

# *CONNECTOR<sup>n</sup>* User Manual

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**Abstract**

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# 1 Data importing by files

The analysis starts from two distinct files:

- A file reporting the discretely sampled curve data. The longitudinal data must be reported in terms of time  $t$  and  $y$ -values  $y$ . Each sample is described by four columns: the first column, named `subjID`, contains a list of subject IDs. The second one, named `value`, contains a list of  $y$  values. The third one, named `time`, includes the lags. The last one, named `measureID`, contains a list of the corresponding measure names. Order is not mandatory. Accepted formats are: Excel, CSV, or `tbl_df`. See Figure 1.
- A file containing the annotations associated with the sampled curves. The first column reports the `subjID`, while the other columns report the features relevant for the analysis, one per column. Note that the `subjID` sample must contain the same ID names that appear in the file of the sampled curves. Accepted formats: Excel, CSV, or `tbl_df`. See Figure 2.

time	value	measureID	subjID
-10.0	207.416776	Parabola	13
-10.0	274.225908	Hyperbola	26
-10.0	28.585194	Sine	3
-10.0	-28.585194	Sine	10
-10.0	28.585194	Sine	15
-10.0	28.585194	Sine	25
-10.0	-28.585194	Sine	26
-10.0	-98.167400	Cosine	13
-10.0	98.167400	Cosine	14
-9.9	-72.514435	Parabola	18
-9.9	249.594127	Hyperbola	16
-9.9	224.939112	Hyperbola	28
-9.9	-23.661854	Sine	16

Figure 1: Example for TimeSeriesFile

subjID	gender	age	treatment_group	baseline_weight	height	comorbidity
1	M	23	Treatment A	74.1	184.1	Diabetes
2	F	77	Treatment B	54.3	169.0	None
3	M	38	Control	81.0	150.1	None
4	M	50	Control	69.0	161.3	None
5	F	47	Treatment A	57.9	183.1	Obesity
6	M	71	Treatment B	56.4	191.9	None
7	F	58	Treatment B	51.4	166.9	Diabetes
8	M	65	Treatment A	99.4	181.5	None
9	M	44	Control	94.4	187.8	Hypertension
10	F	65	Control	90.5	161.5	None

Figure 2: Example for AnnotationFile

## 2 Import

The two files are imported by the **DataImport** function, whose arguments are the file names. In this example, *TimeSeriesFile* and *AnnotationFile*.

```

1 Data<-DataImport(TimeSeries , Annotations)
2 #####
3 Data loaded...
4 Number of curves:120
5 Number of distinct measures:4
6 Average length:14.91667
7 Measure with highest length:# A tibble: 1 x 2
8   curvesID      nTimePoints
9   <chr>          <int>
10 1 26_Hyperbola      24
11
12 Measure with lowest length:# A tibble: 2 x 2
13   curvesID      nTimePoints
14   <chr>          <int>
15 1 23_Hyperbola       9
16 2 9_Cosine           9

```

A CONNECTORData object is created:

```

1 > str(Data)
2 Formal class 'CONNECTORData' [package ".GlobalEnv"] with 4 slots
3 ..@ curves      : tibble [1,790 x 5] (S3: tbl_df/tbl/data.frame)
4 .. ..$ subjID   : Factor w/ 30 levels "1","2","3","4",...: 1 1 1 1
5   1 1 1 1 1 1 ...
6 .. ..$ measureID: chr [1:1790] "Parabola" "Parabola" "Parabola" "
7   Parabola" ...
8 .. ..$ time      : num [1:1790] -6.7 -5.8 -5 -3.6 -1.3 ...
9 .. ..$ curvesID  : chr [1:1790] "1_Parabola" "1_Parabola" "1_
10  Parabola" "1_Parabola" ...
11 .. ..$ value     : num [1:1790] 160.4 130.2 66.1 41.9 30.4 ...
12 ..@ dimension   : tibble [120 x 2] (S3: tbl_df/tbl/data.frame)
13 .. ..$ curvesID  : chr [1:120] "10_Cosine" "10_Hyperbola" "10_
14  Parabola" "10_Sine" ...
15 .. ..$ nTimePoints: int [1:120] 13 14 10 14 14 17 17 13 11 10 ...
16 ..@ annotations: tibble [30 x 7] (S3: tbl_df/tbl/data.frame)
17 .. ..$ subjID    : Factor w/ 30 levels "1","2","3","4",...: 1
18   2 3 4 5 6 7 8 9 10 ...
19 .. ..$ gender    : chr [1:30] "M" "F" "M" "M" ...
20 .. ..$ age       : num [1:30] 23 77 38 50 47 71 58 65 44 65
21   ...
22 .. ..$ treatment_group: chr [1:30] "Treatment A" "Treatment B" "
23   Control" "Control" ...

```

```

17 .. ..$ baseline_weight: num [1:30] 74.1 54.3 81 69 57.9 56.4 51.4
    99.4 94.4 90.5 ...
18 .. ..$ height          : num [1:30] 184 169 150 161 183 ...
19 .. ..$ comorbidity      : chr [1:30] "Diabetes" "None" "None" "None"
    " ...
20 ..@ TimeGrids          :List of 4
21 .. ..$ Cosine          : num [1:179] -10 -9.9 -9.6 -9.5 -9.4 -9.2 -9.1
    -9 -8.9 -8.8 ...
22 .. ..$ Hyperbola       : num [1:190] -10 -9.9 -9.8 -9.7 -9.6 -9.5 -9.4
    -9.3 -9.2 -9.1 ...
23 .. ..$ Parabola        : num [1:181] -10 -9.9 -9.8 -9.7 -9.5 -9.4 -9.3
    -9.2 -9.1 -9 ...
24 .. ..$ Sine            : num [1:176] -10 -9.9 -9.8 -9.7 -9.6 -9.5 -9.4
    -9.3 -9.2 -9.1 ...

```

The components of CONNECTORData are:

- curves: a tibble similar to TimeSeries, but with the addition of curvesID, which is the concatenation of SubjID and measureID.
- dimensions: a tibble that contains the number of points for each curve.
- annotations: a tibble that contains data imported from AnnotationFile.
- TimeGrids: a list containing the time grids for each measure.

### 3 Data visualization

The PlotTimeSeries function generates the plot of the sampled curves, coloured by the selected feature from the AnnotationFile. Figure reports the line plot generated by the following code:

```

1 PlotTimeSeries(Data, feature="treatment_group")

```

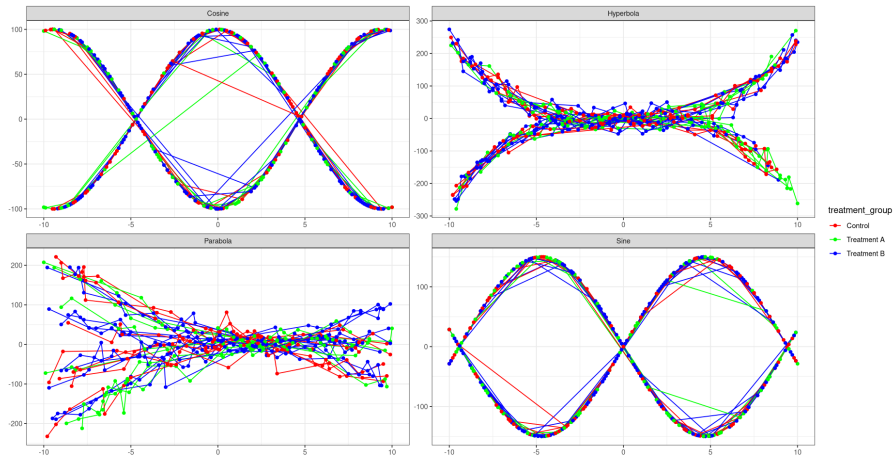


Figure 3: Example for PlotTimeSeries

The DataVisualization function plots the time distribution helping in the inspection of the sparsity of the time points.

```
1 DataVisualization(Data)
```

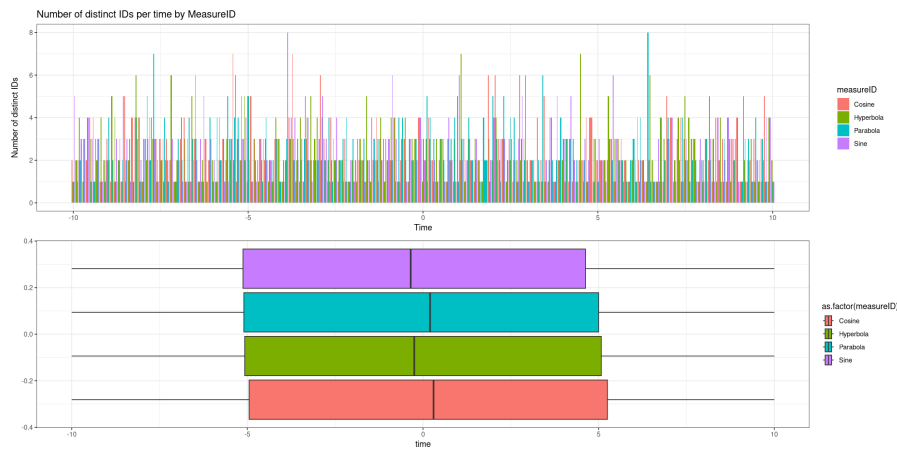


Figure 4: Example for DataVisualization

Setting large as true we can also see the time heat map

```
1 DataVisualization(Data, large=T)
```

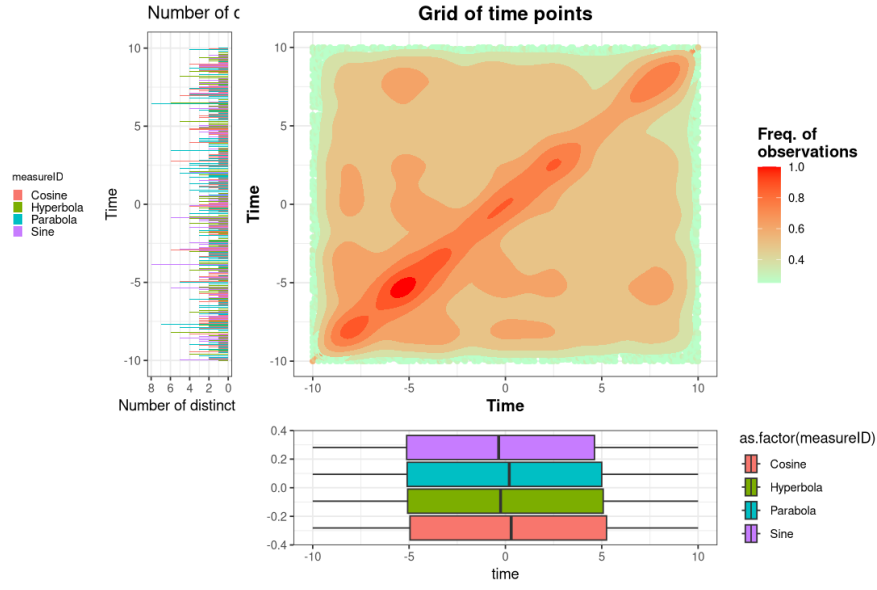


Figure 5: Example for DataVisualization large=T

The DataTruncation function have been developed to truncate the time series at specific time value.

```
1 DataTruncation(Data, feature = "gender",measure = "Parabola",
  truncTime = 5)
```

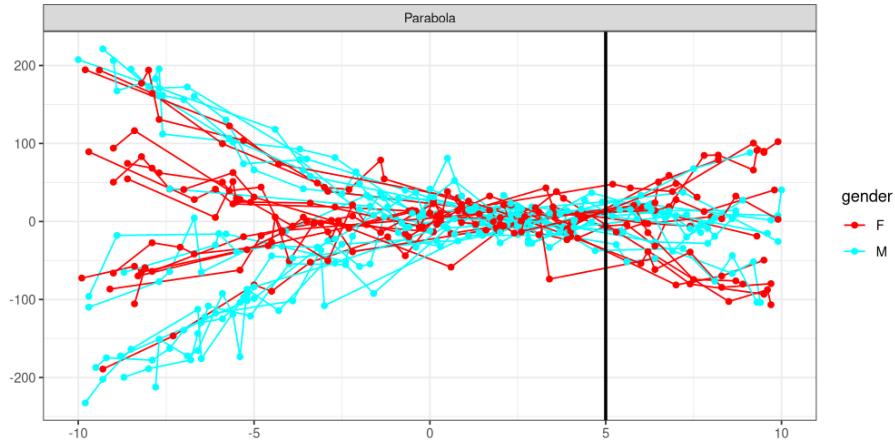


Figure 6: Example for DataTruncation

## 4 Model Selection Tools

Before running the fitting and clustering approach, we have to properly chose the two free parameters:

- the spline basis dimension,  $p$ ;
- the number of clusters,  $G$ .

We developed several functions to enable the user to properly set the free parameters.

#### 4.1 The spline basis dimension - $p$

The dimension of the spline basis can be chose by exploiting the *BasisDimensionChoice* function, by taking the  $p \in [p_{\min}, p_{\max}]$  value corresponding to the largest cross-validated likelihood, as proposed in (James, Hastie, and Sugar 2000), where the interval of values  $[p_{\min}, p_{\max}]$  is given by the user. In particular, a ten-fold cross-validation approach is implemented: firstly the data are split into 10 equal-sized parts, secondly the model is fitted considering 9 parts and the computation of the log-likelihood on the excluded part is performed. A plot is returned: The CrossLogLikePlot return the plot of the mean tested log-likelihoods versus the dimension of the basis, see Figure 7. Each gray dashed line corresponds to the cross-log-likelihood values obtained on different test/learning sets and the solid black line is their mean value. The resulting plot could be used as a guide to choose the largest cross-validated likelihood. Specifically, the optimal value of  $p$  is generally the smallest ensuring the larger values of the mean cross-log-likelihood function. CossLogLikePlot is a list containing a plot for each measure

```
1 CrossLogLikePlot <- BasisDimensionChoice(Data, p=2:10)
```

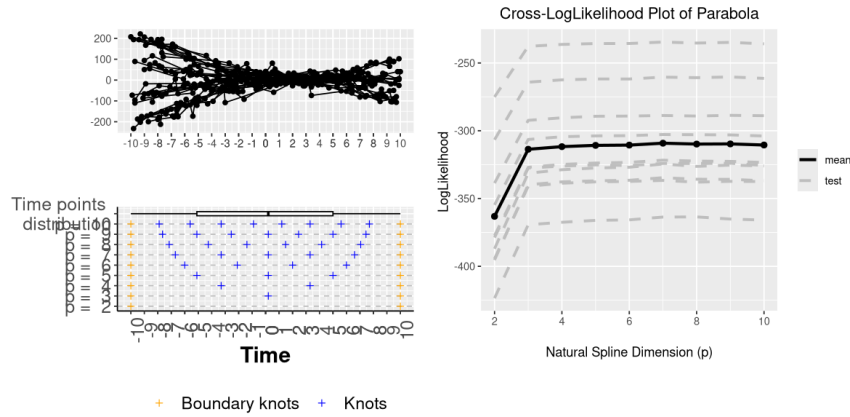


Figure 7: Example for the first element of BasisDimensionChoice

```
1 #set p
2 p <- 3
```

In our example, the optimal value of  $p$  is 3.



## 4.2 The number of clusters - G

Using `ClusterAnalysis` we can choose which G is better based on TT, fDB and silhouette score

```
1 clusters <- ClusterAnalysis(Data, G=2:6, p=c(4,4,6,6), runs=100)
```

The function `IndexesPlotExtrapolation`

```
1 IndexPlotExtrapolation(clusters)
```

generate one plot reported in Figure 8. In details, the fDB, silhouette and tightness indexes are plotted for each value of G. Each value of G is associated with a violin plot created by the distribution of the index values collected from the runs performed. The stars indicates maximum frequency, minimum fDB and maximum silhouette score.

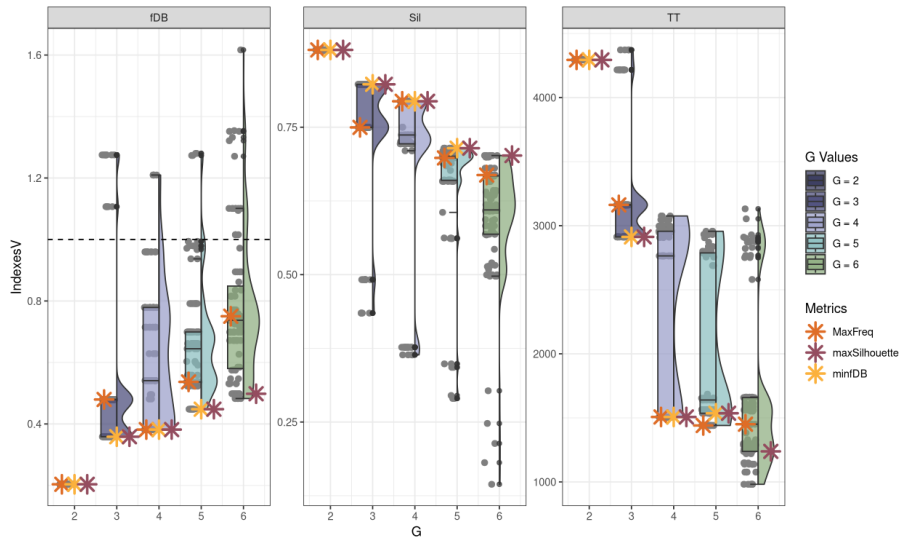


Figure 8: Example for `IndexPlotExtrapolation`

Then we can set our preferred setup by `ConfigSelection`. In our case the best one is G=2

```
1 Set <- ConfigSelection(clusters, G=2, "MinfDB")
```

Then we can plot with `IndexPlotExtrapolation2` for each measure all clusters with their mean, coloring each curve by a feature

```
1 IndexPlotExtrapolation2(Data, ConfigChosen=Set, KData =  
clusters$KData, feature="comorbidity")
```

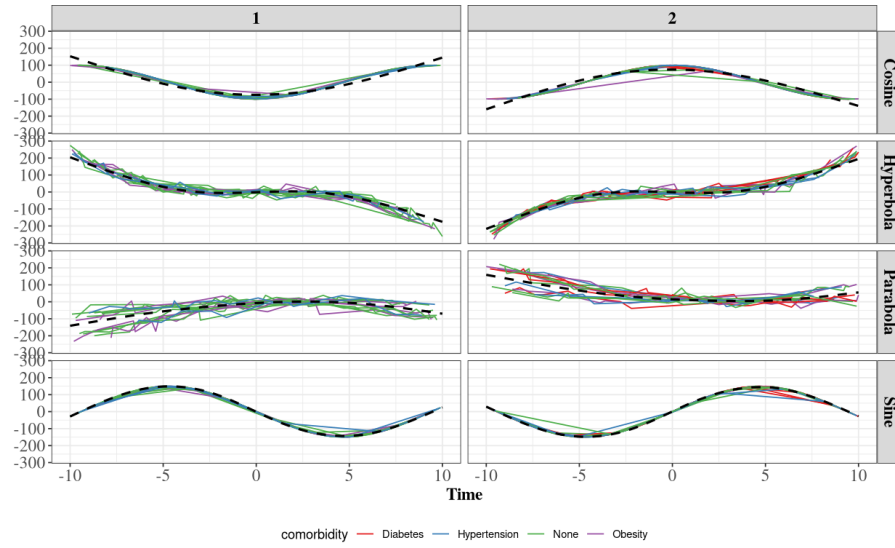


Figure 9: Example for IndexPlotExtrapolation2

You can also plot a Discriminant plot with *DiscriminantPlot*. If the clustering division can be reduced using PCA, the plot dimensions will be automatically adjusted.

```
1 DiscriminantPlot(Data, ConfigChooosen=Set, KData=clusters$KData,
  feature="gender")
```

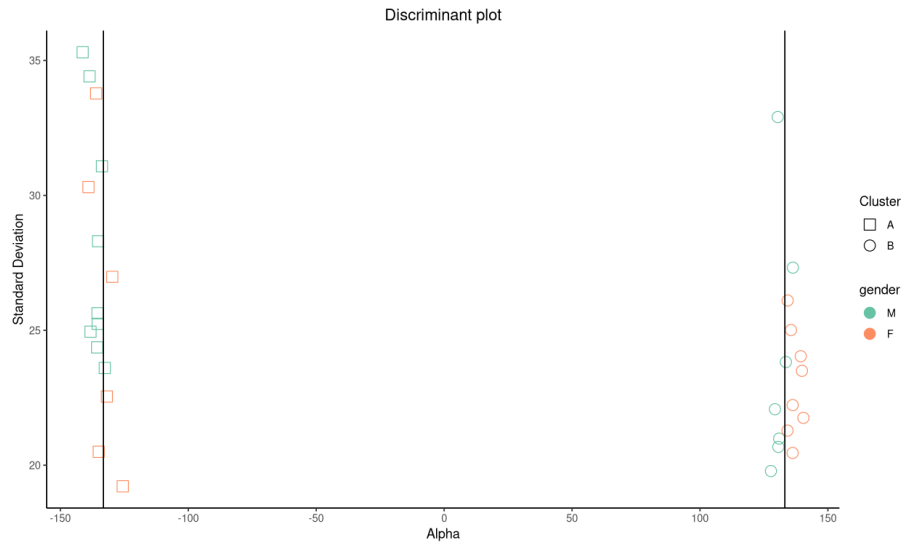


Figure 10: Example for DiscriminantPlot

You can also see entropy and silhouette score for each subject

```
1 SilEntropy(Set, clusters)
```

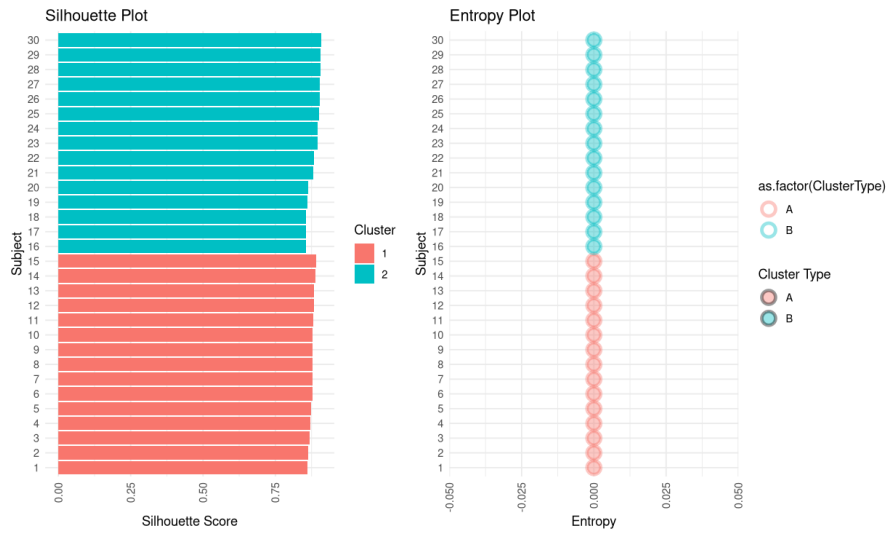


Figure 11: Example for SilEntropy

## 5 SplinePlot

We can recostruct splines using our data as we can see in figure

```
1 splinePlot(KData=clusters$KData, ConfigChosen = Set)[1]
```

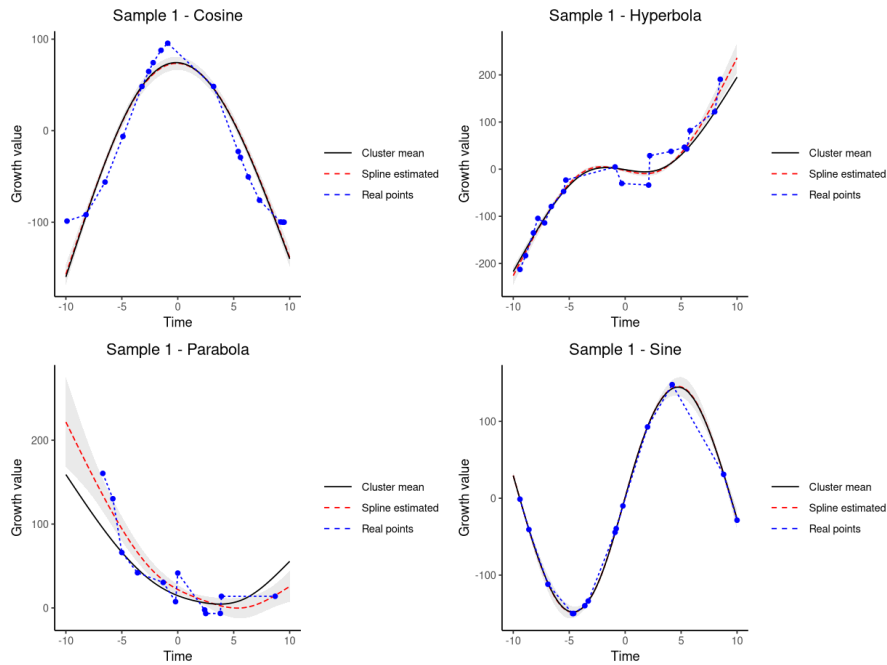


Figure 12: Example for SplinePlot

## 6 MaximumDiscriminationFuncion

With

```
1 MaximumDiscriminationFunction(ConfigChooosen = Set, KData =
clusters$KData)
```

we can plot the next figures:

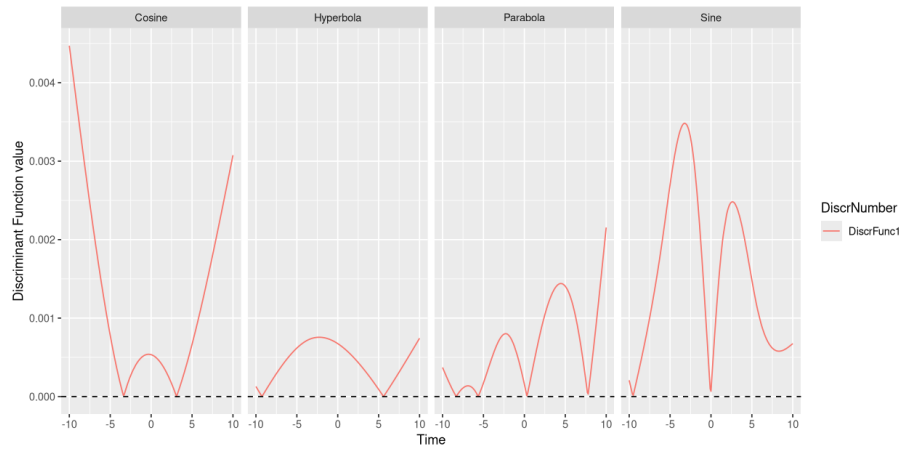


Figure 13: Example for MaximumDiscriminationFuncion[1]

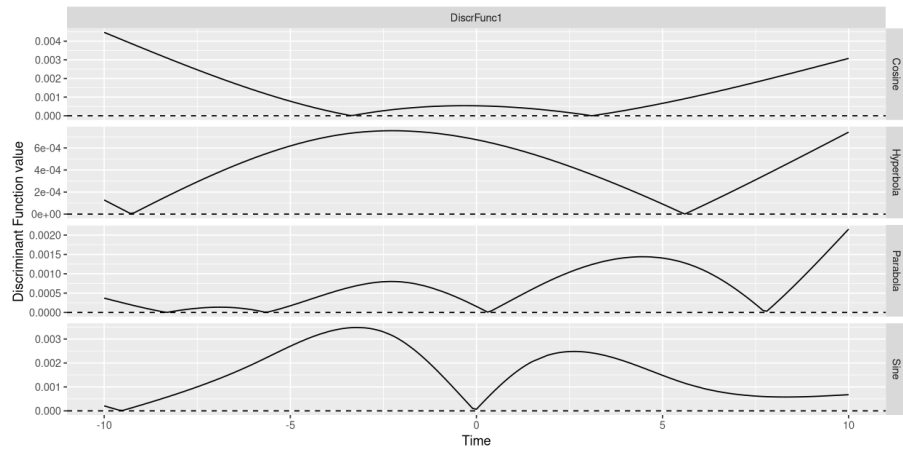


Figure 14: Example for MaximumDiscriminationFuncion[2]