# Package 'bmisc'

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Type Package

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Title Miscellaneous functions

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<b>Description</b> This package has different functions that I have accumulated with time. This is the Alpha version.
Depends car, lattice, zoo, robustbase, methods
License LGPL $>= 3.0$
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att.strp 3

att.strp

Attibute stripper

#### Description

Strips an object of its attributes

### Usage

```
att.strp(x)
```

#### Arguments

х

the name of an object (vector, matrix, data.frame, array or list)

#### **Details**

This function strips an object of its attributes. In the case of a vector, all attributes are removed. For a matrix or an array, only c('dim', 'dimnames') are kept. When att.strp is used on a data.frame, all attributes of the variables are striped and only c('names', 'row.names', 'na.action', 'class') are kept for the data.frame object.

# Value

returns an object of the same class as the original one.

# Author(s)

Benoit Bruneau

```
Creating different objects
    with added attributes (label)
### numerical vector ###
x <- 1:10
attr(x,"label") <- "test1"
attributes(x)
### data frame ###
z=data.frame(x,x)
attr(z,"labels") <- "test2"
attributes(z)
attributes(z[,1])
attributes(z[,2])
### array ###
y=array(x,c(2,2,2))
attr(y,"labels") <- "test3"
attributes(y)
```

4 att.strp

```
attributes(y[,,1])
attributes(y[,,2])
### list containing the vector, ###
### data frame and array
u=list(x,z,y)
attr(u,"labels") <- "test4"
attributes(u)
attributes(u[[1]])
attributes(u[[2]])
attributes(u[[3]])
attribute stripping
x2=att.strp(x)
z2=att.strp(z)
y2=att.strp(y)
u2=att.strp(u)
# verification of the attributes #
   for all stripped objects
### numerical vector ###
attributes(x2)
### data frame ###
attributes(z2)
attributes(z2[,1])
attributes(z2[,2])
### array ###
attributes(y2)
attributes(y2[,,1])
attributes(y2[,,2])
### list containing the vector, ###
### data frame and array
attributes(u2)
attributes(u2[[1]])
                     # vector in the list
attributes(u2[[2]])
                     # data frame in the list
attributes(u2[[2]][,1]) # data frame in the list
attributes(u2[[2]][,2]) # data frame in the list
attributes(u2[[3]]
                     # array in the list
attributes(u2[[3]][,,1]) # array in the list
attributes(u2[[3]][,,2]) # array in the list
```

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bmisc

 $Miscellaneous\ functions$ 

# Description

This package has different functions that I have accumulated with time. I am not the author of all of them even though I have modified most of them. This is the Alpha version.

# **Format**

 $\begin{array}{lll} \mbox{Package:} & \mbox{bmisc} \\ \mbox{Type:} & \mbox{Package} \\ \mbox{Version:} & 0.2\text{-}12 \\ \mbox{Date:} & 29\text{-}07\text{-}2011 \\ \mbox{License:} & \mbox{LGPL} >= 3.0 \end{array}$ 

### Details

For pdf version of the help, write vignette("bmisc").

# Author(s)

Benoit Bruneau

Maintainer: Benoit Bruneau <br/> <br/> denoit.bruneau1@gmail.com>

ceiling.lg

ceiling.lg

 $ceiling\ largest$ 

# Description

Ceiling to largest digit

# Usage

```
ceiling.lg(x)
```

# Arguments

х

Numeric vector

# Details

Gives the ceiling to largest digit (i.e., 54 -> 60).

```
ceiling.lg(250)
ceiling.lg(25000000)
```

clean 7

clean

Clean a Data Frame

### Description

Cleans a data.frame from a starting point with a defined threshold

### Usage

```
clean(data= x, col.start =1, min.val=NULL)
```

# Arguments

data then name of the data.frame

col.start indicate the columns from which to start reading

min.val numeric. Read details

### Details

min.val is the minimum value accepted in a column. Colomns with this value or higher will be kept in the data.frame.

More will be added to this function.

#### Value

returns the data.frame with the clean columns

# Author(s)

Benoit Bruneau

```
x=rnorm(50 , 20, 12)
y=runif(50 )
z=rpois(50, 3)
v=x*y/z
t=z*v
pp=data.frame(aa=x, bb=y, cc=v, dd=z, ee=t)
summary(pp)
pp1 = clean(pp, min.val=0.06)
```

8 corr.perm

corr.	nerm

Pearson Correlation by Permutation

# Description

Tests the Pearson correlation estimate (r) by use of permutation

# Usage

```
corr.perm(x,y,nperm=999)
```

### Arguments

x,y Two vectors of same length used for correlation analysis

nperm Number of permutations (default = 999)

# Value

Correlation	Pearson r
-------------	-----------

 ${\tt P.perm} \qquad \qquad {\tt pvalue \ estimated \ by \ permutations}$ 

P.para parametric pvalue estimated

inf inferior limit of the confidence intervalsup superior limit of the confidence interval

df degree of freedom

```
x <- rnorm(50,0,1)
y <- runif(50,0,1)*x
toto = corr.perm(x, y)</pre>
```

cv 9

cv

Coefficient of Variation (CV)

### Usage

```
cv(x, na.rm=T)
```

### Arguments

x an R object (vector, matrix,...)

na.rm a logical value indicating whether NA values should be stripped before

the computation proceeds

# Details

The coefficient of variation (CV) is the ratio of the standard deviation to the mean. The CV is defined for the absolute value of the mean to ensure it is always positive.

# Examples

x=rnorm(50)
cv(x)

10 day

 ${\tt day} \hspace{1cm} day$ 

# Description

Day of year as decimal number (001-366).

# Usage

day(x)

# Arguments

х

# Examples

# will soon be available

Errbar 11

Errbar	error bars

# Description

Adds error bars on a plot

### Usage

### Arguments

X	numeric vector
У	numeric vector
xinf, xsup	numeric vectors containing the upper (xsup) and/or lower (xinf) limits of the confidence interval for x-axis values.
yinf, ysup	numeric vectors containing the upper (ysup) and/or lower (yinf) limit of the confidence interval for y-axis values.
xCI	numeric vectors containing the confidence intervals for x-axis values.
yCI	numeric vectors containing the confidence intervals for y-axis values.
•••	additional graphical arguments (par) such as $col, lty, lwd$ and/or arguments for $arrows$ .

### Details

If xCI and/or yCI are defined, individually defined limits (ie. xinf, xsup, yinf, ysup) are not used.

### See Also

```
arrows, par
```

```
x <- 1:10
y <- x + rnorm(10)

yci <- runif(10)
xci <- runif(10)

plot(x,y, ylim=c(min(y-yci),max(y+yci)))
Errbar( x, y, yCI=yci)

plot(x,y, xlim=c(min(x-xci),max(x+xci)))
Errbar( x, y, xCI=xci )

plot(x,y, ylim=c(min(y-yci),max(y+yci)), xlim=c(min(x-xci),max(x+xci)))
Errbar( x, y, yCI=yci, xCI=xci )

# Gives an Error message</pre>
```

12 Errbar

fct 13

fct

 $Print\ bmisc\ functions$ 

# Description

Print all functions of bmisc package

# Usage

fct()

14 format.hms

format.hms

 $Format\ seconds\ into\ hours$ 

# Description

Transforms time format

# Usage

format.hms(sec)

# Arguments

sec

time expressed in seconds

### Value

 ${\it returns \ hrs:} {\it min:} {\it sec}$ 

# Examples

format.hms(20000)

gam.Check 15

gam.Check	Some diagnostics for a fitted game	$n \ model$
-----------	------------------------------------	-------------

### Description

Takes a fitted gam object produced by gam() and produces some diagnostic information about the fitting procedure and results. The default is to produce 4 residual plots, and some information about the convergence of the smoothness selection optimization.

### Usage

#### Arguments

```
b a fitted gam object as produced by gam().

main a character vector containing the four titles to be used.

xlab a character vector containing the four x labels to be used.

ylab a character vector containing the four y labels to be used.

text a character or expression vector specifying the text to be written.

args.histplot list of additional arguments to pass to histplot()

... additional text and graphical parameters (see par, mtext)
```

#### **Details**

This function plots 4 standard diagnostic plots, and some other convergence diagnostics. Usually the 4 plots are various residual plots. The printed information relates to the optimization used to select smoothing parameters. For the default optimization methods the information is summarized in a readable way, but for other optimization methods, whatever is returned by way of convergence diagnostics is simply printed.

This is a modified version of gam.check from mgcv-package so that main titles, x labels and y labels can be customized.

### References

Wood S.N. (2006) Generalized Additive Models: An Introduction with R. Chapman and Hall/CRC Press.

16 gam.Check

```
library(mgcv)
set.seed(0)
dat <- gamSim(1,n=200)
b<-gam(y~s(x0)+s(x1)+s(x2)+s(x3),data=dat)
plot(b,pages=1)

gam.check(b)
gam.check(b, main=c("A","B","C","D"))</pre>
```

get.partial.etas 17

 ${\tt get.partial.etas} \qquad \qquad get \ partial \ et as$ 

# Usage

get.partial.etas(model)

# Arguments

model

# Examples

# will soon be available

18 histplot

### Usage

### Arguments

dat one of:

• a numeric vector,

• an object of class "norm" resulting from a call to norm.test

breaks one of:

• a vector giving the breakpoints between histogram cells,

• a single number giving the number of cells for the histogram,

• a character string naming an algorithm to compute the number of cells (see 'Details'),

• a function to compute the number of cells.

In the last three cases the number is a suggestion only.

barc a color to be used to fill the bars.

borc a color to be used for the borders the bars.

fit.norm a logical variable indicating whether to fit a normal density curve (TRUE)

or not (FALSE).

lcol color of the normal density curve

stat the statistic to add on the graph. One of (c("all", "mean", "median")).

Default is NULL.

stat.lab a character vector with the labels for the estimated mean and/or median.

Default is c("Mean", "Median").

rug a logical variable indicating whether to superpose a rug (TRUE) or not

(FALSE).

main the main title of the graph

... additional arguments to be passed to plot (see par)

### **Details**

The default for breaks is "Sturges": see nclass.Sturges. Other names for which algorithms are supplied are "Scott" and "FD" / "Freedman-Diaconis" (with corresponding functions nclass.scott and nclass.FD). Alternatively, a function can be supplied which will compute the intended number of breaks as a function of x.

#### See Also

hist

histplot 19

# Examples

```
x=rnorm(50)
histplot(x)
```

norm.x=norm.test(x)
histplot(norm.x)

20 inv.pred

 ${\tt inv.pred}$ 

 $Inverse\ Predictions\ with\ SE$ 

```
Usage
```

```
xxx( data , , , )
```

# Arguments

data

# Author(s)

Benoit Bruneau

is.even 21

is.even

 $is\ even$ 

# Description

Identifies if a value is even or not

# Usage

is.even(x)

# Arguments

х

numeric vector

# Details

Will returns TRUE if  $\mathtt{roundup}(x)$  is an even number.

# Value

logical

# See Also

is.odd

# Examples

is.even(5)

is.even(6)

is.odd

is.odd

 $is\ odd$ 

# Description

Identifies if a value is odd or not

# Usage

is.odd(x)

# Arguments

х

numeric vector

# Details

Will returns TRUE if roundup(x) is an odd number.

# Value

logical

# See Also

is.even

# Examples

is.odd(5)

is.odd(6)

last 23

last last

Usage

last(x)

Arguments

x

Examples

# will soon be available

24 lev

lev Levene type tests

### Description

Tests heteroscedasticity after an Anova

### Usage

```
lev(y, ...)
## S3 method for class 'formula'
lev(y, data=NULL, ...)
## S3 method for class 'lm'
lev(y, ...)
## Default S3 method:
lev(y, group, data=NULL , trim.alpha = 0.1, type="abs",...)
```

### Arguments

у	response variable for the default method, lm class object for the lm method or formula class object for the formula methode. If y is a linear-model object or a formula, the variables on the right-hand-side of the model must all be factors and must be completely crossed. See details.
group	for the default method, factor (concatenated factor when multiple factors). See details.
data	${\tt data.frame}$ where the dependant variable and the factor(s) are
trim.alpha	Alpha level (percentiles) trimming the data on which the mean will be evaluated
type	Type of transformation made on the residuals. Either "abs" for absolute values or "sq" for sqared values
	arguments to be passed down, e.g., data for the formula method or other options such as type and trim.alpha.

### **Details**

When using the lm method, data doesn't need to be defined. When using the formula or default methods, data can be defined if the data used is in a data.frame.

When group is manually defined in the default method, use paste(x,y,z) or  $\$  interaction(x,y,z) form where "x", "y" and "z" are the factors. There is no restrictions on the number of factors.

O'Brien's (1981) performs test for equality of variances within each group: based on transforming each observation in relation to its group variance and its deviation from its group mean; and performing an ANOVA on these transformed scores (for which the group mean is equal to the variance of the original observations). The procedure is recognised to be robust against violations of normality (unlike F-max).

lev 25

### Value

Model The model

Levene Results for Levene's test

 ${\tt LeveneTrimMean}$ 

Results for Levene's test on the trimmed mean

Brown.Forsythe

Results for Brown-Forsythe's test

OBrien Results for O'Brien's test

#### See Also

```
leveneTest from {car}
```

26 lib.code

lib.code

Retreives the code for lib().

# Description

Will print in the R windows the code for lib() (READ DETAILS).

# Usage

```
lib.code()
lib(pack, install=TRUE, load=TRUE, quietly=TRUE,
    warn.conflicts=FALSE)
```

# Arguments

pack

Character vector specifying which package(s) to load/install.

### **Details**

### USE lib.code() TO GET THE CODE FOR THE FUNCTION lib().

lib.code() prints in R the code for lib(). Copy and paste the code for lib() in the file "C:/Program Files/R/R-2.12.1/etc/Rprofile.site" (Windows) or "~/.Rprofile" (Mac).

lib() will load packages named in a charcater vector. If install is TRUE, packages not yet installed will be installed.

### Author(s)

Benoit Bruneau

# Examples

lib.code()

lsmean 27

 ${\tt lsmean} \hspace{15mm} \textit{Least Squares Means}$ 

### Description

THIS FUNCTION IS FROM PACKAGE pda THAT IS STILL UNDER CONSTRUCTION ON R-Forge. IT HAS BEEN INCLUDED IN bmisc FOR PRACTICAL REASONS.

Caution: This routine is not fully tested for models with nested factors or mixed models. Please check results against another package (e.g. SAS proc mixed). It appears to correctly handle lme objects, but does not work well for aov objects that include Error() type nesting in the formula. Further, it does not properly handle polynomial terms-only the linear term is included. For now, create dummies like x2 = x\*x manually and include x2 in your model.

### Usage

```
lsmean(object, ...)
## Default S3 method:
lsmean(object, ..., factors, effects = FALSE, se.fit = TRUE,
    adjust.covar = TRUE)
## S3 method for class 'lm'
lsmean(object, data, factors, expr, contrast, effects = FALSE,
    se.fit = TRUE, adjust.covar = TRUE, pdiff = FALSE,
    reorder = FALSE, lsd, level = .05, rdf, coef, cov, ...)
## S3 method for class 'lme'
lsmean(object, data, factors, ..., rdf, coef, cov)
## S3 method for class 'lmer'
lsmean(object, data, factors, expr, ..., rdf, coef, cov)
## S3 method for class 'listof'
lsmean(object, data, factors, stratum, expr, contrast, ...)
```

### Arguments

object	response vector (default) or model object (lm).
• • •	factors and covariates (must be same length as y).
data	data frame in which to interpret variables (found from object if missing).
factors	character vector containing names of x.factor and trace.factoras first two entries. Must be in $names(data)$ and $labels(object)$ .Default is all factor names.
effects	drop intercept if TRUE (only works properly with sum-to-zero contrasts).
se.fit	compute pointwise standard errors if T.
adjust.covar	adjust means to average covariate values if T; otherwise use covariate mean for each combination of factors.
pdiff	Include letters to signify significant differences.
reorder	Reorder means from largest to smallest.
lsd	Include average LSD if TRUE (also need pdiff=TRUE).
level	Significance level for pdiff calculations.

28 lsmean

rdf Residual degrees of freedom.

coef Coefficients for fixed effects in object.

cov Covariance matrix for fixed effects.

expr Call expression (formula)

contrast Type of contrasts (default is attribute contrasts of object) stratum Name of stratum for Ismean calculation as character string.

### Value

Data frame containing unique factor levels of factors, predicted response (pred) and standard errors (se). WARNING: Ismean may not function properly if there are empty cells. Standard errors for mixed models using methods lmer and listof are not fully debugged.

### Author(s)

Brian S. Yandell

#### See Also

```
predict.
```

```
## Not run:
lsmean(y,x1,x2)
# the following does the same thing
fit <- lm(y~x1+x2)
data <- data.frame(y,x1,x2)
lsmean(fit,data,factors=c("x1","x2")
## End(Not run)</pre>
```

make.z

make.z make z

# Usage

make.z(x, index = NULL)

# Arguments

X

index

# Examples

# will soon be available

30 mc.long

mc.long Pag	irwise t tests in long format
-------------	-------------------------------

### Description

Calculate pairwise T tests between group levels with corrections for multiple testing presented in long format

### Usage

#### Arguments

	У	response variable for the default method, or lm or formula object. If y is a linear-model object or a formula, the variables on the right-hand-side of the model must all be factors and must be completely crossed.
	group	for the default method, factor (concatenated factor when multiple factors). See details.
	data	data.frame where the dependant variable and the factor(s) are
p.adjust.method		
		$\   \text{method for adjusting p values. Default is Holm's method. (see {\tt P.adjust})}$
	column	new names for the factor(s); this is optional
	digits	controls the number of digits for the presented results presented
	silent	a logical variable indicating whether to indicate the general warning (FALSE) or not (TRUE).
		additional arguments to pass to P.adjust, pairwise.t.test and/or t.test.

### **Details**

When making multiple t tests for all combinations, the n option of P.adjust can be used to identify the number of comparisons that are actually used. This is only to simplify the uses p values corrections on the full output matrix when only some of the comparisons are meaningfull or chosen for hypothesis testing.

When group is manually defined, use paste(x,y,z) or interaction(x,y,z) form; "x", "y" and "z" are the factors. There is no restrictions on the number of factors.

### Value

Object of class "data.frame" containing the results.

mc.long 31

### See Also

```
{\tt P.adjust, pairwise.t.test, pair.diff, DTK.test, Tukey HSD \ and \ glht}
```

32 mse

mse

 $Mean\ square\ error$ 

# Description

Estimates the mean square error (mse)

# Usage

```
mse(model)
```

# Arguments

model

an object containing the results of a model.

# Details

The mean square error is also known as the unexplained variance or the variance of the residuals.

n 33

n Sample size (n)

# Description

Gives n without NA's

# Usage

n(x)

# Arguments

x Vector (numeric or character)

```
x= rep(c(rnorm(30,20,5),NA),3)
n(x)
```

norm.test

norm.test

Normality tests

#### Description

Lilliefors (Kolmogorov-Smirnov), Shapiro-Francia, Shapiro-Wilk, D'Agostino Skewness, Anscombe-Glynn Kurtosis and D'Agostino-Pearson normality tests.

#### Usage

```
## Default S3 method:
  norm.test(norm.test(x, title=NULL, sk=c("G1","b1","mc"), type))
```

### Arguments

numeric vector or an object of class "lm" (i.e. lm, aov, glm, gam, ...).
 title the title at the top of the results. Default is "Normality Tests".
 type of skewness used in D'Agostino skewness test. Can be "G1","b1" or "mc". Read details.
 type type of residuals which should be used. See details.

#### **Details**

This function can be used on objects having "lm" in their class description. For example, class(aov.model) gives "aov" "lm" and class(glm.model) gives "glm" "lm". The type of residuals can be defined. It generally includes c("working", "response", "deviance", "pearson", "partial").

D'Agostino-Pearson's test is more appropriate for analysing a vector with duplicate values in it. The more duplicate values in a vector, the more Shapiro-Wilk will be far from correctly testing the H0 hypothesis.

Given samples from a population, the equation for the sample skewness g1 is a biased estimator of the population skewness. The use of G1 or b1 is advisable. For large samples, the various skewness estimates yield similar results. For small normal distributed samples, b1 is less biased than G1. However, for small non-normal distributed samples, G1 is less biased than b1. These two skewness estimate can be sensitive to outliers in the data (contaminated data). Therefore, the medcouple mc is also an option in type. It has a good performance on uncontaminated data and is robust on contaminated data. For more information on medcouple, please read references and/or type mc (robustbase::mc).

Here, d'Agostino skewness test is based on mc with default settings:

```
g1 = m3/m2(3/2).
where m3 is the sample third central moment, and m2 is the sample variance.
This is the typical definition used in many older textbooks.
```

```
G1 = g1 * [k3/(k2^{(3/2)})] = g1 * [sqrtn(n-1)/(n-2)].
```

where k3 is the unique symmetric unbiased estimator of the third cumulant and k2 is the symmetric unbiased estimator of the second cumulant.

Used in SAS and SPSS.

```
b1 = m3/s^3 = g1((n-1)/n)(3/2).
Used in MINITAB and BMDP.
```

More will be added to this section especially for Anscombe-Glynn Kurtosis test.

norm.test 35

#### Value

A list is returned with the following two components

D	Lilliefor results
W'	Shapiro-Francia results
W	Shapiro-Wilk results
Zb1	D'Agostino Skewness results
Zb2	Anscombe-Glynn Kurtosis results
Chi^2	D'Agostino Pearson results

### Author(s)

Benoit Bruneau

### References

- D. N. Joanes and C. A. Gill (1998), Comparing measures of sample skewness and kurtosis. *The Statistician*, **47**, 183–189.
- G. Brys, M. Hubert and A. Struyf (2003), A Comparison of Some NewMeasures of Skewness. in *Developments in Robust Statistics* **ICORS 2001**, eds. R. Dutter, P. Filzmoser, U. Gather, and P.J. Rousseeuw, Heidelberg: Springer-Verlag, 98–113
- G. Brys, M. Hubert and A. Struyf (2004), A Robust Measure of Skewness; JCGS 13 (4), 996–1017.

### See Also

```
residuals, residuals.lm, residuals.glm, and residuals.gam
```

```
x <- rnorm(300, 50, 10)
y <- 5*(x +10*(rnorm(300,1,2)))

norm.test(x)  ## mc skewness
norm.test(x, type="G1") ## G1 skewness
norm.test(x, type="b1") ## b1 skewness

mod <- lm(y~x)
norm.test(mod)</pre>
```

P.adjust

Adjust P-values for Multiple Comparisons

#### Description

Given a set of p-values, returns p-values adjusted using one of several methods. This is a modified version of p.ajust from stats. It now includes "sidak" correction.

#### Usage

#### Arguments

p vector of p-values (possibly with NAs).

method correction method

n number of pvalues considered for correction; only set this (to non-default)

when you know what you are doing! See details

#### **Details**

The adjustment methods include the Bonferroni correction ("bonferroni") in which the p-values are multiplied by the number of comparisons. Less conservative corrections are also included by Holm (1979) ("holm"), Hochberg (1988) ("hochberg"), Hommel (1988) ("hommel"), Benjamini & Hochberg (1995) ("BH"), and Benjamini & Yekutieli (2001) ("BY"), respectively. A pass-through option ("none") is also included. The P.adjust.methods vector containes the set of correction methods for the benefit of methods that need to have the method as an option and pass it on to P.adjust.

The first five methods are designed to give strong control of the family wise error rate. There seems no reason to use the unmodified Bonferroni correction because it is dominated by Holm's method, which is also valid under arbitrary assumptions.

Hochberg's and Hommel's methods are valid when the hypothesis tests are independent or when they are non-negatively associated (Sarkar, 1998; Sarkar and Chang, 1997). Hommel's method is more powerful than Hochberg's, but the difference is usually small and the Hochberg p-values are faster to compute.

The "BH" and "BY" method of Benjamini, Hochberg, and Yekutieli control the false discovery rate, the expected proportion of false discoveries amongst the rejected hypotheses. The false discovery rate is a less stringent condition than the family wise error rate, so these methods are more powerful than the others.

When making multiple comparisons, n can be used to identify the number of comparisons that are actually used. Correction is then done on the full output matrix when only some of the comparisons are meaningfull or chosen for hypothesis testing. This can be done with the "bonferroni" and "sidak" correction. If other methods are used, exclude the unwanted p.values before applying correction. Unless you know what you are doing, DO NOT modify n if all comparisons are used. Most of the time n should be equal to length(p).

P.adjust 37

Note that you can set n larger than length(p) which means the unobserved p-values are assumed to be greater than all the observed p for "bonferroni" and "holm" methods and equal to 1 for the other methods.

#### Value

A vector of corrected p-values (same length as p).

#### References

Benjamini, Y., and Hochberg, Y. (1995). Controlling the false discovery rate: a practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society Series* B, **57**, 289–300.

Benjamini, Y., and Yekutieli, D. (2001). The control of the false discovery rate in multiple testing under dependency. *Annals of Statistics* **29**, 1165–1188.

Holm, S. (1979). A simple sequentially rejective multiple test procedure. Scandinavian Journal of Statistics,  $\bf 6$ , 65–70.

Hommel, G. (1988). A stagewise rejective multiple test procedure based on a modified Bonferroni test. *Biometrika*, **75**, 383–386.

Hochberg, Y. (1988). A sharper Bonferroni procedure for multiple tests of significance. *Biometrika*, **75**, 800–803.

Shaffer, J. P. (1995). Multiple hypothesis testing. *Annual Review of Psychology*, **46**, 561–576. (An excellent review of the area.)

Sarkar, S. (1998). Some probability inequalities for ordered MTP2 random variables: a proof of Simes conjecture. *Annals of Statistics*, **26**, 494–504.

Sarkar, S., and Chang, C. K. (1997). Simes' method for multiple hypothesis testing with positively dependent test statistics. *Journal of the American Statistical Association*, **92**, 1601–1608.

Wright, S. P. (1992). Adjusted P-values for simultaneous inference. *Biometrics*, **48**, 1005–1013. (Explains the adjusted P-value approach.)

## See Also

```
pairwise.t.test, mc.long, DTK.test, TukeyHSD and glht
```

```
require(graphics)
set.seed(123)
x <- rnorm(50, mean=c(rep(0,25),rep(3,25)))
p <- 2*pnorm( sort(-abs(x)))

round(p, 3)
round(P.adjust(p), 3)
round(P.adjust(p, "BH"), 3)

## or all of them at once (dropping the "fdr" alias):
P.adjust.M <- P.adjust.methods[P.adjust.methods != "fdr"]
p.adj <- sapply(P.adjust.M, function(meth) P.adjust(p, meth))
round(p.adj, 3)
## or a bit nicer:</pre>
```

P.adjust

pack.list 39

pack.list

 $List\ of\ installed\ packages$ 

## Description

Create a text file containing the list of the packages currently installed in R.

# Usage

```
pack.list(n.names=7)
```

## Arguments

n.names

Number of package names to put per line of the output text file.

## Author(s)

Benoit Bruneau

```
pack.list()
pack.list(5)
```

40 pair.diff

	•	٠	_	_
pair	a	٦	t	t

 $Mean\ differences\ matrix\ and\ their\ associated\ standard\ Errors$ 

## Description

Creates two lower triangle matrix: The mean differences and their standard error.

## Usage

```
pair.diff(y, ...)
## S3 method for class 'formula'
pair.diff(y, data=NULL ...)
## S3 method for class 'lm'
pair.diff( y, ...)
## Default S3 method:
pair.diff( y, group, data=NULL, ...)
```

#### Arguments

У	response variable for the default method, or lm or formula object. If y is a linear-model object or a formula, the variables on the right-hand-side of the model must all be factors and must be completely crossed.
group	for the default method, factor (concatenated factor when multiple factors). See details.
data	${\tt data.frame}$ where the dependant variable and the factor(s) are.
	additional arguments to pass to mean and/or sd.

## **Details**

When group is manually defined, use paste(x,y,z) or interaction(x,y,z) form where "x", "y" and "z" are the factors. There is no restrictions on the number of factors.

This function can be usefull with pairwise.t.test since the matrix created are of the same format.

#### Value

Object of class "list" containing two matrices:

diff.m	Mean differences half matrix
diff.se	Standard error associated with the mean differences half matrix

## See Also

Is included in mc.long for the long format of the results.

pair.diff 41

42 performance

 ${\tt performance}$ 

per formance

## Usage

```
performance(expr, samples = 1, gcFirst = TRUE)
```

# Arguments

expr

samples

gcFirst

# Examples

plot.mat 43

 ${\tt plot.mat}$ 

 $Standard\ plot\ for\ maturity\ ogive$ 

# Usage

```
xxx( data , , , )
```

## Arguments

data

# Author(s)

Benoit Bruneau

44 plot.ypr

plot.ypr	Standard Yield per Recruit plot

## Description

Yield per Recruit and Spawning Stock Biomass per Recruit are plotted with standard reference points.

#### Usage

```
## S3 method for class 'ypr'
plot.ypr(object, main, ylab.ypr, ylab.ssb, xlab, col.ypr, col.ssb, ref, legend)
```

#### Arguments

```
object
                 an object of class "ypr" resulting from a call to ypr.1
                 main title for the graph
main
                 a label for the YPR y axis
ylab.ypr
                 a label for the SSB/R y axis
ylab.ssb
xlab
                 a label for the YPR x axis.
                 the color of the the color of the YPR line.
col.ypr
                 the color of the the color of the SSB/R line.
col.ssb
                 logical; if TRUE, standard reference points are added to the plot.
ref
legend
                 logical; if TRUE, a legend is added in the 'topright' corner of the plot.
```

## Details

More to come.

#### See Also

```
ypr.1
```

QQplot 45

QQplot	QQplot		

## Usage

```
QQplot(dat, quant=TRUE,cex.q=2,norm=T, ...)
```

## Arguments

one of:

a numeric vector,
an object of class "norm" resulting from a call to norm.test

quant logical; T for adding quantiles 75, 50 (median) and 25.
cex.q numeric vector giving the amount by which plotting symbols should be magnified relative to the default
norm logical; T adds a line to a normal quantile-quantile plot.
additional arguments to be passed (see par, qqnorm)

```
x=rnorm(50)
QQplot(x)
norm.x=norm.test(x)
QQplot(norm.x)
```

46 r.colors

r.colors

Pie charts of all R character colors

## Description

Creates a pdf file with pie charts of all the 657 basic character colors of R

## Usage

```
r.colors(file)
```

## Arguments

file

the directory in which the pdf file will be created

#### **Details**

Define the directory in which the file should saved by writing file="C:/temp" for example. If file is not defined, it will be saved in "C:/" on windows and in "home" on Mac.

#### Value

None

## Examples

r.colors()

reject.z 47

reject.z

 $reject\ z$ 

## Usage

```
reject.z(x, index = NULL, threshold = 2)
```

# Arguments

x

 ${\tt index}$ 

threshold

# Examples

48 replace.z

```
\verb"replace.z" replace z"
```

# Usage

```
replace.z(x, index = NULL, threshold = 2)
```

# Arguments

x

index

threshold

# Examples

resid.ortho 49

resid.ortho

 $Orthogonal\ residuals$ 

# Usage

```
xxx( data , , , )
```

## Arguments

data

# Author(s)

Benoit Bruneau

50 rivard

rivard

Rivard Weights Calculation

## Description

This function applies Rivard equations to mid-year weight at age data to adjust values to Jan-1 basis.

## Usage

```
rivard(pds, pred=FALSE, K=2, plus.gr=FALSE)
```

## Arguments

data

#### **Details**

More to come. Will be adding interpolation for spawning season.

```
x=rnorm(30,800,10)
rivard(data.frame("2000"=x,"2001"=x*1.2, "2002"=x*0.8,"2003"=x*0.5))
```

rm.levels 51

rm.levels

 $rm\ factor\ levels$ 

## Usage

rm.levels(factor)

## Arguments

factor

## Examples

52 rollmin

rollmin rollmin

# Usage

# Arguments

x
k
na.pad
align
...

## Examples

roundup 53

 $roundup \hspace{3.1cm} roundup$ 

## Description

The "conventional" rounding of 5 to the higher value

## Usage

```
roundup(x, numdigits = 0)
```

## Arguments

x numeric vector.

digits integer indicating the number of decimal places to be used.

#### **Details**

Rounds a 5 to the next value. Therefore roundup(2.5) is 3. This can be usefull when the rounded values are to be presented in a document (eg. table, graph,...).

When rounded values are used in other calculations, round should be used since it follows the IEC 60559 standard.

## Value

numeric vector.

## See Also

round

# Examples

round(2.5)
roundup(2.5)

54 runmax

runmax runmax

# ${\bf Usage}$

runmax(x, window)

# Arguments

Х

window

# Examples

runmean 55

runmean runmean

# ${\bf Usage}$

runmean(x, window)

# Arguments

Х

window

# Examples

runmin

runmin runmin

# Usage

runmin(x, window)

## Arguments

х

window

# Examples

s.an

s.an

 $Simulations\ for\ YPR\ model$ 

# Usage

```
xxx( data , , , )
```

## Arguments

data

# Author(s)

Benoit Bruneau

58 se

se Standard Error

## Usage

```
se(x, na.rm=T)
```

## Arguments

x an R object (vector, matrix,...)

na.rm a logical value indicating whether NA values should be stripped before

the computation proceeds

#### Details

The standard error of the mean is usually estimated by the sample standard deviation divided by the square root of the sample size.

## Examples

x=rnorm(50)
se(x)

show.North 59

s	how.North	North arrow for a map	

## Description

Draws North arrow on a map

#### Usage

#### Arguments

pos	Position of the arrow. Default is 'topright'. See details.
arrow.col	Arrow color.
arrow.fill	Color inside the head of the arrow. NA for no color.
arrow.lwd	Line width of the arrow.
N.cex	Character size for 'N'.
N.family	Font family of 'N'.

#### **Details**

The position of the north arrow is defined by pos and can either be numeric or character.

If pos is numeric, it is a vector of the form c(x,y) where x and y are fractions of the plotting region. If x and y are not in the range of c(0,1), then the north arrow is drawn ouside the bounds of the plotting region and a warning message is given.

If pos is character, it is one of c('topright', 'topleft', 'bottomright', 'bottomleft').

```
plot(1)
show.North()
show.North(c(0.8,0.9))
show.North(c(1.01,0.9)) ### gives a warning
```

60 sort.vdf

sor	t.	vd	f

Sort Data Frames and Vectors

## Description

Single function enabling data.frame and vector sorting

## Usage

```
sort.vdf(x, by, increasing=TRUE)
```

#### Arguments

x data.frame or vector

by A one-sided formula using + for ascending and - for descending. Sorting

is left to right in the formula. This is for data.frame only.

increasing logical. Should the sort be increasing (TRUE) or decreasing (FALSE)? This

is for sorting vectors only.

#### **Details**

See example.

#### Author(s)

Kevin Wright and modified by Benoit Bruneau

summary.ypr 61

summary.ypr

Summarizing the results of YPR models.

## Description

```
summary method for class "ypr".
```

## Usage

```
## S3 method for class 'ypr'
summary(object)
```

## Arguments

object

an object of class "ypr" resulting from a call to ypr.1.

62 ttest.perm

ttest.perm	Student's t-tests by Permutation	
------------	----------------------------------	--

## Description

Performs two sample t-tests or paired t-test by use of permutation

#### Usage

## Arguments

vec1, vec2 two numeric vectors used for Student's t-test analysis nperm number of permutations (default = 999) one of the following: "two.sided", "less" or "greater". alternative var.equal a logical variable indicating whether to treat the two variances as being equal (TRUE) or not (FALSE). a logical variable indicating whether calculation results are printed (FALSE) silent to the R console or not (TRUE). type one of the following: "i" for independant samples or "p" for paired samples. a logical variable indicating whether to perform the exact test (TRUE) or exact

#### Details

The permutational t-test does not require normality of the distributions of each variable. It is also quite robust to heteroscedasticity.

Use exact=TRUE to perform two sample t-test on all the possible combination. This option can only be used when the sum of the sample sizes  $(n_1 + n_2)$  is smaller than 20. It is recommended to use this option when sample sizes are small. It is not implemented yet in the paired t-test.

nperm can not be higher than the maximum number of combination possible  $(n_{comb})$ .

```
n_{comb} = N!/(n_1!n_2!) where n_comb is the number of possible combinations, N! is factorial(n_1 + n_2), n_1! is factorial(n(vec1)) and n_2! is factorial(n(vec2)).
```

There is more to come in this section. plot(x) 5 [ sup ] 7

not (FALSE).

#### Value

ttest.perm 63

```
x <- rnorm(50,0,1)
y <- runif(50,0,1)*x
toto = ttest.perm(x, y) ##independant samples ttest</pre>
```

64 unload

unload

 $Unload\ packages$ 

# Description

Unloads one or multiple packages.

## Usage

unload(pack)

## Arguments

pack

Character vector specifying which packages to unload.

# Author(s)

Benoit Bruneau

# Examples

library(mgcv)
search()
unload(mgcv)
search()

week.1

week.1 week.1

# Description

Week of the year starting on the first of January (01-53)

# Usage

week.1(x)

## Arguments

Х

# Author(s)

Denis Chabot

## Examples

66 week.num

week.num week.num

## Description

Week of the year as decimal number (00-53) using Sunday or Monday as the first day 1 of the week (and typically with the first Sunday of the year as day 1 of week 1).

## Usage

```
week.num(x, day=c("sunday", "monday"))
```

## Arguments

x A vector of dates.

day Either "sunday" or "monday". Default is "sunday".

#### **Details**

Argument day indicates if the week starts on "sunday" or "monday".

*ypr.l* 67

ypr.1

Length Based Yield Per Recruit

## Description

Length based Yield Per Recruit model is define by fishery selectivity and life history parameters related to length.

## Usage

#### Arguments

fsel.type	a list containing the type of fishery selectivity and the values needed for the function related to the type. See 'Details'.
last.age	last age to be considered in the model
age.step	steps used to generate ages (0:last.age). Default is 1.
vonB	a vector containing c(Linf,K) from von Bertalanffy grotwh curve.
1.start	length at the starting age
LW	a vector containing c(alpha, beta) from length-weight curve.
F.max	maximum value of instantaneous rate of fishing mortality $(F)$ . Default is $2$ .
F.incr.YPR	increment for generating the F values to be used for YPR calculation. Default is 0.0001.
M	instantaneous rate of natural mortality (M). Default is 0.2.
mat	a list containing the type of maturity definition and the values needed for the function related to the type. See 'Details'.
f.MSP	reference point defined as the fraction of maximum spawning potential. Default is 0.4.
riv.calc	a logical value indicating whether to use Rivard weights calculation (TRUE) or not. Default is FALSE.
F.f	fraction of instantaneous rate of fishing mortality (F) before spawning.
M.f	fraction of instantaneous rate of natural mortality $(\mathtt{M})$ before spawning.

#### **Details**

More to come.

## Author(s)

Benoit Bruneau

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