

# **Lab6: Imaging tools (Roboflow)**

3099704 AI for Digital Health (2025/2)

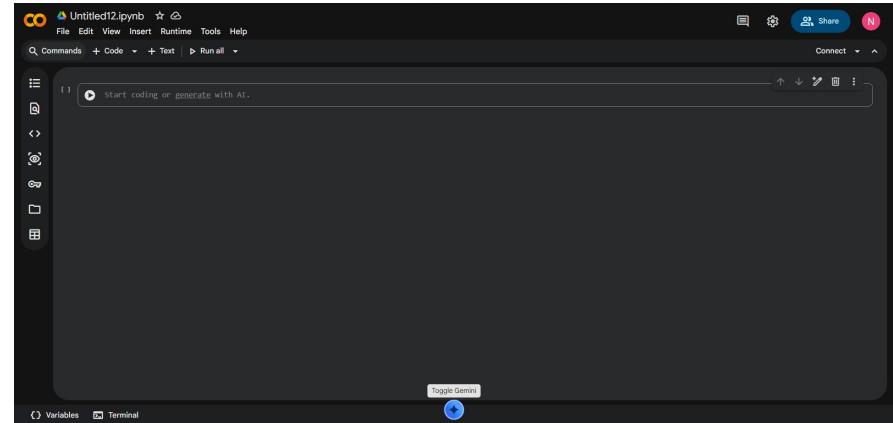
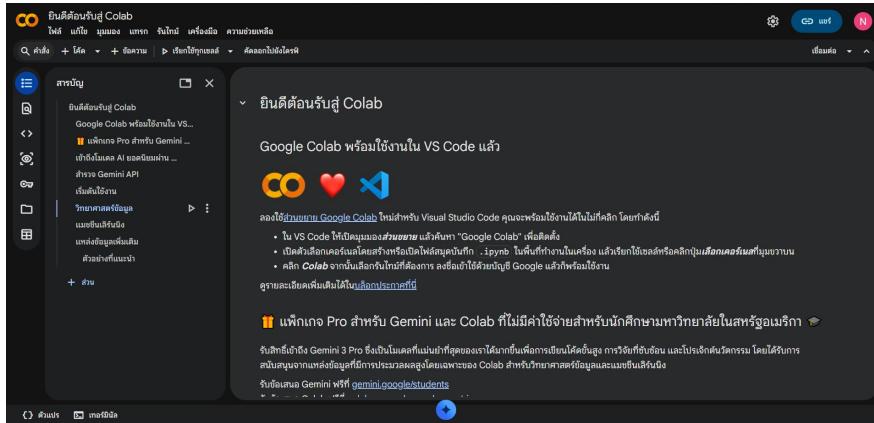
# Objective

- Use the **Roboflow** to create Dataset
- Use created Dataset to train model  
(YOLO,UNet)



# Material

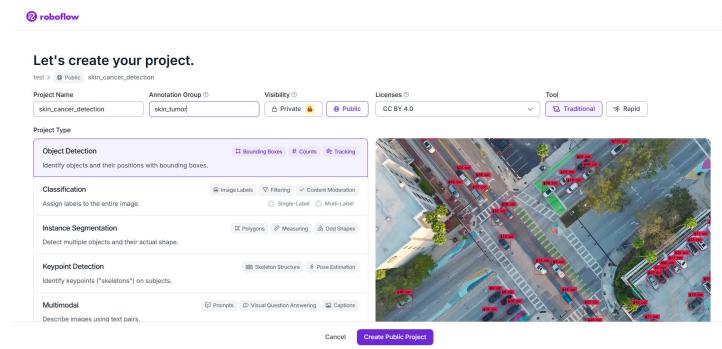
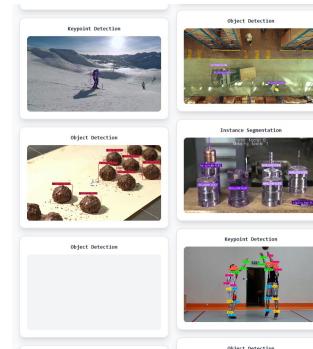
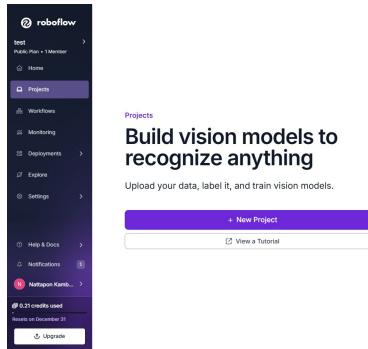
- With **Google Colab**, you don't need to install any software. All you need is a Google account, and you can start using it right away. Simply visit: <https://colab.research.google.com/> or select NEW NOTEBOOK to start a new file.



# Roboflow

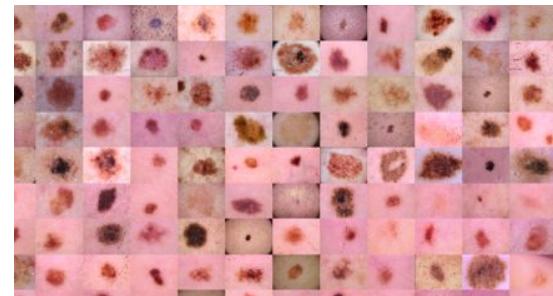
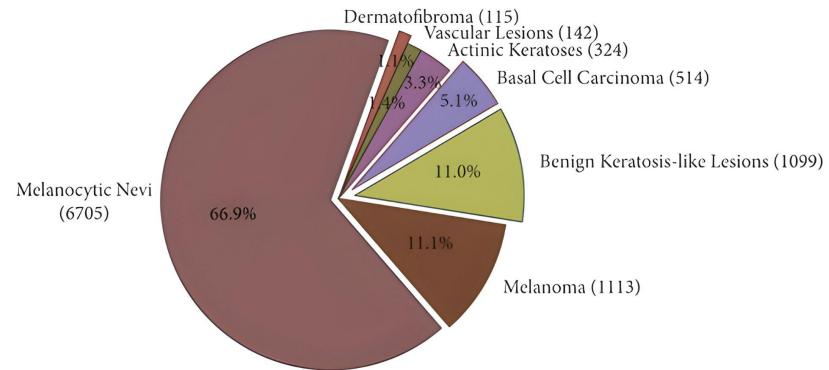


- **Roboflow** is a web-based platform that helps users easily prepare datasets. It can be accessed through a web browser at <https://roboflow.com/> by signing in with a Google or GitHub account.
- Users can upload images, annotate data, apply preprocessing and data augmentation, and export datasets in formats ready for training models such as YOLO.



# Dataset: Skin Cancer MNIST (HAM10000)

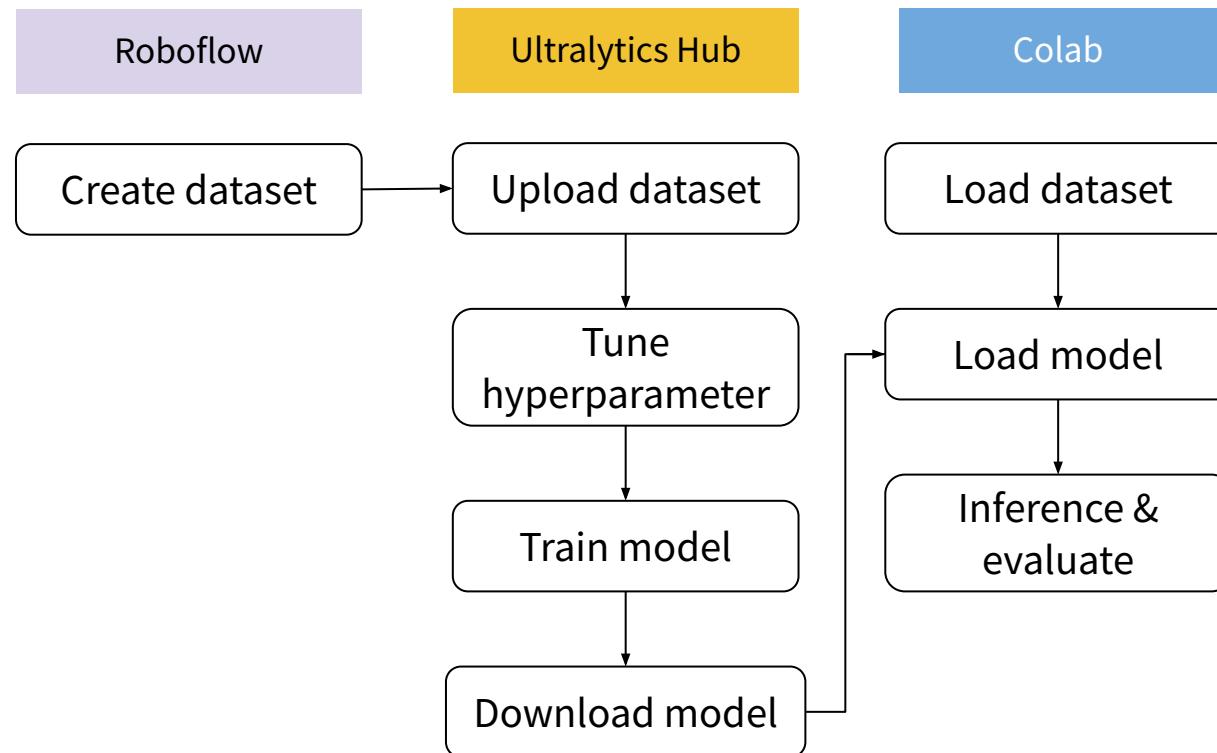
- The dataset consists of 10015 images with 10013 labeled objects belonging to 7 skin cancer classes.
- The data contains image in JPG format and documents in JSON format



# Lab6.1: Object detection dataset

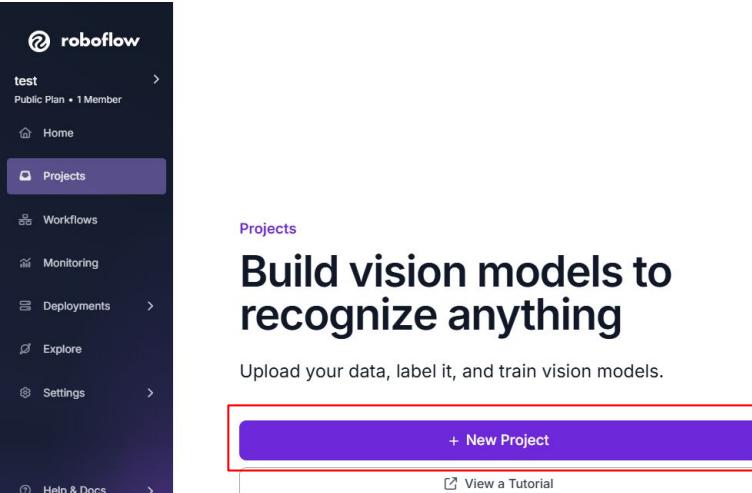
In this lab, we will repeat the experiment from **Lab 4.1 (YOLOv8n)**, but the dataset will be created using Roboflow instead.

- 1) Load image from github
- 2) Create Object detection dataset in Roboflow
- 3) Train YOLO model in ultralytics hub
- 4) evaluate YOLO in [Lab 6.1 ultralyricshub](#)

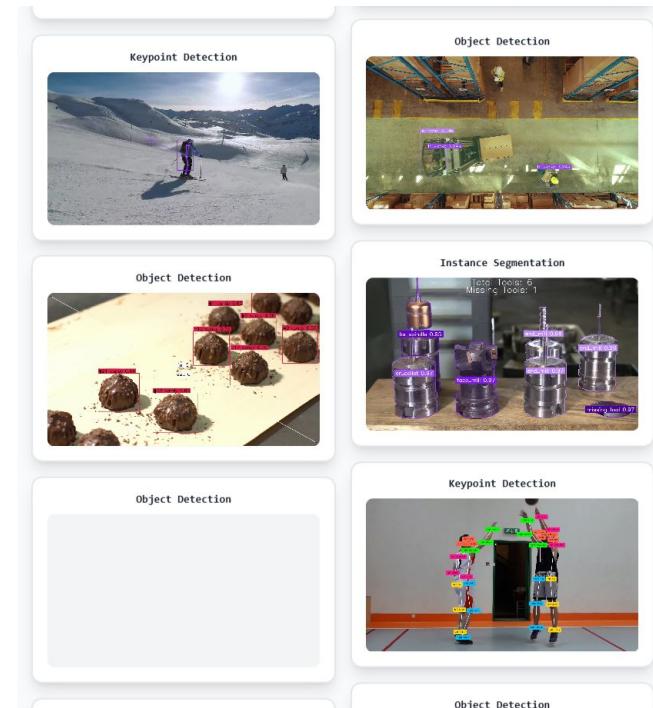


# Lab6.1: Object detection dataset

## 1) Sign in [Roboflow](#)



The image shows the Roboflow web interface. On the left is a dark sidebar with various navigation links: Home, Projects (which is selected and highlighted in purple), Workflows, Monitoring, Deployments, Explore, Settings, Help & Docs, Notifications (with a '1' notification badge), and a user profile for Nattapon Kamb... Below the sidebar, it says '0.21 credits used' and 'Resets on December 31'. At the bottom is a 'Upgrade' button. The main area has a heading 'Build vision models to recognize anything' and a sub-instruction 'Upload your data, label it, and train vision models.' A prominent purple button labeled '+ New Project' is centered at the bottom of this section, with a red rectangular box drawn around it to indicate it as the target for step 1.



# Lab6.1: Object detection dataset

## 2) Create new project (**Object Detection**)

roboflow

Let's create your project.

test > Public skin\_cancer\_detection

Project Name: skin\_cancer\_detection  
Annotation Group: skin\_tumor  
Visibility: Private (Public selected)  
Licenses: CC BY 4.0  
Tool: Traditional (Rapid selected)

Project Type:

- Object Detection**: Identify objects and their positions with bounding boxes.
- Classification**: Assign labels to the entire image.
- Instance Segmentation**: Detect multiple objects and their actual shape.
- Keypoint Detection**: Identify keypoints ("skeletons") on subjects.
- Multimodal**: Describe images using text pairs.

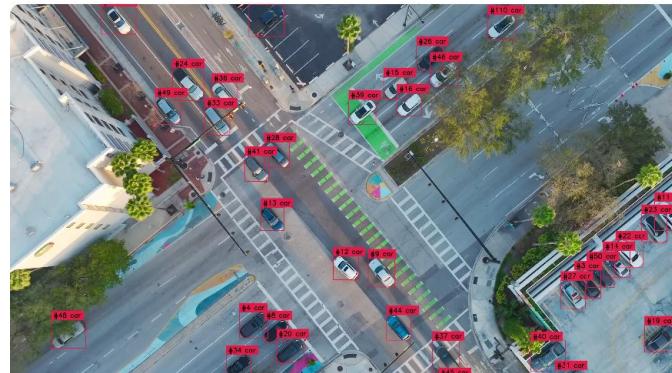
Bounding Boxes # Counts Tracking

Image Labels Filtering Content Moderation Single-Label Multi-Label

Polygons Measuring Odd Shapes

Skeleton Structure Pose Estimation

Captions



Cancel Create Public Project

# Lab6.1: Object detection dataset

- 3) Download the dataset from [GitHub](#), then upload in **Roboflow**.

The screenshot shows the Roboflow web interface. On the left, a sidebar navigation bar includes icons for user profile, dashboard, datasets, annotations, versions, analytics, classes & tags, models, visualizations, deployments, and a search bar. The main area displays a project titled "skin\_cancer\_d..." under "Object Detection". The central part of the screen is the "Upload" page. It features a large input field with a circular "Upload" button containing an upward arrow. Below this is the text "Drag and drop file(s) to upload, or:". Underneath are two buttons: "Select File(s)" and "Select Folder". A section titled "Supported Formats" lists "Images" (formats: .jpg, .png, .bmp, .webp, .avif), "Annotations" (formats: in 26 formats), "Videos" (formats: .mov, .mp4), and "PDFs". At the bottom of this section is a note: "\*Max size of 20MB and 16,400 x 16,900 pixels.". To the right of the main upload area, there are three informational boxes: 1) "Need images to get started?" with a "Upload data from your phone" section and a QR code; 2) "Search on Roboflow Universe" with a search bar; 3) "Bulk Upload Images" with a description of how to upload 1k+ images using Python SDK, REST API, and CLI.

# Lab6.1: Object detection dataset

- 3) Download the dataset from [GitHub](#), then upload in **Roboflow**.

The screenshot shows the Roboflow web interface. On the left is a sidebar with navigation links: TEST, skin\_cancer\_d..., DATA (with Upload Data highlighted), Annotate, Dataset, Versions (Train selected), Analytics, Classes & Tags, MODELS, Models, Visualize, DEPLOY, Deployments, and a circular icon with 'N'. The main area is titled 'Upload' under 'TEST'. It shows a 'Batch Name' field containing 'skin\_cancer\_d...' and a timestamp 'Uploaded on 12/26/25 at 1:18 pm'. There is a 'Tags' field with a placeholder 'Search or add tags for images...'. A checkbox for 'Create batch instantly' is unchecked. Below these are buttons for 'All Images' (16), 'Annotated' (0), and 'Not Annotated' (16). A large central area is labeled 'Drag and drop images, annotations, and videos.' and specifies supported formats: '.jpg, .png, .bmp, .webp, .avif' (in 26 formats) and '.mov, .mp4'. It also notes a max size of 200B and 16,400 x 10,900 pixels. At the bottom of this area are 'Select Files' and 'Select Folder' buttons, and a prominent purple 'Save and Continue →' button. Below this is a grid of 16 small images of medical procedures, each with a unique file name. At the bottom is a section titled 'Want to add similar images?' with a sub-section 'Powered by Objects365' and a row of nine thumbnail images of surgical scenes.

# Lab6.1: Object detection dataset

## 4) Check “Label Myself”

The screenshot shows the Roboflow interface for a dataset named "TEST". On the left, there's a sidebar with various navigation options like "Object Detection", "Upload Data", "Annotate", "Dataset", "Versions", "Train", "Analytics", "Classes & Tags", "Models", "Visualize", "Deploy", and "Deployments". The main area displays a batch of images uploaded on 12/26/25 at 1:18 pm. A red box highlights the "Label Myself" option under the heading "How do you want to label your images?", which includes a sub-section for "Label With My Team" and "Hire Outsourced Labelers".

TEST

Annotate > Batch  
Uploaded on 12/26/25 at 1:18 pm  
Uploaded Dec 26, 2025 (1:18 PM)

Upload More Rename

skin\_cancer\_d... Object Detection

DATA

Upload Data Annotate

Dataset Versions Train

Analytics

Classes & Tags

MODELS

Models Visualize

DEPLOY

Deployments

How do you want to label your images?

Auto-Label Entire Batch Use your own custom model or a zero-shot model to automatically label your entire batch.  
Try with SAM3

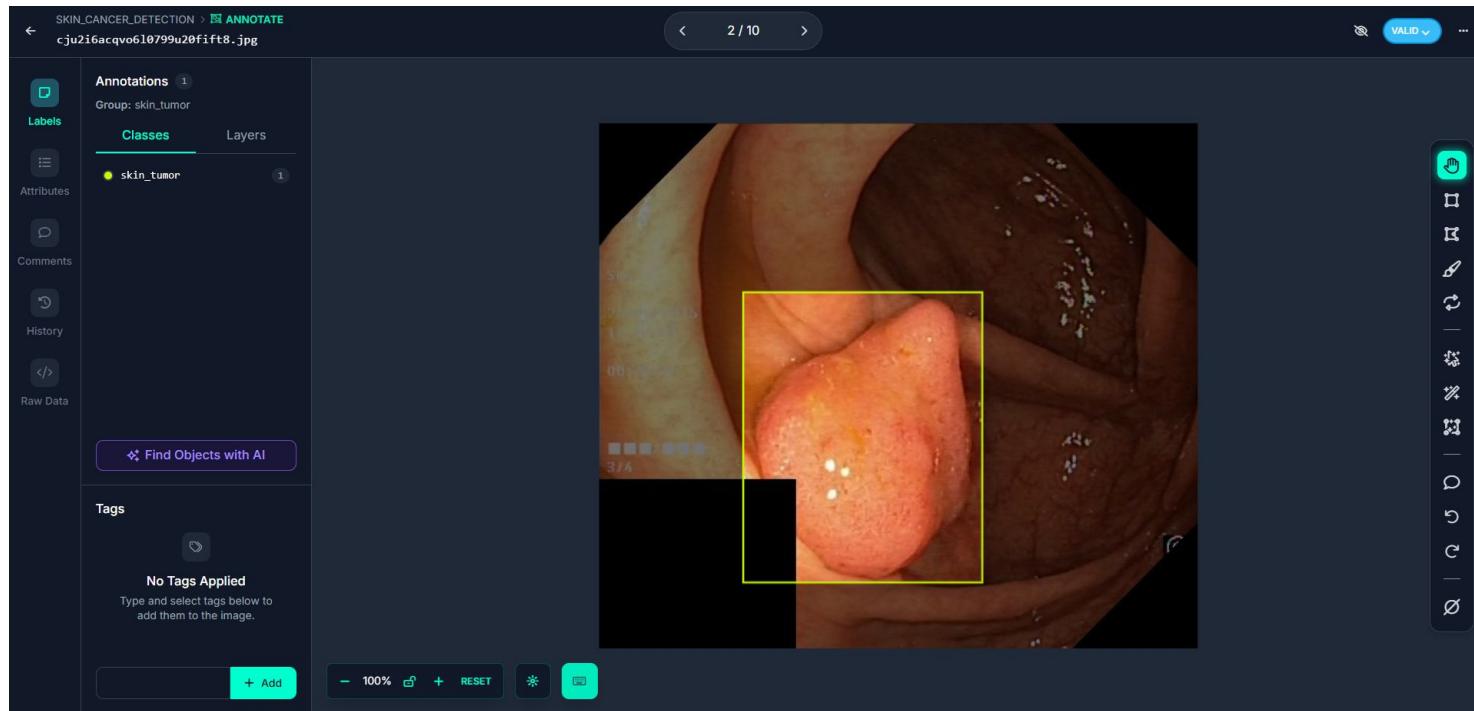
**Label Myself** Label images with our AI labeling tools.

Label With My Team Split up the labeling work across your team.

Hire Outsourced Labelers Work with a professional labeling team vetted by Roboflow. Upgrade

# Lab6.1: Object detection dataset

## 5) Label polyp



# Lab6.1: Object detection dataset

- 6) Split Train:60%, Validation:20%, Test:20%

The screenshot shows a user interface for managing datasets, specifically for an 'Object Detection' task. On the left, a sidebar provides navigation links for TEST, DATA, MODELS, and DEPLOY. The main area displays a preview of an image from the 'skin\_cancer\_d...' dataset, which is currently set to 'TEST'. A 'Versions' tab is selected, showing a list of existing versions and a form to create a new one.

**Versions**

Version Name: 2025-12-26 1:21pm

**Source Images**  
Images: 10  
Classes: 1  
Unannotated: 0

**Train/Test Split** (highlighted with a red box)  
Training Set: 6 images  
Validation Set: 2 images  
Testing Set: 2 images

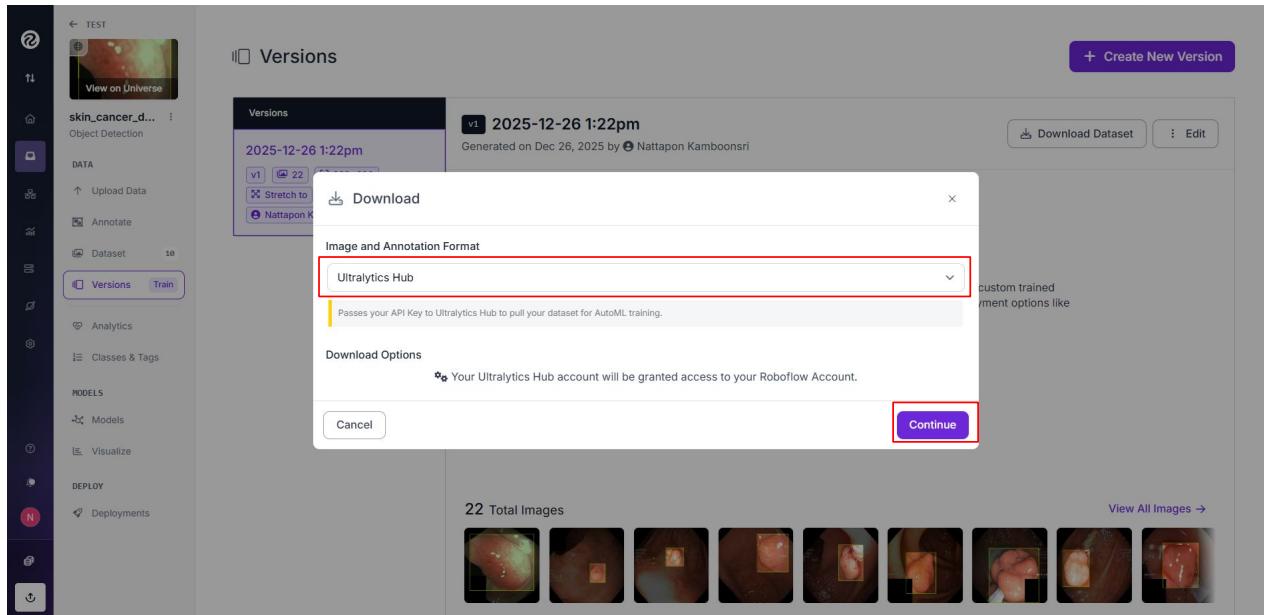
**Preprocessing**  
Auto-Orient: Applied  
Resize: Stretch to 320x320

**Augmentation**  
Flip: Horizontal, Vertical  
Noise: Up to 0.1% of pixels

**Create** (highlighted with a red box)  
Review your selections and select a version size to create a moment-in-time snapshot of your dataset with the applied transformations.

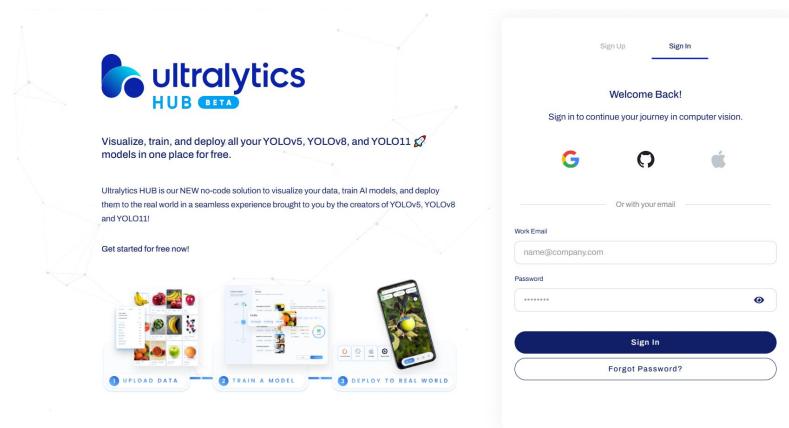
# Lab6.1: Object detection dataset

- 7) Download the dataset in two formats: (1) Ultralytics HUB format for training the model, and (2) YOLOv8 format for evaluation using a Google Colab notebook.

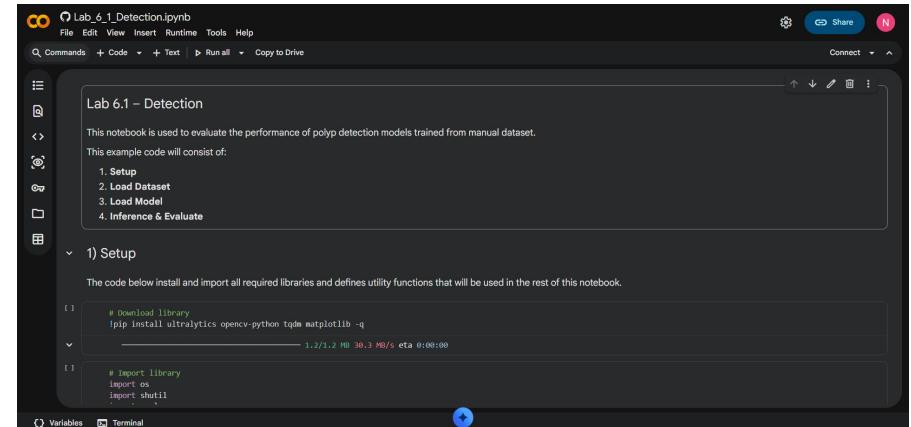


# Lab6.1: Object detection dataset

- 8) Train YOLOv8n in Ultralytics Hub
- 9) Load model to run in [Lab\\_6\\_1\\_Detection](#) (in **colab**)



The image shows two screenshots of the Ultralytics Hub interface. On the left is the homepage, featuring the Ultralytics logo, a 'HUB BETA' badge, and a brief description: 'Visualize, train, and deploy all your YOLOv5, YOLOv8, and YOLOv11 models in one place for free.' It includes sections for 'Upload Data', 'Train a Model', and 'Deploy to Real World'. On the right is the sign-in screen, which has 'Sign Up' and 'Sign In' buttons at the top. Below is a 'Welcome Back!' message and a 'Sign in to continue your journey in computer vision.' section. It features social login buttons for Google, GitHub, and Apple, and fields for 'Work Email' and 'Password'.



The image shows a Google Colaboratory (Colab) notebook titled 'Lab\_6\_1\_Detection.ipynb'. The title bar includes 'File Edit View Insert Runtime Tools Help' and 'Share' and 'Connect' buttons. The main area contains the following text and code:

Lab 6.1 – Detection  
This notebook is used to evaluate the performance of polyp detection models trained from manual dataset.  
This example code will consist of:

1. Setup
2. Load Dataset
3. Load Model
4. Inference & Evaluate

**1) Setup**  
The code below install and import all required libraries and defines utility functions that will be used in the rest of this notebook.

```
# Download library  
!pip install ultralytics opencv-python tqdm matplotlib -q
```

```
# Import library  
import os  
import shutil
```

At the bottom, there are 'Variables' and 'Terminal' tabs.

# Lab6.1: Object detection dataset

5) Open [Lab\\_4\\_1\\_ultralyricshub](#) (in **colab**)

The screenshot shows a Google Colab interface with the following details:

- Title:** Lab\_4\_1\_ultralyricshub.ipynb
- Toolbar:** File, Edit, View, Insert, Runtime, Tools, Help.
- Search Bar:** Commands, + Code, + Text, Run all, Copy to Drive.
- Share Button:** Share, Connect.
- Notebook Content:**
  - Section:** Lab 4.1 – Polyp detection: YOLO8 (Ultralytics hub)
  - Description:** This notebook is used to evaluate the performance of polyp detection models trained on Ultralytics hub.
  - Text:** This example code will consist of:
    1. Setup
    2. Load Dataset
    3. Load Model
    4. Inference & Evaluate
  - Section:** 1) Setup (highlighted with a blue border)
  - Description:** The code below install and import all required libraries and defines utility functions that will be used in the rest of this notebook.
  - Code:**

```
# Download library
!pip install ultralytics opencv-python tqdm matplotlib -q
```

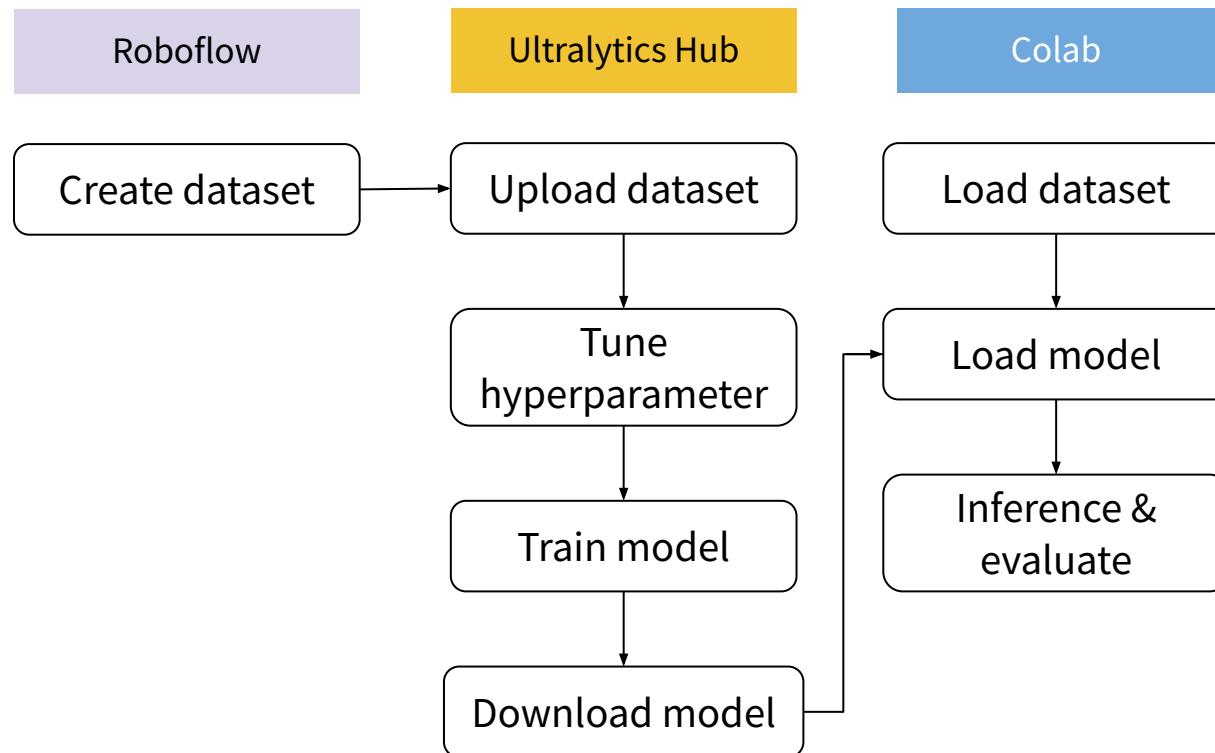
Output: 1.2/1.2 MB 30.3 MB/s eta 0:00:00

```
# Import library
import os
import shutil
```

# Lab6.1: Object detection dataset

In this lab, we will repeat the experiment from **Lab 4.1 (YOLOv8n)**, but the dataset will be created using Roboflow instead.

- 1) Load image from github
- 2) Create Object detection dataset in Roboflow
- 3) Train YOLO model in ultralytics hub
- 4) evaluate YOLO in [Lab 6.1 ultralyricshub](#)



# Lab6.2: Segmentation dataset

Roboflow

Colab

In this lab, we will repeat the experiment from **Lab 5.1 (2DUNet)**, but the dataset will be created using Roboflow instead.

- 1) Load image from [GitHub](#)
- 2) Create segmentation dataset in Roboflow
- 3) Download the dataset in segment formats
- 4) Upload the dataset to the [Lab 6.2 Segmentation](#), then train and evaluate the model.

