Example Analyses: Using the SGBA-5 with Simulated Data

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2024-08-09

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About

The example analyses presented in this document are a demonstration of how sex-and gender-based analysis (SGBA) can be conducted using results from the Sex- and Gender-Based Analysis Tool – 5 item (SGBA-5) by Putman and Dogra [DOI]. The examples in this document demonstrate a descriptive analysis of simulated SGBA-5 responses and SGBA of simulated health outcomes that are continuous, ordered (Likert scale), or categorical (binary) to examine potential interactions with the biological sex item and two of the gendered aspects of health items.

The examples do not cover all potential analyses that could be done using the SGBA-5 but does provide a solid foundation from which researchers can select from and build upon.

1.1 Further Resources

More information on the SGBA-5, instructions for its use, and rationale for are included in the SGBA-5's documentation [SUPPLEMENTARY MATERIAL URL]. Initial reliability and validity testing of the SGBA-5 are reported in the paper by Putman, Cole, & Dogra [DOI] and in A Putman's thesis work.

Data Structure

2.1 Collected SGBA-5 Responses

After collecting responses from the SGBA-5 you will have a dataset that looks something like this:

```
## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
## v dplyr
              1.1.4
                                   2.1.5
                       v readr
## v forcats
              1.0.0
                       v stringr
                                   1.5.1
## v ggplot2
             3.5.1
                       v tibble
                                   3.2.1
## v lubridate 1.9.3
                                   1.3.1
                       v tidyr
## v purrr
              1.0.2
                                      ## -- Conflicts -----
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()
                 masks stats::lag()
## i Use the conflicted package (<a href="http://conflicted.r-lib.org/">http://conflicted.r-lib.org/</a>) to force all conflicts to become
```

Where:

Table 2.1: Example Data Structure

pt_id	sex	gen_id	gen_exp	gen_role	gen_rel
1	Female	48	99	70	25
2	Male	100	17	99	2
3	Female	64	48	88	40
4	Female	24	46	36	88
5	Male	73	23	19	26

The values in this table are placeholders for the example, not real data

pt_id is the participant identifier.

sex is the SGBA-5 categorical Biological Sex item.

• response options of "male", "female", and "intersex".

gen_id is the SGBA-5 Gender Identity gendered aspect of health item.

• Responses are recorded as ordinal values between 0 to 100 on a feminine to masculine scale (measured in mm if completed on paper).

gen_exp is the SGBA-5 Gender Expression gendered aspect of health item.

• Responses are recorded as ordinal values between 0 to 100 on a feminine to masculine scale (measured in mm if completed on paper).

gen_role is the SGBA-5 Gender Role gendered aspect of health item.

• Responses are recorded as ordinal values between 0 to 100 on a feminine to masculine scale (measured in mm if completed on paper).

gen_rel is the SGBA-5 Gender Relations gendered aspect of health item.

• Responses are recorded as ordinal values between 0 to 100 on a feminine to masculine scale (measured in mm if completed on paper).

2.2 Simulated Data

This example analysis uses simulated data, if you wish to replicate the simulate data the code to do so is included in Appendix A or can be downloaded from this example's github page.

Descriptive Analysis

Note: for conciseness, the following examples will only show results for two of the four gendered aspects of health items from the SGBA-5 (gender identity, and gender roles)

3.1 Visualize Distribution of SGBA-5 Responses

In plot 3.1 we see that there are more participants who report their **biological** sex as assigned as female at birth (n=18) than males (n=12).

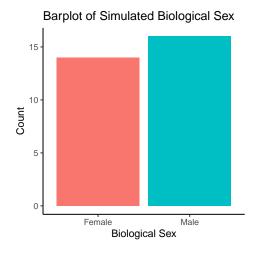


Figure 3.1: Barplot of Biological Sex Responses

Looking at the density plots for **gender identity** and **gender role** (figure 3.2), we see that while both variables are bimodal, the **gender identity** responses is

more strongly bimodal with one peak closer to the feminine side of the feminine-masculine continuum and one peak closer to masculine end of that continuum. Further, we can also see that in general, participants reported their **gender identity** and **roles** as being more feminine, again with the **gender identity** responses showing this trend more strongly than the **gender role** responses.

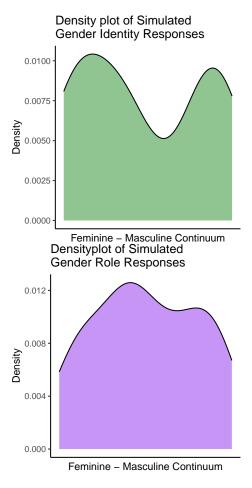


Figure 3.2: Density plots of Gender Identity and Roles

Presently, there is no consensus on what descriptive statistics are most appropriate to report bimodal variables in health research (the typical mean(sd) or median(IQR) will not accurately represent that there is more than one peak in a bimodal variable's frequency distribution). When taken alongside the SGBA-5's assumption that the feminine-masculine continuum doesn't have a true 0 value, it is our suggestion that if researchers decide to report a single variable descriptive statistic for the gendered aspects of health item responses from the SGBA-5, they should provide a

Table 3.1: Potential Interpretation of Sample Means for Gendered Aspect of Health Items.

Mean	Interpretation
>70 55 to 70 45 to 55	"Skews masculine" "More masculine than feminine" "Not strongly skewed"
30 to 45 <30	"More feminine than masculine" "Skews feminine"

This table assumes you have recorded the gendered aspects of health items as 0 being the most feminine score and 100 being the most masculine score.

Table 3.2: Simulated sample characteristics.

SGBA Item	Sample $(n = 30)$
Biological Sex (n(%))	
<i> $>$ Female $<$ /i> $>$	14(47%)
<i $>Intersexi>$	NA
<i $>Malei>$	16(53%)
Gendered Aspect of Health (skew)	
<i>Gender Identity $<$ /i> $<$ i>Gendered Roles $<$ /i>	Not strongly skewed Not strongly skewed

nominal description of skew along the feminine-masculine continuum as their descriptive statistic rather than the numerical average (or other summary statistic). To determine skew of one of the gender variables, the authors suggest calculating the sample's mean score along the feminine-masculine continuum and then classifying the skew using the classifications described in Table 3.1. Please note that these suggested classification guidelines are arbitrary and may not be appropriate in all circumstances.

For the simulated dataset represented in the density plots above, the mean score for the **gender identity** item was 50.2 and 46.8 for the **gender role** item. This means that when reporting descriptive statistics on the simulated sample we could report that: "On the whole, the simulated sample was not strongly skewed on a feminine to masculine continuum for either the gender identity or gender role measures from the SGBA-5".

Taking all these together, an example of a sample characteristics table of the SGBA-5 items in the simulated dataset could be presented as has been displayed in Table 3.2

SGBA of a Continuous Variable

Note: for conciseness, the following examples will only show results for two of the four gendered aspects of health items from the SGBA-5 (gender identity, and gender roles)

4.1 Biological Sex

A good idea is to start by visualizing the continuous variable's distribution disaggregated by sex like the density plot in Figure 4.1. Then calculate disaggregated summary statistics for the continuous variable disaggregated by sex (Table A4), and conduct a statistical test of difference in means (Welch's t- test for this example).

4.1.1 Density Plot

Interpretation: From the above density plots (Figure 4.1) we can see a distinct overlap in the "No interaction example" with suggests that in that example's sample does not have a meaningful difference in the continuous outcome by sex. Conversely, the "Interaction example" density plot has two distinct peaks which suggests that its sample's continuous outcome scores are associated with a participant's sex.

4.1.2 Summary Statistics

Interpretation: As with the density plots, we see that the standard deviations of the continuous variable for both males and females overlap in the "No interaction example" (Table 4.1) - indicating a lack of significant difference by sex. The standard deviations of the continuous variable for both males and females do

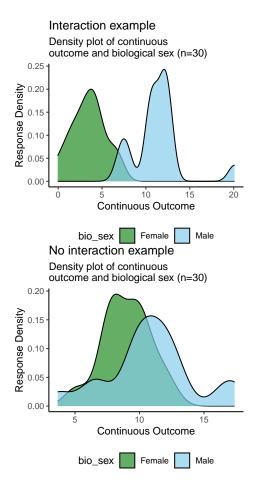


Figure 4.1: Density Plot of Continuous Variable by Biological Sex Examples

Table 4.1: Interation example summary statisitics: Continuous outcome and biological sex

biological sex	n	mean continuous	SD continuous	median continuous	IQR continuous
Female	14	3.5	1.94	4	2
Male	16	11.3	2.95	11	2

Table 4.2: No interation example summary statisities: Continuous outcome and biological sex

biological sex	n	mean continuous	SD continuous	median continuous	IQR continuous
Female	14	9.0	1.84	9	2
Male	16	10.8	3.47	11	3

not overlap in the " $Interaction\ example$ " (Table 4.2) - indicating a potential association between the continuous outcome and sex.

4.1.3 Statistical Test

No interaction: Welch's t-test

Table 4.3: Simulated sample characteristics.

Example	Test	T-score	95% CI	df	p-value
Interaction No interaction	Welch's t-test Welch's t-test		(-3.85, 0.26) (-9.72, -6.02)		0.084 0.000

Appendix

4.2 Simulated Data Creation

Below is the code used to create the simulated data that will be used to create the example SGBA seen throughout the rest of this example analysis.

```
# load libraries -----
library(tidyverse)
## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
## v dplyr 1.1.4 v readr 2.1.5
## v forcats 1.0.0
                     v stringr 1.5.1
## v ggplot2 3.5.1 v tibble 3.2.1
                                1.3.1
## v lubridate 1.9.3
                     v tidyr
## v purrr
            1.0.2
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag() masks stats::lag()
## i Use the conflicted package (<a href="http://conflicted.r-lib.org/">http://conflicted.r-lib.org/</a>) to force all conflicts to become
# set seed for predictability
set.seed(42, kind = "Mersenne-Twister")
# write simulation functions ------
# function for simulating bimodal gender variables
binom_bounded <- function(</pre>
   n, prop, mean1, sd1, mean2, sd2, lim_low = -Inf, lim_up = Inf, round
   ){
 # error checking
 if(prop > 1 || prop< 0){stop("binomial proportion should be between 0 and 1")}</pre>
 if(n < 1){stop("n must be greater than or equal to 1")}</pre>
 if(sd1 < 0 | | sd2 < 0){stop("standard deviations must be non-negative")}
 # create index
```

index <- rbinom(n, size = 1, prob = prop)</pre>

```
# simulate each peak using a bounded `rnorm()`
 binom_sim1 <- index * qnorm(</pre>
   runif(
      n, pnorm(lim_low, mean1, sd1), pnorm(lim_up, mean1, sd1)
     ),
   mean1, sd1)
 binom_sim0 <- (1 - index) * qnorm(</pre>
    runif(
      n, pnorm(lim_low, mean2, sd2), pnorm(lim_up, mean2, sd2)
   mean2, sd2)
 binom sim <- binom sim0 + binom sim1
 binom_sim <- round(binom_sim, round)</pre>
 binom sim
# function for simulating Likert/ordinal outcome variable
ordinal_sim <- function(</pre>
   n, mean, sd, lim_low = -Inf, lim_up = Inf
    ){
 ord_sim <- qnorm(</pre>
   runif(
      n, pnorm(lim_low, mean, sd), pnorm(lim_up, mean, sd)
      ),
   mean, sd)
 ord_sim <- round(ord_sim, 0)</pre>
 ord_sim
# simulate SGBA-5 data with n of 30 -----
# biological sex: categorical (female, intersex, male)
bio_sex <- factor(</pre>
  sample(c('Female', 'Male'), 30, replace=TRUE, prob=c(0.6, 0.4)),
# gender identity: ordered (0 [feminine] to 100 [masculine])
gen_id <- binom_bounded(</pre>
 n = 30, prop = 0.6, mean1 = 20, sd1 = 20, mean2 = 85, sd2 = 15, lim_low = 0,
 lim_up = 100, round = 0
# gender role: ordered (0 [feminine] to 100 [masculine])
```

```
gen_role <- binom_bounded(</pre>
 n = 30, prop = 0.6, mean1 = 25, sd1 = 20, mean2 = 80, sd2 = 20, \lim_{x \to 0} \log x = 0,
 lim_up = 100, round = 0
# simulate example positive outcome data with n of 30 ------
## simulated numerical outcome: -----
# by sex: males (mean = 12, SD = 3), females (mean = 4, sd = 2)
outcome num pos m \leftarrow rnorm(n = 16, mean = 12, sd = 3)
outcome_num_pos_f \leftarrow rnorm(n = 14, mean = 4, sd = 2)
outcome_num_pos_sex <- c(outcome_num_pos_f, outcome_num_pos_m)</pre>
# by g_{id}: high (mean = 12, SD = 3), low (mean = 4, sd = 2)
outcome_num_pos_gid_high <- rnorm(n = 15, mean = 12, sd = 3)
outcome_num_pos_gid_low <- rnorm(n = 15, mean = 4, sd = 2)
outcome_num_pos_gid <- c(outcome_num_pos_gid_high, outcome_num_pos_gid_low)
# by g_{role}: high (mean = 12, SD = 3), low (mean = 4, sd = 2)
outcome_num_pos_grol_high <- rnorm(n = 12, mean = 12, sd = 3)
outcome_num_pos_grol_low <- rnorm(n = 18, mean = 7, sd = 2)
outcome_num_pos_grol <- c(outcome_num_pos_grol_high, outcome_num_pos_grol_low)
## simulated Likert outcome: ------
# by sex males (mean = 2, SD = 1), females (mean = 5, sd = 2)
outcome_ord_pos_m <- ordinal_sim(</pre>
 n = 16, mean = 2, sd = 1, lim_low = 1, lim_up = 7
outcome_ord_pos_f <- ordinal_sim(</pre>
 n = 14, mean = 5, sd = 1, \lim_{\infty} 100 = 1, \lim_{\infty} 100 = 7
outcome_ord_pos_sex <- c(outcome_ord_pos_f, outcome_ord_pos_m) %>%
  factor(., levels = c(1,2,3,4,5,6,7), ordered = TRUE)
# by q_id: high (mean = 5, SD = 2), low (mean = 1, sd = 2)
outcome_ord_pos_high <- ordinal_sim(</pre>
 n = 15, mean = 5, sd = 2, \lim_{\infty} 10w = 1, \lim_{\infty} 10w = 7
outcome_ord_pos_low <- ordinal_sim(</pre>
 n = 15, mean = 1, sd = 2, lim_low = 1, lim_up = 7
```

```
outcome_ord_pos_gid <- c(outcome_ord_pos_high, outcome_ord_pos_low) %>%
 factor(., levels = c(1,2,3,4,5,6,7), ordered = TRUE)
# by q_role: high (mean = 5, SD = 2), low (mean = 1, sd = 2)
outcome_ord_pos_high <- ordinal_sim(</pre>
 n = 12, mean = 5, sd = 2, lim_low = 1, lim_up = 7
outcome_ord_pos_low <- ordinal_sim(</pre>
 n = 18, mean = 1, sd = 2, lim_low = 1, lim_up = 7
outcome_ord_pos_grol <- c(outcome_ord_pos_high, outcome_ord_pos_low) %>%
 factor(., levels = c(1,2,3,4,5,6,7), ordered = TRUE)
## simulated binary outcome: -----
# by sex males (yes = .2, no = .8), females (yes = .8, no = .2)
outcome_bin_pos_m <- sample(</pre>
  c('yes','no'), 16, replace = TRUE, prob = c(.2, .8)
outcome_bin_pos_f <- sample(</pre>
 c('yes','no'), 14, replace = TRUE, prob = c(.8, .2)
outcome_bin_pos_sex <- append(outcome_bin_pos_f, outcome_bin_pos_m) %>%
 factor()
# by g_id: high (yes = .4, no = .6), low (yes = .8, no = .2)
outcome_bin_pos_high <- sample(</pre>
 c('yes','no'), 15, replace = TRUE, prob = c(.4, .6)
outcome_bin_pos_low <- sample(</pre>
 c('yes','no'), 15, replace = TRUE, prob = c(.8, .2)
outcome_bin_pos_gid <- append(outcome_bin_pos_high, outcome_bin_pos_low) %>%
 factor()
# by g_{rol}: high (yes = .3, no = .5), low (yes = .8, no = .2)
outcome_bin_pos_high <- sample(</pre>
 c('yes','no'), 12, replace = TRUE, prob = c(.3, .7)
outcome_bin_pos_low <- sample(</pre>
```

```
c('yes','no'), 18, replace = TRUE, prob = c(.8, .2)
outcome_bin_pos_grol <- append(outcome_bin_pos_high, outcome_bin_pos_low) %>%
 factor()
# simulate example negative outcome data with n of 30 ------
# simulated numerical outcome: continuous (mean = 10, SD = 3)
outcome_num_neg \leftarrow rnorm(n = 30, mean = 10, sd = 3)
# simulated Likert outcome: 7-point Likert scale (mean = 4, SD = 2)
outcome_ord_neg <- ordinal_sim(</pre>
 n = 30, mean = 4, sd = 2, lim_low = 1, lim_up = 7
# simulated binary outcome: categorical (yes, no)
outcome_bin_neg <- factor(</pre>
  sample(c('yes','no'), 30, replace = TRUE, prob = c(0.67, 0.33))
)
# create example data frame -----
# combine into dataframe
sim_data <- tibble(</pre>
 bio_sex, gen_id, gen_role, #outcome_bin_pos,
 outcome_num_neg, outcome_ord_neg, outcome_bin_neg
  ) %>% arrange(., bio_sex) %>%
  cbind(., outcome_num_pos_sex) %>%
  cbind(., outcome_ord_pos_sex) %>%
  cbind(., outcome_bin_pos_sex) %>%
  arrange(., gen_id) %>%
  cbind(., outcome_num_pos_gid) %>%
  cbind(., outcome_ord_pos_gid) %>%
  cbind(., outcome_bin_pos_gid) %>%
  arrange(., gen_role) %>%
  cbind(., outcome_num_pos_grol) %>%
  cbind(., outcome_ord_pos_grol) %>%
  cbind(., outcome_bin_pos_grol)
# save simulated df
write_csv(sim_data, file = "sim-data.csv")
```

R6

4.2.1 Session Info for Data Creation

```
sessioninfo::session info(pkgs = "loaded")
##
   setting value
   version R version 4.4.1 (2024-06-14)
##
##
   os
           macOS Sonoma 14.5
           aarch64, darwin20
##
   system
##
   ui
           X11
##
   language (EN)
  collate en_US.UTF-8
##
##
   ctype
           en_US.UTF-8
##
           America/Toronto
   tz
##
   date
           2024-08-09
##
   pandoc
           3.1.11 @ /Applications/RStudio.app/Contents/Resources/app/quarto/bin/tool
##
## - Packages -----
              * version date (UTC) lib source
   package
                       2022-11-15 [1] CRAN (R 4.4.0)
##
   bit
               4.0.5
               4.0.5
                       2020-08-30 [1] CRAN (R 4.4.0)
##
   bit64
##
  bookdown
               0.40
                       2024-07-02 [1] CRAN (R 4.4.0)
               3.6.3
                       2024-06-21 [1] CRAN (R 4.4.0)
##
              2.1-0
                       2023-01-23 [1] CRAN (R 4.4.0)
##
   colorspace
                       2024-06-20 [1] CRAN (R 4.4.0)
##
   crayon
              1.5.3
              0.6.36 2024-06-23 [1] CRAN (R 4.4.0)
##
   digest
##
   dplyr
              * 1.1.4
                       2023-11-17 [1] CRAN (R 4.4.0)
               0.24.0 2024-06-10 [1] CRAN (R 4.4.0)
## evaluate
## fansi
               1.0.6
                       2023-12-08 [1] CRAN (R 4.4.0)
## fastmap
               1.2.0
                       2024-05-15 [1] CRAN (R 4.4.0)
              * 1.0.0
                       2023-01-29 [1] CRAN (R 4.4.0)
##
  forcats
                       2022-07-05 [1] CRAN (R 4.4.0)
##
   generics
               0.1.3
##
   ggplot2
              * 3.5.1
                       2024-04-23 [1] CRAN (R 4.4.0)
               1.7.0
                       2024-01-09 [1] CRAN (R 4.4.0)
##
   glue
##
   gtable
               0.3.5
                       2024-04-22 [1] CRAN (R 4.4.0)
               1.1.3
                       2023-03-21 [1] CRAN (R 4.4.0)
##
   hms
             0.5.8.1 2024-04-04 [1] CRAN (R 4.4.0)
##
   htmltools
##
   knitr
              1.47
                       2024-05-29 [1] CRAN (R 4.4.0)
   lifecycle
              1.0.4
                       2023-11-07 [1] CRAN (R 4.4.0)
##
             * 1.9.3
                       2023-09-27 [1] CRAN (R 4.4.0)
##
   lubridate
##
   magrittr
               2.0.3
                       2022-03-30 [1] CRAN (R 4.4.0)
##
   munsell
               0.5.1
                       2024-04-01 [1] CRAN (R 4.4.0)
##
   pillar
               1.9.0
                       2023-03-22 [1] CRAN (R 4.4.0)
##
               2.0.3 2019-09-22 [1] CRAN (R 4.4.0)
   pkgconfig
## purrr
              * 1.0.2 2023-08-10 [1] CRAN (R 4.4.0)
```

2.5.1 2021-08-19 [1] CRAN (R 4.4.0)

```
## readr
             * 2.1.5
                     2024-01-10 [1] CRAN (R 4.4.0)
            1.1.4 2024-06-04 [1] CRAN (R 4.4.0)
## rlang
## rmarkdown 2.27
                      2024-05-17 [1] CRAN (R 4.4.0)
## rstudioapi 0.16.0 2024-03-24 [1] CRAN (R 4.4.0)
             1.3.0
                     2023-11-28 [1] CRAN (R 4.4.0)
## scales
## sessioninfo 1.2.2 2021-12-06 [1] CRAN (R 4.4.0)
## stringi
             1.8.4 2024-05-06 [1] CRAN (R 4.4.0)
            * 1.5.1 2023-11-14 [1] CRAN (R 4.4.0)
## stringr
            * 3.2.1
                     2023-03-20 [1] CRAN (R 4.4.0)
## tibble
          * 1.3.1
                     2024-01-24 [1] CRAN (R 4.4.0)
## tidyr
## tidyselect 1.2.1 2024-03-11 [1] CRAN (R 4.4.0)
## tidyverse * 2.0.0 2023-02-22 [1] CRAN (R 4.4.0)
## timechange 0.3.0 2024-01-18 [1] CRAN (R 4.4.0)
## tzdb
             0.4.0 2023-05-12 [1] CRAN (R 4.4.0)
              1.2.4 2023-10-22 [1] CRAN (R 4.4.0)
## utf8
## vctrs
              0.6.5 2023-12-01 [1] CRAN (R 4.4.0)
              1.6.5 2023-12-05 [1] CRAN (R 4.4.0)
## vroom
## withr
             3.0.0 2024-01-16 [1] CRAN (R 4.4.0)
## xfun
             0.45
                      2024-06-16 [1] CRAN (R 4.4.0)
              2.3.8 2023-12-11 [1] CRAN (R 4.4.0)
## yaml
##
## [1] /Library/Frameworks/R.framework/Versions/4.4-arm64/Resources/library
## -----
```