



## **fitdistrplus: An R Package for Fitting Distributions**

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### **Abstract**

The package **fitdistrplus** provides functions for fitting univariate distributions to different types of data (continuous censored or non-censored data and discrete data) and allowing different estimation methods (maximum likelihood, moment matching, quantile matching and maximum goodness-of-fit estimation). Outputs of **fitdistr** and **fitdistrplus** functions are S3 objects, for which kind generic methods are provided, including **summary**, **plot** and **quantile**. This package also provides various functions to compare the fit of several distributions to a same data set and can handle bootstrap of parameter estimates. Detailed examples are given in food risk assessment, ecotoxicology and insurance contexts.

*Keywords:* probability distribution fitting, bootstrap, censored data, maximum likelihood, moment matching, quantile matching, maximum goodness-of-fit, distributions, R.

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## **1. Introduction**

Fitting distributions to data is a very common task in statistics and consists in choosing a probability distribution modelling the random variable, as well as finding parameter estimates for that distribution. This requires judgment and expertise and generally needs an iterative process of distribution choice, parameter estimation, and quality of fit assessment. In the R (R Development Core Team 2013) package **MASS** (Venables and Ripley 2010), maximum likelihood estimation is available via the **fitdistr** function; other steps of the fitting process can be done using other R functions (Ricci 2005). In this paper, we present the R package **fitdistrplus** (Delignette-Muller, Pouillot, Denis, and Dutang 2013) implementing several methods for fitting univariate parametric distribution. A first objective in developing this package was to provide R users a set of functions dedicated to help this overall process.

The **fitdistr** function estimates distribution parameters by maximizing the likelihood function using the **optim** function. In some cases, other estimation methods could be preferred, such as maximum goodness-of-fit estimation (also called minimum distance estimation), as pro-

posed in the R package **actuar** with three different goodness-of-fit distances (Dutang, Goulet, and Pigeon 2008). While developping the **fitdistrplus** package, a second objective was to consider various estimation methods in addition to maximum likelihood estimation (MLE). Functions were developped to enable moment matching estimation (MME), quantile matching estimation (QME), and maximum goodness-of-fit estimation (MGE) using eight different distances. Moreover, the **fitdistrplus** package offers the possibility to specify a user-supplied function for optimization, useful in cases where classical optimization techniques, not included in **optim**, are more adequate.

In applied statistics, it is frequent to have to fit distributions to censored data (Klein and Moeschberger 2003; Helsel 2005; Busschaert, Geeraerd, Uyttendaele, and VanImpe 2010; Leha, Beissbarth, and Jung 2011; Commeau, Parent, Delignette-Muller, and Cornu 2012). The **MASS** `fitdistr` function does not enable maximum likelihood estimation with this type of data. Some packages can be used to work with censored data, especially survival data (Therneau 2011; Hirano, Clayton, and Upper 1994; Jordan 2005), but those packages generally focused on specific models, enabling the fit of a restricted set of distributions. A third objective was thus to provide R users a function to estimate univariate distribution parameters from right-, left- and interval-censored data.

Few packages on CRAN provide estimation procedures for any user-supplied parametric distribution and support different types of data. The **distrMod** package (Kohl and Ruckdeschel 2010) provides an object-oriented (S4) implementation of probability models for non-censored data and includes distribution fitting procedures for a given minimization criterion. This package is based on the R-shipped function `mle` of the **stats4** package producing S4 objects of class "mle" for which generic methods are implemented, e.g., `confint` and `logLik`. In **fitdistrplus**, we chose to use the standard S3 class system for its understanding by most R users. When designing the **fitdistrplus** package, we did not forget to implement generic functions also available for S3 classes. Finally, various other packages provide functions to estimate the mode, the moments or the L-moments of a distribution, see the reference manuals of **modeest**, **lmomco** and **Lmoments** packages.

This manuscript reviews the various features of version 1.0-1 of **fitdistrplus**. The package is available from the Comprehensive R Archive Network at <http://cran.r-project.org/package=fitdistrplus>. The development version of the package is located at R-forge as one package of the project "Risk Assessment with R" (<http://r-forge.r-project.org/projects/riskassessment/>). The paper is organized as follows: Section 2 presents tools for fitting continuous distributions to classic non-censored data. Section 3 deals with other estimation methods and other types of data, before Section 4 concludes.

## 2. Fitting distributions to continuous non-censored data

### 2.1. Choice of candidate distributions

For illustrating the use of various functions of the **fitdistrplus** package with continuous non-censored data, we will first use a data set named **groundbeef** which is included in our package. This data set contains pointwise values of serving sizes in grams, collected in a French survey, for ground beef patties consumed by children under 5 years old. It was used in a quantitative risk assessment published by Delignette-Muller, Cornu, and AFSSA-STEC-Study-Group

(2008).

```
R>library("fitdistrplus")
R>data("groundbeef")
R>str(groundbeef)
```

```
'data.frame':      254 obs. of  1 variable:
 $ serving: num   30 10 20 24 20 24 40 20 50 30 ...
```

Before fitting one or more distributions to a data set, it is generally necessary to choose good candidates among a predefined set of distributions. This choice may be guided by the knowledge of stochastic processes governing the modelled variable, or, in the absence of knowledge regarding the underlying process, by the observation of its empirical distribution. To help the user in this choice, we developed functions to plot and characterize the empirical distribution.

First of all, it is common to start with plots of the empirical distribution function and the histogram, which can be obtained with the `plotdist` function of the **fitdistrplus** package. This function provides two plots (see Figure 1): the left-hand plot is the histogram (on a density scale) and the right-hand plot the empirical cumulative distribution function (CDF).

```
R>plotdist(groundbeef$serving)
```

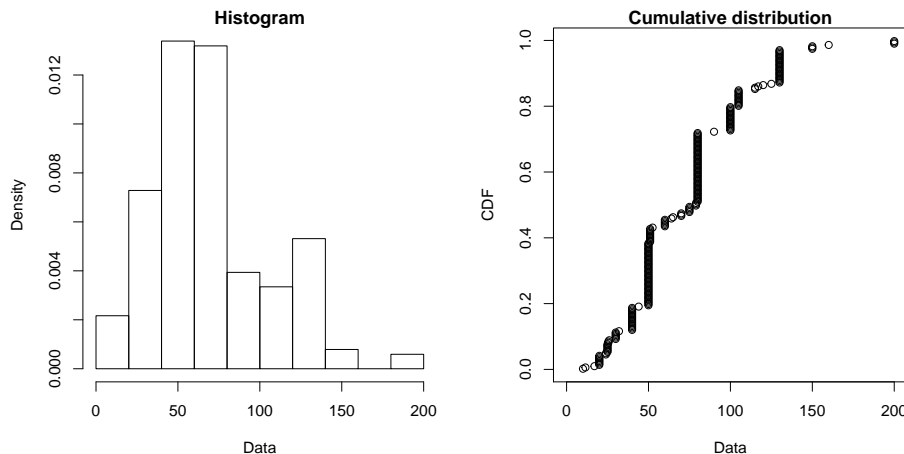


Figure 1: Histogram and CDF plots of an empirical distribution for a continuous variable (serving size from the **groundbeef** data set) as provided by the `plotdist` function.

In addition to empirical plots, descriptive statistics may help to choose candidates to describe a distribution among a set of parametric distributions. Especially the skewness and kurtosis, linked to the third and fourth moments, are useful for this purpose. A non-zero skewness reveals a lack of symmetry of the empirical distribution, while the kurtosis value quantifies the weight of tails in comparison to the normal distribution for which the kurtosis equals 3. The skewness and kurtosis and their corresponding unbiased estimator ([Casella and Berger](#)

2002) from a sample  $(X_i)_i \stackrel{\text{i.i.d.}}{\sim} X$  with observations  $(x_i)_i$  are given by

$$sk(X) = \frac{E[(X - E(X))^3]}{Var(X)^{\frac{3}{2}}}, \quad \widehat{sk} = \frac{\sqrt{n(n-1)}}{n-2} \times \frac{m_3}{m_2^{\frac{3}{2}}}, \quad (1)$$

$$kr(X) = \frac{E[(X - E(X))^4]}{Var(X)^2}, \quad \widehat{kr} = \frac{n-1}{(n-2)(n-3)}((n+1) \times \frac{m_4}{m_2^2} - 3(n-1)) + 3, \quad (2)$$

where  $m_2, m_3, m_4$  denote empirical moments defined by  $m_k = \frac{1}{n} \sum_{i=1}^n (x_i - \bar{x})^k$ , with  $x_i$  the  $n$  observations of variable  $x$  and  $\bar{x}$  their mean value.

The `descdist` function provides classical descriptive statistics (minimum, maximum, median, mean, standard deviation), skewness and kurtosis. By default, unbiased estimations of the three last statistics are provided. Nevertheless, the argument `method` can be changed from "unbiased" (default) to "sample" to obtain them without correction for bias. A skewness-kurtosis plot such as the one proposed by Cullen and Frey (1999) is provided by the `descdist` function for the empirical distribution (see Figure 2 for the `groundbeef` data set). On this plot, values for common distributions are displayed in order to help the choice of distributions to fit to data. For some distributions (normal, uniform, logistic, exponential), there is only one possible value for the skewness and the kurtosis. Thus, the distribution is represented by a single point on the plot. For other distributions, areas of possible values are represented, consisting in lines (as for gamma and lognormal distributions), or larger areas (as for beta distribution).

Skewness and kurtosis are known not to be robust. In order to take into account the uncertainty of the estimated values of kurtosis and skewness from data, a nonparametric bootstrap procedure (Efron and Tibshirani 1994) can be performed by using the argument `boot`. Values of skewness and kurtosis are computed on bootstrap samples (constructed by random sampling with replacement from the original data set) and reported on the skewness-kurtosis plot. The properties of the random variable should be considered, notably its expected value and its range, as a complement to the use of the `plotdist` and `descdist` functions. Below is a call to the `descdist` function to describe the distribution of the serving size from the `groundbeef` data set and to draw the corresponding skewness-kurtosis plot (see Figure 2). Looking at the results on this example with a positive skewness and a kurtosis not far from 3, the fit of three common right-skewed distributions could be considered, Weibull, gamma and lognormal distributions.

```
R>descdist(groundbeef$serving, boot=1000)
```

## 2.2. Fit of distributions by maximum likelihood estimation

Once selected, one or more parametric distributions  $f(\cdot|\theta)$  (with parameter  $\theta \in \mathbb{R}^d$ ) may be fitted to the data set, one at a time, using the `fitdist` function. Under the i.i.d. sample assumption, distribution parameters  $\theta$  are by default estimated by maximizing the likelihood function defined as:

$$L(\theta) = \prod_{i=1}^n f(x_i|\theta) \quad (3)$$

## summary statistics

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```

min: 10    max: 200
median: 79
mean: 73.65
estimated sd: 35.88
estimated skewness: 0.7353
estimated kurtosis: 3.551

```

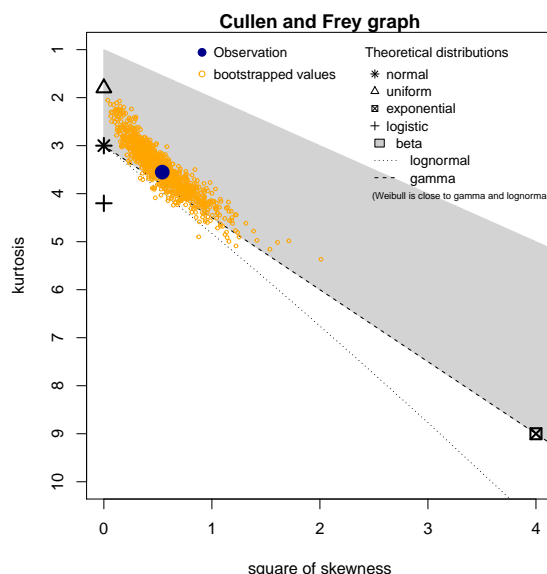


Figure 2: Skewness-kurtosis plot for a continuous variable (serving size from the **groundbeef** data set) as provided by the **descdist** function.

with  $x_i$  the  $n$  observations of variable  $X$  and  $f(\cdot|\theta)$  the density function of the parametric distribution. The other proposed estimation methods are described in Section 3.1.

The **fitdist** function returns an S3 object of class "fitdist" for which **print**, **summary** and **plot** functions are provided. The fit of a distribution using **fitdist** assumes that the corresponding **d**, **p**, **q** functions (standing respectively for the density, the distribution and the quantile functions) are defined. Classical distributions are already defined in that way in the **stats** package, e.g., **dnorm**, **pnorm** and **qnorm** for the normal distribution (see `?Distributions`). Others may be found in various packages (see the CRAN task view: Probability Distributions at <http://cran.r-project.org/web/views/Distributions.html>). Distributions not found in any package must be implemented by the user as **d**, **p**, **q** functions. In the call to **fitdist**, a distribution has to be specified via the argument **dist** either by the character string corresponding to its common root name used in the names of **d**, **p**, **q** functions (e.g., "norm" for the normal distribution) or by the density function itself, from which the root name is extracted (e.g., **dnorm** for the normal distribution). Numerical results returned by the **fitdist** function are (1) the parameter estimates, (2) the estimated standard errors (computed from the estimate of the Hessian matrix at the maximum likelihood solution), (3) the loglikelihood, (4) Akaike and Bayesian information criteria (the so-called AIC and BIC),

and (5) the correlation matrix between parameter estimates. Below is a call to the `fitdist` function to fit a Weibull distribution to the serving size from the `groundbeef` data set.

```
R>fw <- fitdist(groundbeef$serving, "weibull")
R>summary(fw)
```

```
Fitting of the distribution ' weibull ' by maximum likelihood
Parameters :
      estimate Std. Error
shape      2.186      0.1046
scale     83.348      2.5269
Loglikelihood: -1255   AIC:  2514   BIC:  2522
Correlation matrix:
      shape  scale
shape 1.0000 0.3218
scale 0.3218 1.0000
```

The plot of an object of class `"fitdist"` provides four classical goodness-of-fit plots (Cullen and Frey 1999) presented on Figure 3:

- a density plot representing the density function of the fitted distribution along with the histogram of the empirical distribution,
- a CDF plot of both the empirical distribution and the fitted distribution,
- a Q-Q plot representing the empirical quantiles (y-axis) against the theoretical quantiles (x-axis)
- a P-P plot representing the empirical distribution function evaluated at each data point (y-axis) against the fitted distribution function (x-axis).

For CDF, Q-Q and P-P plots, the probability plotting position is defined by default using Hazen's rule, with probability points of the empirical distribution calculated as  $(1:n - 0.5)/n$ , as recommended by Blom (1959). This plotting position can be easily changed (see the reference manual for details (Delignette-Muller *et al.* 2013)).

Unlike the generic `plot` function, the `denscomp`, `cdfcomp`, `qqcomp` and `ppcomp` functions enable to draw separately each of these four plots, in order to compare the empirical distribution and multiple parametric distributions fitted on a same data set. These functions must be called with a first argument corresponding to a list of objects of class `fitdist`, and optionally further arguments to customize the plot (see the reference manual for lists of arguments that may be specific to each plot (Delignette-Muller *et al.* 2013)). In the following example, we compare the fit of a Weibull, a lognormal and a gamma distributions to the `groundbeef` data set (Figure 3).

```
R>fg <- fitdist(groundbeef$serving, "gamma")
R>fln <- fitdist(groundbeef$serving, "lnorm")
R>par(mfrow=c(2, 2))
R>denscomp(list(fw, fln, fg), legendtext=c("Weibull", "lognormal", "gamma"))
```

```
R>qqcomp(list(fw,fln,fg), legendtext=c("Weibull", "lognormal", "gamma"))
R>cdfcomp(list(fw,fln,fg), legendtext=c("Weibull", "lognormal", "gamma"))
R>ppcomp(list(fw,fln,fg), legendtext=c("Weibull", "lognormal", "gamma"))
```

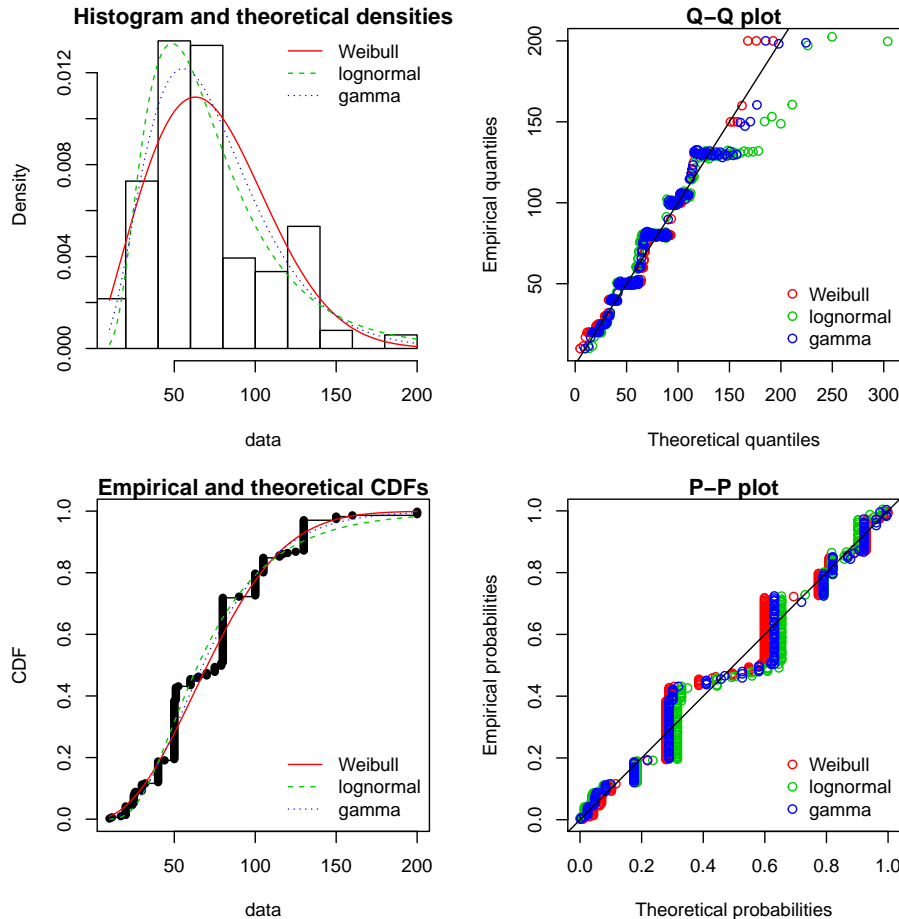


Figure 3: Four Goodness-of-fit plots for various distributions fitted to continuous data (Weibull, gamma and lognormal distributions fitted to serving sizes from the **groundbeef** data set) as provided by functions `denscomp`, `qqcomp`, `cdfcomp` and `ppcomp`.

The density plot and the CDF plot may be considered as the basic classical goodness-of-fits plots. The two other plots are complementary and can be very informative in some cases. The Q-Q plot emphasizes the lack-of-fit at the distribution tails while the P-P plot emphasizes the lack-of-fit at the distribution center. In the present example (in Figure 3), none of the three fitted distributions correctly describes the center of the distribution, but the Weibull and gamma distributions could be preferred for their better description of the right tail of the empirical distribution, especially if this tail is important in the use of the fitted distribution, as it is in the context of food risk assessment.

The data set named **endosulfan** will now be used to illustrate other features of the **fitdistrplus** package. This data set contains acute toxicity values for the organochlorine pesticide endosulfan (geometric mean of LC50 ou EC50 values in  $\mu\text{g.L}^{-1}$ ), tested on Australian and non-Australian laboratory-species (Hose and Van den Brink 2004).



```
R>data("endosulfan")
R>str(endosulfan)

'data.frame':      104 obs. of  3 variables:
 $ ATV           : num  0.6 2.8 182.2 0.8 478 ...
 $ Australian: Factor w/ 2 levels "no","yes": 2 2 2 2 2 2 2 2 1 ...
 $ group        : Factor w/ 3 levels "Arthropods","Fish",...: 1 1 1 1 1 ..
```

In ecotoxicology, a lognormal or a loglogistic distribution is often fitted to such a data set in order to characterize the species sensitivity distribution (SSD) for a pollutant. A low percentile of the fitted distribution, generally the 5% percentile, is then calculated and named the hazardous concentration 5% (HC5). It is interpreted as the value of the pollutant concentration protecting 95% of the species (Posthuma, Suter, and Traas 2010). But the fit of a lognormal or a loglogistic distribution to the whole `endosulfan` data set is rather bad (Figure 4), especially due to a minority of very high values. The two-parameter Pareto distribution and the three-parameter Burr distribution (which is an extension of both the loglogistic and the Pareto distributions) have been fitted. Pareto and Burr distributions are provided in the package **actuar**. Until here, we did not have to define starting values (in the optimization process) as reasonable starting values are implicitly defined within the `fitdist` function for most of the distributions defined in R (see `?fitdist` for details). For other distributions like the Pareto and the Burr distribution, initial values for the distribution parameters have to be supplied in the argument `start`, as a named list with initial values for each parameter (as they appear in the `d`, `p`, `q` functions). Having defined reasonable starting values<sup>1</sup>, various distributions can be fitted and graphically compared. On this example, the function `cdfcomp` can be used to report CDF values in a logscale so as to emphasize discrepancies on the tail of interest while defining an HC5 value (Figure 4).

```
R>ATV <-endosulfan$ATV
R>fendo.ln <- fitdist(ATV, "lnorm")
R>library("actuar")
R>fendo.ll <- fitdist(ATV, "llogis", start=list(shape=1, scale=500))
R>fendo.P <- fitdist(ATV, "pareto", start=list(shape=1, scale=500))
R>fendo.B <- fitdist(ATV, "burr", start=list(shape1=0.3, shape2=1, rate=1))
R>cdfcomp(list(fendo.ln, fendo.ll, fendo.P, fendo.B), xlogscale = TRUE,
+          ylogscale = TRUE, legendtext =
+          c("lognormal", "loglogistic", "Pareto", "Burr"))
```

None of the fitted distribution correctly describes the right tail observed in the data set, but as shown in Figure 4, the left-tail seems to be better described by the Burr distribution. Its use could then be considered to estimate the HC5 value as the 5% quantile of the distribution. This can be easily done using the `quantile` generic function defined for an object of class `"fitdist"`. Below is this calculation together with the calculation of the empirical quantile for comparison.

---

<sup>1</sup>The `plotdist` function can plot any parametric distribution with specified parameter values in argument `para`. It can thus help to find correct initial values for the distribution parameters in non trivial cases, by iterative calls if necessary (see the reference manual for examples (Delignette-Muller *et al.* 2013)).



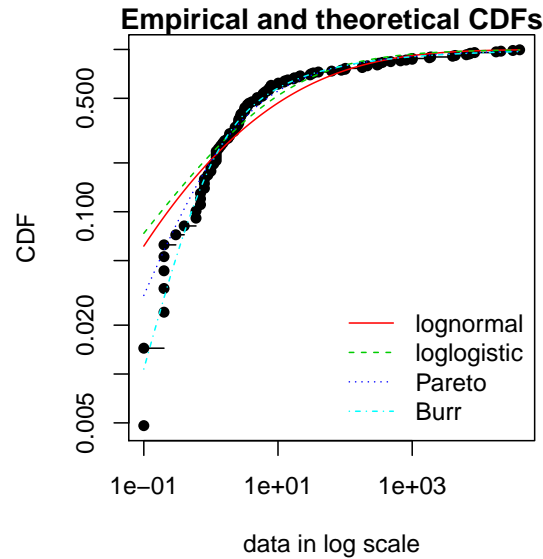


Figure 4: CDF plot to compare the fit of four distributions to acute toxicity values of various organisms for the organochlorine pesticide endosulfan (`endosulfan` data set) as provided by the `cdfcomp` function, with CDF values in a logscale to emphasize discrepancies on the left tail.

```
R>quantile(fendo.B, probs = 0.05)
```

Estimated quantiles for each specified probability (non-censored data)

```
p=0.05
estimate 0.2939
```

```
R>quantile(ATV, probs = 0.05)
```

```
5%
0.2
```

The computation of different goodness-of-fit statistics is proposed in the **fitdistrplus** package in order to further compare fitted distributions. The purpose of goodness-of-fit statistics aims to measure the distance between the fitted parametric distribution and the empirical distribution: e.g., the distance between the fitted cumulative distribution function  $F$  and the empirical distribution function  $F_n$ . When fitting continuous distributions, three goodness-of-fit statistics are classically considered: Cramer-von Mises, Kolmogorov-Smirnov and Anderson-Darling statistics (D’Agostino and Stephens 1986). Naming  $x_i$  the  $n$  observations of a continuous variable  $X$  arranged in an ascending order, Table 1 gives the definition and the empirical estimate of the three considered goodness-of-fit statistics. They can be computed using the function `gofstat` as defined by Stephens (D’Agostino and Stephens 1986).

```
R>gofstat(list(fendo.ln, fendo.ll, fendo.P, fendo.B),
+           fitnames = c("lnorm", "llogis", "Pareto", "Burr"))
```

Goodness-of-fit statistics

	lnorm	llogis	Pareto	Burr
Kolmogorov-Smirnov statistic	0.1672	0.1196	0.08488	0.06155
Cramer-von Mises statistic	0.6374	0.3827	0.13926	0.06803
Anderson-Darling statistic	3.4721	2.8316	0.89206	0.52393

Goodness-of-fit criteria

	lnorm	llogis	Pareto	Burr
Aikake's Information Criterion	1069	1069	1048	1046
Bayesian Information Criterion	1074	1075	1053	1054

Table 1: Goodness-of-fit statistics as defined by Stephens ([D'Agostino and Stephens 1986](#)).

Statistic	General formula	Computational formula
Kolmogorov-Smirnov (KS)	$\sup  F_n(x) - F(x) $	$\max(D^+, D^-)$ with $D^+ = \max_{i=1, \dots, n} \left( \frac{i}{n} - F_i \right)$ $D^- = \max_{i=1, \dots, n} \left( F_i - \frac{i-1}{n} \right)$
Cramer-von Mises (CvM)	$n \int_{-\infty}^{\infty} (F_n(x) - F(x))^2 dx$	$\frac{1}{12n} + \sum_{i=1}^n \left( F_i - \frac{2i-1}{2n} \right)^2$
Anderson-Darling (AD)	$n \int_{-\infty}^{\infty} \frac{(F_n(x) - F(x))^2}{F(x)(1-F(x))} dx$	$-n - \frac{1}{n} \sum_{i=1}^n (2i-1) \log(F_i(1-F_{n+1-i}))$

where  $F_i \triangleq F(x_i)$

As giving more weight to distribution tails, the Anderson-Darling statistic is of special interest when it matters to equally emphasize the tails as well as the main body of a distribution. This is often the case in risk assessment ([Cullen and Frey 1999](#); [Vose 2010](#)). For this reason, this statistics is often used to select the best distribution among those fitted. Nevertheless, this statistics should be used cautiously when comparing fits of various distributions. Keeping in mind that the weighting of each CDF quadratic difference depends on the parametric distribution in its definition (see Table 1), Anderson-Darling statistics computed for several distributions fitted on a same data set are theoretically difficult to compare. Moreover, such a statistic, as Cramer-von Mises and Kolmogorov-Smirnov ones, does not take into account the complexity of the model (i.e., parameter number). It is not a problem when compared distributions are characterized by the same number of parameters, but it could systematically promote the selection of the more complex distributions in the other case. Looking at classical penalized criteria based on the loglikelihood (AIC, BIC) seems thus also interesting, especially to discourage overfitting.

In the previous example, all the goodness-of-fit statistics based on the CDF distance are in favor of the Burr distribution, the only one characterized by three parameters, while AIC and

BIC values respectively give the preference to the Burr distribution or the Pareto distribution. The choice between these two distributions seems thus less obvious and could be discussed. Even if specifically recommended for discrete distributions, the Chi-squared statistic may also be used for continuous distributions (see Section 3.3 and the reference manual for examples (Delignette-Muller *et al.* 2013)).

### 2.3. Uncertainty in parameter estimates

The uncertainty in the parameters of the fitted distribution can be estimated by parametric or nonparametric bootstraps using the `bootdist` function for non-censored data (Efron and Tibshirani 1994). This function returns the bootstrapped values of parameters in an S3 class object which can be plotted to visualize the bootstrap region. The medians and the 95 percent confidence intervals of parameters (2.5 and 97.5 percentiles) are printed in the summary. When inferior to the whole number of iterations (due to lack of convergence of the optimization algorithm for some bootstrapped data sets), the number of iterations for which the estimation converges is also printed in the summary.

The plot of an object of class "bootdist" consists in a scatterplot or a matrix of scatterplots of the bootstrapped values of parameters providing a representation of the joint uncertainty distribution of the fitted parameters. Below is an example of the use of the `bootdist` function with the previous fit of the Burr distribution to the `endosulfan` data set (Figure 5).

```
R>bendo.B <- bootdist(fendo.B, niter=1001)
```

```
R>summary(bendo.B)
```

```
Parametric bootstrap medians and 95% percentile CI
```

	Median	2.5%	97.5%
shape1	0.1998	0.09755	0.3642
shape2	1.5861	1.03452	2.9930
rate	1.4923	0.68657	2.8278

```
The estimation method converged only for 1000 among 1001 iterations
```

```
R>plot(bendo.B)
```

Bootstrap samples of parameter estimates are useful especially to calculate confidence intervals on each parameter of the fitted distribution from the marginal distribution of the bootstrapped values. It is also interesting to look at the joint distribution of the bootstrapped values in a scatterplot (or a matrix of scatterplots if the number of parameters exceeds two) in order to understand the potential structural correlation between parameters (see Figure 5).

The use of the whole bootstrap sample is also of interest in the risk assessment field. Its use enables the characterization of uncertainty in distribution parameters. It can be directly used within a second-order Monte Carlo simulation framework, especially within the package **mc2d** (Pouillot, Delignette-Muller, and Denis 2011b). One could refer to Pouillot and Delignette-Muller (2010) for an introduction to the use of **mc2d** and **fitdistrplus** packages in the context of quantitative risk assessment.

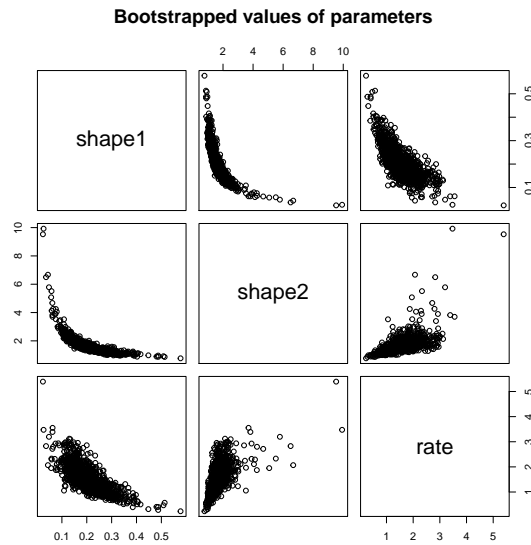


Figure 5: Bootstrapped values of parameters for a fit of the Burr distribution characterized by three parameters (example on the `endosulfan` data set) as provided by the plot of an object of class `"bootdist"`.

The bootstrap method can also be used to calculate confidence intervals on quantiles of the fitted distribution. For this purpose, a generic `quantile` function is provided for class `bootdist`. By default, 95% percentiles bootstrap confidence intervals of quantiles are provided. Going back to the previous example from ecotoxicology, this function can be used to estimate the uncertainty associated to the HC5 estimation, for example from the previously fitted Burr distribution to the `endosulfan` data set.

```
R>quantile(bendo.B, probs = 0.05)
```

```
(original) estimated quantiles for each specified probability (non-censored data)
```

```
  p=0.05
```

```
estimate 0.2939
```

```
Median of bootstrap estimates
```

```
  p=0.05
```

```
estimate 0.3024
```

```
two-sided 95 % CI of each quantile
```

```
  p=0.05
```

```
2.5 % 0.1757
```

```
97.5 % 0.5035
```

The estimation method converged only for 1000 among 1001 bootstrap iterations.

### 3. Advanced topics

#### 3.1. Alternative methods for parameter estimation

This subsection focuses on alternative estimation methods. One of the alternative for continuous distributions is the maximum goodness-of-fit estimation method also called minimum distance estimation method (D'Agostino and Stephens 1986; Dutang *et al.* 2008). In this package this method is proposed with eight different distances: the three classical distances defined in Table 1, or one of the variants of the Anderson-Darling distance proposed by Luceno (2006) and defined in Table 2. The right-tail AD gives more weight to the right-tail, the left-tail AD gives more weight only to the left tail. Either of the tails, or both of them, can receive even larger weights by using second order Anderson-Darling Statistics.

Table 2: Modified Anderson-Darling statistics as defined by Luceno (2006).

Statistic	General formula	Computational formula
Right-tail AD (ADR)	$\int_{-\infty}^{\infty} \frac{(F_n(x)-F(x))^2}{1-F(x)} dx$	$\frac{n}{2} - 2 \sum_{i=1}^n F_i - \frac{1}{n} \sum_{i=1}^n (2i-1) \ln(\bar{F}_{n+1-i})$
Left-tail AD (ADL)	$\int_{-\infty}^{\infty} \frac{(F_n(x)-F(x))^2}{(F(x))} dx$	$-\frac{3n}{2} + 2 \sum_{i=1}^n F_i - \frac{1}{n} \sum_{i=1}^n (2i-1) \ln(F_i)$
Right-tail AD 2nd order (AD2R)	$ad2r = \int_{-\infty}^{\infty} \frac{(F_n(x)-F(x))^2}{(1-F(x))^2} dx$	$ad2r = 2 \sum_{i=1}^n \ln(\bar{F}_i) + \frac{1}{n} \sum_{i=1}^n \frac{2i-1}{\bar{F}_{n+1-i}}$
Left-tail AD 2nd order (AD2L)	$ad2l = \int_{-\infty}^{\infty} \frac{(F_n(x)-F(x))^2}{(F(x))^2} dx$	$ad2l = 2 \sum_{i=1}^n \ln(F_i) + \frac{1}{n} \sum_{i=1}^n \frac{2i-1}{F_i}$
AD 2nd order (AD2)	$ad2r + ad2l$	$ad2r + ad2l$

where  $F_i \triangleq F(x_i)$ ;  $\bar{F}_i \triangleq 1 - F(x_i)$

To fit a distribution by maximum goodness-of-fit estimation, one needs to fix the argument `method` to "mge" in the call to `fitdist` and to specify the argument `gof` coding for the chosen goodness-of-fit distance. This function is intended to be used only with continuous non-censored data.

Maximum goodness-of-fit estimation may be useful to give more weight to data at one tail of the distribution. In the previous example from ecotoxicology, we used a non classical distribution (the Burr distribution) to correctly fit the empirical distribution especially on its left tail. In order to correctly estimate the 5% percentile, we could also consider the fit of the classical lognormal distribution, but minimizing a goodness-of-fit distance giving more weight to the left tail of the empirical distribution. In what follows, the left tail Anderson-Darling distances of first or second order are used to fit a lognormal to `endosulfan` data set (see Figure 6).

```
R>fendo.ln.ADL <- fitdist(ATV,"lnorm",method="mge",gof="ADL")
```

```
R>fendo.ln.AD2L <- fitdistr(ATV,"lnorm",method="mge",gof="AD2L")
R>cdfcomp(list(fendo.ln, fendo.ln.ADL, fendo.ln.AD2L),
+ xlogscale = TRUE, ylogscale = TRUE,
+ main = "Fitting a lognormal distribution",xlegend = "bottomright",
+ legendtext = c("MLE","Left-tail AD", "Left-tail AD 2nd order"))
```

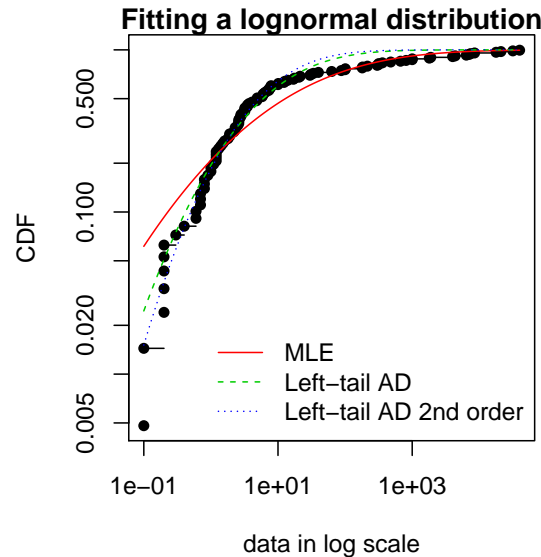


Figure 6: Comparison of a lognormal distribution fitted by MLE and by MGE using two different goodness-of-fit distances : left-tail Anderson-Darling and left-tail Anderson Darling of second order (example with the `endosulfan` data set) as provided by the `cdfcomp` function, with CDF values in a logscale to emphasize discrepancies on the left tail.

Comparing the 5% percentiles (HC5) calculated using these three fits to the one calculated from the MLE fit of the Burr distribution, we can observe, on this example, that fitting the lognormal distribution by maximizing left tail Anderson-Darling distances of first or second order enables to approach the value obtained by fitting the Burr distribution by MLE.

```
R>(HC5.estimates <- c(
+ empirical = as.numeric(quantile(ATV, probs=0.05)),
+ Burr = as.numeric(quantile(fendo.B, probs=0.05)$quantiles),
+ lognormal_MLE = as.numeric(quantile(fendo.ln, probs=0.05)$quantiles),
+ lognormal_AD2 = as.numeric(quantile(fendo.ln.ADL, probs=0.05)$quantiles),
+ lognormal_AD2L = as.numeric(quantile(fendo.ln.AD2L,probs=0.05)$quantiles)))
```

empirical	Burr	lognormal_MLE	lognormal_AD2	lognormal_AD2L
0.20000	0.29393	0.07259	0.19591	0.25877

The moment matching estimation (MME) is another method commonly used to fit parametric distributions (Vose 2010). MME consists in finding the value of the parameter  $\theta$  that equalizes

the first theoretical raw moments of the parametric distribution to the corresponding empirical raw moments as in Equation (4):

$$E(X^k|\theta) = \frac{1}{n} \sum_{i=1}^n x_i^k, \quad (4)$$

for  $k = 1, \dots, d$ , with  $d$  the number of parameters to estimate and  $x_i$  the  $n$  observations of variable  $X$ . For moments of order greater than or equal to 2, it may also be relevant to match centered moments. Therefore, we match the moments given in Equation (5):

$$E(X|\theta) = \bar{x}, \quad E\left((X - E(X))^k|\theta\right) = m_k, \quad \text{for } k = 2, \dots, d, \quad (5)$$

where  $m_k$  denotes the empirical centered moments. This method can be performed by setting the argument `method` to `"mme"` in the call to `fitdist`. The estimate is computed by a closed-form formula for the following distributions: normal, lognormal, exponential, Poisson, gamma, logistic, negative binomial, geometric, beta and uniform distributions. In this case, for distributions characterized by one parameter (geometric, Poisson and exponential), this parameter is simply estimated by matching theoretical and observed means, and for distributions characterized by two parameters, these parameters are estimated by matching theoretical and observed means and variances (Vose 2010). For other distributions, the equation of moments is solved numerically using the `optim` function by minimizing the sum of squared differences between observed and theoretical moments (see the `fitdistrplus` reference manual for technical details (Delignette-Muller *et al.* 2013)).

A classical data set from the Danish insurance industry published in McNeil (1997) will be used to illustrate this method. In `fitdistrplus`, the data set is stored in `danishuni` for the univariate version and contains the loss amounts collected at Copenhagen Reinsurance between 1980 and 1990. In actuarial science, it is standard to consider positive heavy-tailed distributions and have a special focus on the right-tail of the distributions. In this numerical experiment, the lognormal distribution is first considered and then the Pareto type II distribution (Klugman, Panjer, and Willmot 2009).

The lognormal distribution is fitted to `danishuni` data set by matching moments implemented as a closed-form formula. On the left-hand graph of Figure 7, the fitted distribution functions obtained using the moment matching estimation (MME) and maximum likelihood estimation (MLE) methods are compared. The MME method provides a more cautious estimation of the insurance risk as the MME-fitted distribution function (resp. MLE-fitted) underestimates (overestimates) the empirical distribution function for large values of claim amounts.

```
R>data("danishuni")
R>str(danishuni)

'data.frame':      2167 obs. of  2 variables:
 $ Date: Date, format: "1980-01-03" "1980-01-04" ...
 $ Loss: num  1.68 2.09 1.73 1.78 4.61 ...

R>fdanish.ln.MLE <- fitdist(danishuni$Loss, "lnorm")
R>fdanish.ln.MME <- fitdist(danishuni$Loss, "lnorm", method="mme", order=1:2)
```



```
R>cdfcomp(list(fdanish.ln.MLE, fdanish.ln.MME),
+         legend=c("lognormal MLE", "lognormal MME"),
+         main="Fitting a lognormal distribution",
+         xlogscale=TRUE, datapch=20)
```

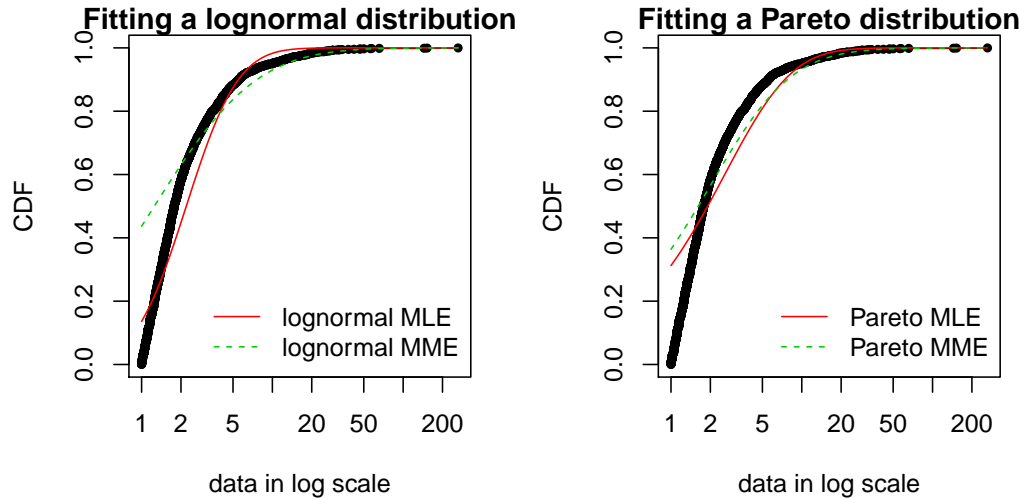


Figure 7: Comparison between MME and MLE when fitting a lognormal or a Pareto distribution to loss data from the `danishuni` data set.

In a second time, a Pareto distribution, which gives more weight to the right-tail of the distribution, is fitted. As the lognormal distribution, the Pareto has two parameters, which allows a fair comparison.

We use the implementation of the **actuar** package providing raw and centered moments for that distribution (in addition to `d`, `p`, `q` and `r` functions (Goulet 2012)). Fitting a heavy-tailed distribution for which the first and the second moments do not exist for certain values of the shape parameter requires some cautiousness. This is carried out by providing, for the optimization process, a lower and an upper bound for each parameter. The code below calls the L-BFGS-B optimization method in `optim`, since this quasi-Newton allows box constraints<sup>2</sup>. We choose match moments defined in Equation (4), and so a function for computing the empirical raw moment (called `memp` in our example) is passed to `fitdist`. For two-parameter distributions (i.e.,  $d = 2$ ), Equations (4) and (5) are equivalent.

```
R>library("actuar")
R>fdanish.P.MLE <- fitdist(danishuni$Loss, "pareto",
+ start=c(shape=10, scale=10), lower=2+1e-6, upper=Inf)
R>memp <- function(x, order) sum(x^order)/length(x)
R>fdanish.P.MME <- fitdist(danishuni$Loss, "pareto", method="mme",
+ order=1:2, memp="memp", start=c(shape=10, scale=10),
+ lower=c(2+1e-6, 2+1e-6), upper=c(Inf, Inf))
R>cdfcomp(list(fdanish.P.MLE, fdanish.P.MME),
```

<sup>2</sup>That is what the B stands for.

```
+ legend=c("Pareto MLE", "Pareto MME"),
+ main="Fitting a Pareto distribution", xlogscale=TRUE, datapch=".")
R>gofstat(list(fdanish.ln.MLE, fdanish.P.MLE,
+ fdanish.ln.MME, fdanish.P.MME),
+ fitnames = c("lnorm.mle","Pareto.mle","lnorm.mle","Pareto.mme"))
```

Goodness-of-fit statistics

	lnorm.mle	Pareto.mle	lnorm.mle	Pareto.mme
Kolmogorov-Smirnov statistic	0.1375	0.3124	0.4368	0.3638
Cramer-von Mises statistic	14.7911	37.7166	88.9503	53.0783
Anderson-Darling statistic	87.1933	208.3139	416.2567	272.4729

Goodness-of-fit criteria

	lnorm.mle	Pareto.mle	lnorm.mle	Pareto.mme
Aikake's Information Criterion	8120	9250	9792	9395
Bayesian Information Criterion	8131	9261	9803	9407

As shown on Figure 7, MME and MLE fits are far less distant (when looking at the right-tail) for the Pareto distribution than for the lognormal distribution on this data set. Furthermore, for these two distributions, the MME method better fits the right-tail of the distribution, which seems logical since empirical moments are influenced by large observed values. In the previous traces, we gave the values of goodness-of-fit statistics. Whatever the statistic considered, the MLE-fitted lognormal always provides the best fit to the observed data.

Maximum likelihood and moment matching estimations are certainly the most commonly used method for fitting distributions (Cullen and Frey 1999). Keeping in mind that these two methods may produce very different results, the user should be aware of its great sensitivity to outliers when choosing the moment matching estimation. This may be seen as an advantage in our example if the objective is to better describe the right tail of the distribution, but it may be seen as a drawback if the objective is different.

Fitting of a parametric distribution may also be done by matching theoretical quantiles of the parametric distributions (for specified probabilities) against the empirical quantiles (Tse (2009)). The equality of theoretical and empirical quantiles is expressed by Equation (6) below, which is very similar to Equations (4) and (5):

$$F^{-1}(p_k|\theta) = Q_{n,p_k} \quad (6)$$

for  $k = 1, \dots, d$ , with  $d$  the number of parameters to estimate (dimension of  $\theta$  if there is no fixed parameters) and  $Q_{n,p_k}$  the empirical quantiles calculated from data for specified probabilities  $p_k$ .

Quantile matching estimation (QME) is performed by setting the argument `method` to "qme" in the call to `fitdist` and adding an argument `probs` defining the probabilities for which the quantile matching is performed. The length of this vector must be equal to the number of parameters to estimate (as the vector of moment orders for MME). Empirical quantiles are computed using the `quantile` function of the `stats` package using `type=7` by default (see `?quantile` and Hyndman and Fan (1996)). But the type of quantile can be easily changed by using the `qty` argument in the call to the `qme` function. The quantile matching is carried out numerically, by minimizing the sum of squared differences between observed and theoretical quantiles.

```

R>fdanish.ln.QME1 <- fitdist(danishuni$Loss, "lnorm", method="qme",
+ probs=c(1/3, 2/3))
R>fdanish.ln.QME2 <- fitdist(danishuni$Loss, "lnorm", method="qme",
+ probs=c(8/10, 9/10))
R>cdfcomp(list(fdanish.ln.MLE, fdanish.ln.QME1, fdanish.ln.QME2),
+ legend=c("MLE", "QME(1/3, 2/3)", "QME(8/10, 9/10)"),
+ main="Fitting a lognormal distribution", xlogscale=TRUE, datapch=20)

```

Above is an example of fitting of a lognormal distribution to `danishuni` data set by matching probabilities ( $p_1 = 1/3, p_2 = 2/3$ ) and ( $p_1 = 8/10, p_2 = 9/10$ ). As expected, the second QME fit gives more weight to the right-tail of the distribution. Compared to the maximum likelihood estimation, the second QME fit best suits the right-tail of the distribution, whereas the first QME fit best models the body of the distribution. The quantile matching estimation is of particular interest when we need a focus around particular quantiles, e.g.,  $p = 99.5\%$  in the Solvency II insurance context or  $p = 5\%$  for the HC5 estimation in the ecotoxicology context.

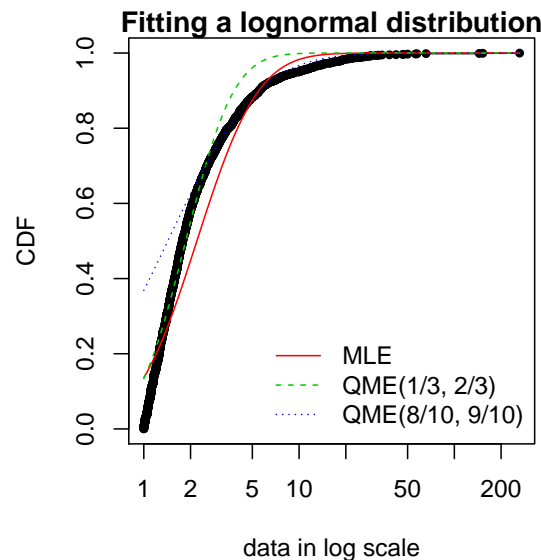


Figure 8: Comparison between QME and MLE when fitting a lognormal distribution to loss data from the `danishuni` data set.

### 3.2. Customization of the optimization algorithm

Each time a numerical minimization is carried out in the `fitdistrplus` package, the `optim` function of the `stats` package is used by default with the "Nelder-Mead" method for distributions characterized by more than one parameter and the "BFGS" method for distributions characterized by only one parameter. Sometimes the default algorithm fails to converge. It is then interesting to change some options of the `optim` function or to use another optimization function than `optim` to minimize the objective function. The argument `optim.method` can be used in the call to `fitdist` or `fitdistcens`. It will internally be passed to `mledist`,

`mmedist`, `mgedist` or `qmedist`, and to `optim` (see `?optim` for details about the different algorithms available).

Below are examples of fits of a gamma distribution  $\mathcal{G}(\alpha, \lambda)$  to the `groundbeef` data set with various algorithms. Note that the conjugate gradient algorithm ("`CG`") needs far more iterations to converge (around 2500 iterations) compared to other algorithms (converging in less than 100 iterations).

```
R>data("groundbeef")
R>fNM <- fitdist(groundbeef$serving, "gamma", optim.method="Nelder-Mead")
R>fBFGS <- fitdist(groundbeef$serving, "gamma", optim.method="BFGS")
R>fSANN <- fitdist(groundbeef$serving, "gamma", optim.method="SANN")
R>fCG <- try(fitdist(groundbeef$serving, "gamma", optim.method="CG",
+   control=list(maxit=10000)))
R>if(class(fCG) == "try-error")
+   fCG <- list(estimate=NA)
```

It is also possible to use another function than `optim` to minimize the objective function by specifying by the argument `custom.optim` in the call to `fitdist`. It may be necessary to customize this optimization function to meet the following requirements. (1) `custom.optim` function must have the following arguments: `fn` for the function to be optimized and `par` for the initialized parameters. (2) `custom.optim` should carry out a MINIMIZATION and must return the following components: `par` for the estimate, `convergence` for the convergence code, `value=fn(par)` and `hessian`. Below is an example of code written to wrap the `genoud` function from the `rgenoud` package in order to respect our optimization “template”. The `rgenoud` package implements the genetic (stochastic) algorithm.

```
R>mygenoud <- function(fn, par, ...)
+ {
+   require(rgenoud)
+   res <- genoud(fn, starting.values=par, ...)
+   standardres <- c(res, convergence=0)
+   return(standardres)
+ }
```

The customized optimization function can then be passed as the argument `custom.optim` in the call to `fitdist` or `fitdistcens`. The following code can for example be used to fit a gamma distribution to the `groundbeef` data set. Note that in this example various arguments are also passed from `fitdist` to `genoud`: `nvars`, `Domains`, `boundary.enforcement`, `print.level` and `hessian`. The code below compares all the parameter estimates ( $\hat{\alpha}$ ,  $\hat{\lambda}$ ) by the different algorithms: shape  $\alpha$  and rate  $\lambda$  parameters are relatively similar on this example, roughly 4.00 and 0.05, respectively.

```
R>fgenoud <- mledist(groundbeef$serving, "gamma", custom.optim= mygenoud,
+   nvars=2, max.generations=10, Domains=cbind(c(0,0), c(10,10)),
+   boundary.enforcement=1, hessian=TRUE, print.level=0, P9=10)
R>cbind(NM = fNM$estimate,
+   BFGS = fBFGS$estimate,
```

```
+ SANN = fSANN$estimate,
+ CG = fCG$estimate,
+ fgenoud = fgenoud$estimate)
```

```
      NM      BFGS      SANN      CG fgenoud
shape 4.00825 4.22848 3.97131 4.12891 4.00834
rate  0.05442 0.05742 0.05392 0.05607 0.05443
```

### 3.3. Fitting distributions to other types of data

Analytical methods often lead to semi-quantitative results which are referred to as censored data. Observations only known to be under a limit of detection are left-censored data. Observations only known to be above a limit of quantification are right-censored data. Results known to lie between two bounds are interval-censored data. These two bounds may correspond to a limit of detection and a limit of quantification, or more generally to uncertainty bounds around the observation. Right-censored data are also commonly encountered with survival data (Klein and Moeschberger 2003). A data set may thus contain right-, left-, or interval-censored data, or may be a mixture of these categories, possibly with different upper and lower bounds. Censored data are sometimes excluded from the data analysis or replaced by a fixed value, which in both cases may lead to biased results. A more recommended approach to correctly model such data is based upon maximum likelihood (Klein and Moeschberger 2003; Helsel 2005).

Censored data may thus contain left-censored, right-censored and interval-censored values, with several lower and upper bounds. Before their use in package **fitdistrplus**, such data must be coded into a dataframe with two columns, respectively named **left** and **right**, describing each observed value as an interval. The **left** column contains either NA for left censored observations, the left bound of the interval for interval censored observations, or the observed value for non-censored observations. The **right** column contains either NA for right censored observations, the right bound of the interval for interval censored observations, or the observed value for non-censored observations. To illustrate the use of package **fitdistrplus** to fit distributions to censored continuous data, we will use another data set from ecotoxicology, included in our package and named **salinity**. This data set contains acute salinity tolerance (LC50 values in electrical conductivity,  $mS.cm^{-1}$ ) of riverine macro-invertebrates taxa from the southern Murray-Darling Basin in Central Victoria, Australia (Kefford, Fields, Clay, and Nuggeoda 2007).

```
R>data("salinity")
R>str(salinity)
```

```
'data.frame':      108 obs. of  2 variables:
 $ left : num  20 20 20 20 20 21.5 15 20 23.7 25 ...
 $ right: num  NA NA NA NA NA 21.5 30 25 23.7 NA ...
```

Using censored data such as those coded in the **salinity** data set, the empirical distribution can be plotted using the **plotdistcens** function. By default, this function uses the Expectation-Maximization approach of Turnbull (1974) to compute the overall empirical cdf

curve with optional confidence intervals, by calls to `survfit` and `plot.survfit` functions from the **survival** package (Figure 10 shows the Turnbull plot of data together with two fitted distributions). A less rigorous but sometimes more illustrative plot can be obtained by fixing the argument `Turnbull` to `FALSE` in the call to `plotdistcens` (see Figure 9 for an example and the help page of Function `plotdistcens` for details). This plot enables to see the real nature of censored data, as points and intervals.

```
R>plotdistcens(salinity,Turnbull = FALSE)
```

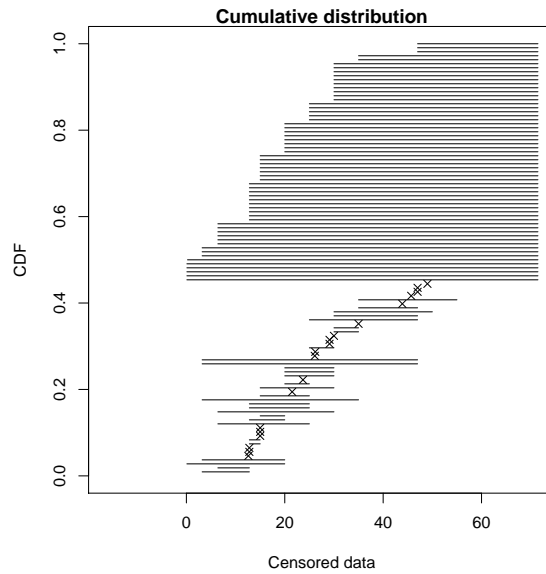


Figure 9: Simple plot of censored raw data (72-hour acute salinity tolerance of riverine macro-invertebrates from the `salinity` data set) as ordered points and intervals.

As for non censored data, one or more parametric distributions can be fitted to the censored data set, one at a time, but using in this case the `fitdistcens` function. This function estimates the vector of distribution parameters  $\theta$  by maximizing the likelihood for censored data defined as:

$$L(\theta) = \prod_{i=1}^{N_{nonC}} f(x_i|\theta) \times \prod_{j=1}^{N_{leftC}} F(x_j^{upper}|\theta) \times \prod_{k=1}^{N_{rightC}} (1 - F(x_k^{lower}|\theta)) \times \prod_{m=1}^{N_{intC}} (F(x_m^{upper}|\theta) - F(x_j^{lower}|\theta)) \quad (7)$$

with  $x_i$  the  $N_{nonC}$  non-censored observations,  $x_j^{upper}$  upper values defining the  $N_{leftC}$  left-censored observations,  $x_k^{lower}$  lower values defining the  $N_{rightC}$  right-censored observations,  $[x_m^{lower}; x_m^{upper}]$  the intervals defining the  $N_{intC}$  interval-censored observations, and  $F$  the cumulative distribution function of the parametric distribution (Klein and Moeschberger 2003; Helsel 2005).

As `fitdist`, `fitdistcens` returns the results of the fit of any parametric distribution to a data set as an S3 class object that can be easily printed, summarized or plotted. For the `salinity` data set, a lognormal distribution or a loglogistic can be fitted as commonly done in ecotoxicology for such data. As with `fitdist`, for some distributions (see Delignette-Muller

[et al. \(2013\)](#) for details), it is necessary to specify initial values for the distribution parameters in the argument `start`. The `plotdistcens` function can help to find correct initial values for the distribution parameters in non trivial cases, by a manual iterative use if necessary.

```
R>fsal.ln <- fitdistcens(salinity, "lnorm")
R>fsal.ll <- fitdistcens(salinity, "llogis", start=list(shape=5, scale=40))
R>summary(fsal.ln)
```

```
FITTING OF THE DISTRIBUTION ' lnorm ' BY MAXIMUM LIKELIHOOD ON CENSORED DATA
PARAMETERS
```

```
      estimate Std. Error
meanlog   3.3854    0.06487
sdlog     0.4961    0.05455
Loglikelihood: -139.1   AIC:  282.1   BIC:  287.5
Correlation matrix:
      meanlog sdlog
meanlog  1.0000 0.2938
sdlog    0.2938 1.0000
```

```
R>summary(fsal.ll)
```

```
FITTING OF THE DISTRIBUTION ' llogis ' BY MAXIMUM LIKELIHOOD ON CENSORED DATA
PARAMETERS
```

```
      estimate Std. Error
shape    3.421    0.4158
scale    29.930    1.9447
Loglikelihood: -140.1   AIC:  284.1   BIC:  289.5
Correlation matrix:
      shape  scale
shape   1.0000 -0.2022
scale  -0.2022  1.0000
```

Computations of goodness-of-fit statistics have not yet been developed for fits using censored data but the quality of fit can be judged using Akaike and Schwarz's Bayesian information criteria (AIC and BIC) and the goodness-of-fit CDF plot, respectively provided when summarizing or plotting an object of class `"fitdistcens"`. Function `cdfcomp` can also be used to compare the fit of various distributions to the same censored data set. Its call is similar to the one of `cdfcomp`. Below is an example of comparison of the two fitted distributions to the `salinity` data set (see [Figure 10](#)).

```
R>cdfcomp(cens(list(fsal.ln, fsal.ll),
+               legendtext=c("lognormal", "loglogistic ")))
```

Function `bootdistcens` is the equivalent of `bootdist` for censored data, except that it only proposes nonparametric bootstrap. Indeed, it is not obvious to simulate censoring within a parametric bootstrap resampling procedure. The generic function `quantile` can also be



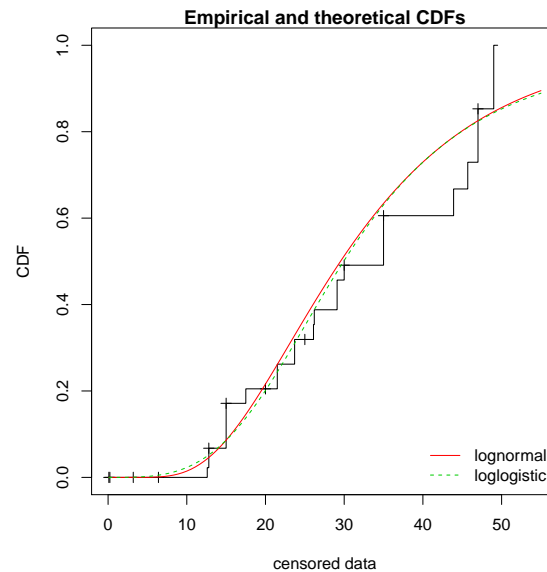


Figure 10: Goodness-of-fit CDF plot for fits of a lognormal and a loglogistic distribution to censored data: LC50 values from the `salinity` data set.

applied to an object of class `"fitdistcens"` or `"bootdistcens"`, as for continuous non-censored data.

In addition to the fit of distributions to censored or non censored continuous data, our package can also accomodate discrete variables, such as count numbers, using the functions developped for continuous non-censored data. These functions will provide somewhat different graphs and statistics, taking into account the discrete nature of the modeled variable. The discrete nature of the variable is automatically recognized when a classical distribution is fitted to data (binomial, negative binomial, geometric, hypergeometric and Poisson distributions) but must be indicated by fixing argument `discrete` to `TRUE` in the call to functions in other cases. The `toxocara` data set included in the package corresponds to the observation of such a discrete variable. Numbers of *Toxocara cati* parasites present in digestive tract are reported from a random sampling of feral cats living on Kerguelen island (Fromont, Morvilliers, Artois, and Pontier 2001). We will use it to illustrate the case of discrete data.

```
R>data("toxocara")
R>str(toxocara)
```

```
'data.frame':      53 obs. of  1 variable:
 $ number: int  0 0 0 0 0 0 0 0 0 0 ...
```

The fit of a discrete distribution to discrete data by maximum likelihood estimation requires the same procedure as for continuous non-censored data. As an example, using the `toxocara` data set, Poisson and negative binomial distributions can be easily fitted.

```
R>(ftoxo.P <- fitdist(toxocara$number, "pois"))
```

Fitting of the distribution ' pois ' by maximum likelihood

Parameters:

	estimate	Std. Error
lambda	8.679	0.4047

```
R>(ftoxo.nb <- fitdist(toxocara$number, "nbinom"))
```

Fitting of the distribution ' nbinom ' by maximum likelihood

Parameters:

	estimate	Std. Error
size	0.3971	0.08289
mu	8.6803	1.93501

For discrete distributions, the plot of an object of class "fitdist" simply provides two goodness-of-fit plots comparing empirical and theoretical distributions in density and in CDF (Figure 11). Function `cdfcomp` can also be used to compare several plots to the same data set, as follows for the previous fits (Figure 12).

```
R>plot(ftoxo.P)
```

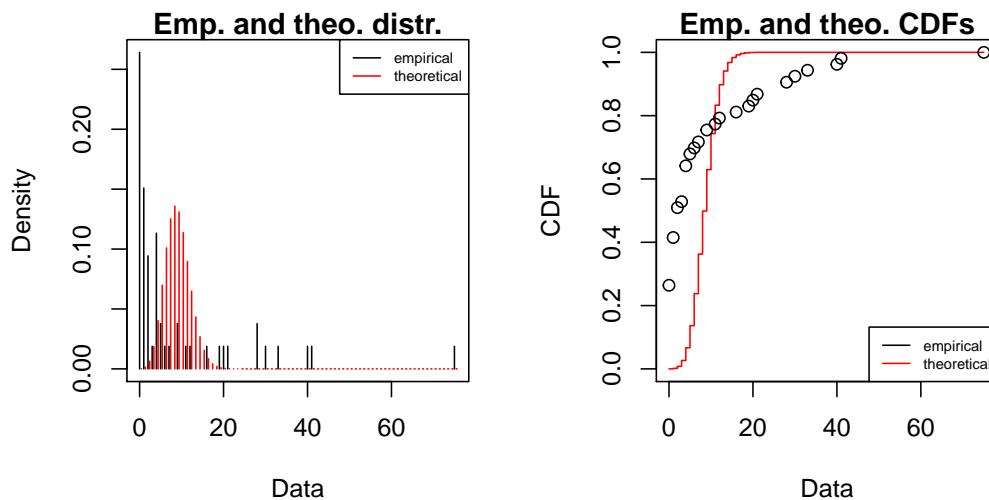


Figure 11: Plot of the fit of a discrete distribution (Poisson distribution fitted to numbers of *Toxocara cati* parasites from the `toxocara` data set.

```
R>cdfcomp(list(ftoxo.P,ftoxo.nb),
+ legendtext=c("Poisson", "negative binomial"))
```

When fitting discrete distributions, the Chi-squared statistic is computed by the `gofstat` function using cells defined by the argument `chisqbreaks` or cells automatically defined from the data in order to reach roughly the same number of observations per cell. This number is roughly equal to the argument `meancount`, or slightly greater if there are some ties. The

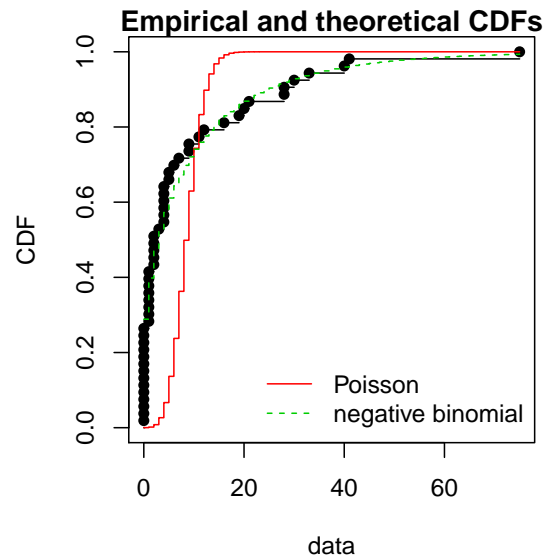


Figure 12: Comparison of the fits of a negative binomial and a Poisson distribution to numbers of *Toxocara cati* parasites from the `toxocara` data set).

choice to define cells from the empirical distribution (data), and not from the theoretical distribution, was done to enable the comparison of Chi-squared values obtained with different distributions fitted on a same data set. If arguments `chisqbreaks` and `meancount` are both omitted, `meancount` is fixed in order to obtain roughly  $(4n)^{2/5}$  cells, with  $n$  the length of the data set (Vose 2010). Using this default option the two previous fits are compared as follows, giving the preference to the negative binomial distribution, from both Chi-squared statistics and information criteria:

```
R>gofstat(list(ftoxo.P,ftoxo.nb),
+         fitnames = c("Poisson","negative binomial"))
```

Chi-squared statistic: 31257 7.486

Degree of freedom of the Chi-squared distribution: 5 4

Chi-squared p-value: 0 0.1123

the p-value may be wrong with some theoretical counts < 5

Chi-squared table:

	obscounts	theo Poisson	theo negative binomial
<= 0	14	0.009014	15.295
<= 1	8	0.078237	5.809
<= 3	6	1.321767	6.845
<= 4	6	2.131298	2.408
<= 9	6	29.827829	7.835
<= 21	6	19.626224	8.271
> 21	7	0.005631	6.537

Goodness-of-fit criteria

	Poisson	negative binomial
Aikake's Information Criterion	1017	322.7
Bayesian Information Criterion	1019	326.6

## 4. Conclusion

The R package **fitdistrplus** allows to easily fit distributions. Our main objective while developing this package was to provide tools for helping R users to fit distributions to data. We have been encouraged to pursue our work by feedbacks from users of our package in various areas as food or environmental risk assessment, epidemiology, ecology, molecular biology, genomics, bioinformatics, hydraulics, mechanics, financial and actuarial mathematics or operations research. Indeed, this package is already used by a lot of practionners and academics for simple MLE fits (Kohl and Ruckdeschel 2010; Jaloustre, Cornu, Morelli, Noel, and Delignette-Muller 2011; Sak and Haksoz 2011; Koch, Yemshanov, Magarey, and Smith 2012; Marquetoux, Paul, Wongnarkpet, Poolkhet, Thanapongtham, Roger, Ducrot, and Chalvet-Monfray 2012; Nordan 2012; Scholl, Nice, Fordyce, Gompert, and Forister 2012; Suuronen, Kallonen, Eik, Puttonen, Serimaa, and Herrmann 2012; González-Varo, López-Bao, and Guitián 2012; Mandl, Monteiro, Vriskoop, and Germain 2013), for MLE fits and goodness-of-fit statistics (Garcia 2012; Tarnczy, Fenyves, and Bcs 2011; Bagaria, Jaravine, Huang, Montelione, and Güntert 2012; Benavides-Piccione, Feraud-Espinosa, Robles, Yuste, and DeFelipe 2012; Breitbach, Böhning-Gaese, Laube, and Schleuning 2012; Poduje 2012; Pouillot and Delignette-Muller 2010), for MLE fits and bootstrap (Croucher, Harris, Barquist, Parkhill, and Bentley 2012; Méheust, Cann, Reponen, Wakefield, and Vesper 2012; Orellano, Reynoso, Grassi, Palmieri, Uez, and Carlino 2012; Tello, Austin, and Telfer 2012; Hoelzer, Pouillot, Gallagher, Silverman, Kause, and Dennis 2012), for MLE fits, bootstrap and goodness-of-fit statistics (Rebuge 2012), for MME fit (Luangkesorn, Norman, Zhuang, Falbo, and Sysko 2012), for censored MLE and bootstrap (Leha *et al.* 2011; Pouillot, Hoelzer, Chen, and Dennis 2012; Jongenburger, Reij, Boer, Zwietering, and Gorris 2012; Commeau *et al.* 2012; Pouillot, Delignette-Muller, and Cornu 2011a), for graphic analysing in (Anand, Yeturu, and Chandra 2012; Rosa 2012), for grouped-data fitting methods (Fu, Steiner, and Costafreda 2012) or more generally (Eling 2012; Busschaert *et al.* 2010; Brooks, Edwards, Sorrell, Srinivasan, and Diehl 2011; Aktaş and Sjöstrand 2011; Eik and Herrmann 2012; Bakos 2011). Many extensions of this package are planned in the future: we target to extend to censored data some methods for the moment only available for non-censored data, especially concerning goodness-of-fit evaluation and fitting methods. We will also enlarge the choice of fitting methods for non-censored data, by proposing new goodness-of-fit distances (e.g., distances based on quantiles) for maximum goodness-of-fit estimation and new types of moments (e.g., limited expected values) for moment matching estimation. At last, we will consider the case of multivariate distribution fitting.

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## References

- Aktaş O, Sjöstrand M (2011). *Cornish-Fisher Expansion and Value-at-Risk Method in Application to Risk Management of Large Portfolios*. Master's thesis, School of Information Science, Computer and Electrical Engineering, Halmstad University.
- Anand P, Yeturu K, Chandra N (2012). "PocketAnnotate: Towards Site-based Function Annotation." *Nucleic Acids Research*, **40**, 1–9.
- Bagaria A, Jaravine V, Huang Y, Montelione G, Güntert P (2012). "Protein Structure Validation by Generalized Linear Model Root-Mean-Square Deviation Prediction." *Protein Science*, **21**(2), 229–238.
- Bakos R (2011). *Poszméh Együttesek Összehasonlító Vizsgálata a Cserépfalusi fás Legelő különböző növényborítású területein*. Ph.D. thesis, Hungarian Veterinary Archive.
- Benavides-Piccione R, Fernaud-Espinosa I, Robles V, Yuste R, DeFelipe J (2012). "Age-based Comparison of Human Dendritic Spine Structure Using Complete Three-Dimensional Reconstructions." *Cerebral Cortex*. doi:10.1093/cercor/bhs154.
- Blom G (1959). *Statistical Estimates and Transformed Beta Variables*. 1st edition. John Wiley & Sons.
- Breitbach N, Böhning-Gaese K, Laube I, Schleuning M (2012). "Short Seed-Dispersal Distances and Low Seedling Recruitment in Farmland Populations of Bird-Dispersed Cherry Trees." *Journal of Ecology*, **100**(6), 1349–1358.
- Brooks J, Edwards D, Sorrell T, Srinivasan S, Diehl R (2011). "Simulating Calls for Service for an Urban Police Department." In *the 2011 Winter Simulation Conference*, pp. 1770–1777.
- Busschaert P, Geeraerd A, Uyttendaele M, VanImpe J (2010). "Estimating Distributions out of Qualitative and (Semi)Quantitative Microbiological Contamination Data for Use in Risk Assessment." *International Journal of Food Microbiology*, **138**, 260–269.
- Casella G, Berger R (2002). *Statistical Inference*. 2nd edition. Duxbury Thomson Learning.
- Commeau N, Parent E, Delignette-Muller ML, Cornu M (2012). "Fitting a Lognormal Distribution to Enumeration and Absence/Presence Data." *International Journal of Food Microbiology*, **155**, 146–152.
- Croucher NJ, Harris SR, Barquist L, Parkhill J, Bentley SD (2012). "A High-Resolution View of Genome-Wide Pneumococcal Transformation." *PLoS Pathogens*, **8**(6), e1002745.
- Cullen A, Frey H (1999). *Probabilistic Techniques in Exposure Assessment*. 1st edition. Plenum Publishing Co.
- D'Agostino R, Stephens M (1986). *Goodness-of-Fit Techniques*. 1st edition. Dekker.
- Delignette-Muller M, Pouillot R, Denis J, Dutang C (2013). *fitdistrplus: Help to Fit of a Parametric Distribution to Non-Censored or Censored Data*. R package version 1.0-1, URL <http://CRAN.R-project.org/package=fitdistrplus>.

- Delignette-Muller ML, Cornu M, AFSSA-STEC-Study-Group (2008). “Quantitative Risk Assessment for *Escherichia coli* O157:H7 in Frozen Ground Beef Patties Consumed by Young Children in French Households.” *International Journal of Food Microbiology*, **128**(1, SI), 158–164.
- Dutang C, Goulet V, Pigeon M (2008). “actuar: an R package for Actuarial Science.” *Journal of Statistical Software*, **25**(7), 1–37.
- Efron B, Tibshirani R (1994). *An Introduction to the Bootstrap*. 1st edition. Chapman & Hall.
- Eik M, Herrmann H (2012). “Raytraced Images for Testing the Reconstruction of Fibre Orientation Distributions.” In *the Estonian Academy of Sciences*, volume 61, pp. 128–136.
- Eling M (2012). “Fitting Insurance Claims to Skewed distributions: Are the Skew-normal and the Skew-student Good Models?” *Insurance: Mathematics and Economics*, **51**(2012), 239–248.
- Fromont E, Morvilliers L, Artois M, Pontier D (2001). “Parasite Richness and Abundance in Insular and Mainland Feral Cats: Insularity or Density?” *Parasitology*, **123**(Part 2), 143–151.
- Fu C, Steiner H, Costafreda S (2012). “Predictive Neural Biomarkers of Clinical Response in Depression: A Meta-Analysis of Functional and Structural Neuroimaging Studies of Pharmacological and Psychological Therapies.” *Neurobiology of Disease*, **52**, 75–83.
- Garcia P (2012). *Analyse Statistique des Pannes du Réseau HTA*. Master’s thesis, Université de Strasbourg.
- González-Varo J, López-Bao J, Guitián J (2012). “Functional Diversity Among Seed Dispersal Kernels Generated by Carnivorous Mammals.” *Journal of Animal Ecology*. doi:10.1111/1365-2656.12024.
- Goulet V (2012). *actuar: An R Package for Actuarial Science*. R package version 1.1-5, URL <http://CRAN.R-project.org/package=actuar>.
- Helsel D (2005). *Nondetects and Data Analysis: Statistics for Censored Environmental Data*. 1st edition. John Wiley & Sons.
- Hirano S, Clayton M, Upper C (1994). “Estimation of and Temporal Changes in Means and Variances of Populations of *Pseudomonas syringae* on Snap Bean Leaflets.” *Phytopathology*, **84**(9), 934–940.
- Hoelzer K, Pouillot R, Gallagher D, Silverman M, Kause J, Dennis S (2012). “Estimation of *Listeria Monocytogenes* Transfer Coefficients and Efficacy of Bacterial Removal Through Cleaning and Sanitation.” *International Journal of Food Microbiology*, **157**(2), 267–277.
- Hose G, Van den Brink P (2004). “Confirming the Species-Sensitivity Distribution Concept for Endosulfan Using Laboratory, Mesocosm, and Field Data.” *Archives of environmental contamination and toxicology*, **47**(4), 511–520.

- Hyndman R, Fan Y (1996). “Sample Quantiles in Statistical Packages.” *American Statistician*, **50**, 361–365.
- Jaloustre S, Cornu M, Morelli E, Noel V, Delignette-Muller M (2011). “Bayesian Modeling of *Clostridium perfringens* Growth in Beef-in-Sauce Products.” *Food microbiology*, **28**(2), 311–320.
- Jongenburger I, Reij M, Boer E, Zwietering M, Gorris L (2012). “Modelling Homogeneous and Heterogeneous Microbial Contaminations in a Powdered Food Product.” *International Journal of Food Microbiology*, **157**(1), 35–44.
- Jordan D (2005). “Simulating the Sensitivity of Pooled-sample Herd Tests for Fecal Salmonella in Cattle.” *Preventive Veterinary Medicine*, **70**(1-2), 59–73.
- Kefford B, Fields E, Clay C, Nuggeoda D (2007). “Salinity Tolerance of Riverine Macroinvertebrates from the Southern Murray-Darling Basin.” *Marine and Freshwater Research*, **58**, 1019–1031.
- Klein J, Moeschberger M (2003). *Survival Analysis: Techniques for Censored and Truncated Data*. 2nd edition. Springer-Verlag.
- Klugman S, Panjer H, Willmot G (2009). *Loss Models: from Data to Decisions*. 3rd edition. John Wiley & Sons.
- Koch F, Yemshanov D, Magarey R, Smith W (2012). “Dispersal of Invasive Forest Insects via Recreational Firewood: A Quantitative Analysis.” *Journal of Economic Entomology*, **105**(2), 438–450.
- Kohl M, Ruckdeschel P (2010). “R package distrMod: S4 Classes and Methods for Probability Models.” *Journal of Statistical Software*, **35**(10), 1–27.
- Leha A, Beissbarth T, Jung K (2011). “Sequential Interim Analyses of Survival Data in DNA Microarray Experiments.” *BMC Bioinformatics*, **12**(127), 1–14.
- Luangkesorn K, Norman B, Zhuang Y, Falbo M, Sysko J (2012). “Practice Summaries: Designing Disease Prevention and Screening Centers in Abu Dhabi.” *Interfaces*, **42**(4), 406–409.
- Luceno A (2006). “Fitting the Generalized Pareto Distribution to Data Using Maximum Goodness-of-fit Estimators.” *Computational Statistics and Data Analysis*, **51**(2), 904–917.
- Mandl J, Monteiro J, Vrisekoop N, Germain R (2013). “T Cell-Positive Selection Uses Self-Ligand Binding Strength to Optimize Repertoire Recognition of Foreign Antigens.” *Immunity*, **38**(2), 263–274.
- Marquetoux N, Paul M, Wongnarkpet S, Poolkhet C, Thanapongtham W, Roger F, Ducrot C, Chalvet-Monfray K (2012). “Estimating Spatial and Temporal Variations of the Reproduction Number for Highly Pathogenic Avian Influenza H5N1 Epidemic in Thailand.” *Preventive Veterinary Medicine*, **106**(2), 143–151.
- McNeil A (1997). “Estimating the Tails of Loss Severity Distributions Using Extreme Value Theory.” *ASTIN Bulletin*, **27**(1), 117–137.



- Méheust D, Cann PL, Reponen T, Wakefield J, Vesper S (2012). “Possible Application of the Environmental Relative Moldiness Index in France: a Pilot Study in Brittany.” *International Journal of Hygiene and Environmental Health*, **216**(3), 333–340.
- Nordan R (2012). *An Investigation of Potential Methods for Topology Preservation in Interactive Vector Tile Map Applications*. Master’s thesis, Norwegian University of Science and Technology.
- Orellano P, Reynoso J, Grassi A, Palmieri A, Uez O, Carlino O (2012). “Estimation of the Serial Interval for Pandemic Influenza (pH1N1) in the Most Southern Province of Argentina.” *Iranian Journal of Public Health*, **41**(12), 26–29.
- Poduje AC (2012). *Bivariate Analysis and Synthesis of Flood Events for the Design of Hydraulic Structures – a Case Study for Argentina*. Master’s thesis, Leibniz Universität Hannover.
- Posthuma L, Suter G, Traas T (2010). *Species Sensitivity Distributions in Ecotoxicology*. Environmental and Ecological Risk Assessment Series. Taylor & Francis.
- Pouillot R, Delignette-Muller M (2010). “Evaluating Variability and Uncertainty Separately in Microbial Quantitative Risk Assessment using two R Packages.” *International Journal of Food Microbiology*, **142**(3), 330–340.
- Pouillot R, Delignette-Muller M, Cornu M (2011a). “Case study: *L. monocytogenes* in Cold-Smoked Salmon.” URL <http://CRAN.R-project.org/web/packages/mc2d/vignettes/mc2dLmEnglish.pdf>.
- Pouillot R, Delignette-Muller M, Denis J (2011b). *mc2d: Tools for Two-Dimensional Monte-Carlo Simulations*. R package version 0.1-12, URL <http://CRAN.R-project.org/package=mc2d>.
- Pouillot R, Hoelzer K, Chen Y, Dennis S (2012). “Estimating Probability Distributions of Bacterial Concentrations in Food Based on Data Generated Using the Most Probable Number (MPN) Method for Use in Risk Assessment.” *Food Control*, **29**(2), 350–357.
- R Development Core Team (2013). *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL <http://www.R-project.org/>.
- Rebuge A (2012). *Business Process Analysis in Healthcare Environments*. Master’s thesis, Universidade Tecnica de Lisboa.
- Ricci V (2005). “Fitting Distributions with **R**.” Contributed Documentation available on CRAN, URL <http://CRAN.R-project.org/doc/contrib/Ricci-distributions-en.pdf>.
- Rosa AS (2012). *Funções de Predição Espacial de Propriedades do Solo*. Master’s thesis, Universidade Federal de Santa Maria.
- Sak H, Haksoz C (2011). “A Copula-based Simulation Model for Supply Portfolio Risk.” *Journal of Operational Risk*, **6**(3), 15–38.

- Scholl C, Nice C, Fordyce J, Gompert Z, Forister M (2012). “Larval Performance in the Context of Ecological Diversification and Speciation in *Lycaeides* Butterflies.” *International Journal of Ecology*. doi:[10.1155/2012/242154](https://doi.org/10.1155/2012/242154).
- Suuronen J, Kallonen A, Eik M, Puttonen J, Serimaa R, Herrmann H (2012). “Analysis of Short Fibres Orientation in Steel Fibre-Reinforced Concrete (SFRC) by X-ray Tomography.” *Journal of Materials Science*, **48**(3), 1358–1367.
- Tarnoczy T, Fenyves V, Bcs Z (2011). “The Business Uncertainty and Variability Management with Real Options Models Combined two Dimensional Simulation.” *International Journal of Management Cases*, **13**(3), 159–167.
- Tello A, Austin B, Telfer T (2012). “Selective Pressure of Antibiotic Pollution on Bacteria of Importance to Public Health.” *Environmental Health Perspectives*, **120**(8), 1100–1106.
- Therneau T (2011). *survival: Survival Analysis, Including Penalized Likelihood*. R package version 2.36-9, URL <http://CRAN.R-project.org/package=survival>.
- Tse Y (2009). *Nonlife Actuarial Models: Theory, Methods and Evaluation*. International Series on Actuarial Science, 1st edition. Cambridge University Press.
- Turnbull B (1974). “Nonparametric Estimation of a Survivorship Function with Doubly Censored Data.” *Journal of the American Statistical Association*, **69**(345), 169–173.
- Venables WN, Ripley BD (2010). *Modern Applied Statistics with S*. 4th edition. Springer-Verlag.
- Vose D (2010). *Quantitative Risk Analysis. A Guide to Monte Carlo Simulation Modelling*. 1st edition. John Wiley & Sons.

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