

## SPATIAL VARIABILITY OF NITRATE CONCENTRATIONS UNDER DIVERSE CONDITIONS IN TRIBUTARIES TO A LAKE WATERSHED<sup>1</sup>

Heather E. Golden, Elizabeth W. Boyer, Michael G. Brown, S. Thomas Purucker, and René H. Germain<sup>2</sup>

**ABSTRACT:** Nitrate-nitrogen ( $\text{NO}_3\text{-N}$ ) concentrations in stream water often respond uniquely to changes in inter-annual conditions (e.g., biological N uptake and precipitation) in individual catchments. In this paper, we assess (1) how the spatial distribution of  $\text{NO}_3\text{-N}$  concentrations varies across a dense network of nonnested catchments and (2) how relationships between multiple landscape factors [within whole catchments and hydrologically sensitive areas (HSAs) of the catchments] and stream  $\text{NO}_3\text{-N}$  are expressed under a variety of annual conditions. Stream  $\text{NO}_3\text{-N}$  data were collected during two synoptic sampling events across >55 tributaries and two synoptic sampling periods with >11 tributaries during summer low flow periods. Sample tributaries drain mixed land cover watersheds ranging in size from 0.150 to 312 km<sup>2</sup> and outlet directly to Cayuga Lake, New York. Changes in  $\text{NO}_3\text{-N}$  concentration ratios between each sampling event suggest a high degree of spatial heterogeneity in catchment response across the Cayuga Lake Watershed, ranging from 0.230 to 61.4. Variations in  $\text{NO}_3\text{-N}$  concentrations within each of the large synoptic sampling events were also high, ranging from 0.040 to 8.7 mg  $\text{NO}_3\text{-N/l}$  (March) and 0.090 to 15.5 mg  $\text{NO}_3\text{-N/l}$  (October). Although Pearson correlation coefficients suggest that this variability is related to multiple landscape factors during all four sampling events, partial correlations suggest percentage of row crops in the catchments as the only similar factor in March and October and catchment area as the only factor during summer low flows. Further, the strength of the relationships is typically lower in the HSAs of catchment. Advancing current understanding of such variations and relationships to landscape factors across multiple catchments – and under a variety of biogeochemical and hydrological conditions – is important, as (1) nitrate continues to be employed as an indicator of regional aquatic ecosystem health and services and (2) a unified framework approach for understanding individual catchment processes is a rapidly evolving focus for catchment-based science and management.

(KEY TERMS: nitrate; spatial variations; multiple catchments; landscape; land cover; Cayuga Lake; New York.)

Golden, Heather E., Elizabeth W. Boyer, Michael G. Brown, S. Thomas Purucker, and René H. Germain, 2009. Spatial Variability of Nitrate Concentrations Under Diverse Conditions in Tributaries to a Lake Watershed. *Journal of the American Water Resources Association* (JAWRA) 45(4):945-962. DOI: 10.1111/j.1752-1688.2009.00338.x

<sup>1</sup>Paper No. JAWRA-08-0119-P of the *Journal of the American Water Resources Association* (JAWRA). Received June 17, 2008; accepted February 19, 2009. © 2009 American Water Resources Association. No claim to original U.S. government works. **Discussions are open until six months from print publication.**

<sup>2</sup>Respectively, Physical Scientist and Ecologist (Golden and Purucker), Ecosystems Research Division, National Environmental Exposure Laboratory, U.S. Environmental Protection Agency, 960 College Station Road, Athens, Georgia 30605; Associate Professor (Boyer), School of Forest Resources and Penn State Institutes of Energy and the Environment, The Pennsylvania State University, 304 Forest Resources Building, University Park, Pennsylvania 16802; Research Associate (Brown), Department of Horticulture, Cornell University, 123 Plant Sciences Bldg., Ithaca, New York 14853; and Associate Professor (Germain), Department of Forest and Natural Resources Management, State University of New York College of Environmental Science and Forestry, Marshall Hall, Syracuse, New York 13210 (E-Mail/Golden: Golden.Heather@epa.gov).

## INTRODUCTION

Nitrogen pollution has emerged as a globally recognized water quality concern, generating interest from scientific, management, and policy communities. Anthropogenic sources of reactive nitrogen (N) to the landscape have resulted in elevated loadings of N to surface waters around the world, from streams and lakes (Carpenter *et al.*, 1998; Driscoll *et al.*, 2001; Aber *et al.*, 2003; Boyer *et al.*, 2006a) to major river basins (Green *et al.*, 2004; Boyer *et al.*, 2006b; Alexander *et al.*, 2007b). The ecological effects of this surplus N delivery to surface waters can be severe, leading to acidification of streams and lakes, and over-enrichment of estuaries (Bricker *et al.*, 1999; Paerl *et al.*, 2002; Driscoll *et al.*, 2003).

In environmental settings ranging from pristine to human-modified, organic forms of N typically dominate fluxes in streams and rivers (Perakis and Hedin, 2002; Schoonover and Lockaby, 2006). However, anthropogenic impacts on the N cycle are often marked by increased contributions of  $\text{NO}_3^-$  to the total loads of N in rivers (Caraco and Cole, 1999; Seitzinger *et al.*, 2002). Nitrate frequently emerges as the dominant form of inorganic N in surface waters (Creed and Band, 1998b; Boyer *et al.*, 2006a) and is typically the primary component of the inorganic N fluxes issuing from watersheds, even in forested settings (Campbell *et al.*, 2004; Ito *et al.*, 2005) due to inputs of anthropogenic nitrogen to otherwise “pristine” locations via atmospheric deposition. Furthermore,  $\text{NO}_3^-$  is assessed as one of a suite of indicators of aquatic ecosystem health, which is often extended to the evaluation of ecosystem services.

Links between the spatial and temporal variations of the amounts of  $\text{NO}_3^-$  in surface water to landscape attributes are well established, often with a focus on surface runoff associated with a particular land use or land cover (Jordan and Weller, 1996; Jordan *et al.*, 1997; Herlihy *et al.*, 1998; Arheimer and Lidén, 2000; Boyer *et al.*, 2002; Franklin *et al.*, 2002; Strayer *et al.*, 2003; Buck *et al.*, 2004; Burns *et al.*, 2005; Cuevas *et al.*, 2006; Poor and McDonnell, 2007). Several studies have also revealed diverse contributions of  $\text{NO}_3^-$  issuing from bedrock to surface waters (Burns *et al.*, 1998b; Holloway *et al.*, 1998; Böhlke, 2003; Puckett and Hughes, 2005), particularly during drought or low flow conditions. These deep subsurface contributions of  $\text{NO}_3^-$  reflect leaching from landscape sources (Jordan *et al.*, 1997; Schilling and Zhang, 2004; Benson *et al.*, 2006) and geological sources of  $\text{NO}_3^-$  (Holloway *et al.*, 1998; Holloway and Dahlgren, 1999). For example, a host of literature demonstrate a particularly strong association between an increased proportion of row crops and  $\text{NO}_3^-$  in both ground and surface water

systems across multiple scales (Jordan *et al.*, 1997; Arheimer and Lidén, 2000; McIsaac *et al.*, 2001; Benson *et al.*, 2006; Schilling and Spooner, 2006; Schoonover and Lockaby, 2006; Johnson *et al.*, 2007; Poor and McDonnell, 2007; Kang *et al.*, 2008). Few studies, however, reveal how relationships between landscape factors and  $\text{NO}_3^-$  in the stream vary throughout an annual cycle (Johnson *et al.*, 1997). Work that begins to advance our understanding of such patterns is needed.

Several detailed small-catchment studies, often conducted in relatively undisturbed forested settings [such as the United States (U.S.) Geological Survey Water, Energy, and Biogeochemical Balance and Hydrologic Benchmark watersheds and other well-instrumented sites], have advanced current understanding of the complex interactions among seasonal variations in water supply and transport, biologically available nitrogen in soils, and amounts of instream nitrate (Mitchell *et al.*, 1996, 2003; Burns, 1998; Petrone *et al.*, 2007). Studies in more diverse land use settings often focus on holding hydrological conditions relatively constant, such as investigating variations in amounts of  $\text{NO}_3^-$  in surface waters and linkages with landscape characteristics during base flow or monthly and annually averaged flows across multiple events (Herlihy *et al.*, 1998; Jones *et al.*, 2001; Wayland *et al.*, 2003; Kang *et al.*, 2008), or assessing export of  $\text{NO}_3^-$  to stream water during storm events (Jordan *et al.*, 1997; Poor and McDonnell, 2007). However, few synoptic or long-term studies specifically examine the spatial distribution of  $\text{NO}_3\text{-N}$  concentrations, with links to landscape attributes, under a variety of seasonal (biogeochemical and hydrological) conditions and across a broad gradient of catchments. Such research is highly relevant for structuring studies related to modeling and monitoring catchment processes and implementing management efforts in multiple catchments issuing to a single water body (e.g., a lake, river, or estuary) and is particularly important as the availability of  $\text{NO}_3^-$  in surface soils – the primary source of  $\text{NO}_3^-$  for hydrologic transport – fluctuates with temperature and moisture controls (Alexander *et al.*, 2007a; Knoepp and Vose, 2007), plant uptake of  $\text{NO}_3^-$  (Arheimer *et al.*, 1996), and applications of nitrogenous fertilizers (Bechtold *et al.*, 2003). Therefore, an assessment of the spatial distribution of  $\text{NO}_3^-$  concentrations in multiple catchments across varying conditions throughout the year warrants further exploration.

In this paper, we investigate (1) how the spatial distribution of  $\text{NO}_3\text{-N}$  concentrations varies across a dense network of nonnested catchments and (2) how relationships between multiple landscape factors and stream  $\text{NO}_3\text{-N}$  are expressed under different biogeochemical and hydrological conditions throughout one year. We assess  $\text{NO}_3\text{-N}$  concentrations using

several synopses – or snapshots – samples at the outlets of a large, diverse subset (from  $n = 11$  to 64) of catchments draining to Cayuga Lake in Central New York State. Spatial variations in stream  $\text{NO}_3\text{-N}$  concentrations, compared among four synoptic sampling events, are examined via changes in concentration ratios, Moran's  $I$  correlograms, and geographic mapping techniques. Pearson correlations and partial correlations are employed to investigate relationships between multiple landscape attributes and stream  $\text{NO}_3\text{-N}$  concentrations during each sampling period. The findings presented here are important for advancing the current understanding of temporal differences in the spatial variations of surface water  $\text{NO}_3\text{-N}$  concentrations across multiple unique catchments. This study is particularly relevant for national and international watershed biogeochemical research and management, as the spatial distribution of  $\text{NO}_3\text{-N}$  in surface waters continues to be linked to broader conclusions, such as the regional status of aquatic ecosystem indicators and services.

## STUDY AREA

Cayuga Lake Watershed (CLW) is located in the Finger Lakes Region of New York State. Cayuga

Lake is the longest (61.5 km), widest (2.8 km wide), and second largest volume freshwater lake in the state outside of the Great Lakes system. Its deepest point reaches  $\sim 40.4$  m. CLW is  $\sim 1,840$   $\text{km}^2$  (Figure 1), excluding the surface area of the lake ( $\sim 173$   $\text{km}^2$ ) and an additional 2,018  $\text{km}^2$  contributing drainage area from Seneca and Keuka Lakes to Cayuga Lake's northern outlet (GFLRPC, 2004). The lake outlet flows northward to the Oswego River system emptying into Lake Ontario, one of the five Great Lakes in the U.S. A dense network of over 100 tributary subcatchments drains CLW. The study subcatchments ( $n = 10\text{--}64$ ) of Cayuga Lake range from 0.15 to 312  $\text{km}^2$  and comprise  $\sim 77\%$  ( $n = 64$ ) of the total CLW drainage area.

The climate of the study area is humid continental, which is characterized by warm summers and long, cold winters (GFLRPC, 2004). Average annual precipitation (rain and snow) is  $\sim 89$  cm and is relatively evenly distributed throughout the year (Ithaca, New York) (NEWA, 2006). The climate of the northern part of the watershed exhibits characteristics strongly influenced by the Laurentian Great Lakes, primarily Lake Ontario (GFLRPC, 2004). Land cover within CLW (minus the lake's area) is 55% agricultural (28% pasture and 27% row crops), 26% forested, 11% urban and agricultural grasses and shrub/scrub, 5% open water and wetlands, 2% developed,  $\sim 1\%$  open water and wetlands, and less than 1% barren

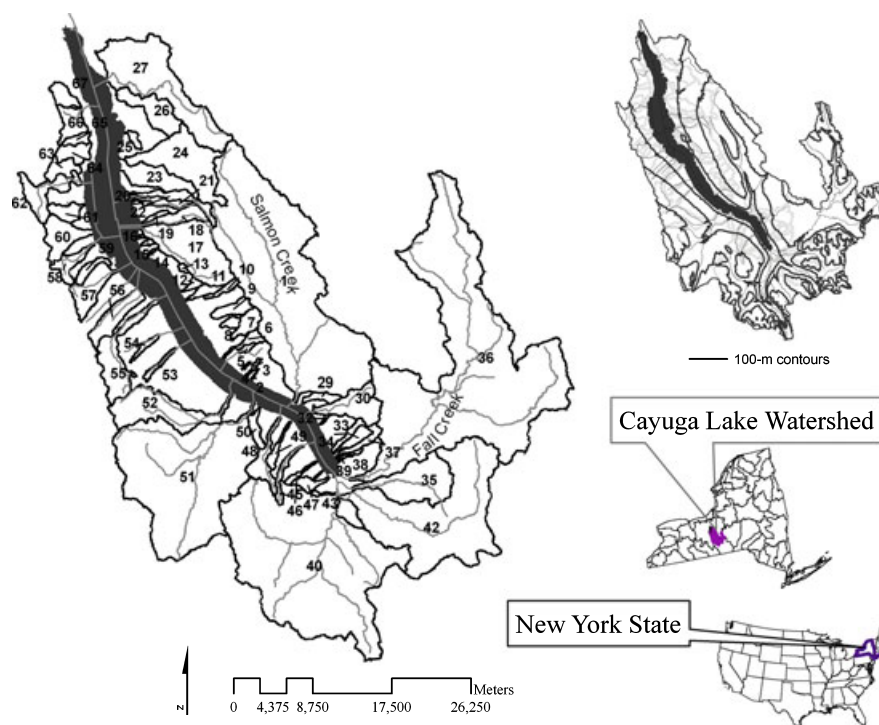


FIGURE 1. Cayuga Lake Watershed, Including the Stream Network, Sample Subcatchments, and Cayuga Lake (in black). Contours are shown in 100 meter increments.



(e.g., rocks and quarries) (MRLC, 2001). Although the land cover proportions vary across sections of the watershed, all subcatchments are predominately a mix of agriculture, forests, and developing areas (Table 1).

Elevations within CLW range from 112 to 638 m above sea level. The lake's current orientation is along a north-south gradient, with relatively low topography in the northern end trending toward a steeper gradient with deeper bedrock and sediment moving southward (Mullins and Hinchey, 1989). Glacially carved valleys in the higher elevations of the watershed create numerous steep waterfalls in the southern part of the watershed. Bedrock geology in the watershed is comprised mostly of limestone in the northern two-thirds of the watershed with Devonian and Silurian shale and sandstone in the southern portion. Surficial geology of the CLW is a variety of unsorted glacial till of varying thickness; however, the majority is comprised of poorly sorted sand-rich diamict, which is typically a coarse layer of till (GFLRPC, 2004). Till moraine is found throughout the watershed, with primarily lacustrine silt and clay of low permeability located in the northeastern portion of the watershed. Parts of the central-eastern section of the lake and southward exhibit exposed

bedrock or bedrock within one meter of the surface. Soils vary in thickness and drainage patterns across the watershed are developed over glacial till, outwash terraces, kames, and sediments (GFLRPC, 2004).

## METHODS

### *Nitrate-N Concentrations*

Synoptic sampling captures a "snapshot" of stream chemistry during a single period in time (Grayson *et al.*, 1997; Wayland *et al.*, 2003), providing a window to the spatial variability of surface water concentrations and fluxes. We collected samples at the outlet of 64 tributaries to Cayuga Lake, comprising ~77% of the CLW, during two complete synoptic sampling periods under varying hydrological and biogeochemical conditions. The sampling events included: (1) March 23-24, 2005 [*Synoptic 1* (S1),  $n = 64$ , vegetative dormancy; early spring, prior to the final snowmelt but with minimal remaining snow (~5-10 cm)], and (2) October 26 and 28 [*Synoptic 4* (S4),  $n = 55$ , a wet period following the growing season, autumn]. We also conducted two additional sampling rounds on major streams in CLW that continued to issue base flow during the summer to capture extreme hydrological conditions, including (1) a summer dry, low flow period [*Synoptic 2* (S2), August 19, 2005,  $n = 11$ ] and (2) a day following a land-based hurricane rainfall event, which provided a brief period of elevated flow (relative to previously dry conditions) in the tributaries [*Synoptic 3* (S3), September 3, 2005,  $n = 16$ ]. During this extremely dry summer, 75-83% of streams previously sampled during the S1 synoptic event lacked surface flow connections from their outlets to Cayuga Lake.

We sampled each tributary at its outlet to Cayuga Lake at points where mixing of lake and tributary chemistry was minimized. Samples were collected from a high velocity midsection of each stream, into a precleaned 1 l high density polyethylene (HDPE) bottle that was rinsed four times with stream water at each field site. Samples were stored at 4°C until analysis. We filtered all samples using 47 mm 0.45  $\mu\text{m}$  hydrophilic polypropylene membrane filters (Pall Gelman GHP, Ann Arbor, Michigan) into 125 ml HDPE bottles (for  $\text{NO}_3^-$ ), and analyzed filtered samples for  $\text{NO}_3^-$  via ion chromatography, using a Dionex DX-500 system (Dionex, Sunnyvale, California). We delineated subcatchments draining directly to our sampling points using ArcGIS 9.0 (ESRI, Redlands, California) with an Albers canonical projection, an equal area method.

TABLE 1. Characteristics of Subcatchments for the Four Synoptic Sampling Periods.

Variable	Mean	Min	Max
Area ( $\text{km}^2$ )	24.1	0.15	312
Subbasin wet areas, HSAs <sup>1</sup> (%)	13.2	1.64	27.1
Average annual wet N deposition (kg N/ha/yr)	7.03	6.47	7.56
Pasture, hay (%)	30.7	0.00	70.0
Row crops (%)	28.6	0.00	64.9
Forested (%)	16.5	0.00	58.3
Developed (%)	3.40	0.00	39.5
Grasses (%)	7.74	0.89	58.9
Scrub-shrub (%)	5.44	0.00	25.9
Wetlands (%)	7.27	0.00	76.4
Open water (%)	0.31	0.00	2.92
Barren land (%)	0.13	0.00	5.84
Hydric soils (%)	14.3	0.00	38.6
Topographic index (TI)	9.17	7.97	10.4
Hydrologic soil group A (%)	1.76	0.00	11.0
Hydrologic soil group B (%)	35.7	0.00	84.2
Hydrologic soil group C (%)	43.6	0.70	97.3
Hydrologic soil group D (%)	9.32	0.00	42.7
Range in elevation (m)	195	31.1	521
Average slope (%)	5.09	1.61	12.4

Notes: CLW, Cayuga Lake Watershed; HSA, hydrologically sensitive area.

<sup>1</sup>Percent of subbasin above mean TI for CLW + 1 SD.

*Synoptic 1* (S1): March 23-24, 2005; *Synoptic 2* (S2): August 19, 2005; *Synoptic 3* (S3): September 3, 2005; *Synoptic 4* (S4): October 26 and 28, 2005.

### Biogeochemical and Hydrological Setting for Sampling Events

Sources and transport of nitrogen within the Fall Creek catchment (subcatchment 35) of CLW have been studied for over 30 years (Elizabeth W. Boyer, 2007, personal communication); records from this research therefore provide a context for assessing variations in the coupling of biogeochemical processes and  $\text{NO}_3\text{-N}$  concentrations in the stream within CLW. Throughout the year, precipitation is fairly evenly distributed (Figure 2, E.W. Boyer) within Fall Creek; however, seasonal controls on  $\text{NO}_3\text{-N}$  are clear, as suggested by the concomitant variations in temperate and in stream  $\text{NO}_3\text{-N}$  concentrations from 2003 to 2004. During the dormant season, plant uptake is relatively low, affording accumulation of available nitrogen in the upper soil horizons. During the summer months, plant uptake is higher, as is loss of nitrogen via denitrification, a temperature controlled process (Alexander *et al.*, 2007a).

Discharge and soil moisture was not measured within each of the 64 subcatchments; therefore, we approximate the hydrological setting throughout CLW during the year of sampling using two

approaches. First, we assessed stream discharge records from a U.S. Geological Survey stream gage closest to the mouth of Cayuga Lake (Fall Creek, Station Number 04234000, subcatchment 35) in the central portion of CLW and the National Weather Service Coop Network (NCDC, 2006) Freeville 1 NE (COOP Station 303050) precipitation station within Fall Creek catchment to represent the relative daily precipitation and discharge regime during one year of sampling within CLW (Figure 3). Although unique rainfall-runoff relationships are specific to each catchment, we confirmed the relative discharge reflected by the hydrograph throughout all catchments during each of four sampling events (e.g., highest flows during October and low to zero base flow during August).

We also evaluated the spatial distribution of total precipitation during the months of each sampling event (Daly *et al.*, 2002) as an estimate of the spatial distribution of antecedent moisture conditions and hydrological setting prior to each sampling event. (Figure 4). Precipitation was estimated using the Parameter-elevations Regressions on Independent Slopes Model (Daly *et al.*, 2002) as a monthly total, which captures the spatial patterns of precipitation at a 4 km resolution. This approach works well for

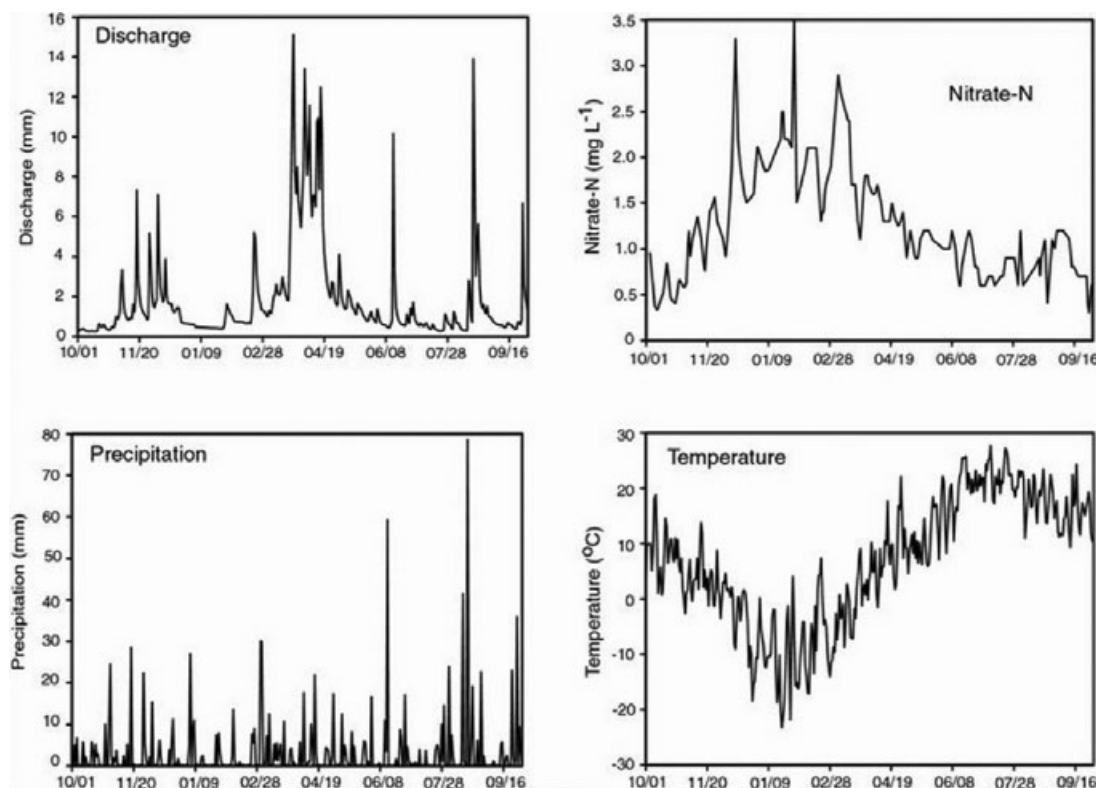


FIGURE 2. Fall Creek, New York, in Cayuga Lake Watershed: Discharge, Precipitation, Temperature, and Stream  $\text{NO}_3\text{-N}$  Measurements During 2003-2004.

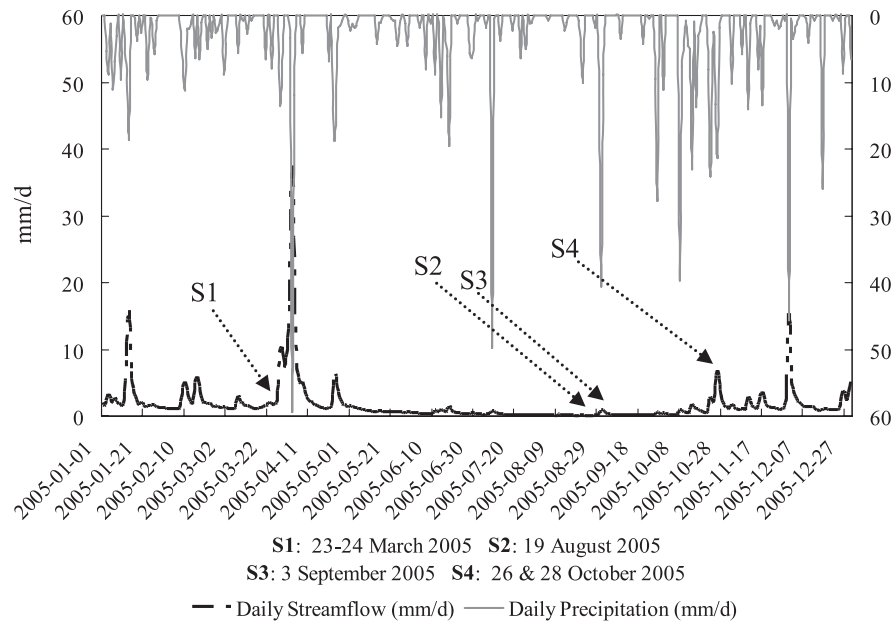


FIGURE 3. Precipitation (Freeville, New York) and Discharge (at U.S. Geological Station: Fall Creek, New York) During the 2005 Synoptic Sampling Period.

March (S1), September (S3), and October (S4) because sampling was conducted either near the end of the month (S1 and S4) or at the very beginning (S3), where the prior month's precipitation rate was assessed (i.e., August precipitation as a hydrological indicator for the September sampling). However, because sampling for S2 occurred mid-month (August 19), the spatial distribution of precipitation was unavailable. Thus, precipitation and discharge from Figure 3 served as a surrogate for hydrological setting throughout the basin during this dry period. Because

March precipitation includes only snow events, precipitation rates in Figure 4 appear lower than other periods. However, qualitatively observed soil moisture conditions during stream sampling were relatively high from the remaining early March snowpack.

#### *Landscape Attributes*

We focused on multiple indicators of landscape sources and sinks of  $\text{NO}_3\text{-N}$  and indices of hydrologi-

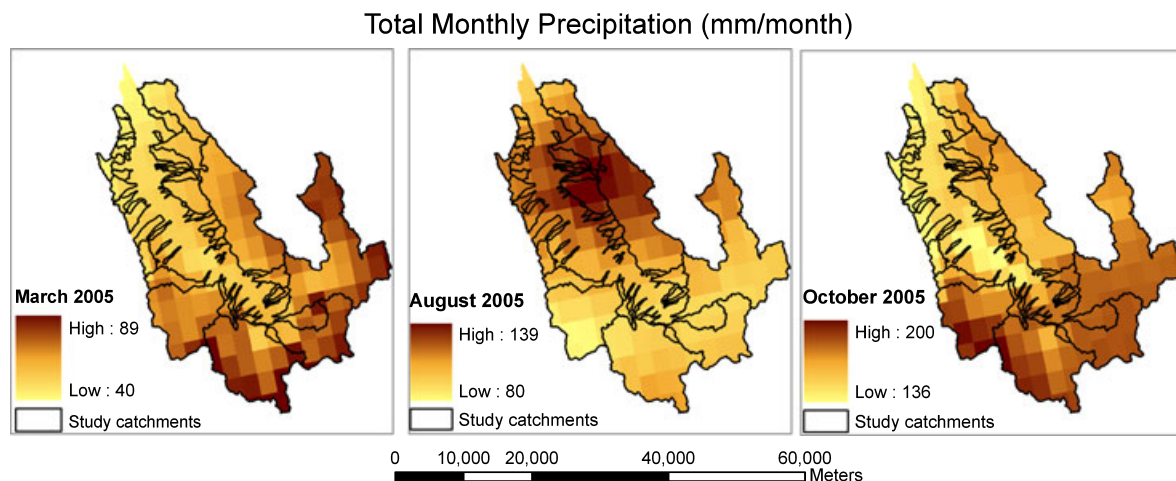


FIGURE 4. Monthly Precipitation Rates (2005) Throughout Cayuga Lake Watershed for March (S1), August (for S3 sampling), and October (S4). Precipitation is derived from modeled Parameter-elevations Regressions on Independent Slopes Model precipitation datasets at a 4 km resolution.

cal wetness and runoff to correlate with snapshots of spatially varying  $\text{NO}_3\text{-N}$  concentrations under diverse conditions. Our methods target landscape connections to stream water  $\text{NO}_3\text{-N}$  concentrations. Therefore, although  $\text{NO}_3^-$  from bedrock sources can potentially influence  $\text{NO}_3\text{-N}$  levels (Holloway *et al.*, 1998; Holloway and Dahlgren, 1999; Böhlke, 2003), particularly during the drier periods of the year, ground water analysis of  $\text{NO}_3\text{-N}$  contributions is not within the scope of this study.

We first estimated land cover data for each catchment as a proxy for sources of  $\text{NO}_3\text{-N}$ . Because most surplus N in forested land cover settings is derived from atmospheric deposition in the northeastern U.S. (Jaworski *et al.*, 1997; Boyer *et al.*, 2002), we considered leakage of  $\text{NO}_3\text{-N}$  from forested areas primarily of atmospheric origin. Other  $\text{NO}_3\text{-N}$  sources included those from fertilizer and fixation of N by crops (row crops), manure (row crops and pasture land cover), human waste sources and urban runoff (urban areas), and point sources [National Pollution Discharge Elimination System (NPDES) permits]. Land cover type for each subcatchment and for the proportion of each subcatchment in hydrologically sensitive areas (HSAs) (see Discussion below) was estimated using the 30-m resolution 2001 National Land Cover Dataset (NLCD) from the Multi-Resolution Land Cover (MRLC) Consortium (MRLC, 2001). Similar to other studies (Strayer *et al.*, 2003; Wayland *et al.*, 2003), we aggregated land cover into nine major categories, including *pasture* (NLCD 81), *row crops* (NLCD 82), *forest* (NLCD 41, 42, and 43), *developed/urban* (NLCD 22, 23, and 24), *wetlands* (NLCD 90 and 95), *grasses* (urban and rural, NLCD 21 and 71), *open water* (NLCD 11), *barren* (NLCD 31 – quarries, strip mines, etc.), and *shrub/scrub* (NLCD 52). Other input sources included estimates of contemporary annual wet N deposition for each catchment (Golden and Boyer, 2008) and sites of point source discharges throughout the basin from the U.S. Environmental Protection Agency's Pollution Compliance System, or NPDES permits (USEPA, 2006).

We investigated whether patterns of soil wetness during synoptic sampling can be correlated to nitrate loss in the landscape. Soil moisture indices have been used in multiple studies to detect areas of denitrification (see Boyer *et al.*, 2006a); therefore, we calculated the percent of hydric soil coverage within each subcatchment using data from the Soil Survey Geographic (SSURGO) database (1:15,480) (USDA-NRCS, 2006).

The mean standard topographic index (TI), which provides a metric describing the theoretical wetness potential for each point within a watershed (Beven and Kirkby, 1979) and has been applied to assess the coupling of hydrological and biogeochemical

processes (Creed and Band, 1998a; Welsch *et al.*, 2001), was calculated using methods put forth by Quinn *et al.* (1995). Calculations were applied to each grid cell of a U.S. Geological Survey National Elevation Dataset (NED) (1 arc second, or  $\sim 30$  m, resolution) clipped to the boundaries of CLW using the formula:

$$\text{TI} = \ln(a/\tan\beta), \quad (1)$$

where  $a$  is the upslope contributing area per unit contour length (or grid cell resolution) and  $\beta$  is the steepest local slope angle among the downslope directions.

The HSAs within each subcatchment were used as an index of the extent, or proportion, of the subcatchment that responds quickly to rainfall events and to examine whether the extent and landscape characteristics, particularly row crops, in hydrologically sensitive zones in the catchments regulate  $\text{NO}_3\text{-N}$  concentrations at different points throughout the year. The HSAs were calculated by (1) adding one standard deviation to the mean TI of the entire CLW to define a “sensitive” TI value and (2) estimating the percent of each subcatchment with a TI greater than this “sensitive” value. We chose to add one standard deviation to the CLW mean TI to define a standard quantitative index value to capture both the wettest areas (i.e., greater than the mean) in each catchment and throughout CLW, and areas that have a relatively high frequency compared with the tail of the normal distribution curve (e.g., data within  $\sim 34\%$  of the mean). The overall goal was to ensure that each unique catchment's hydrologically sensitive zone was included in the analysis, and as such was considered more “site specific” than the mean TI. The proportion of each subcatchment within HSAs ranged from  $\sim 2\%$  to  $27\%$  (Table 1).

We calculated the percent of each hydrologic soil group (HSG) for every subcatchment (whole and HSAs) using the SSURGO soils database (USDA-NRCS, 2006). HSGs are classified into four categories (A, B, C, and D) according to a soil's infiltration rate, subsurface transmissivity, and runoff potential (USDA-NRCS, 1986). HSG A represents soils with the highest infiltration capacity and lowest runoff potential, and D approximates the inverse. Hydrologic soils group B and C soils are bounded sequentially (via infiltrate rate, transmissivity, and runoff potential) between the HSG A and HSG D soils.

We applied other topographic variables to examine influences on  $\text{NO}_3\text{-N}$  transport. Variations in topography among subcatchments were estimated using mean slope and change in elevation values calculated from NED 30-m elevation data. We calculated mean slope for each subcatchment within the whole



catchment and HSA, and range in elevation within the entire subcatchments only.

### *Analysis: Spatial and Statistical*

Analyses of the spatial variation of NO<sub>3</sub>-N concentrations from observed synoptic data proceeded sequentially. We first analyzed the NO<sub>3</sub>-N concentration data via spatial mapping and analysis (ArcGIS 9.1; ESRI) for each synoptic event, developing a qualitative assessment of the spatial variations in NO<sub>3</sub>-N concentrations. To assess changes in NO<sub>3</sub>-N concentration between sampling events, we then assessed variations in concentration ratios between each sampling event at repeated sampling sites. Prior to any statistical analyses, Shapiro-Wilk *W* tests were conducted on NO<sub>3</sub>-N data and suggested that the data were nonnormally distributed. Natural log-transformed results changed all *p*-values from <0.05 to >0.05 ( $\alpha = 0.05$ ); therefore, all statistical analyses were conducted on natural log-transformed concentrations (Sokal and Rohlf, 1995) using SAS 9.2 (SAS Institute, Inc., Cary, North Carolina) and R 2.6.2 (The R Foundation for Statistical Computing, Vienna, Austria).

Our statistical analysis involved multiple steps. We first calculated Pearson's product-moment correlation coefficients between landscape variables, including precipitation, within the whole catchments and HSAs and NO<sub>3</sub>-N concentrations during each sampling event to identify important landscape-NO<sub>3</sub>-N relationships under varying biogeochemical and hydrological conditions. Given that an increase in one land cover factor or HSG results in a decrease of others (Van Sickle, 2003), if landscape variables exhibiting significant relationships with NO<sub>3</sub>-N concentrations were also significantly correlated with each other, we then employed a partial correlation analysis to evaluate the independent relationship of landscape factors and stream NO<sub>3</sub>-N concentrations. We used a minimum correlation of  $r \geq 0.30$  for correlations among landscape variables, a standard minimum for Pearson correlation coefficients below which even statistically significant results are considered to exhibit little – if any – linear relationship among the two variables. The partial correlation, therefore, helped clarify the relationships between landscape factors and surface water NO<sub>3</sub>-N correlations.

After identifying significant relationships between landscape factors and NO<sub>3</sub>-N concentrations, Moran's *I* correlograms and statistics were generated for NO<sub>3</sub>-N concentrations during the large synoptic events (sample size was too small for summer samples) and the significant landscape factors to assess whether and where in the watershed signifi-

cant landscape factors and NO<sub>3</sub>-N concentrations exhibit similar spatial structure. Moran's *I* correlograms measure the degree and significance of spatial autocorrelation within different distance classes. The Moran's *I* statistic used for the correlograms is similar to the Pearson's correlation coefficient; the numerator measures the covariance of all pairs of points within a given distance class, while the denominator is the maximum likelihood estimator of the variance. A significance test is implemented for each distance class (Legendre and Legendre, 1998), and if the Moran's *I* is statistically significant, spatial autocorrelation is detected. Correlations for Moran's *I* typically range from 1 (positive) to -1 (negative) correlation, although it is possible to get results exceeding this range. We analyzed stream NO<sub>3</sub>-N concentrations and significant partial correlation coefficients with distance lags of 1 km. The closer the significant Moran's *I* value is to the absolute value of 1, the higher the correlation between the distance and the variable (e.g., NO<sub>3</sub>-N concentrations). While other techniques exist to analyze the spatial autocorrelation of multiple landscape variable on amounts of NO<sub>3</sub>-N in streams (see King *et al.*, 2005), our study aims to investigate how simple relationships between landscape factors and NO<sub>3</sub>-N vary during different parts of the year.

## RESULTS

### *Snapshots of Nitrate Concentrations Across Sampling Periods*

The upper range of NO<sub>3</sub>-N concentrations during the March sampling event ( $n = 64$ ) (Tables 2 and A1) approached the 10 mg/l Maximum Contaminant Level Goal (MCL) for N established by the Safe Drinking Water Act of 1974 (Phase II and IIb for NO<sub>3</sub>-N, effective July 1992 and January 1994) (USEPA, 1994). Above this level, drinking water supplies are considered to be a threat to human health. During October sampling ( $n = 55$ ), the mean concentration was higher than that of the other sampling periods with four small subcatchments (ranging from 2.0 to 5.9 km<sup>2</sup>) meeting or exceeding MCL guidance levels of 10 mg/l during this period. Concentrations during the summer drought period (August: S2,  $n = 11$ ) were the lowest, averaging 0.32 mg/l (Tables 2 and A1). During the September (S3) sampling period ( $n = 16$ ), which followed a long drought period and was conducted one day after a hurricane-based rainfall event passed across the region resulting in a minor peak in discharge, NO<sub>3</sub>-N concentrations



TABLE 2. Mean, Minimum, Maximum, and SD of NO<sub>3</sub>-N Concentrations During Each Synoptic Sampling Event.

Variable	Sample Size (n)	NO <sub>3</sub> -N (mg/l)			
		Mean	Min	Max	SD
<i>Synoptic 1</i> (March 23-24, 2005)	64	2.11	0.04	8.68	1.99
<i>Synoptic 2</i> (August 19, 2005)	11	0.32	0.09	1.60	0.46
<i>Synoptic 3</i> (September 3, 2005)	16	0.97	0.06	4.10	1.22
<i>Synoptic 4</i> (October 26 and 28, 2005)	55	3.61	0.09	15.50	3.56

averaged slightly higher than the previous sampling event (0.97 mg/l). Daily base flow averaged  $\sim 0.080$  mm/d ( $0.30 \text{ m}^3/\text{s}$ ) in Fall Creek one month prior to the event and peaked at  $0.95 \text{ mm/d}$  ( $3.50 \text{ m}^3/\text{s}$ ) for two days following (Figure 3).

#### *Spatial Variations of Nitrate-N Concentrations Under Different Inter-Annual Settings*

Nitrate concentrations throughout CLW respond dissimilarly to changes in antecedent moisture and seasonal availability soil NO<sub>3</sub>-N during both the large sampling events (S1 and S4) and summer low flow events (S2 and S3). Figure 5 illustrates the magnitude of changes and the spatial distribution of NO<sub>3</sub>-N concentrations around the lake from 30 catchments exhibiting the highest concentrations during the large synoptic events (S1 and S4), and Figure 6 shows the relative change in NO<sub>3</sub>-N concentrations among the 11 catchments sampled during each of the four sampling events. Nine of the 11 catchments sampled during each period (Figure 6) have higher concentrations during March and October compared with summer sampling events; however, two catchments (i.e., catchments 56 and 60) in the northwest portion of CLW, exhibit relatively high concentrations during the summer low flow period ( $\sim 0.070$ – $0.090 \text{ mm/d}$  or  $0.27$ – $0.37 \text{ m}^3/\text{s}$ ).

Among all catchments, ratios of NO<sub>3</sub>-N concentrations from one sampling period to another (i.e., the change in measured NO<sub>3</sub>-N at repeated sampling locations) are spatially variable (Table 3), and these variations were not correlated with the distribution of estimated monthly precipitation (see Nitrate Concentrations and Landscape Factors Under Different Inter-Annual Conditions). From March (S1) to October (S4), NO<sub>3</sub>-N concentrations increase, on average, to approximately two times March concentrations. However, stream NO<sub>3</sub>-N concentrations in some catchments are over seven times March concentrations during October sampling. Other catchments decrease to  $\sim 20\%$  of their March concentrations. Concentration ratios suggest a general decrease in NO<sub>3</sub>-N between March and

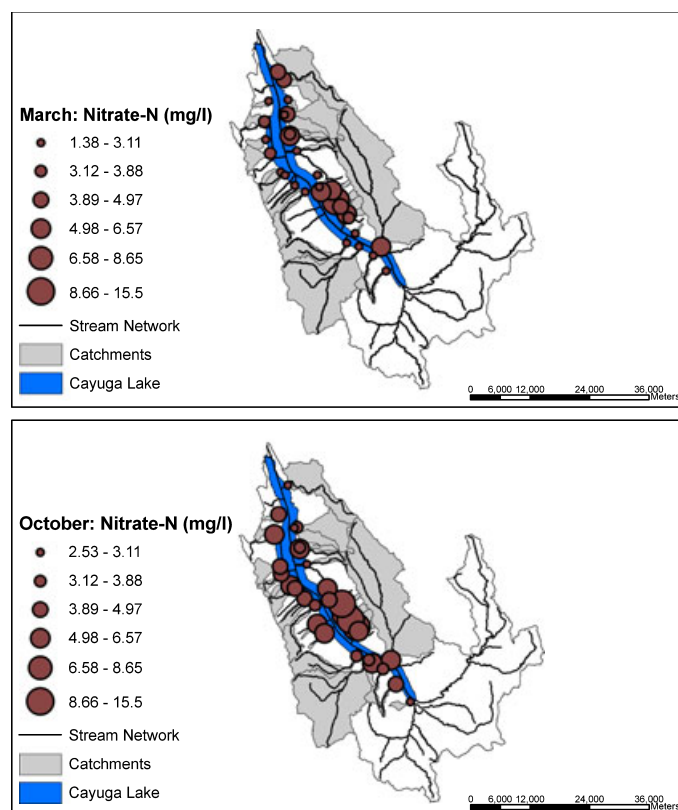


FIGURE 5. Comparison of the Changes in Magnitude and the Spatial Distribution of NO<sub>3</sub>-N Concentrations. The map includes 30 catchments with the highest concentrations during the S1 and S4 sampling events.

August, with ratios ranging from only 0.46% March concentrations to  $\sim 85\%$  (range: 0.010–0.85, mean = 0.23). Changes in concentration ratios between March and September (S3) are more variable. Although concentrations, on average, decrease from S1 to S3 (mean = 0.60), some catchments exhibit over two times measured March NO<sub>3</sub>-N concentrations. Between August (S2) and September (S3), concentrations generally increased in all but one catchment. Nitrate-N concentration in all catchments ( $n = 11$ ) between August (S2) and October (S4) increase (mean ratio = 61); however, ranges vary widely. While NO<sub>3</sub>-N concentrations in some catchments during October are only approximately three times that of August, others increase over two orders of magnitude to greater than 500 times August concentrations. Changes between September (S3) and October (S4) show a similar pattern, with an average overall decrease from S3 to S4 (mean = 36) but with ranges in concentrations from 40% S3 concentrations to over 250 times that of the September sampling event.

Correlograms using Moran's  $I$  statistic further support the spatial variations in NO<sub>3</sub>-N concentrations throughout CLW. Figures 7a and 7b depict the spatial structure for the large (S1 and S4 events); the

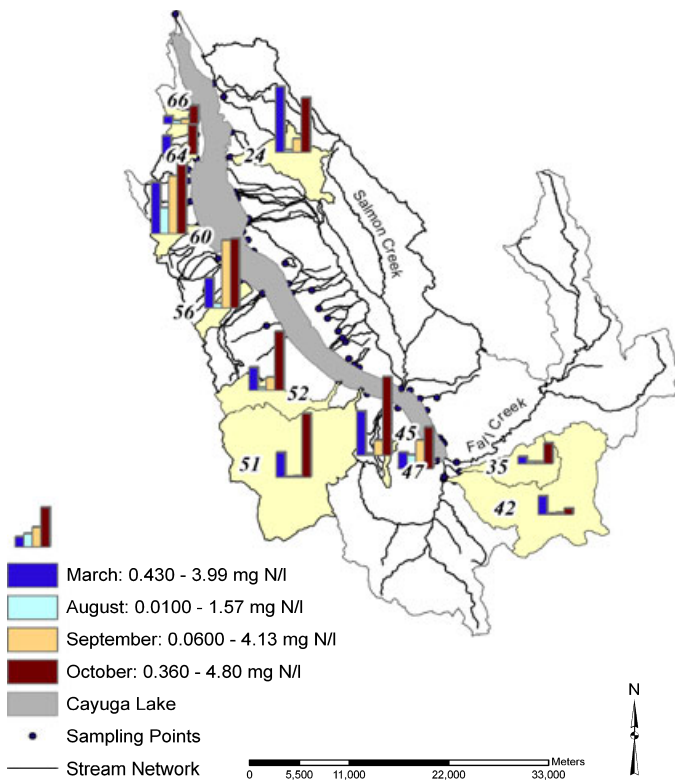


FIGURE 6. Variations in  $\text{NO}_3\text{-N}$  Concentrations Among the 11 Catchments Monitored Across All Four Sampling Events. The scale of the y-axis on all bar charts is zero to 4.80 mg N/l.

TABLE 3. Changes in  $\text{NO}_3\text{-N}$  Concentration Ratios Between Sampling Events.

$\text{NO}_3\text{-N}$ (mg/l)		Ratio of $\text{NO}_3\text{-N}$ Concentration		
From	To	Mean	Min	Max
S1	S4	1.91	0.20	7.15
S1	S2	0.23	0.00	0.85
S1	S3	0.60	0.04	2.27
S2	S3	4.84	0.98	14.7
S2	S4	61.4	2.94	555
S3	S4	35.9	0.39	253

summer sampling events had insufficient sample size to interpret the degree of spatial correlation. The 1 km distance lags for the first 15 km are presented, and two-sided significance tests were performed for each lag to determine if the null hypothesis of no spatial correlation could be rejected. Sample sizes within each lag can vary according to how many sample pairs are separated by each lag distance range (Table 4). Sample sizes for these lags ranged from 28 to 92, and the maximum distance between two points in the dataset is 54 km. For the S1 event, five lags within the first 5 km showed significant positive spatial correlation ( $I = 1.1, 0.51, 0.36, 0.33$ , and  $0.33$ ) and for the S4 event, four of the first six distance lags

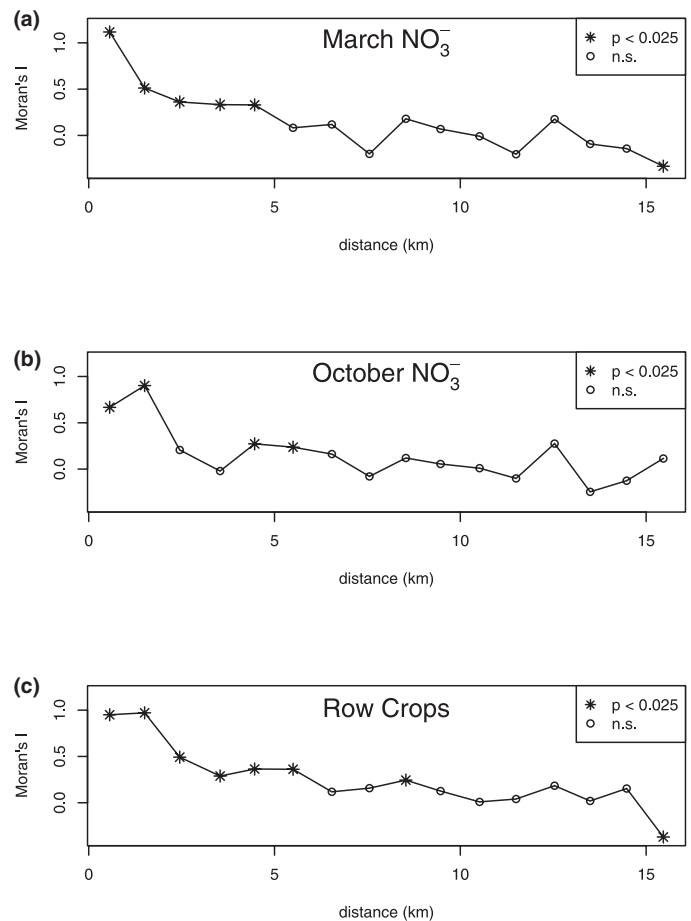


FIGURE 7. Moran's  $I$  Correlograms of Stream  $\text{NO}_3\text{-N}$  Concentrations From March (S1, panel a), October (S4, panel b) Sampling Events, and Percentage Row Crops (panel c); n.s. = Not Significant. The plots show significant positive spatial autocorrelation at lag distances up to 5-6 km for the observed  $\text{NO}_3\text{-N}$  concentrations and for the significant landscape variable - percentage row crops - from the partial correlation analysis. These plots also exhibit a similar spatial structure in terms of the decline in the degree of spatial correlation as a function of lag distance; a general decline in the amount of positive correlation is observed in the first 5-6 km, and beyond this distance significant spatial correlation trends are not observed.

showed significant positive spatial correlation ( $I = 0.67, 0.90, 0.21, -0.020, 0.27$ , and  $0.24$ ). However, two key findings are evident: (1) there is a distinct decrease in the magnitude of correlation with increasing distance and (2) beyond 5-6 km,  $\text{NO}_3\text{-N}$  concentrations throughout CLW show no detectable spatial structure, with some isolated correlations beyond 15 km, which appear to be spurious.

#### Nitrate Concentrations and Landscape Factors Under Different Inter-Annual Conditions

Initial Pearson correlations among landscape factors within the whole catchments and stream  $\text{NO}_3\text{-N}$

TABLE 4. Moran's *I* Coefficients for Lags up to 15 km for March, October, and Row Crops.

Lag Distance (km)	<i>n</i>	March Moran's <i>I</i>	October Moran's <i>I</i>	Row Crop Moran's <i>I</i>
1	28	<b>1.12</b>	<b>0.67</b>	<b>0.95</b>
2	29	<b>0.51</b>	<b>0.90</b>	<b>0.97</b>
3	69	<b>0.36</b>	0.21	<b>0.49</b>
4	62	<b>0.33</b>	-0.02	<b>0.29</b>
5	73	<b>0.33</b>	<b>0.27</b>	<b>0.36</b>
6	92	0.08	<b>0.24</b>	<b>0.36</b>
7	74	0.12	0.16	0.12
8	81	-0.20	-0.08	0.16
9	57	0.18	0.12	0.24
10	62	0.07	0.06	0.13
11	69	-0.01	0.01	0.01
12	58	-0.20	-0.10	0.04
13	56	0.18	0.28	0.18
14	52	-0.09	-0.25	0.02
15	49	-0.14	-0.13	0.15

Note: Bolded values are statistically significant ( $\alpha = 0.025$ ).

concentrations during each sample event revealed many significant relationships, particularly during S1 and S4. Eleven variables were significantly correlated with concentrations in March (all  $p < 0.05$ ), including mean TI ( $r = 0.31$ ), percentage HSG B ( $r = 0.69$ ), percentage HSG C ( $r = -0.69$ ), average slope ( $r = -0.36$ ), percentage water ( $r = -0.33$ ), percentage grasses ( $r = -0.41$ ), percentage developed land ( $r = -0.49$ ), percentage forested land ( $r = -0.44$ ), percentage scrub-shrub land ( $r = -0.35$ ), percentage row crops

( $r = 0.72$ ), and percentage wetlands ( $r = -0.37$ ). Many of the same relationships between landscape factors and NO<sub>3</sub>-N remained during the October sampling, including Mean TI ( $r = 0.010$ ), percentage HSG B ( $r = 0.72$ ), percentage HSG C ( $r = 0.59$ ), percentage water ( $r = -0.52$ ), percentage grasses ( $r = -0.45$ ), percentage developed land ( $r = -0.44$ ), percentage shrub-scrub land ( $r = -0.34$ ), and percentage row crops ( $r = 0.65$ ), all  $p < 0.05$ . However, in S4, percentage HSG D becomes significant with a weak correlation ( $r = -0.04$ ), and percentage forested land and percentage wetlands become nonsignificant.

Partial correlations suggest different results, however, once independent analyses of landscape variables with NO<sub>3</sub>-N concentrations were assessed. For S1, only percentage row crops and percentage developed land remained significant (Table 5). For S4, percentage row crops, percentage water, and percentage HSG B remained significant (Table 5). Figure 7c presents a Moran's *I* correlogram for percentage row crops, the landscape variable significant in the partial correlation analyses of both S1 and S4. It shows a similar degree and range of spatial correlation as the NO<sub>3</sub>-N concentrations in the S1 and S4 events; the first six distance lags show a significant degree of positive spatial correlation (Table 4) ( $I = 0.95, 0.97, 0.49, 0.29, 0.36$ , and  $0.36$ ). Again, a general decline in the amount of positive correlation is observed in the first 5-6 km, and beyond this distance significant spatial correlation trends are not observed.

During the August and September sampling events, fewer correlations between landscape factors

TABLE 5. Partial Correlations of Stream NO<sub>3</sub>-N Concentrations and Landscape Factors Within Whole Catchments and HSAs.

	Whole Catchments: log NO <sub>3</sub> <sup>-</sup> (mg N/l)			
	Partial Correlation, <i>r</i>			
	S1: March <i>n</i> = 64	S2: August <i>n</i> = 11	S3: September <i>n</i> = 16	S4: October <i>n</i> = 55
Catchment area (km <sup>2</sup> )	—	-0.816	—	—
HSG B (%)	—	—	—	0.400
Land cover: water (%)	—	—	—	-0.429
Land cover: developed (%)	-0.279	—	—	—
Land cover: row crops (%)	0.439	—	—	0.496
	HSAs: log NO <sub>3</sub> <sup>-</sup> (mg N/l)			
	Partial Correlation, <i>r</i>			
	S1: March <i>n</i> = 64	S2: August <i>n</i> = 11	S3: September <i>n</i> = 16	S4: October <i>n</i> = 55
HSG B (%)	—	—	—	0.346
Land cover: water (%)	—	—	—	-0.398
Land cover: row crops (%)	0.274	—	—	0.521

Notes: HSA, hydrologically sensitive area; HSG, hydrologic soil group.

All correlations significant at  $p < 0.05$ , unless otherwise stated; — means no significant correlation.

and  $\text{NO}_3\text{-N}$  concentrations were significant within the whole catchments. For S2, only catchment area was significant, and for S3, catchment area, average slope, percentage forested land, and percentage row crops were significant (Table 5). Similar to S1 and S4, partial correlations revealed different results, eliminating all landscape factor- $\text{NO}_3\text{-N}$  concentration correlations in S3, and maintaining catchment area as the single significant factor in August, which is partially attributed to the lack of significantly interfering variables for partial correlation analysis.

Within the HSAs and across all events, Pearson's correlations were relatively similar – but exhibited slightly lower strength – with that of whole catchments. Differences in variables included the addition of percentage forested land and elimination of percentage scrub-shrub land during S1, the addition of percentage wetlands and deletion of average slope for S3, and addition of percentage forested land, percentage wetlands, and monthly precipitation during S4. However, as with whole catchment correlations, partial correlations factored out the influence of other significant factors. Percentage row crops in HSAs remained significant, though markedly lower than Pearson's  $r$ , for S1 (partial  $r = 0.28$ ,  $p < 0.05$ ) and became slightly higher for S4 (partial  $r = 0.52$ ,  $p < 0.05$ ) (Table 5). Only percentage HSG B and percentage water remained significant for S4. No partial correlations between landscape factors and N concentrations were evident during S2 and S3 within the HSAs.

## DISCUSSION

### *Nitrate Variations Among Sampling Events*

Nitrate concentrations were distinctly variable across each sampling event. October exhibited the highest average concentrations (3.6 mg  $\text{NO}_3\text{-N/l}$ ). Ranges in October concentrations were comparable to other studies on the lower end of values (from 0.010 to 0.040 mg  $\text{NO}_3\text{-N/l}$ ), and less than the median value (0.085 mg  $\text{NO}_3\text{-N/l}$ ) of 85 streams draining relatively undeveloped areas of the U.S. (Clark *et al.*, 2000). Maximum concentration values were high, for example, compared with Poor and McDonnell (2007), who report the highest  $\text{NO}_3^-$  concentrations (as N) in an agricultural catchment at  $\sim 1.2$  mg N/l during a storm event and Petry *et al.* (2002), who revealed  $\text{NO}_3^-$  concentrations up to 12.7 mg N/l from samples taken at regular (two weeks) intervals in agricultural catchments. Our catchment with the highest

$\text{NO}_3\text{-N}$  concentration was sampled during the October event (at 15.5 mg N/l, higher than MCL guidance levels) in a slightly smaller row crop dominated catchment compared with the other studies. These higher than average results are potentially related to: (1) elevated antecedent moisture throughout CLW relative to the other sampling periods (Figures 3 and 4), (2) consequent greater connectivity between landscape sources of  $\text{NO}_3\text{-N}$  and surface waters compared with other periods, and (3) an accumulation of  $\text{NO}_3\text{-N}$  in surface soils during dry periods from fertilizers applied during the growing season (Bechtold *et al.*, 2003). This is consistent with findings from other studies (Creed and Band, 1998b; Bechtold *et al.*, 2003), where elevated amounts of  $\text{NO}_3\text{-N}$  from surface soils are released to surface waters immediately before or during the peak in the rising limb of the hydrograph, after which concentrations can become diluted.

Concentrations in March were the second highest (mean = 2.1  $\text{NO}_3\text{-N/l}$ ); however, antecedent hydrological conditions – indicated by precipitation rates – were lower than that of September (Figure 4), suggesting that several important biogeochemical processes and sources of N in the landscape regulate  $\text{NO}_3\text{-N}$  concentrations during this period. For example, the dormant season (March) often reflects a period when soil  $\text{NO}_3\text{-N}$  availability is high in temperate climates resulting, in part, from decreased vegetative uptake of N (Arheimer *et al.*, 1996) and increased soil nitrification following a soil freezing period (Mitchell *et al.*, 1996). Further, transport from preferential elution of  $\text{NO}_3\text{-N}$  from the top of the snowpack during the initial stages of spring melt (Stoddard, 1995) and increased landscape connections with the stream during snowmelt events (Peters *et al.*, 2006) potentially provide transport mechanisms for the accumulated soil  $\text{NO}_3\text{-N}$  to surface waters during this season. Over 30 years of data in Fall Creek support this seasonal trend (Elizabeth W. Boyer, 2007, personal communication; Alexander *et al.*, 2007a), illustrated via an inverse relationship between temperature and  $\text{NO}_3\text{-N}$  concentrations in the stream (Figure 2).

Although extreme low flow conditions ( $\sim 0.070$ – $0.090$  mm/d or  $0.27$ – $0.37$   $\text{m}^3/\text{s}$ ) were occurring during the August sampling event, sources of  $\text{NO}_3\text{-N}$  were present and releasing from ground water supplies from landscape or bedrock sources. Further, the increase in stream  $\text{NO}_3\text{-N}$  concentrations between the August and September sampling events suggests  $\text{NO}_3\text{-N}$  exists in a flush-limited, rather than source-limited, state in the upper soil horizons during this period. This pattern is often observed following long, dry periods when accumulated  $\text{NO}_3\text{-N}$  is greater than that which is utilized by vegetation or denitrified



(Bechtold *et al.*, 2003) and is readily flushed from the soil when wetness and connectivity increase within the catchment (i.e., in response to the hurricane-based rain event the day before S3 sampling).

### *Spatial Variations of NO<sub>3</sub>-N Concentrations*

Changes in concentration ratios (Table 3) among the sampling periods and Moran's *I* correlograms (Table 4) clearly suggest that the response of stream NO<sub>3</sub>-N concentrations to temporally variable biogeochemical and hydrological conditions is nonuniform across all catchments resulting in high spatial variability, similar to other studies (e.g., Peters and Donohue, 2001; Petry *et al.*, 2002). The range of differences in catchment response between the drier periods during the growing season (S2 and S3) and S4, which took place during a wet period near the end of the growing season, was the most extreme. Stream NO<sub>3</sub>-N measured in August compared with October, for example, increased from three to over 500 times August concentrations, suggesting a wide variety of both dynamic (i.e., fate and transport processes) and relatively static (e.g., landscape cover and geomorphic setting) factors uniquely affecting each catchment (see subsequent section) with limited spatial structure to this response.

In general, we found that four of the smallest catchments exhibited the highest NO<sub>3</sub>-N concentrations and responded most rapidly to changes in hydrological conditions, similar to that found by Peters *et al.* (2006) in multiple catchments throughout the U.S. As a result, the distribution of the smaller catchments could partially regulate the spatial variability of NO<sub>3</sub>-N concentration across CLW. Moreover, the riparian areas in many of the smaller catchments in CLW are extremely small or have agricultural practices (i.e., row crops or pasture) extending to the stream edges. As a result, because riparian areas can exhibit higher dissolved inorganic N (primarily NO<sub>3</sub><sup>-</sup>) concentrations than adjacent stream water (Lewis *et al.*, 2007), less NO<sub>3</sub>-N could be retained at the stream edges, possibly resulting in higher NO<sub>3</sub>-N concentrations in streams draining small, agricultural catchments.

### *Variations in Links Between NO<sub>3</sub>-N and Landscape Factors*

Although several relationships between landscape factors and NO<sub>3</sub>-N concentrations were revealed with Pearson correlations, once the influence of competing factors was eliminated, only percentage of row crops shared a significant relationship among the two

largest sampling events, S1 and S4, within the whole catchments and within the HSAs (Table 5). This is supported by the Moran's *I* correlograms for the S1 and S4 NO<sub>3</sub>-N concentration sampling events (Figure 7), which demonstrate similar patterns of spatial correlation as the percentage row crop correlogram: a significant, but declining, degree of spatial correlation for the first 5-6 km with no significant correlation trends beyond this range. These patterns suggest a short-range response between the row crops percentage landscape variable and NO<sub>3</sub>-N concentrations similar to the partial correlation analysis. This fits our conceptual model of landscape-stream NO<sub>3</sub>-N relationships within CLW, based upon multiple studies linking row crops and stream NO<sub>3</sub>-N (Jordan *et al.*, 1997; Arheimer and Lidén, 2000; McIsaac *et al.*, 2001; Benson *et al.*, 2006; Schilling and Spooner, 2006; Schoonover and Lockaby, 2006; Johnson *et al.*, 2007; Poor and McDonnell, 2007; Kang *et al.*, 2008). This is further supported by Johnson *et al.* (2007) who report that corn production for grain (via row crops) was most highly correlated to stream NO<sub>3</sub><sup>-</sup> values in Fall Creek ( $r = 0.53$ ,  $p = 0.01$ ), which is the largest subcatchment in CLW. However, row crops were not a significant factor, as suggested by partial correlations, during the drier sampling periods (S2 and S3). The decomposition of the row crop NO<sub>3</sub>-N relationship, the negative relationship between area and NO<sub>3</sub>-N concentrations during S2, and the lack of relationships between other landscape factors and NO<sub>3</sub>-N during the summer months could be attributed to multiple factors, including low flow conditions, lack of hydrologic connectivity between landscape sources of NO<sub>3</sub>-N and the stream, or to small sample sizes ( $n < 17$ ) during these sampling events.

Other than percentage of row crops, the influence of landscape factors varied across sampling periods, and among catchments, to the extent that no correlations were present during S3. For example, in August, during an extremely dry period, only catchment area exhibited a significant (and strong) negative relationship with NO<sub>3</sub>-N. This is consistent with smaller catchments typically exhibiting "flashier" hydrological and flushing behavior compared with larger catchments in response to rain events (Peters *et al.*, 2006), and with studies suggesting that upstream land cover is more influential on NO<sub>3</sub>-N concentrations in lower order streams (i.e., with a larger contributing area) than smaller, higher order streams (Buck *et al.*, 2004). However, we observed no relationships between landscape factors and NO<sub>3</sub>-N in the HSAs during the two dry sampling periods after controlling for other significant relationships between explanatory variables, suggesting additional explanations for this relationship. In addition to low

flow conditions and lack of hydrologic connectivity between the landscape and stream during the summer, sample sizes during both summer events were substantially lower than the March and October sampling periods, which could also partially contribute to the lack of landscape variable-stream  $\text{NO}_3\text{-N}$  concentrations relationship.

Partial correlation coefficients between  $\text{NO}_3\text{-N}$  and HSG B are also significant during October in both the whole catchment and HSAs (Table 5), potentially suggesting higher connectivity between these typically moderately well-drained to well-drained soils and the tributaries during this wet period. For example, modeled N export coefficients suggest that under “normal” wetness conditions, a lower ratio of atmospherically deposited N is typically delivered to surface water systems in cultivated catchments under HSG B soils compared with those with HSG C or HSG D soils (e.g., see Sheeder *et al.*, 2002). However, this ratio increases under wetter soil conditions, potentially elevating  $\text{NO}_3\text{-N}$  transport from HSG B soils.

Although we used publically available spatial datasets as indicators of the transport and fate of  $\text{NO}_3\text{-N}$  throughout the catchments (e.g., HSGs, estimated monthly precipitation), similar to other studies (Sheeder *et al.*, 2002), only direct measurements would elucidate the actual extent of influence of soil moisture properties on denitrification and delivery of  $\text{NO}_3\text{-N}$  to surface waters. The application of indicator factors, in addition to the use of snapshots of stream data, introduces multiple layers of complexity into estimates of relationships between landscape factors associated with catchment fate and transport of  $\text{NO}_3\text{-N}$  to streams. Further, potential landscape linkages between actual soil wetness and stream  $\text{NO}_3\text{-N}$  concentrations as a result of denitrification, for example, are also highly spatially and temporally variable (Arheimer and Lidén, 2000; Buck *et al.*, 2004). Regardless of the potential limitations of this approach, it is clear that across the four snapshots of stream chemistry within CLW, landscape relationships with  $\text{NO}_3\text{-N}$  vary spatially and among synoptic sampling events, with the exception of row crops during the larger sampling periods.

#### *Implications for Future Watershed Research and Management*

While we know that unique catchment characteristics result in complex interactions and highly individual changes in stream  $\text{NO}_3\text{-N}$  concentrations, assessing this spatial heterogeneity in catchment response across multiple catchments within a single basin and under a variety of seasonal conditions is

important on several fronts. First, spatial variations in stream  $\text{NO}_3\text{-N}$  concentrations and links to landscape characteristics are often evaluated as indicators of regional aquatic ecosystem health and ecosystem services. Understanding variations in catchment response to annual changes in hydrological and biogeochemical conditions is imperative to draw these broader conclusions. Further, most of our knowledge concerning variable hydrological and biogeochemical catchment responses to seasonal changes derives from small-catchment studies in relatively undisturbed forested settings (e.g., Burns *et al.*, 1998a; McHale *et al.*, 2000; Band *et al.*, 2001; Mitchell *et al.*, 2006; Christopher *et al.*, 2008). While these studies are imperative for understanding processes regulating sources, fate, and hydrological transport of solutes through a catchment, details gained from this research are also unique to the catchment from which they originate, further advancing the call for a unifying framework for hydrological (McDonnell *et al.*, 2007) and biogeochemical catchment-based science and supporting the need for conceptualizing a set of principles underpinning this complexity (McDonnell *et al.*, 2007).

A unifying approach to research focusing on multiple catchments is particularly useful for allocating resources more efficiently to assess biogeochemical and hydrological regulators of  $\text{NO}_3\text{-N}$  at the catchment scale to clarify decisions on modeling and monitoring approaches (Mitchell, 2001). This unifying framework could also extend to the watershed management community, decreasing the complexity of each individual management case.

## CONCLUSIONS

Nitrate-N concentrations are highly variable across >55 unique mixed land cover catchments of the CLW, ranging from 0.040 to 8.68 mg  $\text{NO}_3\text{-N/l}$  and 0.090 to 15.5 mg  $\text{NO}_3\text{-N/l}$  during the March and October sampling events, respectively. Summer low flows also exhibited such variability, though the sample size ( $n = 11$ ) and concentration values are considerably lower. Spatial variations across the catchments of CLW exhibit extensive heterogeneity of stream  $\text{NO}_3\text{-N}$  response to diverse seasonal biogeochemical and hydrological settings and unique catchment characteristics, as indicated by changes in  $\text{NO}_3\text{-N}$  concentration ratios and spatial mapping. However,  $\text{NO}_3\text{-N}$  concentrations are generally higher during wetter periods and when available  $\text{NO}_3\text{-N}$  in the soil is elevated (i.e., the dormant season or after extended dry periods with potentially elevated  $\text{NO}_3\text{-N}$  accumulation

in the upper soil horizons). This is consistent with multiple other small-catchment studies.

When competing landscape variables are factored out, row crops exhibit the strongest landscape-NO<sub>3</sub>-N relationship, but only during the larger sampling events, which take place during wetter periods of the year. Further, only one landscape factor – catchment area – is significant during the August summer sampling event after other competing factors are eliminated. However, during the September sampling period, minimal landscape-nitrate relationships exist within whole catchments and within HSAs. Additional research would clarify these low flow-NO<sub>3</sub>-N source relationships.

It is well known that individual catchments respond uniquely to changes in hydrological and biogeochemical conditions. However, research assessing spatial variations of NO<sub>3</sub>-N in surface waters throughout multiple catchments and estimating relationships of NO<sub>3</sub>-N to landscape attributes under seasonal settings that affect both biogeochemical processes (e.g., biological uptake of N) and hydrological conditions is particularly important. For example, stream NO<sub>3</sub>-N concentrations – and often land cover factors affecting NO<sub>3</sub>-N – continue to be employed as indicators of regional aquatic ecosystem health and ecosystem services. Further, the unique response of NO<sub>3</sub>-N across multiple catchments in CLW advances the call for a unifying framework for hydrological and biogeochemical catchment-based science and management.

## APPENDIX

TABLE A1. Nitrate-N (mg/l) Concentrations for Each Catchment During the Sampling Events.  
UNT = unnamed tributary.

Catchment	Catchment Name	Mar-05	Aug-05	Sep-05	Oct-05
1	Salmon Creek	5.83			6.57
2	UNT	0.48			0.34
3	UNT	1.84			1.64
4	UTN	1.70			
5	Morrow Creek	6.71			
6	UNT	3.81			5.62
7	UNT	5.62			15.44
8	UNT	4.53			14.92
9	UNT	8.68			15.53
10	UNT	6.37			9.93
11	UNT	6.18			1.23
12	UNT	1.84			4.96

TABLE A1 (Continued)

Catchment	Catchment Name	Mar-05	Aug-05	Sep-05	Oct-05
13	UNT	1.62			5.43
14	UNT	0.16			0.09
15	UNT	0.04			
16	UNT	1.07			
17	Fames Creek	4.18			
18	Little Creek	2.13			2.99
19	UNT	3.23			
20	UNT	0.60			0.65
21	Dean Creek	6.23			5.05
22	Glen Creek	3.77			3.73
23	UNT	2.61			3.08
24	Great Gully	3.99	0.14	0.82	3.31
25	UNT	2.50			2.39
26	UNT	4.77			1.98
27	Yawger Creek	4.20			3.11
28	Seneca River inlet	0.94	0.03		1.21
29	Minnegar Creek	0.68		0.23	1.89
30	Gulf Creek	0.49			0.87
31	Headwaters of Gulf Creek	0.40			1.18
32	UNT	0.99			
33	UNT	0.21			0.49
34	UNT	0.10			0.18
35	Cascadilla Creek	0.43	0.12	0.12	1.26
36	UNT	0.76		0.11	0.75
37	UNT	0.10			
38	UNT	0.16			
39	UNT				0.54
40	Inlet across from Six Mile Creek in Ithaca	0.44			0.38
42	Six Mile Creek	1.12	0.06	0.11	0.36
43	UNT	0.35			
44	UNT	0.12			
45	Glenwood Creek	2.69	0.11	0.93	4.80
46	Indian Creek	0.50			0.66
47	Williams Brook	0.97	0.83	1.74	2.53
48	Willow Creek	2.36		1.23	3.69
49	UNT	1.41			
50	UNT	0.78		0.13	5.54
51	Taughannock Creek	1.49	0.01	0.06	3.88
52	Trumansburg Creek	1.38	0.15	0.78	3.59
53	Bergen Creek	1.18		0.89	5.34
54	Lively Run	0.98			5.53
55	Sheldrake Creek	1.47			3.43
56	Groves Creek	1.82	0.28	4.13	4.23
57	Mack Creek	1.78			4.23
58	Big Hollow Creek	1.36			4.65
59	UNT	1.81			5.05
60	Hicks Gully	3.12	1.57	3.52	4.63
61	UNT	2.10			1.75
62	Red Creek	0.97			1.14
63	Schuyler Creek	3.19			5.13
64	UNT	1.13	0.31	0.43	1.78
65	UNT	1.99			4.75
66	Canoga Creek	0.45	0.19	0.28	1.09
67	UNT	0.35			0.31



## ACKNOWLEDGMENTS

We appreciate the helpful suggestions from two external reviewers. We are grateful for financial support for this work through grants from the U.S. Environmental Protection Agency Greater Research Opportunities Program, the New York City Watershed Agricultural Council, and the New York Energy and Research Development Authority. We appreciate the helpful advice on this project and manuscript from Doug Burns and Russ Briggs. We also appreciate field assistance from Eric McNeil, Scott Means, Mike Miller, and Rebecca Sauter. Thanks to members of the Cayuga Lake Intermunicipal Organization's technical advisory committee, on which Boyer served, for many useful discussions about landscape characterization in the CLW. This paper has been reviewed in accordance with the U.S. Environmental Protection Agency's peer and administrative review policies and approved for publication. Approval does not signify that the contents necessarily reflect the views and policies of the agency, nor does the mention of trade names or commercial products constitute endorsement or recommendation for use.

## LITERATURE CITED

- Aber, J.D., C.L. Goodale, S.V. Ollinger, M.-L. Smith, A.H. Magill, M.E. Martin, R.A. Hallett, and J.L. Stoddard, 2003. Is Nitrogen Deposition Altering the Nitrogen Status of Northeastern Forests? *BioScience* 53:375-389.
- Alexander, R.B., E.W. Boyer, R.A. Smith, G.E. Schwarz, and R.B. Moore, 2007a. The Role of Headwater Streams in Downstream Water Quality. *Journal of the American Water Resources Association* 43:41-59.
- Alexander, R.B., R.A. Smith, G.E. Schwarz, E.W. Boyer, J.V. Nolan, and J.W. Brakebill, 2007b. Differences in Sources and Recent Trends in Phosphorus and Nitrogen Delivery to the Gulf of Mexico From the Mississippi River and Atchafalaya River Basins. *Environmental Science and Technology* 42:822-830.
- Arheimer, B., L. Andersson, and A. Lepistö, 1996. Variation of Nitrogen Concentration in Forest Streams – Influences of Flow, Seasonality and Catchment Characteristics. *Journal of Hydrology* 179:281-304.
- Arheimer, B. and R. Lidén, 2000. Nitrogen and Phosphorus Concentrations From Agricultural Catchments – Influence of Spatial and Temporal Variables. *Journal of Hydrology* 227:140-159.
- Band, L.E., C. Tague, P. Groffman, and K. Belt, 2001. Forest Ecosystem Processes at the Watershed Scale: Hydrological and Ecological Controls of Nitrogen Export. *Hydrological Processes* 15:2013-2028.
- Bechtold, J.S., R.T. Edwards, and R.J. Naiman, 2003. Biotic Versus Hydrologic Control Over Seasonal Nitrate Leaching in a Floodplain Forest. *Biogeochemistry* 63:53-72.
- Benson, V., J.A. VanLeeuwen, J. Sanchez, I.R. Dohoo, and G.H. Somers, 2006. Spatial Analysis of Land Use Impact on Ground Water Nitrate Concentrations. *Journal of Environmental Quality* 35:421-432.
- Beven, K. and M. Kirkby, 1979. A Physically-Based, Variable Contributing Area Model of Basin Hydrology. *Hydrological Sciences Bulletin* 24:43-69.
- Böhlke, J.-K., 2003. Sources, Transport, and Reaction of Nitrate. In: *Residence Times and Nitrate Transport in Ground Water Discharging to Streams in the Chesapeake Bay Watershed. Water-Resources Investigations*, B.D. Lindsey, S.W. Phillips, C.A. Donnelly, G.K. Speiran, L.N. Plummer, J.-K. Böhlke, M.J. Focazio, W.C. Burton, and E. Busenberg (Editors). U.S. Geological Survey, New Cumberland, Pennsylvania, pp. 25-39. <http://www.pa.water.usgs.gov/reports/wrir03-4035.pdf>, accessed October 8, 2008.
- Boyer, E.W., R.B. Alexander, W.J. Parton, C. Li, K. Butterbach-Bahl, S.D. Donner, R.W. Skaggs, and S.J. Del Grosso, 2006a. Modeling Denitrification in Terrestrial and Aquatic Ecosystems at Regional Scales. *Ecological Applications* 16:2123-2142.
- Boyer, E., C. Goodale, N. Jaworski, and R. Howarth, 2002. Anthropogenic Sources and Relationships to Riverine Nitrogen Export in the Northeastern USA. *Biogeochemistry* 57/58:137-169.
- Boyer, E.W., R.H. Howarth, J.N. Galloway, F. Dentener, P.A. Green, and C.J. Vörösmarty, 2006b. Riverine Nitrogen Export From the Continents to the Coasts. *Global Biogeochem. Cycles* 20, GB1S91, doi: 10.1029/2005GB002537.
- Bricker, S.B., C.G. Clement, D.E. Pirhalla, S.P. Orland, and D.G. Farrow, 1999. National Estuarine Eutrophication Assessment: A Summary of Conditions, Historical Trends, and Future Outlook. National Ocean Service, National Oceanic and Atmospheric Administration, Silver Springs, Maryland. <http://www.ian.umces.edu/nea/>, accessed October 8, 2008.
- Buck, O., D.K. Niyogi, and C.R. Townsend, 2004. Scale-Dependence of Land Use Effects on Water Quality of Streams in Agricultural Catchments. *Environmental Pollution* 130:287-299.
- Burns, D.A., 1998. Retention of  $\text{NO}_3^-$  in an Upland Stream Environment: A Mass Balance Approach. *Biogeochemistry* 40:73-96.
- Burns, D.A., R.P. Hooper, J.J. McDonnell, J.E. Freer, C. Kendall, and K. Beven, 1998a. Base Cation Concentrations in Subsurface Flow From a Forested Hillslope: The Role of Flushing Frequency. *Water Resources Research* 34:3535-3544.
- Burns, D.A., P.S. Murdoch, G.B. Lawrence, and R.L. Michel, 1998b. Effect of Groundwater Springs on  $\text{NO}_3^-$  Concentrations During Summer in Catskill Mountain Streams. *Water Resources Research* 34:1987-1996.
- Burns, D.A., T. Vitvar, J.J. McDonnell, J. Hassett, J. Duncan, and C. Kendall, 2005. Effects of Suburban Development on Runoff Generation in the Croton River Basin, New York, USA. *Journal of Hydrology* 311:266-281.
- Campbell, J.L., J.W. Hornbeck, M.J. Mitchell, M.B. Adams, M.S. Castro, C.T. Driscoll, J.S. Kahl, J.N. Kochenderfer, G.E. Likens, J.A. Lynch, P.S. Murdoch, S.J. Nelson, and J.B. Shanley, 2004. Input-Output Budgets of Inorganic Nitrogen for 24 Forest Watersheds in the Northeastern United States: A Review. *Water, Air, and Soil Pollution* 151:373-396.
- Caraco, N.F., and J.J. Cole, 1999. Human Impact on Nitrate Export: An Analysis Using Major World Rivers. *Ambio* 28:167-170.
- Carpenter, S., N.F. Caraco, D. Correll, R.H. Howarth, A.N. Sharp-ley, and V.H. Smith, 1998. Issues in Ecology, Technical Report: Nonpoint Pollution of Surface Waters With Phosphorus and Nitrogen. *Ecological Applications* 8:559-568.
- Christopher, S.F., M.J. Mitchell, M.R. McHale, E.W. Boyer, D.A. Burns, and C. Kendall, 2008. Factors Controlling Nitrogen Release From Two Forested Catchments With Contrasting Hydrochemical Responses. *Hydrological Processes* 22:46-62.
- Clark, G.M., D.K. Mueller, and M.A. Mast, 2000. Nutrient Concentrations and Yields in Undeveloped Stream Basins of the United States. *Journal of the American Water Resources Association* 36:849-860.
- Creed, I.F., and L.E. Band, 1998a. Exploring Functional Similarity in the Export of Nitrate-N From Forested Catchments: A Mechanistic Modeling Approach. *Water Resources Research* 34:3079-3093.
- Creed, I.F. and L.E. Band, 1998b. Export of Nitrogen From Catchments Within a Temperate Forest: Evidence for a Unifying Mechanism Regulated by Variable Source Area Dynamics. *Water Resources Research* 34:3105-3120.
- Cuevas, J.G., D. Soto, I. Arismendi, M. Pino, A. Lara, and C. Oyarzún, 2006. Relating Land Cover to Stream Properties in



- Southern Chilean Watersheds: Trade-off Between Geographic Scale, Sample Size, and Explicative Power. *Biogeochemistry* 81:313-329.
- Daly, C., W.P. Gibson, G.H. Taylor, G.L. Johnson, and P. Pasteris, 2002. A Knowledge-Based Approach to the Statistical Mapping of Climate. *Climate Research* 22:99-113.
- Driscoll, C.T., G.B. Lawrence, A.J. Bulger, T.J. Butler, C.S. Cronan, C. Eagar, K.F. Lambert, G.E. Likens, J.L. Stoddard, and K.C. Weathers, 2001. Acidic Deposition in the Northeastern United States: Sources and Inputs, Ecosystems Effects, and Management Strategies. *Biogeochemistry* 51:180-198.
- Driscoll, C.T., D. Whitall, A. JD, E.W. Boyer, M. Castro, C. Cronan, C.L. Goodale, P. Groffman, C. Hopkinson, K. Lambert, G.B. Lawrence, and S.V. Ollinger, 2003. Nitrogen Pollution in the Northeastern United States: Sources, Effects, and Management Options. *Biogeochemistry* 53:357-374.
- Franklin, D.H., J.L. Steiner, M.L. Cabrera, and E.L. Usery, 2002. Distribution of Inorganic Nitrogen and Phosphorus Concentrations in Stream Flow of Two Southern Piedmont Watersheds. *Journal of Environmental Quality* 31:1910-1917.
- GFLRPC (Genesee Finger Lakes Regional Planning Council), 2004. Cayuga Lake Watershed Restoration and Protection Plan. Cayuga Lake Watershed Intermunicipal Organization, Lansing, New York. <http://www.cayugawatershed.org/Cayuga%20Lake/RPP/cayindex3.htm>, accessed October 8, 2008.
- Golden, H.E. and E.W. Boyer, 2008. Contemporary Estimates of Atmospheric Nitrogen Deposition to the Watersheds of New York State. *Environmental Monitoring and Assessment*, doi: 10.1007/s10661-10008-10438-10668.
- Grayson, R., C.J. Gippel, B.L. Findlayson, and B.T. Hart, 1997. Catchment-Wide Impacts on Water Quality: The Use of 'Snapshot' Sampling During Stable Flow. *Journal of Hydrology* 199:121-134.
- Green, P., C. Vörösmarty, M. Meybeck, J.N. Galloway, B. Peterson, and E.W. Boyer, 2004. Pre-Industrial and Contemporary Fluxes of Nitrogen Through Rivers: A Global Assessment Based on Typology. *Biogeochemistry* 68:71-105.
- Herlihy, A.T., J.L. Stoddard, and C.B. Johnson, 1998. The Relationship Between Stream Chemistry and Watershed Land Cover Data in the Mid-Atlantic Region, U.S. *Water, Air, and Soil Pollution* 105:377-386.
- Holloway, J.M. and R.A. Dahlgren, 1999. Geologic Nitrogen in Terrestrial Biogeochemical Cycling. *Geology* 27:567-570.
- Holloway, J.M., R.A. Dahlgren, B. Hansen, and W.H. Casey, 1998. Contribution of Bedrock Nitrogen to High Nitrate Concentrations in Stream Water. *Nature* 396:785-787.
- Ito, M., M.J. Mitchell, C.T. Driscoll, and K.M. Roy, 2005. Nitrogen Input-Output Budgets for Lake-Containing Watersheds in the Adirondack Region of New York. *Biogeochemistry* 72:283-314.
- Jaworski, N.A., R.H. Howarth, and L. Hetling, 1997. Atmospheric Deposition of Nitrogen Oxides Onto the Landscape Contributes to Coastal Eutrophication in the Northeast United States. *Environmental Science and Technology* 31:1995-2004.
- Johnson, L.B., C. Richards, G.E. Host, and J.W. Arthur, 1997. Landscape Influences on Water Chemistry in Midwestern Stream Ecosystems. *Freshwater Biology* 37:193-208.
- Johnson, M.S., P.B. Woodbury, A.N. Pell, and J. Lehmann, 2007. Land-Use Change and Stream Water Fluxes: Decadal Dynamics in Watershed Nitrate Exports. *Ecosystems* 10:1182-1196.
- Jones, K., A.C. Neale, M.S. Nash, R.D. Van Remortel, J.D. Wickham, K.H. Riitters, and R.V. O'Neill, 2001. Predicting Nutrient and Sediment Loadings to Streams From Landscape Metrics: A Multiple Watershed Study From the United States Mid-Atlantic Region. *Landscape Ecology* 16:301-312.
- Jordan, T.E., D. Correll, and D. Weller, 1997. Relating Nutrient Discharges From Watersheds to Land Use and Streamflow Variability. *Water Resources Research* 33:2579-2590.
- Jordan, T.E. and D. Weller, 1996. Human Contributions to Terrestrial Nitrogen Flux: Assessing the Sources and Fates of Anthropogenic Fixed Nitrogen. *Biogeochemistry* 46:655-664.
- Kang, S., H. Lin, W.J. Gburek, and G.J. Folmar, 2008. Baseflow Nitrate in Relation to Stream Order and Agricultural Land Use. *Journal of Environmental Quality* 37:808-816.
- King, R.S., M.E. Baker, D.F. Whigham, D. Weller, T.E. Jordan, P.F. Kazyak, and M. Hurd, 2005. Spatial Considerations for Linking Watershed Land Cover to Ecological Indicators in Streams. *Ecological Applications* 15:137-153.
- Knoepp, J.D. and J.M. Vose, 2007. Regulation of Nitrogen Mineralization and Nitrification in Southern Appalachian Ecosystems: Separating the Relative Importance of Biotic vs. Abiotic Controls. *Pedobiologia* 51:89-97.
- Legendre, P. and L. Legendre, 1998. *Numerical Ecology*, Second Edition. Elsevier, Amsterdam, the Netherlands.
- Lewis, D.B., N.B. Grimm, T.K. Harms, and J.D. Schade, 2007. Subsystems, Flowpaths, and the Spatial Variability of Nitrogen in a Fluvial Ecosystem. *Landscape Ecology* 22:911-924.
- McDonnell, J.J., M. Sivapalan, K. Vaché, S. Dunn, G.E. Grant, R. Haggerty, C. Hinz, R.P. Hooper, J. Kirchner, M.L. Roderick, J. Selker, and M. Weiler, 2007. Moving Beyond Heterogeneity and Process Complexity: A New Vision for Watershed Hydrology. *Water Resources Research* 43, W07301, doi: 10.1029/2006WR005467.
- McHale, M.R., M.J. Mitchell, J.J. McDonnell, and C.P. Cirino, 2000. Nitrogen Solutes in an Adirondack Forested Watershed: Importance of Dissolved Organic Nitrogen. *Biogeochemistry* 48:165-184.
- McIsaac, G.F., M.B. David, G.Z. Gertner, and D.A. Goolsby, 2001. Nitrate Flux in the Mississippi River. *Nature* 414:166-167.
- Mitchell, M.J., 2001. Linkages of Nitrate Losses in Watersheds to Hydrological Processes. *Hydrological Processes* 15:3305-3307.
- Mitchell, M.J., C.T. Driscoll, S. Inamdar, G.G. McGee, M. Mbila, and D. Raynal, 2003. Nitrogen Biogeochemistry in the Adirondack Mountains of New York: Hardwood Ecosystems and Associated Surface Waters. *Environmental Pollution* 123:355-364.
- Mitchell, M.J., C.T. Driscoll, J.S. Kahl, G.E. Likens, P.S. Murdoch, and L.H. Pardo, 1996. Climatic Control of Nitrate Loss From Forested Watersheds in the Northeast United States. *Environmental Science and Technology* 30:2609-2612.
- Mitchell, M.J., K.B. Piatek, S.F. Christopher, B. Mayer, C. Kendall, and P. Mchale, 2006. Solute Sources in Stream Water During Consecutive Fall Storms in a Northern Hardwood Forest Watershed: A Combined Hydrological, Chemical and Isotopic Approach. *Biogeochemistry* 78:217-246.
- MRLC (Multi-Resolution Land Cover – National Land Cover Database), 2001. National Land Cover Dataset 2001 (NLCD 2001). Multi-Resolution Land Characteristics Consortium. US Geological Survey, Earth Resources Observation and Science Center, Sioux Falls, South Dakota. <http://www.mrlc.gov/nlcd.php>, accessed October 8, 2008.
- Mullins, H.T. and E.J. Hinchey, 1989. Erosion and Infill of New York Finger Lakes: Implications for Laurentide Ice Sheet Deglaciation. *Geology* 17:622-625.
- NCDC (National Climate Data Center), 2006. National Atmospheric and Oceanic Administration. National Climate Data Center. Asheville, North Carolina. <http://www.ncdc.noaa.gov/oa/ncdc.html>, accessed October 9, 2008.
- NEWA (Network for Environment and Weather Awareness), 2006. New York State Integrated Pest Management Program Network for Environment and Weather Awareness. Cornell University, Ithaca, New York. <http://www.vivo.cornell.edu/lifesci/individual/vivo/individual5456>, accessed October 9, 2008.
- Paerl, H., R. Dennis, and D.R. Whitall, 2002. Atmospheric Deposition of Nitrogen: Implications for Nutrient Over-Enrichment of Coastal Waters. *Estuaries* 25:677-693.

- Perakis, S.S. and L.O. Hedin, 2002. Nitrogen Loss From Unpolluted South American Forests Mainly via Dissolved Organic Compounds. *Nature* 415:416-419.
- Peters, N.E. and R. Donohue, 2001. Nutrient Transport to the Swan-Canning Estuary, Western Australia. *Hydrological Processes* 15:2555-2577.
- Peters, N.E., J.B. Shanley, B.T. Aulenbach, R.M. Webb, D.H. Campbell, R. Hunt, M.C. Larsen, R.F. Stallard, J. Troester, and J.F. Walker, 2006. Water and Solute Mass Balances of Five Small, Relatively Undisturbed Watersheds in the U.S. *Science of the Total Environment* 358:221-242.
- Petrone, K.C., I. Buffam, and H. Laudon, 2007. Hydrologic and Biotic Control of Nitrogen Export During Snowmelt: A Combined Conservative and Reactive Tracer Approach. *Water Resources Research* 43, W06420, doi: 10.1029/2006WR005286.
- Petry, J., C. Soulsby, I.A. Malcolm, and A.F. Youngson, 2002. Hydrologic Controls on Nutrient Concentrations and Fluxes in Agricultural Catchments. *Science of the Total Environment* 294:95-110.
- Poor, C.J. and J.J. McDonnell, 2007. The Effects of Land Use on Stream Nitrate Dynamics. *Journal of Hydrology* 332:54-68.
- Puckett, L.J. and W.B. Hughes, 2005. Transport and Fate of Nitrate and Pesticides: Hydrogeology and Riparian Zone Processes. *Journal of Environmental Quality* 34:2278-2292.
- Quinn, P.F., K.J. Beven, and R. Lamb, 1995. The  $\ln(a/\tan\beta)$  Index: How to Calculate It and How to Use It Within the TOPMODEL Framework. *Hydrological Processes* 9:161-182.
- Schilling, K.E. and J. Spooner, 2006. Effects of Watershed-Scale Land Use Change on Stream Water Nitrate Concentrations. *Journal of Environmental Quality* 36:2132-2145.
- Schilling, K. and Y.-K. Zhang, 2004. Baseflow Contribution to Nitrate-Nitrogen Export From a Large, Agricultural Watershed USA. *Journal of Hydrology* 295:305-316.
- Schoonover, J.E. and B.G. Lockaby, 2006. Land Cover Impacts on Stream Nutrients and Fecal Coliform in the Lower Piedmont of West Georgia. *Journal of Hydrology* 331:371-382.
- Seitzinger, S., A.F. Kroeze, N. Bouwman, N. Caraco, F. Dentener, and R.V. Styles, 2002. Global Patterns of Dissolved Inorganic and Particulate Nitrogen Inputs to Coastal Systems: Recent Conditions and Future Projections. *Estuaries* 25:640-655.
- Sheeder, S.A., J.A. Lynch, and J. Grimm, 2002. Modeling Atmospheric Nitrogen Deposition and Transport in the Chesapeake Bay Watershed. *Journal of Environmental Quality* 31:1194-1206.
- Sokal, R.R. and F.J. Rohlf, 1995. *Biometry: The Principles and Practice of Statistics in Biological Research*. W.H. Freeman and Company, New York.
- Stoddard, J.L., 1995. Episodic Acidification During Snowmelt of High Elevation Lakes in the Sierra Nevada Mountains of California. *Water, Air, and Soil Pollution* 85:353-358.
- Strayer, D.L., R.E. Beighley, L.C. Thompson, S. Brooks, C. Nilsson, G. Pinay, and R.J. Naiman, 2003. Effects of Land Cover on Stream Ecosystems: Roles of Empirical Models and Scaling Issues. *Ecosystems* 6:407-423.
- USDA-NRCS (US Department of Agriculture – National Resources Conservation Service), 1986. *Urban Hydrology for Small Watersheds*, Technical Release 55. USDA-NRCS, Washington, D.C.
- USDA-NRCS (US Department of Agriculture – National Resources Conservation Service), 2006. *Soils Data Mart*. USDA-NRCS, Auburn, Alabama. <http://soildatamart.nrcs.usda.gov/>, accessed October 8, 2008.
- USEPA (US Environmental Protection Agency), 1994. *Water Quality Standards Handbook (Second Edition)*. US Environmental Protection Agency, Office of Water, Washington, D.C. <http://www.epa.gov/waterscience/library/wqstandards/handbook.pdf>, accessed October 9, 2008.
- USEPA (US Environmental Protection Agency), 2006. *Water Discharge Permits – PCS*. US Environmental Protection Agency, Washington, D.C.. [http://www.epa.gov/enviro/html/pcs/pcs\\_query\\_java.html](http://www.epa.gov/enviro/html/pcs/pcs_query_java.html), accessed October 8, 2008.
- Van Sickle, J., 2003. Analyzing Correlations Between Stream and Watershed Attributes. *Journal of the American Water Resources Association* 39:717-726.
- Wayland, K.G., D.T. Long, D.W. Hyndman, B.C. Pijanowski, S.M. Woodhams, and S.K. Haack, 2003. Identifying Relationships Between Baseflow Geochemistry and Land Use With Synoptic Sampling and R-Mode Factor Analysis. *Journal of Environmental Quality* 32:180-190.
- Welsch, D., C. Kroll, J.J. McDonnell, and D.A. Burns, 2001. Topographic Controls on the Chemistry of Subsurface Stormflow. *Hydrological Processes* 15:1925-1938.