

Ensemble approaches for estimating congruence between species delimitation and morphological variation: comparing taxonomic hypotheses for the Pacific Pond Turtle (*Emys marmorata*)

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Abstract

We investigated the morphometric identification of cryptic species using machine learning approaches by examining their implications for a recently proposed cryptic turtle species (*Emys pallida*). We collected landmark-based morphometric data from 532 adult *E. marmorata*/*E. pallida* museum specimens. We assigned a classification to each specimen for six different binning schemes based on geographic occurrence data recorded in museum collection archives. We used an ensemble of supervised machine learning approaches to determine which classification hypothesis was best supported by the data. In addition, we applied the same approach to two clear-cut examples, one consisting of seven unambiguously distinct species closely related to *E. marmorata*, and the other consisting of two subspecies of *Trachemys scripta*. Our results indicate that there is no clear “best” grouping of *E. marmorata*/*E. pallida* based on plastron shape. In contrast, the analyses of the clear-cut examples produced near perfect classifications,

demonstrating that the methods can recover correct results when an appropriate signal exists. Explanations for the lack of grouping in *E. marmorata* include the possibility that genetic differentiation is not associated with plastron shape variation below the species level and/or that local selective pressures (e.g., from hydrological regime) overwhelm morphological differentiation. A reconsideration of the methods used to delimit “*E. pallida*,” the lack of barriers to gene flow, the strong evidence for widespread admixture between lineages, and the fact that plastron shape can be used to delineate other emydine species and sub-species suggest that its lack of diagnosability most likely reflects the non-distinctiveness of this proposed taxon.

INTRODUCTION

Molecular systematics has repeatedly demonstrated the existence of cryptic species that can only be diagnosed using genetic data (Stuart et al. 2006; Bickford et al. 2007; Schilck-Steiner et al. 2007; Pfenninger and Schwenk 2007; Clare 2011; Funk et al. 2012). In attempts to streamline the documentation of biodiversity, several methods of species delimitation that rely almost entirely on genetic data have recently been proposed (Pons et al. 2006; Carstens and Dewey 2010; Hausdorf and Hennig 2010; O’Meara 2010; Yang and Rannala 2010; Huelsenbeck et al. 2011). Although strong caveats on the utility of these methods have been raised (Bauer et al. 2000; Carstens et al. 2013), they are nevertheless being used to name species (Leaché and Fujita 2010; Spinks et al. 2014).

In contrast to those genetically-diagnosed species, the majority of extant taxa, and almost all extinct taxa, are delimited by morphology alone. This disjunction complicates interpretations of variation and diversity in deep time, as apparent morphological stasis may not reflect the true underlying diversity (Eldredge and Gould 1972; Gould and Eldredge 1977; Van Bocxlaer and Hunt 2013). It also has serious implications for our records of modern biodiversity: for many museum specimens of extant taxa (e.g. those preserved in formalin), it is difficult to acquire the genetic data needed for non-morphological species delimitation methods.

These considerations have sparked interest in whether geometric morphometric analyses can capture fine-scale variation that can be used for identifying cryptic species. This would make the task of identifying and maintaining endangered or conserved groups much easier and could contribute to improved classifications of extinct taxa and populations. Most such studies focus on using morphometrics to discover differences between taxa that were identified by other means (Polly 2003; Zelditch et al. 2004; Gaubert et al. 2005; Gündüz et al. 2007; Polly 2007; Demandt and Bergek 2009; Markolf et al. 2013; Fruciano et al. 2016). Additionally, there has been work on automated taxon identification and classification of taxa into groups (Baylac et al. 2003; Dobigny et al. 2003; MacLeod 2007; van den Brink and Bokma 2011; Vitek et al. 2017).

Here, we investigate the morphometric identification of cryptic species using machine learning approaches. We use an ensemble learning approach where multiple methods are used in order to look for consensus among their results. We test our approach on three datasets: plastron

shape of seven species of closely related turtles, plastron shape of two species of closely related turtles, and plastron shape of the *Emys marmorata* species complex. In particular, we ask whether it is possible to determine which among a set of classification hypotheses best aligns with the observed morphology, and examine the implications of our results for the *E. marmorata* complex.

Background and study system

Machine learning is an extension of known statistical methodology (Hastie et al. 2009) that emphasizes high predictive accuracy and generality often at the expense of the interpretability of individual parameters. Basic statistical approaches are supplemented by randomization, sorting, and partitioning algorithms, along with the maximization or minimization of summary statistics, in order to best estimate a general model for all data, both sampled and unsampled (Hastie et al. 2009). Machine learning approaches have found use in medical research, epidemiology, economics, and automated identification of images such as handwritten zip codes (Hastie et al. 2009).

There are two major classes of machine learning method: unsupervised and supervised learning. Unsupervised learning methods are used with unlabeled data where the underlying structure is estimated; they are analogous to clustering and density estimation methods (Kaufman and Rousseeuw 1990). Supervised learning methods are used with labeled data where the final output of data is known and the rules for going from input to output are inferred. These are analogous to classification and regression models (Breiman et al. 1984; Hastie et al. 2009). Our application of the supervised learning approaches used in this study illustrates only a sampling of the various methods available for fitting classification models. The specific methods used in this study were chosen because they are suited for cases with more two or more response classes.

Geometric morphometric approaches to identifying differences in morphological variation between classes, including cryptic species, have mostly relied on methods like linear discriminate analysis and canonical variates analysis (Polly 2003; Zelditch et al. 2004; Gaubert et al. 2005; Gündüz et al. 2007; Polly 2007; Francoy et al. 2009; Sztencel-Jablonka et al. 2009; Mitrovski-Bogdanovic et al. 2013; Dillard 2017). Because of their similarity to multivariate approaches like principal components analysis (PCA), these methods are comparatively straightforward ways of understanding the differences in morphology between classes. They also benefit from producing results that can be easily visualized, which aids in the interpretation and presentation of data and results. Most previous morphometric studies did not assess which amongst a set of alternative classification hypotheses was optimal. For example, studies such as those of Caumul and Polly (2005) and Polly (2007) focused on comparing different aspects of morphology and their fidelity to a classification scheme instead of comparing the fidelity of one aspect of morphology to multiple classification schemes. In this context, the study of Cardini et al. (2009) is noteworthy because they compared morphological variation in marmots at the population, regional, and species level and determined the fidelity of shape to divisions at each of these levels.

Here, we used an ensemble of supervised machine learning methods to compare the congruence of the morphological data to different classification hypotheses. Each of these methods provide different advantages for understanding how to classify taxa, as well as the accuracy of the resulting classifications. Machine learning methods have been combined with geometric morphometric data to study shape variation in a variety of contexts, including automated taxon identification and classification of groups (Baylac et al. 2003; Dobigny et al. 2003; MacLeod 2007; Van Bocxlaer and Schultheiß 2010; van den Brink and Bokma 2011; Navega et al. 2015). In the current study, we not only consider pure classification accuracy but also use a statistic of classification strength that reflects the rate at which taxa are both accurately and inaccurately classified: the area under the Receiver Operating Characteristic curve (Hastie et al. 2009).

We analyzed the problem of whether there are distinct subspecies or cryptic species within the western pond turtle, *Emys marmorata* (Baird and Girard 1852) (formerly *Clemmys marmorata*; see Feldman and Parham 2002). *Emys marmorata* is distributed from northern Washington State, USA to Baja California, Mexico. Traditionally, *E. marmorata* was classified into two named subspecies: the northern *E. marmorata marmorata* and the southern *Emys marmorata pallida* (Seeliger 1945), with a central Californian intergrade zone in between. *Emys marmorata marmorata* is differentiated from *E. marmorata pallida* by the presence of a pair of triangular inguinal scales and darker neck markings. The triangular inguinal plates can sometimes be present in *E. marmorata pallida* although they are considerably smaller. Seeliger (1945) did not formally include the Baja California populations of *E. marmorata* in either taxon, implying the existence of a third distinct but unnamed subspecies.

Previous work on morphological variation in *E. marmorata* has focused primarily on differentiation between populations over a portion of the species' total range (Lubcke and Wilson 2007; Germano and Rathbun 2008; Germano and Bury 2009; Bury et al. 2010); comparatively few studies have included specimens from across the entire range (Holland 1992). Most of these studies considered how local biotic and abiotic factors may contribute to differences in carapace length, and they found that size can vary greatly between different populations (Lubcke and Wilson 2007; Germano and Rathbun 2008; Germano and Bury 2009). There also has been interest in size-based sexual dimorphism in *E. marmorata* (Holland 1992; Lubcke and Wilson 2007; Germano and Bury 2009), with males being on average larger than females based on total carapace length and other linear measurements. However, the quality of size as a classifier of sex can vary greatly between populations (Holland 1992) because of the magnitude of size differences among populations (Lubcke and Wilson 2007; Germano and Bury 2009). The effect of sexual dimorphism on shape, *sensu* Kendall (1977), has not been assessed (Holland 1992; Lubcke and Wilson 2007; Germano and Rathbun 2008).

Of particular relevance in the context of cryptic diversity in *E. marmorata* is the morphometric analysis of carapace shape carried out by Holland (1992), who compared populations of *E. marmorata* from three areas of the species range. Holland concluded that geographic distance was a poor indicator of morphological differentiation, and instead hypothesized that geographic features such as breaks between different drainage basins are probably more important barriers

to dispersal and interbreeding. Additionally, he suggested that morphological differences were more pronounced as the magnitude of barriers and distance increased, but this variation required many variables to adequately capture, implying only very subtle morphological differentiation between putatively distinct populations. Finally, Holland concluded that *E. marmorata* is best classified as three distinct species: a northern species, a southern species, and a Columbia Basin species. This classification is similar to that of Seeliger (1945), except elevated to the species level and without recognition of a distinct Baja species.

More recently, the phylogeography of *E. marmorata* and the possibility of cryptic diversity was investigated using molecular data (Spinks and Shaffer 2005; Spinks et al. 2010, 2014). Based on mitochondrial DNA, Spinks and Shaffer (2005) recognized four subclades within *E. marmorata*, a northern clade, a San Joaquin Valley clade, a Santa Barbara clade, and a southern clade. Analyses with nuclear DNA (Spinks et al. 2010) and single-nucleotide polymorphism (SNP) data suggest a primarily north–south division in *E. marmorata*, although these datasets differed from that of mitochondrial-based results of Spinks and Shaffer (2005) in the location of the break point (Spinks et al. 2014). All three studies discussed the potential taxonomic implications of their results, with Spinks et al. (2014) going so far as to strongly advocate for the recognition of at least two species (*E. marmorata* and *E. pallida*), and a possible third based on populations in Baja California. However, they did not discuss in detail the morphological characters that would help to diagnose these species beyond those specified by Seeliger (1945). Given that these characters are variable within the proposed species, and that Holland (1992) described shell shape variation that might be consistent with this taxonomy, a geometric morphometric analysis of shell shape might provide a reliable way to diagnose groups (whether species or subspecies) within *E. marmorata*.

In this study, we attempt to estimate the best classification scheme of *E. marmorata* based on variation in plastron (ventral shell) shape in order to determine whether this character is consistent with any of the proposed taxonomies of the *E. marmorata* complex.

We choose to analyze plastron shape for multiple reasons. First, it is very easy to collect geometric morphometric data on plastron shape from two-dimensional pictures as the structure is virtually flat. This approach allows both museum specimens and individuals in the field to be analyzed together. Second, previous work has suggested that there are strong differences in plastron shape among traditionally recognized emydine species (Angielczyk and Sheets 2007; Angielczyk et al. 2011; Angielczyk and Feldman 2013). Finally, due to this previous study a large dataset was readily available.

In the case of the *E. marmorata* species complex, we hypothesize that if one or more of the proposed classification schemes are consistent with the morphological data then our ensemble approach fit to those hypotheses will have higher out-of-sample predictive performance than the more inconsistent hypotheses. However, if all of the classification schemes lead to equal out-of-sample predictive performance then we would conclude that the proposed hypotheses are inconsistent with whatever information is present in the morphological data. Because of unclear geographic boundaries between subgroups of *E. marmorata*, we compare multiple

174 permutations of the (Spinks et al. 2010) and Spinks et al. (2014) hypotheses.

MATERIALS AND METHODS

Specimens, sampling, morphometrics

176 Three different geometric morphometric datasets describing turtle plastron variation were
178 assembled for this analysis: 1) specimens from seven distinct emydine species; 2) *T. scripta*
specimens from the two main subspecies (*T. scripta elegans* and *T. scripta scripta*); and 3)
180 *E. marmorata* specimens from across the species' geographic range. The first two datasets
are intended to serve as a test of whether machine learning techniques can differentiate
182 species-level groupings of emydine turtles using plastron shape. We expect that the first case
represents a low complexity dataset because of the high level of plastron shape disparity that
184 exists among these species (Angielczyk et al. 2011), whereas the second dataset should be
relatively higher in complexity and more analogous to the *E. marmorata* example. We predict
186 that the *E. marmorata* dataset should be of the highest complexity and our greatest challenge
given the finding that only very subtle differences existed between geographically-distinct
188 populations (Holland 1992).

The first dataset we analyzed includes 578 total specimens from the following species: *Clemmys*
190 *guttata*, *Emys blandingii*, *Emys orbicularis*, *Glyptemys insculpta*, *Glyptemys muhlenbergii*,
Terrapene coahuila, and *Terrapene ornata*. These specimens are a subset of those used in
192 Angielczyk et al. (2011) and Angielczyk and Feldman (2013).

The second dataset is a compilation of 101 specimens of two subspecies of *T. scripta*: 51
194 specimens of *T. scripta scripta* and 50 specimens of *T. scripta elegans*. These landmark data
are new to this study.

196 The final dataset is of 532 adult *E. marmorata* museum specimens, though not all specimens
were able to be assigned a class for all schemes (Fig. 1). These specimens represent a subset
198 of those included in Angielczyk and Sheets (2007), Angielczyk et al. (2011), and Angielczyk
and Feldman (2013). Because Spinks and Shaffer (2005), Spinks et al. (2010), and Spinks
200 et al. (2014) did not use voucher specimens we were not able to directly sample the
individuals in their studies. Instead, our specimen classifications were based solely on the
202 geographic information and not explicit assignment using molecular data. For each taxonomic
hypothesis, specimens were assigned to one of the possible classes based on geographic
204 occurrence data recorded in museum collections. In cases where precise latitude and longitude
information were not available we estimated for them from other locality information. Because
206 the exact barriers between different biogeographic regions are unknown and unclear, we
represented each hypothesis with multiple possible realizations representing the classification
208 uncertainty for specimens present at the geographic boundaries. The taxonomic hypotheses
and sub-hypotheses for *E. marmorata* used here are presented in Table 1 and Figure 1.

210 For Spinks et al. (2010) we used three binning schemes. All three schemes include a class

Table 1: Table of species delimitation hypotheses for *E. marmorata*

Abbreviation	Number of classes	citation
SP10.1	3	Spinks et al. (2010)
SP10.2	3	Spinks et al. (2010)
SP10.3	4	Spinks et al. (2010)
SP14.1	2	Spinks et al. (2014)
SP14.2	4	Spinks et al. (2014)
Morph	2	Spinks et al. (2010)

for *E. marmorata* specimens from northern populations (marm) as well as a class for those assigned to *E. pallida* (pall) and an intergrade zone in the Central Coast Ranges (CCR). The schemes differ in the assignment of samples from the San Joaquin Valley (Fig. 1). Scheme SP10.1 and SP10.2 differ in the assignment of specimens from the western San Joaquin Valley to either CCR or marm reflecting uncertainty regarding their genetic affinity as explained above. In scheme SP10.3 these specimens are assigned to a San Joaquin class reflecting the mitochondrial distinctiveness shown by Spinks and Shaffer (2005). For Spinks et al. (2014) we used two binning schemes with SP14.1 being based on their phylogenetic network analysis and SP14.2 being based on their Bayesian species delimitation analysis. The latter scheme requires the addition of two new classes, “Baja” and “Foothill,” to accommodate the genetic groupings recovered by the SNP Structure analysis that was used to create the guide tree for the BPP species delimitation analysis in Spinks et al. (2014). Finally, we proposed a conservative morphological hypothesis (“Morph”) in order to compare the molecular hypotheses with something approximating the original taxonomic hypothesis for the group; this scheme is made up solely of the marm and pall classes from the SP10.3 scheme.

Sex was known only for a subset of the total dataset and was not included as a predictor of classification. Instead, we estimated the degree by which specimens cluster morphologically by sex in order to determine how much of a potential biasing factor sexual dimorphism could be for our analysis of the *E. marmorata* species complex (see below).

Following previous work on plastron shape (Angielczyk and Sheets 2007; Angielczyk et al. 2011; Angielczyk and Feldman 2013), we used TpsDig 2.04 (Rohlf 2005) to digitize 19 two-dimensional landmarks (Fig. 2). Seventeen of the landmarks are at the endpoints or intersection of the keratinous plastral scutes that cover the plastron. Twelve of the landmarks were symmetrical across the axis of symmetry. Because damage prevented the digitization of all the symmetric landmarks in some specimens, we reflected landmarks across the axis of symmetry (i.e. midline) prior to analysis and used the average position of each symmetrical pair. In cases where damage or incompleteness prevented symmetric landmarks from being determined, we used only the single member of the pair. We conducted all subsequent analyses on the resulting “half” plastra. We superimposed the plastral landmark configurations using generalized Procrustes analysis (Dryden and Mardia 1998), after which we calculated the principal components (PC) of shape using the `shapes` package for R (R Core Team 2016;

Dryden 2013). All specimens were used for superimposition, after which the subset labeled for each of the schemes were used in model training and testing (see below).

Biasing effects

We estimated the possible effect of digitization error (Arnqvist and Mårtensson 1998; von Cramon-Taubadel et al. 2007; Munoz-Munoz F. and Perpinan D. 2010) on our results by comparing within (replicated) specimen Procrustes distances to the distances between classification scheme centroids. Ten randomly selected *E. marmorata* specimens were each digitized four times, with the original set of digitized coordinates serving as a fifth replicate. These 50 landmark configurations were then Procrustes superimposed. A range of four Procrustes distances was then calculated as the average of the pairwise distances between each of the replicate configurations of a given specimen.

For each specimen, the difference in shape caused by digitization was calculated as the mean of all pairwise Procrustes distances between the five replicates of that specimen. The average distance between any two digitizations was calculated as the mean of all pairwise Procrustes distances between all replicates for all specimens. The ratio between these two values was used to assess the magnitude of variation caused by digitization. The goal of this ratio is to determine if the within group distances are on average smaller than the between individual distances; a value of 0 indicates perfect grouping, a value of 1 indicates no difference between grouping and no grouping, and a value of 1+ indicates that the grouping is counter-intuitive to the data.

Emys marmorata is known to display sexual dimorphism in plastron shape, particularly the presence of a plastron concavity in males (Seeliger 1945). To test for biases resulting from sexual dimorphism in our *E. marmorata* dataset, we used a simple permutation test to determine if the distance between the mean female and male shapes is greater than expected when the sex labels are randomly shuffled. Because not all of our specimens have sex identifications associated with them, this analysis was done using a subset of the data (257 of 532).

Supervised learning approaches

Instead of relying on a single supervised learning method, we chose to use an ensemble approach where multiple model types are used in concert so that any congruence between them increases our support for that conclusion over another (Hastie et al. 2009). The supervised learning methods used here are named in Table 2. Each of these methods makes different assumptions, treats data differently, and can produce different classification results depending on the nature of the data (Hastie et al. 2009). For example, multinomial logistic regression is a type of generalized linear model while random forest is itself an ensemble approach where multiple decision trees are fit to subsets of the full data and then averaged.

The maximum set of possible predictors or features used for any model of our dataset is comprised of the first 25 principal components (PCs), scaled centroid size, and the interaction between scaled centroid size and PC 1. Additional interaction terms were not considered

Table 2: Table of the supervised learning methods used in this analysis.

Method name	abbreviation	R package	citation
multinomial logistic regression	MLR	nnet	Venables and Ripley (2002)
linear discriminate analysis	LDA	MASS	Venables and Ripley (2002)
penalized discriminate analysis	PDA	mda	Hastie et al. (2015)
single-hidden-layer neural network	NN	nnet	Venables and Ripley (2002)
random forests	RF	randomForest	Liaw and Wiener (2002)

because of model complexity/sample size concerns. Size and the interaction between size and PC 1 were included as predictors to account for known ontogenetic variation in plastron shape (Angielczyk and Feldman 2013) as well as potential size differences between classes, even if this is unlikely (Seeliger 1945; Holland 1992). These data constitute a “maximum set” because the best or selected models based on five-fold cross-validation need not, and likely will not, include all predictors possible (see below). Because our supervised learning models use PCs as predictors, this approach is in many ways analogous to PCA regression. PCA regression takes advantage of reduction and orthogonality PCs to improve regression fit (Hastie et al. 2009). Because the PCs of shape are by definition orthogonal, they can easily serve as independent predictors or features of class membership without fear of collinearity.

We adopted a training and testing paradigm for selecting parsimonious models and estimating their overall error rates (Hastie et al. 2009; Kuhn and Johnson 2013). Within-sample model performance is inherently biased upwards, so model evaluation requires overcoming this bias. With very large sample sizes, as in this study, part of the sample can be used as the “training set” and the remainder acts as the “testing set.” In this approach, following all cleaning and vetting, the data are split into a training dataset and a testing dataset. The former is used for fitting the model whereas the later is used for measuring model performance, a process called model generalization. For each scheme, we limited the model training and testing to only those individuals with class labels for that scheme. In this analysis, we randomly divided 80% of samples into the training set and the remaining 20% into the testing set.

In classification studies, such as this one, a common metric of performance is the receiver operating characteristic (ROC) which is the relationship between the false and true positive rates (Hastie et al. 2009). The area under the ROC curve (AUC) is the derived estimate of the model performance; AUC ranges from 0.5 to 1 which correspond to performance similar to random guesses and perfect classification rates, respectively (Hastie et al. 2009). Both ROC and AUC are preferable to simple classification accuracy when class membership is unbalanced, as it is in these analyses (Hastie et al. 2009). The standard ROC and AUC calculations are defined only for binary classifications, which is not the case for our seven species and *Emys* complex datasets. To generalize this approach for situations with multiple response classes, we used an all-against-one strategy where the model AUC is the average of the AUC values from the multiple binary comparisons of one class compared to all others (Hand and Till 2001).

For a given supervised learning method, we compared the fit of 27 models as the average AUC from 10 rounds of five-fold cross-validation. Cross-validation is an approach for estimating the average out-of-sample predictive error of a model by simulating out-of-sample data from the training dataset itself (Hastie et al. 2009). In a single round of k -fold cross-validation, the training data are divided into k blocks where the model is fit to $k - 1$ blocks and the values of the k th block are predicted. This is repeated for all combinations of blocks. Within each round, the predictive performance metrics are averaged across all folds. Finally, the predictive performance metric is the averaged across all rounds of k -fold cross-validation. This process was implemented using the R package `caret` (Kuhn 2013). For a given supervised learning method, the “best” trained model is that with the highest mean AUC as estimated from five-fold cross-validation. The selected or final model, however, is the next most parsimonious model that is within one standard error of the best model; this is a variant on the “one-standard error” rule from Hastie et al. (2009). The purpose of this rule is to ameliorate the chances of selecting an overly complex model that will perform poorly when predicting the classes of out-of-sample data.

RESULTS

Geometric morphometrics

The results of the PCA of plastron shape in both the seven species and *Trachemys* datasets demonstrate strong association between shape and the recognized classification schemes (Fig. 3).

The results of the PCA of plastron shape in the *Emys marmorata* dataset show no clear connection between plastron shape and any of the proposed classification schemes (Fig. 4). The first PC axis of shape variation appears to be primarily structured by differences in individual centroid size (Fig. 4); this was the motivation for including centroid size and its interaction with PC1 as predictors in all of the supervised learning models.

Analysis of the differences between sexes of *E. marmorata* indicates that sex does not appear to strongly structure differences in shape (Fig. 5). The difference in mean shape between the sexes is very small; the sexes overlap about as much as expected given a null distribution based on permuting the sex-labels.

Comparison of the within to between Procrustes distances of the digitization replicates gives an approximate estimate of the error between distinct groupings (Table 3). The ratio of the average within-individual distance to the average distance between individuals for the replicated datasets is 1.11; this indicates that the grouping is slightly counter-intuitive to the data and is consistent with all shapes being very similar regardless of individual identity. This value also provides a baseline by which to understand how distinct the groupings are, where other ratios are compared to the correction ratio 1.11/1.

The results from the seven species and *Trachemys* datasets indicate that both of these classification schemes are more recognizable than not given our estimate of digitization

Table 3: Results from the within-individual to between-individual Procrustes distances for the replicated plastron shape data. Results are presented for all three datasets analyzed here: the *Trachemys* dataset, the seven species dataset, and each of the *Emys marmorata* classification schemes.

Dataset	Scheme	Ratio	Corrected ratio
Replicates		1.11	
Seven species		0.33	0.37
<i>Trachemys</i>		0.76	0.84
<i>E. marmorata</i>	SP10.1	0.99	1.10
	SP10.2	1.00	1.11
	SP10.3	0.94	1.04
	SP14.1	1.01	1.12
	SP14.2	0.93	1.04
	Morph	0.99	1.09

error (Table 3). In contrast, the different *E. marmorata* classification schemes appear to be barely be distinct, with their within:between ratios approximating 1. This indicates that the magnitude of the differences between groupings is approximately the same as the difference as any two random individuals (Table 3).

Supervised learning

Analysis of the seven morphologically and genetically distinct species and the *T. scripta scripta*–*T. scripta elegans* datasets indicate that these classifications are sufficiently morphologically distinct to be differentiated on the basis of plastron shape. Both in-sample and out-of-sample classification have AUC values of approximately 1 for all methods, implying near-perfect classification rates (Fig. 6, 7). For both datasets, the ROC scores from testing datasets are tightly clustered near AUC = 1 (Fig. 7). These results demonstrate that when there are distinctions between the states of the classification schemes (i.e. differences in plastron shape that correlate with the different taxonomic groups), the methods used here can recover them.

AUC-based model selection revealed some important patterns of variation and congruence between the classification schemes and the actual data. Generally, the best performing models tended to include about half the total number of possible PCs (Fig. 8).

Observed AUC values for all of the optimal models are lower for the *E. marmorata* dataset than for the other two datasets (Fig. 6, 8). In most cases the different proposed classification schemes are generally poor descriptors of the observed variation. It appears that the dataset is overwhelmed by noise (likely biological and analytical), making any accurate classifications difficult at best. This observation is cemented with the generalizations of the models to the testing dataset (Fig. 9).

Mean AUC values for the model generalizations, in most cases, are approximately equal to the

observed AUC values from the training dataset (Fig. 8, 9). The cases in which the AUC from the generalizations is less than the observed indicate poor model fit and a poor classification scheme. Comparison of AUC values from the model generalizations do not indicate a clear “best” classification scheme (Fig. 8, 9). Only in the case of the conservative morphological hypothesis (“Morph”) is the mean AUC value potentially distinct from that of other schemes; in this case mean AUC is lower than the average of the other five schemes which indicates that the morphologically-based scheme performs more poorly than the molecularly-based ones. It is important to note, however, that the training and testing dataset for the “Morph” scheme is the smallest of the six schemes which may lead to poorer performance in in-sample and out-of-sample comparisons.

DISCUSSION

As expected, our ensemble approach yields high out-of-sample classification performance for the first two datasets. These results indicate that in cases of clear class separation (Fig. 3) our approach is able to detect this and make good out-of-sample prediction.

In the case of the *E. marmorata* dataset, our results show that none of the proposed taxonomic hypotheses for the *E. marmorata* species complex are more consistent with morphological differentiation than any other proposal (Fig. 9). Both the low out-of-sample AUC values and the significant difference between the correctly and incorrectly classified observations support the conclusion that none of the hypothesized classification schemes are good descriptions of the observed plastral variation within *E. marmorata*. An analytical explanation of this result is that the level of digitization error in the *E. marmorata* dataset is so great as to swamp out any biological signal. We think this is unlikely because all of the specimens considered in our three analyses were digitized by one of us (K.D.A.), and digitization error was not a problem in the seven species or *Trachemys* examples. There are also no features of the plastron of *E. marmorata* that would make it significantly more difficult to accurately digitize than the plastra of the other species.

Biological explanations include the possibility that genetic differentiation is not associated with plastron shape variation and/or that local selective pressures (e.g. from hydrological regime) overwhelm morphological differentiation. Both of these options seem plausible given that shell shape is influenced by selection for both protection and streamlining, but not necessary mate choice (Rivera 2008; Rivera and Stayton 2011; Stayton 2011; Rivera et al. 2014; Polly et al. 2016) and that shell shape in *E. marmorata* is known to vary among populations inhabiting water bodies with different flow regimes (Holland 1992; Lubcke and Wilson 2007; Germano and Bury 2009). Plastron shape does not seem to preserve a strong phylogenetic signal at the interspecific level in emydine turtles, at least compared to the effect of the presence or absence of a plastral hinge (Angielczyk et al. 2011), and our current results suggest that this may be the case for phylogeographic signal within emydine species as well. A final possibility (explored below) is that the proposed classification schemes themselves do not represent significant evolutionary lineages.

Despite the negative result for *E. marmorata*, it is important to note that plastron shape is an extremely effective method for differentiating classes in the additional datasets we investigated. The magnitude of shape differences between the species (measured as Procrustes distance between the seven species' mean shapes) is approximately an order of magnitude greater than the differences between the *E. marmorata* subgroups, and not surprisingly the machine learning methods had no trouble classifying the specimens correctly. However, the magnitude of the shape differences between the *T. scripta* subspecies is comparable to those separating the different *E. marmorata* subgroups, yet even in this case the machine learning methods returned an almost perfect classification. These results demonstrate that plastron shape is normally a good marker for differentiating real subgroups in close relatives of *E. marmorata*, and that our lack of results for *E. marmorata* is not simply a shortcoming of the methods we applied. Indeed, it begs the question of what factors have suppressed morphological differentiation of plastron shape in *E. marmorata* and *E. pallida* if they are distinct species. Invoking issues such as the role of the plastron in protection or the need for streamlining are insufficient because the other species are expected to be subject to similar constraints (Stayton 2011; Polly et al. 2016). Although it may seem counterintuitive that plastron shape is both useful for species delimitation but has weak or absent phylogenetic signal, it is important to remember that these are different goals. While phylogenetically similar species may not be morphologically similar (e.g. compare the box turtles of the genus *Terrapene* to the closely related spotted turtle *Clemmys guttata*), the variation within a species typically is much less than the variation between species. Therefore, the consistent plastron shapes that characterize different emydid species leads to plastron shape being a useful tool for species delimitation, even when other selective factors have overprinted similarities stemming from patterns of descent from common ancestors.

Is there more than one species of Western Pond Turtle?

The lack of morphological support for the distinctiveness of *E. pallida* does not, on its own, preclude the recognition of this taxon. However, this apparent lack of congruence does prompt a reexamination of the methods and concepts that led to that taxonomic revision, especially considering that plastron shape is demonstrably capable of differentiating species and subspecies among other emydids. In other words, before we can assess the significance of the morphological non-diagnosability, it is essential to evaluate the methods and concepts that led to the initial taxonomic revision.

Spinks et al. (2014) elevated *E. pallida* based on a species delimitation analysis of SNP data using BPP (Yang and Rannala 2010). However, Spinks et al. (2014) did not heed the caveats about such species delimitation methods raised by Carstens et al. (2013). In addition to specifically addressing the shortcomings of validation methods such as BPP that rely on guide trees and “should be interpreted with caution,” Carstens et al. (2013) also strongly emphasized that “Inferences regarding species boundaries based on genetic data alone are likely inadequate, and species delimitation should be conducted with consideration of the life history, geographical distribution, morphology and behaviour (where applicable) of the focal

system. . .” These caveats evoke the development of the Unified Species Concept (Dayrat 2005; De Queiroz 2007), Integrative Taxonomy (Padial et al. 2010), and other pluralist approaches to species delimitation. None of these considerations were brought to bear on the *E. marmorata* system until now, and in doing so we find the proposal that *E. pallida* is a distinct species to be lacking in a normally robust morphological marker.

The natural history and geographical distribution of *E. marmorata* and *E. pallida* also make the recognition of these taxa implausible. The data from Spinks et al. (2014) show extensive introgression and admixture in Central California, which is expected because there are no significant barriers to gene flow in this region. Combined with the well-demonstrated ability for testudinoid turtles, including emydids and even *Emys*, to hybridize (e.g. Buskirk et al. 2005; Spinks and Shaffer 2009; Parham et al. 2013) it is hard to imagine how *E. marmorata* and *E. pallida* could maintain their integrity in the face of such admixture. Because the geography, natural history, demonstrated genetic admixture of *E. marmorata*, and comparisons with other morphologically diagnosable species and subspecies conflict with the recognition of *E. pallida*, we hypothesize that our inability to classify the morphological data by proposed species is because *E. pallida* is not a distinct species.

We fully agree with Spinks et al. (2014) that *E. marmorata* (*sensu lato*) is a species deserving of strong conservation efforts, and we do not wish to trivialize this need. Moreover, the genetic diversity uncovered by the analysis of Spinks et al. (2014) should be accounted for explicitly in any conservation plan. Given the apparent lack of morphological distinction combined with the broad range of intergradation and other problems with the species hypothesis outlined above, we recommend that the populations elevated to *E. pallida* by Spinks et al. (2014) are best considered Evolutionary Significant Units or Distinct or Population Segments instead of distinct species.

Finally, it is important to note that the data and analyses we present do not let us definitively say whether the apparent lack of morphological divergence within *E. marmorata* truly reflects the presence of a single species, or if it is an artifact of plastron shape being a poor morphological marker for phylogenetic and phylogeographic divergences in the case of *E. marmorata*. This is because we could not carry out our morphometric analyses on the specimens from which the genetic data were obtained. The comparisons with the other emydid taxa suggest that our negative result is because *E. marmorata* is a single species. However, tests of both our preferred conclusion (*E. marmorata* as a single species) and that of Spinks et al. (2014) should include morphological and molecular analyses of the same set of voucher specimens, as well as additional tests of species delimitation using alternative methods and corroborating evidence as suggested by Carstens et al. (2013). From a morphological standpoint, support for the validity of “*E. pallida*” may come from other aspects of morphology, such as carapace shape or other features. Likewise, further investigation of the phylogeographic utility of plastron shape in other turtle species will help to clarify whether the lack of differentiation seen in *E. marmorata*, and the strong differentiation among the other emydids, is typical or an unusual case.

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BIBLIOGRAPHY

- K. D. Angielczyk and C. R. Feldman. Are diminutive turtles miniaturized? The ontogeny of plastron shape in emydine turtles. *Biological Journal of the Linnean Society*, 108(4): 727–755, apr 2013. ISSN 00244066. doi: 10.1111/bij.12010. URL <http://doi.wiley.com/10.1111/bij.12010>.
- K. D. Angielczyk and H. D. Sheets. Investigation of simulated tectonic deformation in fossils using geometric morphometrics. *Paleobiology*, 33(1):125–148, 2007.
- K. D. Angielczyk, C. R. Feldman, and G. R. Miller. Adaptive evolution of plastron shape in emydine turtles. *Evolution*, 65(2):377–394, feb 2011. ISSN 1558-5646. doi: 10.1111/j.1558-5646.2010.01118.x.
- G. Arnqvist and T. Mårtensson. Measurement error in geometric morphometrics: Empirical strategies to assess and reduce its impact on measures of shape, 1998. ISSN 12178837.
- S. F. Baird and C. Girard. Descriptions of new species of reptiles collected by the U.S. Exploring Expedition under the command of Capt. Charles Wilkes. *Proceedings of the National Academy of Sciences Philadelphia*, 6:174–177, 1852.
- A. M. Bauer, J. F. Parham, R. M. Brown, B. L. Stuart, L. Grismer, T. J. Papenfuss, W. Bohme, J. M. Savage, S. Carranza, J. L. Grismer, P. Wagner, A. Schmitz, N. B. Ananjeva, and R. F. Inger. Availability of new Bayesian-delimited gecko names and the importance of character-based species descriptions. *Proceedings of the Royal Society B: Biological Sciences*, 278:490–492, 2000. ISSN 07331347. doi: 10.2307/1467045. URL <http://www.jstor.org/stable/1467045?origin=crossref>.
- M. Baylac, C. Villemant, and G. Simbolotti. Combining geometric morphometrics with pattern recognition for the investigation of species complexes. *Biological Journal of the Linnean Society*, 80:89–98, 2003.
- D. Bickford, D. J. Lohman, N. S. Sodhi, P. K. L. Ng, R. Meier, K. Winker, K. K. Ingram, and I. Das. Cryptic species as a window on diversity and conservation. *Trends in ecology & evolution*, 22(3):148–55, mar 2007. ISSN 0169-5347. doi: 10.1016/j.tree.2006.11.004. URL <http://www.ncbi.nlm.nih.gov/pubmed/17129636>.

- 528 L. Breiman, J. Friedman, C. J. Stone, and R. A. Olshen. *Classification and regression trees*.
Wadsworth International Group, Belmont, 1984.
- 530 R. B. Bury, D. J. Germano, and G. W. Bury. Population Structure and Growth of the
Turtle *Actinemys marmorata* from the KlamathSiskiyou Ecoregion: Age, Not Size, Matters.
532 *Copeia*, 2010(3):443–451, sep 2010. ISSN 0045-8511. doi: 10.1643/CH-08-096. URL
<http://www.bioone.org/doi/abs/10.1643/CH-08-096>.
- 534 S. W. Buskirk, J. F. Parham, and C. R. Feldman. On the hybridisation between two distantly
related Asian turtles (Testudines: *Scalia* x *Mauremys*). *Salamandra*, 41:21–26, 2005.
- 536 A. Cardini, D. Nagorsen, P. O’Higgins, P. D. Polly, R. W. Thorington Jr, and P. Tongiorgi.
Detecting biological distinctiveness using geometric morphometrics: an example case from
538 the Vancouver Island marmot. *Ethology Ecology & Evolution*, 21:209–223, 2009.
- B. C. Carstens and T. A. Dewey. Species Delimitation Using a Combined Coalescent
540 and Information-Theoretic Approach: An Example from North American *Myotis* Bats.
Systematic Biology, 59(4):400–414, 2010. URL [papers2://publication/doi/10.1093/](http://pubs2://publication/doi/10.1093/sysbio/syq024)
542 [sysbio/syq024](http://pubs2://publication/doi/10.1093/sysbio/syq024).
- B. C. Carstens, T. a. Pelletier, N. M. Reid, and J. D. Satler. How to fail at species delimitation.
544 *Molecular ecology*, 22(17):4369–83, sep 2013. ISSN 1365-294X. doi: 10.1111/mec.12413.
URL <http://www.ncbi.nlm.nih.gov/pubmed/23855767>.
- 546 R. Caumul and P. D. Polly. Phylogenetic and environmental components of morphological
variation: skull, mandible, and molar shape in marmots (*Marmota*, Rodentia). *Evolution;*
548 *international journal of organic evolution*, 59(11):2460–72, nov 2005. ISSN 0014-3820. URL
<http://www.ncbi.nlm.nih.gov/pubmed/16396186>.
- 550 E. L. Clare. Cryptic species? Patterns of maternal and paternal gene flow in eight neotropical
bats. *PloS one*, 6(7):e21460, jan 2011. ISSN 1932-6203. doi: 10.1371/journal.pone.0021460.
- 552 B. Dayrat. Towards integrative taxonomy. *Biological Journal of the Linnean Society*, 85:
407–415, 2005.
- 554 K. De Queiroz. Species concepts and species delimitation. *Systematic Biology*, 56(6):879–86,
dec 2007. ISSN 1063-5157. doi: 10.1080/10635150701701083. URL [http://www.ncbi.nlm.](http://www.ncbi.nlm.nih.gov/pubmed/18027281)
556 [nih.gov/pubmed/18027281](http://www.ncbi.nlm.nih.gov/pubmed/18027281).
- M. H. Demandt and S. Bergek. Identification of cyprinid hybrids by using geometric
558 morphometrics and microsatellites. *Journal of Applied Ichthyology*, 25(6):695–701, dec
2009. ISSN 01758659. doi: 10.1111/j.1439-0426.2009.01329.x. URL [http://doi.wiley.](http://doi.wiley.com/10.1111/j.1439-0426.2009.01329.x)
560 [com/10.1111/j.1439-0426.2009.01329.x](http://doi.wiley.com/10.1111/j.1439-0426.2009.01329.x).
- K. C. Dillard. *A comparative analysis of geometric morphometrics across two *Pseudemys**
562 *turtle species in east central Virginia*. Masters, Virginia Commonwealth University, 2017.

G. Dobigny, L. Granjon, V. Aniskin, K. Ba, and V. Voloboulev. A new sigling species of
Taterillus (Muridae, Gerbillinae) from West Agrica. *Mammalian Biology*, 68:299–316, 2003.

I. L. Dryden. *shapes: Statistical shape analysis*, 2013. URL <http://CRAN.R-project.org/package=shapes>. R package version 1.1-8.

I. L. Dryden and K. Y. Mardia. *Statistical shape analysis*. Wiley, New York, 1998.

N. Eldredge and S. J. Gould. Punctuated equilibria: an alternative to phyletic gradualism. In T. J. M. Schopf, editor, *Models in Paleobiology*, pages 82–115. Freeman Cooper, San Francisco, 1972.

C. R. Feldman and J. F. Parham. Molecular phylogenetics of emydine turtles: taxonomic revision and the evolution of shell kinesis. *Molecular Phylogenetics and Evolution*, 22(3): 388–98, mar 2002. ISSN 1055-7903. doi: 10.1006/mpev.2001.1070. URL <http://www.ncbi.nlm.nih.gov/pubmed/11884163>.

T. M. Francoy, R. A. O. Silva, P. Nunes-Silva, C. Menezes, and V. L. Imperatriz-Fonseca. Gender identification of five genera of stingless bees (Apidae, Meliponini) based on wing morphology. *Genetics and molecular research*, 8(1):207–214, 2009.

C. Fruciano, P. Franchini, F. Raffini, S. Fan, and A. Meyer. Are sympatrically speciating Midas cichlid fish special? Patterns of morphological and genetic variation in the closely related species *Archocentrus centrarchus*. *Ecology and Evolution*, 6(12):4102–4114, 2016. ISSN 20457758. doi: 10.1002/ece3.2184.

W. C. Funk, M. Caminer, and S. R. Ron. High levels of cryptic species diversity uncovered in Amazonian frogs. *Proceedings of the Royal Society B: Biological Sciences*, 279(1734): 1806–14, may 2012. ISSN 1471-2954. doi: 10.1098/rspb.2011.1653.

P. Gaubert, P. J. Taylor, C. a. Fernandes, M. W. Bruford, and G. Veron. Patterns of cryptic hybridization revealed using an integrative approach: a case study on genetids (Carnivora, Viverridae, Genetta spp.) from the southern African subregion. *Biological Journal of the Linnean Society*, 86(1):11–33, aug 2005. ISSN 00244066. doi: 10.1111/j.1095-8312.2005.00518.x. URL <http://doi.wiley.com/10.1111/j.1095-8312.2005.00518.x>.

D. J. Germano and R. B. Bury. Variation in body size, growth, and population structure of *Actinemys marmorata* from lentic and lotic habitats in Southern Oregon. *Journal of Herpetology*, 43(3):510–520, 2009.

D. J. Germano and G. B. Rathbun. Growth, population structure, and reproduction of western pond turtles (*Actinemys marmorata*) on the Central Coast of California. *Chelonian Conservation and Biology*, 7(2):188–194, 2008.

S. J. Gould and N. Eldredge. Punctuated equilibria: the tempo and mode of evolution reconsidered. *Paleobiology*, 3(2):115–151, 1977.

I. Gündüz, M. Jaarola, C. Tez, C. Yeniyurt, P. D. Polly, and J. B. Searle. Multigenic and

morphometric differentiation of ground squirrels (*Spermophilus*, Scuridae, Rodentia) in Turkey, with a description of a new species. *Molecular phylogenetics and evolution*, 43(3):916–35, jun 2007. ISSN 1055-7903. doi: 10.1016/j.ympev.2007.02.021. URL <http://www.ncbi.nlm.nih.gov/pubmed/17500011>.

D. J. Hand and R. J. Till. A Simple Generalisation of the Area Under the ROC Curve for Multiple Class Classification Problems. *Machine Learning*, 45:171–186, 2001.

T. Hastie, R. Tibshirani, and J. Friedman. *The elements of statistical learning: data mining, inference, and prediction*. Springer, New York, 2nd edition, 2009.

T. Hastie, R. Tibshirani, F. Leisch, K. Hornik, and B. D. Ripley. *mda: Mixture and Flexible Discriminant Analysis*, 2015. URL <https://CRAN.R-project.org/package=mda>. R package version 0.4-8.

B. Hausdorf and C. Hennig. Species delimitation using dominant and codominant multilocus markers. *Systematic biology*, 59(5):491–503, oct 2010. ISSN 1076-836X. doi: 10.1093/sysbio/syq039. URL <http://www.ncbi.nlm.nih.gov/pubmed/20693311>.

D. C. Holland. *Level and pattern in morphological variation: a phylogeographic study of the western pond turtle (Clemmys marmorata)*. PhD thesis, University of Southwestern Louisiana, 1992.

J. P. Huelsenbeck, P. Andolfatto, and E. T. Huelsenbeck. Structurama: bayesian inference of population structure. *Evolutionary bioinformatics online*, 7:55–9, jan 2011. ISSN 1176-9343. doi: 10.4137/EBO.S6761. URL <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3118697&tool=pmcentrez&rendertype=abstract>.

L. Kaufman and P. J. Rousseeuw. *Finding groups in data : an introduction to cluster analysis*. Wiley, New York, 1990.

D. G. Kendall. The diffusion of shape. *Advances in Applied Probability*, 9(3):428–430, 1977.

M. Kuhn. *caret: Classification and Regression Training*, 2013. URL <http://CRAN.R-project.org/package=caret>. R package version 5.15-61.

M. Kuhn and K. Johnson. *Applied predictive modeling*. Springer, New York, NY, 2013.

A. D. Leaché and M. K. Fujita. Bayesian species delimitation in West African forest geckos (*Hemidactylus fasciatus*). *Proceedings. Biological sciences / The Royal Society*, 277(1697):3071–7, oct 2010. ISSN 1471-2954. doi: 10.1098/rspb.2010.0662. URL <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=2982061&tool=pmcentrez&rendertype=abstract>.

A. Liaw and M. Wiener. Classification and regression by randomforest. *R News*, 2(3):18–22, 2002. URL <http://CRAN.R-project.org/doc/Rnews/>.

G. M. Lubcke and D. S. Wilson. Variation in shell morphology of the Western Pond

634 Turtle (*Actinemys marmorata* Baird and Giarard) from three aquatic habitats in Northern California. *Journal of Herpetology*, 41(1):107–114, 2007.

636 N. MacLeod. *Automated taxon identification in systematics: theory, approaches and applications*. CRC Press, Boca Raton, 2007.

638 M. Markolf, H. Rakotonirina, C. Fichtel, P. von Grumbkow, M. Brameier, and P. M. Kappeler. True lemurs... true species - species delimitation using multiple data sources in the brown
640 lemur complex. *BMC Evolutionary Biology*, 13(1):233, 2013. ISSN 1471-2148. doi:
10.1186/1471-2148-13-233. URL [http://bmcevolbiol.biomedcentral.com/articles/](http://bmcevolbiol.biomedcentral.com/articles/10.1186/1471-2148-13-233)
642 10.1186/1471-2148-13-233.

A. Mitrovski-Bogdanovic, A. Petrovic, M. Mitrovic, A. Ivanovic, V. Žikic, P. Starý, C. Vorburger, and Ž. Tomanovic. Identification of two cryptic species within the *Praon abjectum* group (Hymenoptera: Braconidae: Aphidiinae) using molecular markers and geometric
644 morphometrics. *Annals of the entomological society of America*, 106(2):170–180, 2013.

Munoz-Munoz F. and Perpignan D. Measurement error in morphometric studies: comparison
648 between manual and computerized methods. *Ann. Zool.*, 47(1):46–56, 2010.

D. Navega, R. Vicente, D. N. Vieira, A. H. Ross, and E. Cunha. Sex estimation from the
650 tarsal bones in a Portuguese sample: a machine learning approach. *International Journal of Legal Medicine*, 129(3):651–659, 2015. ISSN 14371596. doi: 10.1007/s00414-014-1070-5.

652 B. C. O’Meara. New heuristic methods for joint species delimitation and species tree inference. *Systematic biology*, 59(1):59–73, jan 2010. ISSN 1076-836X. doi: 10.1093/sysbio/syp077.
654 URL <http://www.ncbi.nlm.nih.gov/pubmed/20525620>.

J. M. Padiá, A. Miralles, I. De la Riva, and M. Vences. The integrative future of taxonomy.
656 *Frontiers in Zoology*, 7(16):1–14, 2010.

J. F. Parham, T. J. Papenfuss, P. P. V. Dijk, B. S. Wilson, C. Marte, L. R. Schettino, and
658 W. Brian Simison. Genetic introgression and hybridization in Antillean freshwater turtles (*Trachemys*) revealed by coalescent analyses of mitochondrial and cloned nuclear markers.
660 *Molecular phylogenetics and evolution*, 67(1):176–87, apr 2013. ISSN 1095-9513. doi: 10.1016/j.ympev.2013.01.004. URL <http://www.ncbi.nlm.nih.gov/pubmed/23353072>.

662 M. Pfenninger and K. Schwenk. Cryptic animal species are homogeneously distributed among taxa and biogeographical regions. *BMC evolutionary biology*, 7:121, jan 2007. ISSN
664 1471-2148. doi: 10.1186/1471-2148-7-121. URL <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1939701&tool=pmcentrez&rendertype=abstract>.

666 P. D. Polly. Paleophylogeography of *Sorex araneus*: molar shape as a morphological marker for fossil shrews. *Mammalia*, 68(2):233–243, 2003.

668 P. D. Polly. Phylogeographic differentiation in *Sorex araneus*: morphology in relation to geography and karyotype. *Russian Journal of Theriology*, 6(1):73–84, 2007.

- P. D. Polly, C. T. Stayton, E. R. Dumont, S. E. Pierce, E. J. Rayfield, and K. D. Angielczyk. Combining geometric morphometrics and finite element analysis with evolutionary modeling: towards a synthesis. *Journal of Vertebrate Paleontology*, 4634(March), 2016. ISSN 0272-4634. doi: 10.1080/02724634.2016.1111225.
- J. Pons, T. Barraclough, J. Gomez-Zurita, A. Cardoso, D. Duran, S. Hazell, S. Kamoun, W. Sumlin, and A. Vogler. Sequence-Based Species Delimitation for the DNA Taxonomy of Undescribed Insects. *Systematic Biology*, 55(4):595–609, aug 2006. ISSN 1063-5157. doi: 10.1080/10635150600852011. URL <http://sysbio.oxfordjournals.org/cgi/doi/10.1080/10635150600852011>.
- R Core Team. *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria, 2016. URL <http://www.R-project.org/>.
- G. Rivera. Ecomorphological variation in shell shape of the freshwater turtle *Pseudemys concinna* inhabiting different aquatic flow regimes. *Integrative and comparative biology*, 48(6):769–87, dec 2008. ISSN 1540-7063. doi: 10.1093/icb/icn088. URL <http://www.ncbi.nlm.nih.gov/pubmed/21669831>.
- G. Rivera and C. T. Stayton. Finite element modeling of shell shape in the freshwater turtle *Pseudemys concinna* reveals a trade-off between mechanical strength and hydrodynamic efficiency. *Journal of morphology*, 272(10):1192–203, oct 2011. ISSN 1097-4687. doi: 10.1002/jmor.10974. URL <http://www.ncbi.nlm.nih.gov/pubmed/21630321>.
- G. Rivera, J. N. Davis, J. C. Godwin, and D. C. Adams. Repeatability of Habitat-Associated Divergence in Shell Shape of Turtles. *Evolutionary Biology*, pages 29–37, jul 2014. ISSN 0071-3260. doi: 10.1007/s11692-013-9243-6. URL <http://link.springer.com/10.1007/s11692-013-9243-6>.
- F. J. Rohlf. TpsDig 2.04, 2005.
- B. C. Schilck-Steiner, B. Seifert, C. Stauffer, E. Christian, R. H. Crozier, and F. M. Steiner. Without morphology, cryptic species stay in taxonomic crypsis following discovery. *Trends in ecology & evolution*, 22(8):391–392, aug 2007. ISSN 0169-5347. doi: 10.1016/j.tree.2007.05.003. URL <http://www.ncbi.nlm.nih.gov/pubmed/17573147>.
- L. M. Seeliger. Variation in the Pacific Mud Turtle. *Copeia*, 1945(3):150–159, 1945.
- P. Q. Spinks and H. B. Shaffer. Range-wide molecular analysis of the western pond turtle (*Emys marmorata*): cryptic variation, isolation by distance, and their conservation implications. *Molecular ecology*, 14(7):2047–64, jun 2005. ISSN 0962-1083. doi: 10.1111/j.1365-294X.2005.02564.x. URL <http://www.ncbi.nlm.nih.gov/pubmed/15910326>.
- P. Q. Spinks and H. B. Shaffer. Conflicting mitochondrial and nuclear phylogenies for the widely disjunct *Emys* (Testudines: Emydidae) species complex, and what they tell us about biogeography and hybridization. *Systematic biology*, 58(1):1–20, feb 2009. ISSN 1076-836X. doi: 10.1093/sysbio/syp005. URL <http://www.ncbi.nlm.nih.gov/pubmed/20525565>.

- P. Q. Spinks, R. C. Thomson, and H. B. Shaffer. Nuclear gene phylogeography reveals the historical legacy of an ancient inland sea on lineages of the western pond turtle, *Emys marmorata* in California. *Molecular ecology*, 19(3):542–56, feb 2010. ISSN 1365-294X. doi: 10.1111/j.1365-294X.2009.04451.x. URL <http://www.ncbi.nlm.nih.gov/pubmed/20051011>.
- P. Q. Spinks, R. C. Thomson, and H. Bradley Shaffer. The advantages of going large: genome wide SNPs clarify the complex population history and systematics of the threatened western pond turtle. *Molecular Ecology*, pages n/a–n/a, mar 2014. ISSN 09621083. doi: 10.1111/mec.12736. URL <http://doi.wiley.com/10.1111/mec.12736>.
- C. T. Stayton. Biomechanics on the half shell: functional performance influences patterns of morphological variation in the emydid turtle carapace. *Zoology (Jena, Germany)*, 114(4):213–23, sep 2011. ISSN 1873-2720. doi: 10.1016/j.zool.2011.03.002. URL <http://www.ncbi.nlm.nih.gov/pubmed/21820295>.
- B. L. Stuart, R. F. Inger, and H. K. Voris. High level of cryptic species diversity revealed by sympatric lineages of Southeast Asian forest frogs. *Biology letters*, 2(3):470–4, sep 2006. ISSN 1744-9561. doi: 10.1098/rsbl.2006.0505. URL <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1686201&tool=pmcentrez&rendertype=abstract>.
- A. Sztencel-Jablonka, G. Jones, and W. Bogdanowicz. Skull Morphology of Two Cryptic Bat Species: *Pipistrellus pipistrellus* and *P. pygmaeus* A 3D Geometric Morphometrics Approach with Landmark Reconstruction. *Acta Chiropterologica*, 11(1):113–126, jun 2009. ISSN 1508-1109. doi: 10.3161/150811009X465730. URL <http://www.bioone.org/doi/abs/10.3161/150811009X465730>.
- B. Van Bocxlaer and G. Hunt. Morphological stasis in an ongoing gastropod radiation from Lake Malawi. *Proceedings of the National Academy of Sciences*, aug 2013. ISSN 0027-8424. doi: 10.1073/pnas.1308588110. URL <http://www.pnas.org/cgi/doi/10.1073/pnas.1308588110>.
- B. Van Bocxlaer and R. Schultheiß. Comparison of morphometric techniques for shapes with few homologous landmarks based on machine-learning approaches to biological discrimination. *Paleobiology*, 36(3):497–515, 2010.
- V. van den Brink and F. Bokma. Morphometric shape analysis using learning vector quantization neural networks an example distinguishing two microtine vole species. *Annales Zoologici Fennici*, 48:359–364, 2011.
- W. N. Venables and B. D. Ripley. *Modern Applied Statistics with S*. Springer, New York, fourth edition, 2002. URL <http://www.stats.ox.ac.uk/pub/MASS4>.
- N. S. Vitek, C. L. Manz, T. Gao, J. I. Bloch, S. G. Strait, and D. M. Boyer. Semi-supervised determination of pseudocryptic morphotypes using observer-free characterizations of anatomical alignment and shape. *Ecology and Evolution*, 7(14):5041–5055, 2017. ISSN 20457758. doi: 10.1002/ece3.3058. URL <http://doi.wiley.com/10.1002/ece3.3058>.

- 746 N. von Cramon-Taubadel, B. C. Frazier, and M. M. Lahr. The problem of assessing landmark
error in geometric morphometrics: theory, methods, and modifications. *American journal
of physical anthropology*, 132(4):535–544, 2007. ISSN 00029483. doi: 10.1002/ajpa.
- 748 Z. Yang and B. Rannala. Bayesian species delimitation using multilocus sequence data.
Proceedings of the National Academy of Sciences, 107(20):9264–9, may 2010. ISSN
750 1091-6490. doi: 10.1073/pnas.0913022107. URL [http://www.pubmedcentral.nih.gov/
articlerender.fcgi?artid=2889046&tool=pmcentrez&rendertype=abstract](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=2889046&tool=pmcentrez&rendertype=abstract).
- 752 M. L. Zelditch, D. L. Swiderski, and H. D. Sheets. *Geometric morphometrics for biologists: a
primer*. Elsevier Academic Press, Amsterdam, 2004.

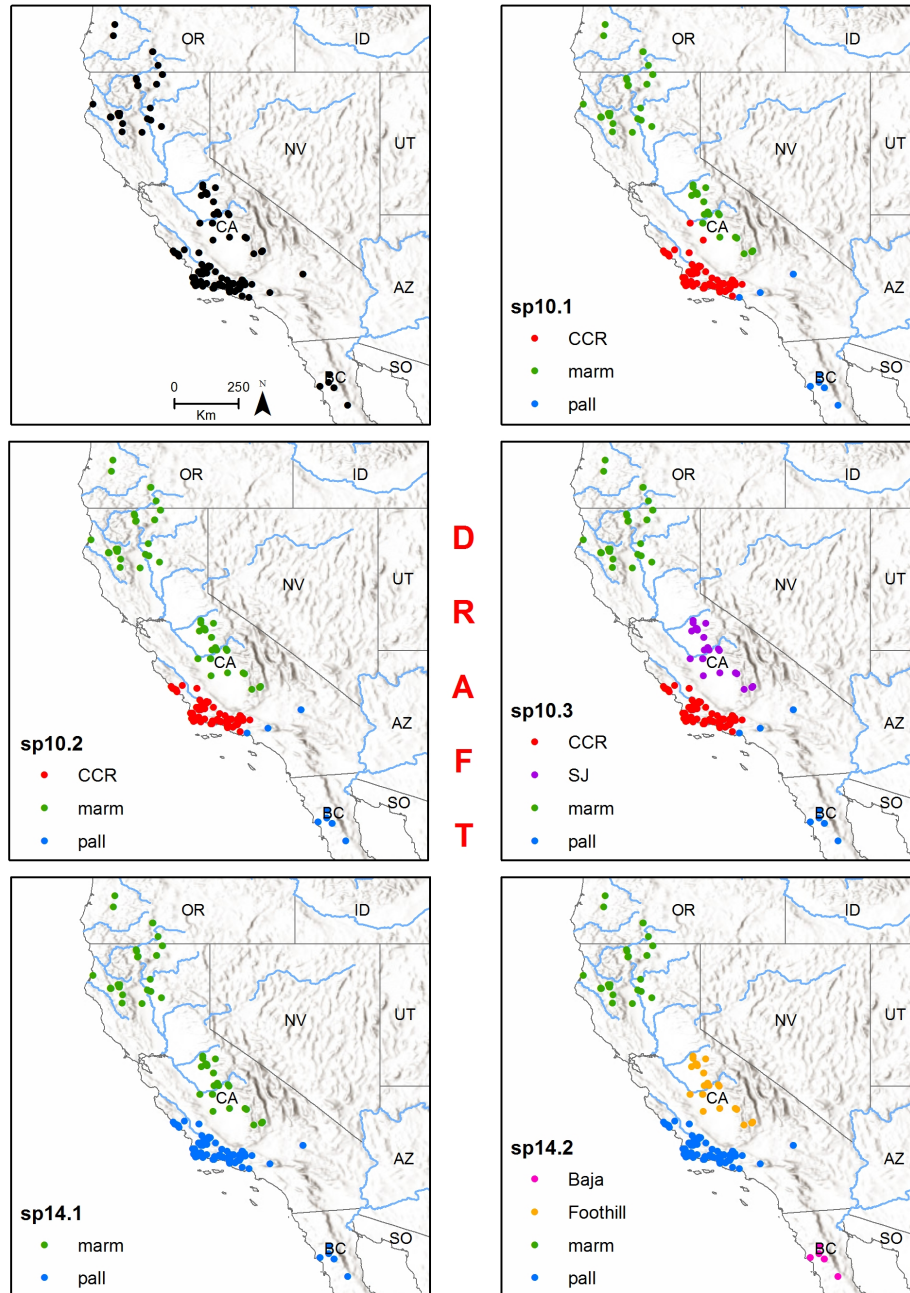


Figure 1: Geographic distribution of specimens sampled for comparing the hypothesized subdivisions of *Emys marmorata*. Each hypothesized scheme has two or more possible classes. Sample size differs between schemes because of our ability to confidently assign museum specimens to the various schemes.

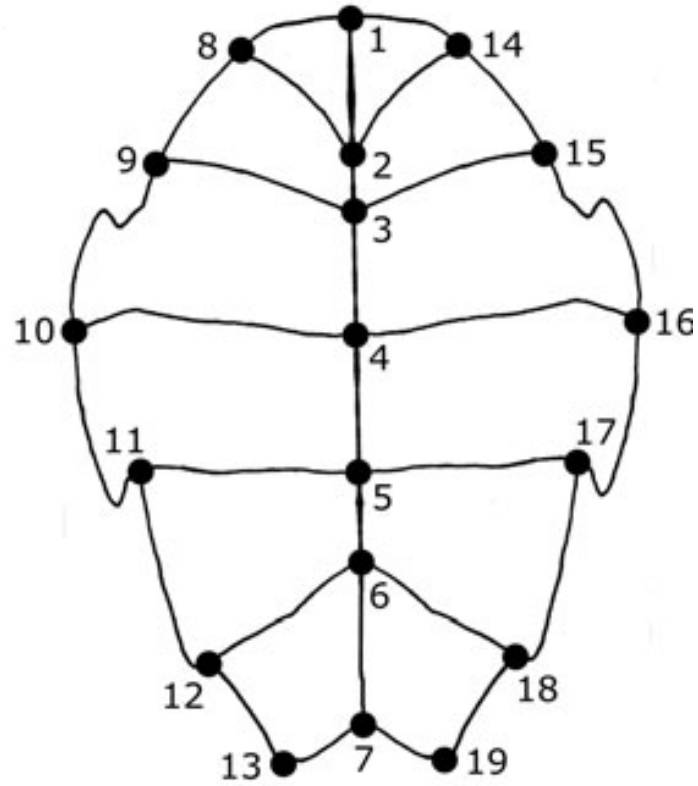
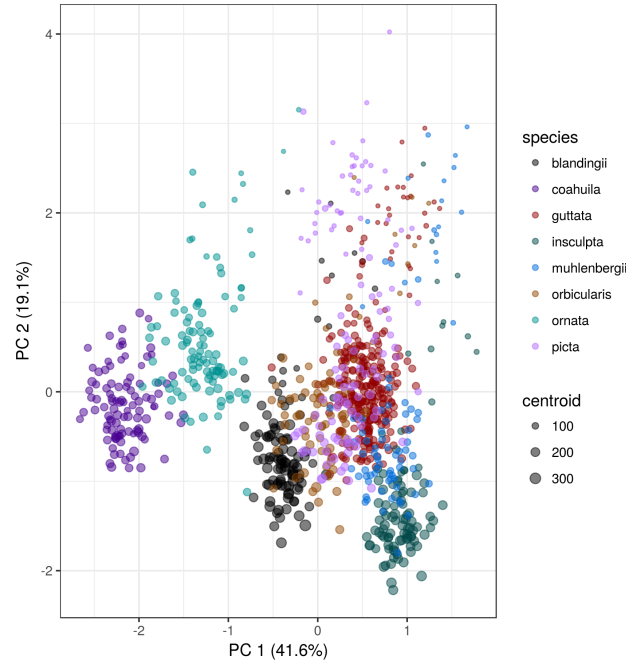


Figure 2: Depiction of general plastral shape of *E. marmorata* and position of the 19 landmarks used in this study. Anterior is towards the top of the figure.

(a)



(b)

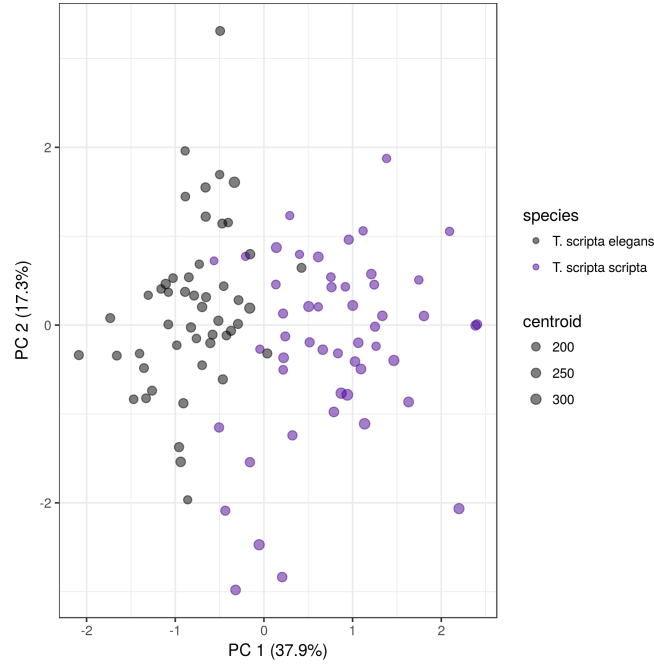


Figure 3: Two scatterplots of morphological differences from two of the three datasets analyzed in this study. (a) Scatterplot of the first two PCA axes from the landmarks from the seven different species dataset, and (b) the first two axes of variation from two subspecies of *Trachemys* dataset. Point colors correspond to the categories within each dataset while point size is proportional to individual centroid size. In parentheses next to the axis labels are the percent of total variation accounted for by that axis. For both datasets there are clear distinctions between the different categories.

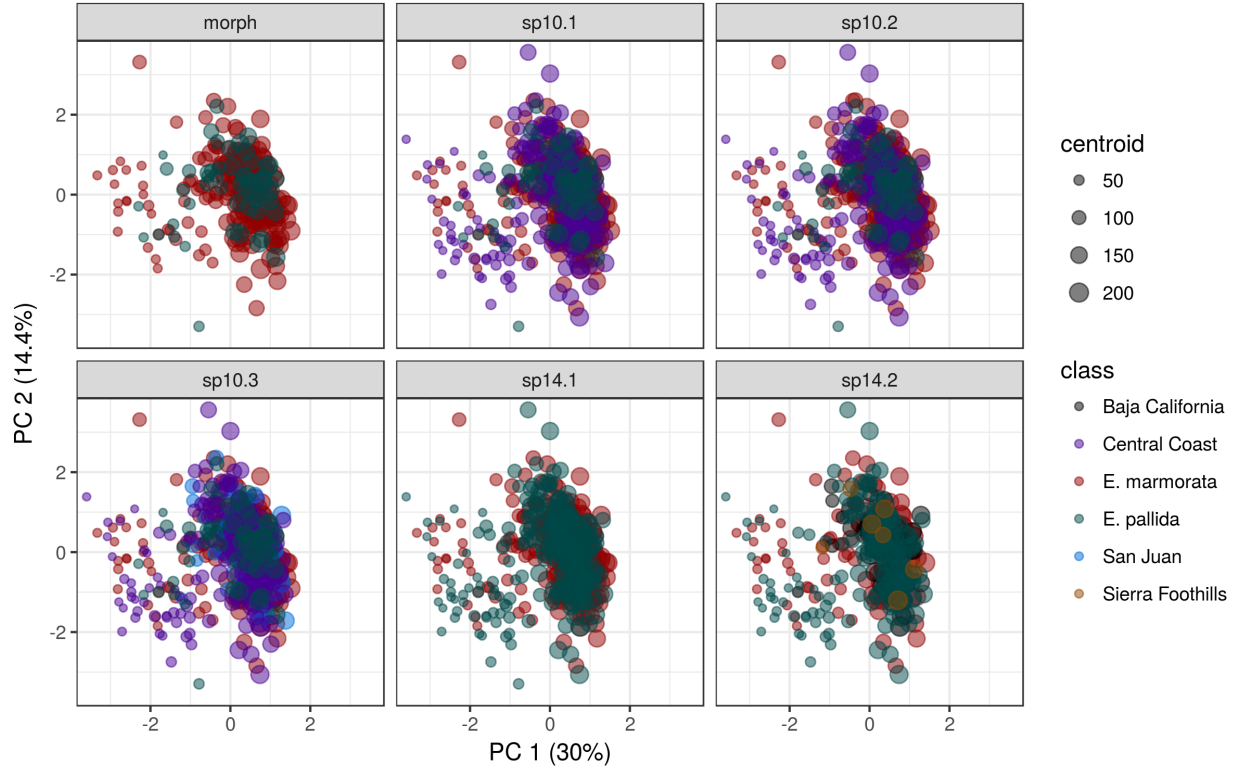


Figure 4: Scatterplot of the first two axes of morphological variation in the *Emys marmorata* dataset. Each panel corresponds to one of the six different classification schemes analyzed as part of this study (Tab. 1). Point color corresponds to the categories within each scheme, and the class names correspond to geographic regions. Point size is proportional to centroid size of that specimen and the numbers in parentheses next to the axis labels are the percent of total variation accounted for along that axis.

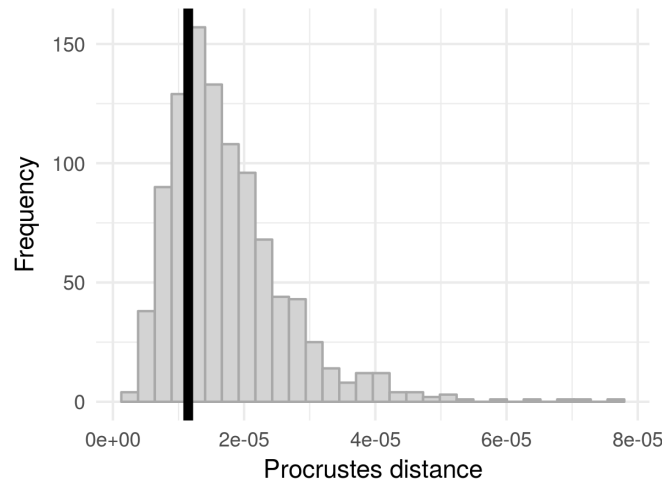


Figure 5: Comparison of observed Procrustes distance between the centroids of each sex (vertical line) to a null distribution generated from 1000 permutations of the sex-labels. This result indicates that the difference between the centroids is as small/smaller than expected by random.

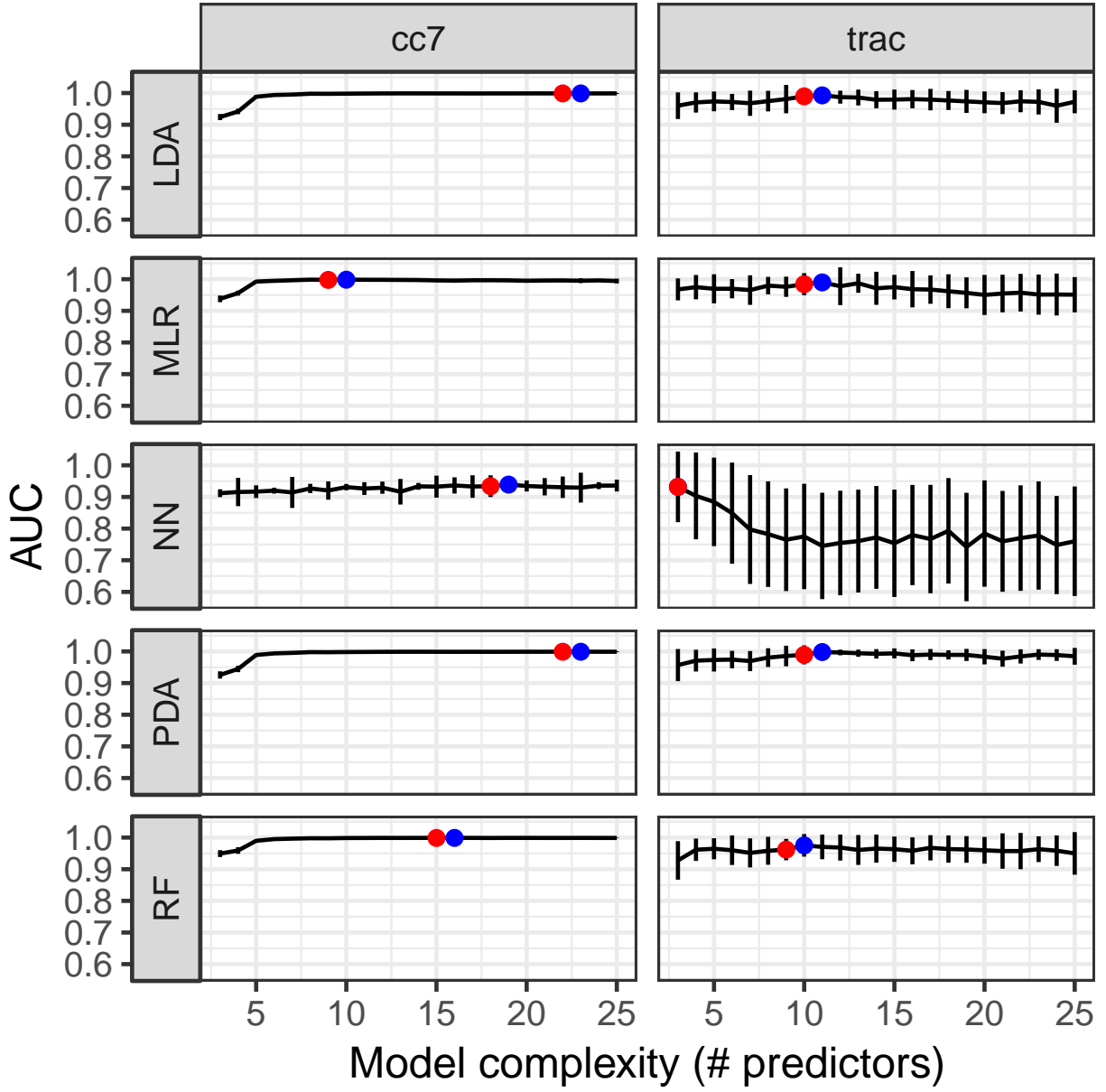


Figure 6: Comparisons of model fit to the training dataset for each of the supervised learning methods applied to the first two datasets; the results from the seven species dataset are presented in the left column, while those from the *Trachemys* dataset are presented in the right column. Models were fit to datasets of varying complexity, with the number of parameters listed along the x-axis. Model fit is measured as the area under the receiver operating characteristic (AUC), which ranges from 0.5 to 1. Error bars correspond to one standard error estimated from 10 rounds of 5-fold cross-validation. The red dot corresponds to the model fit with the highest mean AUC while the blue dot corresponds to the model selected for further analysis. In some cases, there is no difference in complexity between the best and selected models.

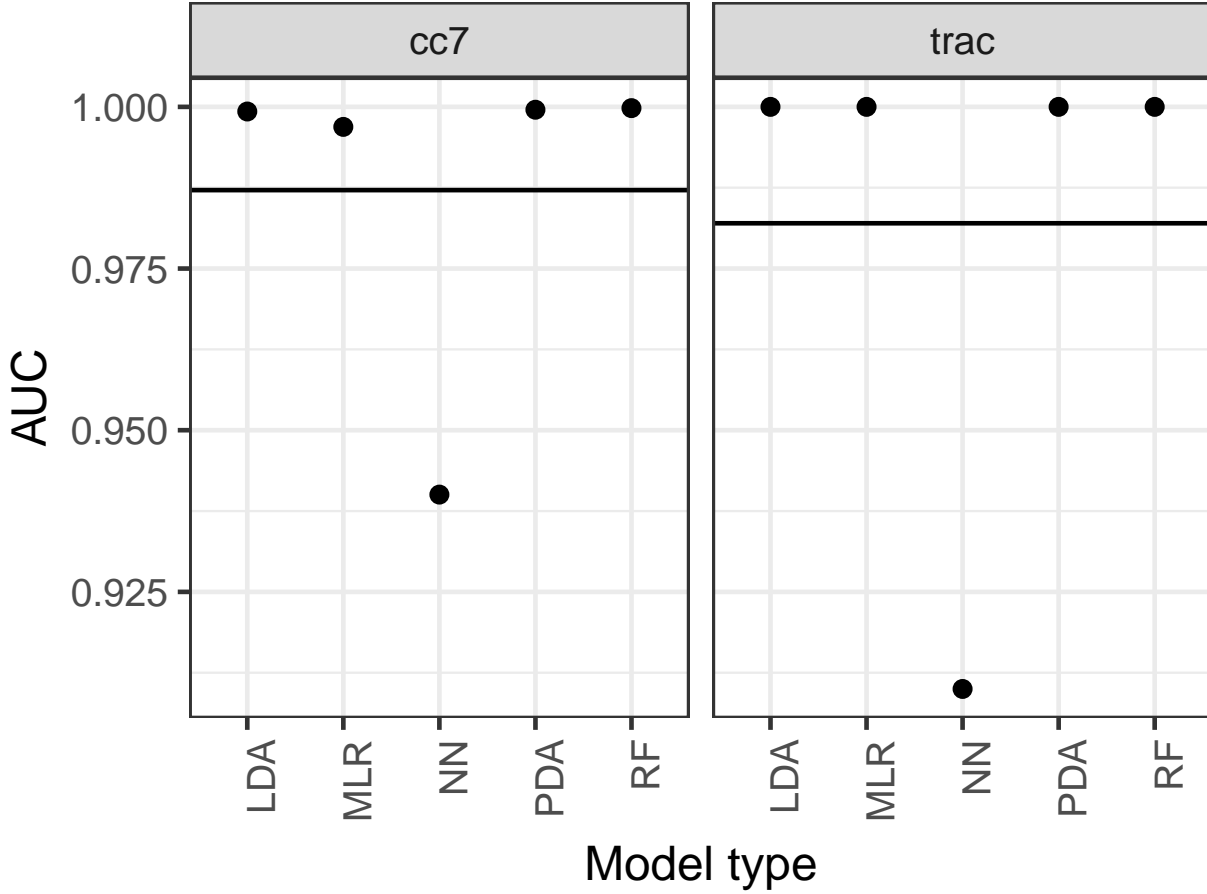


Figure 7: The results of out-of-sample predictive performance of the selected models for both the seven species (left) and *Trachemys* datasets. Predictive performance is measured as the area under the receiver operating characteristic (AUC), which ranges from 0.5 to 1. Points correspond to the individual out-of-sample predictive performance of the specific model, indicated along the x-axis. The horizontal bars correspond the average out-of-sample predictive performance of all the models.

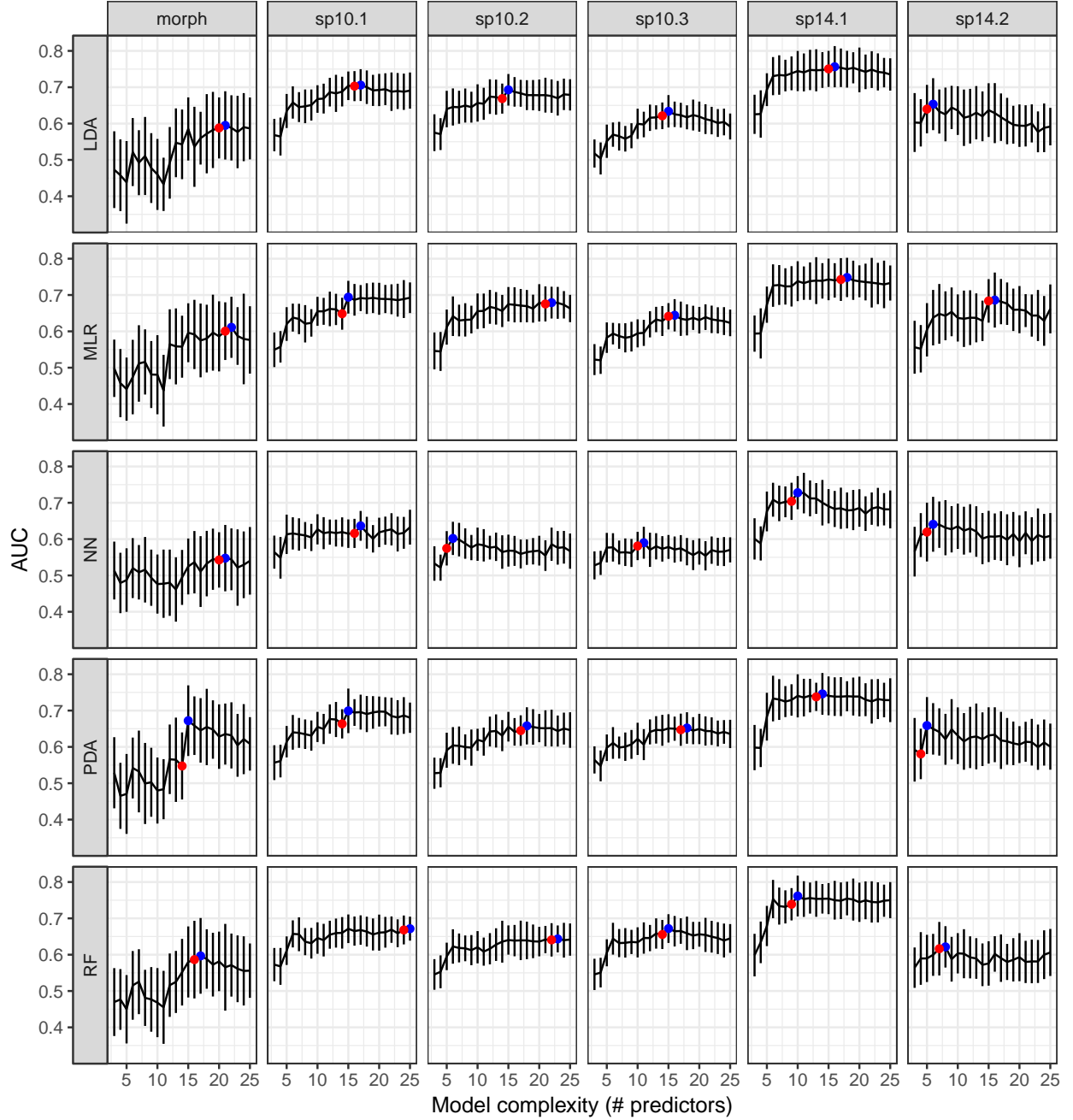


Figure 8: AUC values for models of varying complexity fit to the *Emys marmorata* training datasets for each classification scheme. The x-axis corresponds to the total number of predictors included in each model, while the y-axis corresponds to the AUC value which is a measure of goodness of fit for classification datasets. A model with a high AUC value corresponds to better classification performance than a model with a lower AUC value. Standard errors on AUC estimates are calculated from 10 rounds of 5-fold cross-validation. Indicated are the best performing and the selected models, in red and blue respectively. In some cases, there is no difference in complexity between the best and selected models.

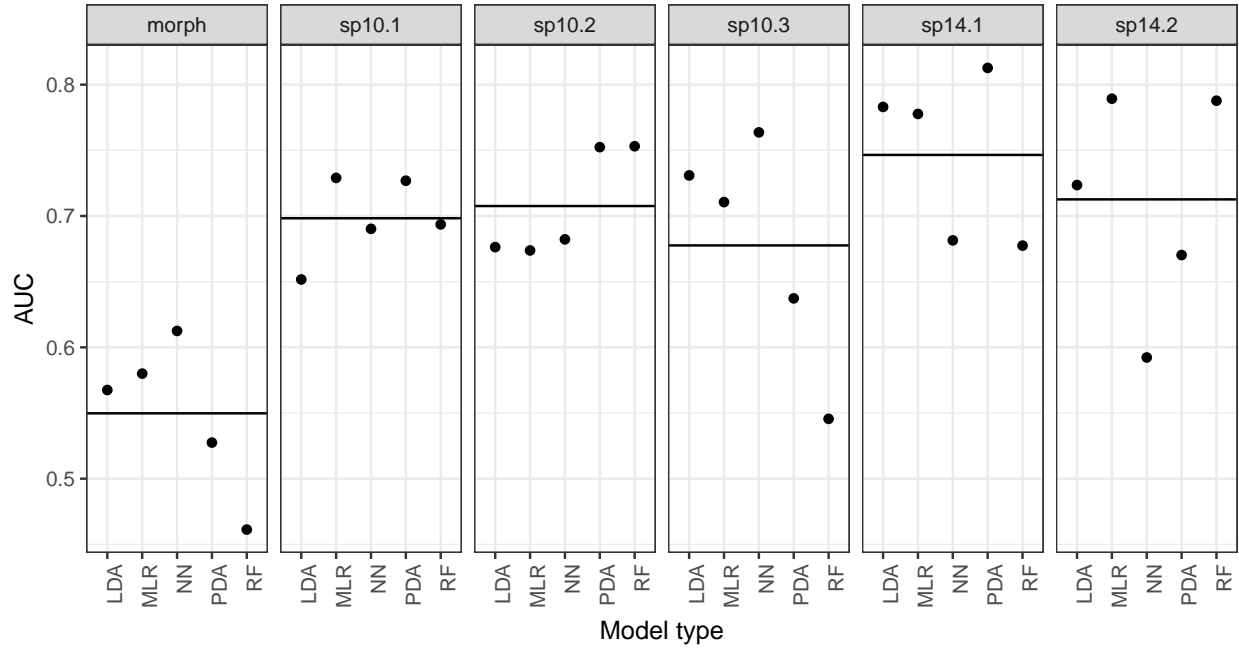


Figure 9: Comparison of out-of-sample AUC estimates from the predictions of selected models (Fig. 8), grouped by classification scheme. The horizontal line in each panel corresponds to the average AUC value across all models of that classification scheme.