

Trapnell Data Set - Differential Expression (cuffdiff)

BIOL550 - Lab 3 Weekly Report (Week 3)

What I accomplished since the previous report

I produced a set of STAR BAM files that include the `xs` strand tag required by `cuffdiff`, and then ran `cuffdiff` on all six samples (3 C1 replicates and 3 C2 replicates) to complete the Trapnell differential expression step. I exported the `cuffdiff` output directory locally, summarized gene-level results from `gene_exp.diff` using `q_value <= 0.05`, and generated two sanity-check figures (volcano plot + top-DE bar chart).

Results summary

```
Lab 3: Cuffdiff gene-level DE summary
OK tests: 8289
Significant genes (q<=0.05): 265
Top up (by log2FC): Fatp, crc, scf, CTPsyn, Df31
Top down (by log2FC): Nep2, RpS19b, Amy-d, CG6847, Aplip1
```

Methods used (commands + parameters)

STAR was used to generate sorted coordinate BAMs, and `cuffdiff` was used for differential expression on the aligned reads. Then, downstream filtering and plotting were done locally from the `cuffdiff` output tables.

STAR re-alignment (to ensure xs tags)

STAR was re-run for all six samples with `--outSAMstrandField intronMotif` so that spliced alignments include `xs` tags.

```
STAR \
--genomeDir /home/pzg8794/star_index/classref_trapnell_zip_bdgp6_84_v2 \
--runThreadN 4 \
--sjdbGTFfile "/home/pzg8794/BIOL550/Lab1/Trapnell_Data/Trapnell Data/Drosophila reference/Drosophila_melanogaster.BDGP6.84.gtf" \
\
--readFilesIn \
"/home/pzg8794/BIOL550/Lab1/Trapnell_Data/Trapnell Data/Raw reads/GSM794483_C1_R1_1.fq.gz" \
"/home/pzg8794/BIOL550/Lab1/Trapnell_Data/Trapnell Data/Raw reads/GSM794483_C1_R1_2.fq.gz" \
--readFilesCommand zcat \
--outSAMtype BAM SortedByCoordinate \
--outSAMstrandField intronMotif \
--limitBAMsortRAM 600000000 \
--outFileNamePrefix /home/pzg8794/BIOL550/Lab1/star_align_classref_v2_all_xs/GSM794483_C1_R1/
```

Differential expression (`cuffdiff`)

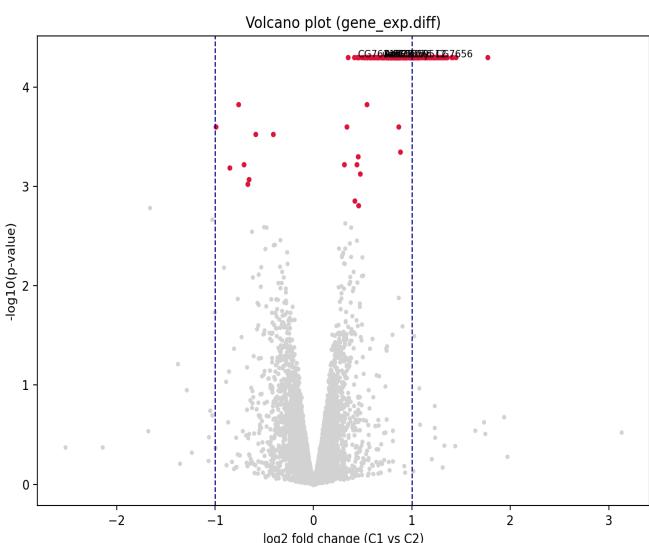
The Drosophila reference directory includes both a GTF and a GFF3; the GTF (`Drosophila_melanogaster.BDGP6.84.gtf`) was used because `cuffdiff` expects GTF2 annotation (GFF3 would require conversion before use). `Cuffdiff` was run on the XS-tagged BAMs (3 replicates per condition) with bias correction enabled via `-b`.

```
/usr/local/bin/cufflinks/cuffdiff \
-o /home/pzg8794/BIOL550/Lab1/cuffdiff_classref_v2_xs \
-p 4 \
-L C1,C2 \
-b /home/pzg8794/refs/classref_bdgp6_84_ids_v2.fa \
"/home/pzg8794/BIOL550/Lab1/Trapnell_Data/Trapnell Data/Drosophila reference/Drosophila_melanogaster.BDGP6.84.gtf" \
\
/home/pzg8794/BIOL550/Lab1/star_align_classref_v2_all_xs/GSM794483_C1_R1/Aligned.sortedByCoord.out.bam,/home/pzg8794/BIOL550/Lab1/star_align_classref_v2_all_xs/GSM794484_C1_R2/Aligned.sortedByCoord.out.bam,/home/pzg8794/BIOL550/Lab1/star_align_classref_v2_all_xs/GSM794485_C1_R3/Aligned.sortedByCoord.out.bam \
\
/home/pzg8794/BIOL550/Lab1/star_align_classref_v2_all_xs/GSM794486_C2_R1/Aligned.sortedByCoord.out.bam,/home/pzg8794/BIOL550/Lab1/star_align_classref_v2_all_xs/GSM794487_C2_R2/Aligned.sortedByCoord.out.bam,/home/pzg8794/BIOL550/Lab1/star_align_classref_v2_all_xs/GSM794488_C2_R3/Aligned.sortedByCoord.out.bam
```

Downstream summary (local)

From the file `gene_exp.diff`, I filtered valid tests by keeping rows where `status == "OK"`, and defined significance at `q_value <= 0.05`. Then, I generated a volcano plot (log2FC vs. $-\log_{10}$ p-value) and a top-gene bar chart (ranked by log2FC) as sanity checks on effect-size distribution and signal presence.

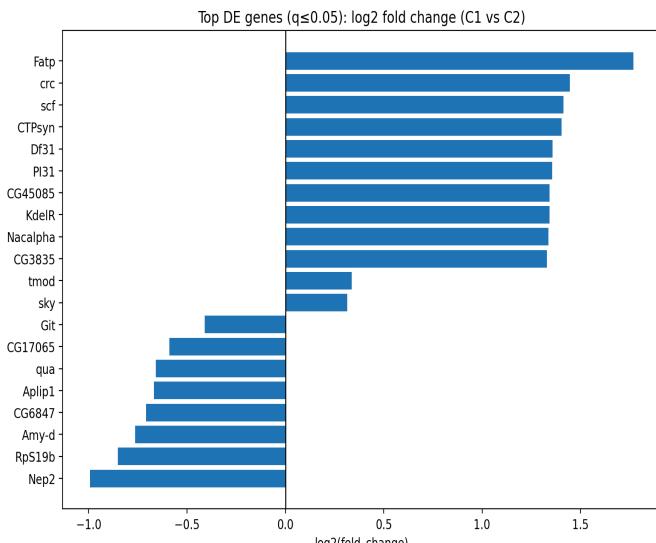
Volcano plot



Each gene is a point: $x = \log_2$ fold-change (effect size), $y = -\log_{10}(p)$ (significance score), so “outliers” pop into corners.

Volcano plot: gene_exp.diff

Top 20 DE genes bar chart



Ranking genes by signed log₂FC while showing the biggest movers up vs. down as bars.

Top DE genes (by log2FC)

Problems encountered

The first issue was annotation format: the reference folder includes both a GTF and a GFF3 file. The GFF3 file uses `ID=/Parent=` attributes, whereas the GTF uses the `gene_id/transcript_id` fields. After inspecting both files, I used the GTF (`Drosophila_melanogaster.BDGP6.84.gtf`) because it was the most compatible with STAR and `cuffdiff` with minimal changes. The second issue was that the initial STAR BAMs from Lab 2 did not contain `xs` strand tags on spliced reads, which caused `cuffdiff` to fail. This required going back and re-aligning all six samples with `--outSAMstrandField intronMotif` to produce XS-tagged BAMs.

Goals for the coming week

Next week I will (1) further explore and improve the visualization workflow (e.g., volcano and MA-style plots) to better interpret the `cuffdiff` outputs, (2) run the FASTQ “dump” workflow for our zebrafish project data using the SRA Toolkit (`prefetch + fasterq-dump`) and validate our wrapper script for downloading an arbitrary number of runs, and (3) run `FastQC` on the downloaded FASTQs and begin reviewing the QC reports to guide downstream analysis.