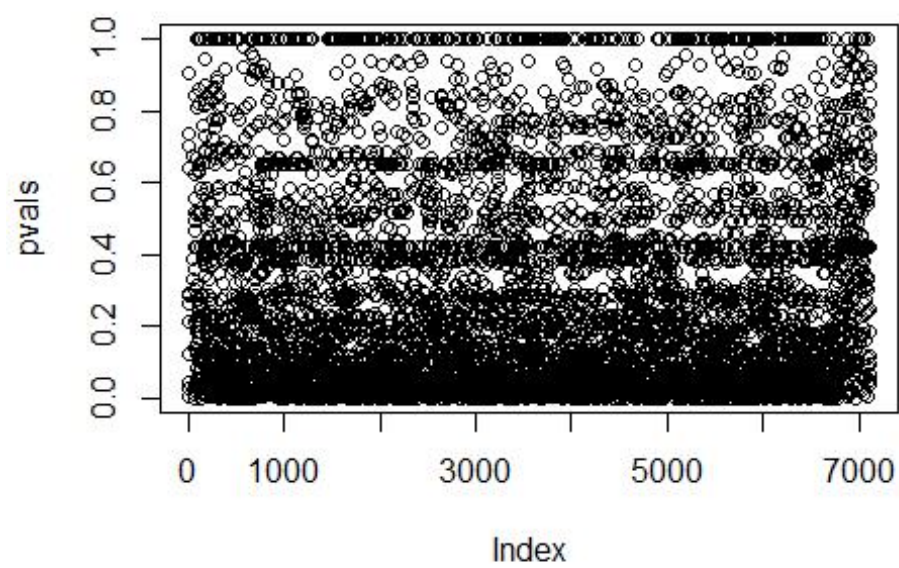
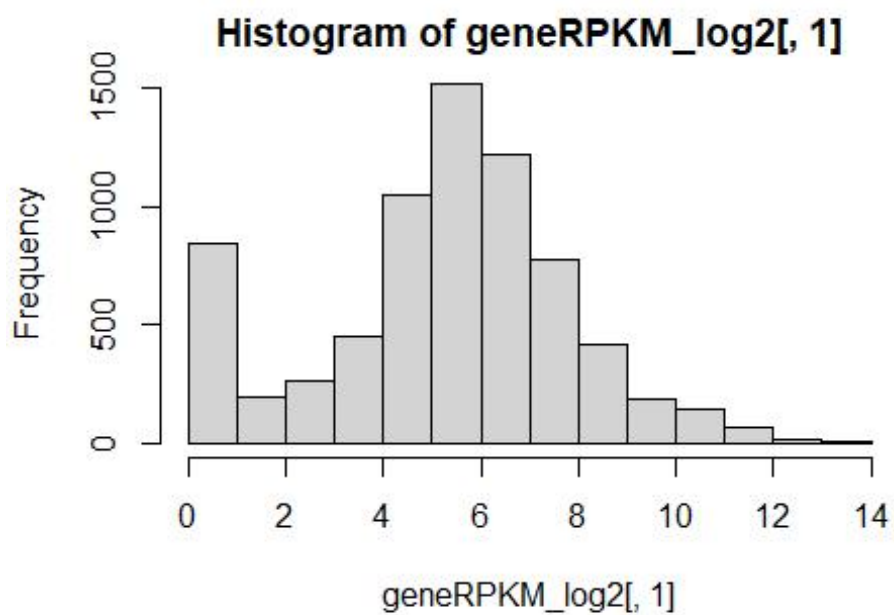
| ## | AMB+LF | AMB+LF | AMB+LF | control 1 | control 2 |
|--------------|--------|--------|--------|-----------|-----------|
| ## ETS1-1 | 59.4 | 59.5 | 35.5 | 61.0 | 96.2 |
| ## ETS1-2 | 60.3 | 53.8 | 36.0 | 51.8 | 104.8 |
| ## ETS2-1 | 61.8 | 32.4 | 27.4 | 15.6 | 20.0 |
| ## ETS2-2 | 53.4 | 33.9 | 25.2 | 21.9 | 19.3 |
| ## HRA1 | 0.2 | 0.3 | 0.3 | 0.3 | 0.0 |
| ## ICR1 | 4.4 | 4.7 | 3.8 | 6.2 | 4.9 |
| ## IRT1 | 13.1 | 11.0 | 9.9 | 12.9 | 11.7 |
| ## ITS1-1 | 7.9 | 11.0 | 7.8 | 6.2 | 7.9 |
| ## ITS1-2 | 15.4 | 22.9 | 13.4 | 14.9 | 20.9 |
| ## ITS2-1 | 33.9 | 47.3 | 31.7 | 40.6 | 40.2 |
| ## control 3 | | | | | |
| ## ETS1-1 | 66.4 | | | | |
| ## ETS1-2 | 74.5 | | | | |
| ## ETS2-1 | 10.3 | | | | |
| ## ETS2-2 | 11.5 | | | | |
| ## HRA1 | 0.2 | | | | |
| ## ICR1 | 7.0 | | | | |
| ## IRT1 | 13.1 | | | | |
| ## ITS1-1 | 11.4 | | | | |
| ## ITS1-2 | 16.0 | | | | |
| ## ITS2-1 | 35.3 | | | | |

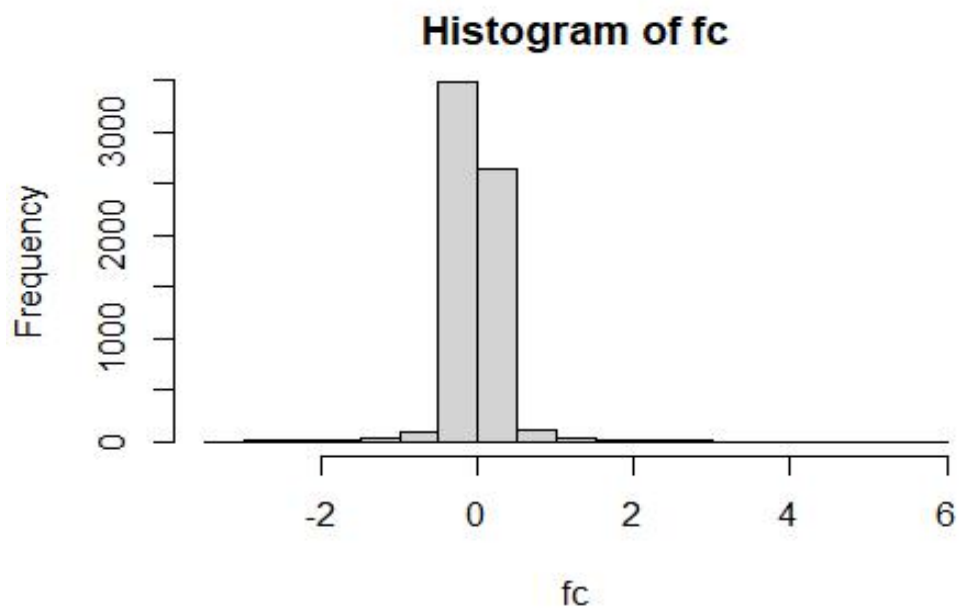
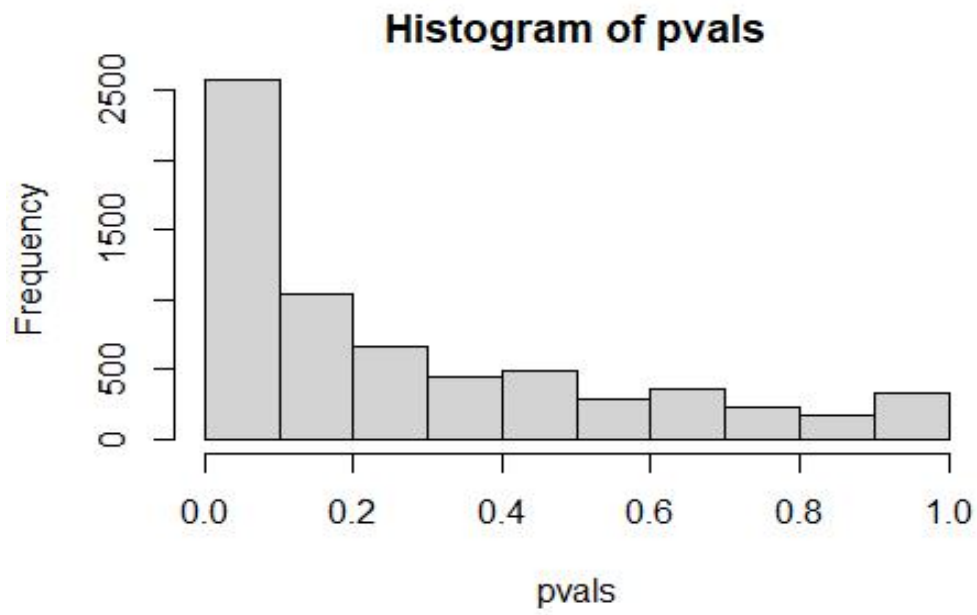
Including Plots

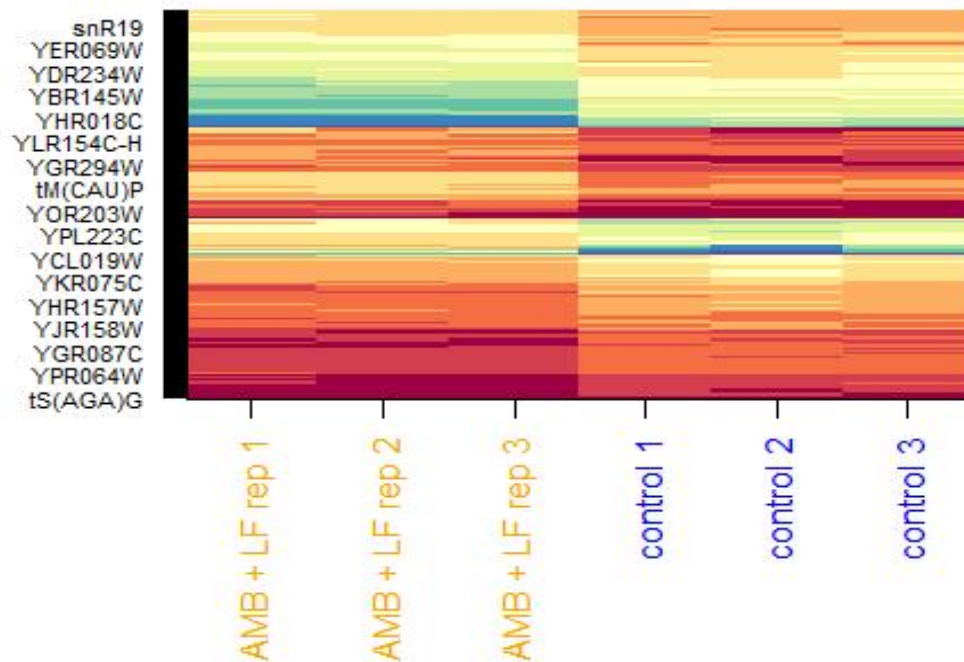
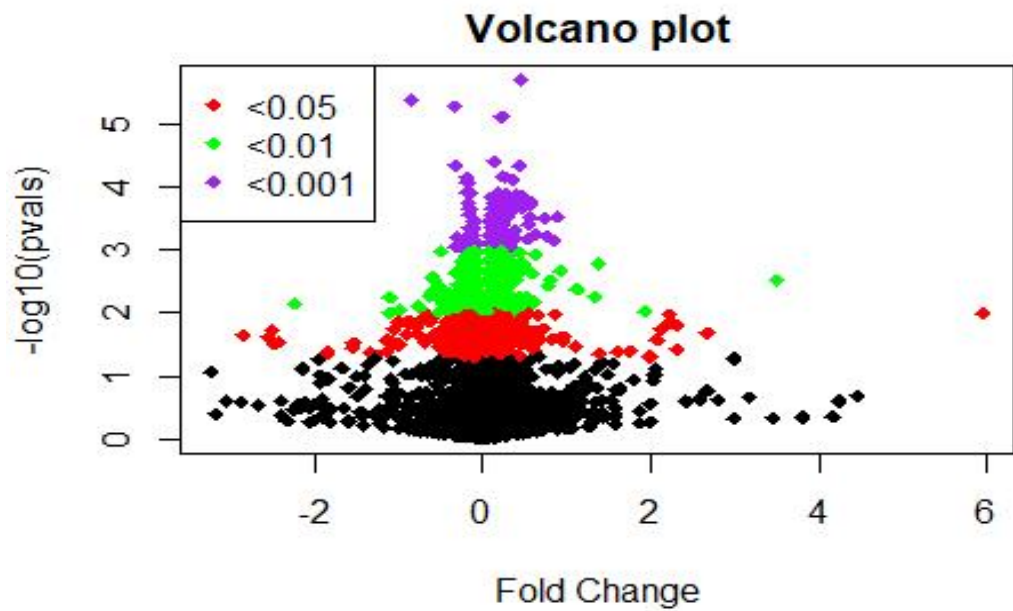












| ## | Control | Control(SD) | AMB + LF | AMB + LF(SD) | FC | P |
|-----------|---------|-------------|----------|--------------|------|----------|
| ## ETS1-1 | 6.1 | 0.4 | 5.7 | 0.4 | -0.1 | 0.291413 |
| ## ETS1-2 | 6.2 | 0.6 | 5.7 | 0.3 | -0.1 | 0.261761 |
| ## ETS2-1 | 3.9 | 0.5 | 5.3 | 0.6 | 0.5 | 0.029178 |

| | | | | | | | |
|----|--------------|-----------|---------|----------|---------|-------------|----------|
| ## | ETS2-2 | 4.1 | 0.5 | 5.2 | 0.5 | 0.4 | 0.043726 |
| ## | HRA1 | 0.0 | 0.0 | 0.0 | 0.0 | NaN | NaN |
| ## | ICR1 | 2.6 | 0.4 | 2.1 | 0.1 | -0.3 | 0.121094 |
| ## | IRT1 | 3.6 | 0.1 | 3.5 | 0.1 | 0.0 | 0.287864 |
| ## | ITS1-1 | 3.0 | 0.4 | 3.1 | 0.4 | 0.1 | 0.701983 |
| ## | ITS1-2 | 4.1 | 0.3 | 4.1 | 0.3 | 0.0 | 0.906643 |
| ## | ITS2-1 | 5.2 | 0.1 | 5.3 | 0.3 | 0.0 | 0.735499 |
| ## | GeneName | Chr | Start | End | Control | Control(SD) | AMB + LF |
| ## | tS(AGA)G | VII | 561662 | 561743 | 0.8 | 0.2 | 0.0 |
| ## | YMR322C | SN04 XIII | 918366 | 919079 | 1.2 | 0.3 | 0.2 |
| ## | YNL109W | XIV | 418964 | 419509 | 1.2 | 0.3 | 0.0 |
| ## | YIR021W-A | IX | 398514 | 398726 | 1.4 | 0.5 | 0.2 |
| ## | tD(GUC)J1 | X | 204735 | 204806 | 1.4 | 0.5 | 0.0 |
| ## | YNR073C | MAN2 XIV | 774792 | 776300 | 2.0 | 0.4 | 0.4 |
| ## | YFL058W | THI5 VI | 12929 | 13951 | 1.7 | 0.5 | 0.3 |
| ## | YPR007C | REC8 XVI | 569336 | 571378 | 1.7 | 0.1 | 0.3 |
| ## | YDR442W | IV | 1345647 | 1346039 | 1.9 | 0.5 | 0.8 |
| ## | YBR180W | DTR1 II | 589741 | 591459 | 1.5 | 0.1 | 0.9 |
| ## | AMB + LF(SD) | FC | P | | | | |
| ## | tS(AGA)G | 0.0 | -Inf | 0.011015 | | | |
| ## | YMR322C | 0.3 | -2.5 | 0.024172 | | | |
| ## | YNL109W | 0.0 | -Inf | 0.024384 | | | |
| ## | YIR021W-A | 0.3 | -2.8 | 0.022259 | | | |
| ## | tD(GUC)J1 | 0.0 | -Inf | 0.044467 | | | |
| ## | YNR073C | 0.6 | -2.4 | 0.028682 | | | |
| ## | YFL058W | 0.4 | -2.5 | 0.018640 | | | |
| ## | YPR007C | 0.4 | -2.5 | 0.030434 | | | |
| ## | YDR442W | 0.5 | -1.3 | 0.040943 | | | |
| ## | YBR180W | 0.2 | -0.7 | 0.010154 | | | |

Conclusion

This study provides valuable insights into the complex mechanism of synergy between AMB and LF in *S. cerevisiae*. The unexpected involvement of oxidative stress response pathways and the pivotal role of transcription factors Aft1p and Zap1p highlight the multifaceted nature of the synergistic interaction. The implications of our findings extend beyond *S. cerevisiae*, suggesting novel avenues for the development of enhanced antifungal therapies with broader applicability in the fight against invasive fungal infections.