



# RNA-seq analysis Analysis in R

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## Overview of the day

- RNA-seq quantification from raw data to an expression table
- **♥** (RNA-seq analysis in R from an expression table to pathway analysis)
- ▼ Visual gene expression analysis in Phantasus

- Materials and slides are available at Google Drive
- ✓ Dockerfile and the scripts are available at <a href="https://github.com/ctlab/sysbio-training/tree/master/tomsk-scs-2021">https://github.com/ctlab/sysbio-training/tree/master/tomsk-scs-2021</a>



#### **Prepare**

- **♥** Go to <a href="https://ctlab.itmo.ru/rstudio-sbNN/">https://ctlab.itmo.ru/rstudio-sbNN/</a>
- ✓ login: student
- password: sysbiopass
- Open the project from the previous module
- Open do\_deseq2.R



# **Export expression values**

- ▼ Run steps 0 & 1
- Export:
  - counts.txt
  - es.gct



#### **ExpressionSet**

- A single place to store both expression data and metadata
- exprs(es) expression matrix
- pData(es) or phenoData(es) sample metadata
- fData(es) or featureData(es) gene metadata
- experimentData(es) experiment metadata



#### Org.db packages

- Org.Mm.eg.db, Org.Hs.eg.db, ...
- Contain gene annotation data for an organism
- Functions:
  - mapIds()
  - columns()
  - keys()
  - select()



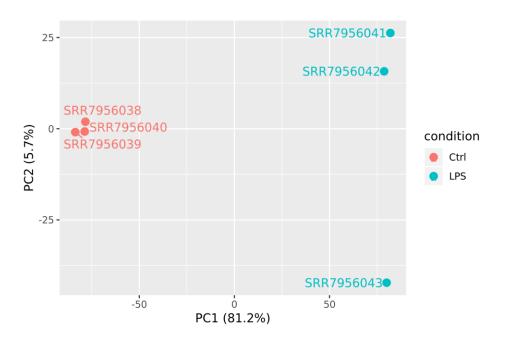
#### **Normalization**

- Run step 2
- Possible normalization for RNA-seq:
  - log2 + quantile
  - divide by median expression
  - DESeq2::getVarianceStabilizedData
  - DESeq2::rlog
  - log2 + limma::voom
- The saved gct file can be opened in Phantasus



# **PCA plot**

Run step 3





## Differential expression for RNA-seq

- DESeq2
- EdgeR
- kallisto/sleuth
- **...**
- Run step 4

#### Method Highly accessed Open Access Comprehensive evaluation of differential gene expression analysis methods for RNA-seg data Franck Rapaport<sup>1</sup>, Raya Khanin<sup>1</sup>, Yupu Liang<sup>1</sup>, Mono Pirun<sup>1</sup>, Azra Krek<sup>1</sup>, Paul Zumbo<sup>23</sup>, Christopher E Mason<sup>23</sup>, Nicholas D Socci<sup>1</sup> and Doron Betel<sup>34</sup>\* \* Corresponding author: Doron Betel dob2014@med.cornell.edu Author Affiliations 1 Bioinformatics Core, Memorial Sloan-Kettering Cancer Center, New York, NY, 10065, USA 2 Department of Physiology and Biophysics, Weill Cornell Medical College, New York, NY, 10021, USA 3 Institute for Computational Biomedicine, Weill Cornell Medical College, New York, NY, 10021, USA 4 Division of Hematology/Oncology, Department of Medicine, Weill Cornell Medical College, New York, NY, 10021, USA For all author emails, please log on. Genome Biology 2013, 14:R95 doi:10.1186/gb-2013-14-9-r95



## **Pathway databases**

- msigdbr
- reactome.db with fgsea::reactomePathways()
- a gmt file with fgsea::gmtPathways()
- KEGG pathways via KEGGREST
- Enrichr pathways <a href="http://amp.pharm.mssm.edu/Enrichr/#stats">http://amp.pharm.mssm.edu/Enrichr/#stats</a>
- Gene Ontology via gage or Org.db packages



## **Pathway analysis**

- fgsea
- DOSE/clusterProfiler:
  - fgsea-based and hypergeometric
- limma:
  - camera
  - roast
- gage
- Run step 5



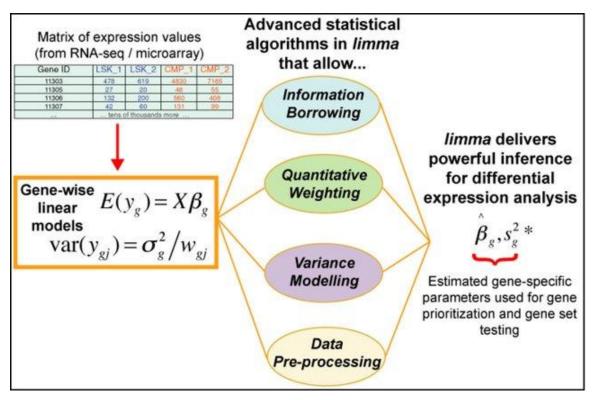
#### **GEOquery**

- Works only for microarrays
- Not for all arrays there is "annotated" (i.e. curated) annotation
- RNA-seq datasets result in an empty matrix
  - Data can be loaded from ARCHS4 file

- Open do\_limma.R
- Run everything



#### limma





#### **Exercises**

- Plot PCA
- Add batch information to the design, calculate differential expression. Are the results differ?
- Do pathway analysis with fgsea
- Do pathway analysis with camera()
  - compare results to fgsea



#### Summary

- There are several common RNA-seq pipelines: alignment-based and kallisto-like
- Multiple tools for downstream analysis
- Visualize and QC your data