Introduction

Falling costs of genetic sequencing have allowed sequencing and annotation of the genomes of non-model organism. In annotating nonmodel genomes, mRNA-seq data has great potential to improve annotation quality. For example the Asian sea bass (L. calcarifer) genome annotation effort drew on previously assembled mRNA-seq data provided by the Temasek Life Sciences Laboratory (TLL). At the South African National Bioinformatics Institute (SANBI) we undertook gene annotation on the Asian seabass genome using a pipeline built out of custom scripts. Simultaneously, a team at Saint Petersburg State University undertook the same task using MAKER2. Comparing the results of this annotation highlighted the impact of tool and parameter choice in gene prediction.

The Galaxy¹ framework allows workflows to be constructed in a high level workflow that hides the systemlanguage specific details their implementation. We Due to the share amount of sequences to align, a two step approach for qualifying sequences was applied. Thi implemented genome annotation workflows in Galaxy, blasta -pg tblastn -q fish.\$SGE_TASK_ID.fa -t \$DBASE/seabass.masked.fa -p T -o \$OUTdir/fis demonstrating suitability for constructing an annotation The disconnect between workflow (in flowcharts) description workflow implementation languages) scripting hampers adaption and reproduction of results. Training of Lates calcarifer (seabass AUGUSTUS gene prediction **TEMASEK**

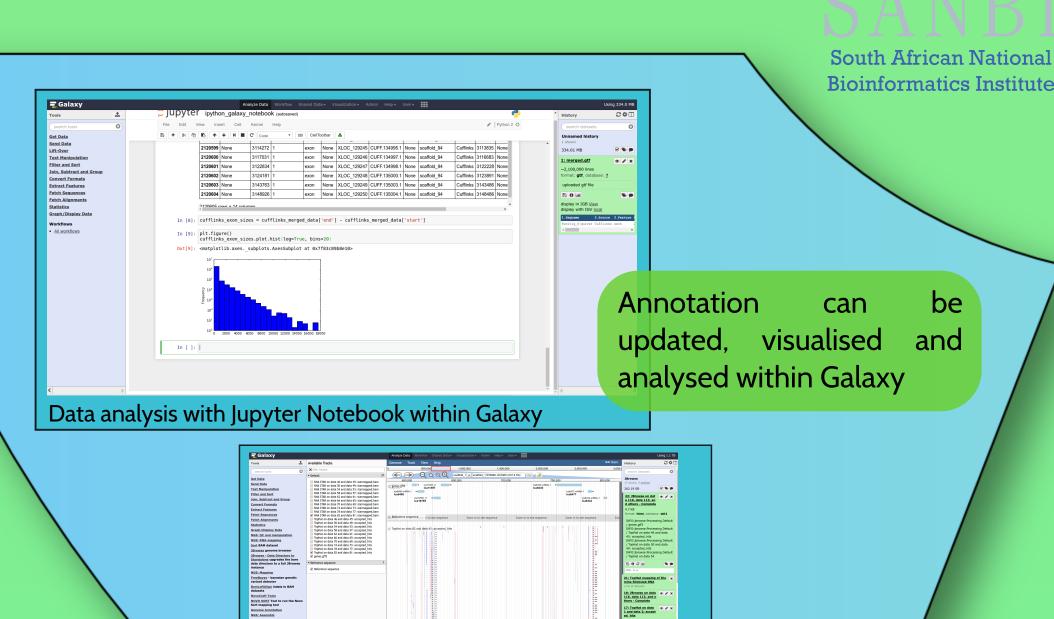
An Extensible Genome Annotation Workbench based on the Galaxy Platform

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JBrowse Genome browser within Galaxy

Asian Seabass Genome Project

SANBI ON A Rainer National Section of the Control o

Bass Explorer database browser

Conclusion

- Repeatable genome analysis workflows allow reuse of methods and reproducibility of results
- Galaxy workflows allow workflow construction in a flowchart-like idiom similar to the way in which workflows are documented
- We demonstrate the construction of two workflows as part of a larger annotation workbench and their use in the annotation of the L. calcarifer genome
- Exposing results through Jupyter notebooks and export to browsers such as JBrowse and the (SANBI-authored) Bass Explorer allows results to be examined seamlessly within Galaxy

Future work

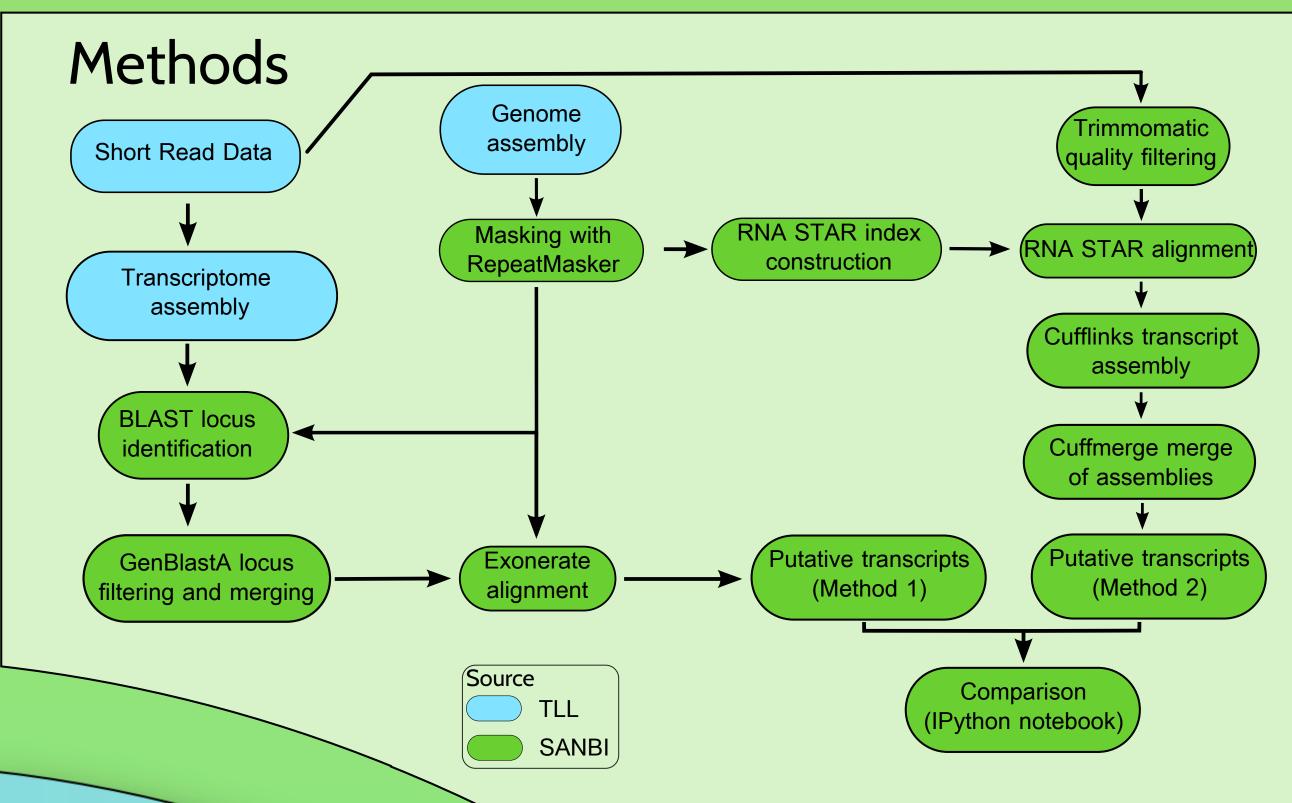
- We intend to implement further workflows to provide a complete Galaxy-based eukaryotic genome annotation workbench
- We will enhance tools that export to an annotation database browser modelled on the Bass **Explorer**

Bibliography and Acknowledgements

1. The Galaxy Team. The Galaxy Project: Online bioinformatics analysis for everyone. [cited 2016 Mar 29]. Available from: https://www.galaxyproject.org/

2. Thevasagayam NM, et al. Transcriptome Survey of a Marine Food Fish: Asian Seabass (Lates calcarifer). J Mar Sci Eng. 2015 Jun 2;3(2):382-400.

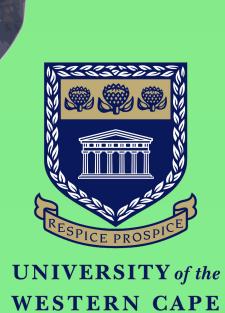
Special thanks to the Galaxy Intergalactic Utilities Commission (IUC) members, especially Eric Rasche (for JBrowse wrapper) and Björn Grüning (for Interactive Environments) and SANBI software developers Thoba Lose (for Bass Explorer) and Ziphozakhe Mashologu (for RNA STAR index builder). This work was supported by the South African Research Chairs Initiative of the Department of Science and Technology and National Research Foundation of SA.



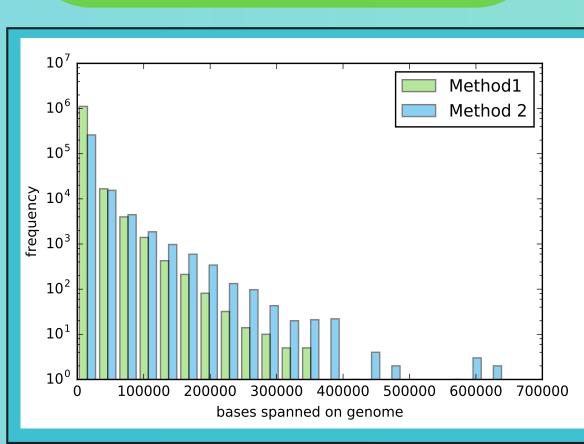
Four datasets were obtained from Temasek Life Sciences Laboratory: assembled² transcript data, comprising 1,184,879 contigs totalling 890 assembled L. calcarifer genome (668 Mb, N50 1,191,366, in press) and two sets of Illumina HiSeq RNAseq reads (486 million reads after filtering for quality length). Analysis conducted using Galaxy workflows, **IPython** notebook custom scripts.

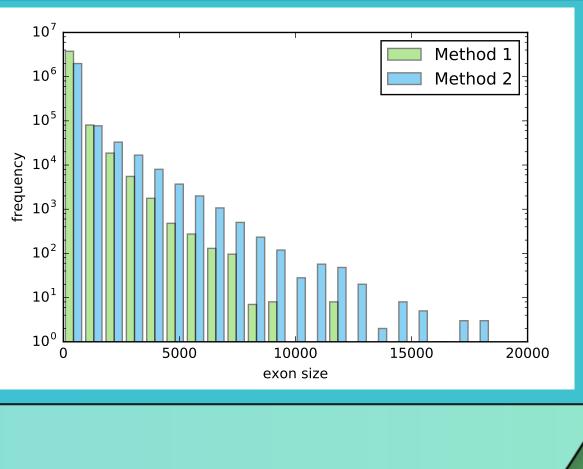
Input FASTA files(s Optional ASN.1 file(s) containing □Nucleotide query sequence Nucleotide BLAST database output1 (tabular, txt, html, blastxn 🔑 Split Fasta 🗶 Portion of Method 1 workflow

workflow annotation implemented in a idiom close to that in which workflows are described. Cufflinks ☐SAM or BAM file fastq_out_paired ☐RNA-Seq FASTQ/FASTA file Global model (for us output_log (txt) Mask File chimeric_junctions (interval) chimeric_reads (bam)🗀 splice_junctions (interval) Portion of Method 2 workflow



the faster method, generates some artificially long isoforms but 96% of exons predicted overlap with those predicted by Method 1.





Results and Discussion

Method	Max Parallelism	Run time	
1. Align assembled transcripts	40 way data split	12 days	
2. Map reads & assemble transcripts	14 way data split	5 days	

The tools in the two transcript-alignment and reconstruction workflows make use of different degrees of multithreading, but in both workflows extra parallelism was achieved by using Galaxy dataset collections. Even with parallelisation the BLAST, GenBlastA and Exonerate steps of the workflow, the RNA STAR and Cufflinks based Method 2 was more than twice as fast as Method 1.

The assembled transcripts used in Method 1 were assembled independently in different libraries, resulting in overlapping transcript alignments and a substantially higher number of aligned transcripts compared to Method 2, which merged transcript alignments from multiple libraries into final transcripts. Method2 predicted significantly (according to Mann-Whitney test) larger exons and on-genome isoforms, with 460 spanning more than 200Kb. We suspect that these suspiciously large isoforms are artifacts possibly caused by matches to unmasked repeats. Despite these suspicious artifacts, 2,040,178 out of 2,120,605 (96%) exons predicted by Method 2 overlapped with exons predicted by Method 1, suggesting that Method 2 is worth further investigation.

Method	No. of transcripts	No. of exons	Exons per transcript	Exon size mean / stddev	Isoform size mean / stddev	
1. Align assembled transcripts	1,137,181	3,853,944	3.4	210 / 291	3718 / 9624	
2. Map reads & assemble transcripts	284,104	2,120,605	7.5	295 / 587	11308 / 21862	