How cryptic is cryptic diversity? Machine learning approaches to classifying morphological variation in the Pacific Pond Turtle (*Emys marmorata*)

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We investigate the morphometric identification of cryptic species using machine learning approaches by examining their implications for a recently proposed cryptic turtle species (*Emys pallida*). We collected landmark-based morphometric data from 354 adult *E. marmorata/“pallida”* museum specimens. We assigned a classification to each specimen for six different binning schemes based on geographic occurrence data recorded in museum collection archives. We used multiple machine learning methods, both unsupervised and supervised, to compare different classification hypotheses and asked whether it is possible to determine which amongst a set of classification hypotheses is best. In addition, we applied the above approach to two additional datasets: the subspecies of *Trachemys scripta* and a set of seven unambiguously distinct species closely related to *E. marmorata*. The results of this study indicate that there is no clear grouping of *E. marmorata/“pallida”* based on plastron shape. In contrast, the analysis of the other datasets demonstrate a near perfect classification, which demonstrates that the methods can recover correct results when they exist. Explanations for the lack of grouping in *E. marmorata* include that possibility that genetic differentiation is not associated with plastron shape variation in this species complex and/or that local selective pressures (e.g., from hydrological regime) overwhelm morphological differentiation. A reconsideration of the methods used to delimit *E. “pallida,”* the lack of barriers to gene flow, the strong evidence for widespread admixture between lineages, and the fact that plastron shape can be used to differentiate other emydine species suggest that its lack of diagnosability most likely reflects the non-distinctiveness of this proposed taxon.

# Introduction

Molecular systematics has repeatedly demonstrated the existence of cryptic species that can only be diagnosed using genetic data (Stuart, Inger, and Voris 2006; Bickford et al. 2007; Schilck-Steiner et al. 2007; Pfenninger and Schwenk 2007; Clare 2011; Funk, Caminer, and Ron 2012). In attempts to streamline the documentation of biodiversity, several methods of species delimitation that rely almost entirely on genetic data have recently been proposed (Pons et al. 2006; B.C. Carstens and Dewey 2010; Hausdorf and Hennig 2010; O’Meara 2010; Yang and Rannala 2010; J. P. Huelsenbeck, Andolfatto, and Huelsenbeck 2011). Although strong caveats on the utility of these methods have been raised (Bauer et al. 2000; Bryan C. Carstens et al. 2013), they are nevertheless being used to name species (Leaché and Fujita 2010; Spinks, Thomson, and Bradley Shaffer 2014).

The majority of extant taxa, and almost all extinct taxa, are delimited by morphology alone. This disjunction complicates interpretations of variation and diversity in deep time, as apparent morphological stasis may not reflect the true underlying diversity (Eldredge and Gould 1972; Gould and Eldredge 1977; G. Hunt 2008; Van Bocxlaer and Hunt 2013). Similarly, for many museum specimens of extant taxa (e.g. those preserved in formalin), it is difficult to acquire the genetic data needed for non-morphological species delimitation methods.

These considerations have sparked interest in whether geometric morphometric analyses can capture fine-scale variation that can be used for identifying cryptic species. This would make the task of identifying and maintaining endangered or conserved groups much easier and could contribute to improved classifications of extinct taxa and populations. Most such studies focus on using morphometrics to discover differences between taxa that were identified by other means (P.David Polly 2003; Zelditch, Swiderski, and Sheets 2004; Gaubert et al. 2005; Gündüz et al. 2007; P.David Polly 2007; Demandt and Bergek 2009). Additionally, there has been work on automated taxon identification and classification of taxa into groups (Baylac, Villemant, and Simbolotti 2003; Dobigny et al. 2003; MacLeod 2007; van den Brink and Bokma 2011).

Here, we investigate the morphometric identification of cryptic species using machine learning approaches. In particular, we ask whether it is possible to determine which amongst a set of classification hypotheses is best and examine the implications of the results for a recently proposed set of cryptic turtle species.

## Background and study system

Machine learning is an extension of known statistical methodology (Hastie, Tibshirani, and Friedman 2009) that emphasizes high predictive accuracy and generality at the expense of the interpretability of individual parameters. The basic statistical mechanics are supplemented by randomization, sorting, and partitioning algorithms, along with the maximization or minimization of summary statistics, in order to best estimate a general model for all data, both sampled and unsampled (Hastie, Tibshirani, and Friedman 2009). Machine learning approaches have found use in medical research, epidemiology, economics and automated identification of images such as handwritten zip codes (Hastie, Tibshirani, and Friedman 2009). The two major classes of machine learning methods are unsupervised and supervised learning. Unsupervised learning methods are used with unlabeled data where the underlying structure is estimated; they are analogous to clustering and density estimation methods (Kaufman and Rousseeuw 1990). Supervised learning methods are used with labeled data where the final output of data is known and the rules for going from input to output are inferred. These are analogous to classification and regression models (Breiman et al. 1984). Our application of the approaches used in this study illustrates only a sampling of the various methods available for clustering observations and fitting classification models.

Geometric morphometric approaches to identifying differences in morphological variation between different classes, including cryptic species, mostly have used methods like linear discriminate analysis and canonical variates analysis (P.David Polly 2003; Zelditch, Swiderski, and Sheets 2004; Gaubert et al. 2005; Gündüz et al. 2007; P.David Polly 2007; Francoy et al. 2009; Sztencel-Jabłonka, Jones, and BogdanowicZ 2009; Mitrovski-Bogdanovic et al. 2013). Because of their similarity to multivariate approaches like principal components analysis (PCA), these methods are comparatively straightforward ways of understanding the differences in morphology between classes. They also benefit from producing results that can be easily visualized, which aids in the interpretation and presentation of data and results. Most previous morphometric studies did not assess which amongst a set of alternative classification hypotheses was optimal. For example, studies such as those of Caumul and Polly (2005) and P.David Polly (2007) focused on comparing different aspects of morphology and their fidelity to a classification scheme instead of comparing the fidelity of one aspect of morphology to multiple classification schemes. In this context, the study of Cardini et al. (2009) is noteworthy because they compared morphological variation in marmots at the population, regional, and species level and determined the fidelity of shape to divisions at each of these levels.

Here, we used multiple machine learning methods, both unsupervised and supervised, to compare different classification hypotheses. These methods provide different advantages for understanding how to classify taxa, as well as the accuracy of the resulting classifications. Although machine learning methods such as neural networks have been applied to studying shape variation (Baylac, Villemant, and Simbolotti 2003; Dobigny et al. 2003; MacLeod 2007; van den Brink and Bokma 2011), including in the context of automated taxon identification and classification of groups, the number of cases remains limited. In the current study, we not only consider pure classification accuracy but also use a statistic of classification strength that reflects the rate at which taxa are both accurately and inaccurately classified.

We analyzed the problem of whether there are distinct subspecies or cryptic species within the western pond turtle, *Emys marmorata* (Baird and Girard 1852) (formerly *Clemmys marmorata*; see (Feldman and Parham 2002)). *Emys marmorata* is distributed from northern Washington State, USA to Baja California, Mexico. Traditionally, *E. marmorata* was classified into two named subspecies: the northern *E. marmorata marmorata* and the southern *Emys marmorata pallida* (Seeliger 1945), with a central Californian intergrade zone in between. *Emys marmorata marmorata* is differentiated from *E. marmorata pallida* by the presence of a pair of triangular inguinal scales and darker neck markings. The triangular inguinal plates can sometimes be present in *E. marmorata pallida* although they are considerably smaller. Seeliger (1945) did not formally include the Baja California populations of *E. marmorata* in either taxon, implying the existence of a third distinct but unnamed subspecies.

Previous work on morphological variation in *E. marmorata* has focused primarily on differentiation between populations over a portion of the species’ total range (Lubcke and Wilson 2007; Germano and Rathbun 2008; Germano and Bury 2009; Bury, Germano, and Bury 2010); comparatively few studies have included specimens from across the entire range (Holland 1992). Most of these studies considered how local biotic and abiotic factors may contribute to differences in carapace length and found that size can vary greatly between different populations (Lubcke and Wilson 2007; Germano and Rathbun 2008; Germano and Bury 2009). There also has been interest in size-based sexual dimorphism in *E. marmorata* (Holland 1992; Lubcke and Wilson 2007; Germano and Bury 2009), with males being on average larger than females based on total carapace length and other linear measurements. However, the quality of size as a classifier of sex can vary greatly between populations (Holland 1992) because of the magnitude of size differences among populations (Lubcke and Wilson 2007; Germano and Bury 2009). However, the effect of sexual dimorphism on shape, *sensu* David G. Kendall (1977), has not been assessed (Holland 1992; Lubcke and Wilson 2007; Germano and Rathbun 2008).

Of particular relevance in the context of cryptic diversity in *E. marmorata* is the morphometric analysis of carapace shape carried out by Holland (1992), who compared populations of *E. marmorata* from three areas of its range. This study concluded that geographic distance was a poor indicator of morphological differentiation, and instead hypothesized that geographic features such as breaks between different drainage basis are probably more important barriers to dispersal and interbreeding. Additionally, Holland (1992) suggested that morphological differences were more pronounced as the magnitude of barriers and distance increased, but this variation required many variables to adequately capture, implying only very subtle morphological differentiation between putatively distinct populations. Finally, Holland concluded that *E. marmorata* is best classified as three distinct species: a northern species, southern species, and a Columbia Basin species. This classification is similar to that of Seeliger (1945), except elevated to the species level and without recognition of a distinct Baja species.

More recently, the phylogeography of *E. marmorata* and the possibility of cryptic diversity was investigated using molecular data (Spinks and Shaffer 2005; Spinks, Thomson, and Shaffer 2010; Spinks, Thomson, and Bradley Shaffer 2014). Based on mitochondrial DNA, Spinks and Shaffer (2005) recognized four subclades within *E. marmorata*, a northern clade, a San Joaquin Valley clade, a Santa Barbara clade, and a southern clade. Analyses with nuclear DNA (Spinks, Thomson, and Shaffer 2010) and single-nucleotide polymorphism (SNP) data suggest a primarily north–south division in *E. marmorata*, although the dataset differed from that of Spinks and Shaffer (2005) in the location of this break point. These studies discussed the potential taxonomic implications of their results, with Spinks, Thomson, and Bradley Shaffer (2014) going so far as to strongly advocate for the recognition of at least two species (*E. marmorata* and *E. pallida*), and a possible third based on populations in Baja California. However, they did not discuss in detail the morphological characters that would help to diagnose these species beyond those specified by Seeliger (1945). Given that these characters are somewhat variable within the proposed species, and that Holland (1992) described shell shape variation that might be consistent with this taxonomy, a geometric morphometric analysis of shell shape might provide a reliable way to diagnose groups (whether species or subspecies) within *E. marmorata*.

In this study, we attempt to estimate the best classification scheme of *E. marmorata* based on variation in plastron (ventral shell) shape in order to determine whether this character is consistent with any of the past divisions based on other morphological features or molecular data. We are particularly interested in whether any aspect of plastron shape can be used to reliably diagnose the proposed species of Spinks, Thomson, and Bradley Shaffer (2014), and if so, the nature of that shape variation.

Because of unclear geographic boundaries between subgroups of *E. marmorata*, we compare multiple hypotheses of morphologically– and molecularly–based classification. We hypothesize that if morphological variation corresponds to class assignment, it should be possible to determine the best classification hypothesis of *E. marmorata* from amongst multiple candidate hypotheses. However, if morphological variation does not correspond to any of the standing hypothesis, then supervised learning model generalization performance will be poor.

In order to test the efficacy of our approach, we analyzed two additional datasets: the two subspecies of *Trachemys scripta scripta* and *Trachemys scripta elgans*, and seven distinct species from the Emydidae family. The former dataset represents a situation similar to the *E. marmorata* scenario except there are two valid subspecies CITATIONS. The latter dataset represents a set of morphologically distinct species. Both of these datasets help constrain the effectiveness of the methods used here and can help in interpreting the *E. marmorata* scenario.

# Materials and Methods

## Specimens, sampling, morphometrics

Three different landmark-based morphometric datasets describing plastron variation were assembled for this analysis: *E. marmorata* specimens from across its geographic range, *T. scripta* specimens from both subspecies, and specimens from seven distinct species from the Emydidae family. We chose to focus on adults because significant changes in plastron shape occur over the course of ontogeny in *E. marmorata* and other emydines (K. D. Angielczyk and Feldman 2013).

The first dataset included 354 adult *E. marmorata* museum specimens; a subset of those included in K. D. Angielczyk and Sheets (2007), K. D. Angielczyk, Feldman, and Miller (2011), and K. D. Angielczyk and Feldman (2013). We assigned a classification to each specimen for the different binning schemes based on geographic occurrence data recorded in museum collection archives. When precise latitude and longitude information were not available we estimated them from locality information. Because Spinks and Shaffer (2005), Spinks, Thomson, and Shaffer (2010), and Spinks, Thomson, and Bradley Shaffer (2014) did not use vouchered specimens we were not able to directly sample individuals in their studies. Therefore our specimen classifications were based solely on the geographic information, not explicit assignment using molecular data. Because the exact barriers between different biogeographic regions are unknown and unclear, we represented each hypothesis with two schemes for a total of six different schemes. The schemes differed based on where geographic boundaries were assigned. This changes the classification of certain individuals near the boundaries between groups, providing a test of the robustness of the classification schemes. Sex information was only know for a subset of the total dataset and was not included as a predictor of classification. Sex information was used to determine if observations cluster by sex or not.

The second dataset is a compilation of XXX specimens of *T. scripta* with approximately equal representation of both the *T. scripta scripta* and *T. scripta elegans* subspecies. This landmark data was collected specifically for this study. For the third dataset, we analyzed 578 total specimens from the following species: *Emys blandigii*, *Terrapene coahuila*, *Clemmys guttata*, *Glyptemys insculpta*, *Glyptemys muhlenbergii*, *Emys orbicularis*, and *Terrapene ornata*. Like the first data set, these specimens are a subset of those used in K. D. Angielczyk, Feldman, and Miller (2011) and K. D. Angielczyk and Feldman (2013).

[h] image [fig:plastra]

Following previous work on plastron variation (K. D. Angielczyk and Sheets 2007; K. D. Angielczyk, Feldman, and Miller 2011; K. D. Angielczyk and Feldman 2013), we used TpsDig 2.04 (Rohlf 2005) to digitize 19 landmarks (Fig. [fig:plastra]). Seventeen of the landmarks are at the endpoints or intersection of the keratinous plastral scutes that cover the platron. Twelve of the landmarks were symmetrical across the axis of symmetry and, in order to limit issues surrounding asymmetry and incomplete specimens (Klingenberg, Barluenga, and Meyer 2002), we reflected these landmarks across the axis of symmetry (i.e. midline) prior to analysis and used the average position of each symmetrical pair. In cases where damage or incompleteness prevented symmetric landmarks from being determined, we used only the single member of the pair. We conducted all subsequent analyses on the resulting “half” plastra. We superimposed the plastral landmark configurations using generalized Procrustes analysis (Dryden and Mardia 1998), after which, we calculated the principal components (PC) of shape using the shapes package for R ([CSL BIBLIOGRAPHIC DATA ERROR: reference "2015" not found.]; [CSL BIBLIOGRAPHIC DATA ERROR: reference "Dryden2013" not found.]).

## Machine learning analyses

### Unsupervised learning

In order to preserve the relationship between all landmark configurations in shape space, we measured the dissimilarity between observations using Kendall’s Riemannian shape distance or (D.G. Kendall 1984; Dryden and Mardia 1998). We chose this metric because shape space, or the set of all possible shape configurations following Procrustes superimposition, is a Riemannian manifold and thus non-Euclidean (Dryden and Mardia 1998). varies between 0 and when there is no reflection invariance, which should not be a concern in the case of the half plastral landmark configurations used in the study.

We divisively clustered the shape dissimilarity matrix using partitioning around mediods clustering (PAM), a method similar to *k*-means clustering except that instead of minimizing the sum of squared Euclidean distances between observations and centroids, the sum of squared dissimilarities between observations and mediods is minimized (Kaufman and Rousseeuw 1990). Because the optimal number of clusters of shape configurations in the study was unknown, being possibly three, four, or some other value, we estimated clustering solutions in which the number of clusters varied between one and eight. We compared clustering solutions using the gap statistic, which is a measure of goodness of clustering (Tibshirani, Walther, and Hastie 2001). This method has been applied to morphometric data before and represents a comparatively simple approach for estimating latent classification (Claude 2008).

Only the *E. marmorata* dataset was analyzed using unsupervised approaches. We conducted this analysis using the cluster package for R ([CSL BIBLIOGRAPHIC DATA ERROR: reference "Maechler2013" not found.]).

### Supervised learning

We used three different supervised learning, or classification, approaches: linear discriminate analysis, multinomial logistic regression, and random forests. Linear discriminate analysis, also known as canonical variate analysis, is commonly used in studies of geometric morphometric data (Zelditch, Swiderski, and Sheets 2004; Claude 2008; Mitteroecker and Bookstein 2011). The other two methods, however, are not. The set of possible predictors included the centroid size of each specimen along with the value of that specimen along all 22 PC axes. In all cases, the optimal subset of PCs included in the final model was chosen via maximum within-sample receiver operating characteristic values, explained below.

In classification studies, such as this one, a common metric of performance is area under the receiver operating characteristic curve (AUC). AUC is an estimate of the relationship between the false positive and true positive rates, as opposed to just the true positive rate (accuracy). This relationship is especially useful in cases such as this study where misclassification needs to be minimized just as much as an accurate classification must be obtained. AUC ranges between 0.5 and 1, with 0.5 indicating classification no better than random and 1 indicating perfect classification (Hastie, Tibshirani, and Friedman 2009).

The standard AUC calculation is defined for binary classifications, however in our application there are multiple categories. The alternative calculation that we used follows an all-against-one strategy where the individual AUC values for each class versus all others are averaged to produce a multiclass AUC (Hand and Till 2001). To estimate confidence intervals on the out-of-sample AUC values, we performed a nonparametric bootstrap in which the true and estimated classifications were resampled with replacement 1000 times.

Linear discriminate analysis (LDA) attempts to find a linear combination of predictors that best model two or more classes. LDA is very similar to PCA except that instead of finding the linear combination of features that maximize the amount of explained variance in the data, LDA maximizes the differences between classes. The results from this approach produce a transformation matrix by which the original features can be transformed to reflect the best discrimination between the classes. In this study, we applied LDA to the eigenscores from a subset of the total number of PCs, ranging from two to six in increasing order of complexity. In total, this produced nine different LDA scaling matrices.

Multinomial logistic regression is an extension of logistic regression, where instead of a binary response there are three or more response classes (Venables and Ripley 2002). Similar to the odds ratios calculated from the coefficients of a logistic regression, the relative risk of a classification can be determined from the coefficients of the model. Multinomial logistic regression models were implemented using the “nnet” R package (Venables and Ripley 2002).

Random forest models are an extension of classification and regression trees (CART) (Breiman et al. 1984; Breiman 2001). In a random forest model, many CARTs are built from a random subsample of both the features and the observations (specimens). This process is then repeated many times and the parameters of the final model are chosen as the mode of estimates from the distribution of CARTs (Breiman 2001). In addition to classifying the observations, this procedure allows for the features to be ranked in order of importance. This is a generally useful property for studies in which the goal is to describe and model the differences between classes and the relative importance of different predictors. Random forest models were fit using the “randomForest” R package ([CSL BIBLIOGRAPHIC DATA ERROR: reference "Liaw2002" not found.]). In this analysis, we used 1000 subtrees to estimate the random forest model parameters.

We estimated the best set of predictors in the random forest model using a recursive feature selection algorithm, with AUC as our optimality criterion. Following the iterative backwards selection algorithm implemented in caret ([CSL BIBLIOGRAPHIC DATA ERROR: reference "KuhnMAN2013" not found.]), the maximum number of features possible were included in the initial model, their importance ranked, and the AUC of the model calculated. The lowest-ranked feature was then removed, and the AUC of the model recalculated. Because PCs were kept in order of importance and not in relation to the amount of variance each PC described, the PCs are not included in the order of ascending eigenvalue.

The ultimate measure of model fit is accurately predicting the values of unobserved samples (Hastie, Tibshirani, and Friedman 2009; Kuhn and Johnson 2013). Within-sample performance is inherently biased upwards, so model evaluation requires overcoming this bias. With very large sample sizes, as in this study, part of the sample can be used as the “training set” and the remainder acts as the “testing set.” The former is used for fitting the model where as the later is used for measuring model performance, a process called model generalization. In this analysis, we used 75% of samples as the training set while the remaining 25% were used as the testing set. Testing and training sets are determined completely at random within each class and with respect to shape. Results were not affected by the individual specimen class assignments to the testing or training sets.

Supervised learning methods were applied to all three datasets.

# Results

## Unsupervised learning

Gap statistic values from the PAM clustering are mostly equivalent for all potential clusters, indicating that none of the proposed clusters are statistically different from each other (Fig. [fig:gap]). No clustering result is statistically greater than that with only a single cluster, because of this we favor the interpretation that there is no latent structure in the data detected in this analysis. While the solution with two groupings does have the greatest mean gap statistic, there is no geographical signal in the results of this clustering solution (Fig. [fig:gapmap]). Additionally, there is no correspondence between these two clusters and the sex of the specimens ( 0.7685, df = 1, *p*-value 0.38). Because of this, we assert that there is no means of naturally partitioning *E. marmorata* plastron shape into distinct subgroups without reference to external information.

[h] image [fig:gap]

[h] image [fig:gapmap]

## Supervised learning

AUC–based model selection revealed some important patterns of variation and congruence between the classification schemes and the actual data. Generally, the best performing models tended to include as many PCs as possible (Fig. [fig:sel]). Note that the best random forest models were determined via recursive feature selection, so PCs were not included in order of percent variance explained. That almost all LDA and multinomial logistic regression models were as complex as possible indicates that the differences between the different groups within each classification scheme are very small.

[h] image [fig:varimp]

As part of fitting a random forest model, a ranking of variable importance also is determined. Interestingly, the order of variable importance is not the same as the order of the PCs (Fig. [fig:varimp]). This means that the variance describing the differences between the classes does not align with the major axes of variance (i.e. the PCs). Such a result is expected if variation between classes was extremely fine grained and not a part of the principal form or function of the plastron. Moreover, this result is consistent with the results from the AUC–based model selection for the multinomial logistic regression and LDA models.

[ht] image [fig:genhist]

Observed AUC values for all of the optimal models are not exceptionally high (Fig. [fig:sel]). In most cases the different proposed classification schemes are generally poor descriptors of the observed variation. It appears that the data set is overwhelmed by noise, making any accurate classifications difficult at best. This observation is cemented with the generalizations of the models to the testing data set (Fig. [fig:genhist]).

[ht] image [fig:sel]

[ht]

l | c c | c c | c c & & &Scheme & Observed & Generalized & Observed & Generalized & Observed & GeneralizedMorph 1 & 0.63 & 0.73 & 0.75 & 0.79 & 0.75 & 0.80Morph 2 & 0.61 & 0.58 & 0.76 & 0.77 & 0.76 & 0.77Mito 1& 0.63 & 0.62 & 0.75 & 0.63 & 0.75 & 0.63Mito 2 & 0.77 & 0.67 & 0.80 & 0.64 & 0.80 & 0.63Mito 3 & 0.56 & 0.64 & 0.71 & 0.74 & 0.71 & 0.73Nuclear & 0.56 & 0.67 & 0.74 & 0.62 & 0.74 & 0.77

[tab:comp]

Mean AUC values for the model generalizations, in most cases, are approximately equal to the observed AUC values from the training data set (Table [tab:comp]). The cases in which the AUC from the generalizations is less than the observed indicate poor model fit and a poor classification scheme. Comparison of AUC values from the model generalizations do not indicate a clear “best” classification scheme (Fig. [fig:genhist]).

## Results from other datasets

In contrast to the muddled results from the analysis of *E. marmorata*, the analysis of *T. scripta scripta* to *T. scripta elegans* indicates that these two subspecies are morphologically distinct enough to be classified with near perfect accuracy (Table [tab:secondres], with both in-sample and out-of-sample classification having AUC values of approximately 1 for all methods. The analysis of the seven morphologically-distinct species also demonstrates near perfect classification of both the in-sample and out-of-sample datasets (Table [tab:secondres]).

For both datasets, the distribution of ROC scores from 1000 bootstrap replicates are tightly clustered near AUC = 1 (Fig. [fig:otherboot]), in contrast to the results from the *E. marmorata* case (Fig. [fig:genhist]). These results demonstrate that when there are strong distinctions between the states of the classification schemes, the methods used here can recover them.

[ht]

[t]

l r r r & # predictors & In-sample AUC & Out-of-sample AUCRF & 10 & 0.965 & 1.000LDA & 7 & 1.000 & 0.993MnL & 5 & 1.000 & 0.979

[t]

l r r r & # predictors & In-sample AUC & Out-of-sample AUCRF & 11 & 1.000 & 0.999LDA & 10 & 1.000 & 1.000MnL & 6 & 1.000 & 0.998

[tab:secondres]

[ht]

[h]0.45 image [fig:trachboot]

[h]0.45 image [fig:sevenboot]

[fig:otherboot]

# Discussion

The results of this study indicate that there is no clear grouping of *E. marmorata* based on plastron shape.

The unsupervised learning results indicate no grouping is optimal or particularly better than a single grouping, which is consistent with the results from the generalizations of the supervised learning models. The classification schemes used in the supervised learning models correspond, loosely, to unsupervised learning solutions with multiple groups. Because unsupervised learning solutions with multiple groups are poor descriptors of the observed variation, it is important to see this generally supported by the supervised learning results.

The results from fitting the various supervised learning models to each of the classification schemes generally shows that no one scheme is “best.” Possible explanations include that the genetic differentiation is not associated with plastron shape variation and/or that local selective pressures (e.g. from hydrological regime) overwhelm morphological differentiation. This makes sense given that the plastron is involved in both protection and hydrodynamics, and not necessary mate choice (Rivera 2008; Rivera and Stayton 2011; Stayton 2011; Rivera et al. 2014) and that shell shape in *E. marmorata* is known to vary among populations inhabiting water bodies with different flow regimes (Holland 1992; Lubcke and Wilson 2007; Germano and Bury 2009). Plastron shape does not seem to preserve a strong phylogenetic signal at the interspecific level in emydine turtles (K. D. Angielczyk, Feldman, and Miller 2011), and our current results suggest that this may be the case for phylogeographic signal within emydine species as well. A final possibility (explored below) is that the proposed classification schemes themselves do not represent significant evolutionary lineages.

Both the low AUC values () and the significant difference between the correctly and incorrectly classified observations support the conclusion that none of the hypothesized classification schemes are good descriptors of the observed plastral variation within *E. marmorata*.

Nevertheless, it is important to note that plastron shape is an extremely effective method for differentiating members of the other seven species we investigated. The magnitude of shape differences between the species (measured as Procrustes distance between species’ mean shapes) is approximately an order of magnitude greater than the differences between the *E. marmorata* subgroups, and the machine learning methods had no trouble accurately classifying the specimens correctly. These results demonstrate that plastron shape is normally a good marker for species delineation in the closest relatives of *E. marmorata*, and that our lack of results for *E. marmorata* is not simply a shortcoming of the methods we applied. Indeed, they beg the question of what factors have suppressed morphological differentiation of plastron shape in *E. marmorata* and *E. pallida* if they are distinct species. Invoking issues such as the role of the plastron in protection or the need for streamlining are insufficient because the other species are expected to be subject to similar constraints (Stayton 2011; P.D. Polly et al.).

## Is there more than one species of Western Pond Turtle?

The lack of morphological support for the distinctiveness of *E. pallida* does not, on its own, preclude the recognition of this taxon. However, this apparent lack of congruence does prompt a reexamination of the methods and concepts that led to that taxonomic revision, especially considering that plastron shape is normally a strong indicator of species differences among emydines. In other words, before we can assess the significance of the morphological non-diagnosablity, it is essential to evaluate the methods and concepts that led to the initial taxonomic revision.

Spinks, Thomson, and Bradley Shaffer (2014) elevated *E. pallida* based on a species delimitation analysis of SNP data using BPP (Yang and Rannala 2010). However, Spinks, Thomson, and Bradley Shaffer (2014) did not heed the caveats about species delimitation methods raised by Bryan C. Carstens et al. (2013). In addition to specifically addressing the shortcomings of validation methods such as BPP that rely on guide trees and “should be interpreted with caution,” Bryan C. Carstens et al. (2013) also strongly emphasize that “Inferences regarding species boundaries based on genetic data alone are likely inadequate, and species delimitation should be conducted with consideration of the life history, geographical distribution, morphology and behaviour (where applicable) of the focal system…” These caveats evoke the development of the Unified Species Concept (Dayrat 2005; De Queiroz 2007), Integrative Taxonomy (Padial et al. 2010), and other pluralist approaches to species delimitation. None of these considerations were brought to bear on the *E. marmorata* system until now, and in doing so we find the proposal that *E. pallida* is a distinct species to be lacking in a normally robust morphological marker.

The natural history and geographical distribution of *E. marmorata* and *E. pallida* also make the recognition of these taxa implausible. The data from Spinks, Thomson, and Bradley Shaffer (2014) show extensive introgression and admixture in Central California, which is expected because there are no significant barriers to gene flow in this region. Combined with the well-demonstrated ability for testudinoid turtles, including emydids and even *Emys*, to hybridize (e.g. Buskirk, Parham, and Feldman (2005; Spinks and Shaffer 2009; Parham et al. 2013)) it is hard to imagine how *E. marmorata* and *E. pallida* could maintain their integrity in the face of such admixture. Because the geography, natural history, demonstrated genetic admixture of *E. marmorata*, and comparisons with other morphologically diagnosable species conflict with the recognition of *E. pallida*, we hypothesize that our inability to classify the morphological data by proposed species is because *E. pallida* is not a distinct species.

We fully agree with Spinks, Thomson, and Bradley Shaffer (2014) that *E. marmorata* (*sensu lato*) is a species deserving of strong conservation efforts, and we do not wish to trivialize this need. Moreover, the genetic diversity uncovered by the analysis of Spinks, Thomson, and Bradley Shaffer (2014) should be explicitly accounted for in any conservation plan. Given the apparent lack of morphological distinction, however, we consider that this diversity should be considered Evolutionary Significant Units or Distinct Population Segments instead of distinct species.

Finally, it is important to note that the data and analyses we present do not let us definitively say whether the apparent lack of morphological divergence within *E. marmorata* truly reflects the presence of a single species, or if it is an artifact of plastron shape being a poor morphological marker for phylogenetic and phylogeographic divergences, because we could not carry out our morphometric analyses on the specimens from which the genetic data were obtained. The comparisons with the other emydine species suggests it is because *E. marmorata* is a single species. However, tests of both our preferred conclusion (*E. marmorata* as a single species) and that Spinks, Thomson, and Bradley Shaffer (2014) should include morphological and molecular analyses of the same set of voucher specimens, as well as additional tests of species delimitation using alternative methods and corroborating evidence as suggested by Bryan C. Carstens et al. (2013). From a morphological standpoint, support for the validity of “*E. pallida*” may come from other aspects of morphology, such as carapace shape or other features. Likewise, further investigation of the phylogeographic utility of plastron shape in other turtle species will help to clarify whether the lack of differentiation seen in *E. marmoarata*, and the strong differentiation among the other emydines, is typical or an unusual case.

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Key: #1Annotation: #2

Angielczyk, K. D. and C. R. Feldman. 2013. Are diminutive turtles miniaturized? The ontogeny of plastron shape in emydine turtles. Biological Journal of the Linnean Society 108:727–755.

Angielczyk, K. D., C. R. Feldman, and G. R. Miller. 2011. Adaptive evolution of plastron shape in emydine turtles. Evolution 65:377–394.

Angielczyk, K. D. and H. D. Sheets. 2007. Investigation of simulated tectonic deformation in fossils using geometric morphometrics. Paleobiology 33:125–148.

Baird, S. F. and C. Girard. 1852. Descriptions of new species of reptiles collected by the U.S. Exploring Expedition under the command of Capt. Charles Wilkes. Proceedings of the National Academy of Sciences Philadelphia 6:174–177.

Bauer, A. M., J. F. Parham, R. M. Brown, B. L. Stuart, L. Grismer, T. J. Papenfuss, W. Bohme, J. M. Savage, S. Carranza, J. L. Grismer, P. Wagner, A. Schmitz, N. B. Ananjeva, and R. F. Inger. 2000. Availability of new Baysian-delimited gecko names and the importance of character-based species descriptions. Proceedings of the Royal Society B: Biological Sciences 278:490–492.

Baylac, M., C. Villemant, and G. Simbolotti. 2003. Combining geometric morphometrics with pattern recognition for the investigation of species complexes. Biological Journal of the Linnean Society 80:89–98.

Bickford, D., D. J. Lohman, N. S. Sodhi, P. K. L. Ng, R. Meier, K. Winker, K. K. Ingram, and I. Das. 2007. Cryptic species as a window on diversity and conservation. Trends in ecology & evolution 22:148–55.

Breiman, L. 2001. Random Forests. Machine Learning 45:5–32.

Breiman, L., J. Friedman, C. J. Stone, and R. A. Olshen. 1984. Classification and regression trees. Wadsworth International Group, Belmont.

Bury, R. B., D. J. Germano, and G. W. Bury. 2010. Population Structure and Growth of the Turtle Actinemys marmorata from the Klamath–Siskiyou Ecoregion: Age, Not Size, Matters. Copeia 2010:443–451.

Buskirk, S. W., J. F. Parham, and C. R. Feldman. 2005. On the hybridisation between two distantly related Asian turtles (Testudines: Scalia x Mauremys). Salamandra 41:21–26.

Cardini, A., D. Nagorsen, P. O’Higgins, P. D. Polly, R. W. Thorington Jr, and P. Tongiorgi. 2009. Detecting biological distinctiveness using geometric morphometrics: an example case from the Vancouver Island marmot. Ethology Ecology & Evolution 21:209–223.

Carstens, B. C. and T. A. Dewey. 2010. Species Delimitation Using a Combined Coalescent and Information-Theoretic Approach: An Example from North American Myotis Bats. Systematic Biology 59:400–414.

Carstens, B. C., T. a. Pelletier, N. M. Reid, and J. D. Satler. 2013. How to fail at species delimitation. Molecular ecology 22:4369–83.

Caumul, R. and P. D. Polly. 2005. Phylogenetic and environmental components of morphological variation: skull, mandible, and molar shape in marmots (Marmota, Rodentia). Evolution; international journal of organic evolution 59:2460–72.

Clare, E. L. 2011. Cryptic species? Patterns of maternal and paternal gene flow in eight neotropical bats. PloS one 6:e21460.

Dayrat, B. 2005. Towards integrative taxonomy. Biological Journal of the Linnean Society 85:407–415.

De Queiroz, K. 2007. Species concepts and species delimitation. Systematic Biology 56:879–86.

Demandt, M. H. and S. Bergek. 2009. Identification of cyprinid hybrids by using geometric morphometrics and microsatellites. Journal of Applied Ichthyology 25:695–701.

Dobigny, G., L. Granjon, V. Aniskin, K. Ba, and V. Voloboulev. 2003. A new sigling species of Taterillus (Muridae, Gerbillinae) from West Agrica. Mammalian Biology 68:299–316.

Dryden, I. L. 2013. shapes: Statistical shape analysis. R package version 1.1-8.

Dryden, I. L. and K. Y. Mardia. 1998. Statistical shape analysis. Wiley, New York.

Eldredge, N. and S. J. Gould. 1972. Punctuated equilibria: an alternative to phyletic gradualism. Pages 82–115 *in* Models in Paleobiology (T. J. M. Schopf, ed.). Freeman Cooper, San Francisco.

Feldman, C. R. and J. F. Parham. 2002. Molecular phylogenetics of emydine turtles: taxonomic revision and the evolution of shell kinesis. Molecular Phylogenetics and Evolution 22:388–98.

Francoy, T. M., R. A. O. Silva, P. Nunes-Silva, C. Menezes, and V. L. Imperatriz-Fonseca. 2009. Gender identification of five genera of stingless bees (Apidae, Meliponini) based on wing morphology. Genetics and molecular research 8:207–214.

Funk, W. C., M. Caminer, and S. R. Ron. 2012. High levels of cryptic species diversity uncovered in Amazonian frogs. Proceedings of the Royal Society B: Biological Sciences 279:1806–14.

Gaubert, P., P. J. Taylor, C. a. Fernandes, M. W. Bruford, and G. Veron. 2005. Patterns of cryptic hybridization revealed using an integrative approach: a case study on genets (Carnivora, Viverridae, Genetta spp.) from the southern African subregion. Biological Journal of the Linnean Society 86:11–33.

Germano, D. J. and R. B. Bury. 2009. Variation in body size, growth, and population structure of Actinemys marmorata from lentic and lotic habitats in Southern Oregon. Journal of Herpetology 43:510–520.

Germano, D. J. and G. B. Rathbun. 2008. Growth, population structure, and reproduction of western pond turtles (Actinemys marmorata) on the Central Coast of California. Chelonian Conservation and Biology 7:188–194.

Gould, S. J. and N. Eldredge. 1977. Punctuated equilibria: the tempo and mode of evolution reconsidered. Paleobiology 3:115–151.

Gündüz, I., M. Jaarola, C. Tez, C. Yeniyurt, P. D. Polly, and J. B. Searle. 2007. Multigenic and morphometric differentiation of ground squirrels (Spermophilus, Scuiridae, Rodentia) in Turkey, with a description of a new species. Molecular phylogenetics and evolution 43:916–35.

Hand, D. J. and R. J. Till. 2001. A Simple Generalisation of the Area Under the ROC Curve for Multiple Class Classification Problems. Machine Learning 45:171–186.

Hastie, T., R. Tibshirani, and J. Friedman. 2009. The elements of statistical learning: data mining, inference, and prediction. 2nd ed. Springer, New York.

Hausdorf, B. and C. Hennig. 2010. Species delimitation using dominant and codominant multilocus markers. Systematic biology 59:491–503.

Holland, D. C. 1992. Level and pattern in morphological variation: a phylogeographic study of the western pond turtle (Clemmys marmorata). Ph.D. thesis University of Southwestern Louisiana.

Huelsenbeck, J. P., P. Andolfatto, and E. T. Huelsenbeck. 2011. Structurama: bayesian inference of population structure. Evolutionary bioinformatics online 7:55–9.

Hunt, G. 2008. Gradual or pulsed evolution: when should punctuational explanations be preferred? Paleobiology 34:360–377.

Kaufman, L. and P. J. Rousseeuw. 1990. Finding groups in data : an introduction to cluster analysis. Wiley, New York.

Kendall, D. G. 1977. The diffusion of shape. Advances in Applied Probability 9:428–430.

Kendall, D. G. 1984. Shape Manifolds, Procrustean Metrics, and Complex Projective Spaces. Bulletin of the London Mathematical Society 16:81–121.

Klingenberg, C. P., M. Barluenga, and A. Meyer. 2002. Shape analysis of symetric structures: quantifying variation among individuals and asymmetry. Evolution 56:1909–1920.

Kuhn, M. 2013. caret: Classification and Regression Training. R package version 5.15-61.

Kuhn, M. and K. Johnson. 2013. Applied predictive modeling. Springer, New York, NY.

Leaché, A. D. and M. K. Fujita. 2010. Bayesian species delimitation in West African forest geckos (Hemidactylus fasciatus). Proceedings. Biological sciences / The Royal Society 277:3071–7.

Lubcke, G. M. and D. S. Wilson. 2007. Variation in shell morphology of the Western Pond Turtle (Actinemys marmorata Baird and Giarard) from three aquativ habitats in Northern California. Journal of Herpetology 41:107–114.

MacLeod, N. 2007. Automated taxon identification in systematics: theory, approaches and applications. CRC Press, Boca Raton.

Maechler, M., P. Rousseeuw, A. Struyf, M. Hubert, and K. Hornik. 2013. cluster: Cluster Analysis Basics and Extensions. R package version 1.14.4.

Mitrovski-Bogdanovic, A., A. Petrovic, M. Mitrovic, A. Ivanovic, V. ikic, P. Starý, C. Vorburger, and v. Tomanovic. 2013. Identification of two cryptic species within the Praon abjectum group (Hymenoptera: Braconidae: Aphidiinae) using molecular markers and geometric morphometrics. Annals of the entomological society of America 106:170–180.

Mitteroecker, P. and F. Bookstein. 2011. Linear Discrimination, Ordination, and the Visualization of Selection Gradients in Modern Morphometrics. Evolutionary Biology 38:100–114.

O’Meara, B. C. 2010. New heuristic methods for joint species delimitation and species tree inference. Systematic biology 59:59–73.

Padial, J. M., A. Miralles, I. De la Riva, and M. Vences. 2010. The integrative future of taxonomy. Frontiers in Zoology 7:1–14.

Parham, J. F., T. J. Papenfuss, P. P. V. Dijk, B. S. Wilson, C. Marte, L. R. Schettino, and W. Brian Simison. 2013. Genetic introgression and hybridization in Antillean freshwater turtles (Trachemys) revealed by coalescent analyses of mitochondrial and cloned nuclear markers. Molecular phylogenetics and evolution 67:176–87.

Pfenninger, M. and K. Schwenk. 2007. Cryptic animal species are homogeneously distributed among taxa and biogeographical regions. BMC evolutionary biology 7:121.

Polly, P. D. 2003. Paleophylogeography of Sorex araneus: molar shape as a morphological marker for fossil shrews. Mammalia 68:233–243.

Polly, P. D. 2007. Phylogeographic differentiation in Sorex araneus: morphology in relation to geography and karyotype. Russian Journal of Theriology 6:73–84.

Polly, P. D., C. T. Stayton, E. R. Dumont, S. E. Pierce, E. J. Rayfield, and K. D. Angielczyk. ???? Combining geometric morphometrics and finite element analysis with evolutionary modeling: towards a synthesis. Journal of Vertebrate Paleontology .

Pons, J., T. Barraclough, J. Gomez-Zurita, A. Cardoso, D. Duran, S. Hazell, S. Kamoun, W. Sumlin, and A. Vogler. 2006. Sequence-Based Species Delimitation for the DNA Taxonomy of Undescribed Insects. Systematic Biology 55:595–609.

R Core Team. 2015. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing Vienna, Austria.

Rivera, G. 2008. Ecomorphological variation in shell shape of the freshwater turtle Pseudemys concinna inhabiting different aquatic flow regimes. Integrative and comparative biology 48:769–87.

Rivera, G., J. N. Davis, J. C. Godwin, and D. C. Adams. 2014. Repeatability of Habitat-Associated Divergence in Shell Shape of Turtles. Evolutionary Biology Pages 29–37.

Rivera, G. and C. T. Stayton. 2011. Finite element modeling of shell shape in the freshwater turtle Pseudemys concinna reveals a trade-off between mechanical strength and hydrodynamic efficiency. Journal of morphology 272:1192–203.

Rohlf, F. J. 2005. TpsDig 2.04.

Schilck-Steiner, B. C., B. Seifert, C. Stauffer, E. Christian, R. H. Crozier, and F. M. Steiner. 2007. Without morphology, cryptic species stay in taxonomic crypsis following discovery. Trends in ecology & evolution 22:391–392.

Seeliger, L. M. 1945. Variation in the Pacific Mud Turtle. Copeia 1945:150–159.

Spinks, P. Q. and H. B. Shaffer. 2005. Range-wide molecular analysis of the western pond turtle (Emys marmorata): cryptic variation, isolation by distance, and their conservation implications. Molecular ecology 14:2047–64.

Spinks, P. Q. and H. B. Shaffer. 2009. Conflicting mitochondrial and nuclear phylogenies for the widely disjunct Emys (Testudines: Emydidae) species complex, and what they tell us about biogeography and hybridization. Systematic biology 58:1–20.

Spinks, P. Q., R. C. Thomson, and H. Bradley Shaffer. 2014. The advantages of going large: genome wide SNPs clarify the complex population history and systematics of the threatened western pond turtle. Molecular Ecology Pages n/a–n/a.

Spinks, P. Q., R. C. Thomson, and H. B. Shaffer. 2010. Nuclear gene phylogeography reveals the historical legacy of an ancient inland sea on lineages of the western pond turtle, Emys marmorata in California. Molecular ecology 19:542–56.

Stayton, C. T. 2011. Biomechanics on the half shell: functional performance influences patterns of morphological variation in the emydid turtle carapace. Zoology (Jena, Germany) 114:213–23.

Stuart, B. L., R. F. Inger, and H. K. Voris. 2006. High level of cryptic species diversity revealed by sympatric lineages of Southeast Asian forest frogs. Biology letters 2:470–4.

Sztencel-Jabłonka, A., G. Jones, and W. BogdanowicZ. 2009. Skull Morphology of Two Cryptic Bat Species: Pipistrellus pipistrellus and P. pygmaeus — A 3D Geometric Morphometrics Approach with Landmark Reconstruction. Acta Chiropterologica 11:113–126.

Tibshirani, R., G. Walther, and T. Hastie. 2001. Estimating the number of clusters in a data set via the gap statistic. Journal of the Royal Statistical Society: Series B (Statistical Methodology) 63:411–423.

Van Bocxlaer, B. and G. Hunt. 2013. Morphological stasis in an ongoing gastropod radiation from Lake Malawi. Proceedings of the National Academy of Sciences .

van den Brink, V. and F. Bokma. 2011. Morphometric shape analysis using learning vector quantization neural networks — an example distinguishing two microtine vole species. Annales Zoologici Fennici 48:359–364.

Venables, W. and B. D. Ripley. 2002. Modern applied statistics with S. 4th ed. Springer, New York.

Law, A. and M. Wiener. 2002. Classification and Regression by randomForest. R News 2(3):18–22.

Yang, Z. and B. Rannala. 2010. Bayesian species delimitation using multilocus sequence data. Proceedings of the National Academy of Sciences 107:9264–9.

Zelditch, M. L., D. L. Swiderski, and H. D. Sheets. 2004. Geometric morphometrics for biologists: a primer. Elsevier Academic Press, Amsterdam.

Claude, J. 2004. Morphometrics with R. Springer-Verlang, New York.

Angielczyk, Kenneth D., Chris R. Feldman, and Gretchen R. Miller. 2011. “Adaptive evolution of plastron shape in emydine turtles.” *Evolution* 65 (2) (feb): 377–394. doi:10.1111/j.1558-5646.2010.01118.x.

Angielczyk, Kenneth D., and Chris R. Feldman. 2013. “Are diminutive turtles miniaturized? The ontogeny of plastron shape in emydine turtles.” *Biological Journal of the Linnean Society* 108 (4) (apr): 727–755. doi:10.1111/bij.12010. <http://doi.wiley.com/10.1111/bij.12010>.

Angielczyk, Kenneth D., and H. David Sheets. 2007. “Investigation of simulated tectonic deformation in fossils using geometric morphometrics.” *Paleobiology* 33 (1): 125–148.

Anon. “2015 not found!”

Anon. “Dryden2013 not found!”

Anon. “Maechler2013 not found!”

Anon. “Liaw2002 not found!”

Anon. “KuhnMAN2013 not found!”

Baird, S. F., and C. Girard. 1852. “Descriptions of new species of reptiles collected by the U.S. Exploring Expedition under the command of Capt. Charles Wilkes.” *Proceedings of the National Academy of Sciences Philadelphia* 6: 174–177.

Bauer, Aaron M., James F. Parham, Rafe M. Brown, Bryan L. Stuart, Lee Grismer, Theodore J. Papenfuss, W. Bohme, et al. 2000. “Availability of new Baysian-delimited gecko names and the importance of character-based species descriptions.” *Proceedings of the Royal Society B: Biological Sciences* 278: 490–492. doi:10.2307/1467045. <http://www.jstor.org/stable/1467045?origin=crossref>.

Baylac, Michel, Claire Villemant, and Giuseppina Simbolotti. 2003. “Combining geometric morphometrics with pattern recognition for the investigation of species complexes.” *Biological Journal of the Linnean Society* 80: 89–98.

Bickford, David, David J. Lohman, Navjot S. Sodhi, Peter K. L. Ng, Rudolf Meier, Kevin Winker, Krista K. Ingram, and Indraneil Das. 2007. “Cryptic species as a window on diversity and conservation.” *Trends in ecology & evolution* 22 (3) (mar): 148–55. doi:10.1016/j.tree.2006.11.004. <http://www.ncbi.nlm.nih.gov/pubmed/17129636>.

Breiman, Leo. 2001. “Random Forests.” *Machine Learning* 45 (1): 5–32.

Breiman, Leo, Jerome Friedman, Charles J. Stone, and R. A. Olshen. 1984. *Classification and regression trees*. Belmont: Wadsworth International Group.

van den Brink, Valentijn, and Folmer Bokma. 2011. “Morphometric shape analysis using learning vector quantization neural networks — an example distinguishing two microtine vole species.” *Annales Zoologici Fennici* 48: 359–364.

Bury, R. Bruce, David J. Germano, and Gwendolynn W. Bury. 2010. “Population Structure and Growth of the Turtle Actinemys marmorata from the Klamath–Siskiyou Ecoregion: Age, Not Size, Matters.” *Copeia* 2010 (3) (sep): 443–451. doi:10.1643/CH-08-096. <http://www.bioone.org/doi/abs/10.1643/CH-08-096>.

Buskirk, S. W., James F. Parham, and Chris R. Feldman. 2005. “On the hybridisation between two distantly related Asian turtles (Testudines: Scalia x Mauremys).” *Salamandra* 41: 21–26.

Cardini, A., D. Nagorsen, P. O’Higgins, P. David Polly, R. W. Thorington Jr, and P. Tongiorgi. 2009. “Detecting biological distinctiveness using geometric morphometrics: an example case from the Vancouver Island marmot.” *Ethology Ecology & Evolution* 21: 209–223.

Carstens, B. C., and T. A. Dewey. 2010. “Species Delimitation Using a Combined Coalescent and Information-Theoretic Approach: An Example from North American Myotis Bats.” *Systematic Biology* 59 (4): 400–414. <papers2://publication/doi/10.1093/sysbio/syq024>.

Carstens, Bryan C., Tara a Pelletier, Noah M. Reid, and Jordan D. Satler. 2013. “How to fail at species delimitation.” *Molecular ecology* 22 (17) (sep): 4369–83. doi:10.1111/mec.12413. <http://www.ncbi.nlm.nih.gov/pubmed/23855767>.

Caumul, Radhekshmi, and P. David Polly. 2005. “Phylogenetic and environmental components of morphological variation: skull, mandible, and molar shape in marmots (Marmota, Rodentia).” *Evolution; international journal of organic evolution* 59 (11) (nov): 2460–72. <http://www.ncbi.nlm.nih.gov/pubmed/16396186>.

Clare, Elizabeth L. 2011. “Cryptic species? Patterns of maternal and paternal gene flow in eight neotropical bats.” *PloS one* 6 (7) (jan): e21460. doi:10.1371/journal.pone.0021460.

Claude, Julien. 2008. *Morphometrics with R*. New York: Springer-Verlang.

Dayrat, Benoît. 2005. “Towards integrative taxonomy.” *Biological Journal of the Linnean Society* 85: 407–415.

De Queiroz, Kevin. 2007. “Species concepts and species delimitation.” *Systematic Biology* 56 (6) (dec): 879–86. doi:10.1080/10635150701701083. <http://www.ncbi.nlm.nih.gov/pubmed/18027281>.

Demandt, M. H., and S. Bergek. 2009. “Identification of cyprinid hybrids by using geometric morphometrics and microsatellites.” *Journal of Applied Ichthyology* 25 (6) (dec): 695–701. doi:10.1111/j.1439-0426.2009.01329.x. <http://doi.wiley.com/10.1111/j.1439-0426.2009.01329.x>.

Dobigny, G., L. Granjon, V. Aniskin, K. Ba, and V. Voloboulev. 2003. “A new sigling species of Taterillus (Muridae, Gerbillinae) from West Agrica.” *Mammalian Biology* 68: 299–316.

Dryden, Ian L., and Kanti Y. Mardia. 1998. *Statistical shape analysis*. New York: Wiley.

Eldredge, Niles, and Stephen Jay Gould. 1972. “Punctuated equilibria: an alternative to phyletic gradualism.” In *Models in Paleobiology*, ed. Thomas J. M. Schopf, 82–115. San Francisco: Freeman Cooper.

Feldman, Chris R., and James Ford Parham. 2002. “Molecular phylogenetics of emydine turtles: taxonomic revision and the evolution of shell kinesis.” *Molecular Phylogenetics and Evolution* 22 (3) (mar): 388–98. doi:10.1006/mpev.2001.1070. <http://www.ncbi.nlm.nih.gov/pubmed/11884163>.

Francoy, T. M., R. A. O. Silva, P. Nunes-Silva, C. Menezes, and V. L. Imperatriz-Fonseca. 2009. “Gender identification of five genera of stingless bees (Apidae, Meliponini) based on wing morphology.” *Genetics and molecular research* 8 (1): 207–214.

Funk, W. Chris, Marcel Caminer, and Santiago R. Ron. 2012. “High levels of cryptic species diversity uncovered in Amazonian frogs.” *Proceedings of the Royal Society B: Biological Sciences* 279 (1734) (may): 1806–14. doi:10.1098/rspb.2011.1653.

Gaubert, Philippe, Peter J. Taylor, Carlos a Fernandes, Michael W. Bruford, and Geraldine Veron. 2005. “Patterns of cryptic hybridization revealed using an integrative approach: a case study on genets (Carnivora, Viverridae, Genetta spp.) from the southern African subregion.” *Biological Journal of the Linnean Society* 86 (1) (aug): 11–33. doi:10.1111/j.1095-8312.2005.00518.x. <http://doi.wiley.com/10.1111/j.1095-8312.2005.00518.x>.

Germano, David J., and R. Bruce Bury. 2009. “Variation in body size, growth, and population structure of Actinemys marmorata from lentic and lotic habitats in Southern Oregon.” *Journal of Herpetology* 43 (3): 510–520.

Germano, David J., and Galen B. Rathbun. 2008. “Growth, population structure, and reproduction of western pond turtles (Actinemys marmorata) on the Central Coast of California.” *Chelonian Conservation and Biology* 7 (2): 188–194.

Gould, Stephen Jay, and Niles Eldredge. 1977. “Punctuated equilibria: the tempo and mode of evolution reconsidered.” *Paleobiology* 3 (2): 115–151.

Gündüz, Islam, Maarit Jaarola, Coskun Tez, Can Yeniyurt, P. David Polly, and Jeremy B. Searle. 2007. “Multigenic and morphometric differentiation of ground squirrels (Spermophilus, Scuiridae, Rodentia) in Turkey, with a description of a new species.” *Molecular phylogenetics and evolution* 43 (3) (jun): 916–35. doi:10.1016/j.ympev.2007.02.021. <http://www.ncbi.nlm.nih.gov/pubmed/17500011>.

Hand, David J., and Robert J. Till. 2001. “A Simple Generalisation of the Area Under the ROC Curve for Multiple Class Classification Problems.” *Machine Learning* 45: 171–186.

Hastie, Trevor, Robert Tibshirani, and Jerome Friedman. 2009. *The elements of statistical learning: data mining, inference, and prediction*. 2nd ed.. New York: Springer.

Hausdorf, Bernhard, and Christian Hennig. 2010. “Species delimitation using dominant and codominant multilocus markers.” *Systematic biology* 59 (5) (oct): 491–503. doi:10.1093/sysbio/syq039. <http://www.ncbi.nlm.nih.gov/pubmed/20693311>.

Holland, Dan C. 1992. “Level and pattern in morphological variation: a phylogeographic study of the western pond turtle (Clemmys marmorata).” University of Southwestern Louisiana.

Huelsenbeck, John P., Peter Andolfatto, and Edna T. Huelsenbeck. 2011. “Structurama: bayesian inference of population structure.” *Evolutionary bioinformatics online* 7 (jan): 55–9. doi:10.4137/EBO.S6761. <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3118697\&tool=pmcentrez\&rendertype=abstract>.

Hunt, Gene. 2008. “Gradual or pulsed evolution: when should punctuational explanations be preferred?” *Paleobiology* 34 (3): 360–377. doi:10.1666/07073.1.

Kaufman, L., and P. J. Rousseeuw. 1990. *Finding groups in data : an introduction to cluster analysis*. New York: Wiley.

Kendall, D. G. 1984. “Shape Manifolds, Procrustean Metrics, and Complex Projective Spaces.” *Bulletin of the London Mathematical Society* 16 (2) (mar): 81–121. doi:10.1112/blms/16.2.81. <http://blms.oxfordjournals.org/cgi/doi/10.1112/blms/16.2.81>.

Kendall, David G. 1977. “The diffusion of shape.” *Advances in Applied Probability* 9 (3): 428–430.

Klingenberg, Christian Peter, Marta Barluenga, and Axel Meyer. 2002. “Shape analysis of symetric structures: quantifying variation among individuals and asymmetry.” *Evolution* 56 (2002): 1909–1920.

Kuhn, Max, and Kjell Johnson. 2013. *Applied predictive modeling*. New York, NY: Springer.

Leaché, Adam D., and Matthew K. Fujita. 2010. “Bayesian species delimitation in West African forest geckos (Hemidactylus fasciatus).” *Proceedings. Biological sciences / The Royal Society* 277 (1697) (oct): 3071–7. doi:10.1098/rspb.2010.0662. <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=2982061\&tool=pmcentrez\&rendertype=abstract>.

Lubcke, Glen M., and Dawn S. Wilson. 2007. “Variation in shell morphology of the Western Pond Turtle (Actinemys marmorata Baird and Giarard) from three aquativ habitats in Northern California.” *Journal of Herpetology* 41 (1): 107–114.

MacLeod, Norman. 2007. *Automated taxon identification in systematics: theory, approaches and applications*. Boca Raton: CRC Press.

Mitrovski-Bogdanovic, Ana, Andjeljko Petrovic, Milana Mitrovic, Ana Ivanovic, Vladimir Žikic, Petr Starý, Christoph Vorburger, and Željko Tomanovic. 2013. “Identification of two cryptic species within the Praon abjectum group (Hymenoptera: Braconidae: Aphidiinae) using molecular markers and geometric morphometrics.” *Annals of the entomological society of America* 106 (2): 170–180.

Mitteroecker, Philipp, and Fred Bookstein. 2011. “Linear Discrimination, Ordination, and the Visualization of Selection Gradients in Modern Morphometrics.” *Evolutionary Biology* 38 (1) (feb): 100–114. doi:10.1007/s11692-011-9109-8. <http://link.springer.com/10.1007/s11692-011-9109-8>.

O’Meara, Brian C. 2010. “New heuristic methods for joint species delimitation and species tree inference.” *Systematic biology* 59 (1) (jan): 59–73. doi:10.1093/sysbio/syp077. <http://www.ncbi.nlm.nih.gov/pubmed/20525620>.

Padial, Jose M., Aurelien Miralles, Ignacio De la Riva, and Miguel Vences. 2010. “The integrative future of taxonomy.” *Frontiers in Zoology* 7 (16): 1–14.

Parham, James F., Theodore J. Papenfuss, Peter Paul Van Dijk, Byron S. Wilson, Cristian Marte, Lourdes Rodriguez Schettino, and W. Brian Simison. 2013. “Genetic introgression and hybridization in Antillean freshwater turtles (Trachemys) revealed by coalescent analyses of mitochondrial and cloned nuclear markers.” *Molecular phylogenetics and evolution* 67 (1) (apr): 176–87. doi:10.1016/j.ympev.2013.01.004. <http://www.ncbi.nlm.nih.gov/pubmed/23353072>.

Pfenninger, Markus, and Klaus Schwenk. 2007. “Cryptic animal species are homogeneously distributed among taxa and biogeographical regions.” *BMC evolutionary biology* 7 (jan): 121. doi:10.1186/1471-2148-7-121. <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1939701\&tool=pmcentrez\&rendertype=abstract>.

Polly, P. D., C. T. Stayton, E. R. Dumont, S. E. Pierce, E. J. Rayfield, and K. D. Angielczyk. “Combining geometric morphometrics and finite element analysis with evolutionary modeling: towards a synthesis.” *Journal of Vertebrate Paleontology*.

Polly, P. David. 2003. “Paleophylogeography of Sorex araneus: molar shape as a morphological marker for fossil shrews.” *Mammalia* 68 (2): 233–243.

———. 2007. “Phylogeographic differentiation in Sorex araneus: morphology in relation to geography and karyotype.” *Russian Journal of Theriology* 6 (1): 73–84.

Pons, Joan, Timothy Barraclough, Jesus Gomez-Zurita, Anabela Cardoso, Daniel Duran, Steaphan Hazell, Sophien Kamoun, William Sumlin, and Alfried Vogler. 2006. “Sequence-Based Species Delimitation for the DNA Taxonomy of Undescribed Insects.” *Systematic Biology* 55 (4) (aug): 595–609. doi:10.1080/10635150600852011. <http://sysbio.oxfordjournals.org/cgi/doi/10.1080/10635150600852011>.

Rivera, Gabriel. 2008. “Ecomorphological variation in shell shape of the freshwater turtle Pseudemys concinna inhabiting different aquatic flow regimes.” *Integrative and comparative biology* 48 (6) (dec): 769–87. doi:10.1093/icb/icn088. <http://www.ncbi.nlm.nih.gov/pubmed/21669831>.

Rivera, Gabriel, Joi N. Davis, James C. Godwin, and Dean C. Adams. 2014. “Repeatability of Habitat-Associated Divergence in Shell Shape of Turtles.” *Evolutionary Biology* (jul): 29–37. doi:10.1007/s11692-013-9243-6. <http://link.springer.com/10.1007/s11692-013-9243-6>.

Rivera, Gabriel, and C. Tristan Stayton. 2011. “Finite element modeling of shell shape in the freshwater turtle Pseudemys concinna reveals a trade-off between mechanical strength and hydrodynamic efficiency.” *Journal of morphology* 272 (10) (oct): 1192–203. doi:10.1002/jmor.10974. <http://www.ncbi.nlm.nih.gov/pubmed/21630321>.

Rohlf, F. James. 2005. “TpsDig 2.04.”

Schilck-Steiner, Brigit C., B. Seifert, C. Stauffer, E. Christian, R. H. Crozier, and F. M. Steiner. 2007. “Without morphology, cryptic species stay in taxonomic crypsis following discovery.” *Trends in ecology & evolution* 22 (8) (aug): 391–392. doi:10.1016/j.tree.2007.05.003. <http://www.ncbi.nlm.nih.gov/pubmed/17573147>.

Seeliger, L. M. 1945. “Variation in the Pacific Mud Turtle.” *Copeia* 1945 (3): 150–159.

Spinks, Phillip Q., Robert C. Thomson, and H. Bradley Shaffer. 2014. “The advantages of going large: genome wide SNPs clarify the complex population history and systematics of the threatened western pond turtle.” *Molecular Ecology* (mar): n/a–n/a. doi:10.1111/mec.12736. <http://doi.wiley.com/10.1111/mec.12736>.

Spinks, Phillip Q., Robert C. Thomson, and H. Bradley Shaffer. 2010. “Nuclear gene phylogeography reveals the historical legacy of an ancient inland sea on lineages of the western pond turtle, Emys marmorata in California.” *Molecular ecology* 19 (3) (feb): 542–56. doi:10.1111/j.1365-294X.2009.04451.x. <http://www.ncbi.nlm.nih.gov/pubmed/20051011>.

Spinks, Phillip Q., and H. Bradley Shaffer. 2005. “Range-wide molecular analysis of the western pond turtle (Emys marmorata): cryptic variation, isolation by distance, and their conservation implications.” *Molecular ecology* 14 (7) (jun): 2047–64. doi:10.1111/j.1365-294X.2005.02564.x. <http://www.ncbi.nlm.nih.gov/pubmed/15910326>.

———. 2009. “Conflicting mitochondrial and nuclear phylogenies for the widely disjunct Emys (Testudines: Emydidae) species complex, and what they tell us about biogeography and hybridization.” *Systematic biology* 58 (1) (feb): 1–20. doi:10.1093/sysbio/syp005. <http://www.ncbi.nlm.nih.gov/pubmed/20525565>.

Stayton, C. Tristan. 2011. “Biomechanics on the half shell: functional performance influences patterns of morphological variation in the emydid turtle carapace.” *Zoology (Jena, Germany)* 114 (4) (sep): 213–23. doi:10.1016/j.zool.2011.03.002. <http://www.ncbi.nlm.nih.gov/pubmed/21820295>.

Stuart, Bryan L., Robert F. Inger, and Harold K. Voris. 2006. “High level of cryptic species diversity revealed by sympatric lineages of Southeast Asian forest frogs.” *Biology letters* 2 (3) (sep): 470–4. doi:10.1098/rsbl.2006.0505. <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1686201\&tool=pmcentrez\&rendertype=abstract>.

Sztencel-Jabłonka, Anna, Gareth Jones, and Wiesław BogdanowicZ. 2009. “Skull Morphology of Two Cryptic Bat Species: Pipistrellus pipistrellus and P. pygmaeus — A 3D Geometric Morphometrics Approach with Landmark Reconstruction.” *Acta Chiropterologica* 11 (1) (jun): 113–126. doi:10.3161/150811009X465730. <http://www.bioone.org/doi/abs/10.3161/150811009X465730>.

Tibshirani, Robert, Guenther Walther, and Trevor Hastie. 2001. “Estimating the number of clusters in a data set via the gap statistic.” *Journal of the Royal Statistical Society: Series B (Statistical Methodology)* 63 (2) (may): 411–423. doi:10.1111/1467-9868.00293. <http://doi.wiley.com/10.1111/1467-9868.00293>.

Van Bocxlaer, B., and G. Hunt. 2013. “Morphological stasis in an ongoing gastropod radiation from Lake Malawi.” *Proceedings of the National Academy of Sciences* (aug). doi:10.1073/pnas.1308588110. <http://www.pnas.org/cgi/doi/10.1073/pnas.1308588110>.

Venables, William, and B. D. Ripley. 2002. *Modern applied statistics with S*. 4 ed.. New York: Springer.

Yang, Ziheng, and Bruce Rannala. 2010. “Bayesian species delimitation using multilocus sequence data.” *Proceedings of the National Academy of Sciences* 107 (20) (may): 9264–9. doi:10.1073/pnas.0913022107. <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=2889046\&tool=pmcentrez\&rendertype=abstract>.

Zelditch, Miriam Leah, Donald L. Swiderski, and H. David Sheets. 2004. *Geometric morphometrics for biologists: a primer*. Amsterdam: Elsevier Academic Press.