

Homework 3

Insert your name here

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[Link to the Github repository](#)

! Due: Thu, Mar 2, 2023 @ 11:59pm

Please read the instructions carefully before submitting your assignment.

1. This assignment requires you to only upload a PDF file on Canvas
2. Don't collapse any code cells before submitting.
3. Remember to make sure all your code output is rendered properly before uploading your submission.

Please add your name to the author information in the frontmatter before submitting your assignment

For this assignment, we will be using the [Wine Quality](#) dataset from the UCI Machine Learning Repository. The dataset consists of red and white *vinho verde* wine samples, from the north of Portugal. The goal is to model wine quality based on physicochemical tests

We will be using the following libraries:

```
library(readr)
```

Warning: package 'readr' was built under R version 4.0.5

```
library(tidyr)
```

Warning: package 'tidyr' was built under R version 4.0.5

```
library(dplyr)
```

Warning: package 'dplyr' was built under R version 4.0.5

Attaching package: 'dplyr'

The following objects are masked from 'package:stats':

filter, lag

The following objects are masked from 'package:base':

intersect, setdiff, setequal, union

```
library(purrr)
library(glmnet)
```

Warning: package 'glmnet' was built under R version 4.0.5

Loading required package: Matrix

Warning: package 'Matrix' was built under R version 4.0.5

Attaching package: 'Matrix'

The following objects are masked from 'package:tidyr':

expand, pack, unpack

Loaded glmnet 4.1-4

```
#library(car)
```

Question 1

💡 50 points

Regression with categorical covariate and *t*-Test

1.1 (5 points)

Read the wine quality datasets from the specified URLs and store them in data frames `df1` and `df2`.

```
url1 <- "https://archive.ics.uci.edu/ml/machine-learning-databases/wine-quality/winequality-red.csv"
url2 <- "https://archive.ics.uci.edu/ml/machine-learning-databases/wine-quality/winequality-white.csv"

df1 <- read.csv(url1, header = TRUE, sep = ";")
df2 <- read.csv(url2, header = TRUE, sep = ";")
```

1.2 (5 points)

Perform the following tasks to prepare the data frame `df` for analysis:

1. Combine the two data frames into a single data frame `df`, adding a new column called `type` to indicate whether each row corresponds to white or red wine.
2. Rename the columns of `df` to replace spaces with underscores
3. Remove the columns `fixed_acidity` and `free_sulfur_dioxide`
4. Convert the `type` column to a factor
5. Remove rows (if any) with missing values.

```
df1$type <- "red"
df2$type <- "white"
df <- rbind(df1, df2)

colnames(df) <- gsub(" ", "_", colnames(df))
```

```
df <- df[, !(colnames(df) %in% c("fixed.acidity", "free.sulfur.dioxide"))]

df$type <- factor(df$type)

df <- na.omit(df)

dim(df)
```

```
[1] 6497    11
```

Your output to R `dim(df)` should be

```
[1] 6497    11
```

1.3 (20 points)

Recall from STAT 200, the method to compute the t statistic for the the difference in means (with the equal variance assumption)

1. Using `df` compute the mean of `quality` for red and white wine separately, and then store the difference in means as a variable called `diff_mean`.
2. Compute the pooled sample variance and store the value as a variable called `sp_squared`.
3. Using `sp_squared` and `diff_mean`, compute the t Statistic, and store its value in a variable called `t1`.

```
mean_white <- mean(df$quality[df$type == "white"])
mean_red <- mean(df$quality[df$type == "red"])
diff_mean <- mean_white - mean_red

n_white <- length(df$quality[df$type == "white"])
n_red <- length(df$quality[df$type == "red"])
var_white <- var(df$quality[df$type == "white"])
var_red <- var(df$quality[df$type == "red"])
sp_squared <- ((n_white - 1) * var_white + (n_red - 1) * var_red) / (n_white + n_red - 2)

se <- sqrt(sp_squared * (1 / n_white + 1 / n_red))
t1 <- diff_mean / se
```

```
t1
```

```
[1] -9.68565
```

1.4 (10 points)

Equivalently, R has a function called `t.test()` which enables you to perform a two-sample *t*-Test without having to compute the pooled variance and difference in means.

Perform a two-sample *t*-test to compare the quality of white and red wines using the `t.test()` function with the setting `var.equal=TRUE`. Store the *t*-statistic in `t2`.

```
t_test <- t.test(quality ~ type, data = df, var.equal = TRUE)
t2 <- t_test$statistic
```

```
t2
```

```
      t
[1] 9.68565
```

1.5 (5 points)

Fit a linear regression model to predict `quality` from `type` using the `lm()` function, and extract the *t*-statistic for the `type` coefficient from the model summary. Store this *t*-statistic in `t3`.

```
fit <- lm(quality ~ type, data = df)
t3 <- summary(fit)$coefficients[2, "t value"]
```

```
t3
```

```
[1] -9.68565
```

1.6 (5 points)

Print a vector containing the values of `t1`, `t2`, and `t3`. What can you conclude from this? Why?

```
c(t1, t2, t3) # Insert your code here
```

```
      t  
-9.68565  9.68565 -9.68565
```

Question 2

💡 25 points

Collinearity

2.1 (5 points)

Fit a linear regression model with all predictors against the response variable **quality**. Use the `broom::tidy()` function to print a summary of the fitted model. What can we conclude from the model summary?

```
fit_all <- lm(quality ~ ., data = df)  
broom::tidy(fit_all)
```

```
# A tibble: 11 x 5
```

	term <chr>	estimate <dbl>	std.error <dbl>	statistic <dbl>	p.value <dbl>
1	(Intercept)	57.1	9.30	6.15	8.38e-10
2	volatile.acidity	-1.61	0.0806	-20.0	4.07e-86
3	citric.acid	0.0272	0.0783	0.347	7.28e- 1
4	residual.sugar	0.0451	0.00416	10.8	3.64e-27
5	chlorides	-0.964	0.333	-2.90	3.78e- 3
6	total.sulfur.dioxide	-0.000329	0.000262	-1.25	2.10e- 1
7	density	-55.2	9.32	-5.92	3.34e- 9
8	pH	0.188	0.0661	2.85	4.38e- 3
9	sulphates	0.662	0.0758	8.73	3.21e-18
10	alcohol	0.277	0.0142	19.5	1.87e-82
11	typewhite	0.386	0.0549	7.02	2.39e-12

2.2 (10 points)

Fit two **simple** linear regression models using `lm()`: one with only `citric_acid` as the predictor, and another with only `total_sulfur_dioxide` as the predictor. In both models, use `quality` as the response variable. How does your model summary compare to the summary from the previous question?

```
model_citric <- lm(quality ~ citric.acid, data = df)
summary(model_citric)
```

Call:

```
lm(formula = quality ~ citric.acid, data = df)
```

Residuals:

	Min	1Q	Median	3Q	Max
	-2.9938	-0.7831	0.1552	0.2426	3.1963

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	5.65461	0.02602	217.343	<2e-16 ***
citric.acid	0.51398	0.07429	6.918	5e-12 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.8701 on 6495 degrees of freedom

Multiple R-squared: 0.007316, Adjusted R-squared: 0.007163

F-statistic: 47.87 on 1 and 6495 DF, p-value: 5.002e-12

```
model_sulfur <- lm(quality ~ total.sulfur.dioxide, data = df)
summary(model_sulfur)
```

Call:

```
lm(formula = quality ~ total.sulfur.dioxide, data = df)
```

Residuals:

	Min	1Q	Median	3Q	Max
	-2.8866	-0.7971	0.1658	0.2227	3.1965

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	5.8923848	0.0246717	238.831	< 2e-16 ***
total.sulfur.dioxide	-0.0006394	0.0001915	-3.338	0.000848 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.8726 on 6495 degrees of freedom
Multiple R-squared: 0.001713, Adjusted R-squared: 0.001559
F-statistic: 11.14 on 1 and 6495 DF, p-value: 0.000848

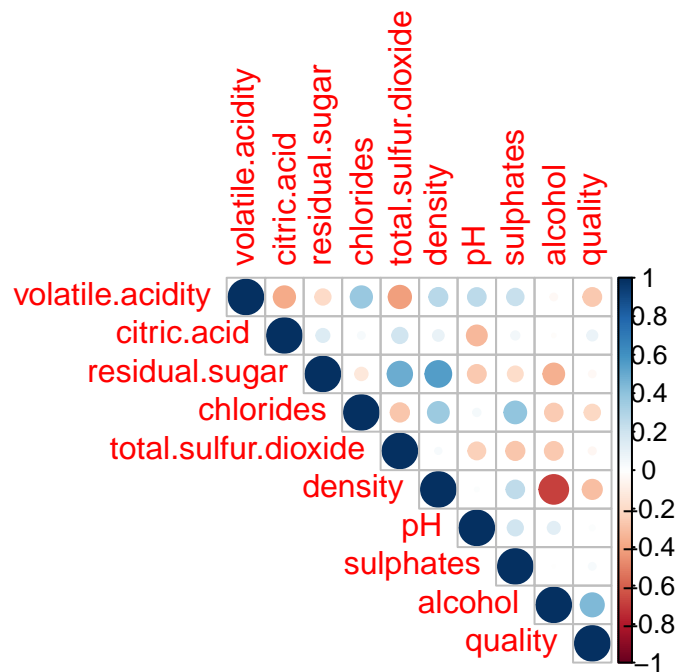
2.3 (5 points)

Visualize the correlation matrix of all numeric columns in `df` using `corrplot()`

```
library(corrplot)
```

corrplot 0.92 loaded

```
correlatoin_matrix <- df %>%
  keep(is.numeric) %>%
  cor()
corrplot(correlatoin_matrix, type = "upper", method = "circle")
```

2.4 (5 points)

Compute the variance inflation factor (VIF) for each predictor in the full model using `vif()` function. What can we conclude from this?

```
#vif_values <- vif(df) # Insert your code here
```

Question 3

💡 40 points

Variable selection

3.1 (5 points)

Run a backward stepwise regression using a `full_model` object as the starting model. Store the final formula in an object called `backward_formula` using the built-in `formula()` function in R

```
full_model <- lm(quality ~ ., df)

backward_formula <- step(full_model, direction = "backward", scope=formula(full_model))
```

Start: AIC=-3953.43

quality ~ volatile.acidity + citric.acid + residual.sugar + chlorides +
total.sulfur.dioxide + density + pH + sulphates + alcohol +
type

	Df	Sum of Sq	RSS	AIC
- citric.acid	1	0.066	3523.6	-3955.3
- total.sulfur.dioxide	1	0.854	3524.4	-3953.9
<none>			3523.5	-3953.4
- pH	1	4.413	3527.9	-3947.3
- chlorides	1	4.559	3528.1	-3947.0
- density	1	19.054	3542.6	-3920.4
- type	1	26.794	3550.3	-3906.2
- sulphates	1	41.399	3564.9	-3879.5
- residual.sugar	1	63.881	3587.4	-3838.7
- alcohol	1	206.860	3730.4	-3584.8
- volatile.acidity	1	216.549	3740.0	-3567.9

Step: AIC=-3955.3

quality ~ volatile.acidity + residual.sugar + chlorides + total.sulfur.dioxide +
density + pH + sulphates + alcohol + type

	Df	Sum of Sq	RSS	AIC
- total.sulfur.dioxide	1	0.818	3524.4	-3955.8
<none>			3523.6	-3955.3
- chlorides	1	4.495	3528.1	-3949.0
- pH	1	4.536	3528.1	-3948.9
- density	1	20.794	3544.4	-3919.1
- type	1	26.943	3550.5	-3907.8
- sulphates	1	41.491	3565.1	-3881.2
- residual.sugar	1	67.371	3590.9	-3834.3
- alcohol	1	235.151	3758.7	-3537.6

```
- volatile.acidity      1    252.565 3776.1 -3507.5
```

Step: AIC=-3955.8

```
quality ~ volatile.acidity + residual.sugar + chlorides + density +
      pH + sulphates + alcohol + type
```

	Df	Sum of Sq	RSS	AIC
<none>			3524.4	-3955.8
- pH	1	4.295	3528.7	-3949.9
- chlorides	1	4.523	3528.9	-3949.5
- density	1	21.540	3545.9	-3918.2
- sulphates	1	40.711	3565.1	-3883.2
- type	1	43.664	3568.0	-3877.8
- residual.sugar	1	66.572	3591.0	-3836.2
- alcohol	1	244.545	3768.9	-3521.9
- volatile.acidity	1	256.695	3781.1	-3501.0

3.2 (5 points)

Run a forward stepwise regression using a `null_model` object as the starting model. Store the final formula in an object called `forward_formula` using the built-in `formula()` function in R

```
null_model <- lm(quality ~ 1, df)

forward_formula <- step(null_model, direction = "forward", scope = formula(full_model))
```

Start: AIC=-1760.04

```
quality ~ 1
```

	Df	Sum of Sq	RSS	AIC
+ alcohol	1	977.95	3975.7	-3186.9
+ density	1	463.41	4490.3	-2396.2
+ volatile.acidity	1	349.71	4604.0	-2233.7
+ chlorides	1	199.47	4754.2	-2025.1
+ type	1	70.53	4883.2	-1851.2
+ citric.acid	1	36.24	4917.4	-1805.7
+ total.sulfur.dioxide	1	8.48	4945.2	-1769.2
+ sulphates	1	7.34	4946.3	-1767.7
+ residual.sugar	1	6.77	4946.9	-1766.9

+ pH	1	1.88	4951.8	-1760.5
<none>			4953.7	-1760.0

Step: AIC=-3186.88
quality ~ alcohol

	Df	Sum of Sq	RSS	AIC
+ volatile.acidity	1	307.508	3668.2	-3707.9
+ residual.sugar	1	85.662	3890.1	-3326.4
+ type	1	54.335	3921.4	-3274.3
+ citric.acid	1	40.303	3935.4	-3251.1
+ chlorides	1	39.696	3936.0	-3250.1
+ total.sulfur.dioxide	1	31.346	3944.4	-3236.3
+ sulphates	1	7.859	3967.9	-3197.7
+ pH	1	5.938	3969.8	-3194.6
<none>			3975.7	-3186.9
+ density	1	0.005	3975.7	-3184.9

Step: AIC=-3707.89
quality ~ alcohol + volatile.acidity

	Df	Sum of Sq	RSS	AIC
+ sulphates	1	48.259	3620.0	-3791.9
+ density	1	38.704	3629.5	-3774.8
+ residual.sugar	1	29.751	3638.5	-3758.8
+ type	1	28.895	3639.3	-3757.3
+ total.sulfur.dioxide	1	5.619	3662.6	-3715.9
+ pH	1	5.533	3662.7	-3715.7
<none>			3668.2	-3707.9
+ chlorides	1	0.162	3668.1	-3706.2
+ citric.acid	1	0.099	3668.1	-3706.1

Step: AIC=-3791.94
quality ~ alcohol + volatile.acidity + sulphates

	Df	Sum of Sq	RSS	AIC
+ residual.sugar	1	43.989	3576.0	-3869.4
+ density	1	18.661	3601.3	-3823.5
+ type	1	6.012	3614.0	-3800.7
+ chlorides	1	4.988	3615.0	-3798.9
+ citric.acid	1	2.031	3617.9	-3793.6
+ pH	1	1.903	3618.1	-3793.4
<none>			3620.0	-3791.9

```
+ total.sulfur.dioxide 1      0.817 3619.2 -3791.4
```

Step: AIC=-3869.37

```
quality ~ alcohol + volatile.acidity + sulphates + residual.sugar
```

	Df	Sum of Sq	RSS	AIC
+ type	1	20.7581	3555.2	-3905.2
+ total.sulfur.dioxide	1	13.3542	3562.6	-3891.7
+ pH	1	6.6430	3569.3	-3879.5
+ citric.acid	1	4.3384	3571.6	-3875.3
+ chlorides	1	1.8907	3574.1	-3870.8
<none>			3576.0	-3869.4
+ density	1	0.0071	3576.0	-3867.4

Step: AIC=-3905.19

```
quality ~ alcohol + volatile.acidity + sulphates + residual.sugar +
      type
```

	Df	Sum of Sq	RSS	AIC
+ density	1	20.4623	3534.8	-3940.7
+ chlorides	1	6.6602	3548.6	-3915.4
+ citric.acid	1	5.2242	3550.0	-3912.7
+ pH	1	3.9477	3551.3	-3910.4
+ total.sulfur.dioxide	1	1.2539	3554.0	-3905.5
<none>			3555.2	-3905.2

Step: AIC=-3940.7

```
quality ~ alcohol + volatile.acidity + sulphates + residual.sugar +
      type + density
```

	Df	Sum of Sq	RSS	AIC
+ chlorides	1	6.0826	3528.7	-3949.9
+ pH	1	5.8541	3528.9	-3949.5
<none>			3534.8	-3940.7
+ citric.acid	1	0.8471	3533.9	-3940.3
+ total.sulfur.dioxide	1	0.5646	3534.2	-3939.7

Step: AIC=-3949.89

```
quality ~ alcohol + volatile.acidity + sulphates + residual.sugar +
      type + density + chlorides
```

	Df	Sum of Sq	RSS	AIC
+ pH	1	4.2945	3524.4	-3955.8

```

<none>                                3528.7 -3949.9
+ total.sulfur.dioxide 1    0.5765 3528.1 -3948.9
+ citric.acid          1    0.2338 3528.4 -3948.3

```

Step: AIC=-3955.8

```

quality ~ alcohol + volatile.acidity + sulphates + residual.sugar +
          type + density + chlorides + pH

```

```

                                Df Sum of Sq    RSS    AIC
<none>                                3524.4 -3955.8
+ total.sulfur.dioxide 1    0.81762 3523.6 -3955.3
+ citric.acid          1    0.02919 3524.4 -3953.9

```

3.3 (10 points)

1. Create a `y` vector that contains the response variable (`quality`) from the `df` dataframe.
2. Create a design matrix `X` for the `full_model` object using the `make_model_matrix()` function provided in the Appendix.
3. Then, use the `cv.glmnet()` function to perform LASSO and Ridge regression with `X` and `y`.

```

y <- df$quality

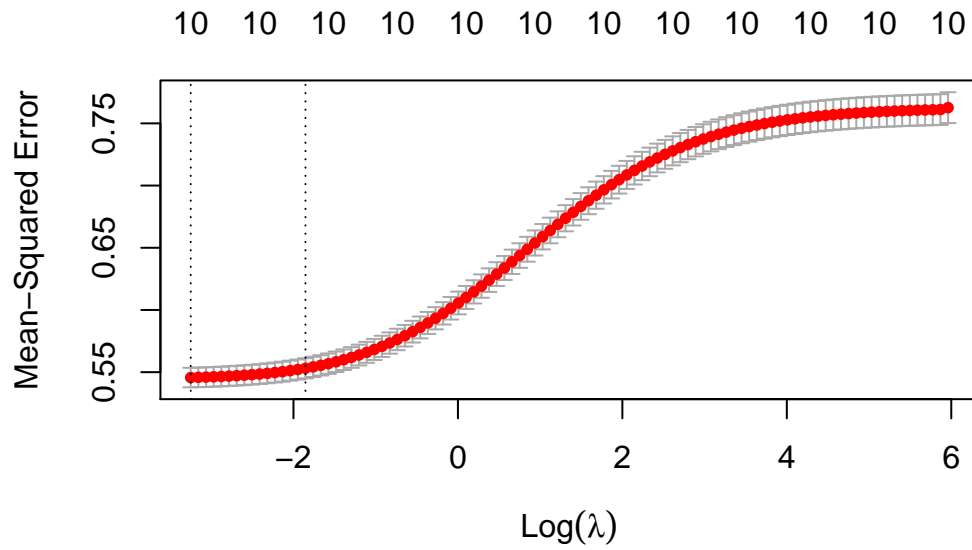
make_model_matrix <- function(formula){
  X <- model.matrix(full_model, df)[, -1]
  cnames <- colnames(X)
  for(i in 1:ncol(X)){
    if(!cnames[i] == "typewhite"){
      X[, i] <- scale(X[, i])
    } else {
      colnames(X)[i] <- "type"
    }
  }
  return(X)
}

cv_ridge <- cv.glmnet(make_model_matrix(forward_formula), y, alpha = 0)
cv_lasso <- cv.glmnet(make_model_matrix(forward_formula), y, alpha = 1)

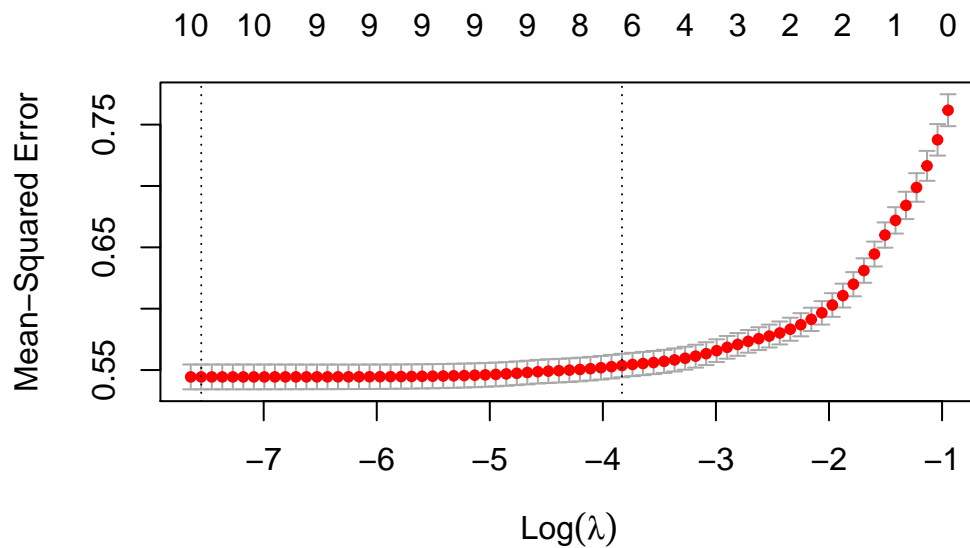
```

Create side-by-side plots of the ridge and LASSO regression results. Interpret your main findings.

```
plot(cv_ridge)
```



```
plot(cv_lasso)
```



3.4 (5 points)

Print the coefficient values for LASSO regression at the `lambda.1se` value? What are the variables selected by LASSO?

Store the variable names with non-zero coefficients in `lasso_vars`, and create a formula object called `lasso_formula` using the `make_formula()` function provided in the Appendix.

```
lasso_coefficient <- coef(cv_lasso, s = "lambda.1se")
lasso_coefficient
```

11 x 1 sparse Matrix of class "dgCMatrix"

```
      s1
(Intercept)      5.816929919
volatile.acidity -0.204848960
citric.acid      .
residual.sugar   0.059537104
chlorides        -0.001916886
total.sulfur.dioxide -0.008115301
density          .
```



```
pH                0.003557628
sulphates         0.067073695
alcohol           0.377201730
type              0.005882625
```

```
lasso_vars <- rownames(lasso_coefficient)[which((lasso_coefficient) > 0)]
lasso_vars
```

```
[1] "(Intercept)"      "residual.sugar" "pH"              "sulphates"
[5] "alcohol"          "type"
```

```
make_formula <- function(x){
  as.formula(
    paste("quality ~ ", paste(x, collapse = " + "))
  )
}

lasso_formula <- make_formula(lasso_vars)
lasso_formula
```

```
quality ~ (Intercept) + residual.sugar + pH + sulphates + alcohol +
  type
<environment: 0x0000000027c1c478>
```

3.5 (5 points)

Print the coefficient values for ridge regression at the `lambda.1se` value? What are the variables selected here?

Store the variable names with non-zero coefficients in `ridge_vars`, and create a formula object called `ridge_formula` using the `make_formula()` function provided in the Appendix.

```
ridge_coefficient <- coef(cv_ridge, s = "lambda.1se")
ridge_coefficient
```

```
11 x 1 sparse Matrix of class "dgCMatix"
```

```
      s1  
(Intercept)      5.79431467  
volatile.acidity -0.18836482  
citric.acid      0.01721478  
residual.sugar   0.11167415  
chlorides        -0.04481638  
total.sulfur.dioxide -0.04027017  
density          -0.08890876  
pH               0.02572467  
sulphates        0.08571325  
alcohol          0.28530948  
type            0.09777210
```

```
ridge_vars <- rownames(ridge_coefficient)[which((ridge_coefficient) > 0)]  
ridge_vars
```

```
[1] "(Intercept)"  "citric.acid"    "residual.sugar" "pH"  
[5] "sulphates"    "alcohol"        "type"
```

```
make_formula <- function(x){  
  as.formula(  
    paste("quality ~ ", paste(x, collapse = " + "))  
  )  
}
```

```
ridge_formula <- make_formula(ridge_vars)  
ridge_formula
```

```
quality ~ (Intercept) + citric.acid + residual.sugar + pH + sulphates +  
  alcohol + type  
<environment: 0x0000000025de69c0>
```

3.6 (10 points)

What is the difference between stepwise selection, LASSO and ridge based on you analyses above?

```
"Stepwise uses 8 variables"
```

```
[1] "Stepwise uses 8 variables"
```


```
"Lasso uses 5 variables, not including the intercept"
```

```
[1] "Lasso uses 5 variables, not including the intercept"
```

```
"Ridge uses 6 variables, not including the intercept"
```

```
[1] "Ridge uses 6 variables, not including the intercept"
```

Question 4

 70 points

Variable selection

4.1 (5 points)

Excluding `quality` from `df` we have 10 possible predictors as the covariates. How many different models can we create using any subset of these 10 covariates as possible predictors? Justify your answer.

```
"The number of different models we can create using any subset of n covariates is 2^n since
```

```
[1] "The number of different models we can create using any subset of n covariates is 2^n since
```

```
"In this case, we have n = 10 covariates, so the total number of different models we can c
```

```
[1] "In this case, we have n = 10 covariates, so the total number of different models we can
```

4.2 (20 points)

Store the names of the predictor variables (all columns except `quality`) in an object called `x_vars`.

```
x_vars <- colnames(df %>% select(-quality))
```

Use:

- the `combn()` function (built-in R function) and
- the `make_formula()` (provided in the Appendix)

to **generate all possible linear regression formulas** using the variables in `x_vars`. This is most optimally achieved using the `map()` function from the `purrr` package.

```
make_formula <- function(x){  
  as.formula(  
    paste("quality ~ ", paste(x, collapse = " + "))  
  )  
}  
  
formulas <- map(  
  1:length(x_vars),  
  function(x){  
    vars <- combn(x_vars, x, simplify = FALSE) # Insert code here  
    map(vars, make_formula) # Insert code here  
  }  
) %>% unlist()
```

If your code is right the following command should return something along the lines of:

```
sample(formulas, 4) %>% as.character()
```

```
[1] "quality ~ chlorides + density + pH + sulphates + alcohol + type"  
[2] "quality ~ type"  
[3] "quality ~ density + pH + alcohol"  
[4] "quality ~ volatile.acidity + citric.acid + residual.sugar + total.sulfur.dioxide + sulphates"
```

```
# Output:
```

```
# [1] "quality ~ volatile_acidity + residual_sugar + density + pH + alcohol"
```

```
# [2] "quality ~ citric_acid"
# [3] "quality ~ volatile_acidity + citric_acid + residual_sugar + total_sulfur_dioxide +
# [4] "quality ~ citric_acid + chlorides + total_sulfur_dioxide + pH + alcohol + type"
```

4.3 (10 points)

Use `map()` and `lm()` to fit a linear regression model to each formula in `formulas`, using `df` as the data source. Use `broom::glance()` to extract the model summary statistics, and bind them together into a single tibble of summaries using the `bind_rows()` function from `dplyr`.

```
models <- map(formulas, ~lm((-quality) ~ ., data=df)) # Insert your code here
summaries <- map(models, broom::glance) # Insert your code here
summaries_tibble <- bind_rows(summaries, .id="formula")
```

4.4 (5 points)

Extract the `adj.r.squared` values from `summaries` and use them to identify the formula with the *highest* adjusted R-squared value.

```
R_squared <- summaries$adj.r.squared

max_adj_index <- which.max(R_squared)

r_form <- formulas[max_adj_index]
```

Store resulting formula as a variable called `rsq_formula`.

```
#rsq_formula <- ... #insert code here
```

4.5 (5 points)

Extract the AIC values from `summaries` and use them to identify the formula with the *lowest* AIC value.

```
... # Insert your code here
```

Store resulting formula as a variable called `aic_formula`.

```
aic_formula <- ... # Insert your code
```

4.6 (15 points)

Combine all formulas shortlisted into a single vector called `final_formulas`.

```
null_formula <- formula(null_model)
full_formula <- formula(full_model)

final_formulas <- c(
  null_formula,
  full_formula,
  backward_formula,
  forward_formula,
  lasso_formula,
  ridge_formula,
  #rsq_formula,
  #aic_formula
)
```

- Are `aic_formula` and `rsq_formula` the same? How do they differ from the formulas shortlisted in question 3?
 - Which of these is more reliable? Why?
 - If we had a dataset with 10,000 columns, which of these methods would you consider for your analyses? Why?
-

4.7 (10 points)

Use `map()` and `glance()` to extract the `sigma`, `adj.r.squared`, `AIC`, `df`, and `p.value` statistics for each model obtained from `final_formulas`. Bind them together into a single data frame `summary_table`. Summarize your main findings.

```
summary_table <- map(
  final_formulas,
  function(x) ... # Insert your code here
) %>% bind_rows()

summary_table %>% knitr::kable()
```

Appendix

Convenience function for creating a formula object

The following function which takes as input a vector of column names `x` and outputs a **formula** object with `quality` as the response variable and the columns of `x` as the covariates.

```
make_formula <- function(x){
  as.formula(
    paste("quality ~ ", paste(x, collapse = " + "))
  )
}

# For example the following code will
# result in a formula object
# "quality ~ a + b + c"
make_formula(c("a", "b", "c"))
```

Convenience function for glmnet

The `make_model_matrix` function below takes a **formula** as input and outputs a **rescaled** model matrix `X` in a format amenable for `glmnet()`

```
make_model_matrix <- function(formula){
  X <- model.matrix(formula, df)[, -1]
  cnames <- colnames(X)
  for(i in 1:ncol(X)){
    if(!cnames[i] == "typewhite"){
      X[, i] <- scale(X[, i])
    } else {
      colnames(X)[i] <- "type"
    }
  }
  return(X)
}
```


Session Information

Print your R session information using the following command

```
sessionInfo()
```

R version 4.0.3 (2020-10-10)

Platform: x86_64-w64-mingw32/x64 (64-bit)

Running under: Windows 10 x64 (build 19044)

Matrix products: default

locale:

[1] LC_COLLATE=English_United States.1252

[2] LC_CTYPE=English_United States.1252

[3] LC_MONETARY=English_United States.1252

[4] LC_NUMERIC=C

[5] LC_TIME=English_United States.1252

attached base packages:

[1] stats graphics grDevices datasets utils methods base

other attached packages:

[1] corrplot_0.92 glmnet_4.1-4 Matrix_1.4-1 purrr_0.3.4 dplyr_1.0.8

[6] tidyr_1.2.0 readr_2.1.2

loaded via a namespace (and not attached):

[1] Rcpp_1.0.8.3 pillar_1.8.1 compiler_4.0.3 iterators_1.0.14

[5] tools_4.0.3 digest_0.6.29 jsonlite_1.8.0 evaluate_0.20

[9] lifecycle_1.0.1 tibble_3.1.6 lattice_0.20-45 pkgconfig_2.0.3

[13] rlang_1.0.2 foreach_1.5.2 cli_3.2.0 rstudioapi_0.14

[17] yaml_2.3.5 xfun_0.20 stringr_1.4.0 knitr_1.30

[21] generics_0.1.0 vctrs_0.4.1 hms_1.1.2 grid_4.0.3

[25] tidyselect_1.1.2 glue_1.6.2 R6_2.5.1 fansi_0.4.2

[29] survival_3.2-7 rmarkdown_2.6 tzdb_0.3.0 magrittr_2.0.3

[33] backports_1.4.1 splines_4.0.3 codetools_0.2-16 ellipsis_0.3.2

[37] htmltools_0.5.1 shape_1.4.6 renv_0.16.0 utf8_1.1.4

[41] stringi_1.7.6 broom_1.0.3