

# **A space travel into the Multiverse**

## **Cognitive Science Arena**

Giulia Calignano    Filippo Gambarota

Department of Developmental Psychology and Socialization, University of Padova

2025-05-02

# **Exploratory Multiverse Analysis (EMA)**

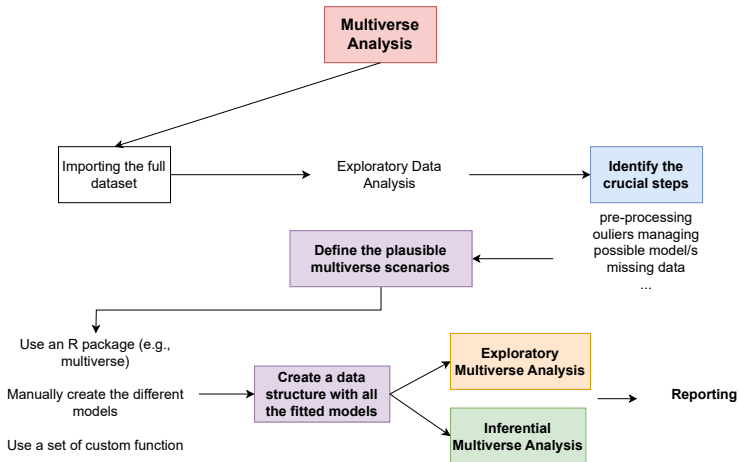
# An example, Statistics and Math Anxiety

McCaughey et al. (2022) explored the relationship between self-efficacy anxiety sensitivity and perfectionism would be related to math/statistics anxiety controlling for gender, university program, and education level.

We used the dataset available at <https://osf.io/nzhq6>.

**We are going to do crazy stuff with this dataset that are not related to the original paper and research question! :)**

# The big picture



# Importing

We did a little bit of pre-processing. The `ms_anxiety.rds` file contains the cleaned version of the original dataset.

```
dat <- readRDS(here("data/ms_anxiety.rds"))
vars <- names(dat)
ys <- vars[grepl("^stat.anx|^math", vars)]
ys
## [1] "stat.anx.tc"      "stat.anx.i"       "stat.anx.ah"      "stat.anx.ws"
## [5] "stat.anx.fst"     "stat.anx.sc"      "math.anx"         "stat.anx.TOT"
## [9] "stat.anx.ANX"     "stat.anx.FEEL"

xs <- vars[!vars %in% ys]
xs
## [1] "self.efficacy"    "asi"              "frost.com"
## [4] "frost.da"         "faculty"          "program.type"
## [7] "gender.category"
```

# Exploring

Let's see the type of variables of the dataset:

```
sapply(dat[ys], class)
```

```
##  stat.anx.tc    stat.anx.i    stat.anx.ah    stat.anx.us    stat.anx.fst  
##    "numeric"    "numeric"    "numeric"    "numeric"    "numeric"  
##  stat.anx.sc    math.anx    stat.anx.TOT    stat.anx.ANX    stat.anx.FEEL  
##    "numeric"    "numeric"    "numeric"    "numeric"    "numeric"
```

```
sapply(dat[xs], class)
```

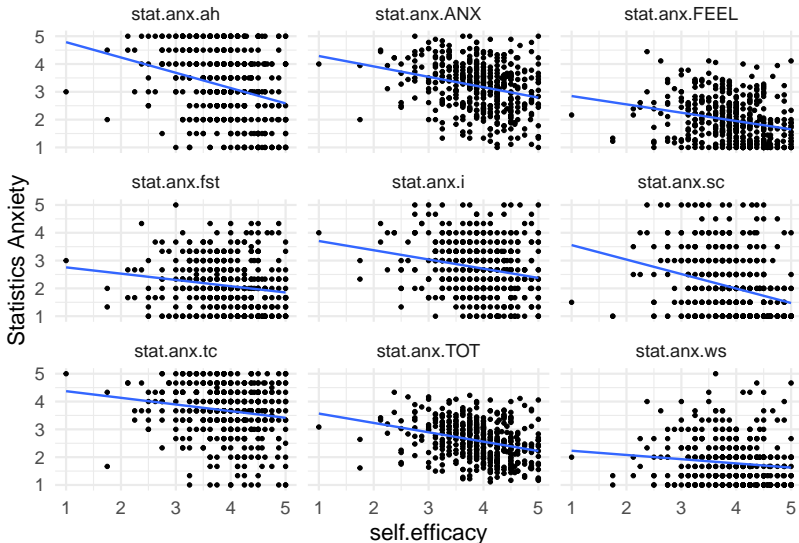
```
##  self.efficacy          asi          frost.com          frost.da  
##    "numeric"          "numeric"          "numeric"          "numeric"  
##    faculty    program.type    gender.category  
##    "factor"    "factor"          "factor"
```

# Main research questions

The main idea of the authors is predicting **math** and **statistics** anxiety with self-efficacy and perfectionism. In particular they pre-registered (see <https://osf.io/b3g7s>):

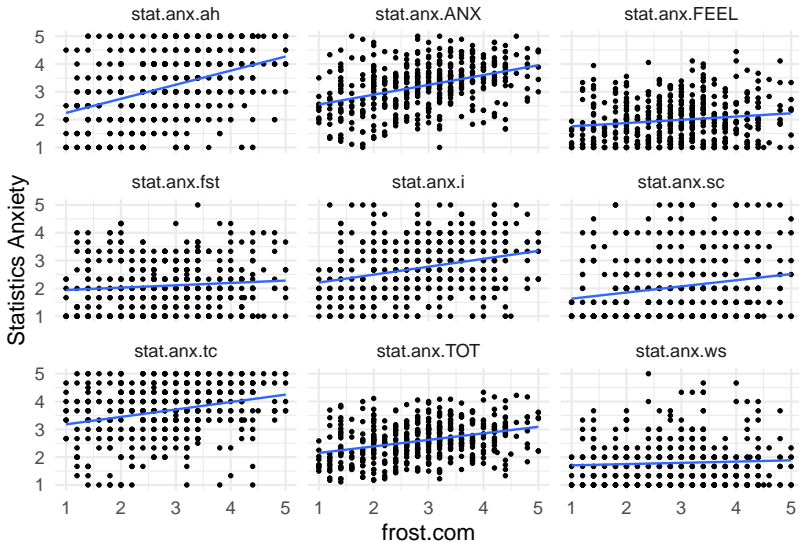
1. self-efficacy will be negatively related to math/statistics anxiety
2. anxiety sensitivity will be positively related to math/statistics anxiety.
3. self-critical perfectionism will be positively related to math/statistics anxiety.
4. the relationships described above will remain when statistically adjusting for gender, university program (arts vs. science) and student status (undergraduate vs. graduate).

# Exploring the relationships

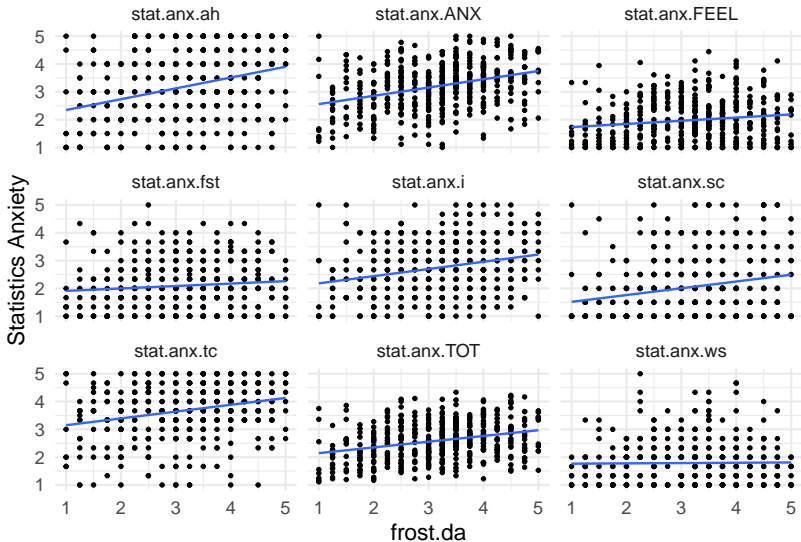




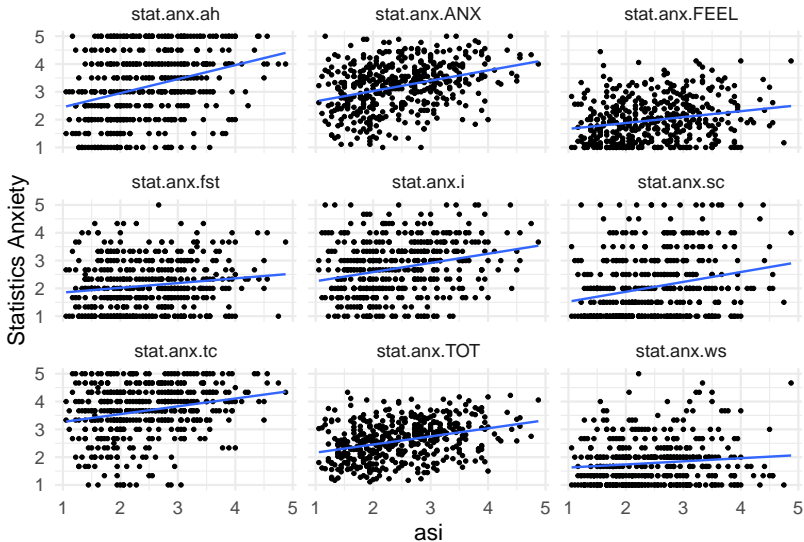
# Exploring the relationships



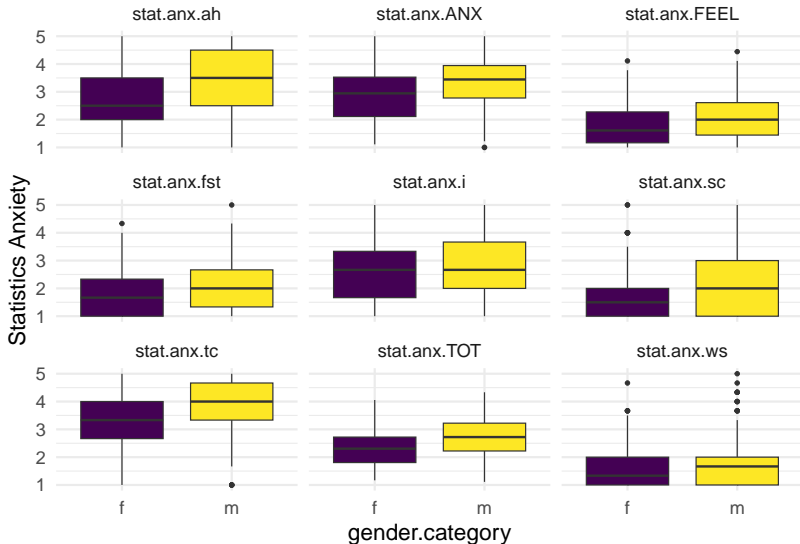
# Exploring the relationships



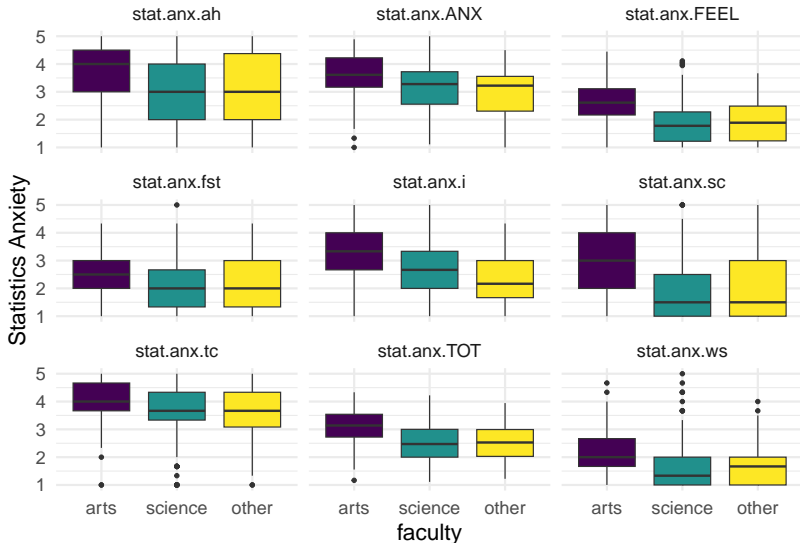
# Exploring the relationships



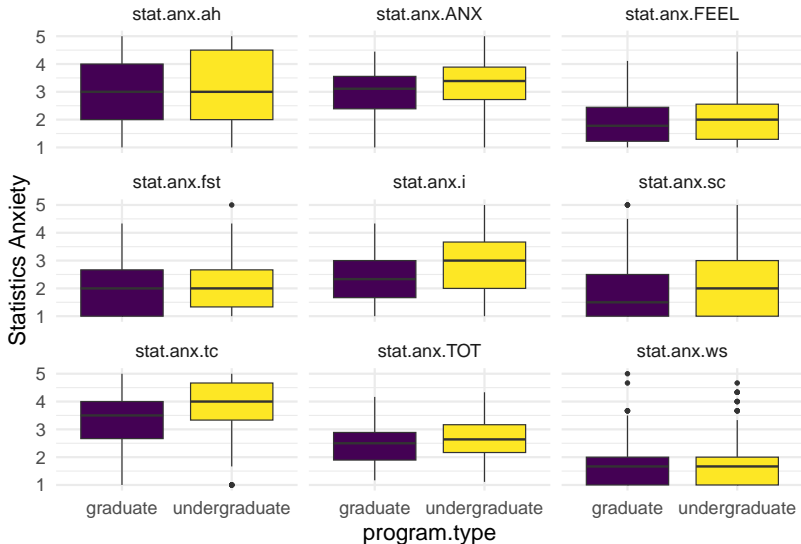
# Exploring the relationships



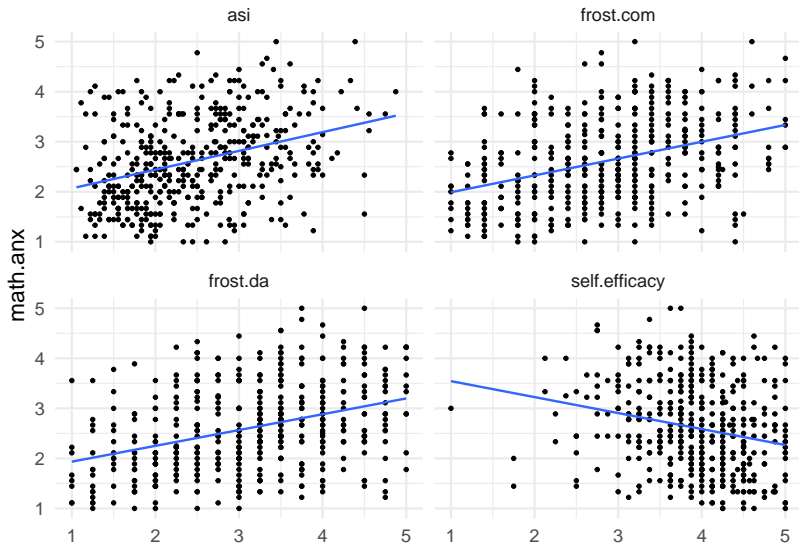
# Exploring the relationships



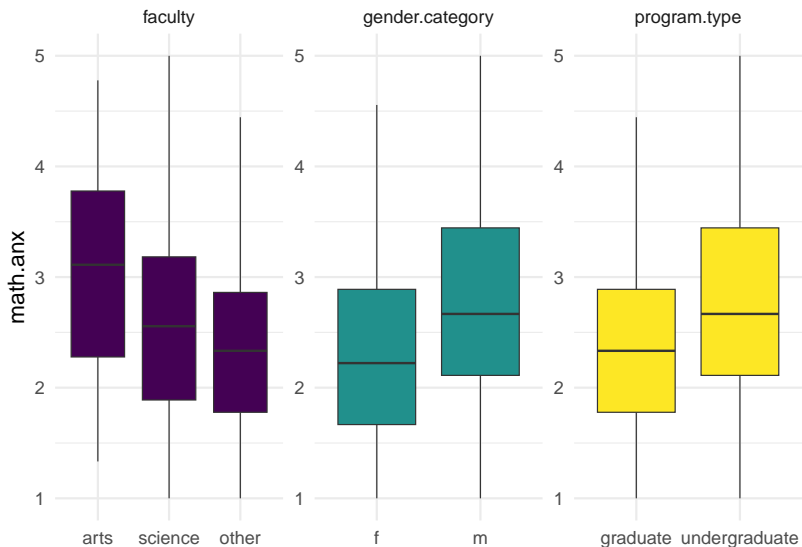
# Exploring the relationships



# Exploring the relationships



# Exploring the relationships





# Selecting a sub-sample

For the purpose of the example, we select a subsample of the dataset to increase the variability and simulate a more uncertain scenario with a lower sample size.

```
set.seed(9386)
N <- 200
selected <- sample(1:nrow(dat), size = N, replace = FALSE)
dat <- dat[selected, ]
```

# Data structure for specifications

When conducting a multiverse in R (or in whatever language) the data structure is very important.

- ▶ how to create and organize the different models?
- ▶ how to easily extract all the informations such as coefficients, standard errors, p-values, etc.
- ▶ ...

# An R list is probably the best

A (named) `list` is flexible, easy to index and can be accessed by other functions to extract information and create other list.

A `list` in R can be easily transformed into a `data.frame` for other models, plots, tables, etc.

You can use the `*apply` family (`sapply`, `lapply`, etc.) to compute complex operations on lists.

# An R list is probably the best

For example, assuming that I have some regression models within a named list:

```
fit1 <- lm(math.anx ~ gender.category + asi, data = dat)
fit2 <- lm(math.anx ~ gender.category + asi + faculty, data = dat)
fit3 <- lm(math.anx ~ gender.category + faculty, data = dat)
fit4 <- lm(math.anx ~ gender.category + asi + program.type, data = dat)

# ... and other thousands of (plausible) models :)

mods <- list(fit1, fit2, fit3, fit4)
names(mods) <- paste0("mod", 1:length(mods))
```

# An R list is probably the best

Then, I want to extract all the asi coefficients and put into a data.frame:

```
get_coef <- function(x, coef = NULL){  
  x <- broom::tidy(x, conf.int = TRUE)  
  if(!is.null(coef)){  
    filter(x, term %in% coef)  
  } else{  
    x  
  }  
}
```

```
get_coef(mods$mod1, "asi")
```

```
# A tibble: 1 x 7
```

	term	estimate	std.error	statistic	p.value	conf.low	conf.high
	<chr>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>
1	asi	0.383	0.0748	5.11	0.000000745	0.235	0.530

# An R list is probably the best

With `lapply` (or `purrr::map()`) and combining the results, you can easily create a nice dataframe with your coefficients:

```
lapply(mods, get_coef, "asi") |>  
  dplyr::bind_rows(.id = "mod")
```

```
# A tibble: 3 x 8  
  mod   term estimate std.error statistic  p.value conf.low conf.high  
  <chr> <chr>   <dbl>    <dbl>    <dbl>   <dbl>   <dbl>   <dbl>  
1 mod1  asi     0.383    0.0748     5.11 7.45e-7  0.235   0.530  
2 mod2  asi     0.386    0.0741     5.21 4.88e-7  0.240   0.532  
3 mod4  asi     0.410    0.0728     5.64 5.89e-8  0.267   0.554
```

# Creating the specifications

There are multiple ways of creating the specifications in practice.  
You can do it from scratch:

```
mod1 <- lm(y ~ x1 + x2)
mod2 <- lm(y ~ log(x1) + log(x2))
mod3 <- lm(y ~ x1 + x2) # removing outliers

mods <- list(mod1 = mod1, mod2 = mod2, mod3 = mod3)
# ...
```

# Creating the specifications

The **multiverse** R Package and the related paper (Götz et al., 2024) provides a very flexible and complex syntax to define different specifications.



# Creating the specifications

For this example we can use some custom functions, in particular the `create_multi()` function. There are no wrong solutions if the results is correct.

```
devtools::load_all()

slog <- function(x) {
  if(any(x == 0)){
    x <- x + 1
  }
  log(x)
}

multi <- create_multi(
  math.anx ~ asi + faculty + stat.anx.TOT, # full model formula
  focal = "asi", # focal predictor, never removed
  nfun = c("slog"), # functions for the numeric variables
  data = dat
)
```

# Creating the specifications

```
$X
      fun          x    type focal .id_fun .id_x
1 identity          asi numeric  TRUE      1      1
2 identity    faculty  factor FALSE      1      2
3 identity stat.anx.TOT numeric FALSE      1      3
5      slog stat.anx.TOT numeric FALSE      2      3

      call
1          asi
2          faculty
3      stat.anx.TOT
5 slog(stat.anx.TOT)

$calls
[1] "~ asi"
[2] "~ asi + faculty"
[3] "~ asi + stat.anx.TOT"
[4] "~ asi + slog(stat.anx.TOT)"
[5] "~ asi + faculty + stat.anx.TOT"
[6] "~ asi + faculty + slog(stat.anx.TOT)"
```

# Creating the specifications

Whatever the method we used, we need:

- ▶ a list of models
- ▶ a way to easily extract the coefficients or other quantities
- ▶ a way to extract a summary of the specifications i.e. if a variable is included or not, the type of transformation, etc.

# Pay attention with interactions!

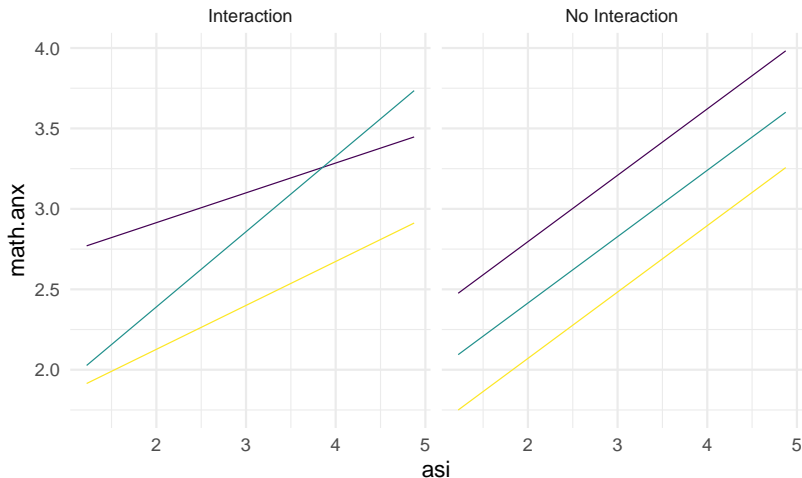
When an interaction is included in the model, the interpretation of the model coefficients completely change, especially if the interaction is consistent. You cannot compare a focal coefficients directly for models with and without interactions.

Let's assume that `asi` is the focal coefficient and we include in the multiverse these two models:

```
fit_int <- lm(math.anx ~ faculty + asi + faculty:asi, data = dat)
fit_no_int <- lm(math.anx ~ faculty + asi, data = dat)
```

# Pay attention with interactions!

The `asi` effect in one case is the overall effect (i.e., main effect) controlling for `faculty`. In the other case is the `asi` effect of the reference value.



# Pay attention with interactions!

One should adjust the contrasts coding of factors and/or the centering of numeric variables.

```
# sum to zero contrasts i.e. estimating the main effect of asi
update(fit_int, contrasts = list(faculty = contr.sum(3)))
```

Call:

```
lm(formula = math.anx ~ faculty + asi + faculty:asi, data = dat,
    contrasts = list(faculty = contr.sum(3)))
```

Coefficients:

(Intercept)	faculty1	faculty2	asi	faculty1:asi
1.8597	0.6839	-0.4048	0.3087	-0.1234

faculty2:asi  
0.1590

```
# with emmeans
```

```
emmeans::emtrends(fit_int, ~1, var = "asi")
```

1	asi.trend	SE	df	lower.CL	upper.CL
overall	0.309	0.11	194	0.0925	0.525

Results are averaged over the levels of: faculty  
Confidence level used: 0.95

# Why exploring is important?

A multiverse analysis increase the complexity of the data analysis.  
**There is no longer a single dataset and result to discuss.**

# Let's create some scenarios :)

Firstly, we use variable transformations directly within the model formula. In this way it is easier to extract the conditions. Thus we define some wrappers:

```
# safe version of log() with 0 variables
slog <- function(x){
  if(any(x == 0)){
    x <- x + 1
  }
  log(x)
}

# function factories, see https://adv-r.hadley.nz/function-factories.html
polyN <- function(degree = 1){
  function(x) poly(x, degree = degree)
}

poly2 <- polyN(2)
poly3 <- polyN(3)
```



# Let's create some scenarios :)

More wrappers:

```
cutN <- function(breaks){  
  function(x){  
    cut(x, breaks = breaks)  
  }  
}
```

```
cut2 <- cutN(2)
```

```
cut4 <- cutN(4)
```

# Let's create some scenarios :)

Then we can identify some univariate/multivariate outliers or some observations that we may consider removing for some reasons.

# Let's create some scenarios :)

```
focal <- "self.efficacy"

multi <- create_multi(
  math.anx ~ self.efficacy + faculty + asi + gender.category +
    program.type + frost.da,
  focal = focal,
  nfun = c("slog", "cut2", "poly2"),
  data = dat
)
```

# Let's fit the models

```
# faster than before
get_coef <- function(x, coef = NULL){
  xs <- data.frame(summary(x)$coefficients)
  if(!is.null(coef)){
    xs <- xs[coef, ]
  }
  xs$param <- rownames(xs)
  return(xs)
}

fitl <- vector(mode = "list", length = length(multi$calls))

for(i in 1:length(multi$calls)){
  form <- paste0("math.anx", multi$calls[i])
  fitl[[i]] <- glm(form, family = gaussian(link = "identity"), data = dat)
}

resl <- lapply(fitl, get_coef, focal)
res <- bind_rows(resl, .id = "mod")
rownames(res) <- NULL
names(res) <- c("mod", "b", "se", "t", "p", "param")
```

# Let's fit the models

Now we have a dataframe with all the model coefficients across the specifications. We can start our multiverse!

```
head(res)
```

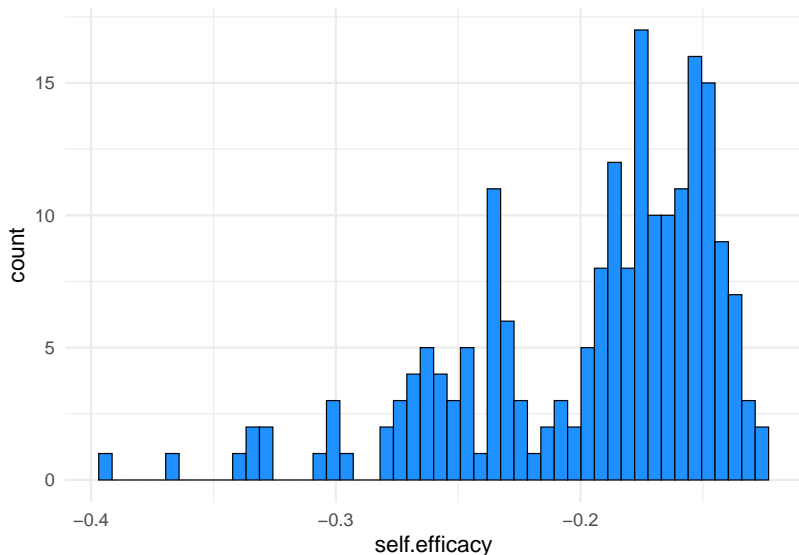
	mod	b	se	t	p	param
1	1	-0.3955213	0.10133460	-3.903122	0.0001301284	self.efficacy
2	2	-0.3315150	0.10439789	-3.175495	0.0017375474	self.efficacy
3	3	-0.3302943	0.09599808	-3.440634	0.0007084586	self.efficacy
4	4	-0.3041665	0.10324396	-2.946095	0.0036060375	self.efficacy
5	5	-0.3335836	0.09874191	-3.378339	0.0008788681	self.efficacy
6	6	-0.2266493	0.10148550	-2.233317	0.0266529679	self.efficacy

# Exploratory tools

- ▶ Marginal/Conditional effects
- ▶ Variation of Effects
- ▶ Specification Curve

# Marginal/Conditional effects

Overall distribution of regression parameters:



# Marginal/Conditional effects

We can combine the model results with a table created by all conditions with the custom `get_info_models()` function:

```
info <- get_info_models(multi)
head(info)
```

```
# A tibble: 6 x 7
  mod x_self.efficacy x_faculty x_asia x_gender.category
<int> <chr>           <chr>    <chr> <chr>
1     1 self.efficacy <NA>    <NA> <NA>
2     2 self.efficacy faculty <NA> <NA>
3     3 self.efficacy <NA>    asi <NA>
4     4 self.efficacy <NA>    <NA> gender.category
5     5 self.efficacy <NA>    <NA> <NA>
6     6 self.efficacy <NA>    <NA> <NA>
# i 2 more variables: x_program.type <chr>, x_frost.da <chr>
```



# Marginal/Conditional effects

Then we can combine the info table with the coefficients table and we have all the important information.

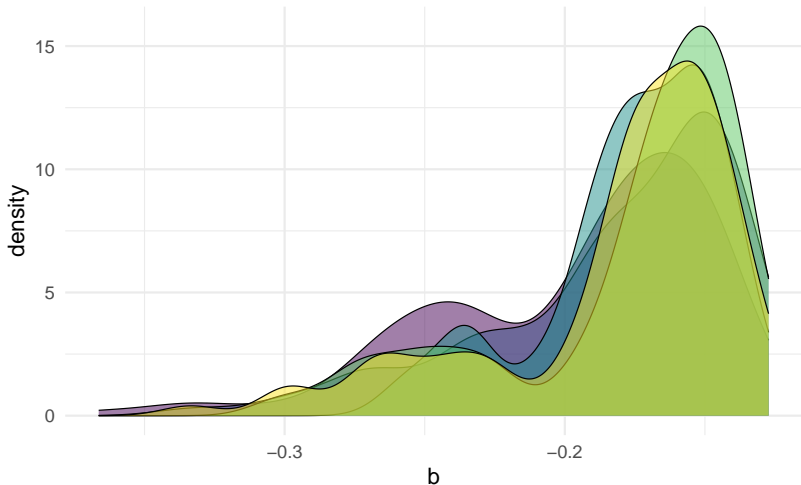
```
# same type
res$mod <- as.numeric(res$mod)
info$mod <- as.numeric(info$mod)
# merging the two tables
multi_res <- left_join(res, info, by = "mod")
head(multi_res)
```

	mod	b	se	t	p	param
1	1	-0.3955213	0.10133460	-3.903122	0.0001301284	self.efficacy
2	2	-0.3315150	0.10439789	-3.175495	0.0017375474	self.efficacy
3	3	-0.3302943	0.09599808	-3.440634	0.0007084586	self.efficacy
4	4	-0.3041665	0.10324396	-2.946095	0.0036060375	self.efficacy
5	5	-0.3335836	0.09874191	-3.378339	0.0008788681	self.efficacy
6	6	-0.2266493	0.10148550	-2.233317	0.0266529679	self.efficacy
	x_self.efficacy	x_faculty	x_asl	x_gender.category	x_program.type	
1	self.efficacy	<NA>	<NA>	<NA>	<NA>	
2	self.efficacy	faculty	<NA>	<NA>	<NA>	
3	self.efficacy	<NA>	asl	<NA>	<NA>	
4	self.efficacy	<NA>	<NA>	gender.category	<NA>	
5	self.efficacy	<NA>	<NA>	<NA>	program.type	
6	self.efficacy	<NA>	<NA>	<NA>	<NA>	

x\_frost.da

# Marginal/Conditional effects

Finally we can plot also the distributions of parameters conditioned on the presence/absence of a particular other predictor:



# Vibration of Effects (VoE) (Patel et al., 2015)



Journal of Clinical Epidemiology 68 (2015) 1046–1058

**Journal of  
Clinical  
Epidemiology**

## Assessment of vibration of effects due to model specification can demonstrate the instability of observational associations

Chirag J. Patel<sup>a</sup>, Belinda Burford<sup>b</sup>, John P.A. Ioannidis<sup>a,c,d,e,f,\*</sup>

<sup>a</sup>Department of Biomedical Informatics, Harvard Medical School, 10 Shattuck St., Room 314A, Boston, MA 02115, USA

<sup>b</sup>Melbourne School of Population and Global Health, Level 4, 207 Bouverie St., The University of Melbourne, Victoria 3010, Australia

<sup>c</sup>Department of Medicine, Stanford Prevention Research Center, Stanford University School of Medicine, Medical School Office Building, Room X306, 1265 Welch Rd, Stanford, CA 94305, USA

<sup>d</sup>Department of Statistics, Stanford University School of Humanities and Sciences, Stanford, CA 94305, USA

<sup>e</sup>Department of Health Research and Policy, Stanford University School of Medicine, Stanford, CA 94305, USA

<sup>f</sup>Meta-Research Innovation Center at Stanford (METRICS), Stanford University, Stanford, CA 94305, USA

Accepted 30 May 2015; Published online 6 June 2015

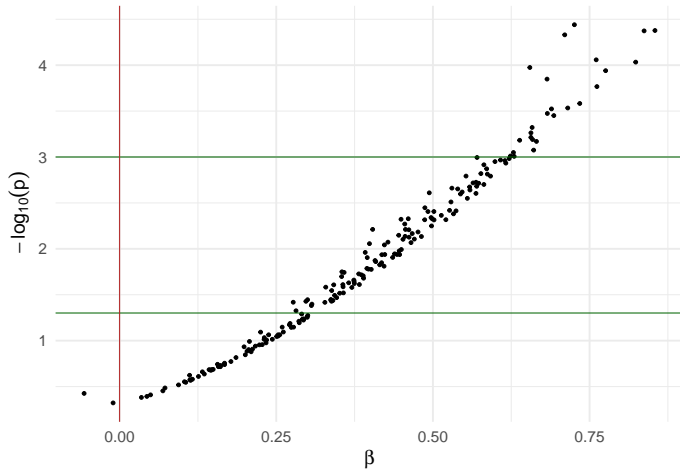
# Vibration of Effects (VoE) (Patel et al., 2015)

The VoE is a statistical approach to evaluate the variability in effect estimates and p value due to different sources of variability (i.e., *vibrations*)

- ▶ **sampling** vibration: subsets of the full dataset
- ▶ **model** vibration: combinations of control variables
- ▶ **pre-processing** vibration: inclusion/exclusion criteria, outliers, etc.

# Vulcano Plot

The Vulcano Plot is the graphical tool used in the VoE as a diagnostic tool.



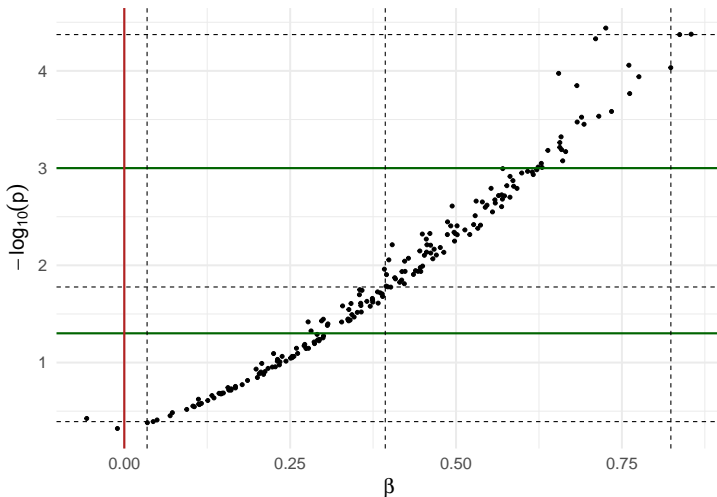
# Vulcano Plot

The x axis is the effect size. Usually a regression coefficient of a *focal* parameter. Can be a raw or standardized regression coefficient or whatever effect size measure.

The y axis is the associated p-value transformed in  $-\log_{10}(p)$  for better interpretation and visualization. Higher transformed p values are smaller raw p values.

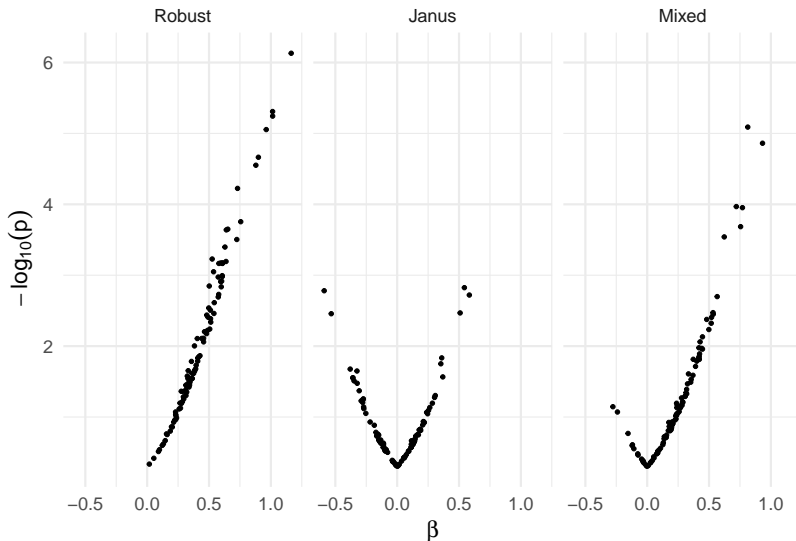
# Vibration of Effects (VoE)

The authors proposed to summarise the VoE using the range of effect sizes and p values. In particular the difference between the 99<sup>th</sup> and 1<sup>st</sup> percentiles.



# Vibration of Effects (VoE)

They identified three usual pattern for a Vulcano Plot:





# Vibration of Effects (VoE)

The **Robust** plot suggests a stable pattern across specifications, with the majority if not the total being positive and significant.

The **Janus**<sup>1</sup> plot suggests the worst scenario where in some conditions the effect is not only not significant but reversed.

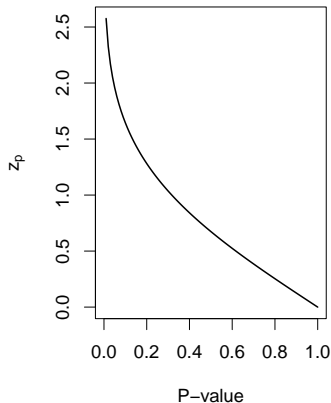
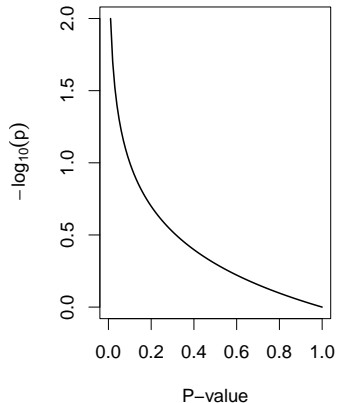
The **Mixed** plot suggests a less robust effect with few effect size reversals in rare specifications.

---

<sup>1</sup>Fun fact: Janus comes from the Roman/Greek god with two faces :)

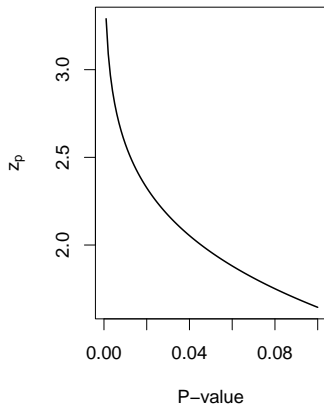
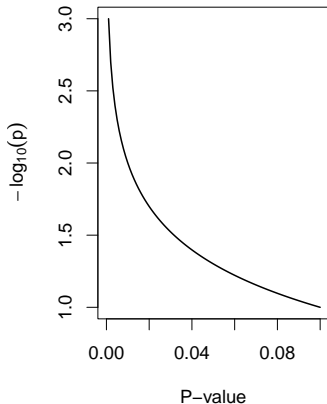
# P-values transformation

There are different ways to transform p-values to improve the interpretation and visualization.



# P-values transformation

Values higher than  $\sim 1.3$  (in  $\log_{10}$ ) or  $\sim 2$  ( $z$  transformation) are significant assuming the traditional  $\alpha = 0.05$ .

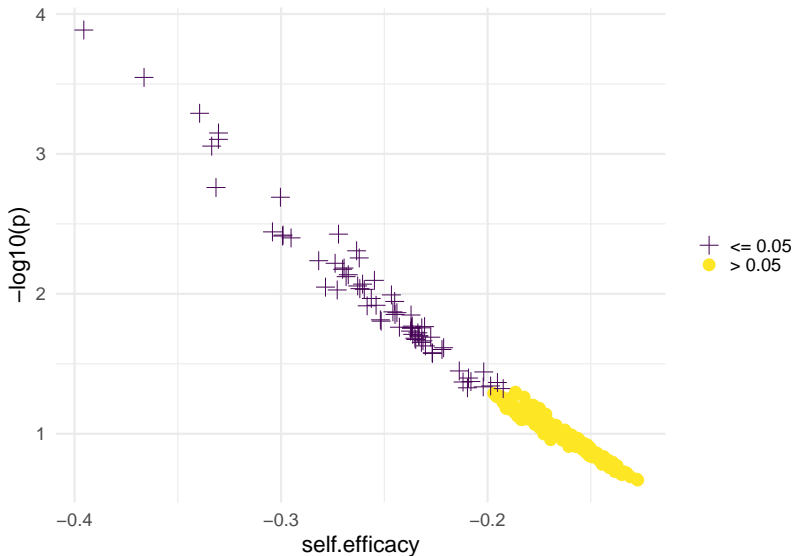


# Vulcano plot with our data

We can create a basic version of the vulcano plot with our dataset:

```
multi_res |>
  mutate(sign = ifelse(p <= 0.05, "<= 0.05", "> 0.05"),
         sign = factor(sign, levels = c("<= 0.05", "> 0.05"))) |>
  ggplot(aes(x = b, y = tp(p, "-log10"))) +
  geom_point(aes(shape = sign, color = sign), size = 5) +
  ylab("-log10(p)") +
  xlab(focal) +
  scale_shape_manual(values = c(3, 16)) +
  theme(legend.title = element_blank())
```

# Vulcano plot with our data



# Marginal/Conditional effects

A way to evaluate the impact of the multiverse scenario could be to use an ANOVA-style way of thinking. We can fit a regression model on the multiverse where the focal coefficient is the response variable and a series of dummy variables to code the inclusion/exclusion of a certain predictor.

Then we can estimate the % of explained variance of each predictor as an index of the impact in the multiverse results.

A more refined version of this approach can be found in Klau et al. (2023)

# Decomposing the multiverse variance

We can create a dataset with dummy variables when a specific predictor is included or not. We are ignoring the transformations of the specific variable.

```
# A tibble: 6 x 6
```

	b	faculty	asi	gender.category	program.type	frost.da
	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>
1	-0.396	0	0	0	0	0
2	-0.332	1	0	0	0	0
3	-0.330	0	1	0	0	0
4	-0.304	0	0	1	0	0
5	-0.334	0	0	0	1	0
6	-0.227	0	0	0	0	1

# Decomposing the multiverse variance

Then we can fit a linear regression and then evaluate the impact of including/excluding a predictor.

```
fit <- lm(b ~ ., data = multi_fit)
summary(fit)
```

Call:

```
lm(formula = b ~ ., data = multi_fit)
```

Residuals:

	Min	1Q	Median	3Q	Max
	-0.071822	-0.013166	0.002804	0.014675	0.037509

Coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	-0.323699	0.004734	-68.375	<2e-16 ***
faculty	0.027851	0.002733	10.190	<2e-16 ***
asi	0.002285	0.003417	0.669	0.504
gender.category	0.037357	0.002733	13.668	<2e-16 ***
program.type	0.028599	0.002733	10.463	<2e-16 ***
frost.da	0.099365	0.003417	29.083	<2e-16 ***

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

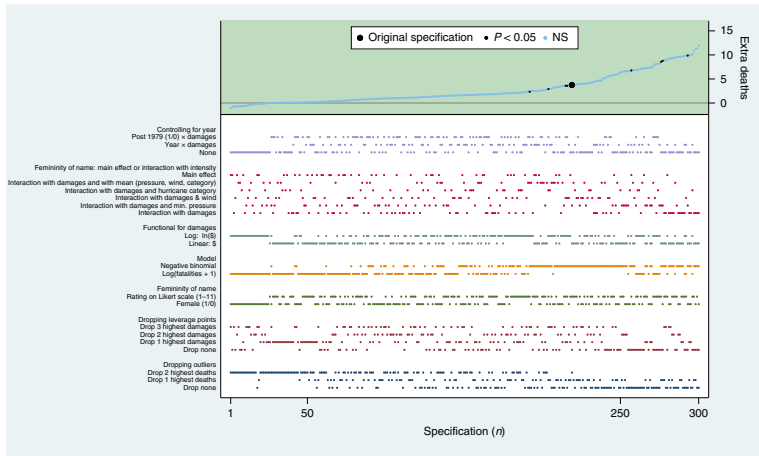


# Decomposing the multiverse variance

```
effectsize::eta_squared(fit, partial = FALSE)
```

# Specification Curve (Simonsohn et al., 2020)

The specification curve is both an inferential and descriptive tool to summarise the results from a multiverse analysis.



# Specification Curve as descriptive tool

Basically from  $M$  specifications we extract the focal coefficient then:

- ▶ we sort the coefficients from the lowest to the highest and assign a progressive index
- ▶ we plot the index as a function of the coefficient value
- ▶ for each scenario we code the corresponding set of conditions/variables
- ▶ we combine the previous plot with a tile-plot (or similar) showing for each scenario the set of variables/choices

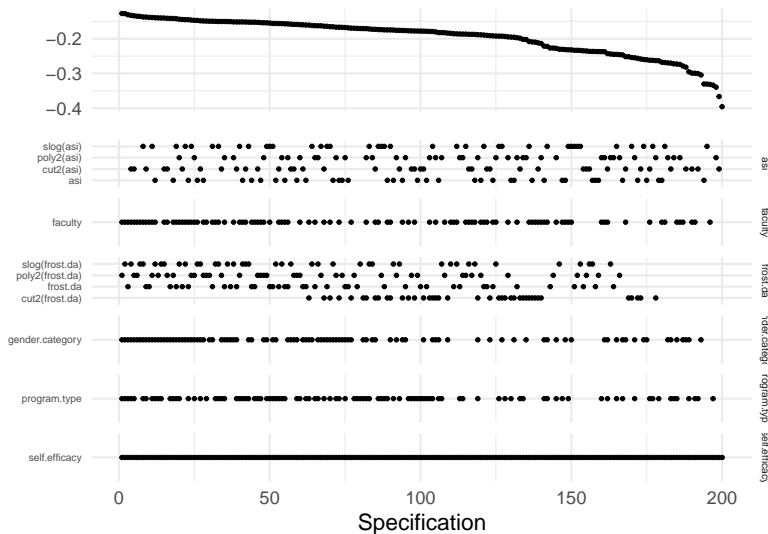
# Specification with the dataset

```
spec_data <- multi_res |>
  # sorting
  arrange(desc(b)) |>
  # index with the order
  mutate(spec = 1:n())

top <- spec_data |>
  # confidence intervals
  mutate(lb = b - se * 2,
         ub = b + se * 2) |>
  ggplot(aes(x = spec, y = b)) +
  geom_point() +
  theme(axis.text.x = element_blank(),
        axis.title.x = element_blank(),
        axis.title.y = element_blank())

bottom <- spec_data |>
  pivot_longer(starts_with("x_")) |>
  drop_na() |>
  mutate(name = gsub("x_", "", name)) |>
  ggplot(aes(x = spec, y = value)) +
  geom_point() +
  theme(axis.title.y = element_blank(),
        strip.text.y = element_text(size = 9),
        axis.text.y = element_text(size = 9)) +
  xlab("Specification") +
  facet_grid(name~., scales = "free")
```

# Specification with the dataset



# Other descriptive tools

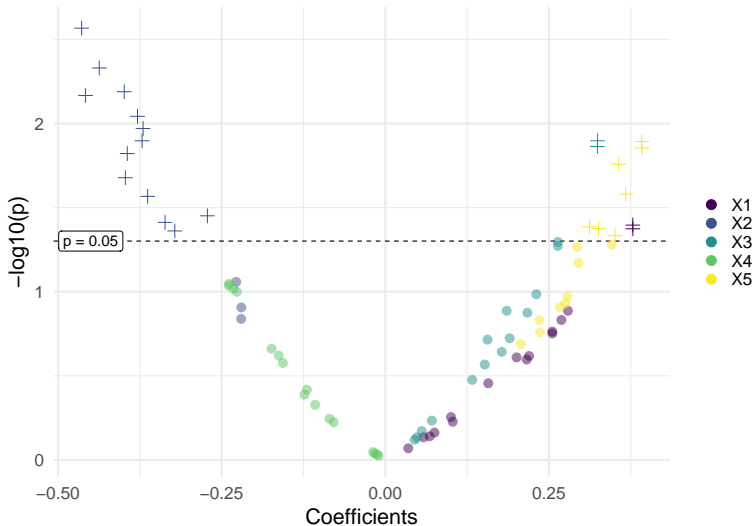
In general, any descriptive statistics can be useful. The main points in a multiverse description are:

- ▶ the range of the estimated effects
- ▶ the impact of the choices
- ▶ the impact on the conclusions (e.g., statistical significance)

**Can the EMA be misleading?**

# Let's have a look to another example

We have a multiverse with 31 scenarios, 50 observations and 5 predictors, this is the vulcano plot. **What do you think?**





## Let's have a look to another example<sup>2</sup>

From the previous multiverse it is clear that something is going on. Some of the coefficients are significant and other not. There is also a little bit of Janus effect.

But, the previous example was a simulated multiverse where all the coefficients  $\beta_j = 0$  (the null hypothesis is true). **All the significant scenarios are false positives (type-1 error)!**

---

<sup>2</sup>Thanks to Livio Finos for the insightful example

## Let's have a look to another example<sup>2</sup>

From the previous multiverse it is clear that something is going on. Some of the coefficients are significant and other not. There is also a little bit of Janus effect.

But, the previous example was a simulated multiverse where all the coefficients  $\beta_j = 0$  (the null hypothesis is true). **All the significant scenarios are false positives (type-1 error)!**

---

<sup>2</sup>Thanks to Livio Finos for the insightful example

# Why? multiple testing problem!

- ▶ A multiverse can be considered as a **multiple testing problem** because we are testing a set of hypotheses with the same dataset. The type-1 error rate ( $\alpha$ ) need to be controlled otherwise the actual level is higher than the nominal level.
- ▶ We can demonstrate this with a simple simulation. We simulate  $k$  variables and a one-sample t-test for each variable. The ground truth is that we have  $\mu_1, \mu_2, \dots, \mu_k = 0$  thus  $H_0$  is true.
- ▶ We repeat the simulation  $B$  times and we count how many times  $p \leq \alpha$  for at least one of the  $k$  tests. This is our estimated type-1 error rate.

# Why? multiple testing problem!

```
k <- 10 # number of variables
n <- 100 # number of observations
R <- 0 + diag(1 - 0, k) # correlation matrix
B <- 1e3

PM <- matrix(NA, B, k)

for(i in 1:B){
  X <- MASS::mvrnorm(n, rep(0, k), R)
  p <- apply(X, 2, function(x) t.test(x)$p.value)
  PM[i, ] <- p
}

# type-1 error for each variable, ignoring multiple testing
apply(PM, 2, function(x) mean(x <= 0.05))

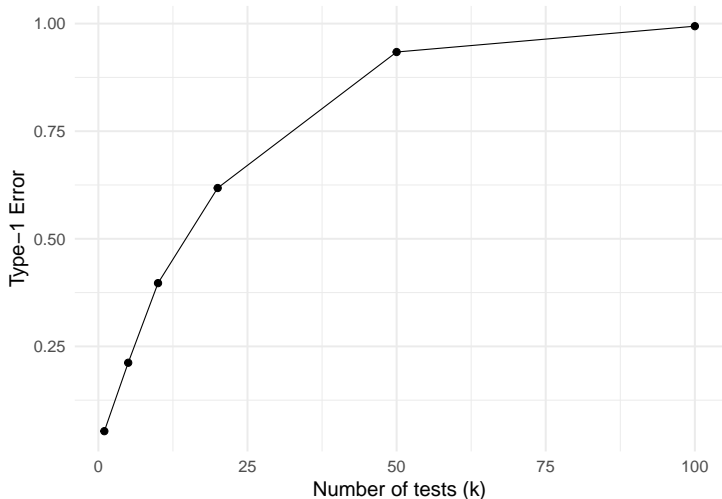
[1] 0.048 0.045 0.055 0.052 0.052 0.061 0.049 0.049 0.056 0.059

# type-1 error considering the k tests (should be alpha)
mean(apply(PM, 1, function(x) any(x <= 0.05)))

[1] 0.427
```

# Why? multiple testing problem!

To have a better overview, we can repeat the simulation for different number of  $k$ . Quite scary right?



# So what? No multiverse?

Exploring is fine and is quite important if not fundamental. But, when we explore the p-values thus the **inferential** results from the single scenarios, we are inflating the type-1 error and our **inferential** conclusions are no longer valid.

If we want an inferential answer from our multiverse (not always the case) we need a proper inferential framework. This is the role of the **inferential multiverse analysis**.

# So what? No multiverse?

Exploring is fine and is quite important if not fundamental. But, when we explore the p-values thus the **inferential** results from the single scenarios, we are inflating the type-1 error and our **inferential** conclusions are no longer valid.

If we want an inferential answer from our multiverse (not always the case) we need a proper inferential framework. This is the role of the **inferential multiverse analysis**.

# **Inferential Multiverse Analysis (IMA)**



# Family-wise error rate (FWER)<sup>3</sup>

		$H_0$		Tot
		False	True	
Test	Rejected	<b>True Positive (S)</b>	<b>False Positive (V)</b>	$R$
	Not rejected	<b>False Negative (T)</b>	<b>True Negative (U)</b>	$m - R$
Tot		$m_1$	$m_0$	$m$

The FWER is the probability of committing type-1 error thus  $P(V > 0)$ . Controlling the FWER (whatever the methods) keep  $P \leq \alpha$ .

There are different procedures for controlling the FWER, such as the Bonferroni or the Holm–Bonferroni method.

---

<sup>3</sup>Thanks to Anna Vesely for the amazing introduction to the multiple testing problem (see the [slides](#))

# Correcting the p-values

The main problem is that the number of tests in a multiverse can be quite large.

As an example, we simulated a series of tests with different effect size to show the impact on the type-1 error rate and the power.

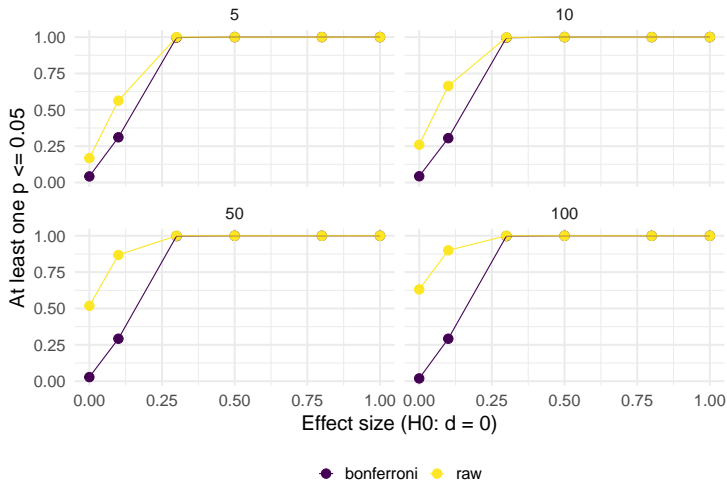
# Correcting the p-values

The main problem is that the number of tests in a multiverse can be quite large.

As an example, we simulated a series of tests with different effect size to show the impact on the type-1 error rate and the power.

# Correcting the p-values

Using a standard method (e.g., Bonferroni) clearly controls the type-1 error but reduces a lot the statistical power. At the same time, without correction the inflation is large.



# Correlation between scenarios is (probably) large

The multiverse scenarios are computed on the same dataset thus the correlation between tests is probably medium-large. For example:

```
x <- runif(100, 5, 10)
y <- x * 0.1 + rnorm(100)

fit1 <- lm(y ~ x)
fit2 <- lm(y ~ cut(x, breaks = 2))
fit3 <- lm(y ~ log(x))
fit4 <- lm(y ~ poly(x, 2))

pp <- sapply(list(fit1, fit2, fit3, fit4), predict)
round(cor(pp), 2)
```

```
      [,1] [,2] [,3] [,4]
[1,] 1.00 0.88 1.00 0.99
[2,] 0.88 1.00 0.88 0.88
[3,] 1.00 0.88 1.00 1.00
[4,] 0.99 0.88 1.00 1.00
```

# A more powerful correction method<sup>4</sup>

The Bonferroni (and similar) methods assume that the tests are independent thus regardless the dependence structure the FWER is under control.

The permutation-based methods (maxT, minP, etc.) take into account the correlation structure providing FWER control under  $H_0$  but a more powerful test under  $H_1$ .

---

<sup>4</sup>Goeman & Solari (2014)

# Permutation testing in a nutshell

Permutation testing requires computing the distribution of the test statistics  $T$  where we know that  $H_0$  is true.

We can force the null to be true permuting the data *removing* the assumed effect. We repeat this process a large number of times  $B$ .

Then we compare the observed test statistics  $T_1$  with the distribution of permuted test statistics obtaining the permutation based p-value  $p = \frac{\#(T_1 \geq T_B)}{B}$ <sup>5</sup>

---

<sup>5</sup> $\#$  is the count function.

# Permutation testing in a nutshell

Permutation testing requires computing the distribution of the test statistics  $T$  where we know that  $H_0$  is true.

We can force the null to be true permuting the data *removing* the assumed effect. We repeat this process a large number of times  $B$ .

Then we compare the observed test statistics  $T_1$  with the distribution of permuted test statistics obtaining the permutation based p-value  $p = \frac{\#(T_1 \geq T_B)}{B}$ <sup>5</sup>

---

<sup>5</sup># is the count function.



# Permutation testing in a nutshell

Permutation testing requires computing the distribution of the test statistics  $T$  where we know that  $H_0$  is true.

We can force the null to be true permuting the data *removing* the assumed effect. We repeat this process a large number of times  $B$ .

Then we compare the observed test statistics  $T_1$  with the distribution of permuted test statistics obtaining the permutation based p-value  $p = \frac{\#(T_1 \geq \mathbf{T}_B)}{B}$ <sup>5</sup>

---

<sup>5</sup># is the count function.

# Permutation testing in a nutshell

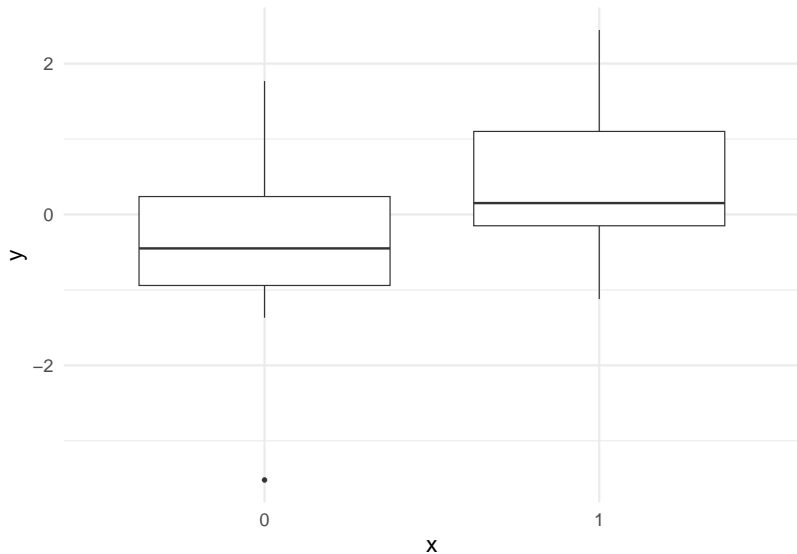
Let's make an example with a two-groups comparison:

```
N <- 30  
d <- 1 # effect size  
x <- rep(c(0, 1), each = N/2) # dummy for the group  
y <- rnorm(N, d * x, 1)  
tapply(y, x, mean)
```

0	1
-0.3933720	0.4743739

# Permutation testing in a nutshell

Let's make an example with a two-groups comparison:



# Permutation testing in a nutshell

We need to flip the group label thus removing the group effect.

```
B <- 1e3 # number of permutations
tp <- rep(NA, B)
tp[1] <- t.test(y ~ x)$statistic # first permutation always the observed data

sample(x) # shuffling the group label
```

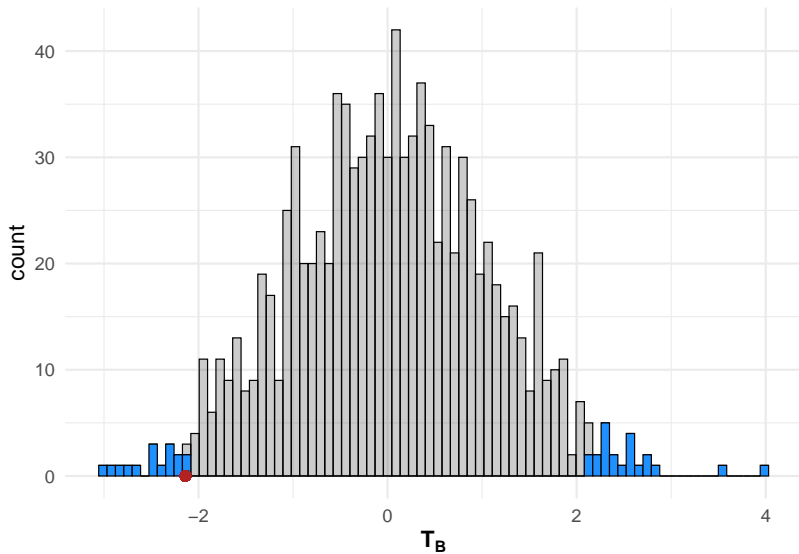
```
[1] 0 1 1 1 1 0 0 0 0 1 0 0 0 0 0 1 1 0 1 0 0 0 1 1 1 1 1 1 0 1

for(i in 2:B){
  xp <- sample(x)
  tp[i] <- unname(t.test(y ~ xp)$statistic)
}

mean(abs(tp) >= abs(tp[1]))
```

```
[1] 0.038
```

# Permutation testing in a nutshell



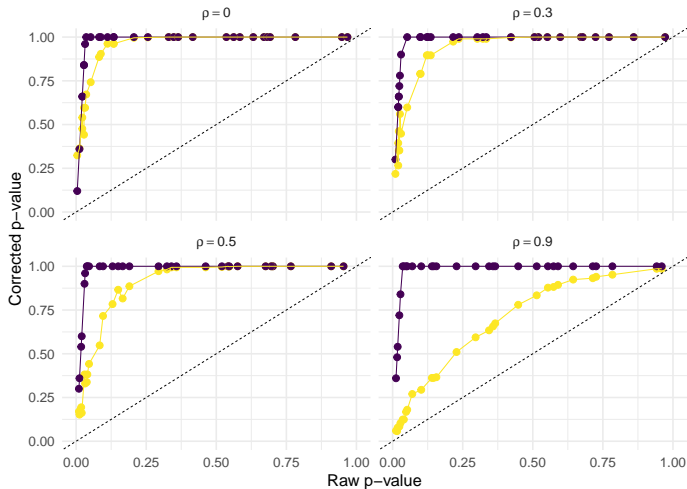
# maxT procedure Westfall & Stanley Young (1993)

The maxT is a permutation-based method to control the FWER. With the method we can obtain:

- ▶ overall inference across  $M$  tests with *weak* control of FWER
- ▶ individually adjusted p-values for each test (i.e, *strong* FWER control)

# maxT with correlated variables

Beyond the actual method and algorithm, the advantage of the maxT approach is taking into account the correlation between tests.



# Inferential Methods

- ▶ Specification Curve
- ▶ Post-Selection Inference in Multiverse Analysis (PIMA)



# Specification Curve

The specification curve (Simonsohn et al., 2020) is the first attempt to build an inferential framework for multiverse analysis.

- ▶ provides only *weak* control of type-1 error
- ▶ is not directly applicable to GLMs (only standard linear models, see Girardi et al., 2024)
- ▶ is computationally expensive

# The PIMA recipe... (Girardi et al., 2024)

PIMA provides *weak* and *strong* type-1 error control with a powerful method based on permutations ( $\max T$ ) and applicable to whatever GLM (Logistic, Poisson, etc.).

For constructing the inferential approach with PIMA we need:

- ▶ a flexible modelling framework: **Generalized Linear Models**
- ▶ a permutation-based inferential approach: **Flipscores**
- ▶ a permutation-based and powerful method for weak and strong FWER control:  **$\max T$**

# The core of PIMA, the **flipscores** method

- ▶ The formal part of the **flipscores** method is quite complex and beyond our scope and expertise. But a detailed description can be found in Hemerik et al. (2020) and Girardi et al. (2024).
- ▶ Essentially the **flipscores** method is an alternative way of doing inference for parameters of a GLM based on permutations.
- ▶ The idea is conceptually the same as the two-groups example, but can work for multiple regression models with covariates and interactions.

# Intution of flipscores

This method can be extended to whatever GLM and to any number of predictors/confounders.

The actual permutation test is obtained flipping the sign of the scores/residuals thus obtaining the distribution under the null hypothesis of the test statistics.

Everything is implemented into the `flipscores` package (Hemerik et al., 2020) and on CRAN

<https://cran.r-project.org/web/packages/flipscores/index.html>.

# flipscores package

With the `flipscores` function is very easy to fit a linear model with permutations-based p-values.

```
library(flipscores)
fit <- flipscores(Sepal.Length ~ Petal.Width + Species, data = iris)
summary(fit)
```

Call:

```
flipscores(formula = Sepal.Length ~ Petal.Width + Species, data = iris)
```

Coefficients:

	Estimate	Score	Std. Error	z value	Part. Cor
(Intercept)	4.78044	160.25913	13.50622	11.86558	0.979
Petal.Width	0.91690	5.64500	1.27732	4.41941	0.365
Speciesversicolor	-0.06025	-0.26260	1.00098	-0.26234	-0.022
Speciesvirginica	-0.05009	-0.09030	0.64372	-0.14028	-0.012

Pr(>|z|)

(Intercept)	0.0002 ***
Petal.Width	0.0002 ***
Speciesversicolor	0.7954
Speciesvirginica	0.9014

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for gaussian family taken to be 0.2313718)

Null deviance: 102.17 on 149 degrees of freedom  
Residual deviance: 33.78 on 146 degrees of freedom  
AIC: 212.07

Number of Fisher Scoring iterations: 2

# Intuition of PIMA

The idea of PIMA is to extend the `flipscores` method to  $M$  models (where  $M$  is the number of scenarios) and perform inference at the multiverse level.

Using the `maxT` approach we can combine the  $M$  tests into a single test with weak control of FWER. The global null hypothesis is:

$$\mathcal{H} = \bigcap_{m=1}^M \mathcal{H}_m : \beta_m = 0 \text{ for all } m = 1, \dots, M.$$

In addition, we can correct the individual p-values with strong FWER control using the `maxT` method.

# The pima package

We are implementing everything into the `pima` package that is under development. You can try it but there could be bugs and breaking changes in the near future.

You can explore the package here

<https://github.com/livioivil/pima>. The package mainly depends on `jointest` that is the actual package for combining multiple (correlated) tests and correcting them.

# The pima package

The package has a main function called `pima::pima()` that takes a list of models (of class `glm`) and compute the global test and the correction for individual scenarios.

```
# fitl is the list of fitted models
# tested_coeffs is the focal variable, other are confounders

library(pima)

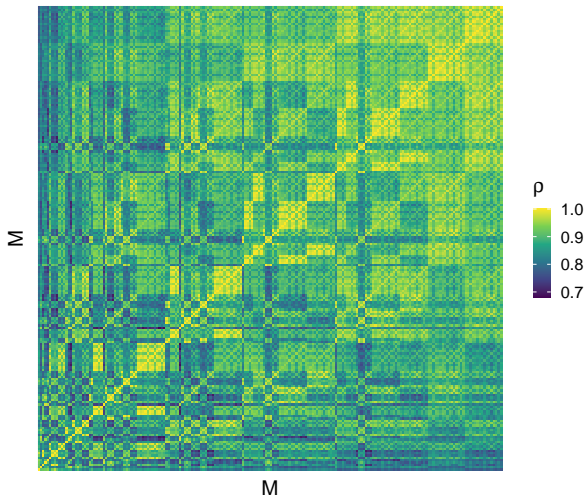
# this is a bug/part to be improved, ignore
for(i in 1:length(fitl)){
  fitl[[i]]$call$formula <- as.formula(fitl[[i]]$formula)
}

multi_pima <- pima(fitl, tested_coeffs = "self.efficacy")
```



# The pima results

The correlations between the scenarios is very high, the maxT method will be powerful!



# The pima results

```
# overall test, weak control FWER
summary(pima::global_tests(multi_pima))
```

	Model	Coeff	Stat	nTests	S	p
1	Overall	self.efficacy	maxT	200	30.26	4e-04

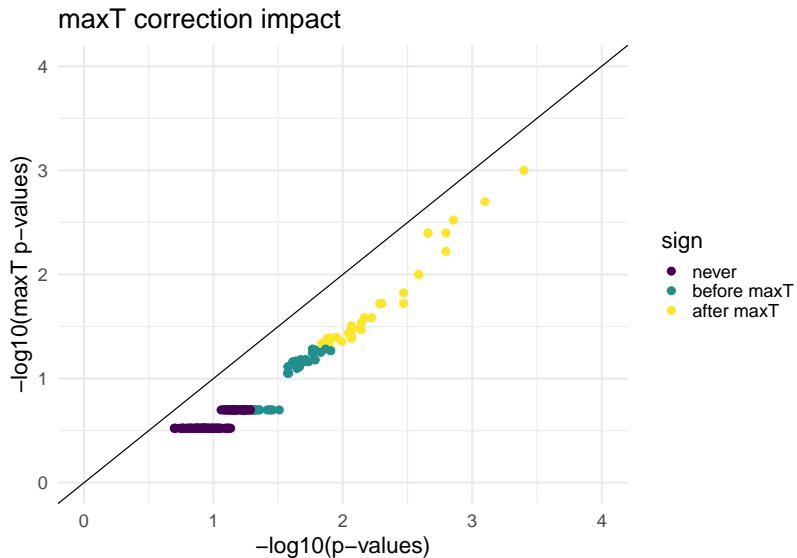
```
# adjusted p values, strong control FWER
head(summary(multi_pima))
```

	Model	.assign	Coeff	Estimate	Score	Std. Error
1	Model1	1 self.efficacy	-0.3955213	-30.26473	8.026511	
2	Model2	1 self.efficacy	-0.3315150	-23.30487	7.506267	
3	Model3	1 self.efficacy	-0.3302943	-24.84790	7.416916	
4	Model4	1 self.efficacy	-0.3041665	-21.44863	7.420203	
5	Model5	1 self.efficacy	-0.3335836	-24.92025	7.567967	
6	Model6	1 self.efficacy	-0.2266493	-15.42204	6.974630	

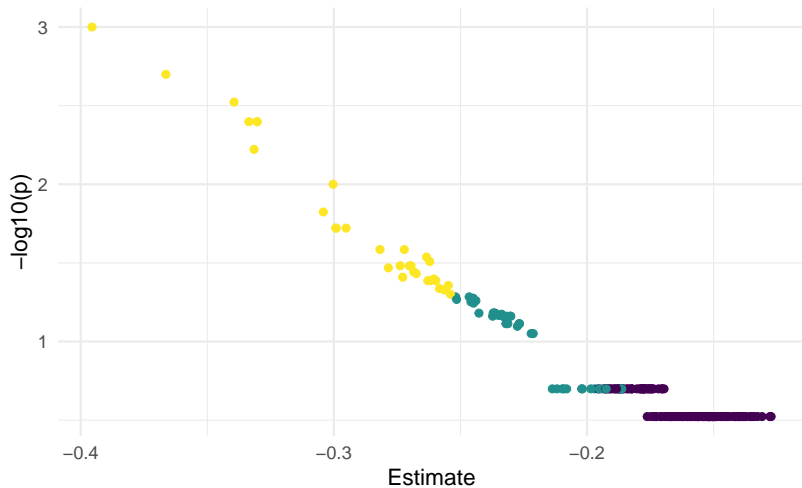
  

	z value	Part. Cor	p	p.adj.maxT
1	-3.770596	-0.2672905	0.0004	0.001
2	-3.104722	-0.2212023	0.0016	0.006
3	-3.350167	-0.2380860	0.0022	0.004
4	-2.890572	-0.2054240	0.0034	0.015
5	-3.292859	-0.2340133	0.0016	0.004
6	-2.211162	-0.1571404	0.0266	0.077

# maxT correction



# Improved vulcano plot



$p \leq 0.05$    never   before maxT   after maxT

# References

- Girardi, P., Vesely, A., Lakens, D., Altoè, G., Pastore, M., Calcagnì, A., & Finos, L. (2024). Post-selection inference in multiverse analysis (PIMA): An inferential framework based on the sign flipping score test. *Psychometrika*, 89, 542–568. <https://doi.org/10.1007/s11336-024-09973-6>
- Goeman, J. J., & Solari, A. (2014). Multiple hypothesis testing in genomics. *Statistics in Medicine*, 33, 1946–1978. <https://doi.org/10.1002/sim.6082>
- Götz, M., Sarma, A., & O'Boyle, E. H. (2024). The multiverse of universes: A tutorial to plan, execute and interpret multiverses analyses using the r package multiverse. *International Journal of Psychology: Journal International de Psychologie*, 59, 1003–1014. <https://doi.org/10.1002/ijop.13229>
- Hemerik, J., Goeman, J. J., & Finos, L. (2020). Robust testing in generalized linear models by sign flipping score contributions. *Journal of the Royal Statistical Society. Series B, Statistical Methodology*, 82, 841–864. <https://doi.org/10.1111/rssb.12369>
- Klau, S., Felix, Patel, C. J., Ioannidis, J. P. A., Boulesteix, A.-L., & Hoffmann, S. (2023). Comparing the vibration of effects due to model, data pre-processing and sampling uncertainty on a large data set in personality psychology. *Meta-Psychology*, 7. <https://doi.org/10.15626/mp.2020.2556>

# References (cont.)

- McCaughey, N. J., Hill, T. G., & Mackinnon, S. P. (2022). The association of self-efficacy, anxiety sensitivity, and perfectionism with statistics and math anxiety. *Personality Science*, 3. <https://doi.org/10.5964/ps.7091>
- Patel, C. J., Burford, B., & Ioannidis, J. P. A. (2015). Assessment of vibration of effects due to model specification can demonstrate the instability of observational associations. *Journal of Clinical Epidemiology*, 68, 1046–1058. <https://doi.org/10.1016/j.jclinepi.2015.05.029>
- Simonsohn, U., Simmons, J. P., & Nelson, L. D. (2020). Specification curve analysis. *Nature Human Behaviour*, 4, 1208–1214. <https://doi.org/10.1038/s41562-020-0912-z>
- Westfall, P. H., & Stanley Young, S. (1993). *Resampling-based multiple testing: Examples and methods for p-value adjustment*. John Wiley & Sons.