Homework 3

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Link to the Github repository

Due: Thu, Mar 2, 2023 @ 11:59pm

Please read the instructions carefully before submitting your assignment.

- 1. This assignment requires you to only upload a PDF file on Canvas
- 2. Don't collapse any code cells before submitting.
- 3. Remember to make sure all your code output is rendered properly before uploading your submission.

Please add your name to the author information in the frontmatter before submitting your assignment

For this assignment, we will be using the Wine Quality dataset from the UCI Machine Learning Repository. The dataset consists of red and white *vinho verde* wine samples, from the north of Portugal. The goal is to model wine quality based on physicochemical tests

We will be using the following libraries:

```
rm(list=ls())
  library(readr)
  library(tidyr)
  library(dplyr)
Attaching package: 'dplyr'
The following objects are masked from 'package:stats':
    filter, lag
The following objects are masked from 'package:base':
    intersect, setdiff, setequal, union
  library(purrr)
  library(car)
Loading required package: carData
Attaching package: 'car'
The following object is masked from 'package:purrr':
    some
The following object is masked from 'package:dplyr':
    recode
  library(glmnet)
Loading required package: Matrix
Attaching package: 'Matrix'
```

```
The following objects are masked from 'package:tidyr':
    expand, pack, unpack

Loaded glmnet 4.1-6

library(corrplot)

corrplot 0.92 loaded

library(janitor)

Attaching package: 'janitor'

The following objects are masked from 'package:stats':
    chisq.test, fisher.test
```

Appendix

Convenience function for creating a formula object

The following function which takes as input a vector of column names x and outputs a formula object with quality as the response variable and the columns of x as the covariates.

```
make_formula <- function(x){
   as.formula(
      paste("quality ~ ", paste(x, collapse = " + "))
   )
}

# For example the following code will
# result in a formula object
# "quality ~ a + b + c"
   make_formula(c("a", "b", "c"))

quality ~ a + b + c
<environment: 0x7f88f8a58958>
```

Convenience function for glmnet

The make_model_matrix function below takes a formula as input and outputs a rescaled model matrix X in a format amenable for glmnet()

```
make_model_matrix <- function(formula){
    X <- model.matrix(formula, df)[, -1]
    cnames <- colnames(X)
    for(i in 1:ncol(X)){
        if(!cnames[i] == "typewhite"){
            X[, i] <- scale(X[, i])
        } else {
            colnames(X)[i] <- "type"
        }
    }
    return(X)
}</pre>
```

Question 1



Regression with categorical covariate and t-Test

1.1 (5 points)

Read the wine quality datasets from the specified URLs and store them in data frames df1 and df2.

```
url1 <- "https://archive.ics.uci.edu/ml/machine-learning-databases/wine-quality/winequalit
url2 <- "https://archive.ics.uci.edu/ml/machine-learning-databases/wine-quality/winequalit
df1 <- read.table(url1,sep=";") # Insert your code here
df1 <- df1 %>%
    row_to_names(row_number = 1)
df2 <- read.table(url2,sep=";") # Insert your code here
df2 <- df2 %>%
    row_to_names(row_number = 1)
```

1.2 (5 points)

Perform the following tasks to prepare the data frame df for analysis:

- 1. Combine the two data frames into a single data frame df, adding a new column called type to indicate whether each row corresponds to white or red wine.
- 2. Rename the columns of df to replace spaces with underscores
- 3. Remove the columns fixed_acidity and free_sulfur_dioxide
- 4. Convert the type column to a factor
- 5. Remove rows (if any) with missing values.

```
red = "red"
  white = "white"
  df1$type <- white
  df2$type <- red
  df <- rbind(df1,df2) # Insert your code here</pre>
  names(df) <- gsub(" ", "_", names(df))</pre>
  df <- df %>%
    select(!c(fixed_acidity,free_sulfur_dioxide))
  invisible(as.factor(df$type))
  df <- df%>%
    drop_na()
  dim(df)
[1] 6497
           11
Your output to R dim(df) should be
[1] 6497
           11
```

1.3 (20 points)

Recall from STAT 200, the method to compute the t statistic for the the difference in means (with the equal variance assumption)

- 1. Using df compute the mean of quality for red and white wine separately, and then store the difference in means as a variable called diff_mean.
- 2. Compute the pooled sample variance and store the value as a variable called sp_squared.
- 3. Using sp_squared and diff_mean, compute the t Statistic, and store its value in a variable called t1.

```
#1
df$quality <- as.numeric(df$quality)
df13 <- aggregate(x= df$quality,by = list(df$type),FUN = mean)
diff_mean <- df13[2,2]-df13[1,2]
#2
n1 <- nrow(df[df$type=="white",])
n2 <- nrow(df[df$type=="red",])
var1 <- var(df$quality[df$type=="white"])
var2 <- var(df$quality[df$type=="red"])
sp_squared <- ((n1-1)*var1 + (n2-1)*var2) / (n1+n2-2)
#3
t1 <- (diff_mean-0)/(sqrt(sp_squared)*sqrt((1/n1)+(1/n2)))
t1</pre>
```

[1] 9.68565

1.4 (10 points)

Equivalently, R has a function called t.test() which enables you to perform a two-sample t-Test without having to compute the pooled variance and difference in means.

Perform a two-sample t-test to compare the quality of white and red wines using the t.test() function with the setting var.equal=TRUE. Store the t-statistic in t2.

```
df14white <- df %>%
  filter(type == "white") %>%
  pull(quality)
df14red <- df %>%
  filter(type == "red") %>%
  pull(quality)
t_test <- t.test(df14white, df14red,var.equal=TRUE)
t2 <- t_test$statistic</pre>
```

1.5 (5 points)

Fit a linear regression model to predict quality from type using the lm() function, and extract the t-statistic for the type coefficient from the model summary. Store this t-statistic in t3.

```
fit <- lm(df$quality~df$type)
t3 <- tail(coef(summary(fit))[, "t value"],1)</pre>
```

1.6 (5 points)

Print a vector containing the values of t1, t2, and t3. What can you conclude from this? Why?

Calculating t statistic by all mesures should yield same result. Because all three methods are using the same function in the background.

Question 2



2.1 (5 points)

Fit a linear regression model with all predictors against the response variable quality. Use the broom::tidy() function to print a summary of the fitted model. What can we conclude from the model summary?

We can conclude that variables volatile_acidity,residual_sugar,chlorides,density,ph,sulphates,alcohol,quality are significant, while the others are not. Total_sulfur_dioxide and citric_acid are not good predictors of the quality, given all other variables are accounted for.

```
df$volatile_acidity <- as.numeric(df$volatile_acidity)</pre>
  df$citric_acid <- as.numeric(df$citric_acid)</pre>
  df$residual_sugar <- as.numeric(df$residual_sugar)</pre>
  df$chlorides <- as.numeric(df$chlorides)</pre>
  df$total_sulfur_dioxide <- as.numeric(df$total_sulfur_dioxide)</pre>
  df$density <- as.numeric(df$density)</pre>
  df$pH <- as.numeric(df$pH)</pre>
  df$sulphates <- as.numeric(df$sulphates)</pre>
  df$alcohol <- as.numeric(df$alcohol)</pre>
  df$quality <- as.numeric(df$quality)</pre>
  df$type <- as.factor(df$type)</pre>
  sapply(df, class)
    volatile_acidity
                               citric_acid
                                                  residual_sugar
           "numeric"
                                 "numeric"
                                                       "numeric"
           chlorides total_sulfur_dioxide
                                                         density
           "numeric"
                                 "numeric"
                                                       "numeric"
                                 sulphates
                                                         alcohol
                  рΗ
                                 "numeric"
                                                       "numeric"
           "numeric"
             quality
                                      type
           "numeric"
                                  "factor"
  full_model <- lm(quality~.,df)</pre>
  broom::tidy(full_model)
# A tibble: 11 x 5
                           estimate std.error statistic p.value
  term
   <chr>
                              <dbl>
                                        <dbl>
                                                   <dbl>
                                                            <dbl>
                          57.5
                                     9.33
                                                   6.17 7.44e-10
 1 (Intercept)
2 volatile acidity
                          -1.61
                                     0.0806
                                                 -20.0 4.07e-86
                                                   0.347 7.28e- 1
3 citric_acid
                          0.0272
                                     0.0783
4 residual_sugar
                          0.0451
                                     0.00416
                                                  10.8 3.64e-27
5 chlorides
                          -0.964
                                     0.333
                                                  -2.90 3.78e- 3
6 total_sulfur_dioxide -0.000329 0.000262
                                                  -1.25 2.10e- 1
7 density
                         -55.2
                                     9.32
                                                  -5.92 3.34e- 9
                          0.188
                                                   2.85 4.38e- 3
Hq 8
                                     0.0661
                                                   8.73 3.21e-18
9 sulphates
                          0.662
                                     0.0758
10 alcohol
                          0.277
                                     0.0142
                                                  19.5 1.87e-82
11 typewhite
                          -0.386
                                     0.0549
                                                  -7.02 2.39e-12
```

```
2.2 (10 points)
```

Fit two **simple** linear regression models using lm(): one with only citric_acid as the predictor, and another with only total_sulfur_dioxide as the predictor. In both models, use quality as the response variable. How does your model summary compare to the summary from the previous question?

The two variables alone are significant predictors of quality. The previous question concludes that they are not.

```
model_citric <- lm(quality~citric_acid,df)</pre>
  summary(model_citric)
Call:
lm(formula = quality ~ citric_acid, data = df)
Residuals:
             1Q Median
                             3Q
    Min
                                    Max
-2.9938 -0.7831 0.1552 0.2426 3.1963
Coefficients:
            Estimate Std. Error t value Pr(>|t|)
(Intercept) 5.65461
                        0.02602 217.343
                                          <2e-16 ***
citric_acid 0.51398
                        0.07429
                                  6.918
                                           5e-12 ***
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 0.8701 on 6495 degrees of freedom
Multiple R-squared: 0.007316, Adjusted R-squared: 0.007163
F-statistic: 47.87 on 1 and 6495 DF, p-value: 5.002e-12
  model_sulfur <- lm(quality~total_sulfur_dioxide,df)</pre>
  summary(model_sulfur)
Call:
lm(formula = quality ~ total_sulfur_dioxide, data = df)
Residuals:
             1Q Median
                             3Q
                                    Max
-2.8866 -0.7971 0.1658 0.2227 3.1965
```

Coefficients:

```
Estimate Std. Error t value Pr(>|t|)
(Intercept) 5.8923848 0.0246717 238.831 < 2e-16 ***
total_sulfur_dioxide -0.0006394 0.0001915 -3.338 0.000848 ***
---
Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
```

Residual standard error: 0.8726 on 6495 degrees of freedom Multiple R-squared: 0.001713, Adjusted R-squared: 0.001559 F-statistic: 11.14 on 1 and 6495 DF, p-value: 0.000848

1 Statistic. 11.14 on 1 and 0430 bi, p value. 0.000040

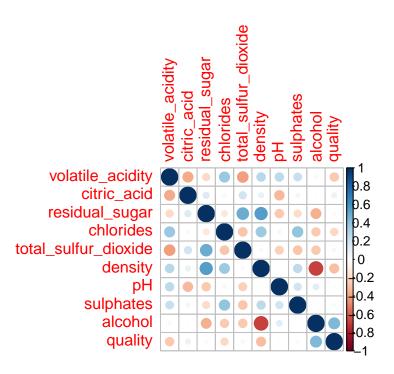
2.3 (5 points)

Visualize the correlation matrix of all numeric columns in df using corrplot()

```
sapply(df, class)
```

residual_sugar	citric_acid	volatile_acidity
"numeric"	"numeric"	"numeric"
density	total_sulfur_dioxide	chlorides
"numeric"	"numeric"	"numeric"
alcohol	sulphates	рН
"numeric"	"numeric"	"numeric"
	type	quality
	"factor"	"numeric"

df23 <- df%>%
 select(!c(type))
dfcor = cor(df23)
corrplot(dfcor)



2.4 (5 points)

Compute the variance inflation factor (VIF) for each predictor in the full model using vif() function. What can we conclude from this?

Most predictors have a low vif value, where as type and density has a vif value of more than 5, meaning that hey are highly correlated and multicollinearity should be considered and the variables should be dropped.

vif(full_model)

volatile_acidity	citric_acid	residual_sugar
2.103853	1.549248	4.680035
chlorides	total_sulfur_dioxide	density
1.625065	2.628534	9.339357
рН	sulphates	alcohol
1.352005	1.522809	3.419849
type		
6.694679		

Question 3



Variable selection

3.1 (5 points)

Run a backward stepwise regression using a full_model object as the starting model. Store the final formula in an object called backward_formula using the built-in formula() function in R

```
backward <- step(full_model,direction='backward', scope=formula(full_model), trace=0)
backward_formula <- formula(backward)
backward_formula

quality ~ volatile_acidity + residual_sugar + chlorides + density +
    pH + sulphates + alcohol + type</pre>
```

3.2 (5 points)

Run a forward stepwise regression using a null_model object as the starting model. Store the final formula in an object called forward_formula using the built-in formula() function in R

3.3 (10 points)

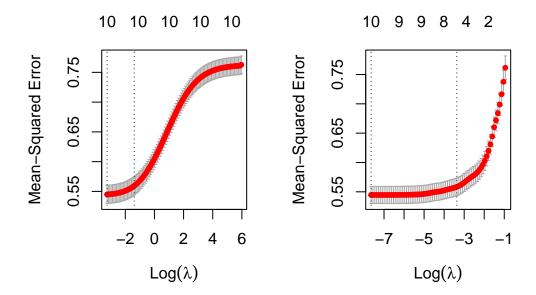
- 1. Create a y vector that contains the response variable (quality) from the df dataframe.
- 2. Create a design matrix X for the full_model object using the make_model_matrix() function provided in the Appendix.
- 3. Then, use the cv.glmnet() function to perform LASSO and Ridge regression with X and y.

```
y <- df$quality
X <- make_model_matrix(full_model)
lasso <- cv.glmnet(X,y,alpha=1)
ridge <- cv.glmnet(X,y,alpha=0)</pre>
```

Create side-by-side plots of the ridge and LASSO regression results. Interpret your main findings.

I have found that The minimum MSE is achieved when λ =0.03879736 for ridge regression, and the minimum MSE is achieved when λ =0.0006323003 for LASSO regression.

```
par(mfrow=c(1, 2))
plot(ridge)
plot(lasso)
```



ridge\$lambda.min

[1] 0.03879736

lasso\$lambda.min

[1] 0.000478312

3.4 (5 points)

Print the coefficient values for LASSO regression at the lambda.1se value? What are the variables selected by LASSO?

Store the variable names with non-zero coefficients in lasso_vars, and create a formula object called lasso_formula using the make_formula() function provided in the Appendix.

```
lasso$lambda.1se
```

[1] 0.034538

```
lasso_vars <- coef(lasso, s = "lambda.1se")
lasso_vars</pre>
```

11 x 1 sparse Matrix of class "dgCMatrix"

(Intercept) 5.81837771 volatile_acidity -0.18833502

citric_acid

residual_sugar 0.03434200

chlorides .
total_sulfur_dioxide .
density .
pH .

 sulphates
 0.04909968

 alcohol
 0.35886559

type .

```
lasso_formula <- make_formula(lasso_vars)</pre>
```

3.5 (5 points)

Print the coefficient values for ridge regression at the lambda.1se value? What are the variables selected here?

Store the variable names with non-zero coefficients in ridge_vars, and create a formula object called ridge_formula using the make_formula() function provided in the Appendix.

```
ridge$lambda.1se
```

[1] 0.2493923

type

```
ridge_vars <- coef(ridge, s = "lambda.1se")
ridge_vars</pre>
```

11 x 1 sparse Matrix of class "dgCMatrix"

(Intercept) 5.85958721 volatile_acidity -0.16471804 citric_acid 0.02178118 residual_sugar 0.08858405 chlorides -0.04745954 total_sulfur_dioxide -0.03952828 -0.08354208 density рΗ 0.02378773 sulphates 0.07807954 alcohol 0.25818761

```
ridge_formula <- make_formula(ridge_vars)</pre>
```

-0.05466274

3.6 (10 points)

What is the difference between stepwise selection, LASSO and ridge based on you analyses above?

The stepwise regressions generated the same 8-predictor model, Lasso formula included the least terms, Ridge formula included all 8 predictors, but many of them has very small slope.

```
summary(backward)
```

Call:

```
lm(formula = quality ~ volatile_acidity + residual_sugar + chlorides +
    density + pH + sulphates + alcohol + type, data = df)
```

Residuals:

```
Min 1Q Median 3Q Max -3.3317 -0.4695 -0.0422 0.4563 3.1248
```

Coefficients:

```
Estimate Std. Error t value Pr(>|t|)
                                    6.539 6.67e-11 ***
(Intercept)
                57.225176
                          8.751451
volatile_acidity -1.626323 0.074814 -21.738 < 2e-16 ***
                          0.003998 11.070 < 2e-16 ***
residual_sugar
                0.044254
chlorides
                density
               -54.876246 8.714620 -6.297 3.23e-10 ***
                 0.175885 0.062554
рΗ
                                    2.812 0.00494 **
sulphates
                 0.652336
                          0.075353
                                    8.657 < 2e-16 ***
                 0.280731
alcohol
                          0.013231 21.217 < 2e-16 ***
                -0.417595
                          0.046578 -8.966 < 2e-16 ***
typewhite
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Residual standard error: 0.737 on 6488 degrees of freedom Multiple R-squared: 0.2885, Adjusted R-squared: 0.2877 F-statistic: 328.9 on 8 and 6488 DF, p-value: < 2.2e-16

```
summary(forward)
```

Call:

```
lm(formula = quality ~ alcohol + volatile_acidity + sulphates +
    residual_sugar + type + density + chlorides + pH, data = df)
```

```
ЗQ
           1Q Median
   Min
                               Max
-3.3317 -0.4695 -0.0422 0.4563 3.1248
Coefficients:
               Estimate Std. Error t value Pr(>|t|)
(Intercept)
               57.225176 8.751451 6.539 6.67e-11 ***
alcohol
                volatile_acidity -1.626323 0.074814 -21.738 < 2e-16 ***
                sulphates
residual_sugar
               -0.417595 0.046578 -8.966 < 2e-16 ***
typewhite
              -54.876246 8.714620 -6.297 3.23e-10 ***
density
chlorides
               -0.950667
                          0.329461 -2.886 0.00392 **
Нα
                0.175885
                          0.062554 2.812 0.00494 **
Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
Residual standard error: 0.737 on 6488 degrees of freedom
Multiple R-squared: 0.2885,
                           Adjusted R-squared: 0.2877
F-statistic: 328.9 on 8 and 6488 DF, p-value: < 2.2e-16
  lasso_formula
quality ~ 5.81837771279066 + -0.188335020240248 + 0 + 0.0343420015225683 +
   0 + 0 + 0 + 0 + 0 + 0.0490996756976685 + 0.358865586117007 +
<environment: 0x7f88e4a4c8d0>
  ridge_formula
quality ~ 5.8595872105273 + -0.164718041565637 + 0.0217811819274036 +
   0.0885840504798778 + -0.0474595383202974 + -0.0395282781377516 +
   -0.0835420820358288 + 0.0237877298703364 + 0.0780795437330479 +
   0.258187612588152 + -0.0546627412811334
<environment: 0x7f88ea4ddca0>
```

Residuals:

Question 4



9 70 points

Variable selection

4.1 (5 points)

Excluding quality from df we have 10 possible predictors as the covariates. How many different models can we create using any subset of these 10 coavriates as possible predictors? Justify your answer.

There could be 3628801 models. (Factorial of 10 gives the permutation of the variables, and +1 for the null model)

```
factorial(10)
```

[1] 3628800

4.2 (20 points)

Store the names of the predictor variables (all columns except quality) in an object called x_vars.

```
x_vars <- colnames(df %>% select(-quality))
```

Use:

- the combn() function (built-in R function) and
- the make_formula() (provided in the Appendix)

to generate all possible linear regression formulas using the variables in x_vars. This is most optimally achieved using the map() function from the purrr package.

```
length42 <- 1:length(x_vars)</pre>
formulas <- map(</pre>
  length42,
  (x)
```

```
vars <- combn(x_vars,x) # Insert code here
   map(split(vars,rep(1:ncol(vars),each=nrow(vars))), make_formula) # Insert code here
}
) %>% unlist()
```

If your code is right the following command should return something along the lines of:

```
sample(formulas, 4) %>% as.character()

[1] "quality ~ chlorides + total_sulfur_dioxide + density"
[2] "quality ~ residual_sugar + chlorides + total_sulfur_dioxide + density + pH + type"
[3] "quality ~ citric_acid"
[4] "quality ~ citric_acid + chlorides + pH + type"

# Output:
# [1] "quality ~ volatile_acidity + residual_sugar + density + pH + alcohol"
# [2] "quality ~ citric_acid"
# [3] "quality ~ volatile_acidity + citric_acid + residual_sugar + total_sulfur_dioxide + # [4] "quality ~ citric_acid + chlorides + total_sulfur_dioxide + pH + alcohol + type"
```

4.3 (10 points)

Use map() and lm() to fit a linear regression model to each formula in formulas, using df as the data source. Use broom::glance() to extract the model summary statistics, and bind them together into a single tibble of summaries using the bind_rows() function from dplyr.

```
models <- map(formulas,~lm(quality~.,data=df)) # Insert your code here
#summaries <- map(data.frame(models), bind_rows(broom::glance)) # Insert your code here</pre>
```

4.4 (5 points)

Extract the adj.r.squared values from summaries and use them to identify the formula with the *highest* adjusted R-squared value.

```
#summaries%>%
    #select(adj.r.squared)
```

Store resulting formula as a variable called rsq_formula.

```
rsq_formula <- ... # Insert your code
```

4.5 (5 points)

Extract the AIC values from summaries and use them to identify the formula with the *lowest* AIC value.

```
... # Insert your code here
```

Store resulting formula as a variable called aic_formula.

```
aic_formula <- ... # Insert your code</pre>
```

4.6 (15 points)

Combine all formulas shortlisted into a single vector called final_formulas.

```
null_formula <- formula(null_model)
full_formula <- formula(full_model)

final_formulas <- c(
   null_formula,
   full_formula,
   backward_formula,
   forward_formula,
   ridge_formula,
   ridge_formula,
   #rsq_formula,
   #aic_formula
)</pre>
```

- Are aic_formula and rsq_formula the same? How do they differ from the formulas shortlisted in question 3?
- Which of these is more reliable? Why?
- If we had a dataset with 10,000 columns, which of these methods would you consider for your analyses? Why?

4.7 (10 points)

Use map() and glance() to extract the sigma, adj.r.squared, AIC, df, and p.value statistics for each model obtained from final_formulas. Bind them together into a single data frame summary_table. Summarize your main findings.

```
summary_table <- map(
  final_formulas,
  \(x) ... # Insert your code here
) %>% bind_rows()

summary_table %>% knitr::kable()
```

Appendix

Convenience function for creating a formula object

The following function which takes as input a vector of column names x and outputs a formula object with quality as the response variable and the columns of x as the covariates.

```
make_formula <- function(x){
   as.formula(
     paste("quality ~ ", paste(x, collapse = " + "))
   )
}

# For example the following code will
   # result in a formula object
   # "quality ~ a + b + c"
   make_formula(c("a", "b", "c"))

quality ~ a + b + c
<environment: 0x7f88e5f42680>
```

Convenience function for glmnet

The make_model_matrix function below takes a formula as input and outputs a rescaled model matrix X in a format amenable for glmnet()

```
make_model_matrix <- function(formula){
    X <- model.matrix(formula, df)[, -1]
    cnames <- colnames(X)
    for(i in 1:ncol(X)){
        if(!cnames[i] == "typewhite"){
            X[, i] <- scale(X[, i])
        } else {
            colnames(X)[i] <- "type"
        }
    }
    return(X)
}</pre>
```

i Session Information

Print your R session information using the following command

```
sessionInfo()
```

```
R version 4.2.1 (2022-06-23)

Platform: x86_64-apple-darwin17.0 (64-bit)

Running under: macOS Big Sur ... 10.16

Matrix products: default

BLAS: /Library/Frameworks/R.framework/Versions/4.2/Resources/lib/libRblas.0.dylib

LAPACK: /Library/Frameworks/R.framework/Versions/4.2/Resources/lib/libRlapack.dylib
```

locale:

[1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8

attached base packages:

[1] stats graphics grDevices datasets utils methods base

other attached packages:

- [1] janitor_2.2.0 corrplot_0.92 glmnet_4.1-6 Matrix_1.4-1 car_3.1-1
- [6] carData_3.0-5 purrr_1.0.1 dplyr_1.1.0 tidyr_1.3.0 readr_2.1.4

loaded via a namespace (and not attached):

```
[1] Rcpp_1.0.10
                     pillar_1.8.1
                                      compiler_4.2.1
                                                       iterators_1.0.14
[5] tools_4.2.1
                     digest_0.6.31
                                      timechange_0.2.0 lubridate_1.9.2
[9] jsonlite_1.8.4
                     evaluate_0.20
                                      lifecycle_1.0.3 tibble_3.1.8
[13] lattice_0.20-45 pkgconfig_2.0.3 rlang_1.0.6
                                                       foreach_1.5.2
[17] cli_3.6.0
                     yaml_2.3.7
                                      xfun_0.37
                                                       fastmap_1.1.1
[21] withr_2.5.0
                                      knitr_1.42
                     stringr_1.5.0
                                                       generics_0.1.3
[25] vctrs_0.5.2
                     hms_1.1.2
                                      grid_4.2.1
                                                       tidyselect_1.2.0
[29] snakecase_0.11.0 glue_1.6.2
                                      R6_2.5.1
                                                       fansi_1.0.4
[33] survival_3.3-1
                     rmarkdown_2.20
                                      tzdb_0.3.0
                                                       magrittr_2.0.3
[37] backports_1.4.1 splines_4.2.1
                                      codetools_0.2-18 ellipsis_0.3.2
[41] htmltools_0.5.4 abind_1.4-5
                                      shape_1.4.6
                                                       renv_0.16.0-53
[45] utf8_1.2.3
                     stringi_1.7.12
                                      broom_1.0.3
```