# DNA Storage Toolkit: A Modular End-to-End DNA Data Storage Codec and Simulator



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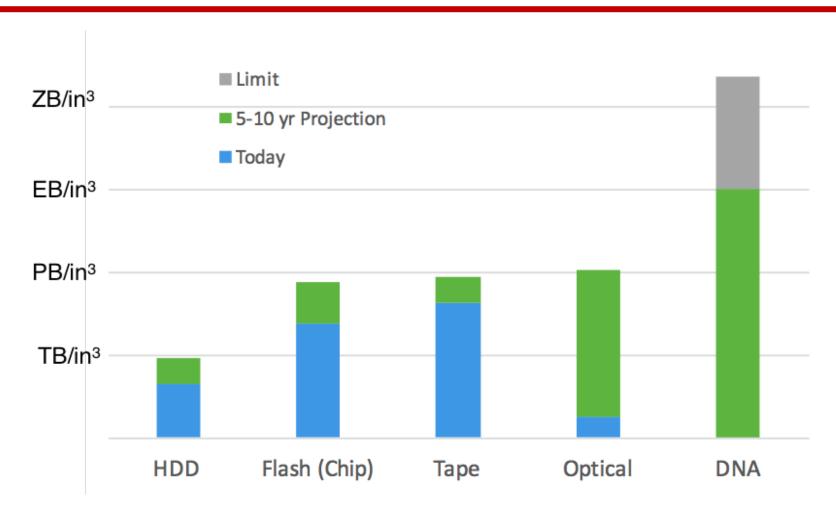
## Why storing data in DNA molecules?

- 1. Incredible density
  - 6-7 orders of magnitude ahead of best alternatives!



- 2. Unmatched durability
  - Thousands/millions/billions of years (vs. 3-5 years for disks/flash)
- 3. Never obsolete: R/W interfaces will only improve with time
- 4. Efficient random access
- 5. Convenient for many data-parallel & near-data computations

## Storage Density Projections\*



<sup>\*</sup>Credit: Luis Ceze & Karin Strauss, Microsoft

## Key Problems with DNA Storage

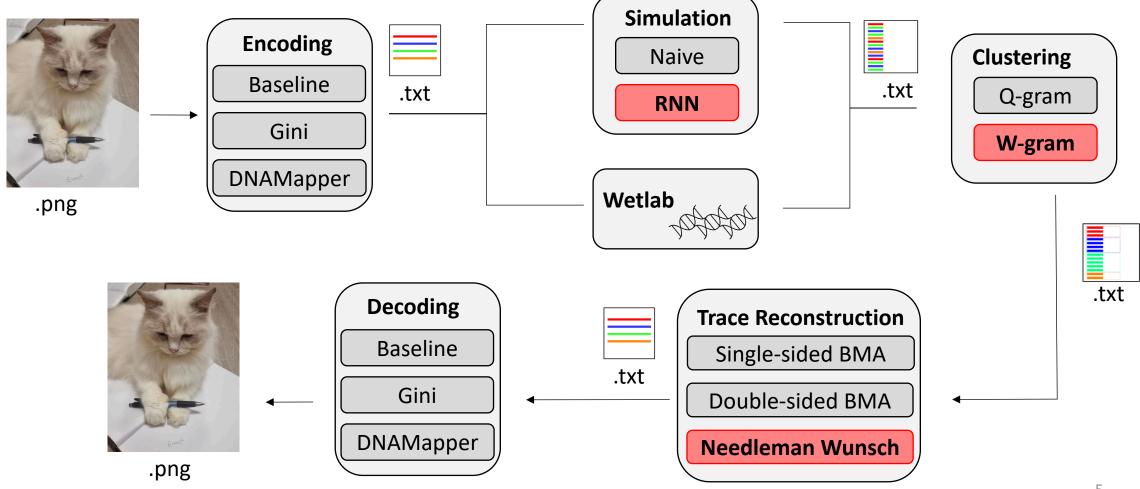
- 1. Expensive R/W interfaces
  - Writing cost: \$1K \$10K/MiB, reading cost: \$10 \$10K/MiB
  - → High cost also makes research very challenging



- Takes days to write DNA, hours (to days) to read (OK for archival storage)
- → But this delay slows down research
- 3. Extremely error-prone interfaces
  - Errors are very peculiar and hard to simulate
- 4. Requires expensive equipment and wetlab expertise
- 5. No complete open-source codec available



## Our DNA Storage Pipeline



## Outline

- Introduction
- DNA Storage Basics
- Our Toolkit
  - Encoding
  - Simulation
  - Clustering
  - Trace Reconstruction
  - Decoding
- Conclusion

#### **DNA Molecules**

#### 4 nucleotides

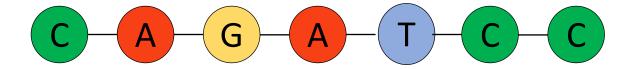








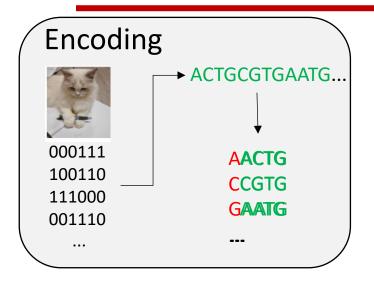
#### Synthetic DNA molecule



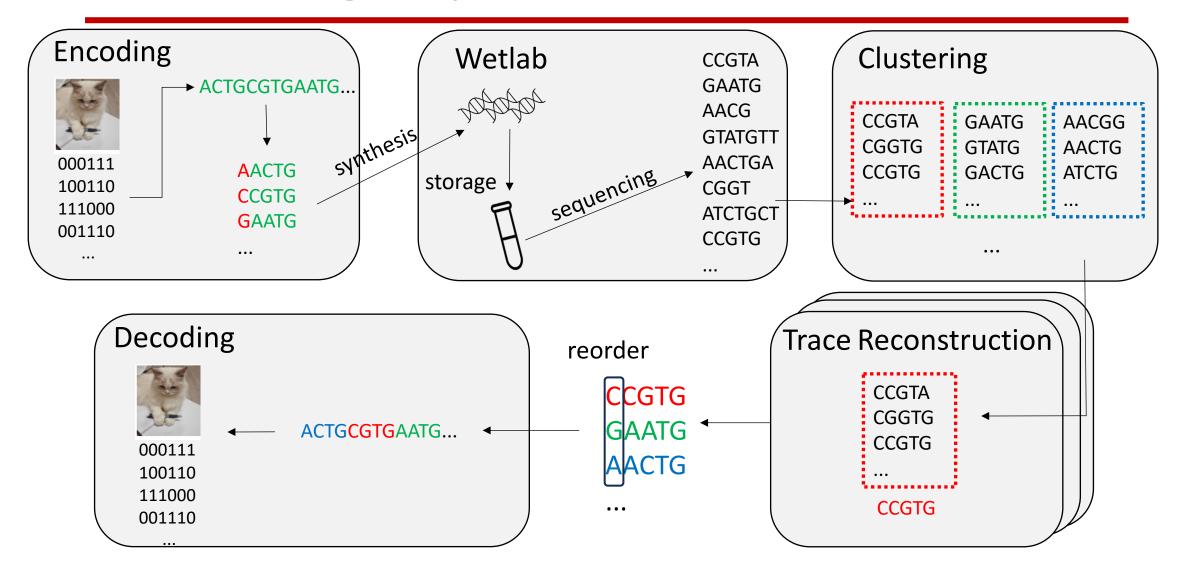
- Artificially created string of nucleotides
- No biological meaning

 $log_2$  | {A, C, G, T}| = 2 bits of data per nucleotide

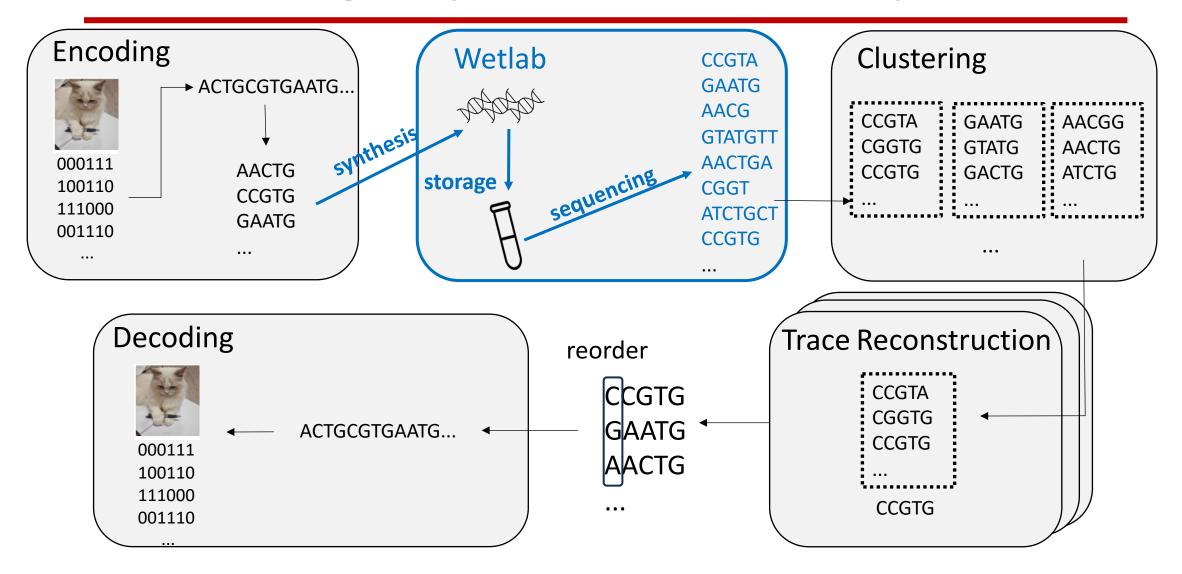
## **DNA Storage Pipeline**



## **DNA Storage Pipeline**



## DNA Storage Pipeline – wetlab steps



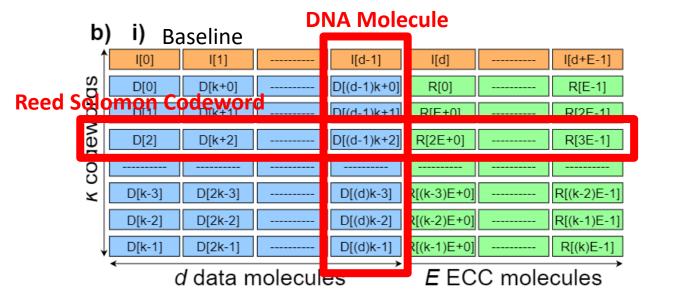
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## Encoding

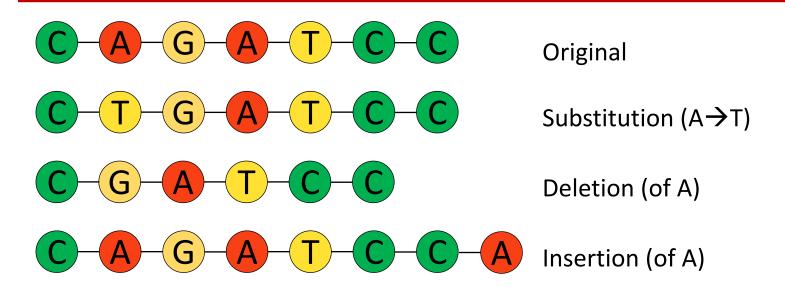
3 encoding schemes provided.

- i) Baseline: Introduced by Organick et al. [1]
- ii) Gini: Introduced by Lin et al. [2]
- iii) DNAMapper: Introduced by Lin et al. [2]

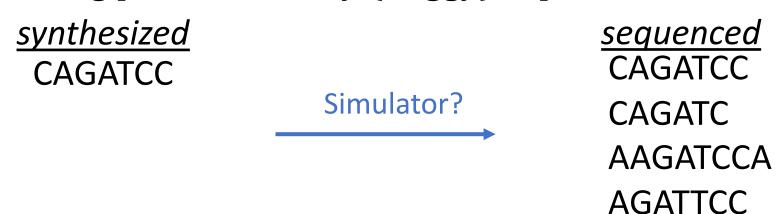


ii) Gini										
I[0]	I[1]		I[d-1]	l[d]		I[d+E-1]				
D[0]	D[k+0]		D[(d-1)k+0]	R[0]		R[E-1]				
D[1]	D[k+1]		D[(d-1)k+1]	R[E+0]		R[2E-1]				
D[2]	D[k+2]		D[(d-1)k+2]	R[2E+0]		R[3E-1]				
D[k-3]	D[2k-3]		D[(d)k-3]	R[(k-3)E+0]		R[(k-2)E-1]				
D[k-2]	D[2k-2]		D[(d)k-2]	R[(k-2)E+0]		R[(k-1)E-1]				
D[k-1]	D[2k-1]		D[(d)k-1]	R[(k-1)E+0]		R[(k)E-1]				
iii) DNA Mapper										
I[0]	I[1]		I[d-1]	l[d]		I[d+E-1]				
P[d+0]	P[d+1]		P[2d-1]	R[0]		R[E-1]				
P[3d+0]	P[3d+1]		P[4d-1]	R[E+0]		R[2E-1]				
P[5d+0]	P[5d+1]		P[6d-1]	R[2E+0]		R[3E-1]				
P[4d+0]	P[4d+1]		P[5d-1]	R[(k-3)E+0]		R[(k-2)E-1]				
P[2d+0]	P[2d+1]		P[3d-1]	R[(k-2)E+0]		R[(k-1)E-1]				
P[0]	P[1]		P[d-1]	R[(k-1)E+0]		R[(k)E-1]				

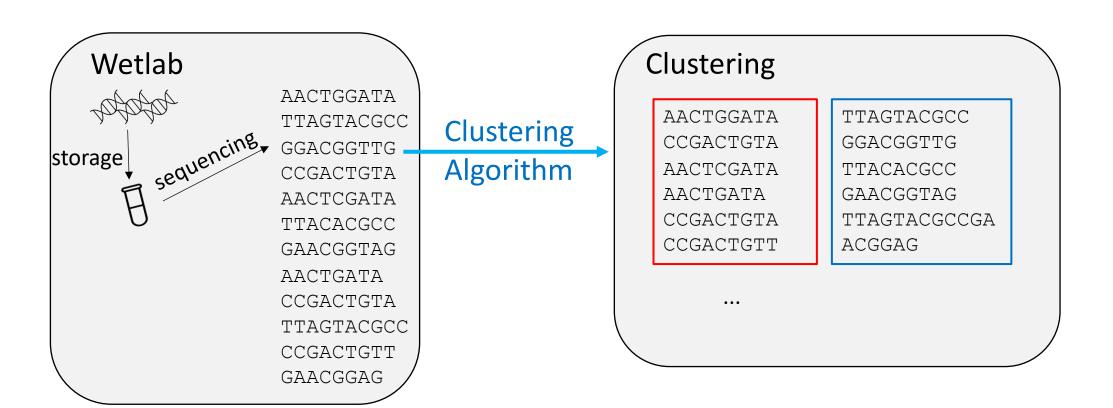
#### Wetlab



Sequencing produces many (buggy) copies of each molecule:



## Clustering



## Clustering

- Using edit distance for clustering is too slow.
- We can approximate using q-gram binary signatures and Hamming distance.

	CT	GA	GT	
AC <b>GT</b> C <b>GA</b> AC	0	1	1	
AC <b>GT</b> CC <b>GA</b> AC	0	1	1	Low Hamming distance
AC <b>GT</b> C <b>GA</b> AC	0	1	1	
TACCTATTCC	1	0	0	High Hamming distance

## Autotuning the threshold parameters

The threshold parameters for the binary signature Hamming distance need to be tuned based on dataset.

Take a tiny sample and plot the Hamming distance for them:



**Hamming Distance** 

GTACCAGTCGAGTAAAGC
GCCGTGCGTAAGCT
GTACAATGTCGTGTAAC
GTCATGGTCAGTAAGC
GTACAGTCCGTAAAGC
TACGTGTATAGC
GATACAGCACGTGAAGC

What is the consensus?

**G**TACCAGTCGAGTAAAGC

**G**CCGTGCGTAAGCT

**G**TACAATGTCGTGTAAC

**G**TCATGGTCAGTAAGC

**G**TACAGTCCGTAAAGC

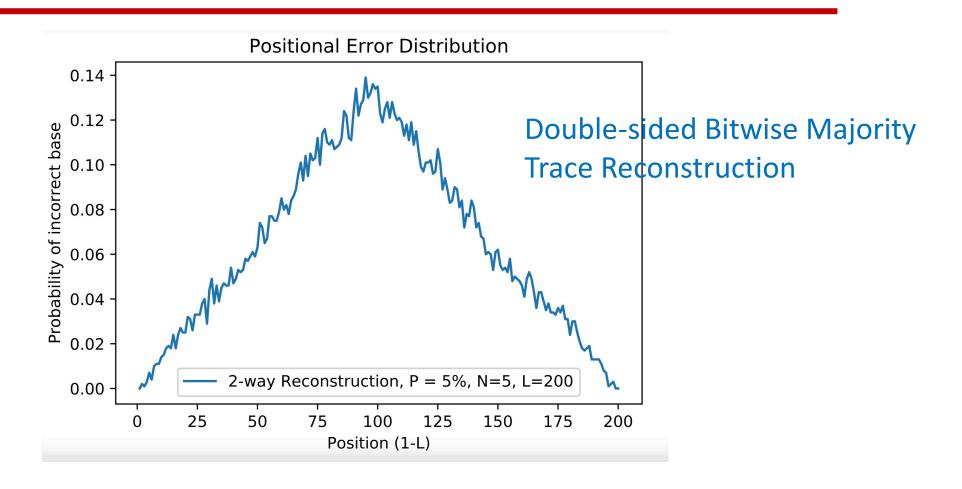
TACGTGTATAGC

**G**ATACAGCACGTGAAGC

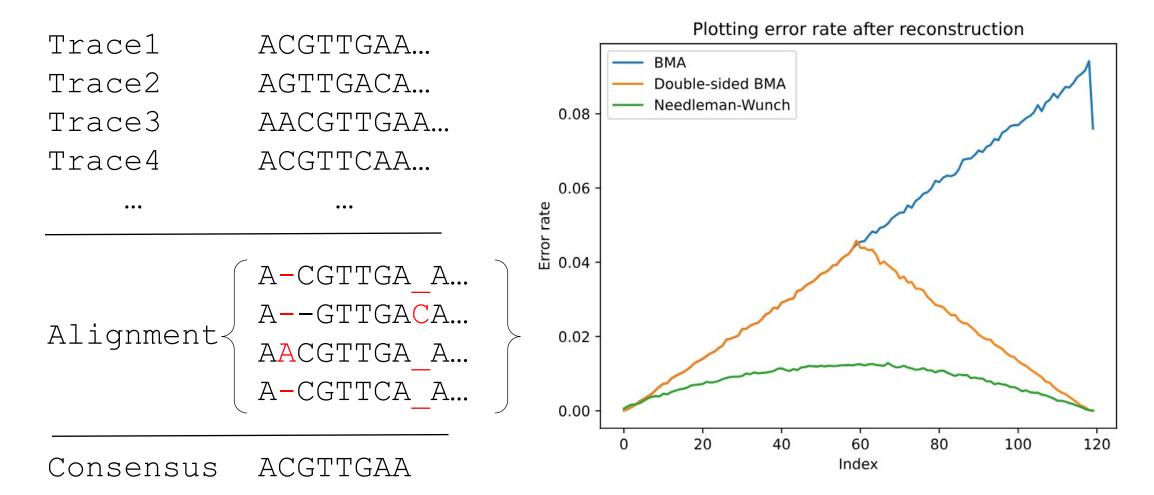
G

- GTACCAGTCGAGTAAAGC
- GCCGTGCGTAAGCT
- GTACAATGTCGTGTAAC
- GTCATGGTCAGTAAGC
- GTACAGTCCGTAAAGC
- -**T**ACGTGTATAGC
- GATACAGCACGTGAAGC

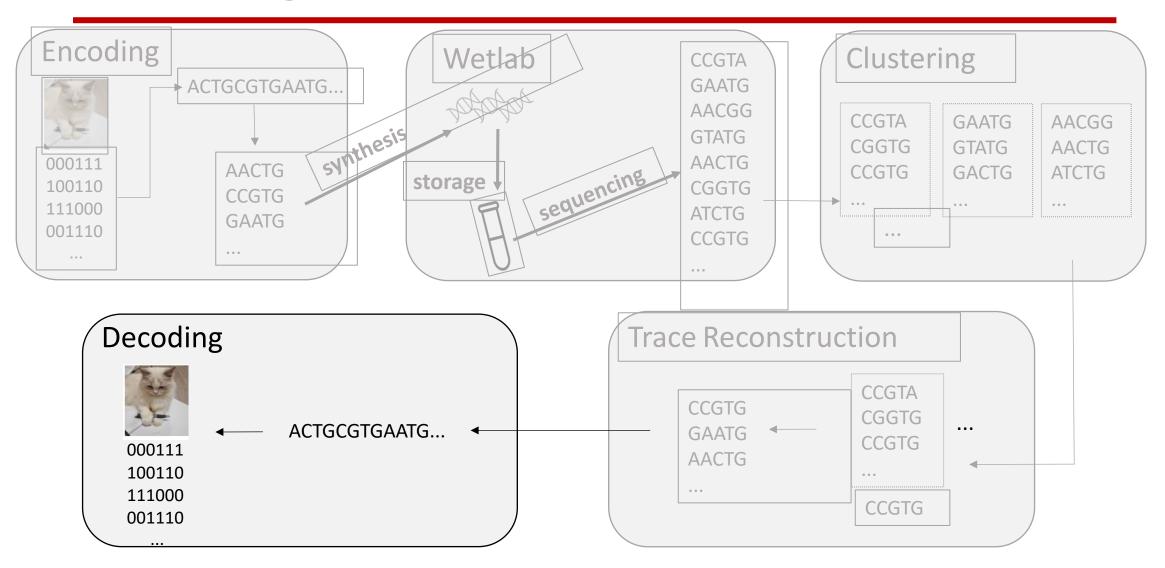
GT



## Needleman-Wunsch



## Decoding



#### Wetlab Simulation

Sequencing produces many (buggy) copies of each molecule:

synthesized CAGATCC

Simulator?

sequenced CAGATCC

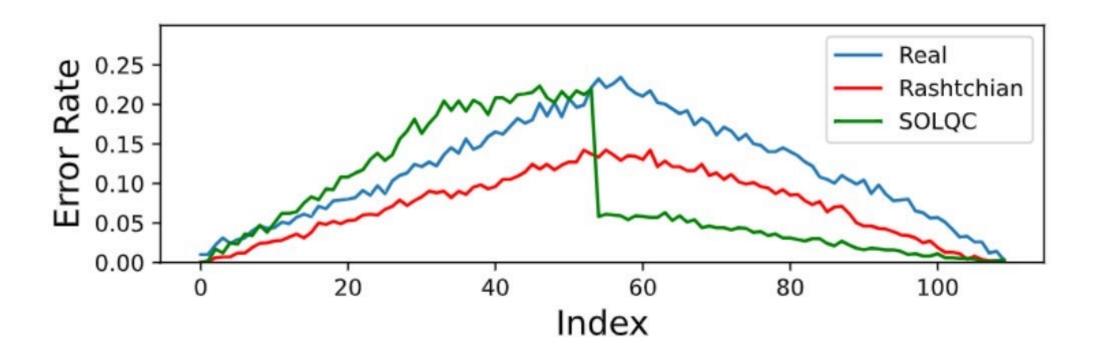
CAGATC

**AAGATCCA** 

**AGATTCC** 

#### Wetlab Simulation

We evaluate the simulation by comparing how the Trace Reconstruction module performs on the simulated data.



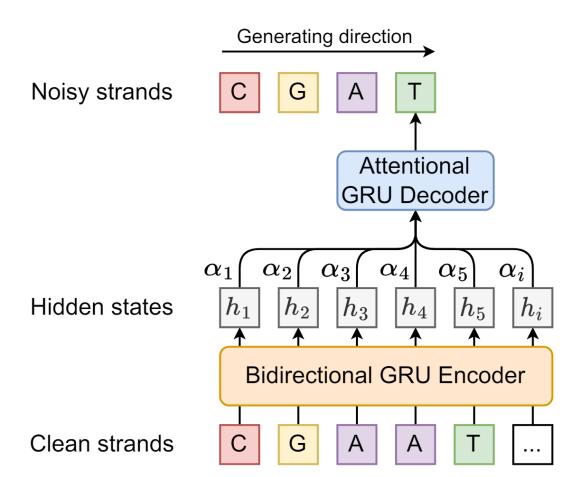
#### **Our Simulation**

Sequence-to-sequence problem.

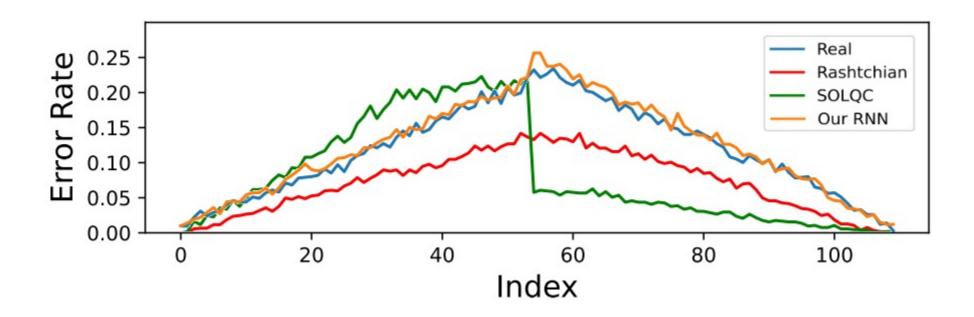
Model based on basic NMT model [3].

Attention based encoder-decoder.

[Hidden layer size = 128, Greedy sampling.]



## **Our Simulation**

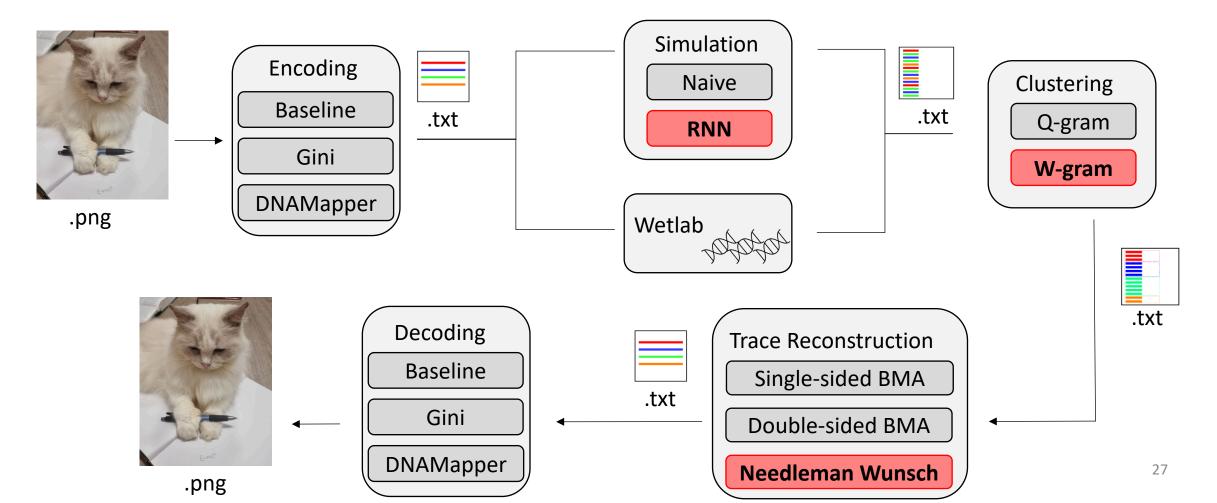


The performance of the reconstruction module on our simulated data is very similar to real data.

We can evaluate later modules of this pipeline using our simulated data!

## **Toolkit**

https://github.com/prongs1996/DNAStorageToolkit/



#### Conclusion

- First open end-to-end DNA storage toolkit
- Very accurate simulator for wetlab steps



Repository: https://github.com/prongs1996/DNAStorageToolkit/

## Thank you! Questions?



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