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## Introduction

qWizard is a web-application embedded into the QBiC portal (<https://portal.qbic.uni-tuebingen.de>) allowing users to create a full-factorial experimental design. In a full-factorial design every experimental variable, e.g. genotype, different tissue types, treatments etc., as given by the user is multiplied by the number of replicates, leading to the number and type of samples. To give an example illustrating this principles: Selecting two different tissues from one patient will result in two samples, while selecting two different tissues from two patients will results in four samples. To this end, we see several key advantages in using qWizard:

- (a) it speeds up experimental design
- (b) it helps to perform statistically sound experiments

In reality, full-factorial designs are rare, so qWizard enables users to remove unnecessary samples as well as specify.

An additional consideration is that qWizard uses a multi-tier data model. This means that there are (currently) three different main levels to input information about experimental design and samples. These levels are the sample source (or patient) level, the tissue (or cell culture) level and the analyte (measured samples or "test samples") level. Further levels exist, for example for specific experimental steps like in MHC Ligand Extraction or the Mass Spectrometry runs themselves, but information about these is either collected at the end of the wizard or when the data is registered.

# Step-By-Step Explanation

In the first step, a project space is selected. An existing sub-project can be chosen to add experimental steps with samples or a new sub-project can be created. If a new sub-project is created, a description is needed. Other information like a meaningful short name and people involved in the project can be added:

Create Project

Import Project

Update Metadata

Project

BIH\_DEMO

Sub-Project

New Sub-Project

QDEMO

Short name

Name of sub project

Principal Investigator

Andreas Friedrich

Contact Person

Project Manager

Description \*

Demo project

At the bottom of the first step, we can either add a completely new experiment or specify which kind of addition to an existing sub-project we would like to make. We can also create an empty sub-project and add samples later.

If our sub-project is a pilot study, the related box can be checked.

Description \*

Demo project

- ☒ Add new experiment [\[?\]](#)
- ☐ Add sample extraction to existing sample sources
- ☐ Measure existing extracted samples again
- ☐ Create empty sub-project
- ☐ Download existing sample spreadsheet
- ☐ Add similar samples

Pilot Project

☐ [\[?\]](#)

In the next step, the source of our samples has to be selected. In this example, we want to compare a group of healthy humans with a group of influenza patients. We select Homo Sapiens and the experimental variable “Health State”, as well as 3 replicates (each group will consist of 3 people):

Sample Sources [\[?\]](#)

Experimental Step Name

Experimental Variable



Health State



Species \*

Homo sapiens



Contact Person



How many identical biological replicates (e.g. animals) per group are there?

3

Since we selected the “health state” variable, the next step asks us to specify which kind of health states are investigated in this project. After adding this information (one per line) and selecting the type of value (in this case “categorical” information), the amount (which can be changed) and type of patients are summarized:

## Health State

Values \*

healthy  
influenza

☐ Continuous [?](#)☒ Categorical

## Preview of Combinations

Factors	Samples	<a href="#">?</a>
healthy	3	
influenza	3	

In the next step, we can make some last changes to our patient information. We can change names, add a second identifier or delete superfluous samples:

## Sample Sources Tailoring [\[?\]](#)

### 6 Samples

Secondary Name	External DB ID	health_state	Customize
<input type="text" value="healthy"/>	<input type="text"/>	healthy	
<input type="text" value="healthy"/>	<input type="text"/>	healthy	
<input type="text" value="healthy"/>	<input type="text"/>	healthy	
<input type="text" value="influenza"/>	<input type="text"/>	influenza	
<input type="text" value="influenza"/>	<input type="text"/>	influenza	
<input type="text" value="influenza"/>	<input type="text"/>	influenza	

**Remove Secondary Names**

**Remove External IDs**



The following step handles sample extracts: the kind of tissue taken from patients for this experiment. For special cases (often cell-cultures), isotope labeling can be selected. In this case we only select that blood samples are taken. Since we draw only one blood sample per patient, we leave the number of replicates at 1:

Sample Extracts [\[?\]](#)

Experimental Step Name

Experimental Variable



Isotope Labeling

☐

Tissue \*

Contact Person



Extracted replicates per patient/animal/plant per experimental variable.







We could theoretically add other experimental variables at this step (more on that later), but since we did not, we are presented with the summary step of this (extract) level:

## Sample Extracts Tailoring [\[?\]](#)

### Pool Sample Extracts

☐[\[?\]](#)

### 6 Samples

Secondary Name	External DB ID	health_state	Customize
healthy		healthy	
healthy		healthy	
healthy		healthy	
influenza		influenza	
influenza		influenza	
influenza		influenza	

Remove Secondary NamesRemove External IDs[\[?\]](#)

For cases using isotope labeling, samples can be pooled here. Again, names of the blood samples can be changed or single samples deleted.

In the next step, the analysis method(s) are selected. Let's say we want to sequence DNA as well as measure RNA abundance of different active genes using a Microarray and extract one of each analyte samples from each blood sample (keep "1" as replicate):



Analysis Method [\[?\]](#)

No further preparation of samples?

☐

[\[?\]](#)

Analyte

Analyte

[\[?\]](#)



Replicates

Replicates

[\[?\]](#)

Contact Person

Contact Person

[\[?\]](#)

Pool/Multiplex Samples

☐

Pool/Multiplex Samples

☐

[\[?\]](#)

---

In cases where no Analytes are extracted (e.g. medical imaging), we can stop the experiment process at this point by selecting “No further preparation”. In both cases we are presented with a summary in the next step:

## Sample Registration [\[?\]](#)

### Summary

Type	Content	Samples <a href="#">[?]</a>
Sample Sources	Homo Sapiens	6
Sample Extracts	Whole Blood	6
Split Sample Preparations	RNA, DNA	12

[Download Spreadsheet](#) [\[?\]](#)

- ☒ I am aware of all costs associated with this project, since I previously signed a corresponding agreement and I agree to pay all charges upon receipt of the invoice. This submission is binding.
- ☐ I understand that this experimental design draft will now be submitted for QBiC review. I thereby request a consultancy meeting. There are no costs associated with this submission.

[Register All Samples](#)

We created 6 patient objects, 6 blood samples, and 6 (RNA) + 6 (DNA) samples, leading to 12 analyte samples in total. We can now select to register the experiment(s) and samples we created or send in the study design for review.

---

## Summary and File Upload [?](#)

Your Experimental Design was registered. Project QDEMO now has 6 Sample Sources and 18 samples.  
Project description: Demo project...



All done!

Download Spreadsheets:

Sample Sources

Sample Extracts

Sample Preparations

Show in Project Browser

Upload Additional Files

☐

Download Spreadsheets:

Sample Sources

Sample Extracts

Sample Preparations

Show in Project Browser

Upload Additional Files



Description \*

abc

Attach to...

Experimental Desi



Select File

Durchsuchen...

test.tsv

Add File

## Special Cases

### Experiments on Cell Lines and Single Cell Organisms

The three-tier-model of sample source, tissue and analyte preparation leads to some ambiguous solutions concerning the handling of single cell organisms and cell lines.

For single cell organisms like bacteria, that can be selected, a “tissue” has to be chosen at the second level. Here, selecting “**Full Organism**” is common. When selecting the option “**Other**”, something else can be specified.

Although it can be argued that the genetic makeup of some (cancer) cell lines makes them unique organisms themselves, qWizard models **cell lines on the level of Tissue Extracts**.

This means that an organism has to be chosen in the first step. This can either be the origin of the cell line, e.g. **Homo sapiens** for HeLa cells, or “**Unidentified**” if no further information is available.

When selecting **Cell Line** from the drop-down menu in the **Sample Extraction** step, a cell line can be specified:

[Create Project](#) [Import Project](#) [Update Metadata](#) [Admin Fu](#)

1. Project

2. Sample Sources

3. Summary

4. Sample Extr.

Sample Extracts [\[?\]](#)

Experimental Step Name

Cell Lines

Experimental Variable

+

[?]

-

Isotope Labeling

☒

[?]

SILAC

Tissue \*

Cell Line

Cell Line

HELA cell

Multiple different cell lines can be used in the same experiment by selecting the **Experimental Variable** “**Tissue**”:

Tissues

Tissues 1 \*

Cell Line

Cell Line

NALM-1 cell

Tissues 2 \*

Cell Line

Cell Line

KG-1 cell

Tissues 3 \*

Cell Line

Cell Line

HeLa BC1-CB cell

## Multiple Levels of Exp. Factors

Complex experiments may not only examine a single experimental factor (with several test conditions) but the interplay of factors like genotype, treatment and different time points after the treatment. qWizard always multiplies factors within and between different levels of the experiment, creating all possible permutations of these attributes as a sample (or multiple samples, depending on the number of chosen replicates). Users can then fine-tune the numbers of the different sample types in their experiment.

For this to work, however, care has to be taken to input the conditions at their respective levels.

In this example, 6 mice are analyzed. They differ in genotypes (**3 WT and 3 KO**) and phenotypes (**2 affected and 4 control**) Furthermore, two different extracts (**blood and lymph - one for each mouse**) are sampled.

After adding the general information about the project (see Chapter X), we select the two experimental variables concerning our mice in the **Sample Sources** step. We choose to leave the number of replicates at 1 and to fine-tune this information in the following step.

Create Project

Import Project

Update Metadata

1. Project

2. Sample Sources

3. Biol. Variables

4. Summary

Sample Sources

Experimental Step Name

Experimental Variable

Phenotype

Experimental Variable

Genotype

Species \*

Mus musculus

Contact Person

How many identical biological replicates (e.g. animals) per group are there?

1

After selecting genotype and phenotype, the next step asks for the different **test conditions** of the two types of experimental variable. We add **affected** and **control**, as well as **WT** (wild type) and **KO** (knock out).

Phenotype

Genotype

Values \*

affected

control

☐ Continuous [?](#)

☒ Categorical

Phenotype

Genotype

Values \*

WT

KO

☐ Continuous [?](#)

☒ Categorical

Preview of Combinations [?](#)

Factors	Samples
affected ; WT	1
affected ; KO	1
control ; WT	1
control ; KO	1

The resulting preview of all combinations represents four different types of mice:

**affected+WT; affected+KO; control+WT and control+KO.**

Since we chose only one replicate in the previous step, qWizard assumes that the experiment contains one mouse of each type.

However, since we have two control mice of each genotype, we change the number accordingly:

#### Preview of Combinations

Factors	Samples
affected ; WT	1
affected ; KO	1
control ; WT	2
control ; KO	2







In the following step we see the six mice in our experiment. We can rename the **Secondary Name**, which automatically contains an overview of all experimental factors or we can add



Lab IDs (in this case just “mouse 1” etc.). Both of these infos are used in many other parts of qPortal to make it easier to identify samples or sample sources.  
Experimental Factors are saved after sample registration, even if we change the names of our sources/samples.

## Sample Sources Tailoring [\[?\]](#)

### 6 Samples

Secondary Name	External DB ID	phenotype	genotype	Customize
affected ; WT	mouse 1	affected	WT	
affected ; KO	mouse 2	affected	KO	
control ; WT	mouse 3	control	WT	
control ; WT	mouse 4	control	WT	
control ; KO	mouse 5	control	KO	
control ; KO	mouse 6	control	KO	

Since we only input information about our lab animals up to this point, we still need to specify which samples are taken from them. This happens in the next step: **Sample Extracts**.  
Since we want to extract **blood** as well as **lymph**, we chose **Tissue** as an experimental variable and set the number of tissue types to 2. Again, we leave the number of extracted replicates at 1 for simplicity's sake:

Create ProjectImport ProjectUpdate MetadataAdd

1. Project2. Sample Sources3. Biol. Variables4. Summary5. Sample Extr.

Sample Extracts

Experimental Step Name

Experimental Variable

Tissue

Isotope Labeling

How many different tissue types are there in this sample extraction?

2

Contact Person

Extracted replicates per patient/animal/plant per experimental variable.

1

The next step allows us to choose the different “tissue types”, in this case **Whole Blood** and **Lymph** from a pre-defined vocabulary. Like in the case of different mice, we see the resulting samples below once we have made our choice. In this case, taking one blood and one lymph sample from each of our six mice leads to 12 extract samples:

## Tissues

Tissues 1 \*



Whole Blood



Tissues 2 \*

Lymph



### Preview of Combinations

Factors	Samples
(2) affected ; WT ; Whole Blood	1
(2) affected ; WT ; Lymph	1
(3) affected ; KO ; Whole Blood	1
(3) affected ; KO ; Lymph	1
(4) control ; WT ; Whole Blood	1
(4) control ; WT ; Lymph	1
(5) control ; WT ; Whole Blood	1
(5) control ; WT ; Lymph	1
(6) control ; KO ; Whole Blood	1
(6) control ; KO ; Lymph	1
(7) control ; KO ; Whole Blood	1
(7) control ; KO ; Lymph	1



After selection of Analyte (and no technical replicates), this leads to the following result which we can register in the system or send in for QBiC review:

## Sample Registration [\[?\]](#)

### Summary

Type	Content	Samples <a href="#">[?]</a>
Sample Sources	Mus Musculus	6
Split Sample Extracts	Lymph, Whole Blood	12
Sample Preparations	RNA	12

[Download Spreadsheet](#) [\[?\]](#)

- ☒ I am aware of all costs associated with this project, since I previously signed a corresponding agreement and I agree to pay all charges upon receipt of the invoice. This submission is binding.
- ☐ I understand that this experimental design draft will now be submitted for QBiC review. I thereby request a consultancy meeting. There are no costs associated with this submission.

[Register All Samples](#)

The Batch Upload functionality (see next chapter) automatically recognizes to which tier of the experiment an experimental factor should belong. E.g. if two conditions of the same specified factor are different for the same mouse, then this condition can not be related to the mouse (like, for example, a genotype), but has to belong to different tissue extracts or analytes, like a time point of extraction.

## Batch Upload of Samples

Creating large experiments with many samples and rich metadata by hand can sometimes be a hassle. To simplify and speed up this process, we have created several spreadsheet file formats that can be used to directly upload experiments or update the metadata of existing experiments afterwards (see Chapter X).

### General Format

- Columns are tab-separated
- No quotation marks or additional tabs should be used
- Order of columns can be random

All Batch Upload files must contain the basic information about Sample Sources, Tissues and Analytes (explained in Chapter Y) as well as label the different entities of each of these levels with unique IDs:

Organism	Organism ID	Tissue	Extract ID	Analyte	Analyte ID
Mus musculus	m1	Whole Blood	B1-1	DNA	R3001
Mus musculus	m2	Whole Blood	B2-1	DNA	R3002
Mus musculus	m3	Whole Blood	B3-1	DNA	R3003

In this example experiments, blood samples from 3 mice are taken and DNA is extracted. Since each sample and each mouse is unique, all the identifiers (on all three levels) are unique as well.

To illustrate how **replicates** are handled, we will add two additional DNA measurements: one further blood sample (B2-2) taken from the second mouse (m2), as well as one taken from the existing blood sample (B3-1):

Organism	Organism ID	Tissue	Extract ID	Analyte	Analyte ID
Mus musculus	m1	Whole Blood	B1-1	DNA	R3001
Mus musculus	m2	Whole Blood	B2-1	DNA	R3002
Mus musculus	m3	Whole Blood	B3-1	DNA	R3003
Mus musculus	<b>m2</b>	Whole Blood	<b>B2-2</b>	DNA	R3012
Mus musculus	<b>m3</b>	Whole Blood	<b>B3-1</b>	DNA	R3013

Keep in mind, that these identifiers are just examples. In reality, they can be based on lab-internal labeling, as long as they are unique for different entities.

## Adding additional meta-information

Experimental factors (and other properties TODO) can be added in additional columns:

Organism ID	Extract ID	Analyte ID	Condition:	Condition:	Condition:
-------------	------------	------------	------------	------------	------------

			genotype	extraction_time	protocol
m1	B1-1	R3001	gene+	t1	a
m2	B2-1	R3002	gene+	t1	a
m3	B3-1	R3003	gene-	t1	a
m2	B2-2	R3012	gene+	t2	b
m3	B3-1	R3013	gene-	t1	b

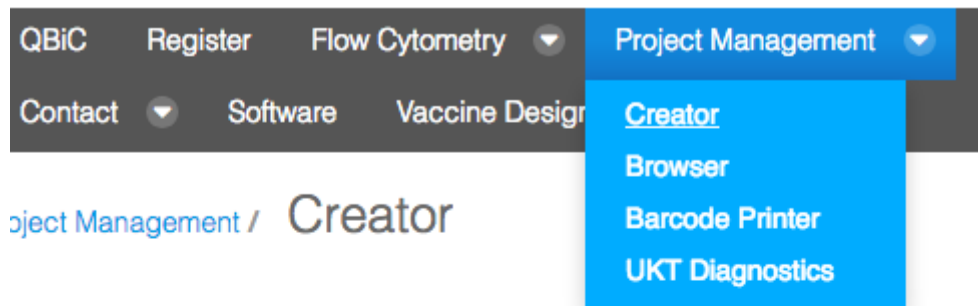
In this example, the conditions genotype (on Organism level), extraction time (of the blood, Extract level) and protocol of DNA preparation was added: Mice m1 and m2 have a certain allele, while m3 is a knockout; all blood samples but B2-2 were taken at the same time and the last two DNA measurements were performed using a new protocol ("b").

Organism, Tissue and Analyte columns have been removed from this example for the sake of clarity.

To keep the basic upload format simple, additional properties can be added with the Metadata Upload functionality (see Chapter X).

## Upload of the Experiment

The upload functionality can be found as part of the Project Creator on qPortal's Project Management Section:

The image shows the 'Import Project' form. At the top, there are three tabs: 'Create Project', 'Import Project' (which is active), and 'Update Metadata'. Under the 'Import Project' tab, there are two columns: 'Import Format' and 'Format Information'. The 'Import Format' column has three radio buttons: 'QBiC', 'Standard', and 'MHC Ligandomics (measured)'. The 'Format Information' column has three blue question mark icons. Below these columns, there is a section titled 'Upload your file here' with a 'Durchsuchen...' button, a 'Keine Datei ausgewählt.' status, and an 'Upload' button.

To register your experiments follow these steps:

1. Create a TSV file using the information above or the example
2. Select "Import Project"
3. Select "Standard" and upload the file
4. Select Project and Sub-project and complete other missing information
5. Register experiments, a download link is created containing one barcode per row of the uploaded file
6. Download file use the first two columns to add the QBiC barcode to each file
7. Transfer the files (more information about this can be found...)