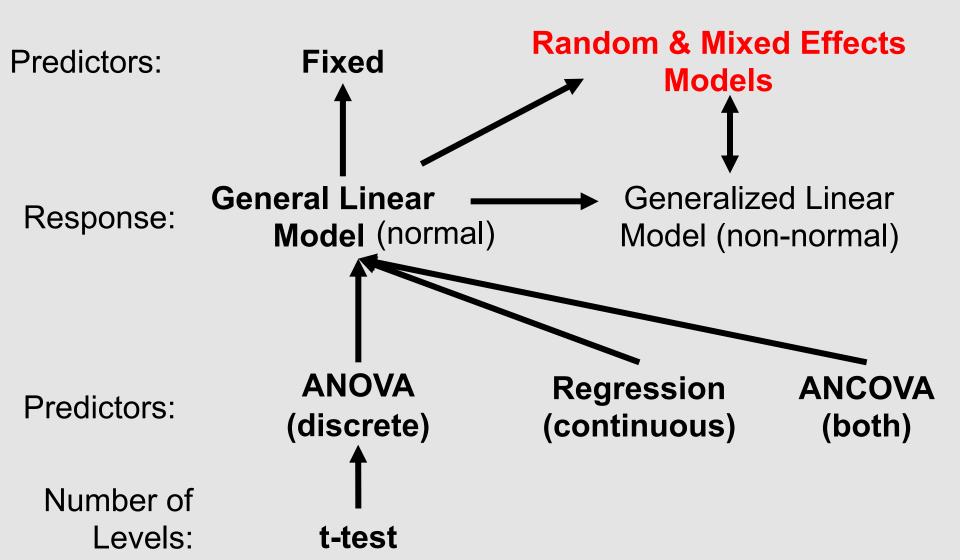
Modeling random effects and mixed effect models part I

SMEE ZOL851 Tuesday Nov 27th 2012

 $Y \sim N(X\beta, \Sigma)$

Continuity of Statistical Approaches

Process Models



Readings

In the Mixed model folder in ANGEL I have put a number of potentially useful readings:

Bolker book: pp. 316-328

Gelman and Hill: Chapters 11-13 (Start with11)

Hadfield: Course notes for Mixed Models using MCMCglmm

Some papers

Bolker et al. 2008. Generalized linear mixed models: a practical guide for ecology and evolution. TREE.

Bates, D. [R] Imer, p-values and all that. (Why p-values are can be so difficult to determine for mixed models, along with df).

Readings

In addition there are a number of books which may be useful to you.

Pinheiro, J. & Bates, D. 2000. Mixed-effects models in S and S-Plus. (E-book, link available in the resource section).

Zuur, A. et al. 2009. Mixed effects Models and Extensions in Ecology with R.

Note: While this book has a lot of potential, it is also full of errors. So please use additional sources for understanding the basics.

The problem

- So far we have assumed that are observations are IID.
- Identically distributed.: The observations come from the same underlying error distribution.
- Independently distributed: The observations are all independent of one another, i.e. cov(y_i,y_i)=0

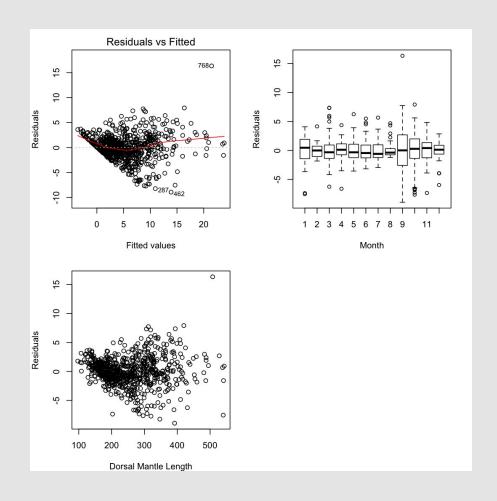
The problem

- However both of these assumptions are violated with many data sets, and can lead to many issues with estimation and inference.
- Thus we need to stop thinking about the residual error as the "dumping ground" for unfit variance, and start incorporating structure into this part of the model as well!!

Outline

- Heterogeneity in residual variances
- Replication and Independence of errors
- Accounting for grouping in observations
- Fixed vs Random effects
- The addition of random effects to our general linear model
 - Familiar Approaches
 - Blocking factor
 - Repeated measures
 - Nested Designs
- Mixed-Effects Models in R

Fitting a model for relating squid size and month to testis size as a measure of sexual maturity.



•When dealing with such heterogeneity we want to model the residual variance.

$$\varepsilon_i = N(0, \sigma^2 \times DML_i)$$

A fixed variance structure where the variance is multiplied by one of the continuous variables.

```
design.M.1 <- model.matrix(~ DML, data=Squid) # Design matrix

NLL.M.1.HET.VAR <- function(b0, b1, sig){
    het.sig <- sig*sqrt(design.M.1[,2]) # sigma as a function
    det <- b0 + b1*design.M.1[,2] # deterministic part like before.
    -sum(dnorm(Squid$Testisweight, mean=det, sd=het.sig, log=T))}

M.1.Het <- mle2(NLL.M.1.HET.VAR , start=list(b0=0, b1=0, sig=0.01))</pre>
```

•When dealing with such heterogeneity we want to model the residual variance.

$$\varepsilon_i = N(0, \, \sigma^2 \times DML_i)$$

A fixed variance structure where the variance is multiplied by one of the continuous variables.

In R we would use the gls() in the nlme library

vf1Fixed <- varFixed(~DML) #specifying the variance structure
M.1.gls <- gls(Testisweight ~ DML*fMONTH, weights = vf1Fixed,
data=Squid) # only difference is incorporating it into the
model.</pre>

There are many other variance structures that can be used

$$\varepsilon_{ij} = N(0,\sigma^2_j)$$
 Different variance for each treatment level of a factor, varident

In R we would use the gls() in the nlme library

```
vf2 <- varIdent(~ 1| fMONTH) #specifying the variance structure
M.1.gls <- gls(Testisweight ~ DML*fMONTH, weights = vf2,
data=Squid) # only difference is incorporating it into the
model.</pre>
```

There are many other variance structures that can be used

| Name of the function in nlme | What it does |
|------------------------------|--|
| VarFixed | Fixed variance based on the covariate |
| Varldent | Different variances per treatment levels |
| VarPower | Power of the variance covariate |
| VarExp | Exponential of variance covariate |
| VarConstPower | Constant + power of variance covariate |
| VarComb | combination |

After Zuur et al. 2009

- There are many other variance structures that can be used
- •For non-linear models there is the gnls() function in nlme
- ?varClasses

Replication and Independence

- Statistics is based on collecting <u>independent samples</u> from a population. The degree to which those samples reflect the true population depends on the number of samples collected
- So, all inferential statistics depend on independence
- Replicates must be more than just unbiased, they must be independent
 - Pairs of samples treated alike are on average no more similar or dissimilar than those treated differently - from Hurlbert, 1984
 - Shading may cause plants to grow taller and two plants experiencing the same degree of shading might both be tall but they are independent if the source of the shading differs between the two plants

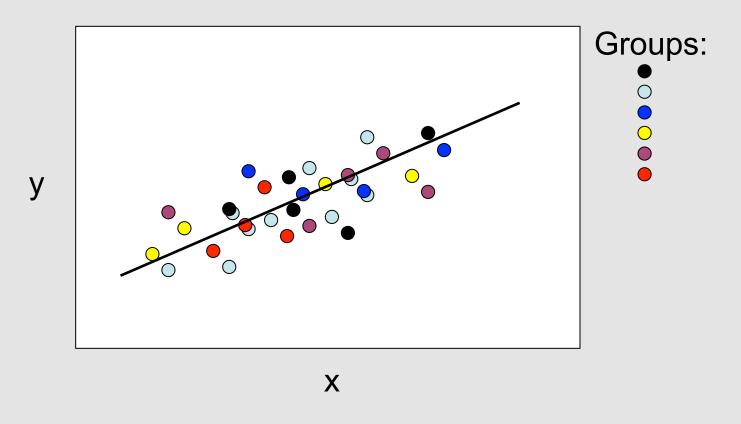
Independence of Errors

- Independence of <u>errors</u> is the key assumption
- Lack of independence comes from some sort of "grouping" of the data.

Examples:

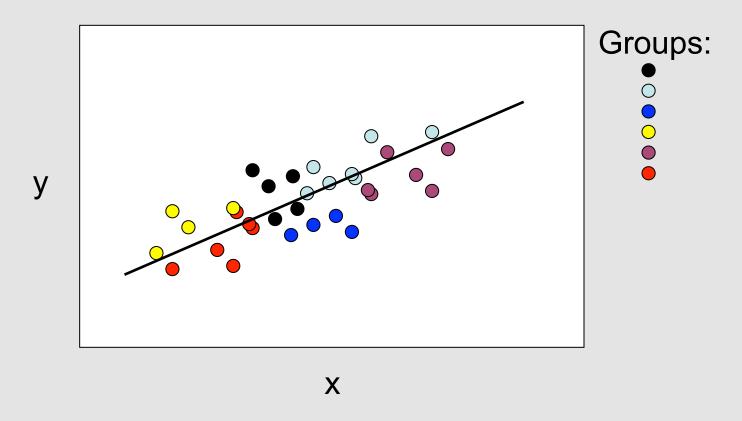
- Multiple measurements made per individual
- Observations clustered in space or time
- Treatment randomly applied to groups of replicates rather than individual replicates
- Some individuals come from the same genetic strain

Underlying Structure



Errors are independent of group structure

Underlying Structure

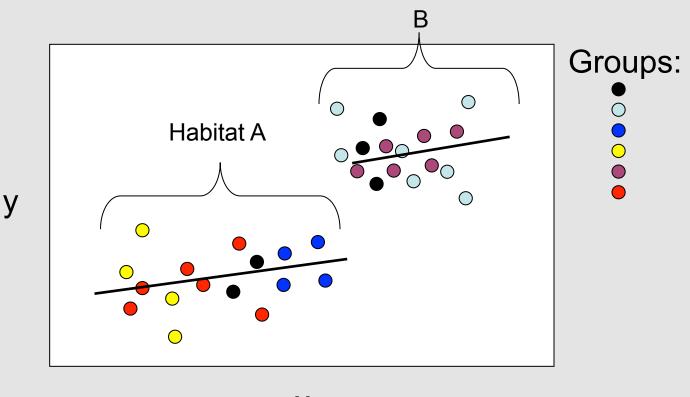


Errors are **not** independent of group structure!!

Accounting for Grouping

- Samples might not be independent but we can and need to account for this lack of independence in our analysis.
- Independence as "virtue" and "truth"

Underlying Structure



Covariance structures for time series

Under the assumption of independence we assume that there is no covariance between years.

$$\varepsilon = \begin{bmatrix} \varepsilon_1 \\ \varepsilon_2 \\ \cdot \\ \varepsilon_n \end{bmatrix}$$

$$Var(\varepsilon) = \sigma_{\varepsilon}^{2} \times \mathbf{I} = \begin{bmatrix} \sigma_{\varepsilon}^{2} & 0 & 0 & 0 \\ 0 & . & 0 & 0 \\ 0 & 0 & . & 0 \\ 0 & 0 & \sigma_{\varepsilon}^{2} \end{bmatrix}$$

For n observations

Covariance structures for time series

- Clearly this assumption is not valid so we should try some other types of covariance structures.
- The simplest is the compound symmetry structure
- Assumes that there is a constant co-variance between errors, regardless of time.

Compound symmetry

- The simplest is the compound symmetry structure
- Assumes that there is a constant co-variance between errors, regardless of time.

$$Var(\varepsilon) = \begin{cases} \theta + \sigma_{\varepsilon}^{2} & \theta & \theta & \theta \\ \theta & \cdot & \theta & \theta \\ \theta & \theta & \cdot & \theta \\ \theta & \theta & \theta & \theta + \sigma_{\varepsilon}^{2} \end{cases}$$

```
# Compound symmetry structure in R using gls()
M.1 <- gls(Birds ~ Rainfall + Year, data=Hawaii,
correlation=corCompSymm(form =~Year))</pre>
```

This adds just one more parameter, theta. It is still pretty simplistic though.

Auto-regressive (AR)-1 auto-correlation

- Auto-regressive order model 1
- Models residual at time s as a function of the residual of time s-1 plus error.

$$Cor(\varepsilon) = \frac{\rho^{1}}{\rho^{2}} \quad \frac{\rho^{2}}{\rho^{1}} \quad \frac{\rho^{3}}{\rho^{1}} \quad \frac{\rho^{2}}{\rho^{1}}$$

$$\frac{\rho^{3}}{\rho^{3}} \quad \frac{\rho^{2}}{\rho^{2}} \quad \frac{\rho^{1}}{\rho^{1}} \quad 1$$

```
# Compound symmetry structure in R using gls()
M.1 <- gls(Birds ~ Rainfall + Year, data=Hawaii,
correlation=corAR1(form =~Year) )</pre>
```

The further you are in time, the less covariation.

ARMA

 Auto-regressive moving average.. Yet more complex models...

Mixed-Effects Models

 General (-ized) Linear Models that contain both fixed and random effects

- So far we have considered only fixed effects
- Models to predict the response to some predictors when the data are grouped according to one or more classification variables (random effects)

Random & Mixed-effects models

 Such models are also referred to as multilevel models (mostly in the social sciences and psychology, but increasingly in biology).

 Also called hierarchical models (which will often refer to a nesting structure).

There may actually be some subtle distinctions implied by these names, but the amount of variation in how people use these terms is far greater than the variation that may be implied by the models...

Fixed effects VS. Random effects

 Break into small groups and come up with a definition of each.

Fixed VS. Random effects

 Break into small groups and come up with a definition of each.

Fixed VS. Random effects: Some definitions

- Fixed effects are constant across individuals, and random effects vary. (Kreft & De Leeuw 1998 p.12).
- Effects are *fixed* if they are interesting in themselves, or *random* if there is interest in the underlying population (Searle, Casella & McCulloch 1992).
- When a sample (close to) exhausts the population, the corresponding variable is *fixed*; when the sample is a small part of the population the corresponding variable is *random* (Green and Tukey 1960).

Fixed VS. Random effects: Some definitions

- If an effect is assumed to be a realized value of a random variable, it is called a *random* effect (LaMotte 1983).
- Fixed effects are estimated using LS or ML and random effects are estimated with shrinkage ("linear unbiased predictions"). This is the common definition for multilevel models.

Fixed vs. Random Factors

Fixed

- Specific levels of factor have some interest / significance
- Levels are fixed and repeatable
- Interested in estimating the means for these levels

 Examples: experimental treatment, habitat, age class, sex, species, year?

Random

- Levels of factor do not have significance
- Repeatability of levels isn't important
- Levels "drawn" at random from a population of possible levels
- Interested in measuring or correcting for variation among levels

Examples: Subject, plot, year?

Thus one of the most difficult parts about mixed effect models is figuring out which variables should be fixed and which should be random.

Gelman & Hill choose not to make a distinction in most cases. They treat all variables as random (which is consistent with a Bayesian philosophy of estimation).

Examples of Common Designs with Random Effect

1. Blocking Factors

- Lump similar (usually spatially close) plots together
- Analogous to the pairwise t-test
- Called "within-subject" in psychology
 - Also equivalent to giving same plant two different treatments and measuring response each time
- Pulls variation into the block and out of the explained error term (ε)

Example: Interested in primary production. 5 fields in different locations. Randomly apply treatments within fields. Include "Field" as a blocking factor to account for variation among fields

Examples of Common Designs with Random Effect

2. Repeated Measures - ANOVAR

- Also called within-subject, longitudinal study
- Record multiple measures on an individual under different conditions
- Similar to blocking but the "plot" is an individual organism and the blocking factor is time
- Many scenarios:
 - Using same individuals to reduce variance
 - Growth, recruitment or other variable changing over time
 - Before/After

Example: Measure body temperature in mice when non-reproductive, pregnant and lactating.

Examples of Common Designs with Random Effect

Nested Designs

- Exploring variation with spatial scale
- Subplots within plots within sites
- When each level of factor A is only observed in one level of a different factor B then A is nested within B

Examples:

- Growth of sea lamprey larvae within streams within lakes.
- Growth of nestlings within clutches within years
- Primary production within subplots within exclosures

Let's remind ourselves of the sex comb teeth data set from *Drosophila*.

- Trying to account for variation in number of SCT on male flies.
- We had numerous factors we were looking at:
- Genotype (2 treatment levels: Dll and wt).
- Temperature (2 treatment levels: 25 & 30°C)
- Line (strain of flies: 27 levels)

Let's remind ourselves of the sex comb teeth data set from *Drosophila*.

 Work in groups to figure out which variables you think should be fixed or random, and why.

Benefits of Mixed-Effects Models

- Allow you to correct for lack of independence of errors
 - Account for underlying group structure in the data
- Handle balanced and unbalanced designs with missing data.
- Utilizes all of the data when making "group" level predictors (I.e. the "best linear unbiased predictions, BLUPs" for random co-efficients).

Let's look at some ways to write a pure random effects models.

In the simplest model we one term that some consider "fixed", the overall population mean (or intercept, although we will see that term gets confusing).

y = a + e
$$e \sim N(0, \sigma_e^2)$$
 Just a population mean or

$$y \sim N(\mu_a, \sigma^2)$$

Let's look at some ways to write a pure random effects models.

In the simplest Random effects model we one term that some consider "fixed", the overall population mean (or intercept, although we will see that term gets confusing). We also have a single **random** (grouping) variable.

$$y = a_j + e$$

$$e \sim N(0, \sigma_e^2)$$

$$a_j \sim N(\mu_a, \sigma_a^2)$$

Random effect

$$y_{ij} \sim N(\mu + a_j, \sigma_e^2)$$

 $a_j \sim N(0, \sigma_a^2)$

The important point is that we are modeling some random variables, so that there are additional sources of variance.

In general these can all be combined into a model like

$$\mathbf{y} = \mathbf{X}\boldsymbol{\beta} + \mathbf{Z}\boldsymbol{u} + \boldsymbol{e}$$

$$\mathbf{y} = \mathbf{X}\boldsymbol{\beta} + \mathbf{Z}\boldsymbol{u} + \boldsymbol{e}$$

X & Z are incidence (X is still called the design matrix) Beta and u are what we are trying to estimate.

$$y_i \sim N(X_i\beta + a_{j[i]}, \sigma_e^2)$$
$$a_i \sim N(0, \sigma_a^2)$$

Sex comb teeth in Drosophila example.

In the simplest case let's say way we wanted to generate a pure random effects model where we estimate the varying intercepts model for line effects.

Mixed Effects Models in R

- Using mixed models requires more complicated calculation approach
 - SAS uses PROC MIXED, glmmixed, nlmixed
 - R has MANY options
 - Ime() is an older function from the nlme package that works well
 - Imer() is a new function in the package Ime4
 - Imer handles both nested and non-nested random factors as well as non-normal error terms
 - MCMCglmm() which we will use.
- Using Imer
 - MCMCglmm(), Imer() work in a similar fashion to Im and glm
 - Takes formula
 - Gives object that goes to summary, print, anova, etc.
 - Specify model in the same way, but random effects are indicated slightly differently
 - e.g. Y~ Fixed.1+Fixed.2+(1|Random.1) Imer
 - Y~ Fixed.1. + Fixed.2, random=~ Random.1

How do we code this in R?

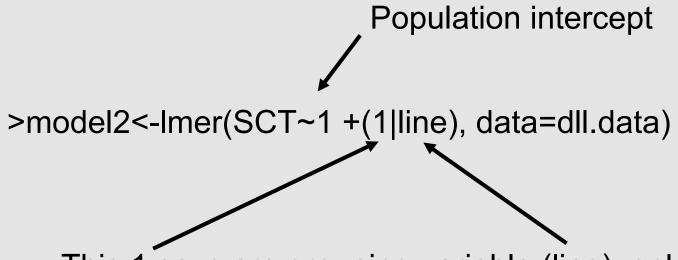
- Load Ime4 package (install first)
- Start with original linear model
 >model<-lm(SCT~1, dll.data)
- Add random effect for "line"

```
>model2<-lmer(SCT~1 +(1|line), data=dll.data)
>model2 <- MCMCglmm(SCT ~1, random=~line, data=dll.data)
This model is conceptually similar to
>model3<-lm(SCT~ 0 + line, data=dll.data)
```

Why? Let's dissect this (on the board)

Coding models in Imer() (from Ime4 library)

How do we code this in R?



This 1 says are grouping variable (line), only needs to vary for the intercept term (the 1).

Coding models in MCMCglmm() How do we code this in R?

Population intercept

>model2.MCMC<- MCMCglmm(SCT~1, random=~line, data=dll.data)

This says the grouping variable (line), only needs to vary for the implicit intercept term.

- > model2.MCMC<- MCMCglmm(SCT~1, random=~us(1):line, data=dll.data)
- > model2.MCMC<- MCMCglmm(SCT~1, random=~idh(1):line, data=dll.data)

All three of these are identical models, we will discuss why soon.

The original package for such models was nlme, and it is still useful for a variety of things.

It specifies things in a similar manner to MCMCglmm...

```
require(nlme) # original mixed model library for R/S
lme(SCT ~ 1, data=dll.data, random=~ line)
```

How might we code a more complicated model

 Let's say we wanted to model the effects of tarsus, and account for line level variation? How might we do it?

Add some figures to help this along plots of lines with varying slopes and intercepts

Coding models in MCMCglmm()

How do we code this in R?

What you probably **DO NOT** want is:

```
>model2.MCMC<- MCMCglmm(SCT~1 + tarsus, random=~sct:line, data=dll.data)
```

Coding models in MCMCglmm() and Ime4 How do we code this in R?

More likely you want to consider one of these.

```
model2.MCMC<- MCMCglmm(SCT~1 + tarsus, random=~us(1 + tarsus):line, data=dll.data)
```

model2.lmer<- lmer(SCT~1 + tarsus + (1 + tarsus| line), data=dll.data)

These two are equivalent representations

Or this:

model2.MCMC<- MCMCglmm(SCT~1+ tarsus, random=~idh(1 + tarsus):line, data=dll.data)

model2.lmer<- lmer(SCT ~ 1 + tarsus + (1 | line) + (0 + tarsus| line), data=dll.data) These two are equivalent representations

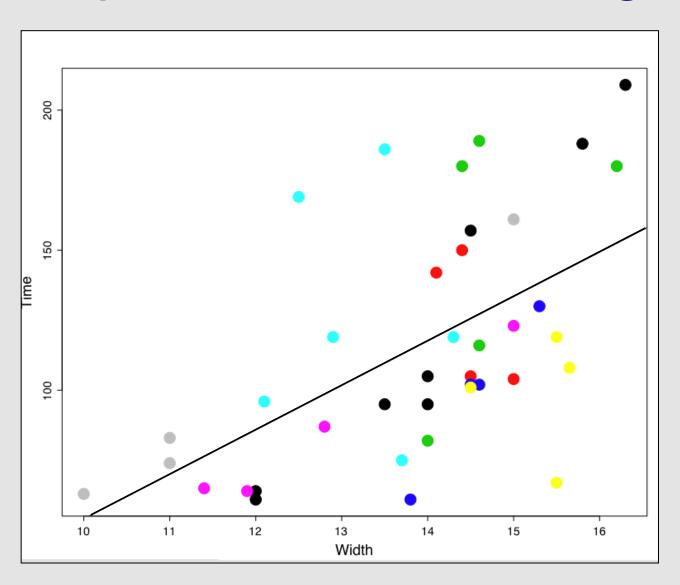
These differ in whether they fit the co-variances between the parameter estimates for random effects

Variance structures

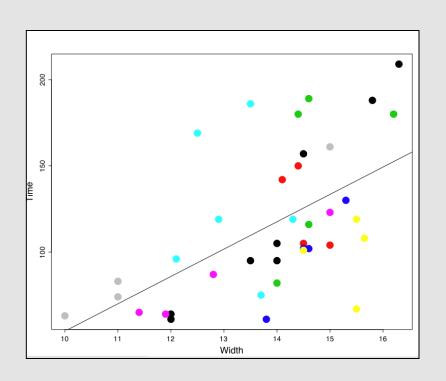
| lmer | MCMCglmm/asreml | No. Parameters | Variance | Correlation |
|---------------------|-----------------|----------------|---|---|
| (1 dam) | dam | 1 | $\left[\begin{array}{ccc} V & V & V \\ V & V & V \\ V & V & V \end{array}\right]$ | $\left[\begin{array}{ccc} 1 & 1 & 1 \\ 1 & 1 & 1 \\ 1 & 1 & 1 \end{array}\right]$ |
| (sex-1 dam) | us(sex):dam | 6 | $\left[\begin{array}{ccc} V_{1,1} & C_{1,2} & C_{1,3} \\ C_{1,2} & V_{2,2} & C_{2,3} \\ C_{1,3} & C_{2,3} & V_{3,3} \end{array}\right]$ | $\left[\begin{array}{ccc}1&r_{1,2}&r_{1,3}\\r_{1,2}&1&r_{2,3}\\r_{1,3}&r_{2,3}&1\end{array}\right]$ |
| (1 sex:dam) | sex:dam | 1 | $\left[\begin{array}{ccc} V & 0 & 0 \\ 0 & V & 0 \\ 0 & 0 & V \end{array}\right]$ | $\left[\begin{array}{ccc} 1 & 0 & 0 \\ 0 & 1 & 0 \\ 0 & 0 & 1 \end{array}\right]$ |
| (1 dam)+(1 sex:dam) | dam+sex:dam | 2 | $\left[\begin{array}{ccc} V_1+V_2 & V_1 & V_1 \\ V_1 & V_1+V_2 & V_1 \\ V_1 & V_1 & V_1+V_2 \end{array}\right]$ | $\left[\begin{array}{ccc}1&r&r\\r&1&r\\r&r&1\end{array}\right]$ |
| - | idh(sex):dam | 3 | $\left[\begin{array}{ccc} V_{1,1} & 0 & 0 \\ 0 & V_{2,2} & 0 \\ 0 & 0 & V_{3,3} \end{array}\right]$ | $\left[\begin{array}{ccc} 1 & 0 & 0 \\ 0 & 1 & 0 \\ 0 & 0 & 1 \end{array}\right]$ |
| - | corh(sex):dam | 4 | $\left[\begin{array}{ccc} V_{1,1} & rV_{1,1}V_{2,2} & rV_{1,1}V_{2,2} \\ rV_{1,1}V_{2,2} & V_{2,2} & rV_{2,2}V_{2,3} \\ rV_{1,1}V_{3,3} & rV_{2,2}V_{3,3} & V_{3,3} \end{array}\right]$ | $\left[\begin{array}{ccc}1&r&r\\r&1&r\\r&r&1\end{array}\right]$ |
| - | cor(sex):dam | 3 | $\left[\begin{array}{ccc} 1 & r_{1,2} & r_{1,3} \\ r_{1,2} & 1 & r_{2,3} \\ r_{1,3} & r_{2,3} & 1 \end{array}\right]$ | $\left[\begin{array}{ccc} 1 & r_{1,2} & r_{1,3} \\ r_{1,2} & 1 & r_{2,3} \\ r_{1,3} & r_{2,3} & 1 \end{array}\right]$ |

From Hadfield 2010, pg 68, course notes for MCMCglmm

Squirrel Cone Handling



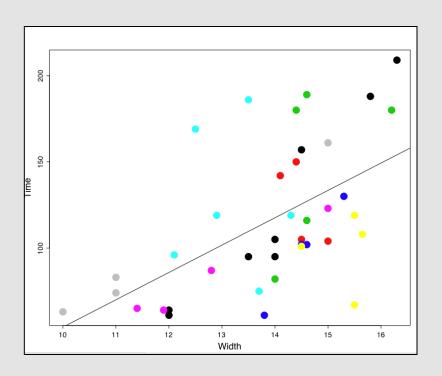
Regression Model



$$Time_{ij} = \beta_0 + \beta_1 Width + \varepsilon_{ij}$$

Errors (e_{ij}) are not independent

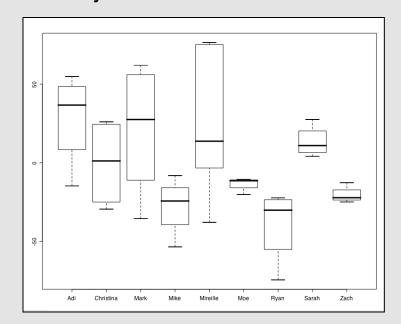
Regression Model



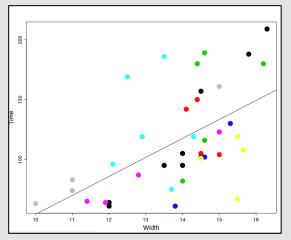
Account for this lack of independence by including a random "Squirrel" effect in the model.

$$Time_{ij} = \beta_0 + \beta_1 Width + \varepsilon_{ij}$$

Errors (e_{ij}) are not independent

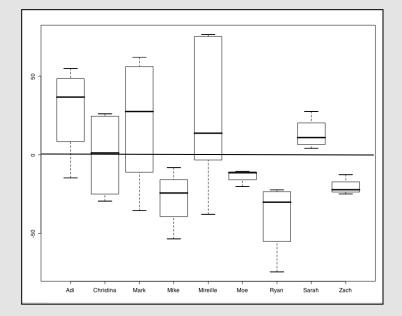


What do the random effects do?



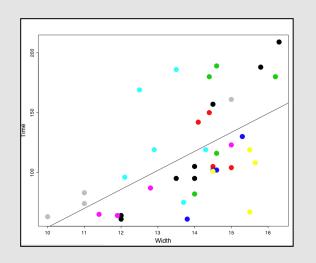
$$Time_{ij} = \beta_0 + \beta_1 Width + \varepsilon_{ij}$$

$$Time_{ij} = \beta_0 + \beta_1 Width + b_j + \varepsilon_{ij}$$



 b_i is the random effect

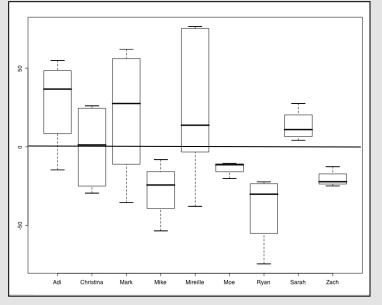
What do the random effects



do?

$$Time_{ij} = \beta_0 + \beta_1 Width + \varepsilon_{ij}$$

$$Time_{ij} = \beta_0 + \beta_1 Width + b_j + \varepsilon_{ij}$$



 b_i is the random effect

$$b_j \sim N(0, \sigma_b^2)$$

$$b_{Adi} = 18.4$$

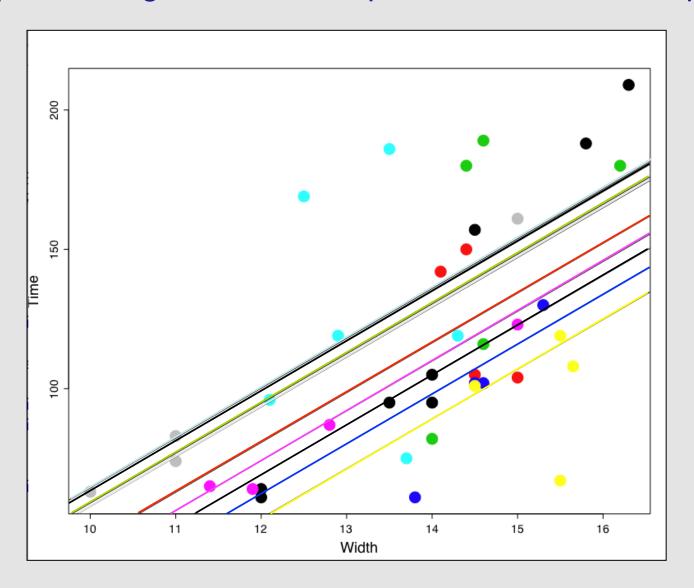
$$b_{Christina} = -0.1$$

$$b_{Mark} = 13.87$$

$$b_{Mike} = -18.5$$

• •

This is the same as fitting separate regressions for each squirrel using a common slope but different intercepts



We now have within group errors and random effects

Remember that our model is:

$$Time_{ij} = \beta_0 + \beta_1 Width + b_j + \varepsilon_{ij}$$

Random effects

Within group errors

$$\begin{vmatrix} b_j \sim N(0, \sigma_b^2) \\ \varepsilon_{ij} \sim N(0, \sigma^2) \end{vmatrix}$$

Yet more complex models

 What if we wanted to model group level variation for the effect of "width" (I.e. group level variation for the slope)?

>model3.MCMC <-MCMCglmm(Time~ Width+ random=~Squirrel, data=cone.handling)

>model3<-lmer(Time~ Width+(1|Squirrel), data=cone.handling)

How would we modify this?

Yet more complex models

 What if we wanted to model group level variation for the effect of "width" (I.e. group level variation for the slope)?

```
>model4.MCMC <-MCMCglmm(Time~ Width + random=~idh(1+ Width):Squirrel, data=cone.handling)
```

>model4<-Imer(Time~ Width+(Width|Squirrel), data=cone.handling)

This tells us that width needs to incorporate group (random) level variation, like a separate slopes model. There is an implicit intercept for lmer (Width|Squirrel) = (1+Width|Squirrel).

Variance structures

| lmer | MCMCglmm/asreml | No. Parameters | Variance | Correlation |
|---------------------|-----------------|----------------|---|---|
| (1 dam) | dam | 1 | $\left[\begin{array}{ccc} V & V & V \\ V & V & V \\ V & V & V \end{array}\right]$ | $\left[\begin{array}{ccc} 1 & 1 & 1 \\ 1 & 1 & 1 \\ 1 & 1 & 1 \end{array}\right]$ |
| (sex-1 dam) | us(sex):dam | 6 | $\left[\begin{array}{ccc} V_{1,1} & C_{1,2} & C_{1,3} \\ C_{1,2} & V_{2,2} & C_{2,3} \\ C_{1,3} & C_{2,3} & V_{3,3} \end{array}\right]$ | $\left[\begin{array}{ccc}1&r_{1,2}&r_{1,3}\\r_{1,2}&1&r_{2,3}\\r_{1,3}&r_{2,3}&1\end{array}\right]$ |
| (1 sex:dam) | sex:dam | 1 | $\left[\begin{array}{ccc} V & 0 & 0 \\ 0 & V & 0 \\ 0 & 0 & V \end{array}\right]$ | $\left[\begin{array}{ccc} 1 & 0 & 0 \\ 0 & 1 & 0 \\ 0 & 0 & 1 \end{array}\right]$ |
| (1 dam)+(1 sex:dam) | dam+sex:dam | 2 | $\left[\begin{array}{ccc} V_1+V_2 & V_1 & V_1 \\ V_1 & V_1+V_2 & V_1 \\ V_1 & V_1 & V_1+V_2 \end{array}\right]$ | $\left[\begin{array}{ccc}1&r&r\\r&1&r\\r&r&1\end{array}\right]$ |
| - | idh(sex):dam | 3 | $\left[\begin{array}{ccc} V_{1,1} & 0 & 0 \\ 0 & V_{2,2} & 0 \\ 0 & 0 & V_{3,3} \end{array}\right]$ | $\left[\begin{array}{ccc} 1 & 0 & 0 \\ 0 & 1 & 0 \\ 0 & 0 & 1 \end{array}\right]$ |
| - | corh(sex):dam | 4 | $\left[\begin{array}{ccc} V_{1,1} & rV_{1,1}V_{2,2} & rV_{1,1}V_{2,2} \\ rV_{1,1}V_{2,2} & V_{2,2} & rV_{2,2}V_{2,3} \\ rV_{1,1}V_{3,3} & rV_{2,2}V_{3,3} & V_{3,3} \end{array}\right]$ | $\left[\begin{array}{ccc}1&r&r\\r&1&r\\r&r&1\end{array}\right]$ |
| - | cor(sex):dam | 3 | $\left[\begin{array}{ccc} 1 & r_{1,2} & r_{1,3} \\ r_{1,2} & 1 & r_{2,3} \\ r_{1,3} & r_{2,3} & 1 \end{array}\right]$ | $\left[\begin{array}{ccc} 1 & r_{1,2} & r_{1,3} \\ r_{1,2} & 1 & r_{2,3} \\ r_{1,3} & r_{2,3} & 1 \end{array}\right]$ |

From Hadfield 2010, pg 68, course notes for MCMCglmm

Best linear unbiased predictor's (BLUP's)

This is where the shrinkage comes in.

For a model where all we were estimating is the group level "means", the use of a mixed model framework would result in.

$$\hat{\alpha}_{j}^{BLUP} \approx \frac{\frac{n_{j}}{\sigma_{e}^{2}} \overline{y}_{j} + \frac{1}{\sigma_{\alpha}^{2}} \overline{y}_{all}}{\frac{n_{j}}{\sigma_{e}^{2}} + \frac{1}{\sigma_{\alpha}^{2}}}$$

Shrinkage in mixed models

$$\hat{\alpha}_{j}^{BLUP} \approx \frac{\frac{n_{j}}{\sigma_{e}^{2}} \overline{y}_{j} + \frac{1}{\sigma_{\alpha}^{2}} \overline{y}_{all}}{\frac{n_{j}}{\sigma_{e}^{2}} + \frac{1}{\sigma_{\alpha}^{2}}}$$

 $\hat{\alpha}_{i}^{BLUP}$ The new predicted value for group j (instead of the mean)

 \overline{y}_j The (LS) mean for group j (unpooled estimate)

The mean over all observations (pooled estimate)

 σ_e^2 Within group variance

 \mathcal{H}_{lpha}^{2} Between group variance \mathcal{H}_{i}^{2} Sample size in group j

Shrinkage in mixed models

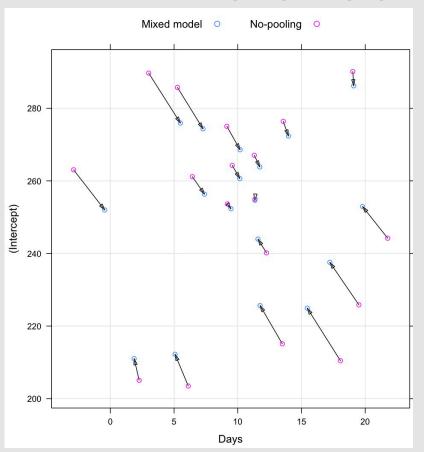
$$\hat{\alpha}_{j}^{BLUP} \approx \frac{\frac{n_{j}}{\sigma_{e}^{2}} \overline{y}_{j} + \frac{1}{\sigma_{\alpha}^{2}} \overline{y}_{all}}{\frac{n_{j}}{\sigma_{e}^{2}} + \frac{1}{\sigma_{\alpha}^{2}}}$$

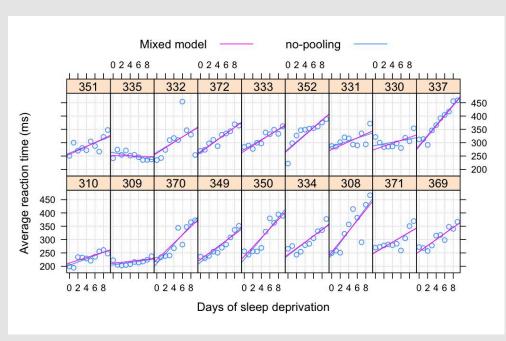
What this means is that group averages E(y_j) with small sample sizes contain less information, and the weighting pulls the BLUP's closer to the overall average.

When sample size is larger for a group the estimate will be closer to the expected value $E(y_i)$.

You can consider this a kind of "partial pooling"

The effects of shrinkage





Example for longitudinal (repeated measure) data

ML vs. REML in Mixed-Effects Models

- Maximum likelihood estimates of variance components maximize the likelihood of the data
 - Do not take into account the fixed effect structure of a model
 - Assume that the fixed effects are known without error
 - Are, therefore, biased downward (similar to the bias in sample variance)
- Restricted ML address this bias by considering only the residuals of the fixed effects in the model
 - Maximize only the part of the likelihood that does not depend on the fixed effects
- This is helpful for coming up with good estimates of variance components, <u>but it</u> <u>causes problems</u> when we want to use **LRT** to assess a change in the fixed effects between two models.s

Assumptions of Mixed-Effects Models

- 1. Within group errors (e_{ij}) are independent, do not differ among groups, have a mean of zero and are normally distributed
- 2. Random effects (b_j) are normally distributed, mean of zero and are independent of one another

Mixed Effects Models Summary

- Fixed Effects estimate means for levels within a factor, random effects estimate variation among levels
- The inclusion of random effects allows us to account for structure in the data
 - Independence of errors
 - Corrects degrees of freedom
- Flexible framework for the analysis of blocked, nested and repeated measures designs
- Handle missing data
- In more advanced scenarios you can model both the response and the variance in random effects

Benefits of Mixed-Effects Models

- Allow you to correct for lack of independence of errors
 - Account for underlying group structure in the data
- Handle balanced and unbalanced designs with missing data.
- Utilizes all of the data when making "group" level predictors (I.e. the "best linear unbiased predictions, BLUPs" for random coefficients).

Software for (generalized) linear Mixed models

Table 1. Capabilities of different software packages for GLMM analysis: estimation methods, scope of statistical models that can be fitted and available inference methods

| | | Penalized quasilikelihood | Laplace | Gauss- Hermite quadrature | random | Wald χ^2 or Wald F tests | Degrees of freedom | MCMC sampling | | Overdispersion |
|----------|--------------|------------------------------|-----------------------|---------------------------------|------------|-------------------------------|--------------------|------------------|----------|----------------|
| SAS | PROC GLIMMIX | ✓ | ∕ ^a | ✓ a | ✓ | ✓ | BW, S, KR | | ∠ | QL |
| | PROC NLMIXED | | | | | / | BW, S, KR | | | Dist |
| R | glmmPQL | ∠ | | | | | BW | | | QL |
| | glmmML | | | | | | | | | |
| | glmer | | 1 | () | | | | (•••) | | QL |
| | glmmADMB | | | | | | | | | Dist |
| | GLMM | ✓ | | | ▶ ? | / | | | - | QL |
| GenStat/ | | | 1 | _ | ✓ | | | ✓ | | Dist |
| ASREML | | | | | | | | | | |
| AD Model | | ✓ | 1 | | / | | | | | ✓ |
| Builder | | | | | | | | | | |
| HLM | | | | _ | | | | | | |
| GLLAAMM | | | | | | | | ✓ | | Dist |
| (Stata) | | | | | | | | | | |
| WinBUGS | | | | | _ | | | _ | | |

Abbreviations: BW, between-within; dist, specified distribution (e.g. negative binomial); KR, Kenward-Roger; QL, quasilikelihood; S, Satterthwaite.

Bolker et al. 2009

Also add RLRsim, Imm, MCMCglmm, sabreR in R and JAGS (Gibbs Sampler), wombat

For general linear mixed models SAS also has PROC MIXED

^aVersion 9.2 only.

Estimation

Table I. Techniques for GLMM parameter estimation, their advantages and disadvantages and the software packages that implement them

| Technique | Advantages | Disadvantages | Software |
|---------------------------|-----------------------------------|--|-----------------------------------|
| Penalized quasilikelihood | Flexible, widely implemented | Likelihood inference inappropriate; | PROC GLIMMIX (SAS), GLMM |
| | | biased for large variance or small means | (Genstat), glmmPQL (R), glmer (R) |
| Laplace approximation | More accurate than PQL | Slower and less flexible than PQL | PROC GLIMMIX [56], glmer (R), |
| | | | glmm.admb (R), AD Model Builder, |
| | | | HLM |
| Gauss-Hermite quadrature | More accurate than Laplace | Slower than Laplace; limited to | PROC GLIMMIX [56], PROC |
| | | 2–3 random effects | NLMIXED (SAS), glmer (R), glmmML |
| | | | (R) |
| Markov chain Monte Carlo | Highly flexible, arbitrary number | Very slow, technically challenging, | WinBUGS, JAGS, MCMCpack, (R), |
| | of random effects; accurate | Bayesian framework | AD Model Builder |

MCMCglmm (R)

Making inferences for mixed models

Options

Parametric Bootstrap (Monte Carlo Simulations) – available in Imer(), maybe in others?

Non-Parametric Bootstrapping (must carefully consider sampling design, within and between random effects. In particular you war to keep the numbers of levels of your random effect the same as with the observed data. However you may also want to sample within groups as well... Depends on the data).

MCMC (options available in MCMCglmm, glmmADMB and to some extent lmer via mcmcsamp, and arm).

Decision tree

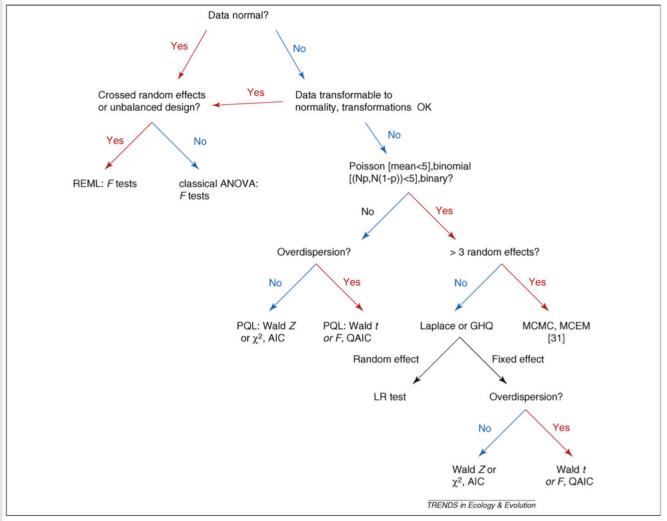


Figure 1. Decision tree for GLMM fitting and inference. Conditions on the Poisson and binomial distributions along the right branch refer to penalized quasilikelihood (PQL) rules of thumb [30]: to use PQL, Poisson distributions should have mean > 5 and binomial distributions should have the minimum of the number of successes and failures > 5. MCEM = Monte Carlo expectation-maximization [40].