

# Package ‘forensim’

July 18, 2009

**Type** Package

**Title** Statistical tools for the interpretation of forensic DNA mixtures

**Version** 1.1-1

**Date** 2009-01-19

**Author** Hinda Haned <haned@biomserv.univ-lyon1.fr>

**Maintainer** Hinda Haned <haned@biomserv.univ-lyon1.fr>

**Suggests** gdata,gtools,MASS,mvtnorm,genetics,tcltk

**Depends** methods

**Description** Statistical methods and simulation tools for the interpretation of forensic DNA mixtures

**License** (>=2)

**LazyLoad** yes

**Collate** classes\_definitions.R classes\_constructors.R accessors.R simufreqD.R simupopD.R zzz.R  
AuxFunc.R changepop.R PE.R likelihood.R likestim.R mincontri.R Pevd2.R LR.R RMP.R  
A2.simu.R A3.simu.R A4.simu.R mastermix.R

## R topics documented:

forensim-package . . . . .	2
A2.simu . . . . .	2
A3.simu . . . . .	4
A4.simu . . . . .	5
Accessors . . . . .	6
changepop . . . . .	7
Cmn . . . . .	7
comb . . . . .	8
dataL . . . . .	9
findfreq . . . . .	10
findmax . . . . .	11
lik . . . . .	11
lik.loc . . . . .	13
likestim . . . . .	14
likestim.loc . . . . .	15

LR . . . . .	17
mastermix . . . . .	18
mincontri . . . . .	20
naomitab . . . . .	20
nball . . . . .	21
PE . . . . .	22
Pevd2 . . . . .	23
RMP . . . . .	24
simufreqD . . . . .	26
simugeno . . . . .	28
simugeno constructor . . . . .	29
simumix . . . . .	30
simumix constructor . . . . .	31
simupopD . . . . .	32
strusa . . . . .	33
strveneto . . . . .	34
tabfreq . . . . .	35
tabfreq constructor . . . . .	36
Tu . . . . .	37
virtualClasses . . . . .	37
<b>Index</b>	<b>38</b>

---

forensim-package      *The forensim package*

---

## Description

forensim is dedicated to the interpretation of forensic DNA mixtures through statistical methods. It relies on three S4 classes that facilitate the manipulation and the storage of genetic data produced in forensic casework: [tabfreq](#), [simugeno](#) and [simumix](#).

[tabfreq](#) objects are used to store allele frequencies, [simugeno](#) objects are used to store genotypes and [simumix](#) objects are used to store DNA mixtures.

For more information about these classes type 'class ?tabfreq', 'class ?simugeno' and 'class ?simumix'.

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

---

`A2.simu`*A Tcl/Tk graphical user interface for simple DNA mixtures resolution using allele peak heights or areas information when two alleles are observed at a given locus*

---

## Description

The `A2.simu` function launches a Tcl/Tk graphical interface with functionalities devoted to two-person DNA mixtures resolution, when two alleles are observed at a given locus.

## Usage

```
A2.simu()
```

## Details

When two alleles are observed at a given locus in the DNA stain, seven genotype combinations are possible for the two contributors: (AA,AB), (AB,AB), (AA,BB), (AB,AA), (BB,AA), (AB,BB) and (BB,AB), where A and B are the two observed alleles (in ascending order of molecular weight). Having previously obtained an estimation for the mixture proportion, it is possible to reduce the number of possible genotype combinations by keeping those only supported by the observed data. This is achieved by computing the sum of square differences between the expected allelic ratio and the observed allelic ratio, for all possible mixture combinations. The likelihood of peak heights (or areas), given the combination of genotypes, is high if the residuals are low. Genotype combinations are thus selected according to the peak heights with the highest likelihoods.

The `A2.simu()` function launches a dialog window with three buttons:

- Plot simulations: plot of the residuals of each possible genotype combination for varying values of the mixture proportion across the interval [0.1, 0.9]. The observed mixture proportion is also reported on the plot.

- Simulation details: a matrix containing the simulation results. Simulation details and genotype combinations with the lowest residuals can be saved as a text file by clicking the "Save" button. It is also possible to choose specific paths and names for the save files.

- Genotypes filter: a matrix giving the mixture proportion conditional on the genotype combination. This conditional mixture proportion helps filter the most plausible genotypes among the seven possible combinations. The matrix can be saved as a text file by clicking the "Save" button. It is also possible to choose a specific path and a name for the save file.

## Note

- Linux users may have to download the `libtktable` package to their system before using the `A2.simu` function. This is due to the `Tktable` widget, used in `forensim`, which is not (always) downloaded with the Tcl/Tk package.

- For the computational details, please see `forensim` tutorial at <http://forensim.r-forge.r-project.org/misc/forensim-tutorial.pdf>.

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## References

Gill P, Sparkes P, Pinchin R, Clayton, Whitaker J, Buckleton J. Interpreting simple STR mixtures using allele peak areas. *Forensic Sci Int* 1998;91:41-53.

## See Also

[A3.simu](#): the three-allele model, and [A4.simu](#): the four-allele model

## Examples

```
A2.simu()
```

---

A3.simu

*A Tcl/Tk graphical user interface for simple DNA mixtures resolution using allele peak heights or areas when three alleles are observed at a given locus*

---

## Description

The `A3.simu` function launches a Tcl/Tk graphical interface with functionalities devoted to two-person DNA mixtures resolution, when three alleles are observed at a given locus.

## Usage

```
A3.simu()
```

## Details

When three alleles are observed at a given locus in the DNA stain, twelve genotype combinations are possible for the two contributors: (AA,BC), (BB,AC), (CC,AB), (AB,AC), (BC,AC), (AB,BC), (BC,AA), (AC,BB), (AB,CC), (AC,AB), (AC,BC) and (BC,AB) where A, B and C are the three observed alleles (in ascending order of molecular weights). Having previously obtained an estimation for the mixture proportion, it is possible to reduce the number of possible genotype combinations by keeping those only supported by the observed data. This is achieved by computing the sum of square differences between the expected allelic ratio and the observed allelic ratio, for all possible mixture combinations. The likelihood of peak heights (or areas), given the combination of genotypes, is high if the residuals are low. Genotype combinations are thus selected according to the peak heights with the highest likelihoods.

The `A3.simu()` function launches a dialog window with three buttons:

-Plot simulations: plot of the residuals of each possible genotype combination for varying values of the mixture proportion across the interval [0.1, 0.9]. The observed mixture proportion is also reported on the plot.

-Simulation details: a matrix containing the simulation results. Simulation details and genotype combinations with the lowest residuals can be saved as a text file by clicking the "Save" button. It is also possible to choose specific paths and names for the save files.

-Genotypes filter: a matrix giving the mixture proportion conditional on the genotype combination. This conditional mixture proportion helps filter the most plausible genotypes among the twelve possible combinations. The matrix can be saved as a text file by clicking the "Save" button. It is also possible to choose a specific path and a name for the save file.

**Note**

-Linux users may have to download the `libtktable` package to their system before using the `A3.simu` function. This is due to the `Tktable` widget, used in `forensim`, which is not (always) downloaded with the `Tcl/Tk` package.

-For the computational details, please see `forensim` tutorial at <http://forensim.r-forge.r-project.org/misc/forensim-tutorial.pdf>.

**Author(s)**

Hinda Haned <haned@biomserv.univ-lyon1.fr>

**References**

Gill P, Sparkes P, Pinchin R, Clayton, Whitaker J, Buckleton J. Interpreting simple STR mixtures using allele peak areas. *Forensic Sci Int* 1998;91:41-53.

**See Also**

[A2.simu](#): the two-allele model, and [A4.simu](#): the four-allele model

**Examples**

```
A3.simu()
```

---

<code>A4.simu</code>	<i>A Tcl/Tk graphical user interface for simple DNA mixtures resolution using allele peak heights or areas when four alleles are observed at a given locus</i>
----------------------	--

---

**Description**

The `A4.simu` function launches a `Tcl/Tk` graphical interface with functionalities devoted to two-person DNA mixtures resolution, when four alleles are observed at a given locus.

**Usage**

```
A4.simu()
```

**Details**

When four alleles are observed at a given locus in the DNA stain, six genotype combinations are possible for the two contributors: (AB,CD),(AC,BD),(AD,BC),(BC,AD),(BD,AC) and (CD,AB) where A, B, C and D are the four observed alleles (in ascending order of molecular weights). Having previously obtained an estimation for the mixture proportion, it is possible to reduce the number of possible genotype combinations by keeping those only supported by the observed data. This is achieved by computing the sum of square differences between the expected allelic ratio and the observed allelic ratio, for all possible mixture combinations. The likelihood of peak heights (or areas), given the combination of genotypes, is high if the residuals are low. Genotype combinations are thus selected according to the peak heights with the highest likelihoods.

The `A4.simu()` function launches a dialog window with three buttons:

- Plot simulations: plot of the residuals of each possible genotype combination for varying values of the mixture proportion across the interval [0.1, 0.9]. The observed mixture proportion is also reported on the plot.
- Simulation details: a matrix containing the simulation results. Simulation details and genotype combinations with the lowest residuals can be saved as a text file by clicking the "Save" button. It is also possible to choose specific paths and names for the save files.
- Genotypes filter: a matrix giving the mixture proportion conditional on the genotype combination. This conditional mixture proportion helps filter the most plausible genotypes among the six possible combinations. The matrix can be saved as a text file by clicking the "Save" button. It is also possible to choose a specific path and a name for the save file.

### Note

- Linux users may have to download the `libtktable` package to their system before using the `A4.simu` function. This is due to the `Tktable` widget, used in `forensim`, which is not (always) downloaded with the `Tcl/Tk` package.
- For the computational details, please see `forensim` tutorial at <http://forensim.r-forge.r-project.org/misc/forensim-tutorial.pdf>.

### Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

### References

Gill P, Sparkes P, Pinchin R, Clayton, Whitaker J, Buckleton J. Interpreting simple STR mixtures using allele peak areas. *Forensic Sci Int* 1998;91:41-53.

### See Also

[A2.simu](#): the two-allele model, and [A3.simu](#): the three-allele model

### Examples

```
A4.simu()
```

---

Accessors

*Accessors for forensim objects*

---

### Description

Accessors for `forensim` objects: [simugeno](#), [simumix](#) and [tabfreq](#). "\$" and "\$<-" are used to access the slots of an object, they are equivalent to "@" and "@<-".

### Value

A [simugeno](#), a [simumix](#) or a [tabfreq](#) object.

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## Examples

```
data(strusa)
class(strusa)

strusa@pop.names
#equivalent
strusa$pop.names
```

---

changePop

*Function to change population-related information in forensim objects*

---

## Description

The changePop function changes population-related information in [tabfreq](#), [simugeno](#) and [simumix](#) objects

## Usage

```
changePop(obj, oldpop, newpop)
```

## Arguments

obj	a forensim object, either a <a href="#">tabfreq</a> , a <a href="#">simugeno</a> or a <a href="#">simumix</a> object
oldpop	a character vector giving the population names to be changed
newpop	a character vector giving the new population names

## Value

a [forensim](#) object where the slots containing population-related information have been modified

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## Examples

```
data(strveneto)
tab1 <- simugeno(strveneto, n=100)
tab2 <- changePop(tab1, "Veneto", "VENE")
tab1$pop.names
tab2$pop.names
```

---

Cmn	<i>The number of all possible combinations of <math>m</math> elements among <math>n</math> with repetitions</i>
-----	---

---

## Description

The number of all possible combinations of  $m$  elements among  $n$  with repetitions.

## Usage

```
Cmn (m, n)
```

## Arguments

$m$	the $m$ elements to combine among $n$
$n$	the $n$ elements from which to combine $m$ elements with repetitions

## Details

There are  $(n+m-1)/(m!(n-1)!)$  ways to combine  $m$  elements among  $n$  with repetitions.

## Note

Cmn was implemented as an auxiliary function for the [dataL](#) function which computes the likelihood of the observed alleles in a mixed DNA stain conditional on the number of contributors.

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## See Also

[comb](#) for all possible combinations of  $m$  elements among  $n$  with repetitions

## Examples

```
Cmn (2, 3)
comb (2, 3)
```



---

comb	<i>Generate all possible combinations of m elements among n with repetitions</i>
------	--

---

## Description

Generate all possible combinations of m elements among n with repetitions.

## Usage

```
comb(m, n)
```

## Arguments

m	the number of elements to combine
n	the number of elements from which to combine the m elements

## Details

There are  $(n+m-1)/(m!(n-1)!)$  ways to combine m elements among n with repetitions, `combn` generates all these possible combinations.

## Value

A matrix of  $(n+m-1)/(m!(n-1)!)$  rows, and n columns, each row is a possible combination of m elements among n .

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## See Also

[Cmn](#) for the calculation of the number of all possible combinations of m elements among n with repetitions

## Examples

```
#combine 2 objcets among 3 with repetitions
Cmn(2, 3)
comb(2, 3)
```

dataL

*Generic formula of the likelihood of the observed alleles in a mixture conditional on the number of contributors for a specific locus*

## Description

The function `dataL` gives the likelihood of a set of alleles observed at a specific locus conditional on the number of contributors that gave these alleles. Calculation is based upon the frequencies of the observed alleles.

## Usage

```
dataL(x = 1, p, theta = 0)
```

## Arguments

<code>x</code>	an integer giving the number of contributors
<code>p</code>	a numeric vector giving the frequencies of the observed alleles in the mixture
<code>theta</code>	a float in $[0,1]$ . <code>theta</code> is equivalent to Wright's $F_{st}$ . In case of population subdivision, it allows a correction of the allele frequencies in the subpopulation of interest

## Note

`dataL` function has several similarities with the `Pevid.gen` function of the *forensic* package which computes the probability of the DNA evidence, `dataL` implements a particular case of this probability. Please see <http://cran.r-project.org/web/packages/forensic/>

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## References

Haned H, Pontier D, Lobry J R, Pene L, Dufour AB. Estimating the number of contributors to forensic DNA mixtures: does maximizing the likelihood performs better than the maximum allele count? In prep, 2009.

Curran JM, Triggs CM, Buckleton J, Weir BS. Interpreting DNA Mixtures in Structured Populations. *J Forensic Sci* 1999;44(5): 987-995

## See Also

[lik.loc](#) and [lik](#) for calculating the likelihood of a given `simumix` object

## Examples

```
#likelihood of observing two alleles at frequencies 0.1 and 0.01 when the number of
#contributors is 2, in two cases: theta=0 and theta=0.03
dataL(x=2,p=c(0.1,0.01), theta=0)
dataL(x=2,p=c(0.1,0.01), theta=0.03)
```

---

findfreq	<i>Finds the allele frequencies of a mixture from a tabfreq object</i>
----------	--

---

### Description

The `findfreq` function finds the allele frequencies of a mixture stored in a `simumix` object, from a given `tabfreq` object. If the `tabfreq` object contains multiple populations, a reference population from which to extract the frequencies must be specified.

### Usage

```
findfreq(mix, freq, refpop = NULL)
```

### Arguments

<code>mix</code>	a <code>simumix</code> object
<code>freq</code>	a <code>tabfreq</code> object from which to extract the allele frequencies of the mixture
<code>refpop</code>	a factor giving the reference population in <code>tabfreq</code> from which to extract the allele frequencies

### Value

A list giving the allele frequencies for each locus.

### Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

### See Also

[simumix](#)

### Examples

```
data(strusa)
s2<-simumix(simugeno(strusa,n=c(0,2000,0)),ncontri=c(0,2,0))
findfreq(s2,strusa,refpop="Cauc")
```

---

findmax	<i>Function to find the maximum of a vector and its position</i>
---------	--

---

### Description

The `findmax` function finds the maximum of a vector and its position.

### Usage

```
findmax(vec)
```

**Arguments**

`vec` a numeric vector

**Details**

`findmax` finds the maximum value of a vector and its position.

**Value**

A matrix of two rows:

`max` the position of the maximum in `vec`

`maxval` the maximum

**Note**

`findmax` is an auxiliary function for the `dataL` function, used to compute the likelihood of the observed alleles in a mixed DNA stain given the number of contributors.

**Author(s)**

Hinda Haned <haned@biomserv.univ-lyon1.fr>

**Examples**

```
findmax(1:10)
```

---

`lik`

*Likelihood of the observed alleles at different loci in a DNA mixture conditional on the number of contributors to the mixture*

---

**Description**

The `lik` function computes the likelihood of the observed alleles in a forensic DNA mixture, for a set of loci, conditional on the number of contributors to the mixture. The overall likelihood is computed as the product of loci likelihoods.

**Usage**

```
lik(x = 1, mix, freq, refpop = NULL, theta = NULL, loc=NULL)
```

**Arguments**

<code>x</code>	the number of contributors to the DNA mixture, default is 1
<code>mix</code>	a <code>simumix</code> object which contains the mixture to be analyzed
<code>freq</code>	a <code>tabfreq</code> object from which to extract the allele frequencies
<code>refpop</code>	a factor giving the reference population in <code>tabfreq</code> from which to extract the allele frequencies. This argument is used only if <code>freq</code> contains allele frequencies for multiple populations, otherwise it is by default set to <code>NULL</code>
<code>theta</code>	a float from <code>[0,1[</code> giving Wright's $F_{st}$ coefficient. <code>theta</code> accounts for population subdivision while computing the likelihood of the data
<code>loc</code>	loci for which the overall likelihood shall be computed. Default ( <code>NULL</code> ) corresponds to all loci

## Details

`lik` computes the likelihood of the alleles observed at all loci conditional on the number of contributors. This function implements the general formula for the interpretation of DNA mixtures in case of population subdivision (Curran et al, 1999), in the particular case where all contributors are unknown and belong to the same subpopulation.

The likelihood for multiple loci is computed as the product of loci likelihoods.

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## References

Haned H, Pontier D, Lobry J R, Pene L, Dufour AB. Estimating the number of contributors to forensic DNA mixtures: does maximizing the likelihood performs better than the maximum allele count? In prep, 2009.

Curran JM, Triggs CM, Buckleton J, Weir BS. Interpreting DNA Mixtures in Structured Populations. *J Forensic Sci* 1999;44(5): 987-995

## See Also

[lik.loc](#) for the likelihood per locus, [likestim](#) and [likestim.loc](#) for the estimation of the number of contributors to a DNA mixture through likelihood maximization

## Examples

```
data(strusa)
#simulation of 1000 genotypes from the African American allele frequencies
gen<-simugeno(strusa,n=c(1000,0,0))
#3-person mixture
mix3<-simumix(gen,ncontri=c(3,0,0))
sapply(1:3, function(i) lik(x=i,mix3, strusa, refpop="Afri"))
```

---

`lik.loc`

*Likelihood per locus of the observed alleles in a DNA mixture conditional on the number of contributors to the mixture*

---

## Description

The `lik.loc` function computes the likelihood of the observed data in a forensic DNA mixture, for each of the loci involved, conditional on the number of contributors to the mixture.

## Usage

```
lik.loc(x = 1, mix, freq, refpop = NULL, theta = NULL, loc=NULL)
```

## Arguments

<code>x</code>	the number of contributors to the DNA mixture
<code>mix</code>	a <code>simumix</code> object which contains the mixture to be analyzed
<code>freq</code>	a <code>tabfreq</code> object from which to extract the allele frequencies
<code>refpop</code>	a factor giving the reference population in <code>tabfreq</code> from which to extract the allele frequencies
<code>theta</code>	a float from $[0,1[$ giving Wright's $F_{st}$ coefficient. <code>theta</code> accounts for population subdivision while computing the likelihood of the data.
<code>loc</code>	the loci for which the likelihood shall be computed. Default (set to <code>NULL</code> ) corresponds to all loci.

## Details

`lik.loc` computes the likelihood per locus of the observed alleles. This function implements the general formula for the interpretation of DNA mixtures in case of subdivided populations (Curran et al, 1999), in the particular case where all contributors are unknown and belong to the same subpopulation.

The  $F_{st}$  coefficient given in the `theta` argument allows accounting for population subdivision when all contributors belong to the same subpopulation.

## Value

The function `lik.loc` returns a vector, of length the number of loci in `loc`, giving the likelihood of the data for each locus.

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## References

Haned H, Pontier D, Lobry J R, Pene L, Dufour AB. Estimating the number of contributors to forensic DNA mixtures: does maximizing the likelihood performs better than the maximum allele count? In prep, 2009.

Curran JM, Triggs CM, Buckleton J, Weir BS. Interpreting DNA Mixtures in Structured Populations. *J Forensic Sci* 1999;44(5): 987-995

## See Also

[lik](#) for the overall loci likelihood, [likestim](#) and [likestim.loc](#) for the estimation of the number of contributors to a DNA mixture through likelihood maximization

## Examples

```
data(strusa)
#simulation of 1000 genotypes from the Caucasian allele frequencies
gen<-simugeno(strusa,n=c(0,100,0))

#4-person mixture
mix4 <- simumix(gen,ncontri=c(0,4,0))
```

```
lik.loc(x=2,mix4, strusa, reffpop="Cauc")
lik.loc(x=2,mix4, strusa, reffpop="Afri")
#You may also want to try:
#likestim(mix4,strusa,reffpop="Cauc")
```

---

likestim	<i>Maximum likelihood estimation of the number of contributors to a forensic DNA mixture for a set of loci</i>
----------	--

---

## Description

The `likestim` function gives multiloci estimation of the number of contributors to a forensic DNA mixture using likelihood maximization.

## Usage

```
likestim(mix, freq, reffpop = NULL, theta = NULL, loc=NULL)
```

## Arguments

<code>mix</code>	a <code>simumix</code> object
<code>freq</code>	a <code>tabfreq</code> object containing the allele frequencies to use for the calculation
<code>reffpop</code>	the reference population from which to extract the allele frequencies used in the likelihood calculation. If <code>tabfreq</code> contains more than one population, <code>reffpop</code> must be specified, otherwise, <code>reffpop</code> is set to default (NULL).
<code>theta</code>	a float from [0,1[ giving Wright's $F_{st}$ coefficient. <code>theta</code> accounts for population subdivision while computing the likelihood of the data.
<code>loc</code>	loci to be considered in the estimation. Default (set to NULL) corresponds to all loci.

## Details

The number of contributors which maximizes the likelihood of the data observed in the mixture is searched in the discrete interval [1,6]. In most cases this interval is a plausible range for the number of contributors.

## Value

A matrix, the first row, `max`, gives the maximum likelihood estimation of the number of contributors, the second row gives the corresponding likelihood value `maxvalue`.

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## References

Haned H, Pontier D, Lobry J R, Pene L, Dufour AB. Estimating the number of contributors to forensic DNA mixtures: does maximizing the likelihood performs better than the maximum allele count? In prep, 2009.

Egeland T, Dalen I, Mostad PF. Estimating the number of contributors to a DNA profile. *Int J Legal Med* 2003, 117: 271-275

Curran JM, Triggs CM, Buckleton J, Weir BS. Interpreting DNA Mixtures in Structured Populations. *J Forensic Sci* 1999, 44(5): 987-995

## See Also

[likestim.loc](#) for maximum of likelihood estimations per locus

## Examples

```
data(strusa)
#simulation of 1000 genotypes from the Hispanic allele frequencies
gen<-simugeno(strusa,n=c(0,0,100))
#4-person mixture
mix4 <- simumix(gen,ncontri=c(0,0,4))
likestim(mix4,strusa,refpop="Hisp")
```

---

likestim.loc	<i>Maximum likelihood estimation per locus of the number of contributors to forensic DNA mixtures.</i>
--------------	--

---

## Description

The `likestim.loc` function returns the estimation of the number of contributors, at each locus, obtained by maximizing the likelihood.

## Usage

```
likestim.loc(mix, freq, refpop = NULL, theta = NULL, loc = NULL)
```

## Arguments

<code>mix</code>	a <code>simumix</code> object
<code>freq</code>	a <code>tabfreq</code> object containing the allele frequencies to use for the calculation
<code>refpop</code>	the reference population from which to extract the allele frequencies used in the likelihood calculation. Default set to <code>NULL</code> , if <code>tabfreq</code> contains more than one population, <code>refpop</code> must be specified
<code>theta</code>	a float from <code>[0,1[</code> giving Wright's $F_{st}$ coefficient. <code>theta</code> accounts for population subdivision while computing the likelihood of the data.
<code>loc</code>	loci to be considered in the estimation. Default (set to <code>NULL</code> ) corresponds to all loci.



## Details

The number of contributors which maximizes the likelihood of the data observed in the mixture is searched in the discrete interval [1,6]. In most cases this interval is a plausible range for the number of contributors.

## Value

A matrix of dimension  $2 \times \text{loc}$ . The first row, `max`, gives the maximum likelihood estimation of the number of contributors for each locus in column. The second row, `maxvalue`, gives the corresponding likelihood value.

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## References

Haned H, Pontier D, Lobry J R, Pene L, Dufour AB. Estimating the number of contributors to forensic DNA mixtures: does maximizing the likelihood performs better than the maximum allele count? In prep, 2009.

Egeland T , Dalen I, Mostad PF. Estimating the number of contributors to a DNA profile. *Int J Legal Med* 2003, 117: 271-275

Curran, JM , Triggs CM, Buckleton J , Weir BS. Interpreting DNA Mixtures in Structured Populations. *J Forensic Sci* 1999, 44(5): 987-995

## See Also

[likestim](#) for multiloci estimations

## Examples

```
data(strusa)
#simulation of 1000 genotypes from the Hispanic allele frequencies
gen<-simugeno(strusa,n=c(0,0,100))
#4-person mixture
mix4 <- simumix(gen,ncontri=c(0,0,4))
likestim.loc(mix4,strusa,refpop="Hisp")
```

## Description

The `LR` function calculates the likelihood ratio for a DNA evidence, when two competing hypotheses  $H_d$  and  $H_p$ , respectively the defence and the prosecution hypotheses, are weighted about the origin of the DNA evidence. The evidence can either be a simple or a mixed stain.

**Usage**

```
LR(stain, freq, xp=0, xd=0, Tp=NULL, Vp=NULL, Td=NULL, Vd=NULL, theta=0)
```

**Arguments**

stain	a vector giving the set of (distinct) alleles present in the DNA stain
freq	vector of the corresponding allele frequencies in the global population
xp	the number of unknown contributors to the stain under the prosecuting hypotheses $H_p$ . Default is 0.
xd	the number of unknown contributors to the stain under the defence hypotheses $H_d$ . Default is 0.
Tp	a vector of strings where each string contains two alleles separated by '/', corresponding to one known contributor under the prosecution hypotheses $H_p$ . The length of the vector equals the number of known contributors. Default is NULL.
Vp	a vector of strings where each string contains two alleles separated by '/', corresponding to one known non-contributor under the prosecution hypotheses $H_p$ . The length of the vector equals the number of known non-contributors. Default is NULL.
Td	a vector of strings where each string contains two alleles separated by '/', corresponding to one known contributor under the defence hypotheses $H_d$ . The length of the vector equals the number of known contributors. Default is NULL.
Vd	a vector of strings where each string contains two alleles separated by '/', corresponding to one known non-contributor under the defence hypotheses $H_d$ . The length of the vector equals the number of known non-contributors. Default is NULL.
theta	a float in [0,1]. <code>theta</code> is equivalent to Wright's $F_{st}$ . In case of population subdivision, it allows a correction of the allele frequencies in the subpopulation of interest

**Details**

LR is the implementation of the general formula of Curran et al (1999) for the evaluation of forensic DNA mixtures through likelihood ratios. The likelihood ratio is computed as a ratio of two probabilities of the DNA evidence,  $E$ , conditional on the evaluated hypotheses:

$$LR = \frac{P(E|H_p)}{P(E|H_d)},$$

where  $H_p$  denotes the prosecution hypotheses and  $H_d$  the defence hypotheses.

In case of population subdivision, contributors to the DNA stain are considered to come from the same subpopulation. Allele dependencies within subpopulations are accounted for through Wright's  $F_{st}$  coefficient, denoted here  $\theta$ .

**Note**

Please note that the LR function is based on functions initially implemented in the forensic package by Miriam Marusiakova <http://cran.r-project.org/web/packages/forensic/>

**Author(s)**

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## References

Curran JM, Triggs CM, Buckleton J, Weir BS. Interpreting DNA Mixtures in Structured Populations. *J Forensic Sci* 1999;44(5): 987-995

## See Also

the exclusion probability [PE](#).

## Examples

```
# A rape case in Hong Kong (Hu and Fung, Int J Legal Med 2003)
# The stain shows alleles 14, 15, 17 and 18 at locus D3S1358.
stain =c(14,15,17,18)
# suspect's profile: "14/17"
suspect<-"14/17"
# victim's profile: "15/18"
victim<-"15/18"
# corresponding allele frequencies
freq<-c(0.033,0.331,0.239,0.056)

# Prosecution hypotheses: Contributors were the victim and the suspect
# defence hypotheses: Contributors were the victim and 1 unknown contributor
# Likelihood ratios for DNA evidence for different alternatives:
LR(stain,freq,xp=0,Tp=c(victim,suspect),Vp=NULL,Td=victim,Vd=suspect,xd=1)
```

---

mastermix

*A Tcl/Tk graphical user interface for simple DNA mixtures resolution using allele peak heights/ or areas information*

---

## Description

The `mastermix` function launches a Tcl/Tk graphical user interface dedicated to the resolution of two-person DNA mixtures using allele peak heights/ or areas information. `mastermix` is the implementation of a method developed by Gill et al (see the references section), and previously programmed into an Excel macro by Dr. Peter Gill.

## Usage

```
mastermix()
```

## Details

`mastermix` is a Tcl/Tk graphical user interface implementing a method developed by Gill et al (1998) for simple mixtures resolution, using allele peak heights or areas information.

This method searches through simulation the most likely combination(s) of the contributors' genotypes. Having previously obtained an estimation for the mixture proportion, it is possible to reduce the number of possible genotype combinations by keeping only those supported by the observed data. This is achieved by computing the sum of square differences between the expected allelic ratio and the observed allelic ratio, for all possible mixture combinations. The likelihood of peak heights (or areas), conditional on the combination of genotypes, is high if the residuals are low.

Genotype combinations are thus selected according to the peak heights with the highest (conditioned) likelihoods.

mastermix offers a graphical representation of the simulation for three models:

- The two allele model: at a given locus, two alleles are observed in the DNA stain.
- The three allele model: at a given locus, three alleles are observed in the DNA stain.
- The four allele model: at a given locus, four alleles are observed in the DNA stain.

A left-click on each button launches a simulation dialog window for the corresponding model, while a right-click opens the corresponding help page.

### Note

- Each implemented model can either be launched using the mastermix interface, or the `A2.simu`, `A3.simu` and `A4.simu` functions, depending on the considered model.
- For the computational details, please see forensim tutorial at <http://forensim.r-forge.r-project.org/misc/forensim-tutorial.pdf>.

### Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

### References

Gill P, Sparkes P, Pinchin R, Clayton, Whitaker J, Buckleton J. Interpreting simple STR mixtures using allele peak areas. *Forensic Sci Int* 1998;91:41-5.

### See Also

`A2.simu`, `A3.simu` and `A4.simu`

### Examples

```
mastermix()
```

---

mincontri	<i>Minimum number of contributors required to explain a forensic DNA mixture</i>
-----------	--

---

### Description

mincontri gives the minimum number of contributors required to explain a forensic DNA mixture. This method is also known as the maximum allele count as it relies on the maximum number of alleles showed through all available loci

### Usage

```
mincontri(mix, loc = NULL)
```

**Arguments**

<code>mix</code>	a <code>simumix</code> object
<code>loc</code>	the loci to consider for the calculation of the minimum of contributors, default (NULL) corresponds to all loci

**Author(s)**

Hinda Haned <haned@biomserv.univ-lyon1.fr>

**See Also**

`likestim` for the estimation of the number of contributors through likelihood maximization

**Examples**

```
data(strusa)
#simulation of 1000 genotypes from the African American allele frequencies
gen<-simugeno(strusa,n=c(1000,0,0))
#5-person mixture
mix5<-simumix(gen,ncontri=c(5,0,0))
#compare
likestim(mix5, strusa, refpop="Afri")
mincontri(mix5)
```

---

naomitab

*Handling of missing values in a data frame*


---

**Description**

`naomitab` handles missing values (NA) in a data frame: it returns a list of the columns where NAs have been removed.

**Usage**

```
naomitab(tab)
```

**Arguments**

<code>tab</code>	a data frame
------------------	--------------

**Value**

Returns a list of length the number of columns in `tab` where each component is a column of `tab`, and the values are the corresponding rows where NAs have been removed.

**Note**

This function was designed to handle missing values in data frames in the format of the Journal of Forensic Sciences for population genetic data: allele names are given in the first column, and frequencies for a given allele are read in rows for different loci. When a given allele is not observed, the value is coded NA (originally coded "-" in the journal).

**Author(s)**

Hinda Haned <haned@biomserv.univ-lyon1.fr>

**See Also**

[tabfreq](#)

**Examples**

```
data(Tu)
naomitab(Tu)
```

---

nball	<i>Number of alleles in a mixture</i>
-------	---------------------------------------

---

**Description**

nball gives the number of alleles of a simumix object.

**Usage**

```
nball(mix, byloc = FALSE)
```

**Arguments**

mix	a simumix object
byloc	a logical indicating whether the number of alleles must be calculated by locus or for all loci (default)

**Author(s)**

Hinda Haned <haned@biomserv.univ-lyon1.fr>

**See Also**

[simumix](#)

**Examples**

```
data(strusa)
#simulating 100 genotypes with allele frequencies from the African American population
gaa<-simugeno(strusa,n=c(100,0,0))
#simulating a 4-person mixture
maa4<-simumix(gaa,ncontri=c(4,0,0))
nball(maa4,byloc=TRUE)
```

PE

*The random man exclusion probability***Description**

Computes the random man exclusion probability of a mixture stored in a `simumix` object

**Usage**

```
PE(mix, freq, refpop = NULL, theta = 0, byloc = FALSE)
```

**Arguments**

<code>mix</code>	a <code>simumix</code> object
<code>freq</code>	a <code>tabfreq</code> object giving the allele frequencies from which to compute the exclusion probability
<code>refpop</code>	character giving the reference population, used only if <code>freq</code> contains allele frequencies for multiple populations
<code>theta</code>	a float from $[0,1[$ giving Wright's $F_{st}$ coefficient. <code>theta</code> accounts for population subdivision while computing the likelihood of the data.
<code>byloc</code>	logical, if TRUE, than the exclusion probability is computed per locus, if FALSE (default), the calculations are done for all loci simultaneously

**Details**

PE gives the exclusion probability at a locus, or at several loci when conditions for Hardy Weinberg are met. If this condition is not met in the population, than a value for `theta` must be supplied to take into account dependencies between alleles. The formula of the exclusion probability that allows taking into account departure from Hardy Weinberg proportions due to population subdivision was provided by Bruce Weir, please see the references section.

**Author(s)**

Hinda Haned <haned@biomserv.univ-lyon1.fr>

**References**

Clayton T, Buckleton JS. Mixtures. In: Buckleton JS, Triggs CM, Walsh SJ, editors. Forensic DNA Interpretation. CRC Press 2005;217-74

**Examples**

```
data(strusa)
geno1<-simugeno(strusa,n=c(0,0,100))
mix2 <-simumix(geno1,ncontri=c(0,0,2))
PE(mix2,strusa,"Hisp",byloc=TRUE)
```

PevId2

*Conditional profile probabilities***Description**

Calculates the probability of observing a set of DNA profiles conditional on a given hypotheses specifying who were the contributors to the observed profiles. All the individuals involved in the analyzed case are assumed to come from the same subpopulation with a given coancestry coefficient.

**Usage**

```
PevId2(stain, freq, x, T = NULL, V = NULL, theta = 0)
```

**Arguments**

stain	vector of distinct alleles (from one specific locus) found in the crime sample.
freq	vector of the corresponding allele frequencies in the global population
x	the number of unknown contributors to the mixture
T	object of class <b>genotype</b> (package <b>genetics</b> ), or a vector of strings where each string contains two alleles separated by '/', corresponding to one known contributor. The length of the vector equals the number of known contributors. Default is NULL.
V	object of class <b>genotype</b> (package <b>genetics</b> ), or a vector of strings where each string contains two alleles separated by '/', corresponding to one known non-contributor. The length of the vector equals the number of known non-contributors. Default is NULL.
theta	a float in [0,1[. <i>theta</i> is equivalent to Wright's <i>Fst</i> . In case of population subdivision, it allows a correction of the allele frequencies in the subpopulation of interest

**Note**

Please note that the `PevId2` function is an improved version of the `PevId.gen` function from the forensic package by Miriam Marusiakova (which explains the 2 in the function name). `PevId2` calls external functions in C code.

**Author(s)**

Hinda Haned <haned@biomserv.univ-lyon1.fr>

**References**

Curran JM, Triggs CM, Buckleton J, Weir BS. Interpreting DNA Mixtures in Structured Populations. *J Forensic Sci* 1999;44(5): 987-995

**See Also**

[LR](#)



## Examples

```
# A rape case in Hong Kong (Hu and Fung, Int J Legal Med 2003)
# The stain shows alleles 14, 15, 17 and 18 at locus D3S1358.
stain=c(14,15,17,18)
# suspect's profile: "14/17"
suspect<-"14/17"
# victim's profile: "15/18"
victim<-"15/18"
# corresponding allele frequencies
freq<-c(0.033,0.331,0.239,0.056)

# Prosecution proposition: Contributors were the victim and the suspect
# defence proposition: Contributors were the victim and 1 unknown contributor
# from the same subpopulation as the victim
# Evaluation of the defence proposition, in case of independence between alleles
Pevd2(stain, freq, x=1, T = victim)

# note that if theta=0, the suspect's profile plays no role in the calculation
# and the same result is obtained
Pevd2(stain, freq, x=1, T = victim, V = suspect)
# In case of allele dependencies, measured by theta=0.03
Pevd2(stain, freq, x=1, T = victim, V = suspect, theta = 0.03)
```

---

RMP

---

*The Random Match Probability of DNA evidence (RMP)*


---

## Description

RMP computes the random match probability of DNA evidence given in a matrix (or data frame) or in a text file. Several situations are handled: the suspect and an unknown offender are unrelated, or are members of the same subpopulation with a given coancestry coefficient  $\theta$ , or are close relatives. For the latter case, the relationship is described by the kinship coefficients.

## Usage

```
RMP(suspect=NULL, filename=NULL, freq, k=c(1,0,0), theta=0, reipop=NULL)
```

## Arguments

suspect	a matrix or a data frame of dimension $L \times 2$ , $L$ being the number of loci involved in the DNA evidence. The first column gives the loci names, and the second column gives the suspect's genotype at each locus. A genotype is coded as a character where each string contains two alleles separated by '/'. The DNA evidence can also be given in a text file, see argument <code>filename</code> .
filename	the file name from which the input data should be read. Data must be a matrix of dimension $L \times 2$ , $L$ being the number of loci involved in the DNA evidence. The first column gives the loci names, and the second column gives the suspect's genotype at each locus. A genotype is coded as a character where each string contains two alleles separated by '/'.
freq	a <code>tabfreq</code> object giving the allele frequencies

<code>k</code>	vector of kinship coefficients ( $k_0, k_1, k_2$ ), where $k_i$ is the probability that two people (the suspect and an unknown offender) will share $i$ alleles identical by descent, $i = 0, 1, 2$ .
<code>theta</code>	a float in $[0,1[$ . <code>theta</code> is equivalent to Wright's $F_{st}$ . In case of population subdivision, it allows a correction of the allele frequencies in the subpopulation of interest
<code>refpop</code>	the reference population in <code>freq</code> from which to extract the allele frequencies from the RMP calculation. This argument is obligatory only if <code>freq</code> contains allele frequencies from several populations

### Details

The match probability is derived from Balding and Nichols (1994) and is computed as:

$$k_2 + k_1 Z_1 + k_0 Z_2$$

where  $k_0, k_1, k_2$  are the kinship coefficients,

$Z_1$  is the match probability when the suspect and the unknown offender share one allele identical-by-descent.

$Z_2$  is the match probability in the unrelated case, when the suspect and the unknown offender share 0 allele identical-by-descent.

In the homozygous case, with the allele frequency  $p_i$ :

$$Z_1 = \frac{2\theta + (1 - \theta)p_i}{1 + \theta}$$

$$Z_2 = \frac{[2\theta + (1 - \theta)p_i][3\theta + (1 - \theta)p_i]}{(1 + \theta)(1 + 2\theta)}$$

In the heterozygous case, with allele frequencies  $p_i$  and  $p_j$ :

$$Z_1 = \frac{2\theta + (1 - \theta)(p_i + p_j)}{2(1 + \theta)}$$

$$Z_2 = \frac{2[\theta + (1 - \theta)p_i][\theta + (1 - \theta)p_j]}{(1 + \theta)(1 + 2\theta)}$$

$\theta$  is Wright's  $F_{st}$  coefficient, usually called the coancestry coefficient in forensic studies. Main effects of allele dependencies between loci in the suspect's subpopulation are taken into account through the coancestry coefficient, hence, the match probability at all loci is, to a close approximation, the product of single-locus probabilities.

### Value

RMP returns a list with the following components:

<code>RMP.loc</code>	single-locus match probabilities
<code>RMP</code>	multiloci match probability (product of single-locus match probabilities)

### Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## References

Balding DJ, Nichols RA. DNA profile match probability calculation: How to allow for population stratification, relatedness, database selection and single bands. *Forensic Sci I* 1994;64:125-140.

## See Also

[LR](#) for the evaluation of DNA evidence through likelihood ratio

## Examples

```
# random match probability
# data input

data <- matrix(c("CSF1PO", "FGA", "TH01", "TPOX", "VWA", "D3S1358", "D5S818",
"D7S820", "D8S1179", "D13S317", "D16S539", "D18S51", "D21S11", "D2S1338", "D19S433",
"12/11", "22/19", "6/7", "10/8", "17/18", "18/17", "12/12", "8/8", "13/13", "11/11",
"12/10", "14/15", "33.2/32.2", "23/22", "14/14"), nc=2)
colnames(data) <- c('locus', 'genotype')
#15-locus genotype
data
#allele frequencies are taken from the strusa data set

data(strusa)

RMP(suspect=data, freq=strusa, reffpop="Cauc")

# using a preexisting file from the forensim package
RMP(filename=system.file("files/exprofile.txt", package = "forensim"),
freq=strusa, reffpop="Cauc")
```

---

simufreqD

*Function to simulate allele frequencies for independent loci from a Dirichlet model*

---

## Description

The `simufreqD` function simulate single population allele frequencies for independent loci. Allele frequencies are generated as random deviates from a Dirichlet distribution, the parameters of which control the mean and the variance of the simulated allele frequencies.

## Usage

```
simufreqD(nloc = 1, nal = 2, alpha = 1)
```

## Arguments

<code>nloc</code>	the number of loci to simulate
<code>nal</code>	the numbers of alleles per locus. Either an integer, if the loci have the same number of alleles, or an integer vector, if the number of alleles differ between loci

**alpha** the parameter used to simulate allele frequencies from the Dirichlet distribution. If the `nloc` loci have the same allele number, `alpha` can either be the same for all alleles (default is one: uniform distribution), in this case `alpha` is an integer, or `alpha` can be different between alleles at a given locus, in this case, `alpha` is a matrix of dimension `nal` x `nloc`.

When the number of alleles differ between loci, `alpha` can either be the same or differ between alleles at a given locus. In the first case `alpha` is a vector of length `nloc`, in the second case, `alpha` is a matrix of dimensions `nal` x `nloc` where NAs are introduced for alleles not seen at a given locus.

### Details

Allele frequencies for independent loci are simulated using a Dirichlet distribution with parameter `alpha`. At a given locus `L` with `n` alleles, the allele frequencies are modeled as a vector of random variables  $p=(p_1, \dots, p_n)$ , following a Dirichlet distribution with parameters:  $\alpha = (\alpha_1, \dots, \alpha_n)$  where  $p_1 + \dots + p_n = 1$  and  $\alpha_1, \dots, \alpha_n > 0$ .

### Value

A matrix containing the simulated allele frequencies. The data is presented in the format of the Journal of Forensic Sciences for genetic data: allele names are given in the first column, and frequencies for a given allele are read in rows for the different markers in columns. When an allele is not observed for a given locus, the value is coded NA (instead of "-" in the original format).

### Note

The code used here for the generation of random Dirichlet deviates was previously implemented in the `gttools` library.

### Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

### References

Johnson NL, Kotz S, Balakrishnan N. Continuous Univariate Distributions, vol 2. John Wiley & Sons, 1995.

Wright S. The genetical structure of populations. Ann Eugen 1951;15:323-354.

### See Also

[simupopD](#)

### Examples

```
#simulate alleles frequencies for 5 markers with respectively 2, 3, 4, 5, and 6 alleles
simufreqD(nloc=5,na=c(2,3,4,5,6) , alpha=1)
```

---

simugeno*forensim class for simulated genotypes*

---

## Description

The S4 `simugeno` class is used to store existing or simulated genotypes.

## Slots

**tab.freq:** a list giving allele frequencies for each locus. If there are several populations, `tab.freq` gives allele frequencies in each population

**nind:** integer vector giving the number of individuals. If there are several populations, `nind` gives the numbers of individuals per population

**pop.names:** factor of populations names

**popind:** factor giving the population of each individual

**which.loc:** character vector giving the locus names

**tab.geno:** matrix giving the genotypes (in rows) for each locus (in columns). The genotype of a homozygous individual carrying the allele "12" is coded "12/12". A heterozygous individual carrying alleles "12" and "13" is coded "12/13" or "13/12".

**indID:** character vector giving the individuals ID

## Methods

**names** `signature(x = "simugeno")`: gives the names of the attributes of a `simugeno` object

**show** `signature(object = "simugeno")`: shows a `simugeno` object

**print** `signature(object = "simugeno")`: prints a `simugeno` object

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## See Also

[as.simugeno](#) for the `simugeno` class constructor, [is.simugeno](#), [simumix](#) and [tabfreq](#)

## Examples

```
showClass("simugeno")
```

---

```
simugeno constructor
      simugeno constructor
```

---

## Description

Constructor for [simugeno](#) objects.

The function `simugeno` creates a [simugeno](#) object from a [tabfreq](#) object.

The function `as.simugeno` is an alias for `simugeno` function.

`is.simugeno` tests if an object is a valid `simugeno` object.

Note: to get the manpage about [simugeno](#), please type `'class ? simugeno'`.

## Usage

```
simugeno(tab, which.loc=NULL, n=1)
as.simugeno(tab, which.loc=NULL, n=1)
is.simugeno(x)
```

## Arguments

<code>tab</code>	a <a href="#">tabfreq</a> object created with constructor <a href="#">tabfreq</a>
<code>which.loc</code>	a character vector giving the chosen loci for the genotypes simulation. The default is set to <code>NULL</code> , which corresponds to all the loci of the <a href="#">tabfreq</a> object given in argument
<code>n</code>	integer vector giving the number of individuals. If there are several populations, <code>n</code> gives the numbers of individuals to simulate per population. For a single population, default is 1.
<code>x</code>	an object

## Value

For `simugeno` and `as.simugeno`, a `simugeno` object. For `is.simugeno`, a logical.

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## See Also

["simugeno"](#), and [tabfreq](#) for creating a `tabfreq` object from a data file.

## Examples

```
data(Tu)
tab<-tabfreq(Tu)
#simulation of 3 individual genotypes for the STR marker FGA
geno1 <- simugeno(tab, which.loc='FGA', n =1000)
geno1@tab.geno
```

---

simumix*forensim class for DNA mixtures*

---

## Description

The S4 `simumix` class is used to store DNA mixtures of individual genotypes along with informations about the individuals populations and the loci used to simulate the genotypes.

## Slots

**ncontri:** integer vector giving the number of contributors to the DNA mixture. If there are several populations, `ncontri` gives the number of contributors per population

**mix.prof:** matrix giving the contributors genotypes (in rows) for each locus (in columns). The genotype of a homozygous individual carrying the allele "12" is coded "12/12". A heterozygous individual carrying alleles "12" and "13" is coded "12/13" or "13/12".

**mix.all:** list giving the alleles present in the mixture for each locus

**which.loc:** character vector giving the locus names

**popinfo:** factor giving the population of each contributor

## Methods

**names** `signature(x = "simumix")`: gives the names of the attributes of a `simumix` object

**show** `signature(object = "simumix")`: shows a `simumix` object

**print** `signature(object = "simumix")`: prints a `simumix` object

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## See Also

`simugeno`, `as.simumix`, `is.simumix`, `simugeno` and `tabfreq`

## Examples

```
showClass("simumix")
data(strusa)
```

---

simumix constructor

*simumix constructor*


---

## Description

Constructor for [simumix](#) objects.

The function `simumix` creates a [simumix](#) object from a [tabfreq](#) object.

The function `as.simumix` is an alias for `simumix` function.

`is.simumix` tests if an object is a valid `simumix` object.

Note: to get the manpage about [simumix](#), please type `'class ? simumix'`.

## Usage

```
simumix(tab, which.loc=NULL, ncontri=1)
as.simumix(tab, which.loc=NULL, ncontri=1)
is.simumix(x)
```

## Arguments

<code>tab</code>	a <code>simugeno</code> object created with constructor <code>simugeno</code>
<code>which.loc</code>	a character vector giving the chosen loci for the genotypes simulation. The default is set to <code>NULL</code> , which corresponds to all the loci of the <code>simugeno</code> object given in argument
<code>ncontri</code>	integer vector giving the number of individuals. If there are several populations, <code>ncontri</code> gives the numbers of individuals to simulate per population. Default is one.
<code>x</code>	an object

## Value

For `simumix` and `as.simumix`, a `simumix` object. For `is.simumix`, a logical.

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## See Also

"[simumix](#)", [simugeno](#) for creating a `simugeno` object.

## Examples

```
data(Tu)
tab<-simugeno(tabfreq(Tu),n=1200)
#simulation of a 3-person mixture characterized with markers FGA, TH01 and TPOX
simumix(tab,which.loc=c('FGA','TH01','TPOX'), n =3)
```



---

simupopD	<i>Simulate multi-population allele frequencies for independent loci from a reference population, following a Dirichlet model</i>
----------	---

---

## Description

Simulate multi-population allele frequencies for independent loci, from a given reference population, following a Dirichlet model. Allele frequencies in the populations are generated as random deviates from a Dirichlet distribution, the parameters of which control the deviation of allele frequencies from the values in the reference population.

## Usage

```
simupopD(npop = 1, nloc = 1, na = 2, globalfreq = NULL, which.loc = NULL,
alpha1, alpha2 = 1)
```

## Arguments

<code>npop</code>	the number of populations
<code>nloc</code>	the number of loci
<code>na</code>	an integer vector giving the numbers of alleles per locus
<code>globalfreq</code>	matrix of allele frequencies in the reference population. Data must be given in the format of the Journal of Forensic Sciences for genetic data. Default corresponds to allele frequencies generated from a Dirichlet distribution with parameter <code>alpha2</code> for all allele frequencies.
<code>which.loc</code>	which loci to simulate from the <code>globalfreq</code> matrix, default considers all loci
<code>alpha1</code>	a positive float vector of length <code>npop</code> giving the variance parameter of the Dirichlet distribution used to generate allele frequencies in the <code>npop</code> independent populations
<code>alpha2</code>	a positive float giving the parameter to be used to in the Dirichlet distribution to generate allele frequencies for the reference population

## Details

In the reference population, allele frequencies for independent loci are simulated using a Dirichlet distribution with parameter `alpha2`.

At a given locus  $L$  with  $n$  alleles, the allele frequencies are modeled as a vector of random variables  $p=(p_1, \dots, p_n)$  following a Dirichlet distribution with a parameter vector of length  $n$ , where each component is equal to `alpha2`,  $p_1+\dots+p_n=1$  and `alpha2`  $> 0$ .

Note that a more sophisticated generation of global allele frequencies is possible using the [simufreqD](#) function. Similarly, allele frequencies in the independent populations are simulated using a Dirichlet Distribution. For example, for the first population to simulate, at a given locus  $L$  with  $n$  alleles, the allele frequencies are modeled as a vector of random variables  $p=(p_1, \dots, p_n)$  following a Dirichlet distribution with a parameter vector of length  $n$ :

$(p_1(1-\alpha_1)/\alpha_1[1], \dots, p_n(1-\alpha_1[1])/alpha1[1])$ , where  $p_1+\dots+p_n=1$  and  $\alpha_1[1] > 0$ .

`alpha1[1]` is the variance parameter for population 1 and is equivalent to Wright's  $F_{st}$ . The closest this parameter is to one, the more the population allele frequencies are different from the values of the reference population.

**Value**

The result is stored in a list with two elements :

<code>globfreq</code>	a <code>tabfreq</code> object giving the allele frequencies of the chosen reference population, with the chosen loci.
<code>popfreq</code>	a <code>tabfreq</code> object giving the allele frequencies of the simulated populations.

**Note**

The code used here for the generation of random Dirichlet deviates was previously implemented in the `gtools` library.

**Author(s)**

Hinda Haned <haned@biomserv.univ-lyon1.fr>

**References**

Nicholson G, Smith AV, Jonsson F, Gustafsson O, Stefansson K, Donnelly P. Assessing population differentiation and isolation from single-nucleotide polymorphism data. *J Roy Stat Soc B* 2002;64:695–715

Marchini J, Cardon LR. Discussion on the meeting on "Statistical modelling and analysis of genetic data" *J Roy Stat Soc B*, 2002;64:740-741

Wright S. The genetical structure of populations. *Ann Eugen* 1951;15:323-354

**See Also**

[simufreqD](#)

**Examples**

```
# simulate allele frequencies for two populations
data(Tu)
simupopD(npop=2, globalfreq=Tu, which.loc=c("FGA", "TH01", "TPOX"),
alpha1=c(0.2, 0.3), alpha2=1)
```

---

`strusa`

*Allele frequencies for 15 autosomal short tandem repeats core loci on U.S. Caucasian, African American, and Hispanic populations.*

---

**Description**

Allele frequencies for 15 autosomal short tandem repeats loci on three American populations : Caucasians, African Americans and Hispanics. Among the 15 loci, 13 belong to the core Combined DNA Index System (CODIS) loci used by the Federal Bureau of Investigation (USA), in forensic DNA analysis, and two supplementary loci are more commonly used in Europe, see details.

**Usage**

```
data(strusa)
```

**Format**

strusa is a tabfreq object giving allele frequencies of 15 loci in three American populations.

**Details**

CSF1PO, FGA, TH01, TPOX, vWA, D3S1358, D5S818, D7S820, D8S1179, D13S317, D16S539, D18S51 and D21S11, belong to the core CODIS loci used in the US, whereas D2S1338 and D19S433 belong to the European core loci.

**References**

Butler JM, Reeder DJ. <http://www.cstl.nist.gov/strbase/index.htm>, last visited: May 11th 2009

Butler JM, Schoske R, Vallone MP, Redman JW, Kline MC. Allele frequencies for 15 autosomal STR loci on U.S. Caucasian, African American, and Hispanic populations. *J Forensic Sci* 2003;48(8):908-911.

**Examples**

```
data(strusa)
strusa
#genotypes simulations from each population
geno<- simugeno(strusa,n=c(100,100,100))
geno
#3-person mixture simulation with the contributors from the 3 populations
mix3<- simumix(geno,ncontri=c(1,1,1))
mix3
```

---

strveneto

---

*Population study of three miniSTR loci in Veneto (Italy)*


---

**Description**

Allele frequencies for three short tandem repeats loci D10S1248, D2S441 and D22S1045 in a sample of 198 individuals born in Veneto, Italy. These loci are commonly used in forensic DNA characterization.

**Usage**

```
data(strveneto)
```

**Format**

strveneto is a tabfreq object

## References

Turrina S, Atzei R, De Leo D. Population study of three miniSTR loci in Veneto (Italy). Forensic Sci Int Genetics 2008; 1(1);378-379

## Examples

```
data(strveneto)
#allele frequencies
strveneto@tab
```

---

tabfreq	<i>forensim class for population allele frequencies</i>
---------	---

---

## Description

The S4 `tabfreq` class is used to store allele frequencies, from either one or several populations.

## Slots

**tab:** a list giving allele frequencies for each locus. If there are several populations, `tab` gives allele frequencies in each population

**which.loc:** character vector giving the names of the loci

**pop.names:** factor of populations names (optional)

## Methods

**names** signature(`x = "tabfreq"`): gives the names of the attributes of a `tabfreq` object

**show** signature(`object = "tabfreq"`): shows a `tabfreq` object

**print** signature(`object="tabfreq"`): prints a `tabfreq` object

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## See Also

[as.tabfreq](#), [is.tabfreq](#) and [simugeno](#) for genotypes simulation from allele frequencies stored in a `tabfreq` object

## Examples

```
showClass("tabfreq")
```

---

```
tabfreq constructor
      tabfreq constructor
```

---

## Description

Constructor for [tabfreq](#) objects.

The function `tabfreq` creates a [tabfreq](#) object from a data frame or a matrix giving allele frequencies for a single population in the Journal of Forensic Sciences (JFS) format for population genetic data. When multiple populations are considered, data shall be given as a list, where each element is either a matrix or a data frame in the JFS format, and the populations names must be specified.

The function `as.tabfreq` is an alias for the `tabfreq` function.

`is.tabfreq` tests if an object is a valid `tabfreq` object.

Note: to get the manpage about [tabfreq](#), please type `'class ? tabfreq'`.

## Usage

```
tabfreq(tab, pop.names=NULL)
as.tabfreq(tab, pop.names=NULL)
is.tabfreq(x)
```

## Arguments

<code>tab</code>	either a matrix or a data.frame of markers allele frequencies given in the Journal of Forensic Sciences format for population genetic data
<code>pop.names</code>	(optional) a factor giving the populations names. For a single population in <code>tab</code> , default is set to <code>NULL</code> .
<code>x</code>	an object

## Value

For `tabfreq` and `as.tabfreq`, a `tabfreq` object. For `is.tabfreq`, a logical.

## Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

## See Also

`"tabfreq"`, [simugeno](#) for creating a `simugeno` object from a `tabfreq` object.

## Examples

```
data(Tu)
tabfreq(Tu, pop.names=factor("Tu"))
```

---

Tu	<i>Allele frequencies of 15 autosomal short tandem repeats loci on Chinese Tu ethnic minority group</i>
----	---

---

### Description

Population genetic analysis of 15 STR loci of Chinese Tu ethnic minority group.

### Usage

```
data(Tu)
```

### Format

a data frame presented in the format of the Journal of Forensic Sciences for genetic data: allele names are given in the first column, and frequencies for a given allele are read in rows for the different markers. When a given allele is not observed, value is coded NA (rather than "-" in the original format).

### Details

CSF1PO, FGA, TH01, TPOX, vWA, D3S1358, D5S818, D7S820, D8S1179, D13S317, D16S539, D18S51 and D21S11, belong to the core CODIS loci used in the US, whereas D2S1338 and D19S433 belong to the European core loci.

### References

Zhu B, Yan J, Shen C, Li T, Li Y, Yu X, Xiong X, Muf H, Huang Y, Deng Y. (2008). Population genetic analysis of 15 STR loci of Chinese Tu ethnic minority group. *Forensic Sci Int*; 174: 255-258.

### Examples

```
data(Tu)
tabfreq(Tu)
```

---

virtualClasses	<i>Virtual classes for forensim</i>
----------------	-------------------------------------

---

### Description

Virtual classes that are only for internal use in forensim

### Objects from the Class

A virtual Class: programming tool, not intended for objects creation.

### Author(s)

Hinda Haned <haned@biomserv.univ-lyon1.fr>

# Index

## \*Topic **classes**

simugeno, 28  
 simumix, 30  
 tabfreq, 35  
 virtualClasses, 37

## \*Topic **datagen**

forensim-package, 1  
 simufreqD, 26  
 simugeno, 28  
 simugeno constructor, 29  
 simumix, 30  
 simumix constructor, 31  
 simupopD, 32  
 tabfreq, 35  
 tabfreq constructor, 36

## \*Topic **datasets**

strusa, 33  
 strveneto, 34  
 Tu, 37

## \*Topic **htest**

A2.simu, 2  
 A3.simu, 3  
 A4.simu, 4  
 dataL, 9  
 lik, 11  
 lik.loc, 12  
 likestim, 14  
 likestim.loc, 15  
 LR, 16  
 mastermix, 18  
 mincontri, 19  
 PE, 22  
 Pevd2, 23  
 RMP, 24

## \*Topic **manip**

Accessors, 6  
 changepop, 6  
 forensim-package, 1  
 naomitab, 20  
 simugeno, 28  
 simugeno constructor, 29  
 simumix, 30  
 simumix constructor, 31

tabfreq, 35

tabfreq constructor, 36

## \*Topic **misc**

findfreq, 10  
 findmax, 10  
 nball, 21

## \*Topic **models**

Cmn, 7  
 comb, 8  
 \$, simugeno-method (*Accessors*), 6  
 \$, simumix-method (*Accessors*), 6  
 \$, tabfreq-method (*Accessors*), 6  
 \$<-, simugeno-method (*Accessors*), 6  
 \$<-, simumix-method (*Accessors*), 6  
 \$<-, tabfreq-method (*Accessors*), 6

A2.simu, 2, 4, 5, 19

A3.simu, 3, 3, 5, 19

A4.simu, 3, 4, 4, 19

Accessors, 6

as.simugeno, 28

as.simugeno (*simugeno*  
*constructor*), 29

as.simumix, 30

as.simumix (*simumix constructor*),  
 31

as.tabfreq, 35

as.tabfreq (*tabfreq constructor*),  
 36

changepop, 6

characterOrNULL-class  
 (*virtualClasses*), 37

Cmn, 7, 8

comb, 7, 8

dataL, 7, 9, 11

factorOrNULL-class  
 (*virtualClasses*), 37

findfreq, 10

findmax, 10

forensim, 6

forensim (*forensim-package*), 1

- forensim-package, [1](#)
- is.simugeno, [28](#)
- is.simugeno(*simugeno*  
    *constructor*), [29](#)
- is.simumix, [30](#)
- is.simumix(*simumix constructor*),  
    [31](#)
- is.tabfreq, [35](#)
- is.tabfreq(*tabfreq constructor*),  
    [36](#)
- lik, [9](#), [11](#), [13](#)
- lik.loc, [9](#), [12](#), [12](#)
- likestim, [12](#), [13](#), [14](#), [16](#), [20](#)
- likestim.loc, [12](#), [13](#), [15](#), [15](#)
- listOrdataframe-class  
    (*virtualClasses*), [37](#)
- LR, [16](#), [23](#), [26](#)
- mastermix, [18](#)
- matrixOrdataframe-class  
    (*virtualClasses*), [37](#)
- mincontri, [19](#)
- names, simugeno-method(*simugeno*),  
    [28](#)
- names, simumix-method(*simumix*), [30](#)
- names, tabfreq-method(*tabfreq*), [35](#)
- naomitab, [20](#)
- nball, [21](#)
- PE, [18](#), [22](#)
- Pevid2, [23](#)
- print, simugeno-method(*simugeno*),  
    [28](#)
- print, simumix-method(*simumix*), [30](#)
- print, tabfreq-method(*tabfreq*), [35](#)
- RMP, [24](#)
- show, simugeno-method(*simugeno*),  
    [28](#)
- show, simumix-method(*simumix*), [30](#)
- show, tabfreq-method(*tabfreq*), [35](#)
- simufreqD, [26](#), [32](#), [33](#)
- simugeno, [1](#), [6](#), [28](#), [29–31](#), [35](#), [36](#)
- simugeno(*simugeno constructor*),  
    [29](#)
- simugeno constructor, [29](#)
- simugeno-class(*simugeno*), [28](#)
- simugeno-methods(*simugeno*  
    *constructor*), [29](#)
- simumix, [1](#), [6](#), [10](#), [20–22](#), [28](#), [30](#), [31](#)
- simumix(*simumix constructor*), [31](#)
- simumix constructor, [31](#)
- simumix-class(*simumix*), [30](#)
- simumix-methods(*simumix*  
    *constructor*), [31](#)
- simupopD, [27](#), [32](#)
- strusa, [33](#)
- strveneto, [34](#)
- tabfreq, [1](#), [6](#), [21](#), [28–31](#), [35](#), [36](#)
- tabfreq(*tabfreq constructor*), [36](#)
- tabfreq constructor, [36](#)
- tabfreq-class(*tabfreq*), [35](#)
- tabfreq-methods(*tabfreq*  
    *constructor*), [36](#)
- Tu, [37](#)
- vectorOrdataframe-class  
    (*virtualClasses*), [37](#)
- vectorOrNULL-class  
    (*virtualClasses*), [37](#)
- virtualClasses, [37](#)