

# SeqinR 1.0-7

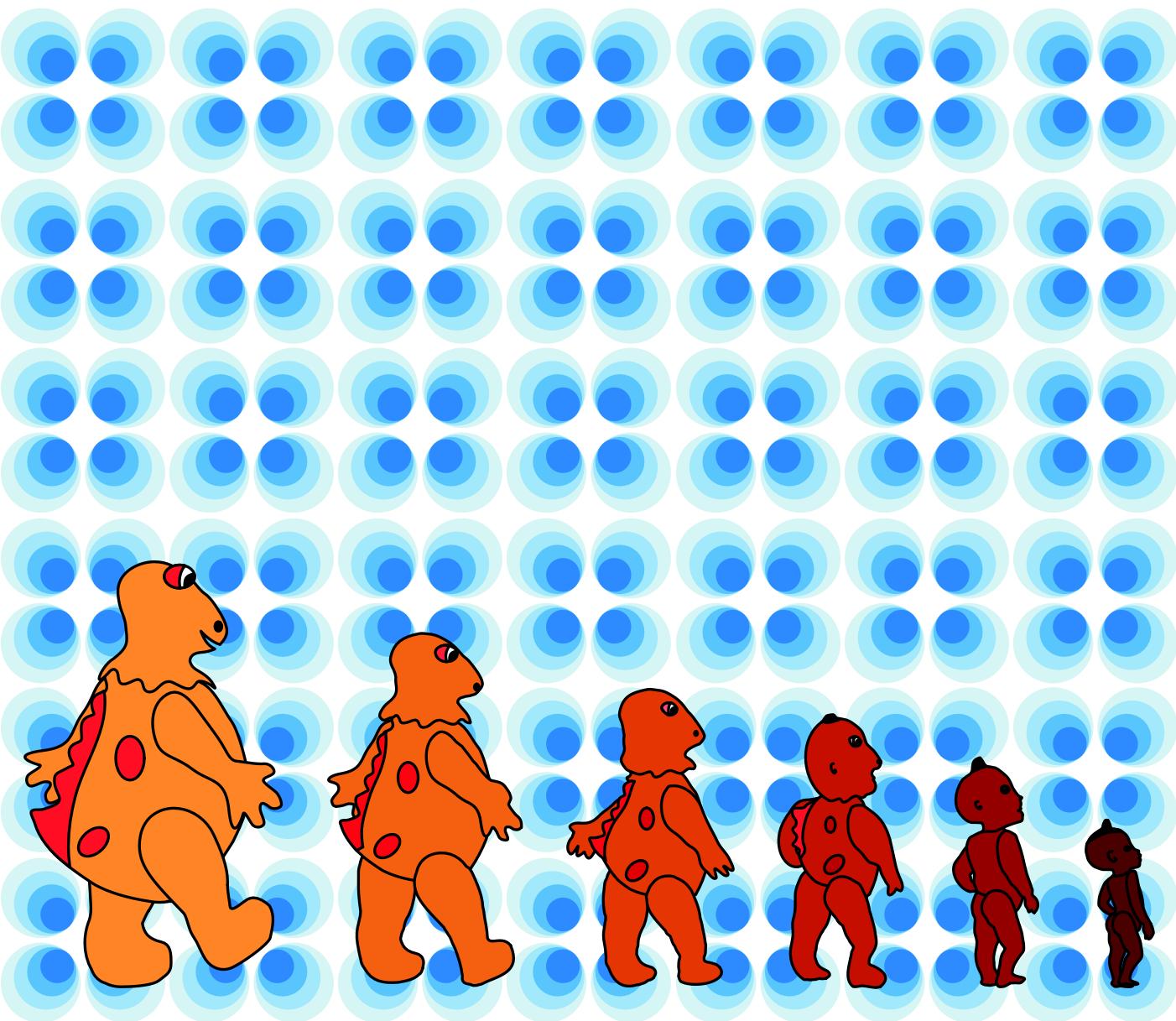




Figure 1: The march of progress icon is very common in popular press. This example is from page 46 of a 1984 summer issue of the tchek edition of *Playboy*.

## The march of progress icon

The cover, an artwork created<sup>1</sup> by Lionel Humblot, is an allusion to what Stephen J. Gould considered as a canonical icon of "[t]he most serious and pervasive of all misconceptions about evolution equates the concept with some notion of progress, usually inherent and predictable, and leading to a human pinnacle" [16]. Some examples of the so-called "march of progress icon" out of hundreds in S.J. Gould's collection from popular press are given in the beginning of his famous book *Wonderful life* [15].

Note that the underlying conception predates Darwin [41]. We know now that evolution does not equal progress, and this is illustrated here in the cover by the unusual **decreasing** size from the initial character (on the left) to the last one (on the right).



*L'île aux enfants.*

## The character on the left

The character on the left is called Casimir, the cult character of the French TV show *l'île aux enfants* (literally Kid's island, a French adaptation of *Sesame Street* from 1974 to 1975 and then an autonomous production until 1982 when it eventually ended). Casimir was a muppet, human-sized, with an actor playing inside, representing an orange dinosaur (the exact taxonomy has never been published) with yellow and red spots. Casimir was symbolically chosen here for two reasons. First, its birth corresponds to one of the earliest paper from our

<sup>1</sup> with Canvas from ACD Systems.

lab about molecular evolution [20]. If you dig into `seqinR` you will find that the data from this more than 30 years old paper are still available<sup>2</sup>:

```
data(aaindex)
grth <- which(sapply(aaindex, function(x) length(grep("Grantham",
  x$A)) != 0))
lapply(aaindex[grth], "[[", "D")

$GRAR740101
[1] "Composition (Grantham, 1974)"

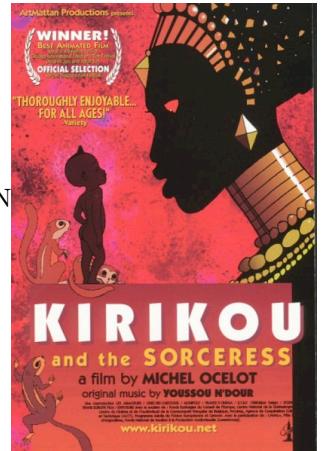
$GRAR740102
[1] "Polarity (Grantham, 1974)"

$GRAR740103
[1] "Volume (Grantham, 1974)"
```

Second, Casimir's life span correspond more or less to the time during which the sequence analysis software called ANALSEQ<sup>3</sup> [26] was under development in our lab. ANALSEQ has never been published as a regular paper (although it is mentioned in one of the ACNUC paper [19]), there is only a reference manual in french [26] also available on-line at <http://biomserv.univ-lyon1.fr/doclogi/docanalsmanuel.html>. ANALSEQ was entirely written in FORTRAN 77, and although you won't find any fossil code from it within `seqinR`, we wanted to credit symbolically ANALSEQ as a kind of spiritual ancestor of `seqinR` with the cover.

## The character on the right

The character on the right is called Kirikou. He is the main character of the animated film *Kirikou et la sorcière* (Kirikou and the sorceress, 1998) and *Kirikou et les bêtes sauvages* (Kirikou and the Wild Beasts, 2005). Kirikou was chosen as a symbol of `seqinR` development time. `SeqinR` started in september 2002 as part of the work of Delphine Charif's master of sciences. The first public presentation of `seqinR` was a seminar (2-JUL-2003, Lausanne University, Swiss) and the first public release on the CRAN<sup>4</sup> was in october 2004.



Kirikou and the sorceress, a film by Michel Ocelot with original music by Youssou N'Dour.

## Technical details

The cover was saved from Canvas into an EPS<sup>5</sup> file. This file was then manually edited to remove non-ASCII characters. It was then converted into RGML<sup>6</sup> format with the following `R` code based on `grid` [50], `XML` [7] and `grImport` [43]:

```
library(grid)
library(XML)
library(grImport)
PostScriptTrace("../figs/couverture.eps", ".../figs/couverture.rgml")
```

<sup>2</sup> thanks to `aaindex` database [30, 61, 44].

<sup>3</sup> not to be confused with the ANALYSEQ program by Rodger Staden [58].

<sup>4</sup> Comprehensive R Archive Network.

<sup>5</sup> Encapsulated Postscript.

<sup>6</sup> RDF (Resource Description Framework) Graph Modeling Language (<http://www.cs.rpi.edu/~puninj/RGML/>).

The picture was then edited to add automatically the current **seqinR** release number:

```
cover <- readPicture("../figs/couverture.rgml")
pdf(file = "../figs/cover.pdf", width = 21/2.54, height = 29.7/2.54)
pushViewport(plotViewport(margins = c(0, 0, 0, 0)))
grid.picture(cover)
grid.text(paste("SeqinR", packageDescription("seqinr")$Version),
          gp = gpar(cex = 5), y = unit(0.72, "npc"))
popViewport()
dev.off()
```

And finally inserted at the begining of the L<sup>A</sup>T<sub>E</sub>X file with:

```
\atxy(0cm,0cm){
  \includegraphics[width=\paperwidth,height=\paperheight]{../figs/cover}
}
```

## Session Informations

This part was compiled under the following  environment:

- R version 2.4.1 (2006-12-18), i386-apple-darwin8.8.1
- Locale: C
- Base packages: base, datasets, grDevices, graphics, grid, methods, stats, utils
- Other packages: MASS 7.2-30, XML 1.4-1, ade4 1.4-2, ape 1.9-3, gee 4.13-12, grImport 0.2-2, lattice 0.14-16, nlme 3.1-78, seqinr 1.0-7, xtable 1.4-3

**SeqinR** 1.0-7: a contributed package to the  
 project for statistical computing devoted to  
biological sequences retrieval and analysis

Charif, D.      Humblot, L.      Lobry, J.R.      Necșulea, A.  
Palmeira, L.      Penel, S.

April 15, 2007



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# CHAPTER 1

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## Introduction

Lobry, J.R.

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### 1.1 About ACNUC

ACNUC<sup>1</sup> was first a database of nucleic acids developed in the early 80's in the same lab (Lyon, France) that issued **seqinR**. ACNUC was first published as a printed book in two volumes [13, 14] whose covers are reproduced in margin there. At about the same time, two other databases were created, one in the USA (GenBank, at Los Alamos and now managed by the NCBI<sup>2</sup>), and another one in Germany (created in Köln by K. Stüber). To avoid duplication of efforts at the european level, a single repository database was initiated in Germany yielding the EMBL<sup>3</sup> database that moved from Köln to Heidelberg, and then to its current location at the EBI<sup>4</sup> near Cambridge. The DDBJ<sup>5</sup> started in 1986

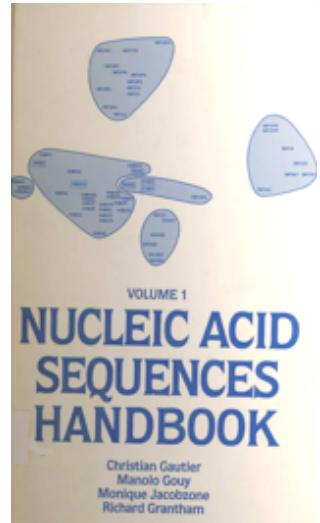
<sup>1</sup> A contraction of ACides NUCléiques, that is *NUCleic ACids* in french (<http://pbil.univ-lyon1.fr/databases/acnuc/acnuc.html>)

<sup>2</sup>National Center for Biotechnology Information

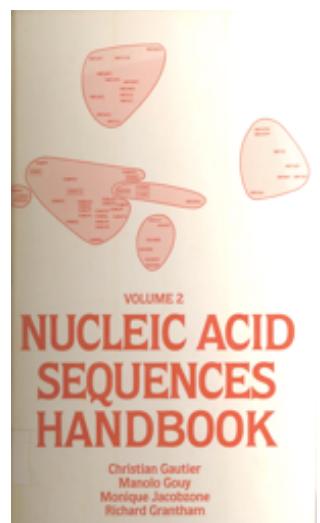
<sup>3</sup>European Molecular Biology Laboratory

<sup>4</sup>European Bioinformatic Institute

<sup>5</sup>DNA Data Bank of Japan



Cover of ACNUC book vol. 1



Cover of ACNUC book vol. 2

at the NIG<sup>6</sup> in Mishima. These three main repository DNA databases are now collaborating to maintain the INSD<sup>7</sup> and are sharing data on a daily basis.

The sequences present in the ACNUC books [13, 14] were all the published nucleic acid sequences of about 150 or more continuous unambiguous nucleotides up to May or June 1981 from the journal given in table 1.1.

Journal name
<i>Biochimie</i>
<i>Biochemistry (ACS)</i>
<i>Cell</i>
<i>Comptes Rendus de l'Académie des Sciences, Paris</i>
<i>European Journal of Biochemistry</i>
<i>FEBS Letters</i>
<i>Gene</i>
<i>Journal of Bacteriology</i>
<i>Journal of Biological Chemistry</i>
<i>Journal of Molecular Biology</i>
<i>Molecular and General Genetics</i>
<i>Nature</i>
<i>Nucleic Acids Research</i>
<i>Proceedings of the National Academy of Sciences of the United States of America</i>
<i>Science</i>

Table 1.1: The list of journals that were manually scanned for nucleic sequences that were included in the ACNUC books [13, 14]



ACNUC books are about 4.5 cm width

The total number of base pair was 526,506 in the two books. They were about 4.5 cm width. We can then compute of much place would it take to print the last GenBank release with the same format as the ACNUC book:

```
acnucbooksize <- 4.5
acnucbp <- 526506
mybank <- choosebank("genbank")
mybank$details
```

[1] "GenBank Rel. 158 (15 February 2007) Last Updated: Apr 13, 2007"
[2] "75,756,334,808 bases; 71,808,730 sequences; 3,939,297 subseqs; 462,456 refers."
[3] "Software by M. Gouy, Lab. Biometrie et Biologie Evolutive, Universite Lyon I"

```
bpbk <- unlist(strsplit(mybank$details[2], split = " "))[1]
bpbk
```

```
[1] "75,756,334,808"
```

```
bpbk <- as.numeric(paste(unlist(strsplit(bpbk, split = ",")),
collapse = ""))
```

```
widthcm <- acnucbooksize * bpbk/acnucbp
```

```
(widthkm <- widthcm/10^5)
```

```
[1] 6.474827
```

It would be about 6.5 kilometer long in ACNUC book format to print to print GenBank today (April 15, 2007).

<sup>6</sup>National Institute of Genetics

<sup>7</sup> International Nucleotide Sequence Database (<http://www.insdc.org/>)

## 1.2 About R and CRAN

R [25, 51] is a *libre* language and environment for statistical computing and graphics which provides a wide variety of statistical and graphical techniques: linear and nonlinear modelling, statistical tests, time series analysis, classification, clustering, etc. Please consult the R project homepage at <http://www.R-project.org/> for further information.

The Comprehensive R Archive Network, CRAN, is a network of servers around the world that store identical, up-to-date, versions of code and documentation for R. At compilation time of this document, there were 70 mirrors available from 29 countries. Please use the CRAN mirror nearest to you to minimize network load, they are listed at <http://cran.r-project.org/mirrors.html>, and can be directly selected with the function `chooseCRANmirror()`.

## 1.3 About this document

In the terminology of the R project [25, 51], this document is a package *vignette*. The examples given thereafter were run under R version 2.4.1 (2006-12-18) on Fri Apr 13 11:29:57 2007 with Sweave [35]. The last compiled version of this document is distributed along with the **seqinR** package in the `/doc` folder. Once **seqinR** has been installed, the full path to the package is given by the following R code :

```
.find.package("seqinr")
[1] "/Users/lobry/Library/R/library/seqinr"
```

## 1.4 About sequin and seqinR

Sequin is the well known software used to submit sequences to GenBank, **seqinR** has definitively no connection with sequin. **seqinR** is just a shortcut, with no google hit, for "Sequences in R".

However, as a mnemotechnic tip, you may think about the **seqinR** package as the **Reciprocal** function of sequin: with sequin you can submit sequences to Genbank, with **seqinR** you can **Retrieve** sequences from Genbank. This is a very good summary of a major functionality of the **seqinR** package: to provide an efficient access to sequence databases under R.

## 1.5 About getting started

You need a computer connected to the Internet. First, install R on your computer. There are distributions for Linux, Mac and Windows users on the CRAN (<http://cran.r-project.org>). Then, install the **ape**, **ade4** and **seqinr** packages. This can be done directly in an R console with for instance the command `install.packages("seqinr")`. Last, load the **seqinR** package with:

```
library(seqinr)
```

The command `lseqinr()` lists all what is defined in the package **seqinR**:

```
lseqinr() [1:9]
```

[1] "AAstat"	"EXP"	"GC"	"GC1"	"GC2"
[6] "GC3"	"SEQINR.UTIL"	"a"	"aaa"	

We have printed here only the first 9 entries because they are too numerous. To get help on a specific function, say `aaa()`, just prefix its name with a question mark, as in `?aaa` and press enter.

## 1.6 About running R in batch mode

Although R is usually run in an interactive mode, some data pre-processing and analyses could be too long. You can run your R code in batch mode in a shell with a command that typically looks like :

```
unix$ R CMD BATCH input.R results.out &
```

where `input.R` is a text file with the R code you want to run and `results.out` a text file to store the outputs. Note that in batch mode, the graphical user interface is not active so that some graphical devices (*e.g.* `x11`, `jpeg`, `png`) are not available (see the R FAQ [23] for further details).

It's worth noting that R uses the XDR representation of binary objects in binary saved files, and these are portable across all R platforms. The `save()` and `load()` functions are very efficient (because of their binary nature) for saving and restoring any kind of R objects, in a platform independent way. To give a striking real example, at a given time on a given platform, it was about 4 minutes long to import a numeric table with 70000 lines and 64 columns with the defaults settings of the `read.table()` function. Turning it into binary format, it was then about 8 *seconds* to restore it with the `load()` function. It is therefore advisable in the `input.R` batch file to save important data or results (with something like `save(mybigdata, file = "mybigdata.RData")`) so as to be able to restore them later efficiently in the interactive mode (with something like `load("mybigdata.RData")`).

## 1.7 About the learning curve

### Introduction

If you are used to work with a purely graphical user interface, you may feel frustrated in the beginning of the learning process because apparently simple things are not so easily obtained (*ce n'est que le premier pas qui coûte !*). In the long term, however, you are a winner for the following reasons.

#### 1.7.1 Wheel (the)

Do not re-invent (there's a patent [31] on it anyway). At the compilation time of this document there were 960 contributed packages available. Even if you don't want to be spoon-feed *à bouche ouverte*, it's not a bad idea to look around there just to check what's going on in your own application field. Specialists all around the world are there.

### 1.7.2 Hotline

There is a very reactive discussion list to help you, just make sure to read the posting guide there: <http://www.R-project.org/posting-guide.html> before posting. Because of the high traffic on this list, we strongly suggest to answer *yes* at the question *Would you like to receive list mail batched in a daily digest?* when subscribing at <https://stat.ethz.ch/mailman/listinfo/r-help>. Some *bons mots* from the list are archived in the  `fortunes` package.

### 1.7.3 Automation

Consider the 178 pages of figures in the additional data file 1 (<http://genomebiology.com/2002/3/10/research/0058/suppl/S1>) from [40]. They were produced in part automatically (with a proprietary software that is no more maintained) and manually, involving a lot of tedious and repetitive manipulations (such as italicising species names by hand in subtitles). In few words, a waste of time. The advantage of the  environment is that once you are happy with the outputs (including graphical outputs) of an analysis for species x, it's very easy to run the same analysis on n species.

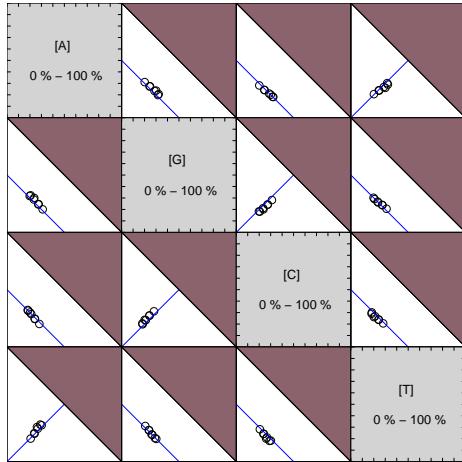
### 1.7.4 Reproducibility

If you do not consider the reproducibility of scientific results to be a serious problem in practice, then the paper by Jonathan Buckheit and David Donoho [3] is a must read. Molecular data are available in public databases, this is a necessary but not sufficient condition to allow for the reproducibility of results. Publishing the R source code that was used in your analyses is a simple way to greatly facilitate the reproduction of your results at the expense of no extra cost. At the expense of a little extra cost, you may consider to set up a RWeb server so that even the laziest reviewer may reproduce your results just by clicking on the "do it again" button in his web browser (*i.e.* without installing any software on his computer). For an example involving the `seqinR` pacakage, follow this link <http://pbil.univ-lyon1.fr/members/lobry/repro/bioinfo04/> to reproduce on-line the results from [4].

### 1.7.5 Fine tuning

You have full control on everything, even the source code for all functions is available. The following graph was specifically designed to illustrate the first experimental evidence [52] that, on average, we have also  $[A]=[T]$  and  $[C]=[G]$  in single-stranded DNA. These data from Chargaff's lab give the base composition of the L (Ligth) strand for 7 bacterial chromosomes.

```
example(chargaff, ask = FALSE)
```



This is a very specialised graph. The filled areas correspond to non-allowed values because the sum of the four bases frequencies cannot exceed 100 %. The white areas correspond to possible values (more exactly to the projection from  $\mathbb{R}^4$  to the corresponding  $\mathbb{R}^2$  planes of the region of allowed values). The lines correspond to the very small subset of allowed values for which we have in addition  $[A]=[T]$  and  $[C]=[G]$ . Points represent observed values in the 7 bacterial chromosomes. The whole graph is entirely defined by the code given in the example of the `chargaff` dataset (`?chargaff` to see it).

Another example of highly specialised graph is given by the function `tablecode()` to display a genetic code as in textbooks :

```
tablecode()
```

Genetic code 1 : standard									
TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys		
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys		
TTA	Leu	TCA	Ser	TAA	Stop	TGA	Stop		
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp		
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg		
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg		
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg		
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg		
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser		
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser		
ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg		
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg		
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly		
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly		
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly		
GTC	Val	GCG	Ala	GAG	Glu	GGG	Gly		

It's very convenient in practice to have a genetic code at hand, and moreover here, all genetic code variants are available :

```
tablecode(numcode = 2)
```

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	TGA	Trp
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Thr	CCT	Pro	CAT	His	CGT	Arg
CTC	Thr	CCC	Pro	CAC	His	CGC	Arg
CTA	Thr	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Thr	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Met	ACA	Thr	AAA	Lys	AGA	Arg
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table 1.2: Genetic code number 3: yeast.mitochondrial.

<b>Genetic code 2 : vertebrate.mitochondrial</b>							
TTT	Phe	TCT	Ser	TAT	Tyr	T GT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	T GC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	T GA	Trp
TTG	Leu	TCG	Ser	TAG	Stop	T GG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Met	ACA	Thr	AAA	Lys	AGA	Stop
ATG	Met	ACG	Thr	AAG	Lys	AGG	Stop
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

As from **seqinR** 1.0-4, it is possible to export the table of a genetic code into a L<sup>A</sup>T<sub>E</sub>X document, for instance table 1.2 and table 1.3 were automatically generated with the following R code:

```
tablecode(numcode = 3, latexfile = "../tables/code3.tex",
          size = "small")
tablecode(numcode = 4, latexfile = "../tables/code4.tex",
          size = "small")
```

The tables were then inserted in the L<sup>A</sup>T<sub>E</sub>X file with:

```
\input{../tables/code3.tex}
```

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	TGA	Trp
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table 1.3: Genetic code number 4: protozoan.mitochondrial+mycoplasma.

\input{../tables/code4.tex}

### 1.7.6 Data as fast moving targets

In research area, data are not always stable. Consider figure 1 from [37] which is reproduced here in figure 1.1. Data have been updated since then, but we can re-use the same  code<sup>8</sup> to update the figure:

```

data <- get.db.growth()
scale <- 1
ltymoore <- 1
date <- data$date
Nucleotides <- data$Nucleotides
Month <- data$Month
plot.default(date, log10(Nucleotides), main = "Update of Fig. 1 from Lobry (2004) LNCS, 3039:679:\nThe e",
            xlab = "Year", ylab = "Log10 number of nucleotides", pch = 19,
            las = 1, cex = scale, cex.axis = scale, cex.lab = scale)
abline(lm(log10(Nucleotides) ~ date), lwd = 2)
lm1 <- lm(log(Nucleotides) ~ date)
mu <- lm1$coef[2]
dbt <- log(2)/mu
dbt <- 12 * dbt
x <- mean(date)
y <- mean(log10(Nucleotides))
a <- log10(2)/1.5
b <- y - a * x
lm10 <- lm(log10(Nucleotides) ~ date)
for (i in seq(-10, 10, by = 1)) if (i != 0) abline(coef = c(b +
  i, a), col = "black", lty = ltymoore)

```

<sup>8</sup> This code was adapted from <http://pbil.univ-lyon1.fr/members/lobry/repro/lncs04/>.

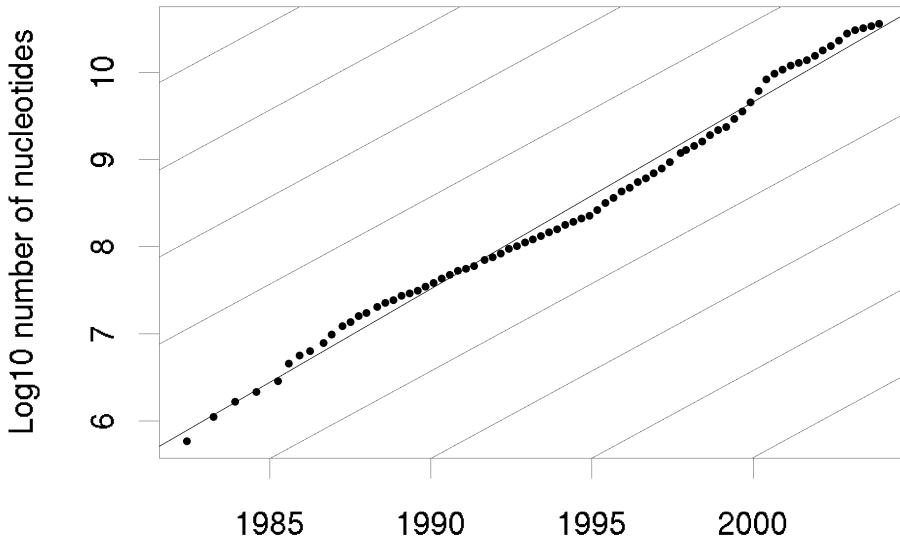
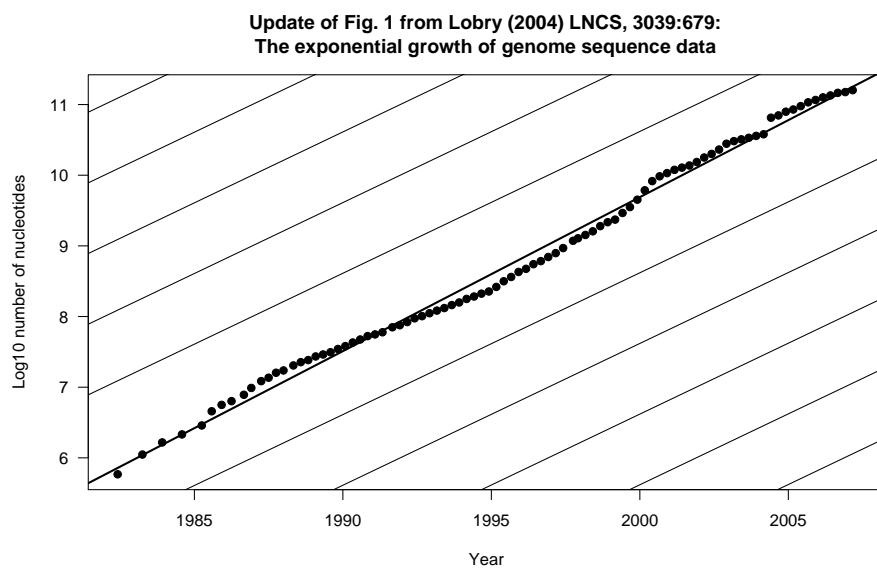


Figure 1.1: Screenshot of figure 1 from [37]. The exponential growth of genomic sequence data mimics Moore's law. The source of data is the december 2003 release note (realnote.txt) from the EMBL database available at <http://www.ebi.ac.uk/>. External lines correspond to what would be expected with a doubling time of 18 months. The central line through points is the best least square fit, corresponding to a doubling time of 16.9 months.



The doubling time is now 16.6 months.

### 1.7.7 `Sweave()` and `xtable()`

For L<sup>A</sup>T<sub>E</sub>X users, it's worth mentioning the fantastic tool contributed by Friedrich Leisch [35] called `Sweave()` that allows for the automatic insertion of R outputs (including graphics) in a L<sup>A</sup>T<sub>E</sub>X document. In the same spirit, there is a package called `xtable` [5] to coerce R data into L<sup>A</sup>T<sub>E</sub>X tables.

## 1.8 Session Informations

This part was compiled under the following R environment:

- R version 2.4.1 (2006-12-18), i386-apple-darwin8.8.1
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: MASS 7.2-30, ade4 1.4-2, ape 1.9-3, gee 4.13-12, lattice 0.14-16, nlme 3.1-78, seqinr 1.0-7, xtable 1.4-3

---

---

# CHAPTER 2

---

## Importing sequences from flat files

Charif, D. Lobry, J.R.

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---

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---

### 2.1 Importing raw sequence data from FASTA files

#### 2.1.1 FASTA files examples

The FASTA format is very simple and widely used for simple import of biological sequences. It was used originally by the FASTA program [47]. It begins with a single-line description starting with a character >, followed by lines of sequence data of maximum 80 character each. Examples of files in FASTA format are distributed with the `seqinR` package in the `sequences` directory:

```
list.files(path = system.file("sequences", package = "seqinr"),
           pattern = ".fasta")

[1] "ATH1_pep_cm_20040228.fasta" "Anouk.fasta"
[3] "bb.fasta"                   "bordetella.fasta"
[5] "ct.fasta"                  "ecolicgpe5.fasta"
[7] "gopher.fasta"              "louse.fasta"
[9] "malM.fasta"                "ortho.fasta"
[11] "seqAA.fasta"               "smallAA.fasta"
```

Here is an example of a FASTA file:

```
cat(readLines(system.file("sequences/seqAA.fasta", package = "seqinr")),
    sep = "\n")
```

```
>AO6852          183 residues
MPLRFSYLLGVWLQLSQLPREIPGQSTNDFIKACGRRELVRLWVEICGSVSWGRTLSLEE
PQLETGPPAETMPSSITKDAEILKMMLEFVNLPQELKATLSERQPSLRELQQSASKDSN
LNFEEFKKIILNRQNEAEDKSLLELKNLGLDKHSRKRLFRMTLSEKCCQVGCIRKDIAR
LC*
```

### 2.1.2 The function `read.fasta()`

The function `read.fasta()` imports sequences from FASTA files into your workspace.

#### DNA file example

The example file looks like:

```
dnafile <- system.file("sequences/malM.fasta", package = "seqinr")
cat(readLines(dnafile), sep = "\n")
```

```
>XYLEECOM.MALM 921 bp ACCESSION E00218, X04477
ATGAAAATGAATAAAAGTCTCATCGCCTCTGTTATCAGCAGGGTTACTGGCAAGCGCG
CTCTGGAATTAGCCTTGCGATGTTAACTACGTACCGCAAACACCCAGCGACGCCAGCC
ATTCATCTGCTGGCTGCAACAACACTCACCTGGACACCGCTCGATCAATCTAAACCCAG
ACCAACCAACTGGCAGCCGGCAACAACTGAACGTTCCCGGCATCAGTGGTCCGGTT
GCTCGTCAAGCGCTCCGGAAACATTGCGGAACTGACCCCTGACCCGTGACCCAGGAAAGTG
AAACAACAAACACCGGTTTGCCTGGCAACGTGCTGATTCTTGATCAGAACATGACCCCA
TCAGCCTCTCCCCAGCAGTTATTCACCTACCAGAACCGAGCTGATGAGTCAGAT
CGGCTGGAAGGGCTTATGGCCTGACACCCGGCTGGGGCAGCAAAAACTTTATGTTCTG
GTCTTTACACGGAAAAGATCTCCAGCACACGCCAACTGCTGACCCGGCTAAAGCC
TATGCCAAGGGCTCGGTAACTCGATCCCGGATATCCCCGATCCTGGTCTCGTCATACC
ACCGATGGCTACTGAAAGTGAACAGAACACTCCAGCTCCAGCTGGTCTGGTAGGA
CCCTTATTGGTCTCCGCTCCAGCTCCGGTTAGGTAGGTAACACGGCGCACAGCT
GTGGCTGACCCGCTCCGGCACCGGTAGAAGAAAGCAGGGCATGCTAACGACACGGAA
AGTATTTAAACCGCGATCAAACAGCTGTCGGAAAGGTGATGTTGATAAGGGTTA
AAACTGCTGATGAAGCTGAACGCTGGGATCGACATCTGCCGTTCCACCTTATCAGC
AGTAAAGGCAAGGGTAA
```

With default arguments the output looks like:

```
read.fasta(file = dnafile)
```

```
$XYLEECOM.MALM
[1] "a" "t" "g" "a" "a" "a" "a" "t" "g" "a" "a" "t" "a" "a" "a" "g" "t"
[19] "c" "t" "c" "a" "t" "c" "g" "t" "c" "t" "c" "t" "g" "t" "t" "t" "t" "a"
[37] "t" "c" "a" "g" "c" "a" "g" "g" "t" "t" "a" "c" "t" "g" "g" "c" "a" "a"
[55] "a" "g" "c" "g" "c" "a" "g" "g" "t" "g" "g" "a" "a" "t" "t" "a" "g" "c"
[73] "c" "t" "t" "g" "c" "c" "g" "a" "t" "g" "t" "t" "a" "a" "c" "t" "a" "c"
[91] "g" "t" "a" "c" "c" "g" "c" "a" "a" "a" "c" "a" "c" "c" "a" "g" "c" "c"
[109] "g" "a" "c" "g" "c" "g" "c" "a" "a" "g" "c" "c" "a" "a" "t" "t" "c" "c" "a"
[127] "t" "t" "c" "g" "t" "g" "c" "g" "t" "t" "g" "c" "a" "a" "c" "a" "a" "a"
[145] "c" "t" "c" "a" "c" "c" "t" "g" "g" "t" "g" "a" "c" "a" "c" "c" "g" "t" "c"
[163] "g" "a" "t" "c" "a" "a" "t" "c" "t" "a" "a" "a" "a" "c" "c" "c" "a" "g"
[181] "a" "c" "c" "a" "c" "c" "a" "a" "c" "t" "g" "g" "c" "g" "a" "c" "c" "c"
[199] "g" "g" "c" "g" "g" "c" "c" "a" "a" "a" "c" "a" "a" "t" "g" "a" "a" "c"
[217] "g" "t" "t" "c" "c" "c" "g" "g" "t" "c" "a" "g" "t" "g" "g" "g" "t"
[235] "c" "c" "g" "g" "t" "t" "g" "c" "t" "g" "c" "g" "t" "a" "c" "a" "g" "c"
[253] "g" "t" "c" "c" "c" "g" "g" "c" "a" "a" "a" "c" "a" "t" "t" "g" "g" "c"
[271] "g" "a" "a" "c" "t" "g" "a" "c" "c" "t" "g" "a" "c" "g" "c" "t" "t" "g"
[289] "a" "c" "c" "a" "g" "c" "g" "a" "a" "g" "t" "g" "a" "a" "c" "a" "a" "a"
[307] "c" "a" "a" "a" "c" "c" "a" "g" "c" "g" "t" "t" "t" "t" "g" "c" "g"
[325] "c" "c" "g" "a" "a" "c" "g" "t" "g" "c" "t" "g" "a" "t" "t" "c" "t" "t"
[343] "g" "a" "t" "c" "a" "g" "a" "c" "a" "t" "g" "a" "c" "c" "c" "c" "a"
[361] "t" "c" "a" "g" "c" "t" "t" "c" "t" "t" "c" "c" "c" "c" "a" "g" "c"
[379] "a" "g" "t" "t" "a" "t" "t" "c" "a" "c" "c" "t" "a" "c" "c" "a" "g"
[397] "g" "a" "a" "c" "c" "a" "g" "g" "c" "g" "t" "g" "a" "t" "g" "a" "g" "t"
[415] "g" "c" "a" "g" "a" "t" "c" "g" "g" "c" "t" "g" "g" "a" "a" "g" "g" "c"
[433] "g" "t" "t" "a" "t" "g" "c" "g" "c" "t" "g" "a" "c" "c" "c" "c" "g"
[451] "g" "c" "g" "t" "t" "g" "g" "g" "c" "a" "g" "c" "a" "a" "a" "a" "a"
```

```
[469] "c" "t" "t" "t" "a" "t" "g" "t" "t" "c" "t" "g" "g" "t" "c" "t" "t" "t"
[487] "a" "c" "c" "a" "c" "g" "g" "a" "a" "a" "a" "g" "a" "t" "c" "t" "c"
[505] "c" "a" "g" "c" "a" "g" "a" "c" "g" "a" "c" "c" "c" "c" "a" "a" "c" "t" "g"
[523] "c" "t" "c" "g" "a" "c" "c" "g" "g" "t" "a" "a" "a" "g" "c" "c"
[541] "t" "a" "t" "g" "c" "c" "a" "g" "g" "g" "c" "g" "t" "c" "g" "g" "t"
[559] "a" "a" "c" "t" "c" "g" "a" "t" "c" "c" "c" "g" "g" "a" "t" "a" "t" "c"
[577] "c" "c" "c" "g" "a" "t" "c" "c" "g" "g" "t" "t" "g" "c" "t" "c" "g" "t"
[595] "c" "a" "t" "a" "c" "c" "a" "c" "g" "a" "t" "g" "g" "c" "t" "t" "a"
[613] "c" "t" "g" "a" "a" "c" "t" "g" "a" "a" "a" "g" "t" "g" "a" "a" "a"
[631] "a" "c" "g" "a" "a" "c" "t" "c" "c" "a" "g" "c" "t" "c" "c" "a" "g" "c"
[649] "g" "t" "g" "t" "t" "g" "g" "t" "a" "g" "g" "a" "c" "c" "t" "t" "a"
[667] "t" "t" "t" "g" "g" "t" "t" "c" "t" "c" "c" "g" "c" "t" "c" "c" "a"
[685] "g" "c" "t" "c" "c" "g" "g" "t" "t" "a" "c" "g" "g" "t" "a" "g" "g" "t"
[703] "a" "a" "c" "a" "c" "g" "g" "c" "g" "g" "c" "a" "c" "c" "a" "g" "c" "t"
[721] "g" "t" "g" "g" "c" "t" "g" "c" "a" "c" "c" "g" "c" "t" "c" "c" "g"
[739] "g" "c" "a" "c" "c" "g" "g" "t" "g" "a" "a" "g" "a" "a" "a" "a" "g" "c"
[757] "g" "a" "g" "c" "c" "g" "a" "t" "g" "c" "t" "c" "a" "a" "c" "g" "a" "c"
[775] "a" "c" "g" "g" "a" "a" "a" "g" "t" "t" "a" "t" "t" "t" "a" "a" "t"
[793] "a" "c" "c" "g" "g" "a" "t" "c" "a" "a" "a" "a" "a" "c" "g" "c" "t"
[811] "g" "t" "c" "g" "c" "g" "a" "a" "a" "g" "g" "t" "g" "a" "t" "g" "t" "t"
[829] "g" "a" "t" "a" "a" "g" "g" "c" "g" "t" "t" "a" "a" "a" "a" "c" "t" "g"
[847] "c" "t" "t" "g" "a" "t" "g" "a" "a" "g" "c" "t" "g" "a" "a" "c" "g" "c"
[865] "t" "t" "g" "g" "g" "a" "t" "c" "g" "a" "c" "a" "t" "c" "t" "g" "c" "c"
[883] "c" "g" "t" "t" "c" "c" "a" "c" "c" "t" "t" "t" "a" "t" "c" "a" "g" "c"
[901] "a" "g" "t" "g" "t" "a" "a" "a" "a" "g" "g" "c" "a" "a" "g" "g" "g" "g"
[919] "t" "a" "a"
attr(,"name")
[1] "XYLEECOM.MALM"
attr(,"Annot")
[1] ">XYLEECOM.MALM 921 bp ACCESSION E00218, X04477"
attr(,"class")
[1] "SeqFastadna"
```

As from **seqinR** 1.0-5 the automatic conversion of sequences into vector of single characters can be neutralized, for instance:

```
read.fasta(file = dnafile, as.string = TRUE)
```

```
$XYLEECOM.MALM
[1] "atgaaaatgaataaaagtctcatcgccctgtttatcagcagggtactggcaagcgccctgaaattgcctgcccgttaactacgtaccgcaaaacaccagcgcacg
attr(,"name")
[1] "XYLEECOM.MALM"
attr(,"Annot")
[1] ">XYLEECOM.MALM 921 bp ACCESSION E00218, X04477"
attr(,"class")
[1] "SeqFastadna"
```

Forcing to lower case letters can be disabled this way:

```
read.fasta(file = dnafile, as.string = TRUE, forceDNAtolower = FALSE)
```

```
$XYLEECOM.MALM
[1] "ATGAAAATGAATAAAAGTCTCATCGCCCTGTATTACAGCAGGGTTACTGGCAAGCGCCCTGAAATTGCCTGCCGTAACTACGTACCACCGCAACACCAGCGACG
attr(,"name")
[1] "XYLEECOM.MALM"
attr(,"Annot")
[1] ">XYLEECOM.MALM 921 bp ACCESSION E00218, X04477"
attr(,"class")
[1] "SeqFastadna"
```

## Protein file example

The example file looks like:

```
aafile <- system.file("sequences/seqAA.fasta", package = "seqinr")
cat(readLines(aafile), sep = "\n")
```

```
>A06852          183 residues
MPRLFSYLLGVWLLSQLPREIPGQSTNDFIKACGRELVRWLVEICGSVSWGRTALSLEE
PQLETGPPAETMPSSITKDAEILKMMLEFVPNLPQELKATLSERQPSLRELQQSASKDSN
LNFEFKKIILNRQNEAEDKSLLELKNLGLDKHSRKRLFRMTLSEKCCQVGCIRKDIAR
LC*
```

Read the protein sequence file, looks like:

```
read.fasta(aafile, seqtype = "AA")
```

```
$A06852
[1] "M" "P" "R" "L" "F" "S" "Y" "L" "L" "G" "V" "W" "L" "L" "S" "Q" "I"
[19] "P" "R" "E" "I" "P" "G" "Q" "S" "T" "N" "D" "F" "I" "K" "A" "C" "G" "R"
[37] "E" "L" "V" "R" "L" "W" "V" "E" "I" "C" "G" "S" "V" "S" "W" "G" "R" "T"
[55] "A" "L" "S" "L" "F" "E" "P" "Q" "L" "E" "T" "G" "P" "P" "A" "E" "T" "M"
[73] "P" "S" "S" "I" "T" "K" "D" "A" "E" "I" "L" "K" "M" "M" "L" "E" "F" "V"
[91] "P" "N" "L" "P" "Q" "E" "L" "K" "A" "T" "L" "S" "E" "R" "Q" "P" "S" "L"
[109] "R" "E" "L" "Q" "O" "S" "A" "S" "K" "D" "S" "N" "L" "N" "F" "E" "E" "F"
[127] "K" "K" "I" "I" "L" "N" "R" "Q" "N" "E" "A" "E" "D" "K" "S" "L" "L" "E"
[145] "L" "K" "N" "L" "G" "L" "D" "K" "H" "S" "R" "K" "R" "L" "F" "R" "M"
[163] "T" "L" "S" "E" "K" "C" "C" "Q" "V" "G" "C" "I" "R" "K" "D" "I" "A" "R"
[181] "L" "C" "*"
attr(,"name")
[1] "A06852"
attr(,"Annot")
[1] ">A06852"          183 residues"
attr(,"class")
[1] "SeqFastaAA"
```

The same, but as string and without attributes setting, looks like:

```
read.fasta(aafile, seqtype = "AA", as.string = TRUE, set.attributes = FALSE)
```

```
$A06852
[1] "MPRLFSYLLGVWLLSQLPREIPGQSTNDFIKACGRELVRWLVEICGSVSWGRTALSLEEPQLETGPPAETMPSSITKDAEILKMMLEFVPNLPQELKAT
```

### 2.1.3 The function `write.fasta()`

This function writes sequences to a file in FASTA format. Read 3 coding sequences sequences from a FASTA file:

```
ortho <- read.fasta(file = system.file("sequences/ortho.fasta",
                                         package = "seqinr"))
length(ortho)
```

```
[1] 3
```

```
ortho[[1]][1:12]
```

```
[1] "a" "t" "g" "g" "c" "t" "c" "a" "g" "c" "g" "g"
```

Select only third codon positions:

```
ortho3 <- lapply(ortho, function(x) x[seq(from = 3, to = length(x),
                                         by = 3)])
ortho3[[1]][1:4]
```

```
[1] "g" "t" "g" "g"
```

Write the modified sequences to a file:

```
tmpf <- tempfile()
write.fasta(sequences = ortho3, names = names(ortho3), nbchar = 80,
           file.out = tmpf)
```

Read them again from the same file and check that sequences are preserved:

```
ortho3bis <- read.fasta(tmpf, set.attributes = FALSE)
identical(ortho3bis, ortho3)
```

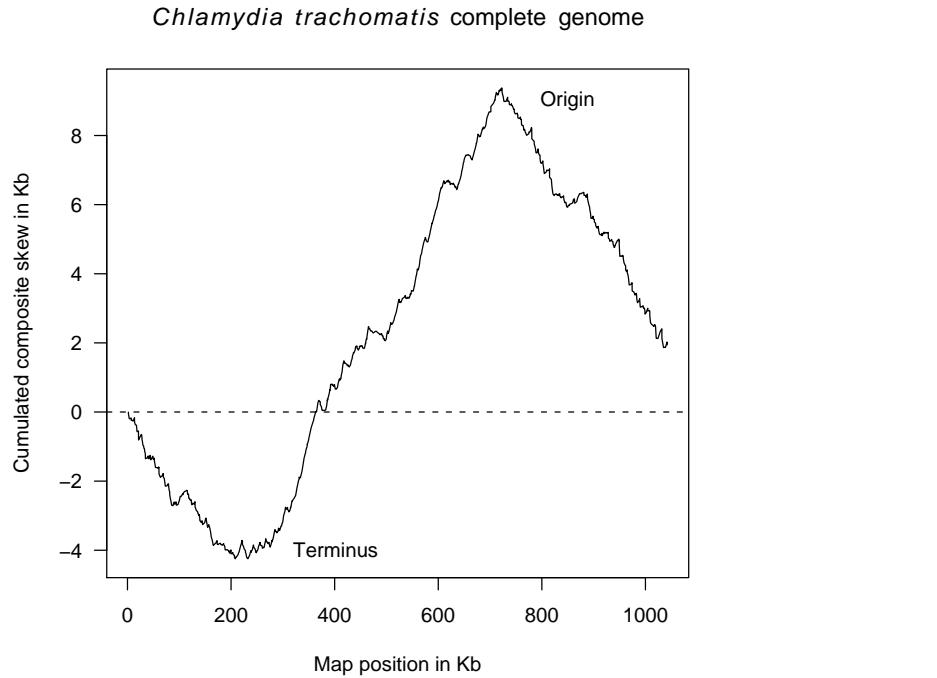
```
[1] TRUE
```

### 2.1.4 Big room examples

#### Oriloc example (*Chlamydia trachomatis* complete genome)

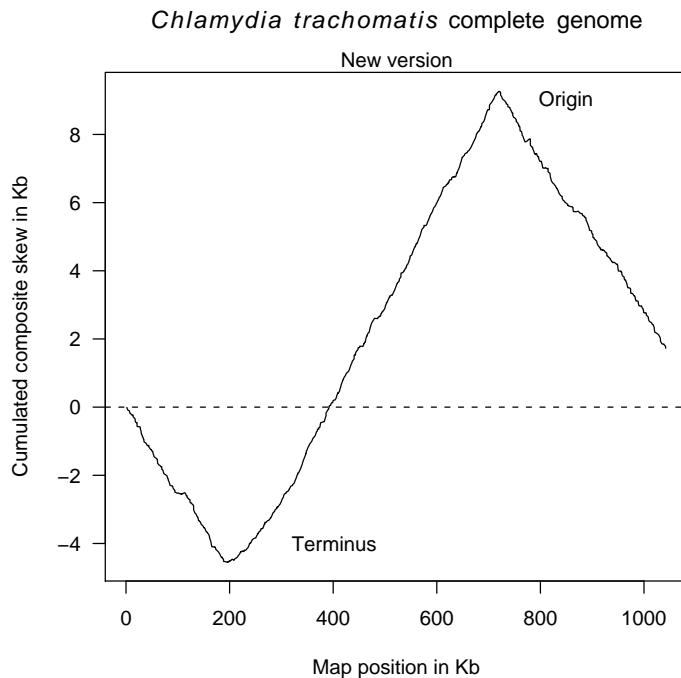
A more consequent example is given in the fasta file `ct.fasta` which contains the complete genome of *Chlamydia trachomatis* that was used in [9]. You should be able to reproduce figure 1b from this paper with the following code:

```
out <- oriloc(seq.fasta = system.file("sequences/ct.fasta",
  package = "seqinr"), g2.coord = system.file("sequences/ct.coord",
  package = "seqinr"), oldoriloc = TRUE)
plot(out$st, out$sk/1000, type = "l", xlab = "Map position in Kb",
  ylab = "Cumulated composite skew in Kb", main = expression(italic(Chlamydia ~
  "trachomatis) ~ "complete ~ "genome), las = 1)
abline(h = 0, lty = 2)
text(400, -4, "Terminus")
text(850, 9, "Origin")
```



Note that the algorithm has been improved since then and that it's more advisable to use the default option `oldoriloc = FALSE` if you are interested in the prediction of origins and terminus of replication from base composition biases (more on this at <http://pbil.univ-lyon1.fr/software/oriloc.html>). See also [42] for a recent review on this topic.

```
out <- oriloc(seq.fasta = system.file("sequences/ct.fasta",
  package = "seqinr"), g2.coord = system.file("sequences/ct.coord",
  package = "seqinr"))
plot(out$st, out$sk/1000, type = "l", xlab = "Map position in Kb",
  ylab = "Cumulated composite skew in Kb", main = expression(italic(Chlamydia ~
  "trachomatis) ~ "complete ~ "genome), las = 1)
mtext("New version")
abline(h = 0, lty = 2)
text(400, -4, "Terminus")
text(850, 9, "Origin")
```



*Arabidopsis thaliana*. Source: wikipedia.

### Example with 21,161 proteins from *Arabidopsis thaliana*

As from `seqinR` 1.0-5 the automatic conversion of sequences into vector of single characters and the automatic attribute settings can be neutralized, for instance :

```
smallAA <- system.file("sequences/smallAA.fasta", package = "seqinr")
read.fasta(smallAA, seqtype = "AA", as.string = TRUE, set.attributes = FALSE)

$smallAA
[1] "SEQINRSEQINRSEQINRSEQINR*
```

This is interesting to save time and space when reading large FASTA files. Let's give a practical example. In their paper [21], Matthew Hannah, Arnd Heyer and Dirk Hincha were working on *Arabidopsis thaliana* genes in order to detect those involved in cold acclimation. They were interested by the detection of proteins called hydrophilins, that had a mean hydrophilicity of over 1 and glycine content of over 0.08 [11], because they are thought to be important for freezing tolerance. The starting point was a FASTA file called `ATH1_pep_cm_20040228` downloaded from the Arabidopsis Information Ressource (TAIR at <http://www.arabidopsis.org/>) which contains the sequences of 21,161 proteins.

```
athfile <- system.file("sequences/ATH1_pep_cm_20040228.fasta",
                       package = "seqinr")
system.time(ath <- read.fasta(athfile, seqtype = "AA", as.string = TRUE,
                           set.attributes = FALSE))

[1] 6.326 0.072 6.480 0.000 0.000
```

It's about one minute here to read 21,161 protein sequences. We save them in XDR binary format to read them faster later at will:

```
save(ath, file = "ath.RData")
system.time(load("ath.RData"))
[1] 0.305 0.013 0.318 0.000 0.000
```

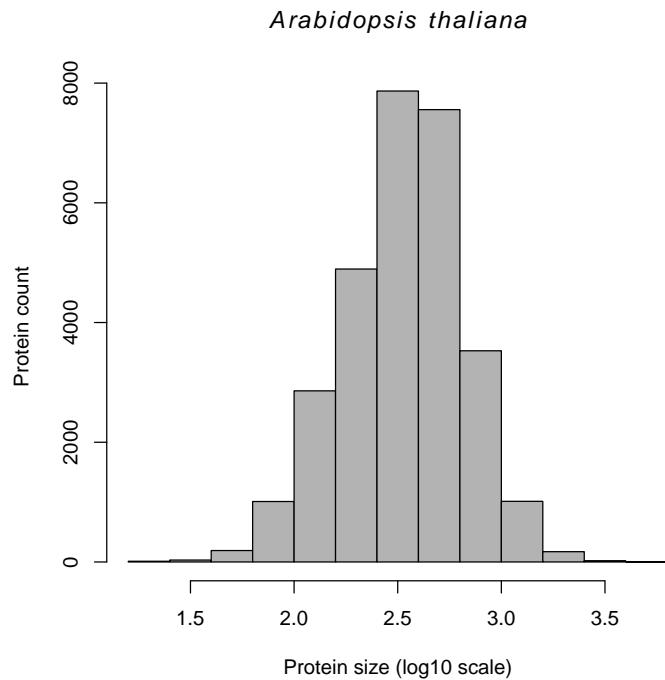
Now it's about one second to load the whole data set thanks to the XDR format. The object size is about 15 Mo in RAM, that is something very close to the flat file size on disk:

```
object.size(ath)/2^20
```

```
[1] 14.65537
file.info(athfile)$size/2^20
[1] 15.89863
```

Using strings for sequence storage is very comfortable when there is an efficient function to compute what you want. For instance, suppose that you are interested by the distribution of protein size in *Arabidopsis thaliana*. There is an efficient vectorized function called `nchar()` that will do the job, we just have to remove one unit because of the stop codon which is translated as a star (\*) in this data set. This is a simple and direct task under R:

```
nres <- nchar(ath) - 1
hist(log10(nres), col = grey(0.7), xlab = "Protein size (log10 scale)",
     ylab = "Protein count", main = expression(italic(Arabidopsis ~
     ^thaliana)))
```



However, sometimes it is more convenient to work with the single character vector representation of sequences. For instance, to count the number of glycine (G), we first play with one sequence, let's take the smallest one in the data set:

```
which.min(nres)

At2g25990.1
9523

ath[[9523]]

[1] "MAGSQREKLKPRTKGSTRC*"

s2c(ath[[9523]])

[1] "M" "A" "G" "S" "Q" "R" "E" "K" "L" "K" "P" "R" "T" "K" "G" "S" "T" "R"
[19] "C" "*"

s2c(ath[[9523]]) == "G"

[1] FALSE FALSE TRUE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
[13] FALSE FALSE TRUE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE

sum(s2c(ath[[9523]]) == "G")

[1] 2
```

We can now easily define a vectorised function to count the number of glycine:

```
ngly <- function(data) {
  res <- sapply(data, function(x) sum(s2c(x) == "G"))
  names(res) <- NULL
  return(res)
}
```

Now we can use `ngly()` in the same way that `nchar()` so that computing glycine frequencies is very simple:

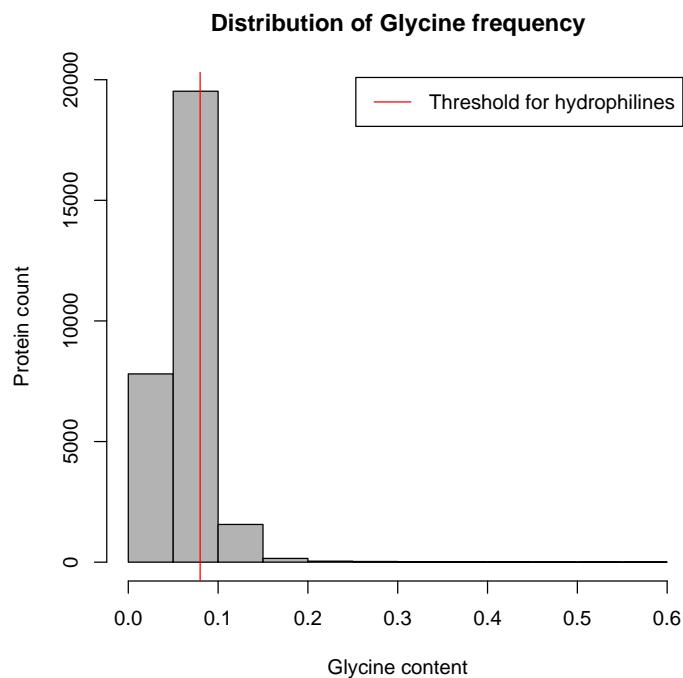
```
ngly(ath[1:10])

[1] 25 5 29 128 8 27 27 26 21 18

fgly <- ngly(ath)/nres
```

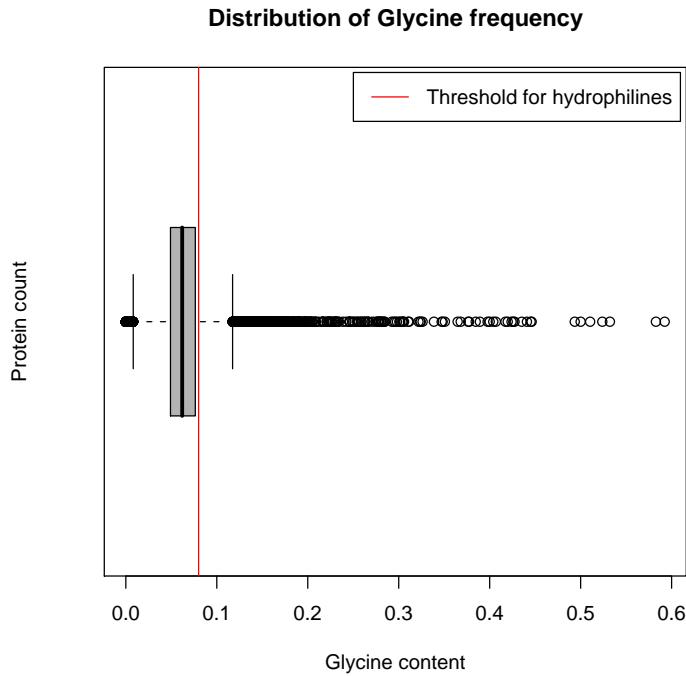
And we can have a look at the distribution:

```
hist(fgly, col = grey(0.7), main = "Distribution of Glycine frequency",
      xlab = "Glycine content", ylab = "Protein count")
abline(v = 0.08, col = "red")
legend("topright", inset = 0.01, lty = 1, col = "red", legend = "Threshold for hydrophilines")
```



Let's use a boxplot instead:

```
boxplot(fgly, horizontal = TRUE, col = grey(0.7), main = "Distribution of Glycine frequency",
        xlab = "Glycine content", ylab = "Protein count")
abline(v = 0.08, col = "red")
legend("topright", inset = 0.01, lty = 1, col = "red", legend = "Threshold for hydrophilines")
```



The threshold value for the glycine content in hydrophilines is therefore very close to the third quartile of the distribution:

```
summary(fgly)
```

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
0.00000	0.04907	0.06195	0.06475	0.07639	0.59240

We want now to compute something relatively more complex, we want the Kyte and Doolittle[33] hydropathy score of our proteins (aka GRAVY score). This is basically a linear form on amino acid frequencies:

$$s = \sum_{i=1}^{20} \alpha_i f_i$$

where  $\alpha_i$  is the coefficient for amino acid number  $i$  and  $f_i$  the relative frequency of amino acid number  $i$ . The coefficients  $\alpha_i$  are given in the KD component of the data set EXP:

```
data(EXP)
EXP$KD
```

```
[1] -3.9 -3.5 -3.9 -3.5 -0.7 -0.7 -0.7 -0.7 -4.5 -0.8 -4.5 -0.8 4.5 4.5
[15] 1.9 4.5 -3.5 -3.2 -3.5 -3.2 -1.6 -1.6 -1.6 -1.6 -4.5 -4.5 -4.5 -4.5 -4.5
[29] 3.8 3.8 3.8 3.8 -3.5 -3.5 -3.5 -3.5 1.8 1.8 1.8 1.8 -0.4 -0.4
[43] -0.4 -0.4 4.2 4.2 4.2 4.2 0.0 -1.3 0.0 -1.3 -0.8 -0.8 -0.8 -0.8
[57] 0.0 2.5 -0.9 2.5 3.8 2.8 3.8 2.8
```

This is for codons in lexical order, that is:

```
words()
```

```
[1] "aaa" "aac" "aag" "aat" "aca" "acc" "acg" "act" "aga" "agc" "agg" "agt"
[13] "ata" "atc" "atg" "att" "caa" "cac" "cag" "cat" "cca" "ccc" "ccg" "cct"
[25] "cga" "cgc" "cgg" "cgt" "cta" "ctc" "ctg" "ctt" "gaa" "gac" "gag" "gat"
[37] "gca" "gcc" "gcg" "gct" "gga" "ggc" "ggg" "ggt" "gta" "gtc" "gtg" "gtt"
[49] "taa" "tac" "tag" "tat" "tca" "tcc" "tcg" "tct" "tga" "tgc" "tgg" "tgt"
[61] "tta" "ttc" "ttg" "ttt"
```

But since we are working with protein sequences here we name the coefficient according to their amino acid :

```
names(EXP$KD) <- sapply(words(), function(x) translate(s2c(x)))
```

We just need one value per amino acid, we sort them in the lexical order, and we reverse the scale so as to have positive values for hydrophilic proteins as in [21] :

```
kdc <- EXP$KD[unique(names(EXP$KD))]
kdc <- -kdc[order(names(kdc))]
kdc
```

*	A	C	D	E	F	G	H	I	K	L	M	N	P	Q
0.0	-1.8	-2.5	3.5	3.5	-2.8	0.4	3.2	-4.5	3.9	-3.8	-1.9	3.5	1.6	3.5
R	S	T	V	W	Y									
4.5	0.8	0.7	-4.2	0.9	1.3									

Now that we have the vector of coefficient  $\alpha_i$ , we need the amino acid relative frequencies  $f_i$ , let's play with one protein first:

```
ath[[9523]]
```

```
[1] "MAGSQREKLKPRTKGSTRC*
```

```
s2c(ath[[9523]])
```

```
[1] "M" "A" "G" "S" "Q" "R" "E" "K" "L" "K" "P" "R" "T" "K" "G" "S" "T" "R"
[19] "C" "*"
```

```
table(s2c(ath[[9523]]))
```

```
* A C E G K L M P Q R S T
1 1 1 1 2 3 1 1 1 3 2 2
```

```
table(factor(s2c(ath[[9523]]), levels = names(kdc)))
```

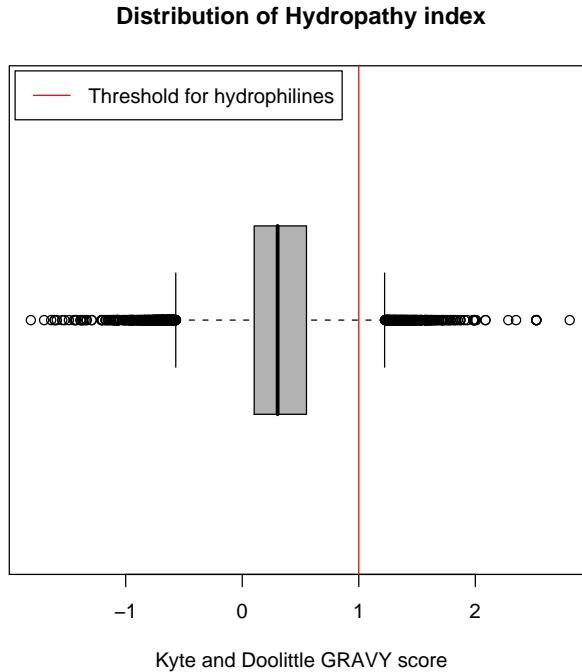
```
* A C D E F G H I K L M N P Q R S T V W Y
1 1 1 0 1 0 2 0 0 3 1 1 0 1 1 3 2 2 0 0 0
```

Now that we know how to count amino acids it's relatively easy thanks to R's matrix operator  $\%*\%$  to define a vectorised function to compute a linear form on amino acid frequencies:

```
linform <- function(data, coef) {
  f <- function(x) {
    aaseq <- s2c(x)
    freq <- table(factor(aaseq, levels = names(coef)))/length(aaseq)
    return(coef %*% freq)
  }
  res <- sapply(data, f)
  names(res) <- NULL
  return(res)
}
kdath <- linform(ath, kdc)
```

Let's have a look at the distribution:

```
boxplot(kdath, horizontal = TRUE, col = grey(0.7), main = "Distribution of Hydropathy index",
       xlab = "Kyte and Doolittle GRAVY score")
abline(v = 1, col = "red")
legend("topleft", inset = 0.01, lty = 1, col = "red", legend = "Threshold for hydrophilines")
```



The threshold is therefore much more stringent here than the previous one on glycine content. Let's define a vector of logicals to select the hydrophilines:

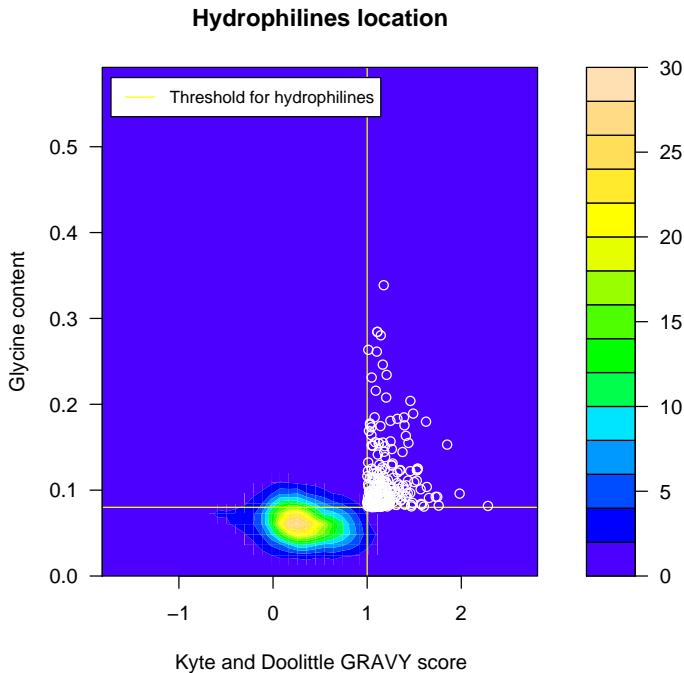
```
hydrophilines <- fgly > 0.08 & kdath > 1
names(ath)[hydrophilines]
```

```
[1] "At1g02840.1" "At1g02840.2" "At1g02840.3" "At1g03320.1" "At1g03820.1"
[6] "At1g04450.1" "At1g12810.1" "At1g13020.1" "At1g13290.1" "At1g13890.1"
[11] "At1g15270.1" "At1g15280.1" "At1g15280.2" "At1g15400.1" "At1g15400.2"
[16] "At1g20220.1" "At1g21520.1" "At1g23530.1" "At1g23860.1" "At1g23860.2"
[21] "At1g26210.1" "At1g27650.1" "At1g31750.1" "At1g42430.1" "At1g44478.1"
[26] "At1g47970.1" "At1g48920.1" "At1g49680.1" "At1g50300.1" "At1g51510.1"
[31] "At1g52300.1" "At1g52690.1" "At1g52690.2" "At1g52710.1" "At1g53160.1"
[36] "At1g53160.2" "At1g53260.1" "At1g53645.1" "At1g54410.1" "At1g54926.1"
[41] "At1g55310.1" "At1g56085.1" "At1g60650.1" "At1g60650.2" "At1g62045.1"
[46] "At1g63310.1" "At1g64360.1" "At1g64370.1" "At1g67325.1" "At1g67870.1"
[51] "At1g72050.1" "At1g72080.1" "At1g73350.1" "At1g75560.1" "At1g76010.1"
[56] "At1g77030.1" "At1g77765.1" "At1g80000.1" "At1g80000.2" "At2g02160.1"
[61] "At2g03440.1" "At2g04380.1" "At2g04870.1" "At2g07675.1" "At2g11910.1"
[66] "At2g11910.2" "At2g13125.1" "At2g17950.1" "At2g19750.1" "At2g20430.1"
[71] "At2g20760.1" "At2g21490.1" "At2g22080.1" "At2g22100.1" "At2g22820.1"
[76] "At2g23110.1" "At2g24590.1" "At2g25990.1" "At2g27100.1" "At2g32710.1"
[81] "At2g32710.2" "At2g33460.1" "At2g33690.1" "At2g34310.1" "At2g34310.2"
[86] "At2g34310.3" "At2g35230.2" "At2g37340.1" "At2g37340.2" "At2g37340.3"
[91] "At2g39855.2" "At2g40040.1" "At2g40080.1" "At2g40170.1" "At2g41730.1"
[96] "At2g42190.1" "At2g43370.1" "At2g43970.2" "At2g44710.1" "At2g44820.1"
[101] "At2g44820.2" "At2g46270.2" "At3g02480.1" "At3g05220.1" "At3g05220.2"
[106] "At3g07195.1" "At3g10020.1" "At3g13570.1" "At3g13780.1" "At3g15270.1"
[111] "At3g16040.1" "At3g16080.1" "At3g17160.1" "At3g17190.1" "At3g20910.1"
```

```
[116] "At3g21270.1" "At3g21290.1" "At3g22660.1" "At3g23140.1" "At3g23380.1"
[121] "At3g23390.1" "At3g24100.1" "At3g24740.1" "At3g25590.1" "At3g26400.1"
[126] "At3g26420.1" "At3g29075.1" "At3g29470.1" "At3g29600.1" "At3g47120.1"
[131] "At3g48180.1" "At3g49430.1" "At3g50320.1" "At3g50610.1" "At3g50670.1"
[136] "At3g50690.1" "At3g50970.1" "At3g50980.1" "At3g51810.1" "At3g51940.1"
[141] "At3g53500.1" "At3g53500.2" "At3g55460.1" "At3g56720.1" "At3g57930.1"
[146] "At3g58070.1" "At3g60230.1" "At3g62790.1" "At3g63100.1" "At3g63400.1"
[151] "At3g63400.2" "At4g02425.1" "At4g02430.2" "At4g07940.1" "At4g09840.1"
[156] "At4g09850.1" "At4g09980.1" "At4g12610.1" "At4g12670.1" "At4g13560.1"
[161] "At4g13615.1" "At4g14020.1" "At4g14320.1" "At4g16830.1" "At4g17520.1"
[166] "At4g22740.1" "At4g22740.2" "At4g24500.1" "At4g24500.2" "At4g24680.1"
[171] "At4g25340.1" "At4g25500.1" "At4g25500.2" "At4g26640.1" "At4g26640.2"
[176] "At4g28870.1" "At4g28990.1" "At4g29390.1" "At4g31580.1" "At4g36600.1"
[181] "At4g36730.1" "At4g36730.2" "At4g38710.1" "At5g02020.1" "At5g03990.1"
[186] "At5g04290.1" "At5g11260.1" "At5g11970.1" "At5g12840.1" "At5g12840.2"
[191] "At5g12840.3" "At5g12840.4" "At5g16470.1" "At5g17650.1" "At5g19960.1"
[196] "At5g22040.1" "At5g22650.1" "At5g22650.2" "At5g26890.1" "At5g27690.1"
[201] "At5g28610.1" "At5g28630.1" "At5g44200.1" "At5g47210.1" "At5g50830.1"
[206] "At5g51840.1" "At5g52200.1" "At5g52310.1" "At5g52530.1" "At5g52530.2"
[211] "At5g53820.1" "At5g55430.1" "At5g55640.1" "At5g55670.1" "At5g56670.1"
[216] "At5g58040.1" "At5g58470.1" "At5g58470.2" "At5g59080.1" "At5g59950.1"
[221] "At5g59950.3" "At5g60800.1" "At5g60880.1" "At5g61890.1" "At5g64130.1"
[226] "At5g66400.1" "At5g66780.1" "At5g67190.1" "At5g67490.1" "AtMg00880"
```

Check with a simple graph that there is no mistake here:

```
library(MASS)
dst <- kde2d(kdath, fgly, n = 50)
filled.contour(x = dst, color.palette = topo.colors, plot.axes = {
  axis(1)
  axis(2)
  title(xlab = "Kyte and Doolittle GRAVY score", ylab = "Glycine content",
    main = "Hydrophilines location")
  abline(v = 1, col = "yellow")
  abline(h = 0.08, col = "yellow")
  points(kdath[hydrophilines], fgly[hydrophilines], col = "white")
  legend("topleft", inset = 0.02, lty = 1, col = "yellow",
    bg = "white", legend = "Threshold for hydrophilines",
    cex = 0.8)
})
```



Everything seems to be OK, we can save the results in a data frame:

```
athres <- data.frame(list(name = names(ath), KD = kdath, Gly = fgly))
head(athres)
```

	name	KD	Gly	
1	At1g01010.1	At1g01010.1	0.7297674	0.05827506
2	At1g01020.1	At1g01020.1	-0.1674419	0.03906250
3	At1g01030.1	At1g01030.1	0.8136490	0.08100559
4	At1g01040.1	At1g01040.1	0.4159686	0.06705081
5	At1g01050.1	At1g01050.1	0.4460094	0.03773585
6	At1g01060.1	At1g01060.1	0.7444272	0.04186047

We want to check now that the results are consistent with those reported previously. The following table is extracted from the file `pge0.0010026.st003.xls` provided as the supplementary material table S3 in [21] and available at <http://www.ncbi.nlm.nih.gov/picrender.fcgi?artid=1189076&blobname=pge0.0010026.st003.xls>. Only the protein names, the hydrophilicity and the glycine content were extracted:

```
hannah <- read.table(system.file("sequences/hannah.txt", package = "seqinr"),
                      sep = "\t", header = TRUE)
head(hannah)
```

	AGI	Hydrophilicity	Glycine
1	At2g19570	-0.10	0.07
2	At2g45290	-0.25	0.09
3	At4g29570	-0.05	0.07
4	At4g29580	-0.10	0.06
5	At4g29600	-0.14	0.06
6	At5g28050	-0.11	0.08

The protein names are not exactly the same because they have no extension. As explained in [21], when multiple gene models were predicted only the first was one used. Then:

```

hannah$AGI <- paste(hannah$AGI, "1", sep = ".")
head(hannah)

```

	AGI	Hydrophilicity	Glycine
1	At2g19570.1	-0.10	0.07
2	At2g45290.1	-0.25	0.09
3	At4g29570.1	-0.05	0.07
4	At4g29580.1	-0.10	0.06
5	At4g29600.1	-0.14	0.06
6	At5g28050.1	-0.11	0.08

We join now the two data frames thanks to their common key:

```

join <- merge(hannah, athres, by.x = "AGI", by.y = "name")
head(join)

```

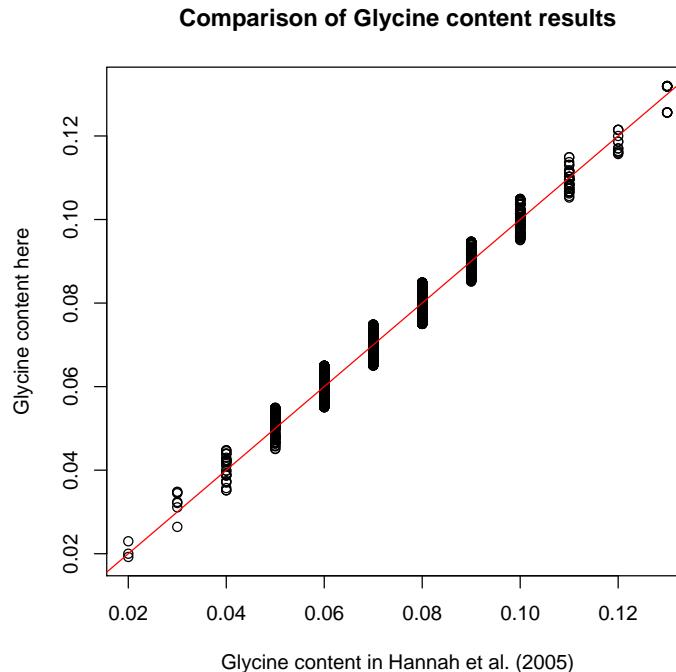
	AGI	Hydrophilicity	Glycine	KD	Gly
1	At1g01120.1	-0.10	0.06	0.10699433	0.05871212
2	At1g01390.1	0.02	0.06	0.00914761	0.06458333
3	At1g01390.1	0.02	0.06	0.00914761	0.06458333
4	At1g01420.1	-0.05	0.07	0.06203320	0.07276507
5	At1g01420.1	-0.05	0.07	0.06203320	0.07276507
6	At1g01480.1	-0.20	0.07	0.20080483	0.06653226

Let's compare the glycine content :

```

plot(join$Glycine, join$Gly, xlab = "Glycine content in Hannah et al. (2005)",
     ylab = "Glycine content here", main = "Comparison of Glycine content results")
abline(c(0, 1), col = "red")

```

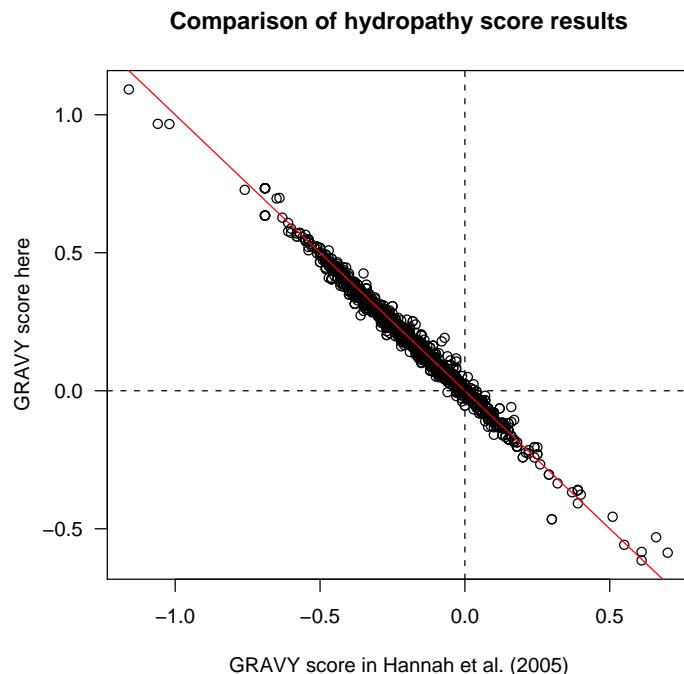


The results are consistent, we have just lost some resolution because there are only two figures after the decimal point in the Excel<sup>1</sup> file. Let's have a look at the GRAVY score now:

---

<sup>1</sup> this software is a real **pain** for the reproducibility of results. This is well documented, see [http://www.burns-stat.com/pages/Tutor/spreadsheet\\_addiction.html](http://www.burns-stat.com/pages/Tutor/spreadsheet_addiction.html) and references therein.

```
plot(join$Hydrophilicity, join$KD, xlab = "GRAVY score in Hannah et al. (2005)",
      ylab = "GRAVY score here", main = "Comparison of hydropathy score results",
      las = 1)
abline(c(0, -1), col = "red")
abline(v = 0, lty = 2)
abline(h = 0, lty = 2)
```



The results are consistent, it's hard to say whether the small differences are due to Excel rounding errors or because the method used to compute the GRAVY score was not exactly the same (in [21] they used the mean over a sliding window).

## 2.2 Importing aligned sequence data

### 2.2.1 Aligned sequences files examples

#### mase

Mase format is a flatfile format used by the SeaView multiple alignment editor [10], developed by Manolo Gouy and available at <http://pbil.univ-lyon1.fr/software/seaview.html>. The mase format is used to store nucleotide or protein multiple alignments. The beginning of the file must contain a header containing at least one line (but the content of this header may be empty). The header lines must begin by `;;`. The body of the file has the following structure: First, each entry must begin by one (or more) commentary line. Commentary lines begin by the character `;`. Again, this commentary line may be empty. After the commentaries, the name of the sequence is written on a separate line. At last, the sequence itself is written on the following lines.

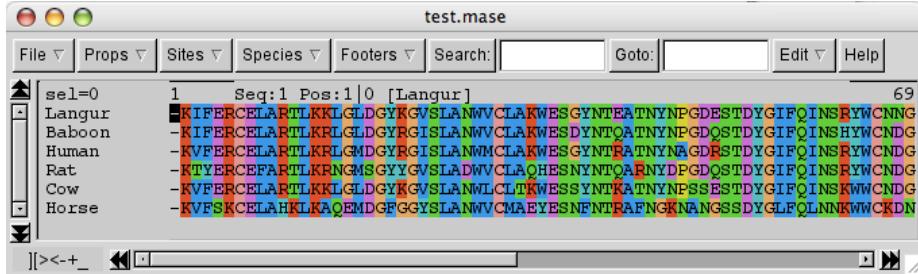


Figure 2.1: The file `test.mase` under SeaView. This is a graphical multiple sequence alignment editor developed by Manolo Gouy [10]. SeaView is able to read and write various alignment formats (NEXUS, MSF, CLUSTAL, FASTA, PHYLIP, MASE). It allows to manually edit the alignment, and also to run DOT-PLOT or CLUSTALW programs to locally improve the alignment.

```
masef <- system.file("sequences/test.mase", package = "seqinr")
cat(readLines(masef), sep = "\n")
```

```
;;Aligned by clustal on Tue Jun 30 17:36:11 1998
;empty description
Langur
-KIFERCELARTLKKLGLDGYKGVLANWVCLAKWESGYNTEATNYNPGDESTDYGIFQINSRYWCNNKGPGAVDACHISCALLQNNIADAVACAKRVVSDQGIRAWVAWRNHCQN
;
Baboon
-KIFERCELARTLKRLGLDGYRGISLANWVCLAKWESDYNTQATNYNPGDQSTDYGIFQINSHYWCNDGKPGAVNACHISCNALLQDNITDAVACAKRVVSDQGIRAWVAWRNHCQN
;
Human
-KVFERCELARTLKRLGMDGYRGISLANWMCLAKWESGYNTRATNYNAGDRSTDYGIFQINSRYWCNDGKPGAVNACHLSCSALLQDNIADAVACAKRVVRDQGIRAWVAWRNRCQN
;
Rat
-KTYERCEFARTLKRNGMSYYGVSLADWVCLAQHESNYNTQARNYDPGDQSTDYGIFQINSRYWCNDGKPRAKNACGIPCSALLQDDITQAIQCAKRVVRDQGIRAWVAWRHCKN
;
Cow
-KVFERCELARTLKKLGLDGYKGVLANWLCLTKWESSYNTKATNYNPSSESTDYGIFQINSKWWCNDGKPNAVDGCHVSCSELMENDIAKAVACAKKIVSEQGITAWVAKSHCRD
;
Horse
-KVFSKCELAHKLKAQEMDGFGGYSLANWVCMAYESENFTAFNGKNANGSSDYGLFQLNNKWWCKDNKRSSSNACNIMCSKLLDENIDDDISCAKRVVRDKGMSAWKAWVKHCKD
```

A screenshot copy of the same file as seen under SeaView is given in figure 2.1.

### clustal

The CLUSTAL format (\*.aln) is the format of the ClustalW multialignment tool output [22, 62]. It can be described as follows. The word CLUSTAL is on the first line of the file. The alignment is displayed in blocks of a fixed length, each line in the block corresponding to one sequence. Each line of each block starts with the sequence name (maximum of 10 characters), followed by at least one space character. The sequence is then displayed in upper or lower cases, '-' denotes gaps. The residue number may be displayed at the end of the first line of each block.

```
clustalf <- system.file("sequences/test.aln", package = "seqinr")
cat(readLines(clustalf), sep = "\n")
```

## CLUSTAL W (1.82) multiple sequence alignment

FOSB_MOUSE	MFQAFPGDYDSGRCSSPSAESQYLSSVDSFGSPPTAAASQEAGLGEPMGSFVPTVTA	60
FOSB_HUMAN	MFQAFPGDYDSGRCSSPSAESQYLSSVDSFGSPPTAAASQEAGLGEPMGSFVPTVTA	60
*****		
FOSB_MOUSE	ITTSQDLQWLVQPTLISSMAQSQQPLASQPPAVDPYDMPGTSYSTPGLSAYSTGGASGS	120
FOSB_HUMAN	ITTSQDLQWLVQPTLISSMAQSQQPLASQPPVVDPYDMPGTSYSTPGMSGYSSGGASGS	120
*****		
FOSB_MOUSE	GGPSTTTSGPVSARPARARPRPREELTPEEEKRRVRERNKLAAKCRNRRRELT	180
FOSB_HUMAN	GGPSTGTSGPVSARPARARPRPREELTPEEEKRRVRERNKLAAKCRNRRRELT	180
*****		
FOSB_MOUSE	DRLQAETDQLEEEKAELSEIAELQKEKERLEFVLVAHKPGCKIPYEEGPGPGLAEVRD	240
FOSB_HUMAN	DRLQAETDQLEEEKAELSEIAELQKEKERLEFVLVAHKPGCKIPYEEGPGPGLAEVRD	240
*****		
FOSB_MOUSE	LPGSTSAKEDGFGWLLPPPPPLPFQSSRDAPPNLTASLFTHSEVQLGDPFPVVSPTY	300
FOSB_HUMAN	LPGSAPAKEDGFSWLLPPPPPLPFQTSDAPPNLTASLFTHSEVQLGDPFPVNPSPY	300
*****		
FOSB_MOUSE	TSSFVLTCPVEVSAFAGAQRTSGSEQPSDPLNSPSLLL 338	
FOSB_HUMAN	TSSFVLTCPVEVSAFAGAQRTSGSDQPSDPLNSPSLLL 338	
*****		

## phylip

PHYLIP is a tree construction program [8]. The format is as follows: the number of sequences and their length (in characters) is on the first line of the file. The alignment is displayed in an interleaved or sequential format. The sequence names are limited to 10 characters and may contain blanks.

```
phylipf <- system.file("sequences/test.phylip", package = "seqinr")
cat(readLines(phylipf), sep = "\n")
```

```
      42
Turkey   AAGCTNGGGC ATTCAGGGT
Salmo    gairAAGCCTTGGC AGTCAGGGT
H. SapiensACCGGTTGGC CGTTCAGGGT
Chimp    AAACCCCTTGC CGTTACGCTT
Gorilla   AAACCCCTTGC CGGTACGCTT

GAGCCCGGGC AATACAGGGT AT
GAGCCGTGGC CGGGCACGGT AT
ACAGGTTGGC CGTTCAGGGT AA
AAACCGAGGC CGGGACACTC AT
AAACCATTGC CGGTACGCTT AA
```

## msf

MSF is the multiple sequence alignment format of the GCG sequence analysis package (<http://www.accelrys.com/products/gcg/index.html>). It begins with the line (all uppercase) !!NA\_MULTIPLE\_ALIGNMENT 1.0 for nucleic acid sequences or !!AA\_MULTIPLE\_ALIGNMENT 1.0 for amino acid sequences. Do not edit or delete the file type if its present (optional). A description line which contains informative text describing what is in the file. You can add this information to the top of the MSF file using a text editor (optional). A dividing line which contains the number of bases or residues in the sequence, when the file was created, and importantly, two dots (..) which act as a divider between the descriptive information and the following sequence information (required). msf files contain some other information: the Name/Weight, a Separating Line which must include two slashes (//) to divide the name/weight

information from the sequence alignment (required) and the multiple sequence alignment.

```
msff <- system.file("sequences/test.msf", package = "seqinr")
cat(readLines(msff), sep = "\n")

PileUp of: @Pi3k.Fil
Symbol comparison table: GenRunData:Pileuppep.Cmp CompCheck: 1254
  GapWeight: 3.000
  GapLengthWeight: 0.100

Pi3k.Msf MSF: 377 Type: P July 12, 1996 10:40 Check: 167 ..
Name: Tor1_Yeast Len: 377 Check: 7773 Weight: 1.00
Name: Tor2_Yeast Len: 377 Check: 8562 Weight: 1.00
Name: Frap_Human Len: 377 Check: 9129 Weight: 1.00
Name: Esr1_Yeast Len: 377 Check: 8114 Weight: 1.00
Name: Tel1_Yeast Len: 377 Check: 1564 Weight: 1.00
Name: Pi4k_Human Len: 377 Check: 8252 Weight: 1.00
Name: Stt4_Yeast Len: 377 Check: 9117 Weight: 1.00
Name: Pik1_Yeast Len: 377 Check: 3455 Weight: 1.00
Name: P3k1_Soybn Len: 377 Check: 4973 Weight: 1.00
Name: P3k2_Soybn Len: 377 Check: 4632 Weight: 1.00
Name: Pi3k_Arath Len: 377 Check: 3585 Weight: 1.00
Name: Vp34_Yeast Len: 377 Check: 5928 Weight: 1.00
Name: P11a_Human Len: 377 Check: 6597 Weight: 1.00
Name: P11b_Human Len: 377 Check: 8486 Weight: 1.00

//
```

Tor1_Yeast	.....GHE	DIRQDSLVMQ	LFGLVNTLLK	NDSECFKRHL	DIQQYPAIPL	50
Tor2_Yeast	.....GHE	DIRQDSLVMQ	LFGLVNTLLQ	NDAECFRRHL	DIQQYPAIPL	
Frap_Human	.....GHE	DLRQDERVMQ	LFGLVNTLLA	NDPTSLRKNL	SIQRYAVIPL	
Esr1_Yeast	.....KKE	DVRQDNQYMQ	FATTMDFLLS	KDIASRKRSL	GINIYSVLSL	
Tel1_Yeast	.KALMKGSND	DLRQDAIMEQ	VFQQVNKLQ	NDKVLRLNLDL	GIRTYKVPL	
Pi4k_Human	..AAIFKVGD	DCRQDMLALQ	IIDLFKNIFQ	LV....GLDL	VFVPYRVVAT	
Stt4_Yeast	..AAIFKVGD	DCRQDVLALQ	LISLFRTIWS	S1....GLDV	YVFPYRVAT	
Pik1_Yeast	..VIAKTGD	DLRQEAFAYQ	MIQAMANIWV	KE....KVDV	WVKRMKILIT	
P3k1_Soybn	TCKIIFKKGD	DLRQDQLVVQ	MVSLSMDRLLK	LE....NLDL	HLTPYKVLAT	
P3k2_Soybn	..IFKKGD	DLRQDQLVVQ	MVSLSMDRLLK	LE....NLDL	HLTPYKVLAT	
Pi3k_Arath	..KLIFKKGD	DLRQDQLVVQ	MWLSMDRLLK	LE....NLDL	CLTPYKVLAT	
Vp34_Yeast	..YHLMFKVGD	DLRQDQLVVQ	IISLMNELLK	NE....NVDL	KLTPYKILAT	
P11a_Human	..IIFKNGD	DLRQDMMLTLQ	IIRIMENIWQ	NQ....GLDL	RMLPYGCLSI	
P11b_Human	..VIFKNGD	DLRQDMMLTLQ	MLRLMDLLWK	EA....GLDL	RMLPYGCLAT	
51						
Tor1_Yeast	SPKGGLLGWV	PNSDTFHVLI	REHRDAKKIP	LNIEHWVMLQ	MAPDYENLTL	100
Tor2_Yeast	SPKGGLLGWV	PNSDTFHVLI	REHREAKKIP	LNIEHWVMLQ	MAPDYDNLTL	
Frap_Human	STNSGLIGWV	PHCDTLHALI	RDYREKKKIL	LNIEHRIMLR	MAPDYDHLLT	
Esr1_Yeast	REDCGILEMV	PNVVTLRSIL	STKYESLKK	Y....SLKS	LHDRWQHTAV	
Tel1_Yeast	GPKAGIIEFV	ANSTSLHQIL	SKLHTNDKIT	FDQARKGMKA	VQTTSN ...	
Pi4k_Human	APCCGVIECI	PDCTS.....	.....	RDQLGRQTDF	GMYDYFTRQY	
Stt4_Yeast	APCGVIDVL	PNSVS.....	.....	RDMLGREAQN	GLYEYFTSKF	
Pik1_Yeast	SANTGLVTF	TNSAMSVHSIK	KALTKKMIED	AELDDKGGA	SLNDHFLRAF	
P3k1_Soybn	GGDEGMLEFI	P.SRSLAQI.	.....	.LSENRSII	SYLQ.....	
P3k2_Soybn	GGDEGMLEFI	P.SRSLAQI.	.....	.LSENRSII	SYLQ.....	
Pi3k_Arath	GHDEGMLEFI	P.NDTLASI.	.....	.LSEHRSIT	SYLQ.....	
Vp34_Yeast	GPOEGAIEFI	P.NDTLASI.	.....	.LSKYHGIL	CYLK .....	
P11a_Human	GDCVGLIEVV	RNSHTIMQI.	.....	.Q.CKGGLK	GALQFNSHTL	
P11b_Human	GDRSGLIEVV	STSETIADI.	.....	.QLNSSNVA	AAAIFNKDAL	

## FASTA

Sequence in fasta format begins with a single-line description (distinguished by a greater-than (>) symbol), followed by sequence data on the next line.

```
fastaf <- system.file("sequences/Anouk.fasta", package = "seqinr")
cat(readLines(fastaf), sep = "\n")
```

```
>LmjF01.0030
ATGATGTGCGGCCGAGCCGGCTCGCAGCGTACATCAGCAGCTGCTGCGCGGTAC
CAGCTGGAGCGCTTCAGTGTGCTTGATCGAGCATGACCATCAAGGACCTCTCGCC
CTGCAGCCAGAGGACTTAACCGTACGGCGTCTAGAGGGATGGACATTGCGGCTG
CGTAGACGCCATCGACTACAGCTAACCGCTCCCGCTCCGGCTCGGAGTGAC
GTGCTGACAACGACGGCACGGGACAGCTACAGCCGGAGGGAAAGGGGG
TGCCTGACGGAGCCGGCGGAGCTACACAGCAGCGGAACACAGTCCTTGGCGTCTG
ACCGACACGGCCGGAGGGATGAAAGCGCAAGAGCCATCTCGTCCGAAAGCGT
CCGCTCAGGGCGGGAGCAGCGAACGGCTTACGGACATCATGGACGCCAACAGC
GGGAGATTGTCTGAAGGAGCCAAGGGTAAGGTGACCTCCCAAGTACACCCAGTG
CACCGCTTCTCGAGGAGTTTCGACGGCTGACGACAACGTCAGCGTGTACAAC
CGCGCTGCCGGCGCTGATCGAACCGTCTCGACGGCGTGGCGACATGCTCGCC
TATGGACAGACAGGGAGCGCAAGACACACAGATGCTGGCAAGGGCCCGAGCGGGC
CTCTACGACTCGCCGAAAGACATGTTGACGCCCTACCGAGCGAACACCGCATCGT
GTTTCTTCTACAGATCTACAGGGGAAGCTCTTGACTTGTCAACGGCGGCCACCC
CTGCGAGCCCTCGAGGAGCACAAGGGCCGGTGAACATCGCGGCTCACCGAACACTGC
TCTACCGAGGTGGAGGACCTCATGACGATCATGACCAACGGGAGCGGTGTTGCACTGC
GGCTCACCGGGCGCATGACACAAGCTCCCGCTCCACGCCATTCTCGAGATCAAGCTC
AAGCGAACAGGACGCTGAAGGAGCGCAAGTTCAGCTCACGACCTCGTGGAGAC
GAGCGCGGGCTGACACGGTGAAGCTGCGCCGACAGCACGCCCTCGAAGGGCGGAGATC
AACAGAGCGTACTCGCGCTGAAGGAGTGCATTGTTAGATCAGAACAGGAAGC
GTCCTGTTCCGGCTCGAGCTGAAGCTGACTGAGGTGCTCCGACTCTTATGGCAACTGC
CGCACGGTATGCTGGCCGCTCTCCCTCAACAAATGCCAGCACCGTGAAC
ACGCTGGCTACGGCGATCGTCAAGGGAGCTGAAGCGAACGCCACGGAGCGGGCACT
GTGTCATGCCGACGACAGGAAAGGGCTTCTTGACACGACCGAGACGAGGCCACCG
TCGGGAGGAGCAGCAAACCTGGCTTCTACGGGCCGGCTTTCTCGGCTCTCGGACG
GCTGCGCAGCCTAGGACTACAGCTGAGGAGTGCATTGTTAGATCAGAACAGGAC
TCGTCGAGGCCAGTGAACCTCGTCAACCGGAGCGCTCCGGCATCGGACTCGG
GACATGGTGTGCACTAACGGGCCCGCACTGACAGAGAACGGGAGCACACGGTGAAC
GGCGGGAGGAGCTGGCGCCAAAGCTTCAAGCGCTTGAGAGCGGGGGAGCTTGTGCG
GCCAGCGCAGTCCGCTATTGACCAATAACCGCTACCTCGAGACGGACATGAAGTGT
ATCAAGGAGGAGTACCAAGGTGAAGTACGACGAGCAGATGAACGCCAACACGGCAGC
TTGTGGAGGAGCTGCTGTTGAGCGAGAACGGGCCGATGGAGTCTTCTA
ACCGAGCTGGAGGAGCTGACAAGATCGCGCAGCAGGTGCCGACATCACGCCCTTCAG
CAGCACCTGCCGCAACG

>LinJ01.0030
ATGATGTGCGGCCGAGCCGGCTCGCAGCGTACATCAGCAGCTGCTGCGCGGTAC
CAGCTGGAGCGCTTCAGGTTCTTGATCGAGCATGACCATCAAGGACCTCTCGCC
CTGCAGCCAGAGGACTTAACCGTACGGCGTCTAGAGGGATGGACATTGCGGCTG
CGGACGCCATCGACTACAGCCAAACCGCTCCCGCTCGGCTCGGACTGAC
GTGCTGACAACGACGGGACGGGACGGGACAGTACGCCGGAGGGGGAGGGGG
TGCCTGACGGAGCCGGCAACGGGACTACAGCCGGAAACCCACGGCTCTTGGGGTCTG
ACCGACACGGGGAGGAGCTGAAGGGCAAGGCCATCATGTCCTCACGCCAAGCGT
CCGCTCAGGCCGGGGAGCAGCGAACGGCTTACGGACATCATGGACGCCAACAAC
GGGAGATTGTCTGAAGGAGCCAAGGGTAAGGTGACCTCCCGAAGTACACCCAGTG
CACCGCTTCTCTGACCAAGGTTTCTGACGGGGTGTGACAAACCTCGAGGTGACAAC
CGCGCTGCCGGCGCTGATCGACACCCTTCTGACGGGGTGTGCGACATGCTCGCC
TATGGGAGACAGGGAGCGGCAAGACACACAGATGCTGGCAAGGGCCCGAGCGGGC
CTGTCAGCCTCGGCGGAAAGACATGTTGACCCCTCAGAGCGACACGCCATCGTT
GTTTCTTCTACAGGATCTACAGGGGAAGCTTGTACTTGTCAACGGCGGCCACCA
CTGCGAGGAGTGCATGACGATCATCGACCGAGCGGGAGCGGGCTCCGAGCTGC
TCTACAGCGTGGAGGACCTCATGACGATCATCGACCGAGCGGGAGCGGGCTCCGAGCTGC
GGCTCACCGGGCCAACGACACAGAGCTCCGCTCCACGCCATTCTCGAGATCAAGCTC
AAGCGAACAGGAGCTGAAGGAGCGCAAGGAGCTAACATTCATCGACCTCGTGGAGC
GAGCGGGCCGGCGCACAGCGTGGGATTCGCGCGACAGACGCCCTCGAAGGGGGAGGATT
AACAGAGCTACTCGCTCTGAAGGAGTGCATTGTTAGATCAGAACAGGAAGC
GTCCTGCTCGCGCTGAAGCTGACTGAAGGTGCTCCGACTCTTATGGCAACTGC
CGCACGGTGTGCGTAAAGCGGCCGCTCTCCGTCACCCGCAAGGCAACTGCGAGCACACGGTGAAC
ACGTCGCTCACCGGATCGCTCAAGGAGCTGAAGCGAACGCCACGGAGCGGGCACC
GTGTCGCTGCCAACGACCCAGGAAGGGCTTCTTGACACGACCCAGAGCGAGGCCACCG
TCGGGAGGAGCACAACCTGGCTTCTGCGCGACAGACGCCCTCGAAGGGGGAGGATT
GCTGCCGAGGAGCTGCTCAAGGAGCTGCTCAGCACCGCTCCGCTAACACACTCTCGCC
TCGTCGCAAGGCAAGTGCAGCTCTGTCACCCGCAAGGCAACTGCGGAGATCGGACTCCG
GACATGGTGTGCGTAAAGCGGCCGACTCACCGGAAGCGGGAGAGCGAAGTGTG
GCCGGCGGGAGTGGCGCCCAAGCTTCAAGCGCTTGAGGGCGGCCAGCTCGGCG
GCCAGGCCAGTGTGCAACACCGCTTACCTCGAGACGGACATGAAGTGT
ATCAAGGAGGAGTACCGCTGAGGAGTACGACGCGAGCAGATGAACGCCAACACGGCACC
TTGTGAGGCCAACGGCTGCTGTTGAGCGAGAACGGGCCGAGCTGGAGTCTTCTA
ACCGAGCTGGAGGAGCTGATAAGATCGCGCAGCAGGTGCCAGCATCACGCCCTTCAG
CAGCACCTGCCGCAACG
```

## 2.2.2 The function `read.alignment()`

Aligned sequence data are very important in evolutionary studies, in this representation all vertically aligned positions are supposed to be homologous, that is sharing a common ancestor. This is a mandatory starting point for comparative studies. There is a function in `seqinR` called `read.alignment()` to read

aligned sequences data from various formats (`mase`, `clustal`, `phylip`, `fasta` or `msf`) produced by common external programs for multiple sequence alignment.

```
example(read.alignment)

rd.lgn> mase <- read.alignment(file = system.file("sequences/test.mase",
  package = "seqinr"), format = "mase")
rd.lgn> clustal <- read.alignment(file = system.file("sequences/test.aln",
  package = "seqinr"), format = "clustal")
rd.lgn> phylip <- read.alignment(file = system.file("sequences/test.phylip",
  package = "seqinr"), format = "phylip")
rd.lgn> msf <- read.alignment(file = system.file("sequences/test.msf",
  package = "seqinr"), format = "msf")
rd.lgn> fasta <- read.alignment(file = system.file("sequences/Anouk.fasta",
  package = "seqinr"), format = "fasta")
```

### 2.2.3 A simple example with the louse-gopher data

Let's give an example. The gene coding for the mitochondrial cytochrome oxidase I is essential and therefore often used in phylogenetic studies because of its ubiquitous nature. The following two sample tests of aligned sequences of this gene (extracted from ParaFit [34]), are distributed along with the `seqinR` package:

```
louse <- read.alignment(system.file("sequences/louse.fasta",
  package = "seqinr"), format = "fasta")
louse$nam

[1] "gi|548117|gb|L32667.1|GYDCYTOXIB" "gi|548119|gb|L32668.1|GYDCYTOXIC"
[3] "gi|548121|gb|L32669.1|GYDCYTOXID" "gi|548125|gb|L32671.1|GYDCYTOXIF"
[5] "gi|548127|gb|L32672.1|GYDCYTOXIG" "gi|548131|gb|L32675.1|GYDCYTOXII"
[7] "gi|548133|gb|L32676.1|GYDCYTOXIJ" "gi|548137|gb|L32678.1|GYDCYTOXIL"

gopher <- read.alignment(system.file("sequences/gopher.fasta",
  package = "seqinr"), format = "fasta")
gopher$nam

[1] "gi|548223|gb|L32683.1|PPGCYTOXIA" "gi|548197|gb|L32686.1|OGOCYTOXIA"
[3] "gi|548199|gb|L32687.1|OGOCYTOXIB" "gi|548201|gb|L32691.1|OGOCYTOXIC"
[5] "gi|548203|gb|L32692.1|OGOCYTOXID" "gi|548229|gb|L32693.1|PPGCYTOXID"
[7] "gi|548231|gb|L32694.1|PPGCYTOXIE" "gi|548205|gb|L32696.1|OGOCYTOXIE"
```

The aligned sequences are now imported in your  environment. The 8 genes of the first sample are from various species of louse (insects parasitic on warm-blooded animals) and the 8 genes of the second sample are from their corresponding gopher hosts (a subset of rodents), see figure 2.2 :

```
l.names <- readLines(system.file("sequences/louse.names",
  package = "seqinr"))
l.names

[1] "G.chapini" "G.cherriei" "G.costaric" "G.ewingi" "G.geomys" "
[6] "G.oklahome" "G.panamens" "G.setzeri" "
```

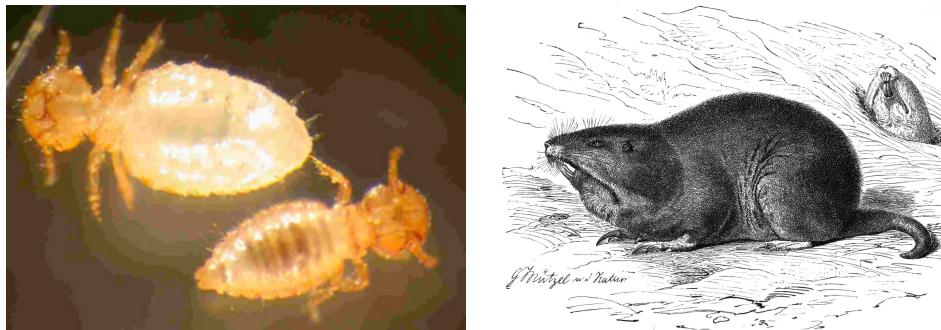


Figure 2.2: Louse (left) and gopher (right). Images are from the wikipedia (<http://www.wikipedia.org/>). The picture of the chewing louse *Damalinia limbata* found on Angora goats was taken by Fiorella Carnevali (ENEA, Italy). The gopher drawing is from Gustav Müntzel, Brehms Tierleben, Small Edition 1927.

```

g.names <- readLines(system.file("sequences/gopher.names",
  package = "seqinr"))
g.names

[1] "G.brevicep" "O.cavator"  "O.cherriei" "O.underwoo" "O.hispidus" "
[6] "G.burs1"    "G.burs2"   "O.heterodu"

```

**SeqinR** has very few methods devoted to phylogenetic analyses but many are available in the **ape** package. This allows for a very fine tuning of the graphical outputs of the analyses thanks to the power of the R facilities. For instance, a natural question here would be to compare the topology of the tree of the hosts and their parasites to see if we have congruence between host and parasite evolution. In other words, we want to display two phylogenetic trees face to face. This would be tedious with a program devoted to the display of a single phylogenetic tree at time, involving a lot of manual copy/paste operations, hard to reproduce, and then boring to maintain with data updates.

How does it looks under R? First, we need to *infer* the tree topologies from data. Let's try as an *illustration* the famous neighbor-joining tree estimation of Saitou and Nei [53] with Jukes and Cantor's correction [28] for multiple substitutions.

```

library(ape)
louse.JC <- dist.dna(x = lapply(louse$seq, s2c), model = "JC69")
gopher.JC <- dist.dna(x = lapply(gopher$seq, s2c), model = "JC69")
l <- nj(louse.JC)
g <- nj(gopher.JC)

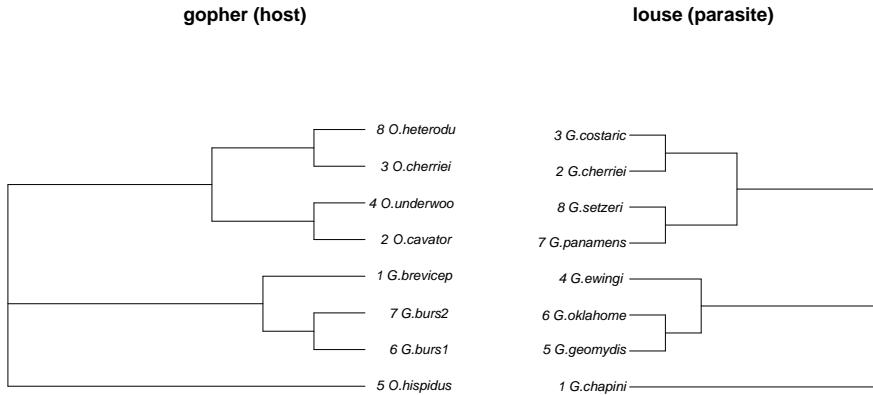
```

Now we have an estimation for *illustrative* purposes of the tree topology for the parasite and their hosts. We want to plot the two trees face to face, and for this we must change R graphical parameters. The first thing to do is to save the current graphical parameter settings so as to be able to restore them later:

```
op <- par(no.readonly = TRUE)
```

The meaning of the `no.readonly = TRUE` option here is that graphical parameters are not all settable, we just want to save those we can change at will. Now, we can play with graphics :

```
g$tip.label <- paste(1:8, g.names)
l$tip.label <- paste(1:8, l.names)
layout(matrix(data = 1:2, nrow = 1, ncol = 2), width = c(1.4,
  1))
par(mar = c(2, 1, 2, 1))
plot(g, adj = 0.8, cex = 1.4, use.edge.length = FALSE, main = "gopher (host)",
  cex.main = 2)
plot(l, direction = "l", use.edge.length = FALSE, cex = 1.4,
  main = "louse (parasite)", cex.main = 2)
```



We now restore the old graphical settings that were previously saved:

```
par(op)
```

OK, this may look a little bit obscure if you are not fluent in programming, but please try the following experiment. In your current working directory, that is in the directory given by the `getwd()` command, create a text file called `essai.r` with your favourite text editor, and copy/paste the previous R commands, that is :

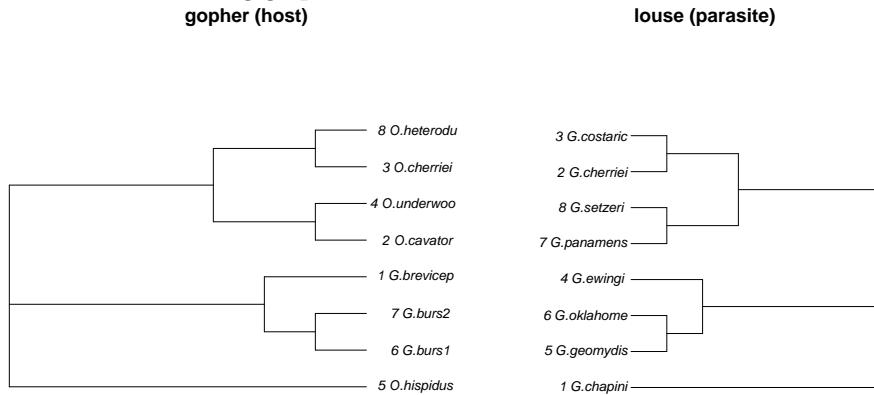
```
louse <- read.alignment(system.file("sequences/louse.fasta", package = "seqinr"), format = "fasta")
gopher <- read.alignment(system.file("sequences/gopher.fasta", package = "seqinr"), format = "fasta")
l.names <- readlines("http://pbil.univ-lyon1.fr/software/SequinR/Datasets/louse.names")
g.names <- readlines("http://pbil.univ-lyon1.fr/software/SequinR/Datasets/gopher.names")
louse.JC <- dist.dna(x = lapply(louse$seq, s2c), model = "JC69" )
gopher.JC <- dist.dna(x = lapply(gopher$seq, s2c), model = "JC69" )
l <- nj(louse.JC)
g <- nj(gopher.JC)
g$tip.label <- paste(1:8, g.names)
l$tip.label <- paste(1:8, l.names)
layout(matrix(data = 1:2, nrow = 1, ncol = 2), width=c(1.4, 1))
par(mar=c(2,1,2,1))
plot(g, adj = 0.8, cex = 1.4, use.edge.length=FALSE,
  main = "gopher (host)", cex.main = 2)
plot(l,direction="l", use.edge.length=FALSE, cex = 1.4,
  main = "louse (parasite)", cex.main = 2)
```

Make sure that your text has been saved and then go back to console to enter the command :

```
source("essai.r")
```

This should reproduce the previous face-to-face phylogenetic trees in your graphical device. Now, your boss is unhappy with working with the Jukes and

Cantor's model [28] and wants you to use the Kimura's 2-parameters distance [32] instead. Go back to the text editor to change `model = "JC69"` by `model = "K80"`, save the file, and in the  console `source("essai.r")` again, you should obtain the following graph :



Nice congruence, isn't it? Now, something even worst, there was a error in the aligned sequence set : the first base in the first sequence in the file `louse.fasta` is not a C but a T. To locate the file on your system, enter the following command:

```
system.file("sequences/louse.fasta", package = "seqinr")
```

```
[1] ".../.../.../.../seqinr.Rcheck/seqinr/sequences/louse.fasta"
```

Open the `louse.fasta` file in your text editor, fix the error, go back to the  console to `source("essai.r")` again. That's all, your graph is now consistent with the updated dataset.

## 2.3 Session Informations

This part was compiled under the following  environment:

- R version 2.4.1 (2006-12-18), i386-apple-darwin8.8.1
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: MASS 7.2-30, ade4 1.4-2, ape 1.9-3, gee 4.13-12, lattice 0.14-16, nlme 3.1-78, seqinr 1.0-7, xtable 1.4-3

---

---

# CHAPTER 3

---

## Importing sequences from ACNUC databases

Charif, D. Lobry, J.R.

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---

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### 3.1 Introduction

As a rule of thumb, after compression one nucleotide needs one octet of disk space storage (because you need also the annotations corresponding to the sequences), so that most likely you won't have enough space on your computer to work with a local copy of a complete DNA database. The idea is to import under  only the subset of sequences you are interested in. This is done in three steps:

1. Choose the bank you want to work with.
2. Select the sequences you are interested in.
3. Retrieve sequences from server into your workspace.

We now give a full example of those three steps under the ACNUC system [13, 14, 19, 17, 18].

#### 3.1.1 Choose a bank

Select the database from which you want to extract sequences with the `choosebank()` function. This function initiates a remote access to an ACNUC database. Called without arguments, `choosebank()` returns the list of available databases:

```
choosebank()

[1] "genbank"      "embl"        "emblwgs"      "swissprot"
[5] "ensembl"      "refseq"       "nrsub"       "nbrf"
[9] "hobacnucl"    "hobacprot"    "hovernucl"   "hoverprot"
[13] "hogennucl"    "hogenprot"    "hoverclnu"   "hoverclpr"
[17] "homolensprot" "homolensnucl" "greview"     "emplib"
[21] "HAMAPnucl"    "HAMAPprot"   "hoppsigen"   "nurebnucl"
[25] "nurebprot"    "taxobacgen"
```

Biological sequence databases are fast moving targets, and for publication purposes it is recommended to specify on which release you were working on when you made the job. To get more informations about available databases on the server, just set the `infobank` parameter to TRUE. For instance, here is the result for the three first databases on the default server at the compilation time (April 15, 2007) of this document:

```
choosebank(infobank = TRUE)[1:3, ]

      bank status
1 genbank  on
2 embl    on
3 emblwgs on

           info
1 GenBank Rel. 158 (15 February 2007) Last Updated: Mar 10, 2007
2 EMBL Library Release 89 (December 2006) Last Updated: Mar 8, 2007
3 EMBL Whole Genome Shotgun sequences Release 89 (December 2006)
```

Note that there is a `status` column because a database could be unavailable for a while during updates. If you try call `choosebank(bank = "bankname")` when the bank called `bankname` is off from server, you will get an explicit error message stating that this bank is temporarily unavailable, for instance:

```
choosebank("off")
```

```
Error in choosebank("off") : Database with name -->off<-- is currently off for maintenance,
please try again later.
```

Some special purpose databases are not listed by default. These are *tagged* databases that are only listed if you provide an explicit `tagbank` argument to the `choosebank()` function. Of special interest for teaching purposes is the TP tag, an acronym for *Travaux Pratiques* which means "practicals", and corresponds to *frozen* databases so that you can set up a practical whose results are stable from year to year. Currently available frozen databases at the default server are:

```
choosebank(tagbank = "TP", infobank = TRUE)
```

```
      bank status
1     emblTP  on
2 swissprotTP on
3 hoverprotTP on
4 hovernuclTP on
5     trypano on

           info
1          EMBL Library Release 78 WITHOUT ESTs (March 2004)
2 UniProt Rel. 1 (SWISS-PROT 43 + TrEMBL 26 + NEW): Last Updated: May 3, 2004
3          HOVERGEN - Release 45 (Jan 22 2004) Last Updated: Jan 22, 2004
4          HOVERGEN - Release 45 (Jan 22 2004) Last Updated: Jan 22, 2004
5          trypano Rel. 1 (27 Janvier 2004) Last Updated: Jan 27, 2004
```

Now, if you want to work with a given database, say GenBank, just call `choosebank()` with "genbank" as its first argument and store the result in a variable in the workspace, called for instance `mybank` in the example thereafter:

```
mybank <- choosebank("genbank")
str(mybank)
```

```
List of 8
$ socket  :Classes 'sockconn', 'connection'  int 21
$ bankname: chr "genbank"
$ totseqs : chr "71980262"
$ totspecs: chr "437097"
$ totkeys : chr "6375122"
$ release : chr "GenBank Rel. 158 (15 February 2007) Last Updated: Mar 10, 2007"
$ status  :Class 'AsIs'  chr "on"
$ details : chr [1:3] "GenBank Rel. 158 (15 February 2007) Last Updated: Mar 10, 2007" "72,371,122,329 bases; 68,158
```

The list returned by `choosebank()` here means that in the database called `genbank` at the compilation time of this document there were 71,980,262 sequences from 437,097 species and a total of 6,375,122 keywords. The status of the bank was `on`, and the release information was `GenBank Rel. 158 (15 February 2007) Last Updated: Mar 10, 2007`. For specialized databases, some relevant informations are also given in the `details` component, for instance:

```
moreinfo <- choosebank("taxobacgen")$details
cat(moreinfo, sep = "\n")
```

```
TaxoBacGen Rel. 7 (September 2005)
1,151,149,763 bases; 254,335 sequences; 847,767 subseqs; 63,879 refers.
Data compiled from GenBank by Gregory Devulder
Laboratoire de Biometrie & Biologie Evolutive, Univ Lyon I
-----
This database is a taxonomic genomic database.
It results from an expertise crossing the data nomenclature database DSMZ
[http://www.dsmz.de/species/bacteria.htm Deutsche Sammlung von
Mikroorganismen und Zellkulturen GmbH, Braunschweig, Germany]
and GenBank.
- Only contains sequences described under species present in
Bacterial Nomenclature Up-to-date.
- Names of species and genus validly published according to the
Bacteriological Code (names with standing in nomenclature) is
added in field "DEFINITION".
- A keyword "type strain" is added in field "FEATURES/source/strain" in
GenBank format definition to easily identify Type Strain.
Taxobacgen is a genomic database designed for studies based on a strict
respect of up-to-date nomenclature and taxonomy.
```

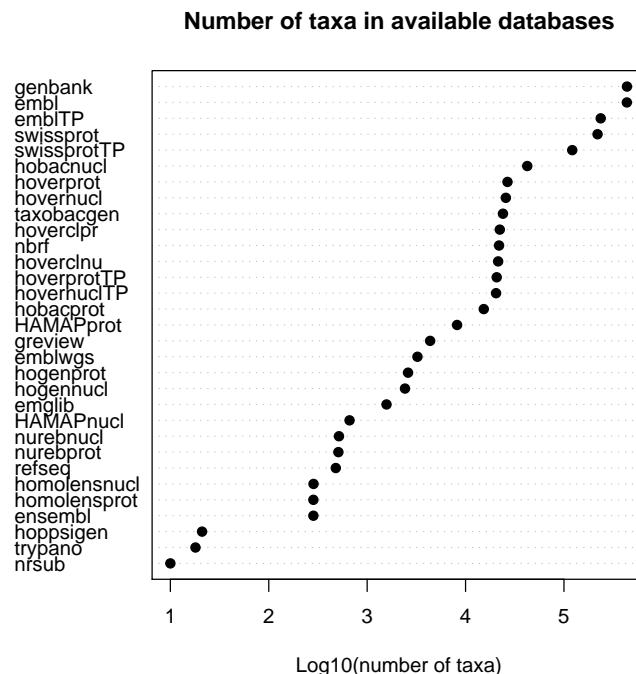
The previous command has a side-effect that is worth mentioning. As from `seqinR` 1.0-3, the result of the `choosebank()` function is automatically stored in a global variable named `banknameSocket`, so that if no socket argument is given to the `query()` function, the last opened database will be used by default for your requests. This is just a matter of convenience so that you don't have to explicitly specify the details of the socket connection when working with the last opened database. You have, however, full control of the process since `choosebank()` returns (invisibly) all the required details. There is no trouble to open *simultaneously* many databases. You are just limited by the number of simultaneous connections your build of `R` is allowed<sup>1</sup>.

---

<sup>1</sup> As from `R` 2.4.0 the maximum number of open connections has been increased from 50 to 128. Note also that there is a very convenient function called `closeAllConnections()` in the `R` base package if you want to close all open connections at once.

For advanced users who may wish to access to more than one database at time, a good advice is to close them with the function `closebank()` as soon as possible so that the maximum number of simultaneous connections is never reached. In the example below, we want to display the number of taxa (*i.e.* the number of nodes) in the species taxonomy associated with each available database (including frozen databases). For this, we loop over available databases and close them as soon as the information has been retrieved.

```
banks <- c(choosebank(), choosebank(tagbank = "TP"))
ntaxa <- numeric(0)
for (i in banks) {
  ntaxa[i] <- as.numeric(choosebank(i)$totspecs)
  closebank()
}
dotchart(log10(ntaxa[order(ntaxa)]), pch = 19, main = "Number of taxa in available databases",
         xlab = "Log10(number of taxa)")
```



### 3.1.2 Make your query



*Felis catus*. Source: wikipedia

For this section, set up the default bank to GenBank, so that you don't have to provide the sockets details for the `query()` function:

```
choosebank("genbank")
```

Then, you have to say what you want, that is to compose a query to select the subset of sequences you are interested in. The way to do this is documented under `?query`, we just give here a simple example. In the query below, we want to select all the coding sequences (`t=cds`) from cat (`AND sp=felis catus`) that are not (AND NOT) partial sequences (`k=partial`). We want the result to be stored in an object called `completeCatsCDS`.

```
query("completeCatsCDS", "sp=felis catus AND t=cds AND NOT k=partial")
```

Now, there is in the workspace an object called `completeCatsCDS`, which does not contain the sequences themselves but the *sequence names* (and various relevant informations such as the genetic code and the frame) that fit the query. They are stored in the `req` component of the object, let's see the name of the first ten of them:

```
sapply(completeCatsCDS$req[1:10], getName)
```

```
[1] "AB000483.PE1"      "AB000484.PE1"      "AB000485.PE1"      "AB004237"
[5] "AB004238"         "AB009279.PE1"      "AB009280.PE1"      "AB010872.UGT1A1"
[9] "AB011965.SDF-1A"   "AB011966.SDF-1B"
```

The first sequence that fit our request is `AB000483.PE1`, the second one is `AB000484.PE1`, and so on. Note that the sequence name may have an extension, this corresponds to *subsequences*, a specificity of the ACNUC system that allows to handle easily a subsequence with a biological meaning, typically a gene. The list of available subsequences in a given database is given by the function `getType()`, for example the list of available subsequences in GenBank is given in table 3.1.

	Type	Description
1	CDS	.PE protein coding region
2	LOCUS	sequenced DNA fragment
3	MISC_RNA	.RN other structural RNA coding region
4	RRNA	.RR mature ribosomal RNA
5	SCRNA	.SC small cytoplasmic RNA
6	SNRNA	.SN small nuclear RNA
7	TRNA	.TR mature transfer RNA

Table 3.1: Available subsequences in genbank

The component `call` of `completeCatsCDS` keeps automatically a trace of the way you have selected the sequences:

```
completeCatsCDS$call
```

```
query(listname = "completeCatsCDS", query = "sp=felis catus AND t=cds AND NOT k=partial")
```

At this stage you can quit your  session saving the workspace image. The next time an  session is opened with the workspace image restored, there will be an object called `completeCatsCDS`, and looking into its `call` component will tell you that it contains the names of complete coding sequences from *Felis catus*.

In practice, queries for sequences are rarely done in one step and are more likely to be the result of an iterative, progressively refining, process. An important point is that a list of sequences can be re-used. For instance, we can re-use `completeCatsCDS` to get only the list of sequences that were published in 2004:

```
query("ccc2004", "completeCatsCDS AND y=2004")
length(ccc2004$req)
```

```
[1] 58
```

Hence, there were 58 complete coding sequences published in 2004 for *Felis catus* in GenBank.

As from release 1.0-3 of the **seqinR** package, there is new parameter **virtual** which allows to disable the automatic retrieval of information for all list elements. This is interesting for list with many elements, for instance :

```
query("allcds", "t=cds", virtual = TRUE)
allcds$nelem
```

```
[1] 4114605
```

There are therefore 4,114,605 coding sequences in this version of GenBank<sup>2</sup>. It would be long to get all the informations for the elements of this list, so we have set the parameter **virtual** to TRUE and the **req** component of the list has not been documented:

```
allcds$req
```

```
[[1]]
[1] NA
```

However, the list can still be re-used<sup>3</sup>, for instance we may extract from this list all the sequences from, say, *Mycoplasma genitalium*:

```
query("small", "allcds AND sp=mycoplasma genitalium", virtual = TRUE)
small$nelem
```

```
[1] 916
```

There are then 916 elements in the list **small**, so that we can safely repeat the previous query without asking for a virtual list:

```
query("small", "allcds et sp=mycoplasma genitalium")
sapply(small$req, getName)[1:10]
```

```
[1] "AY191424" "AY386807" "AY386808" "AY386809" "AY386810" "AY386811"
[7] "AY386812" "AY386813" "AY386814" "AY386815"
```

Here are some illustrations of using virtual list to answer simple questions about the current GenBank release.

**Man.** How many sequences are available for our species?

```
query("man", "sp=homo sapiens", virtual = T)
man$nelem
```

```
[1] 10771591
```

There are 10,771,591 sequences from *Homo sapiens*.

---

<sup>2</sup> which is stored in the **release** component of the object **banknameSocket** and current value is today (April 15, 2007): **banknameSocket\$release** = GenBank Rel. 158 (15 February 2007) Last Updated: Mar 10, 2007.

<sup>3</sup> of course, as long as the socket connection with the server has not been lost: virtual lists details are only known by the server.

**Sex.** How many sequences are annotated with a keyword starting by sex?

```
query("sex", "k=sex@", virtual = T)
sex$nelem
```

```
[1] 1169
```

There are 1,169 such sequences.

**tRNA.** How many complete tRNA sequences are available?

```
query("trna", "t=trna AND NOT k=partial", virtual = T)
trna$nelem
```

```
[1] 210285
```

There are 210,285 complete tRNA sequences.

**Nature vs. Science.** In which journal were the more sequences published?

```
query("nature", "j=nature", virtual = T)
nature$nelem
```

```
[1] 1533935
```

```
query("science", "j=science", virtual = T)
science$nelem
```

```
[1] 1281344
```

There are 1,533,935 sequences published in *Nature* and 1,281,344 sequences published in *Science*, so that the winner is *Nature*.

**Smith.** How many sequences have Smith (last name) as author?

```
query("smith", "au=smith", virtual = T)
smith$nelem
```

```
[1] 1860052
```

There are 1,860,052 such sequences.

**YK2.** How many sequences were published after year 2000 (included)?

```
query("yk2", "y>2000", virtual = T)
yk2$nelem
```

```
[1] 56700047
```

There are 56,700,047 sequences published after year 2000.

**Organelle contest.** Do we have more sequences from chloroplast genomes or from mitochondrion genomes?

```
query("chloro", "o=chloroplast", virtual = T)
chloro$nelem
```

```
[1] 131872
```

```
query("mito", "o=mitochondrion", virtual = T)
mito$nelem
```

```
[1] 446692
```

There are 131,872 sequences from chloroplast genomes and 446,692 sequences from mitochondrion genomes, so that the winner is mitochondrion.

### 3.1.3 Extract sequences of interest

#### Introduction

There are two functions to get the sequences. The first one, `getSequence()`, uses regular socket connections, the second one, `extractseq()`, uses zlib compressed sockets, which is faster but the function is experimental and has not been extensively tested.

#### Extracting sequences with `getSequence()`

For this section we set up the bank to `emb1TP` which is a frozen subset of EMBL database to allow for the reproducibility of results.

```
choosebank("emb1TP")
```

We suppose that the sequences we are interested in are all the complete coding sequences from *Felis catus* :

```
query("completeCatsCDS", "sp=felis catus AND t=cds AND NOT k=partial")
(nseq <- completeCatsCDS$nelem)
```

```
[1] 257
```

Thus, there were 257 complete CDS from *Felis catus* in this release of EMBL.

The sequences are obtained with the function `getSequence()`. For example, the first 50 nucleotides of the first sequence of our request are:

```
myseq <- getSequence(completeCatsCDS$req[[1]])
myseq[1:50]
```

```
[1] "a" "t" "g" "a" "c" "a" "a" "c" "a" "a" "t" "t" "t" "c" "g" "a" "a" "a" "a"
[19] "t" "c" "a" "c" "a" "c" "c" "c" "c" "c" "t" "t" "a" "c" "c" "a" "a" "a"
[37] "a" "t" "t" "a" "t" "a" "a" "t" "c" "a" "c" "t" "c"
```

They can also be coerced as string of character with the function `c2s()`:

```
c2s(myseq[1:50])
```

```
[1] "atgaccaacattcgaaaatcacaccccttaccaaattattaatcactc"
```

Note that what is done by `getSequence()` is much more complex than a substring extraction because subsequences of biological interest are not necessarily contiguous or even on the same DNA strand. Consider for instance the following coding sequence from sequence AE003734:

```
query("trs", "N=AE003734.PE35")
annots <- getAnnot(trs$req[[1]])
cat(annots, sep = "\n")

FT      CDS          join(complement(153944..154157),complement(153727..153866),
FT                      complement(152185..153037),138523..138735,138795..138955)
FT                      /codon_start=1
FT                      /db_xref="FLYBASE:FBgn0002781"
FT                      /db_xref="GOA:Q86B86"
FT                      /db_xref="TrEMBL:Q86B86"
FT                      /note="mod(mdg4) gene product from transcript CG32491-RZ;
FT                      trans splicing"
FT                      /gene="mod(mdg4)"
```

```

FT          /product="CG32491-PZ"
FT          /locus_tag="CG32491"
FT          /protein_id="AA041581.1"
FT          /translation="MADDEQFSLCWNNFNTNLSAGFHESLCRGDLVDVSLAAEGQIVKA
FT          HRLVLSVCSPFFRKMFQTQMPNSNTHAIVFLNNVSHSALKDLIQFMYCGEVNVKQDALPAF
FT          ISTAESLQIKGLTDMDPAPQPQESSPPAAAPHVQQQIPAQRVQRQPRASARYKLET
FT          VDDGLGDEKQSTTQIVIQTAAAPQATIVQQQPQQAAQQIQSQQLQTGTTTATLVSTN
FT          KRSAQRSSLT PASSSAGVKSRTSTSANVMDPLDSTTEGATTAAQLVPQQITVQTSVV
FT          SAAEAKLHQQSPQQVRQEEAEYIDLPMELPTKSEPDYSEDHGDAAGDAEGTYVEDDTYG
FT          DMRYDDSYFTENEDAGNQTAANTSQQGVATTSSKAVVKQQSQNYSESSFVDTSGDQGNT
FT          EAQVITQHVNRNCGPQMFLISRKGGLTINNNFVYRSNLKFFGKSNILYWECVQNRSVKC
FT          RSRLLTIGDDLYVTNDVHNHMGDNKRIEAAKAGMLIHKKLSSLTAADKIQGSWKMDE
FT          GNPDHLPKM"

```

To get the coding sequence manually you would have to join 5 different pieces from AE003734 and some of them are in the complementary strand. With `getSequence()` you don't have to think about this. Just make a query with the sequence name:

```

query("transspliced", "N=AE003734.PE35")
length(transspliced$req)

```

```

[1] 1

getName(transspliced$req[[1]])

[1] "AE003734.PE35"

```

Ok, now there is in your workspace an object called `transspliced` which `req` component is of length one (because you have asked for just one sequence) and the name of the single element of the `req` component is AE003734.PE35 (because this is the name of the sequence you wanted). Let see the first 50 base of this sequence:

```

getSequence(transspliced$req[[1]])[1:50]

```

```

[1] "a" "t" "g" "c" "g" "a" "c" "g" "a" "c" "g" "a" "g" "c" "a" "a"
[19] "t" "t" "c" "a" "g" "c" "t" "t" "g" "t" "g" "c" "t" "g" "g" "a" "a" "c"
[37] "a" "a" "c" "t" "t" "c" "a" "a" "c" "a" "c" "g" "a" "a" "a"

```

All the complex transsplicing operations have been done here. You can check that there is no in-frame stop codons<sup>4</sup> with the `getTrans()` function to translate this coding sequence into protein:

```

getTrans(transspliced$req[[1]])[1:50]

```

```

[1] "M" "A" "D" "D" "E" "Q" "F" "S" "L" "C" "W" "N" "N" "F" "N" "T" "N" "L"
[19] "S" "A" "G" "F" "H" "E" "S" "L" "C" "R" "G" "D" "L" "V" "D" "V" "S" "L"
[37] "A" "A" "E" "G" "Q" "I" "V" "K" "A" "H" "R" "L" "V" "L"

```

```

table(getTrans(transspliced$req[[1]]))

```

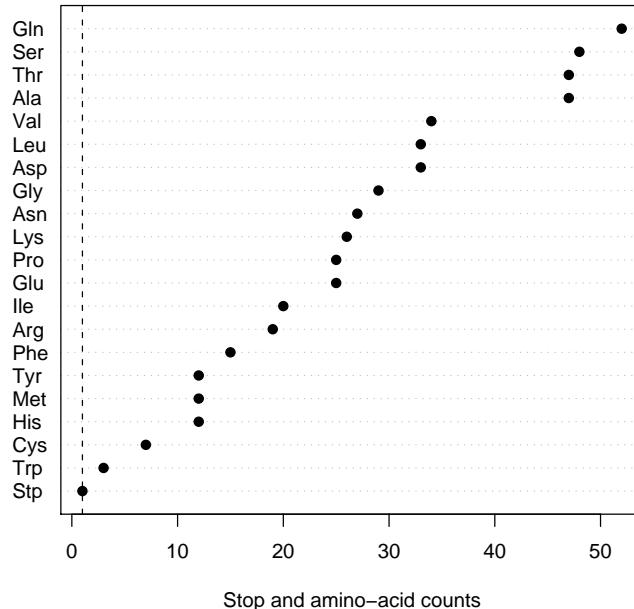
	A	C	D	E	F	G	H	I	K	L	M	N	P	Q	R	S	T	V	W	Y
1	47	7	33	25	15	29	12	20	26	33	12	27	25	52	19	48	47	34	3	12

In a more graphical way:

---

<sup>4</sup> Stop codons are represented by the character \* when translated into protein.

```
aaccount <- table(getTrans(transspliced$req[[1]]))
aaccount <- aaccount[order(aaccount)]
names(aaccount) <- aaa(names(aaccount))
dotchart(aaccount, pch = 19, xlab = "Stop and amino-acid counts",
         main = "There is only one stop codon in AE003734.PE35")
abline(v = 1, lty = 2)
```

**There is only one stop codon in AE003734.PE35**

Note that the relevant variant of the genetic code was automatically set up during the translation of the sequence into protein. This is because the `transspliced$req[[1]]` object belongs to the `SeqAcnucWeb` class:

```
class(transspliced$req[[1]])
[1] "SeqAcnucWeb"
```

Therefore, when you are using the `getTrans()` function, you are automatically redirected to the `getTrans.SeqAcnucWeb()` function which knows how to take into account the relevant frame and genetic code for your coding sequence.

### Extracting sequences with `extractseqs()` (experimental)

The timings here were from an home-ADSL connection, and are only indicative. For this section we set up the bank to `emblTP` which is a frozen subset of EMBL database to allow for the reproducibility of results.

```
(tcb <- system.time(choosebank("emblTP")))
[1] 0.101 0.004 0.858 0.000 0.000
```

It was then about 1 seconds to select the relevant database. We suppose that the sequences we are interested in are all the complete coding sequences from *Homo sapiens* that are encoded in the nucleus (we don't want sequences from human mitochondrion). We use a virtual query to speed up things:

```
(tqu <- system.time(query("hsCDS", "sp=Homo sapiens AND t=cds AND o=nuclear AND NOT k=partial",
                           virtual = TRUE)))
```

```
[1] 0.002 0.000 10.628 0.000 0.000
```

```
(nseq <- hsCDS$nelem)
```

```
[1] 78573
```

It was then about 11 seconds to create on the server a list of 78573 sequences. We download now the sequences in zlib compressed mode:

```
(tex <- system.time(mycds <- extractseqs("hsCDS")))
```

```
[1] 7.920 1.099 150.114 0.000 0.000
```

It was then about 150 seconds to dowload the sequences in the object mycds, which looks like :

```
cat(head(mycds), sep = "\n")
```

```
>A00127.PE1          2217 residues
ATGCGGGGTCCGAGCGGGGCTCTGGCTGCTCTGGCTCTGCGCACCGTGCCTGGAGGC
ATGGAGGTGGGTGGCGCACCTCGGACCCAGACAGCACAAGTGGCAACATGAGC
GAGGCCCTCCGGAAAGCGGGCATCCAGCCCTCCCTCTGCGTCGGGGCACCTCGGC
GACCACTGGCTCCAGCTCATCGCGCCAGGAGGCTGACGCCATCAGCTGGATGGAGGA
GCCATCTATGAGGCCGGAAAGGAGCACGGCCTGAACGGCTGGTGGCGAAGTGTACGAT
```

```
cat(tail(mycds), sep = "\n")
```

```
ATCACTGGCCCCAGAGAGAGAGGGCATAGGCCACGGCGGCCCAAGCTATGCTGCACA
CTGAGCTCCCTCAGCTCCGTGCTGAGACTGCGCCGGACCCGCTGACAGCGAGGAGGAG
GCAACCAGCGGCCAGGATGAACTGGCTGAACCCGCCTCCCGGGCCAGTTTCT
TCCCTCTCAGCCAGGGATGCTCGAGCAGCCACAGGGCAGGAACCTCTGACTGCCATC
CTGCTGCTGCTGGAGCTGGATGCAAGAGGGCTGGAGGCCGTGAGCAGACTGTGGC
AGCCGGCTGCAGGCCCTGCGTGGGAAGAGGTGCAGGAGCACGCCAGTGA
```

We save now the sequences in a local FASTA file for future use:

```
(twl <- system.time(writeLines(mycds, "mycds.fasta")))
```

```
[1] 0.830 0.867 2.977 0.000 0.000
```

It was then about 3 seconds to dump the sequences on a local file. We read the sequences as strings without setting attributes to save time:

```
(trf <- system.time(mycdss <- read.fasta("mycds.fasta", as.string = TRUE,
                                         set.attributes = FALSE))
```

```
[1] 29.173 0.404 29.588 0.000 0.000
```

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It was then about 30 seconds to read the sequences as strings. We save them in XDR format:

```
(tsrd <- system.time(save(mycdss, file = "mycdss.RData")))
```

```
[1] 38.850 0.271 39.531 0.000 0.000
```

It was then about 40 seconds to save the sequences in XDR format. How long is it to load the sequences from XDR format?

```
(tlrd <- system.time(load("mycdss.RData")))
```

```
[1] 1.183 0.043 1.226 0.000 0.000
```

It was then about 1 seconds to load the sequences from an XDR formated file. Now, we also want the corresponding proteins. We download the translated CDS from the server:

```
(texp <- system.time(myprot <- extractseqs("hsCDS", operation = "translate")))
```

```
[1] 2.725 0.641 159.953 0.000 0.000
```

It was then about 160 seconds to get the protein sequences from the server. The object `myprot` looks like:

```
cat(head(myprot), sep = "\n")
```

```
>A00127.PE1 739 residues
MRGPSGALWLLLARLRTVLGGMEVRWCATSDPEQHKCGNMSEAFREAGIQPSLLCVRGTSA
DHCVQLIAAQEADAITLDGGAIYEAGKEHGLKPVVGEVYDQEVTTSYYAVAVRRSSHVT
IDTLKGVKSCHTGINRTVGNVPVGYLVESGRSLVMGCDVLAWSYFGGSCVPGAGETS
YSESLCRLCRGDSSEGEGVCDKSPLERYYYDYSGAFRCLAEAGAGDVAFVKHSTVLNTDGKT
LPSWGQALLSQDFELLCRDGSRADVTEWRQCHLARVPAHAVVRAVDTDGGLIFRLLNEGQ
```

```
cat(tail(myprot), sep = "\n")
```

```
>Z93322.PE1 257 residues
MKLTRKMLTRAKASELHSVRKLMCWGSRLTDISICQEMPSLEVITLSVNSISTLEPVSR
CQRLESEYLRRNRIPSLAELFYLKGLPRLRVWLAEAPCCGTSPHYRMTVLRTLPRLQK
LDNQAVTEEEELSRLASEGEETAAPEREGIGHGCPKLCCTLSSISSAAETGRDPLDSEE
ATSGAQDERGLKPPSRGQFPLSARDASSSHRGRNVLTAILLLRELDAEGLAEVQQTVE
SRLQALRGEEVQEHAE*
```

We save the protein sequences in a local FASTA file for future use:

```
(tw12 <- system.time(writeLines(myprot, "myprot.fasta")))
```

```
[1] 0.305 0.306 0.610 0.000 0.000
```

It was then about 1 seconds to dump the protein sequences on a local file. We read the sequences as strings without setting attributes to save time:

```
(trf2 <- system.time(myprots <- read.fasta("myprot.fasta",
  as.string = TRUE, set.attributes = FALSE))
```

```
[1] 17.026 0.100 17.163 0.000 0.000
```

It was then about 17 seconds to read the protein sequences as strings. We save them in XDR format:

```
(tsrd2 <- system.time(save(myprots, file = "myprots.RData")))
```

```
[1] 3.779 0.160 3.939 0.000 0.000
```

It was then about 4 seconds to save the protein sequences in XDR format. How long is it to load the protein sequences from XDR format?

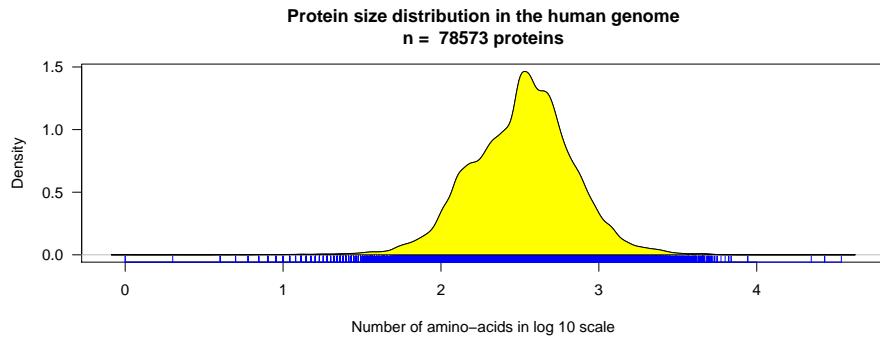
```
(tlrd2 <- system.time(load("myprots.RData")))
```

```
[1] 0.830 0.025 0.855 0.000 0.000
```

It was then about 1 seconds to load the protein sequences from an XDR formated file.

As a quick sanity check, we plot the distribution of protein size:

```
x <- log10(nchar(myprots) - 1)
dstx <- density(x)
plot(dstx, main = paste("Protein size distribution in the human genome\nn = ",
  length(myprots), "proteins"), xlab = "Number of amino-acids in log 10 scale",
  las = 1)
rug(x, col = "blue")
polycurve <- function(x, y, base.y = min(y), ...) polygon(x = c(min(x),
  x, max(x)), y = c(base.y, y, base.y), ...)
polycurve(dstx$x, dstx$y, col = "yellow")
```



## 3.2 The query language

### 3.2.1 Where to find information

The last version of the documentation for the query language is available online at <http://pbil.univ-lyon1.fr/databases/acnuc/cfonctions.html#QUERYLANGUAGE>. This documentation has been imported within the documentation of the `query()` function, but the last available update is the online version.

### 3.2.2 Case sensitivity and ambiguities resolution

The query language is case insensitive, for instance:

```
choosebank("emblTP")
query("lowercase", "sp=escherichia coli", virtual = TRUE)
query("uppercase", "SP=Escherichia coli", virtual = TRUE)
lowercase$nelem == uppercase$nelem
```

```
[1] TRUE
```

```
closebank()
```

Three operators (AND, OR, NOT) can be ambiguous because they can also occur within valid criterion values. Such ambiguities can be solved by encapsulating elementary selection criteria between escaped double quotes. For example:

```
choosebank("emblTP")
query("ambig", "\"sp=Beak and feather disease virus\" AND \"au=ritchie\"", 
      virtual = T)
ambig$nelem
```

```
[1] 18
```

```
closebank()
```

### 3.2.3 Selection criteria

#### Introduction

Selection criteria are in the form `c=something` (without space before the = sign) or `list_name` where `list_name` is a previously constructed list.

`SP=taxon`

This is used to select sequences attached to a given taxon or any other below in the tree. The at sign @ substitutes as a wildcard character for any zero or more characters. Here are some examples:

```
choosebank("emblTP")
query("bb", "sp=Borrelia burgdorferi", virtual = T)
bb$nelem
```

```
[1] 1682
```

```
query("borrelia", "sp=Borrelia", virtual = T)
borrelia$nelem
```

```
[1] 3173
```

```
closebank()
```

Here is an example of use of the wildcard @ to look for sapiens species:

```

choosebank("emb1TP")
query("sapiens", "sp=@sapiens@", virtual = T)
sapiens$nelem

[1] 2216556

query("sapienspecies", "PS sapiens")
sapply(sapienspecies$req, getName)

[1] "HOMO SAPIENS"
[2] "HOMO SAPIENS NEANDERTHALENSIS"
[3] "HOMO SAPIENS X HUMAN PAPILLOMAVIRUS TYPE"
[4] "HOMO SAPIENS X SIMIAN VIRUS 40"
[5] "HOMO SAPIENS X HUMAN ENDOGENOUS RETROVIR"
[6] "HOMO SAPIENS X HUMAN T-CELL LYMPHOTROPIC"
[7] "HEPATITIS B VIRUS X HOMO SAPIENS"
[8] "HOMO SAPIENS X HEPATITIS B VIRUS"
[9] "HOMO SAPIENS X HUMAN IMMUNODEFICIENCY VI"
[10] "SYNTHETIC CONSTRUCT X HOMO SAPIENS"
[11] "HUMAN PAPILLOMAVIRUS X HOMO SAPIENS"
[12] "MUS SP. X HOMO SAPIENS"
[13] "HOMO SAPIENS X HUMAN PAPILLOMAVIRUS"
[14] "HOMO SAPIENS X HUMAN ADENOVIRUS TYPE 5"
[15] "HOMO SAPIENS X HERV-H/ENV62"
[16] "HOMO SAPIENS X HERV-H/ENV60"
[17] "HOMO SAPIENS X HERV-H/ENV59"
[18] "EXPRESSION VECTOR PTH-HIN X HOMO SAPIENS"
[19] "ADENO-ASSOCIATED VIRUS 2 X HOMO SAPIENS"
[20] "SIMIAN VIRUS 40 X HOMO SAPIENS"
[21] "HOMO SAPIENS X MUS MUSCULUS"
[22] "HOMO SAPIENS X INFLUENZA B VIRUS (B/LEE/"
[23] "MUS MUSCULUS X HOMO SAPIENS"
[24] "CRICETULUS GRISEUS X HOMO SAPIENS"
[25] "TRYPANOSOMA CRUZI X HOMO SAPIENS"
[26] "HOMO SAPIENS X TRYPANOSOMA CRUZI"

closebank()

```

TID=id

This is used to select sequences attached to a given numerical NCBI's taxonomy ID. For instance, the taxonomy ID for *Homo sapiens neanderthalensis* is 63221:

```

choosebank("genbank")
query("hsn", "TID=63221", virtual = T)
hsn$nelem

[1] 1335

query("hsnsp", "PS hsn")
sapply(hsnsp$req, getName)

[1] "HOMO SAPIENS NEANDERTHALensis"

closebank()

```



*Homo neanderthalensis.*  
wikipedia

Source:

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### K=keyword

This is used to select sequences attached to a given keyword or any other below in the tree. The at sign @ substitutes as a wildcard character for any zero or more characters. Example:

```
choosebank("emblTP")
query("ecoliribprot", "sp=escherichia coli AND k=rib@ prot@", 
      virtual = T)
ecoliribprot$nelem
```

```
[1] 105
```

```
closebank()
```

### T=type

This is used to select sequences of specified type. The list of available type for the currently opened database is given by function `getType()`:

```
choosebank("emblTP")
getType()
```

sname	libel
2661 CDS	.PE protein coding region
2662 ID	.Locus entry
2663 MISC_RNA	.RN other structural RNA coding region
2664 RRNA	.RR Ribosomal RNA coding gene
2665 SCRNA	.SC small cytoplasmic RNA
2666 SNRNA	.SN small nuclear RNA
2667 tRNA	.TR Transfer RNA coding gene

```
closebank()
```

For instance, to select all coding sequences from *Homo sapiens* we can use:

```
choosebank("emblTP")
query("hscds", "sp=Homo sapiens AND t=cds", virtual = T)
hscds$nelem
```

```
[1] 150513
```

```
closebank()
```

### J=journal\_name

This is used to select sequences published in journal specified using defined journal code. For instance to select all sequences published in *Science*:

```
choosebank("emblTP")
query("allseqsfromscience", "J=Science", virtual = TRUE)
allseqsfromscience$nelem
```

```
[1] 930397
```

```
closebank()
```

The list of available journal code can be obtained from the `readsmj()` function this way:

```
choosebank("emblTP")
nl <- readfirstrec(type = "SMJ")
smj <- readsmj(nl = nl, all.add = TRUE)
head(smj[!is.na(smj$nature) & smj$nature == "journal", c("sname",
  "libel")])
```

sname	libel
ABP	Acta Biochim. Pol.
ABSTR-SOCNEUROSCI	Abstr. - Soc. Neurosci.
ABSTRGENMEETAMSOCM	Abstr. Gen. Meet. Am. Soc. Microbiol.
ABSTRMIDWINTERESM	Abstr. Midwinter Res. Meet. Assoc. Res. Otolaryngol.
ACTAAGRICSCANDAANI	Acta Agric. Scand. A Anim. Sci.
ACTABIOCHIMBIOPHYS	Acta Biochim. Biophys. Sin.

```
closebank()
```

#### R=refcode

This is used to select sequences from a given bibliographical reference specified as `jcode/volume/page`. For instance, to select sequences associated with the first publication [1] of the complete genome of *Rickettsia prowazekii*, we can use:

```
choosebank("emblTP")
query("rpro", "R=Nature/396/133")
sapply(rpro$req, getName)
```

```
[1] "RPDNAOMPB" "RPXX01"      "RPXX02"      "RPXX03"      "RPXX04"
```

```
closebank()
```

#### AU=name

This is used to select sequences having a specified author (only last name, no initial).

```
choosebank("emblTP")
query("Graur", "AU=Graur")
Graur$nelem
```

```
[1] 48
```

```
closebank()
```

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### AC=accession\_no

This is used to select sequences attached to specified accession number. For instance if we are looking for sequences attached to the accession number AY382159:

```

choosebank("emblTP")
query("ACexample", "AC=AY382159")
getName(ACexample$req[[1]])

[1] "AY382159"

annotations <- getAnnot(ACexample$req[[1]])
cat(annotations, sep = "\n")

ID  AY382159 standard; genomic DNA; PRO; 783 BP.
XX
AC  AY382159;
XX
SV  AY382159.1
XX
DT  08-OCT-2003 (Rel. 77, Created)
DT  08-OCT-2003 (Rel. 77, Last updated, Version 1)
XX
DE  Borrelia burgdorferi strain FP1 OspA gene, partial cds.
XX
KW .
XX
OS  Borrelia burgdorferi (Lyme disease spirochete)
OC  Bacteria; Spirochaetes; Spirochaetales; Spirochaetaceae; Borrelia;
OC  Borrelia burgdorferi group.
XX
RN  [1]
RP  1-783
RA  Hao Q., Wan K. ;
RT ;
RL  Submitted (03-SEP-2003) to the EMBL/GenBank/DDBJ databases.
RL  Department of Lyme Spirochetosis, CDC, Beijing 102206, China
XX
FH  Key          Location/Qualifiers
FH
FT  source       1..783
FT  /db_xref="taxon:139"
FT  /mol_type="genomic DNA"
FT  /organism="Borrelia burgdorferi"
FT  /strain="FP1"
FT  CDS           <1..>783
FT  /codon_start=1
FT  /transl_table=11
FT  /product="OspA"
FT  /protein_id="AAQ89576.1"
FT  /translation="ALIACKQNVSSLDEKNSASVDPGEMKVLSKEKDGDGYSLKAT
FT  VDKLELKGTSDKNNGSGTLEGEKTDKSKAKLTISDDLSKTTFEVKEDGKTLVSRKVSS
FT  KDKTSTDEMNEKGELSAKTMTRENGTKLEYTEMKSDGTGKTKEVLKNTLLEGRVANDK
FT  VTLEVKEGTVTLSKEIAKSGEVTVALNDTNTTQATKKTGAWSKTSLLTISVNSKKTTQ
FT  LVFTKQDTITVQYDSAGTNLEGTAVEIKTLDELKNALK"
XX
SQ  Sequence 783 BP; 342 A; 124 C; 145 G; 172 T; 0 other;

closebank()

```

### N=seq\_name

This is used to select sequences of a given name<sup>5</sup>. Sequences names are not necessarily stable, so that it's almost always better to work with accession numbers. Anyway, the distinction between sequence names and accession numbers is on a vanishing way because they tend more and more to be the same thing (as in the example just below). The use of the at sign @ to substitute as a wildcard character for any zero or more characters is possible here.

<sup>5</sup> i.e. what is documented in the ID or the LOCUS field

```

choosebank("emblTP")
query("Nexample", "N=AY382159")
getName(Nexample$req[[1]])

[1] "AY382159"

annotations <- getAnnot(Nexample$req[[1]])
cat(annotations, sep = "\n")

ID  AY382159    standard; genomic DNA; PRO; 783 BP.
XX
AC  AY382159;
XX
SV  AY382159.1
XX
DT  08-OCT-2003 (Rel. 77, Created)
DT  08-OCT-2003 (Rel. 77, Last updated, Version 1)
XX
DE  Borrelia burgdorferi strain FP1 OspA gene, partial cds.
XX
KW  .
XX
OS  Borrelia burgdorferi (Lyme disease spirochete)
OC  Bacteria; Spirochaetes; Spirochaetales; Spirochaetaceae; Borrelia;
OC  Borrelia burgdorferi group.
XX
RN  [1]
RP  1-783
RA  Hao Q., Wan K. ;
RT  ;
RL  Submitted (03-SEP-2003) to the EMBL/GenBank/DDBJ databases.
RL  Department of Lyme Spirochetosis, CDC, Beijing 102206, China
XX
FH  Key          Location/Qualifiers
FH
FT  source       1..783
FT  /db_xref="taxon:139"
FT  /mol_type="genomic DNA"
FT  /organism="Borrelia burgdorferi"
FT  /strain="FP1"
FT  <1..>783
FT  /codon_start=1
FT  /transl_table=11
FT  /product="OspA"
FT  /protein_id="AAQ89576.1"
FT  /translation="ALIACKQNVSSLDEKNSASVDPGEMKVLVSKEKDGDGYSLKAT
FT  VDKLELKGTSDKNNGSGTLEGEKTDKSKAKLTISDDLSTKTFEVFKEDGKTLVSRKVSS
FT  KDKTSTDEMPEKGELSAKTMRENGTKLEYTEMKSDGTGKTKEVLKNFTLEGRVANDK
FT  VTLEVKEGTVTLSKEIAKSGEVTVALNDNTTQATKKTGAWSKTSTLTISVNSKKTTQ
FT  LVFTKQDITVQKYDSAGTNLEGTAIVEIKTLDELKNALK"
XX
SQ  Sequence 783 BP; 342 A; 124 C; 145 G; 172 T; 0 other;

closebank()

```

**Y=year or Y>year or Y<year**

This is used to select sequences published in a given year (**Y=year**), or in a given year and after this year (**Y>year**), or in a given year and before this year (**Y<year**).

```

choosebank("emblTP")
query("Yexample", "Y=1999", virtual = TRUE)
Yexample$nelem

[1] 955274

closebank()

```

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### O=organelle

This is used to select sequences from specified organelle named following defined code (*e.g.*, chloroplast). The list of available organelle codes can be obtained from the `readsmj()` function this way:

```
choosebank("genbank")
nl <- readfirstrec(type = "SMJ")
smj <- readsmj(nl = nl, all.add = TRUE)
smj[!is.na(smj$nature) & smj$nature == "organelle", c("sname",
"libel")]

      sname          libel
3455  CHLOROPLAST    Chloroplast genome
3456  MITOCHONDRION  Mitochondrial genome
3457  NUCLEOMORPH    Nucleomorph genome
3458    PLASTID      non-green plastid genome

closebank()
```

To select for instance all sequences from chloroplast genome we can use:

```
choosebank("emblTP")
query("Oexample", "O=chloroplast", virtual = TRUE)
Oexample$nelem

[1] 65011

closebank()
```

### M=molecule

This is used to select sequences according to the chemical nature of the sequenced molecule<sup>6</sup>. The list of available organelle code can be obtained from the `readsmj()` function this way:

```
choosebank("genbank")
nl <- readfirstrec(type = "SMJ")
smj <- readsmj(nl = nl, all.add = TRUE)
smj[!is.na(smj$nature) & smj$nature == "molecule", c("sname",
"libel")]

      sname          libel
4    CRNA          <NA>
5    DNA   Sequenced molecule is DNA
6   MRNA  sequenced molecule is mRNA
7    RNA   Sequenced molecule is RNA
8   RRNA  sequenced molecule is rRNA
9  SCRNA sequenced molecule is small cytoplasmic RNA
10 SNORNA sequenced molecule is small nucleolar RNA
11 SNRNa sequenced molecule is small nuclear RNA
12  TRNA  sequenced molecule is tRNA

closebank()
```

To select for instance all sequences sequenced from DNA we can use:

```
choosebank("emblTP")
query("Mexample", "M=DNA", virtual = TRUE)
Mexample$nelem

[1] 7421752

closebank()
```

---

<sup>6</sup>as named in ID or LOCUS annotation records

**ST=status**

This is used to select sequences from specified data class (EMBL) or review level (UniProt). The list of status codes can be obtained from the `readsmj()` function this way:

```

choosebank("embl")
nl <- readfirstrec(type = "SMJ")
smj <- readsmj(nl = nl, all.add = TRUE)
smj[!is.na(smj$nature) & smj$nature == "status", c("sname",
  "libel")]

  sname          libel
1 EST      Expressed Sequence Tags data class
2 GSS      Genome Survey Sequence data class
3 HTC      High Throughput cDNA data class
4 HTG High Throughput Genome sequencing data class
5 PAT      Patent data class
6 STD      standard data class
7 STS      Sequence Tagged Site data class
8 TPA      Third Party Annotation data class

closebank()
choosebank("swissprot")
nl <- readfirstrec(type = "SMJ")
smj <- readsmj(nl = nl, all.add = TRUE)
smj[!is.na(smj$nature) & smj$nature == "status", c("sname",
  "libel")]

  sname          libel
1 REVIEWED Entry was reviewed and annotated by UniProtKB curators
2 UNREVIEWED Computer-annotated entry

closebank()

```

To select for instance all fully annotated sequences from Uniprot we can use:

```

choosebank("swissprot")
query("STexample", "ST=REVIEWED", virtual = TRUE)
STexample$nelem

```

[1] 257964

```
closebank()
```

**F=file\_name**

This is used to select sequences whose names are in a given file, one name per line. This is not directly implemented in seqinR, you have to use the function `crelistfromclientdata()` or its short form `clfc()` for this purpose. Here is an example with a file of sequence names distributed with the seqinR package:

```

choosebank("emblTP")
fileSQ <- system.file("sequences/bb.mne", package = "seqinr")
cat(readLines(fileSQ), sep = "\n")

```

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```

A04009.OSPA
A04009.OSPB
A22442
A24006
A24008
A24010
A24012
A24014
A24016
A33362
A67759.PE1
AB011063
AB011064
AB011065
AB011066
AB011067
AB035616
AB035617
AB035618
AB041949.VLSE

clfcfd("listSQ", file = fileSQ, type = "SQ")
sapply(listSQ$req, getName)

[1] "A04009.OSPA"    "A04009.OSPB"    "A22442"        "A24006"
[5] "A24008"         "A24010"        "A24012"        "A24014"
[9] "A24016"         "A33362"        "A67759.PE1"    "AB011063"
[13] "AB011064"       "AB011065"       "AB011066"       "AB011067"
[17] "AB035616"       "AB035617"       "AB035618"       "AB041949.VLSE"

closebank()

```

FA=file\_name

This is used to select sequences whose accession numbers are in a given file, one name per line. This is not directly implemented in seqinR, you have to use the function `crelistfromclientdata()` or its short form `clfcfd()` for this purpose. Here is an example with a file of sequence accession numbers distributed with the seqinR package:

```

choosebank("emb1TP")
fileAC <- system.file("sequences/bb.acc", package = "seqinr")
cat(readLines(fileAC), sep = "\n")

```

```

AY382159
AY382160
AY491412
AY498719
AY498720
AY498721
AY498722
AY498723
AY498724
AY498725
AY498726
AY498727
AY498728
AY498729
AY499181
AY500379
AY500380
AY500381
AY500382
AY500383

```

```

clfcfd("listAC", file = fileAC, type = "AC")
sapply(listAC$req, getName)

```

```
[1] "AY382159" "AY382160" "AY491412" "AY498719" "AY498720" "AY498721"
[7] "AY498722" "AY498723" "AY498724" "AY498725" "AY498726" "AY498727"
[13] "AY498728" "AY498729" "AY499181" "AY500379" "AY500380" "AY500381"
[19] "AY500382" "AY500383"
```

```
closebank()
```

**FK=file\_name**

This is used to produces the list of keywords named in given file, one keyword per line. This is not directly implemented in seqinR, you have to use the function `crelistfromclientdata()` or its short form `clfcfd()` for this purpose. Here is an example with a file of keywords distributed with the seqinR package:

```
choosebank("emblTP")
fileKW <- system.file("sequences/bb.kwd", package = "seqinr")
cat(readLines(fileKW), sep = "\n")

PLASMID
CIRCULAR
PARTIAL
5'-PARTIAL
3'-PARTIAL
MOTA GENE
MOTB GENE
DIVISION PRO
GYRB GENE
JOINING REGION
FTSA GENE
RPOB GENE
RPOC GENE
FLA GENE
DNAJ GENE
TUF GENE
PGK GENE
RUVA GENE
RUVB GENE
PROMOTER REGION

clfcfd("listKW", file = fileKW, type = "KW")
sapply(listKW$req, getName)

[1] "PLASMID"           "CIRCULAR"          "PARTIAL"            "5'-PARTIAL"
[5] "3'-PARTIAL"         "MOTA GENE"          "MOTB GENE"          "DIVISION PRO"
[9] "GYRB GENE"          "JOINING REGION"    "FTSA GENE"          "RPOB GENE"
[13] "RPOC GENE"          "FLA GENE"           "DNAJ GENE"          "TUF GENE"
[17] "PGK GENE"           "RUVA GENE"          "RUVB GENE"          "PROMOTER REGION"

closebank()
```

**FS=file\_name**

This is used to produces the list of species named in given file, one species per line. This is not directly implemented in seqinR, you have to use the function `crelistfromclientdata()` or its short form `clfcfd()` for this purpose. Here is an example with a file of species names distributed with the seqinR package:

```
choosebank("emblTP")
fileSP <- system.file("sequences/bb.sp", package = "seqinr")
cat(readLines(fileSP), sep = "\n")
```

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```
BORRELIA ANSERINA
BORRELIA CORIACEAE
BORRELIA PARKERI
BORRELIA TURICATAE
BORRELIA HERMSII
BORRELIA CROCIDURAE
BORRELIA LONESTARI
BORRELIA HISPANICA
BORRELIA BARBOURI
BORRELIA THEILERI
BORRELIA DUTTONII
BORRELIA MIYAMOTOI
BORRELIA PERSICA
BORRELIA RECURRENTIS
BORRELIA BURGDORFERI
BORRELIA AFZELII
BORRELIA GARINII
BORRELIA ANDERSONII
BORRELIA VALAISIANA
BORRELIA JAPONICA
```

```
clfcfd("listSP", file = fileSP, type = "SP")
sapply(listSP$req, getName)
```

```
[1] "BORRELIA ANSERINA"      "BORRELIA CORIACEAE"    "BORRELIA PARKERI"
[4] "BORRELIA TURICATAE"     "BORRELIA HERMSII"      "BORRELIA CROCIDURAE"
[7] "BORRELIA LONESTARI"      "BORRELIA HISPANICA"    "BORRELIA BARBOURI"
[10] "BORRELIA THEILERI"      "BORRELIA DUTTONII"     "BORRELIA MIYAMOTOI"
[13] "BORRELIA PERSICA"        "BORRELIA RECURRENTIS" "BORRELIA BURGDORFERI"
[16] "BORRELIA AFZELII"        "BORRELIA GARINII"      "BORRELIA ANDERSONII"
[19] "BORRELIA VALAISIANA"    "BORRELIA JAPONICA"
```

```
closebank()
```

```
list_name
```

A list name can be re-used, for instance:

```
choosebank("emblTP")
query("MyFirstListName", "Y=2000", virtual = TRUE)
MyFirstListName$nelem
```

```
[1] 885225
```

```
query("MySecondListName", "SP=Borrelia burgdorferi", virtual = TRUE)
MySecondListName$nelem
```

```
[1] 1682
```

```
query("MyThirdListName", "MyFirstListName AND MySecondListName",
      virtual = TRUE)
MyThirdListName$nelem
```

```
[1] 131
```

```
closebank()
```

### 3.2.4 Operators

AND

This is the binary operator for the logical and: a sequence belongs to the resulting list if, and only if, it is present in both operands. To select for instance sequences from *Borrelia burgdorferi* that are also coding sequences we can use:

```
choosebank("emblTP")
query("ANDexample", "SP=Borrelia burgdorferi AND T=CDS", virtual = TRUE)
ANDexample$nelem

[1] 3218

closebank()
```

OR

This is the binary operator for the logical or: a sequence belongs to the resulting list if it is present in at least one of the two operands. To select for instance sequences from *Borrelia burgdorferi* or from *Escherichia coli* we can use:

```
choosebank("emblTP")
query("ORexample", "SP=Borrelia burgdorferi OR SP=Escherichia coli",
      virtual = TRUE)
ORexample$nelem

[1] 28584

closebank()
```

NOT

This is the unary operator for the logical negation. To select for instance sequences from *Borrelia burgdorferi* that are not partial we can use:

```
choosebank("emblTP")
query("NOTexample", "SP=Borrelia burgdorferi AND NOT K=PARTIAL",
      virtual = TRUE)
NOTexample$nelem

[1] 3266

closebank()
```

PAR

This is a unary operator to compute the list of parent sequences of a list of sequences. The reciprocal operator is SUB. To check the reciprocity we can use for instance:

```
choosebank("emblTP")
query("A", "T=TRNA", virtual = TRUE)
query("B", "PAR A", virtual = TRUE)
query("C", "SUB B", virtual = TRUE)
query("D", "PAR C", virtual = TRUE)
query("emptySet", "B AND NOT D", virtual = TRUE)
emptySet$nelem

[1] 0

closebank()
```

**SUB**

This is a unary operator to add all subsequences of members of the single list operand.

```
choosebank("emblTP")
query("SUBexample", "AC=AE000783", virtual = T)
SUBexample$nelem
```

```
[1] 70
```

```
query("SUBexample2", "SUB SUBexample", virtual = T)
SUBexample2$nelem
```

```
[1] 943
```

```
closebank()
```

**PS**

This unary operator is used to get the list of species attached to member sequences of the operand list.

```
choosebank("emblTP")
query("PSexample", "K=hyperthermo@", virtual = T)
query("PSexample2", "PS PSexample")
sapply(PSexample2$req, getName)
```

```
[1] "BACILLUS LICHENIFORMIS" "DESULFUROCOCCUS"
[3] "PYROCOCCUS FURIOSUS"
```

```
closebank()
```

**PK**

This unary operator is used to get the list of keywords attached to member sequences of the operand list.

```
choosebank("emblTP")
query("PKexample", "AC=AE000783", virtual = T)
query("PKexample2", "PK PKexample")
sapply(PKexample2$req, getName)
```

```
[1] "DIVISION PRO" "CDS"           "RRNA"          "TRNA"
[5] "SOURCE"        "RELEASE 75"
```

```
closebank()
```

## UN

This unary operator is used to get the list of sequences attached to a list of species or keywords.

```
choosebank("emb1TP")
fileSP <- system.file("sequences/bb.sp", package = "seqinr")
cat(readLines(fileSP), sep = "\n")
```

```
BORRELIA ANSERINA
BORRELIA CORIACEAE
BORRELIA PARKERI
BORRELIA TURICATAE
BORRELIA HERMSII
BORRELIA CROCIDURAE
BORRELIA LONESTARI
BORRELIA HISPANICA
BORRELIA BARBOURI
BORRELIA THEILERI
BORRELIA DUTTONII
BORRELIA MIYAMOTOI
BORRELIA PERSICA
BORRELIA RECURRENTIS
BORRELIA BURGDORFERI
BORRELIA AFZELII
BORRELIA GARINII
BORRELIA ANDERSONII
BORRELIA VALAISIANA
BORRELIA JAPONICA
```

```
clfc("listSP", file = fileSP, type = "SP")
query("UNexample", "UN listSP", virtual = TRUE)
UNexample$nelem
```

```
[1] 2786
```

```
closebank()
```

## SD

This unary operator computes the list of species placed in the tree below the members of the species list operand.

```
choosebank("emb1TP")
query("hominidae", "SP=Hominidae", virtual = T)
query("hsp", "PS hominidae", virtual = T)
hsp$nelem
```

```
[1] 19
```

```
query("SDexample", "SD hsp")
sapply(SDexample$req, getName)
```

[1]	"HOMINIDAE"	"PONGO"
[3]	"PONGO PYGMAEUS"	"PONGO PYGMAEUS ABELII"
[5]	"PONGO PYGMAEUS PYGMAEUS"	"PONGO SP."
[7]	"HOMO/PAN/GORILLA GROUP"	"GORILLA"
[9]	"GORILLA GORILLA"	"GORILLA GORILLA BERINGEI"
[11]	"GORILLA GORILLA GRAUERI"	"GORILLA GORILLA GORILLA"
[13]	"GORILLA GORILLA UELLENSIS"	"PAN"
[15]	"PAN TROGLODYTES"	"PAN TROGLODYTES SCHWEINFURTHII"
[17]	"PAN TROGLODYTES TROGLODYTES"	"PAN TROGLODYTES VERUS"
[19]	"PAN TROGLODYTES VELLEROSUS"	"PAN PANISCUS"
[21]	"HOMO"	"HOMO SAPIENS"
[23]	"HOMO SAPIENS NEANDERTHALensis"	

```
closebank()
```

**KD**

This unary operator computes the list of keywords placed in the tree below the members of the keywords list operand.

```
choosebank("emblTP")
query("cat", "SP=Felis catus", virtual = TRUE)
query("catkw", "PK cat", virtual = TRUE)
catkw$nelem
```

```
[1] 540
```

```
query("KDexample", "KD catkw", virtual = TRUE)
KDexample$nelem
```

```
[1] 572
```

```
closebank()
```

### 3.3 Session Informations

This part was compiled under the following  environment:

- R version 2.4.1 (2006-12-18), i386-apple-darwin8.8.1
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: MASS 7.2-30, ade4 1.4-2, ape 1.9-3, gee 4.13-12, lattice 0.14-16, nlme 3.1-78, seqinr 1.0-7, xtable 1.4-3

---

---

# CHAPTER 4

---

## How to deal with sequences

Charif, D. Lobry, J.R.

### Contents

---

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<b>4.4</b>	<b>Session Informations</b>	<b>69</b>

---

### 4.1 Sequence classes

There are currently three classes of sequences, depending on the way they were obtained:

- **seqFasta** is the class for the sequences that were imported from a fasta file
- **seqAcnucWeb** is the class for the sequences coming from an ACNUC database server
- **seqFrag** is the class for the sequences that are fragments of other sequences

### 4.2 Generic methods for sequences

All sequence classes are sharing a common interface, so that there are very few method names we have to remember. In addition, all classes have their specific `as.ClassName` method that return an instance of the class, and `is.ClassName` method to check whether an object belongs or not to the class. Available methods are:

Methods	Result	Type of result
<code>getFrag</code>	a sequence fragment	a sequence fragment
<code>getSequence</code>	the sequence	vector of characters
<code>getName</code>	the name of a sequence	string
<code>getLength</code>	the length of a sequence	numeric vector
<code>getTrans</code>	translation into amino-acids	vector of characters
<code>getAnnot</code>	sequence annotations	vector of string
<code>getLocation</code>	position of a Sequence on its parent sequence	list of numeric vector

## 4.3 Internal representation of sequences

The current mode of sequence storage is done with vectors of characters instead of strings. This is very convenient for the user because all  tools to manipulate vectors are immediately available. The price to pay is that this storage mode is extremely expensive in terms of memory. There are two utilities called `s2c()` and `c2s()` that allows to convert strings into vector of characters, and *vice versa*, respectively.

### 4.3.1 Sequences as vectors of characters

In the vectorial representation mode, all the very convenient  tools for indexing vectors are at hand.

1. Vectors can be indexed by a vector of *positive* integers saying which elements are to be selected. As we have already seen, the first 50 elements of a sequence are easily extracted thanks to the binary operator `from:to`, as in:

```
dnafile <- system.file("sequences/malM.fasta", package = "seqinr")
myseq <- read.fasta(file = dnafile)[[1]]
1:50
```

```
[1] 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24
[25] 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48
[49] 49 50
```

```
myseq[1:50]
```

```
[1] "a" "t" "g" "a" "a" "a" "a" "t" "g" "a" "a" "t" "a" "a" "a" "a" "g" "t"
[19] "c" "t" "c" "a" "t" "c" "g" "t" "c" "t" "c" "t" "g" "t" "t" "t" "a"
[37] "t" "c" "a" "g" "c" "a" "g" "g" "g" "t" "t" "a" "c" "t"
```

The `seq()` function allows to build more complexe integer vectors. For instance in coding sequences it is very common to focus on third codon positions where selection is weak. Let's extract bases from third codon positions:

```
tcp <- seq(from = 3, to = length(myseq), by = 3)
tcp[1:10]
```

```
[1] 3 6 9 12 15 18 21 24 27 30
```

```
myseqtcp <- myseq[tcp]
myseqtcp[1:10]

[1] "g" "a" "g" "t" "a" "t" "c" "c" "c" "c"
```

2. Vectors can also be indexed by a vector of *negative* integers saying which elements have to be removed. For instance, if we want to keep first and second codon positions, the easiest way is to remove third codon positions:

```
-tcp[1:10]

[1] -3 -6 -9 -12 -15 -18 -21 -24 -27 -30

myseqfscp <- myseq[-tcp]
myseqfscp[1:10]

[1] "a" "t" "a" "a" "a" "t" "a" "a" "a" "a"
```

3. Vectors are also indexable by a vector of *logicals* whose TRUE values say which elements to keep. Here is a different way to extract all third coding positions from our sequence. First, we define a vector of three logicals with only the last one true:

```
ind <- c(F, F, T)
ind

[1] FALSE FALSE TRUE
```

This vector seems too short for our purpose because our sequence is much more longer with its 921 bases. But under  $\mathbb{R}$  vectors are automatically *recycled* when they are not long enough:

```
(1:30)[ind]

[1] 3 6 9 12 15 18 21 24 27 30

myseqtcp2 <- myseq[ind]
```

The result should be the same as previously:

```
identical(myseqtcp, myseqtcp2)

[1] TRUE
```

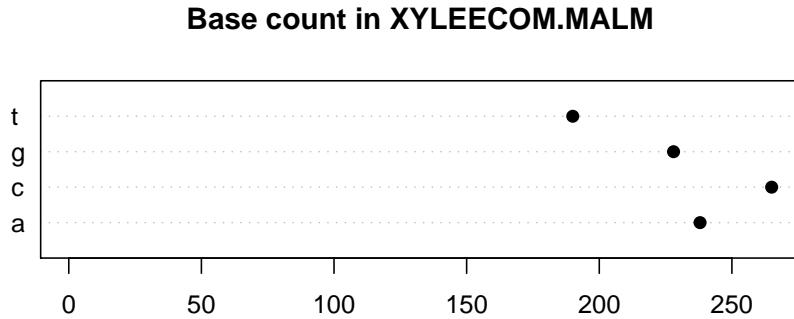
This recycling rule is extremely convenient in practice but may have surprising effects if you assume (incorrectly) that there is a stringent dimension control for  $\mathbb{R}$  vectors as in linear algebra.

Another advantage of working with vector of characters is that most  $\mathbb{R}$  functions are vectorized so that many things can be done without explicit looping. Let's give some very simple examples:

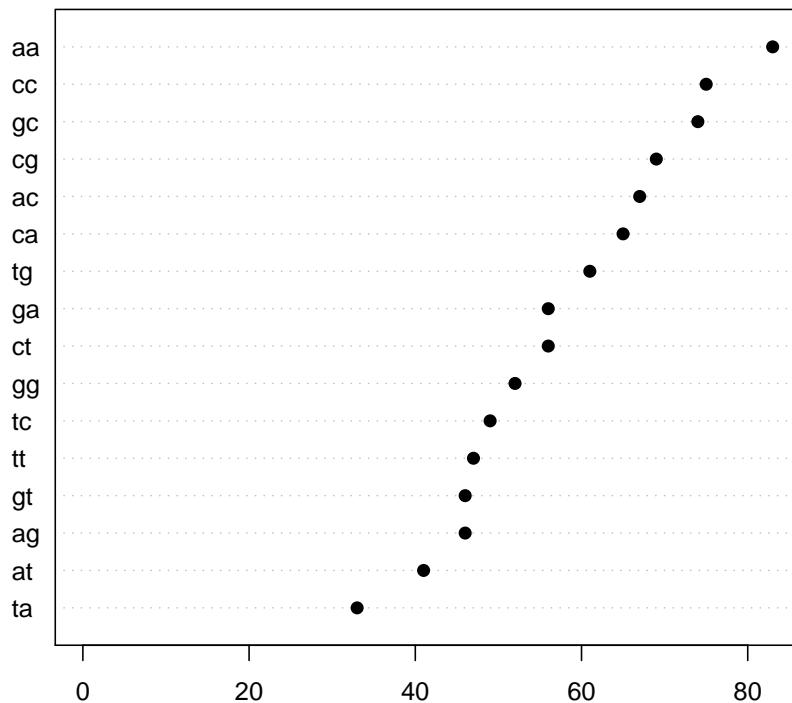
```
tota <- sum(myseq == "a")
```

The total number of a in our sequence is 238. Let's compare graphically the different base counts in our sequence :

```
basecount <- table(myseq)
myseqname <- getName(myseq)
dotchart(basecount, xlim = c(0, max(basecount)), pch = 19,
         main = paste("Base count in", myseqname))
```

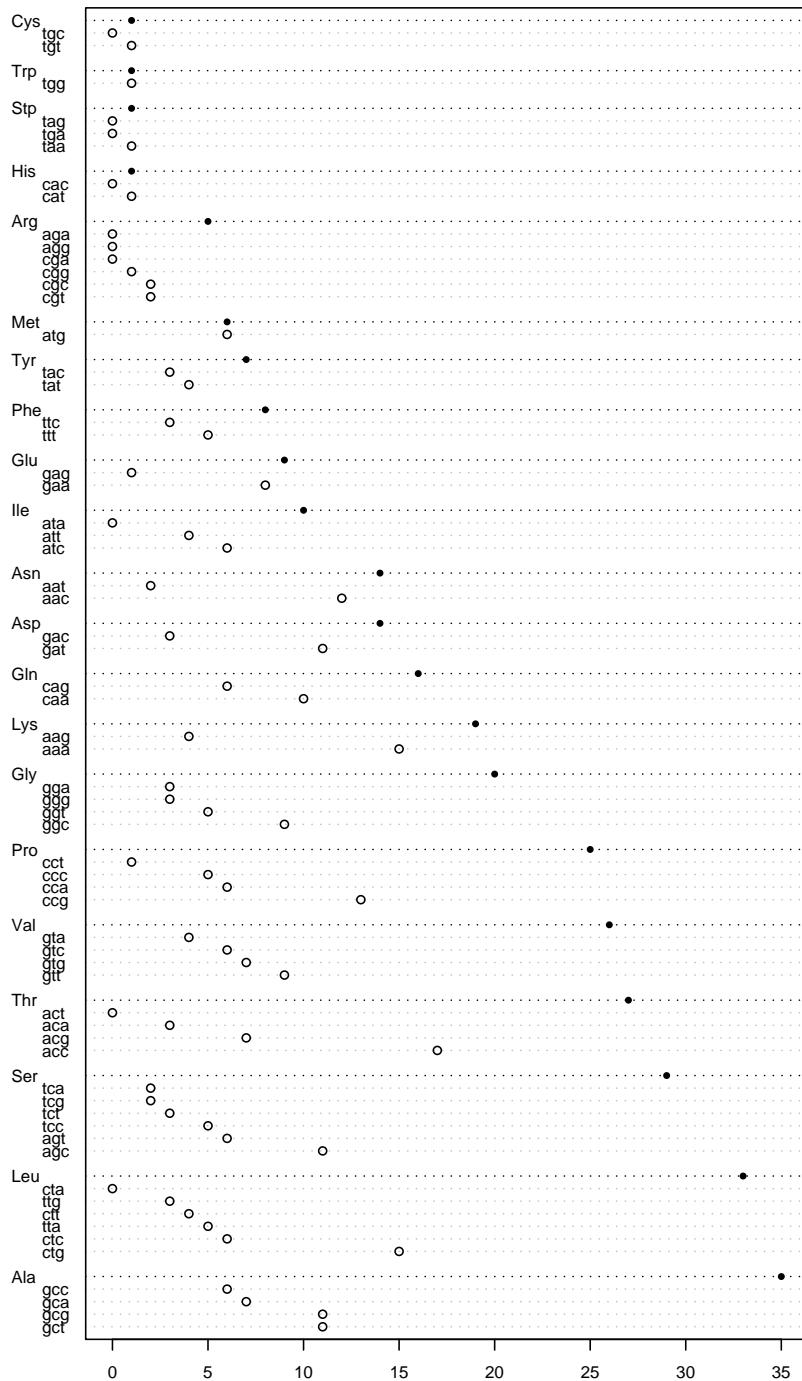


```
dinuclcount <- count(myseq, 2)
dotchart(dinuclcount[order(dinuclcount)], xlim = c(0, max(dinuclcount)),
          pch = 19, main = paste("Dinucleotide count in", myseqname))
```

**Dinucleotide count in XYLEECOM.MALM**

```
codonusage <- uco(myseq)
dotchart.uco(codonusage, main = paste("Codon usage in", myseqname))
```

Codon usage in XYLEECOM.MALM



### 4.3.2 Sequences as strings

If you are interested in (fuzzy) pattern matching, then it is advisable to work with sequence as strings to take advantage of *regular expression* implemented in . The function `words.pos()` returns the positions of all occurrences of a given regular expression. Let's suppose we want to know where are the trinucleotides "cgt" in a sequence, that is the fragment CpGpT in the direct strand:

```
mystring <- c2s(myseq)
words.pos("cgt", mystring)

[1] 24 90 216 245 252 315 330 405 432 452 552 592 648 836 883
```

We can also look for the fragment CpGpTpY to illustrate fuzzy matching because Y (IUPAC code for pyrimidine) stands C or T:

```
words.pos("cgt[ct]", mystring)

[1] 24 216 252 315 432 452 552 592 836 883
```

To look for all CpC dinucleotides separated by 3 or 4 bases:

```
words.pos("cc.{3,4}cc", mystring)

[1] 72 119 176 177 539 577 578 638 677 682 730 731 736 881 882
```

Virtually any pattern is easily encoded with a regular expression. This is especially useful at the protein level because many functions can be attributed to short linear motifs.

## 4.4 Session Informations

This part was compiled under the following  environment:

- R version 2.4.1 (2006-12-18), i386-apple-darwin8.8.1
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: MASS 7.2-30, ade4 1.4-2, ape 1.9-3, gee 4.13-12, lattice 0.14-16, nlme 3.1-78, seqinr 1.0-7, xtable 1.4-3



---

---

# CHAPTER 5

---

## Multivariate analyses

Lobry, J.R.

### Contents

---

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---

### 5.1 Correspondence analysis

This is the most popular multivariate data analysis technique for amino-acid and codon count tables, its application, however, is not without pitfalls [49]. Its primary goal is to transform a table of counts into a graphical display, in which each gene (or protein) and each codon (or amino-acid) is depicted as a point. Correspondence analysis (CA) may be defined as a special case of principal components analysis (PCA) with a different underlying metrics. The interest of the metrics in CA, that is the way we measure the distance between two individuals, is illustrated bellow with a very simple example (Table 5.1 inspired from [12]) with only three proteins having only three amino-acids, so that we can represent exactly on a map the consequences of the metric choice.

```
data(toyaa)
toyaa
```

	Ala	Val	Cys
1	130	70	0
2	60	40	0
3	60	35	5

Let's first use the regular Euclidian metrics between two proteins  $i$  and  $i'$ ,

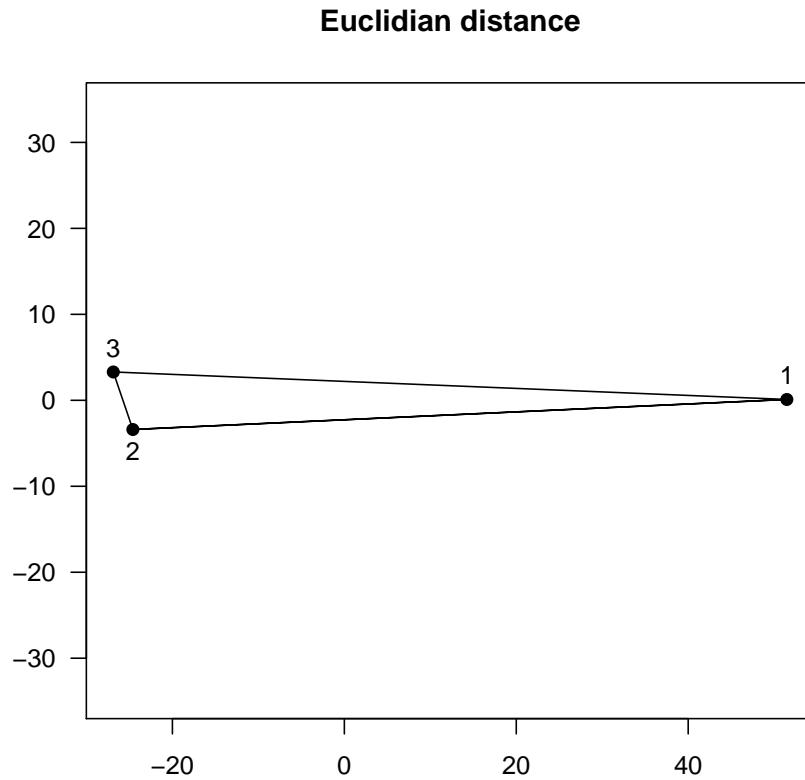
$$d^2(i, i') = \sum_{j=1}^J (n_{ij} - n_{i'j})^2 \quad (5.1)$$

	Ala	Val	Cys
1	130	70	0
2	60	40	0
3	60	35	5

Table 5.1: A very simple example of amino-acid counts in three proteins to be loaded with `data(toyaa)`.

to visualize this small data set:

```
library(ade4)
pco <- dudi.pco(dist(toyaa), scann = F, nf = 2)
myplot <- function(res, ...) {
  plot(res$li[, 1], res$li[, 2], ...)
  text(x = res$li[, 1], y = res$li[, 2], labels = 1:3, pos = ifelse(res$li[, 2] < 0, 1, 3))
  perm <- c(3, 1, 2)
  lines(c(res$li[, 1], res$li[perm, 1]), c(res$li[, 2], res$li[perm, 2]))
}
myplot(pco, main = "Euclidian distance", asp = 1, pch = 19,
       xlab = "", ylab = "", las = 1)
```



From this point of view, the first individual is far away from the two others. But thinking about it, this is a rather trivial effect of protein size:

```
rowSums(toyaa)
```

1	2	3
200	100	100

With 200 amino-acids, the first protein is two times bigger than the others so that when computing the Euclidian distance (5.1) its  $n_{ij}$  entries are on average bigger, sending it away from the others. To get rid of this trivial effect, the first obvious idea is to divide counts by protein lengths so as to work with *protein profiles*. The corresponding distance is,

$$d^2(i, i') = \sum_{j=1}^J \left( \frac{n_{ij}}{n_{i\bullet}} - \frac{n_{i'j}}{n_{i'\bullet}} \right)^2 \quad (5.2)$$

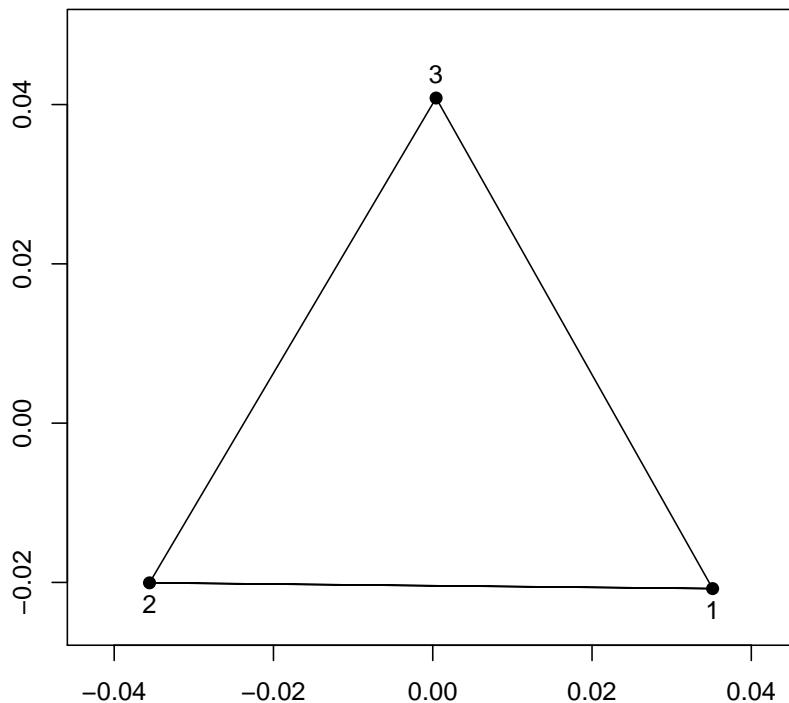
where  $n_{i\bullet}$  and  $n_{i'\bullet}$  are the total number of amino-acids in protein  $i$  and  $i'$ , respectively.

```
profile <- toyaa/rowSums(toyaa)
profile
```

	Ala	Val	Cys
1	0.65	0.35	0.00
2	0.60	0.40	0.00
3	0.60	0.35	0.05

```
pco1 <- dudi.pco(dist(profile), scann = F, nf = 2)
myplot(pco1, main = "Euclidian distance on protein profiles",
       asp = 1, pch = 19, xlab = "", ylab = "", ylim = range(pco1$li[, 2]) * 1.2)
```

### Euclidian distance on protein profiles

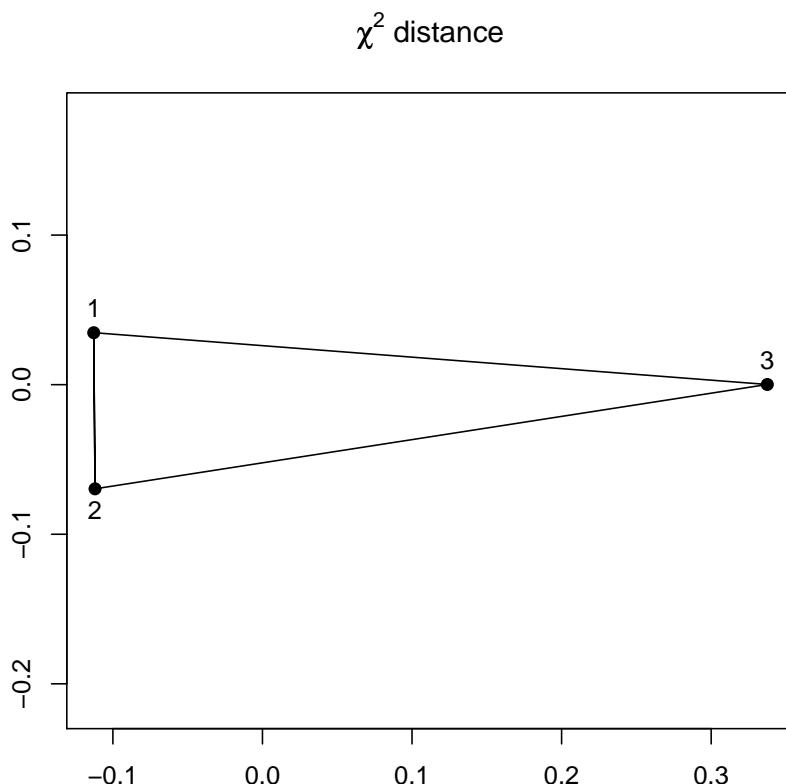


The pattern is now completely different with the three protein equally spaced. This is normal because in terms of relative amino-acid composition they are all differing two-by-two by 5% at the level of two amino-acids only. We have clearly removed the trivial protein size effect, but this is still not completely satisfactory. The proteins are differing by 5% for all amino-acids but the situation is somewhat different for Cys because this amino-acid is very rare. A difference of 5% for a rare amino-acid has not the same significance than a difference of 5% for a common amino-acid such as Ala in our example. To cope with this, CA make use of a variance-standardizing technique to compensate for the larger variance in high frequencies and the smaller variance in low frequencies. This is achieved with the use of the *chi-square distance* ( $\chi^2$ ) which differs from the previous Euclidean distance on profiles (5.2) in that each square is weighted by the inverse of the frequency corresponding to each term,

$$d^2(i, i') = n_{\bullet\bullet} \sum_{j=1}^J \frac{1}{n_{\bullet j}} \left( \frac{n_{ij}}{n_{i\bullet}} - \frac{n_{i'j}}{n_{i'\bullet}} \right)^2 \quad (5.3)$$

where  $n_{\bullet j}$  is the total number of amino-acid of kind  $j$  and  $n_{\bullet\bullet}$  the total number of amino-acids. With this point of view, the map is now like this:

```
coa <- dudi.coa(toyaa, scann = FALSE, nf = 2)
myplot(coa, main = expression(paste(chi^2, " distance")),
asp = 1, pch = 19, xlab = "", ylab = "")
```

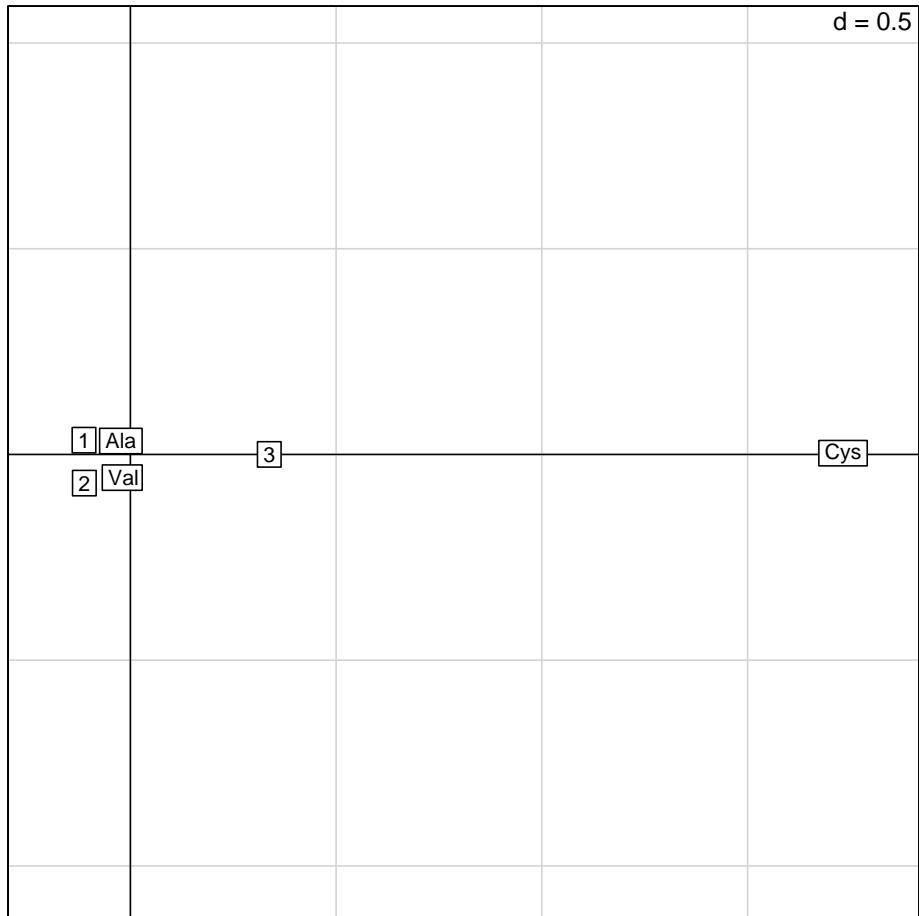


The pattern is completely different with now protein number 3 which is far away from the others because it is enriched in the rare amino-acid Cys as compared to others.

The purpose of this small example was to demonstrates that the metric choice is not without dramatic effects on the visualisation of data. Depending on your objectives, you may agree or disagree with the  $\chi^2$  metric choice, that's not a problem, the important point is that you should be aware that there is an underlying model there, *chacun a son goût ou chacun à son goût*, it's up to you.

Now, if you agree with the  $\chi^2$  metric choice, there's a nice representation that may help you for the interpretation of results. This is a kind of "biplot" representation in which the lines and columns of the dataset are simultaneously represented, in the right way, that is as a graphical *translation* of a mathematical theorem, but let's see how does it look like in practice:

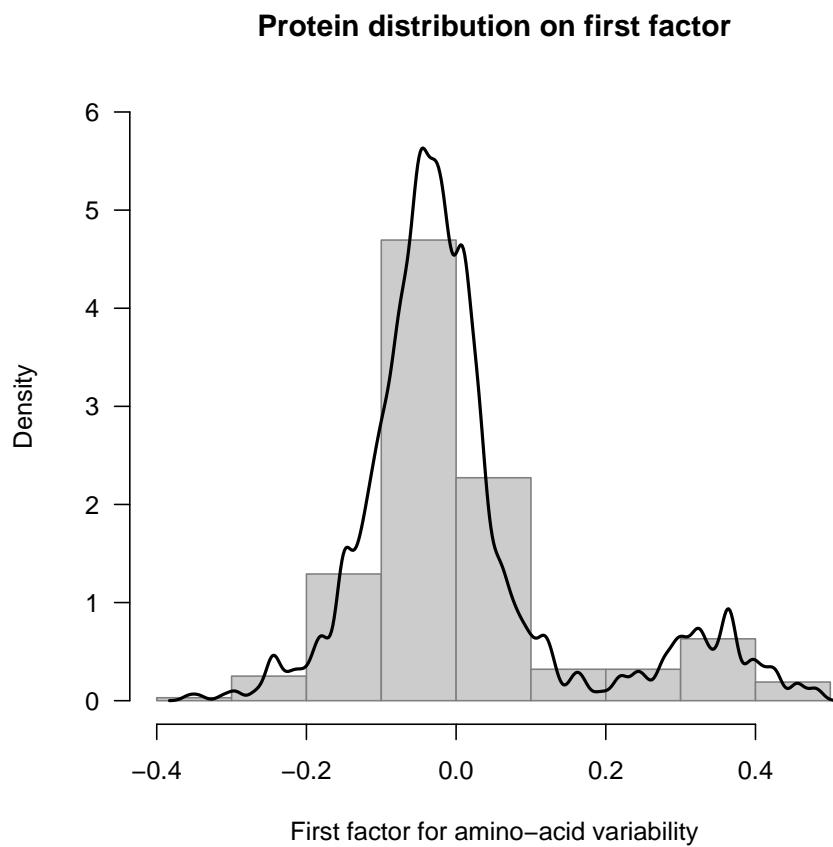
```
scatter(coa, clab.col = 0.8, clab.row = 0.8, posi = "none")
```



What is obvious is that the Cys content has a major effect on protein variability here, no scoop. Please note how the information is well summarised here: protein number 3 differs because it's enriched in Cys ; protein number 1 and 2 are almost the same but there is a small trend protein number 1 to be enriched in Ala. As compared to table 5.1 this graph is of poor information here, so let's try a more big-room-sized example (with 20 columns so as to illustrate the dimension reduction technique).

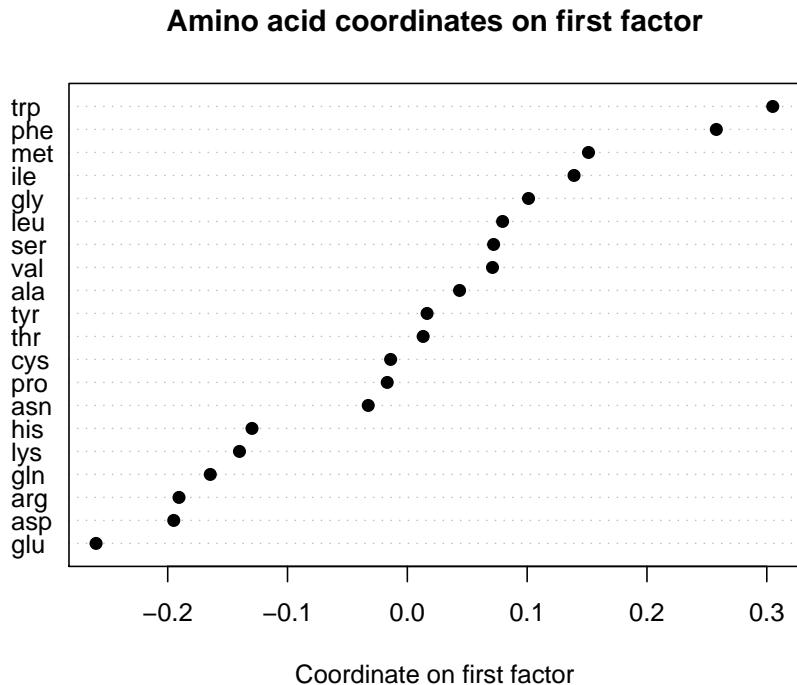
Data are from [39], a sample of the proteome of *Escherichia coli*. According to the title of this paper, the most important factor for the between-protein variability is hydrophilic - hydrophobic gradient. Let's try to reproduce this assertion :

```
download.file(url = "ftp://pbil.univ-lyon1.fr/pub/datasets/NAR94/data.txt",
             destfile = "data.txt")
ec <- read.table(file = "data.txt", header = TRUE, row.names = 1)
ec.coa <- dudi.coa(ec, scann = FALSE, nf = 1)
F1 <- ec.coa$li[, 1]
hist(F1, proba = TRUE, xlab = "First factor for amino-acid variability",
      col = grey(0.8), border = grey(0.5), las = 1, ylim = c(0,
      6), main = "Protein distribution on first factor")
lines(density(F1, adjust = 0.5), lwd = 2)
```



There is clearly a bimodal distribution of proteins on the first factor. What are the the amino-acid coordinates on this factor?

```
aacoo <- ec.co$co[, 1]
names(aacoo) <- rownames(ec.co$co)
aacoo <- sort(aacoo)
dotchart(aacoo, pch = 19, xlab = "Coordinate on first factor",
         main = "Amino acid coordinates on first factor")
```



Aliphatic and aromatic amino-acids have positive values while charged amino-acids have negative values<sup>1</sup>. Let's try to compute the GRAVY score (*i.e.* the Kyte and Doolittle hydropathic index[33]) of our proteins to compare this with their coordinates on the first factor. We need first the amino-acid *relatives* frequencies in the proteins, for this we divide the all the amino-acid counts by the total by row:

```
ecfr <- ec/rowSums(ec)
ecfr[1:5, 1:5]
```

```
      arg      leu      ser      thr      pro
FOLE 0.05829596 0.10313901 0.06278027 0.08520179 0.03587444
MSBA 0.06529210 0.10309278 0.08591065 0.06185567 0.02233677
NARV 0.06637168 0.12831858 0.06637168 0.05752212 0.03539823
NARW 0.05627706 0.16450216 0.05627706 0.03030303 0.04329004
NARY 0.06614786 0.06420233 0.05058366 0.03891051 0.06031128
```

We need also the coefficients corresponding to the GRAVY score:

```
gravy <- read.table(file = "ftp://pbil.univ-lyon1.fr/pub/datasets/NAR94/gravy.txt")
gravy[1:5, ]
```

```
      V1     V2
1 Ala  1.8
2 Arg -4.5
3 Asn -3.5
4 Asp -3.5
5 Cys  2.5
```

---

<sup>1</sup>The physico-chemical classes for amino acids are given in the component AA.PROPERTY of the SEQINR.UTIL object.

```
coef <- gravy$V2
```

The coefficient are given in the alphabetical order of the three letter code for the amino acids, that is in a different order than in the object **ecfr**:

```
names(ecfr)
```

```
[1] "arg" "leu" "ser" "thr" "pro" "ala" "gly" "val" "lys" "asn" "gln" "his"
[13] "glu" "asp" "tyr" "cys" "phe" "ile" "met" "trp"
```

We then re-order the columns of the data set and check that everthing is OK:

```
ecfr <- ecfr[, order(names(ecfr))]
ecfr[1:5, 1:5]
```

```
      ala      arg      asn      asp      cys
FOLE 0.08520179 0.05829596 0.04035874 0.05381166 0.008968610
MSBA 0.08247423 0.06529210 0.03608247 0.05154639 0.003436426
NARV 0.05309735 0.06637168 0.01769912 0.02212389 0.013274336
NARW 0.09090909 0.05627706 0.02597403 0.09090909 0.017316017
NARY 0.06225681 0.06614786 0.03891051 0.05642023 0.035019455
```

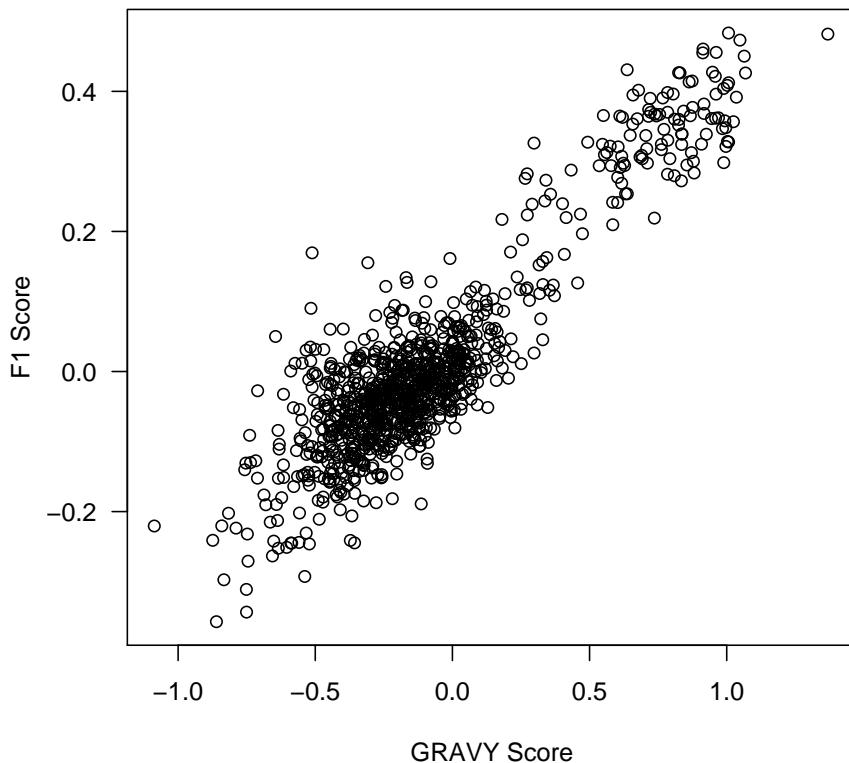
```
all(names(ecfr) == tolower(as.character(gravy$V1)))
```

```
[1] TRUE
```

Now, thanks to R build-in matrix multiplication, it's only one line to compute the GRAVY score:

```
gscores <- as.matrix(ecfr) %*% coef
plot(gscores, F1, xlab = "GRAVY Score", ylab = "F1 Score",
     las = 1, main = "The first factor is protein hydrophaty")
```

### The first factor is protein hydrophathy



The proteins with high GRAVY scores are integral membrane proteins, and those with low scores are cytoplasmic proteins. Now, suppose that we want to adjust a mixture of two normal distributions to get an estimate of the proportion of cytoplasmic and integral membrane proteins. We first have a look on the predefined distributions (Table 5.2), but there is apparently not an out of the box solution. We then define our own probability density function and then use `fitdistr` from package MASS to get a maximum likelihood estimate of the parameters:

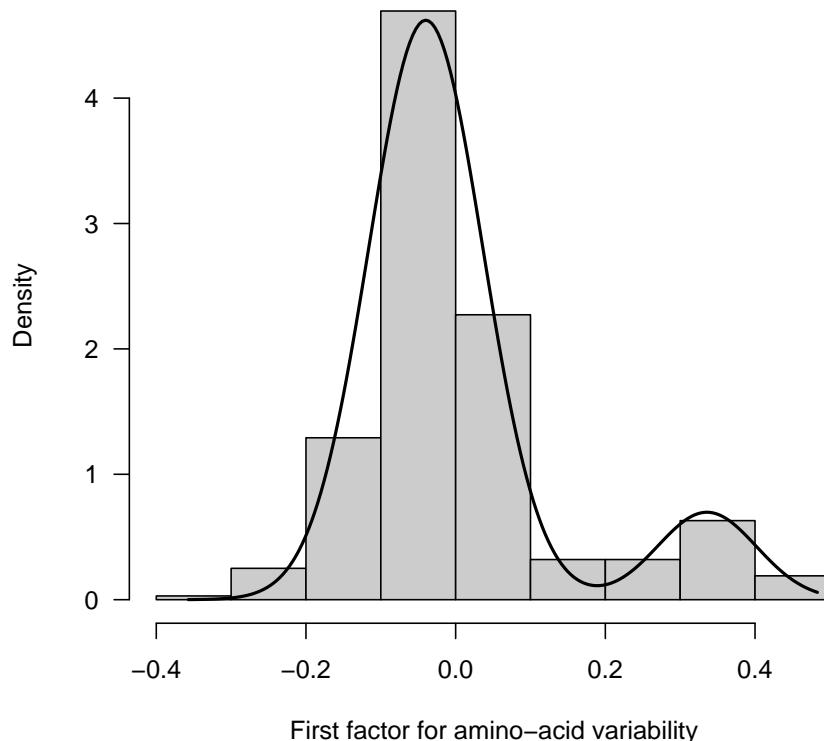
```
dmixnor <- function(x, p, m1, sd1, m2, sd2) {
  p * dnorm(x, m1, sd1) + (1 - p) * dnorm(x, m2, sd2)
}
library(MASS)
e <- fitdistr(F1, dmixnor, list(p = 0.88, m1 = -0.04, sd1 = 0.076,
  m2 = 0.34, sd2 = 0.07))$estimate
e

  p      m1      sd1      m2      sd2
0.88405009 -0.03989489  0.07632235  0.33579162  0.06632259

hist(F1, proba = TRUE, col = grey(0.8), main = "Ajustement with a mixture of two normal distributions",
  xlab = "First factor for amino-acid variability", las = 1)
xx <- seq(from = min(F1), to = max(F1), length = 200)
lines(xx, dmixnor(xx, e[1], e[2], e[3], e[4], e[5]), lwd = 2)
```

	d	p	q	r
beta	dbeta	pbeta	qbeta	rbeta
binom	dbinom	pbinom	qbinom	rbinom
cauchy	dcauchy	pcauchy	qcauchy	rcauchy
chisq	dchisq	pchisq	qchisq	rchisq
exp	dexp	pexp	qexp	rexp
f	df	pf	qf	rf
gamma	dgamma	pgamma	qgamma	rgamma
geom	dgeom	pgeom	qgeom	rgeom
hyper	dhyper	phyper	qhyper	rhyper
lnorm	dlnorm	plnorm	qlnorm	rlnorm
logis	dlogis	plogis	qlogis	rlogis
nbinom	dnbinom	pnbinom	qnbinom	rnbnom
norm	dnorm	pnorm	qnorm	rnorm
pois	dpois	ppois	qpois	rpois
signrank	dsignrank	psignrank	qsignrank	rsignrank
t	dt	pt	qt	rt
unif	dunif	punif	qunif	runif
weibull	dweibull	pweibull	qweibull	rweibull
wilcox	dwilcox	pwilcox	qwilcox	rwilcox

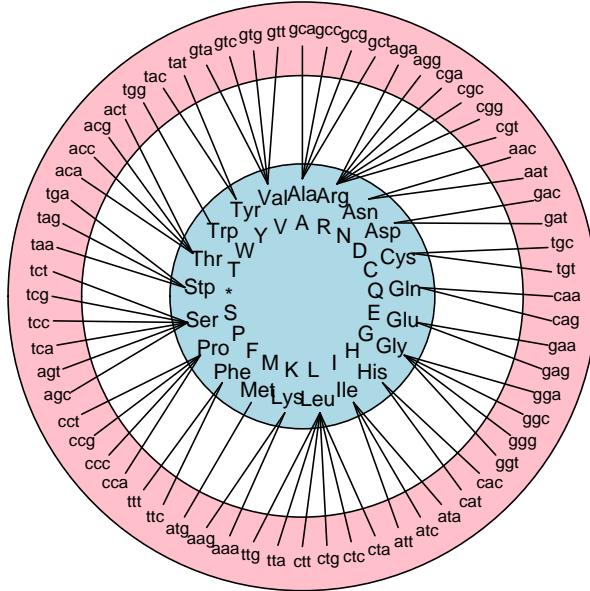
Table 5.2: Density, distribution function, quantile function and random generation for the predefined distributions under R

**Ajustement with a mixture of two normal distributions**

## 5.2 Synonymous and non-synonymous analyses

Genetic codes are surjective applications from the set codons ( $n = 64$ ) into the set of amino-acids ( $n = 20$ ) :

**The surjective nature of genetic codes**  
**Genetic code number 1**



Adapted from insert 2 in Lobry & Chessel (2003) JAG 44:235

Two codons encoding the same amino-acid are said synonymous while two codons encoding a different amino-acid are said non-synonymous. The distinction between the synonymous and non-synonymous level are very important in evolutionary studies because most of the selective pressure is expected to work at the non-synonymous level, because the amino-acids are the components of the proteins, and therefore more likely to be subject to selection.

$K_s$  and  $K_a$  are an estimation of the number of substitutions per synonymous site and per non-synonymous site, respectively, between two protein-coding genes [36]. The  $\frac{K_a}{K_s}$  ratio is used as tool to evaluate selective pressure (see [24] for a nice back to basics). Let's give a simple illustration with three orthologous genes of the thioredoxin family from *Homo sapiens*, *Mus musculus*, and *Rattus norvegicus* species:

```
ortho <- read.alignment(system.file("sequences/ortho.fasta",
  package = "seqinr"), format = "fasta")
kaks.ortho <- kaks(ortho)
kaks.ortho$ka/kaks.ortho$ks
```

HSU78678.PE1	AK002358.PE1	HSU78678.PE1
	0.1243472	
RNU73525.PE1		0.1356036

The  $\frac{K_a}{K_s}$  ratios are less than 1, suggesting a selective pressure on those proteins during evolution.

For transversal studies (*i.e.* codon usage studies in a genome at the time it was sequenced) there is little doubt that the strong requirement to distinguish between synonymous and non-synonymous variability was the source of many mistakes [49]. We have just shown here with a scholarship example that the metric choice is not neutral. If you consider that the  $\chi^2$  metric is not too bad, with respect to your objectives, and that you want to quantify the synonymous and non-synonymous variability, please consider reading this paper [38], and follow this link <http://pbil.univ-lyon1.fr/members/lobry/repro/jag03/> for on-line reproducibility.

Let's now use the toy example given in table 5.3 to illustrate how to study synonymous and non-synonymous codon usage.

```
data(toycodon)
toycodon
```

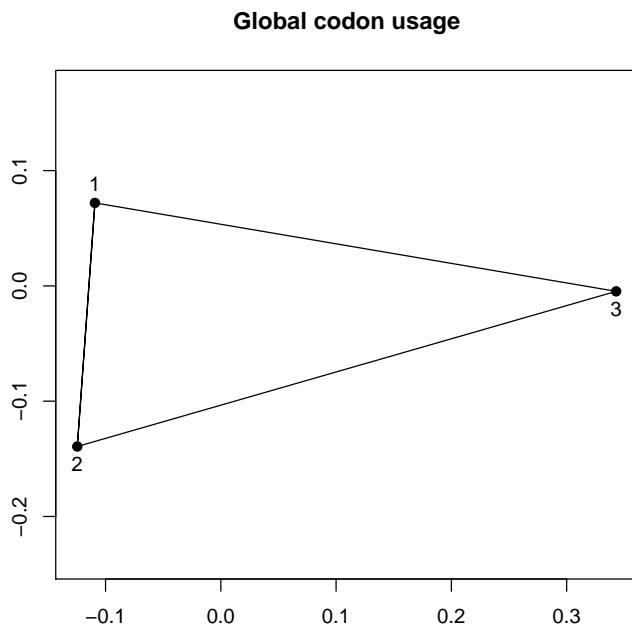
	gca	gcc	gct	gtc	gtg	gtt	tgt	tgc		
1	33	32	32	33	18	17	17	18	0	0
2	13	17	17	13	8	12	12	8	0	0
3	16	14	14	16	8	9	10	8	3	2

	gca	gcc	gct	gtc	gtg	gtt	tgt	tgc		
1	33	32	32	33	18	17	17	18	0	0
2	13	17	17	13	8	12	12	8	0	0
3	16	14	14	16	8	9	10	8	3	2

Table 5.3: A very simple example of codon counts in three coding sequences to be loaded with `data(toycodon)`.

Let's first have a look to global codon usage, we do not take into account the structure of the genetic code:

```
global <- dudi.coa(toycodon, scann = FALSE, nf = 2)
myplot(global, asp = 1, pch = 19, xlab = "", ylab = "", main = "Global codon usage")
```



From a global codon usage point of view, coding sequence number 3 is away. To take into account the genetic code structure, we need to know for which amino-acid the codons are coding. The codons are given by the names of the columns of the object `toycodon`:

```
names(toycodon)
```

```
[1] "gca" "gcc" "gcg" "gct" "gta" "gtc" "gtg" "gtt" "tgt" "tgc"
```

Put all codon names into a single string:

```
c2s(names(toycodon))
```

```
[1] "gcagccgcggctgttagtcgtggttttgc"
```

Transform this string as a vector of characters:

```
s2c(c2s(names(toycodon)))
```

```
[1] "g" "c" "a" "g" "c" "c" "g" "c" "g" "g" "c" "t" "g" "t" "a" "g" "t" "c"
[19] "g" "t" "g" "g" "t" "t" "g" "t" "g" "t" "g" "c"
```

Translate this into amino-acids using the default genetic code:

```
translate(s2c(c2s(names(toycodon))))
```

```
[1] "A" "A" "A" "A" "V" "V" "V" "C" "C"
```

Use the three letter code for amino-acid instead:

```

aaa(translate(s2c(c2s(names(toycodon)))))

[1] "Ala" "Ala" "Ala" "Ala" "Val" "Val" "Val" "Val" "Cys" "Cys"

```

Make this a factor:

```

facaa <- factor(aaa(translate(s2c(c2s(names(toycodon))))))

facaa

```

```

[1] Ala Ala Ala Ala Val Val Val Val Cys Cys
Levels: Ala Cys Val

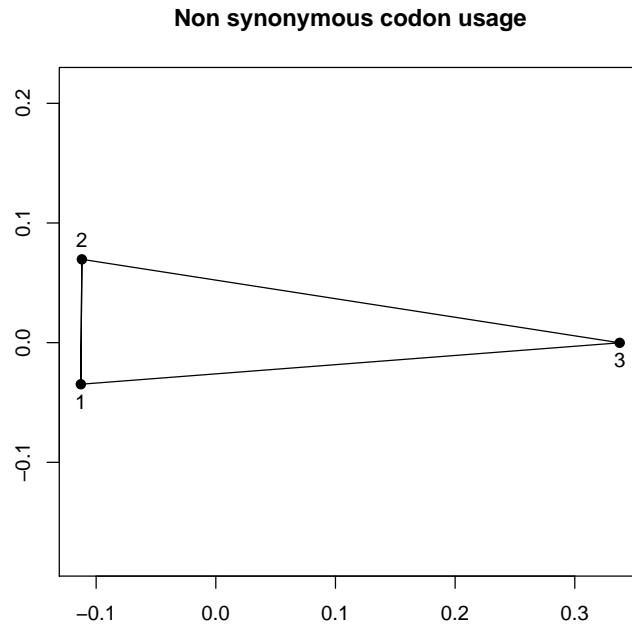
```

The non synonymous codon usage analysis is the between amino-acid analysis:

```

nonsynonymous <- t(between(dudi = t(global), fac = facaa,
  scann = FALSE, nf = 2))
myplot(nonsynonymous, asp = 1, pch = 19, xlab = "", ylab = "",
  main = "Non synonymous codon usage")

```



This is reminiscent of something, let's have a look at amino-acid counts:

```
by(t(toycodon), facaa, colSums)
```

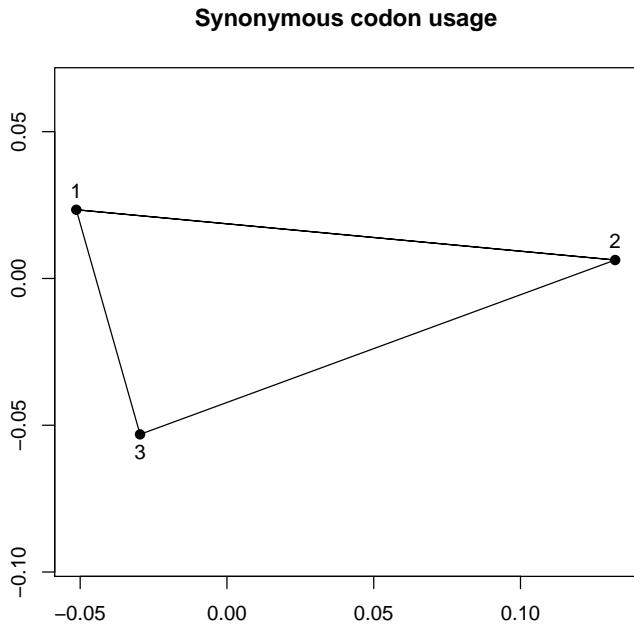
```

INDICES: Ala
1 2 3
130 60 60
-----
INDICES: Cys
1 2 3
0 0 5
-----
INDICES: Val
1 2 3
70 40 35

```

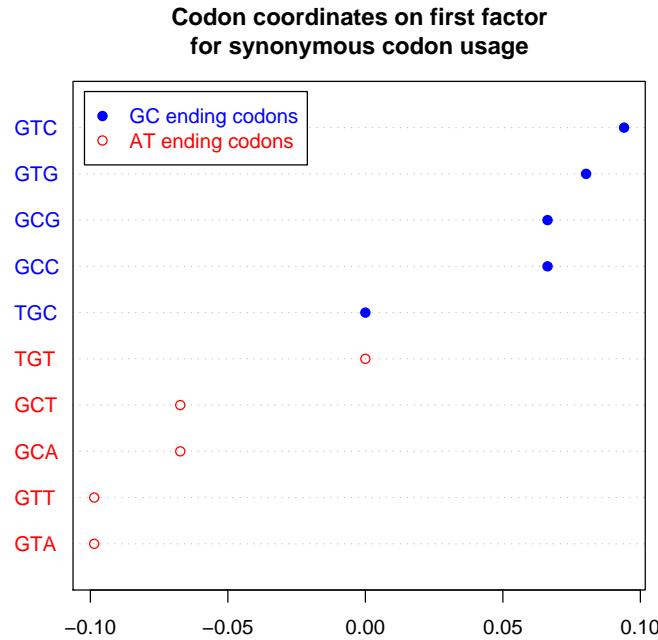
This is exactly the same data set that we used previously (table 5.1) at the amino-acid level. The non synonymous codon usage analysis is exactly the same as the amino-acid analysis. Coding sequence number 3 is far away because it codes for many Cys, a rare amino-acid. Note that at the global codon usage level, this is also the major visible structure. To get rid of this amino-acid effect, we use the synonymous codon usage analysis, that is the within amino-acid analysis:

```
synonymous <- t(within(dudi = t(global), fac = facaa, scann = FALSE,
  nf = 2))
myplot(synonymous, asp = 1, pch = 19, xlab = "", ylab = "",
  main = "Synonymous codon usage")
```



Now, coding sequence number 2 is away. When the amino-acid effect is removed, the pattern is then completely different. To interpret the result we look at the codon coordinates on the first factor of synonymous codon usage:

```
tmp <- synonymous$co[, 1, drop = FALSE]
tmp <- tmp[order(tmp$Axis1), , drop = FALSE]
colcod <- sapply(rownames(tmp), function(x) ifelse(substr(x,
  3, 3) == "c" || substr(x, 3, 3) == "g", "blue", "red"))
pchcod <- ifelse(colcod == "red", 1, 19)
dotchart(tmp$Axis1, labels = toupper(rownames(tmp)), color = colcod,
  pch = pchcod, main = "Codon coordinates on first factor\nfor synonymous codon usage")
legend("topleft", inset = 0.02, legend = c("GC ending codons",
  "AT ending codons"), text.col = c("blue", "red"), pch = c(19,
  1), col = c("blue", "red"), bg = "white")
```



At the synonymous level, coding sequence number 2 is different because it is enriched in GC-ending codons as compared to the two others. Note that this is hard to see at the global codon usage level because of the strong amino-acid effect.

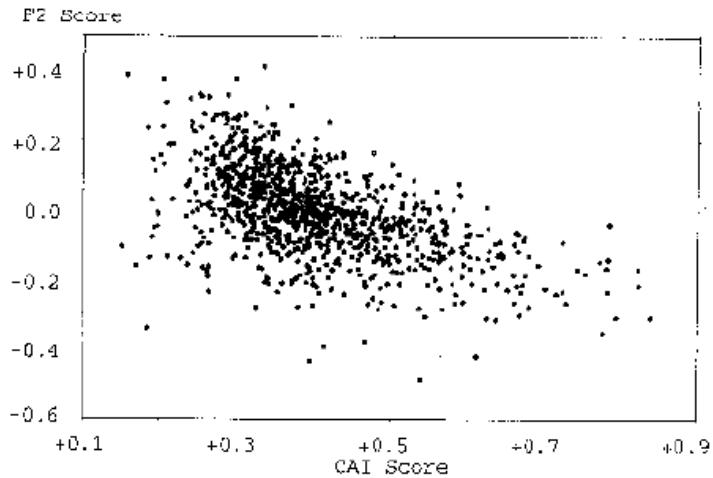
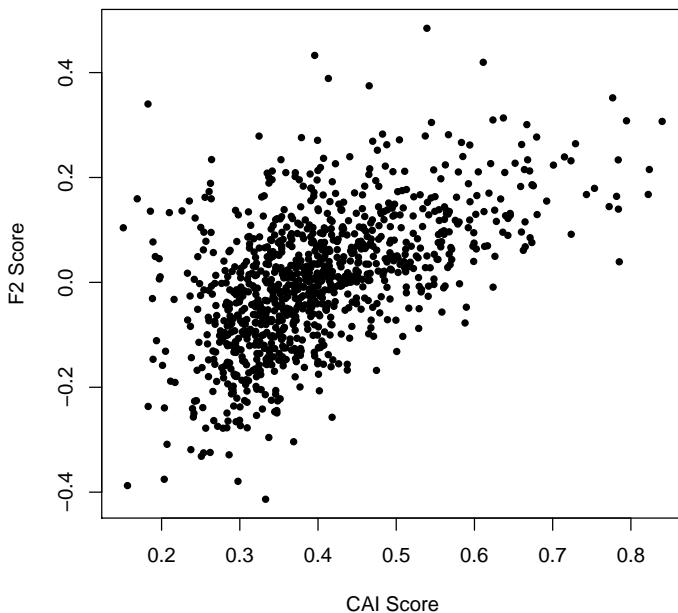


Figure 5.1: Screenshot of figure 5 from [39]. Each point represents a protein. This was to show the correlation between the codon adaptation index (CAI Score) with the second factor of correspondence analysis at the amino-acid level (F2 Score). Highly expressed genes have a high CAI value.

To illustrate the interest of synonymous codon usage analyses, let's use now a more realistic example. In [39] there was an assertion stating that selection for translation optimisation in *Escherichia coli* was also visible at the amino-acid level. The argument was in figure 5 of the paper (*cf* fig 5.1), that can be reproduced<sup>2</sup> with the following R code:

```
ec <- read.table(file = "ftp://pbil.univ-lyon1.fr/pub/datasets/NAR94/data.txt",
  header = TRUE, row.names = 1)
ec.coa <- dudi.coa(ec, scann = FALSE, nf = 3)
tmp <- read.table(file = "ftp://pbil.univ-lyon1.fr/pub/datasets/NAR94/ecoli999.cai")
caj <- exp(tmp$V2)
plot(caj, ec.coa$li[, 2], pch = 20, xlab = "CAI Score", ylab = "F2 Score",
  main = "Fig 5 from Lobry & Gautier (1994) NAR 22:3174")
```

**Fig 5 from Lobry & Gautier (1994) NAR 22:3174**



So, there was a correlation between the CAI (Codon Adaptation Index [56]) and the second factor for amino-acid composition variability. However, this is not completely convincing because the CAI is not completely independent of the amino-acid composition of the protein. Let's use within amino-acid correspondence analysis to remove the amino-acid effect. Here is a commented step-by-step analysis:

```
data(ec999)
class(ec999)
```

```
[1] "list"

names(ec999)[1:10]
```

---

<sup>2</sup> the code to reproduce all figures from [39] is available at <http://pbil.univ-lyon1.fr/members/lobry/repro/nar94/>.

```
[1] "ECFOLE.FOLE"      "ECMSBAG.MSBA"      "ECNARZYW-C.NARV" "ECNARZYW-C.NARW"
[5] "ECNARZYW-C.NARY" "ECNARZYW-C.NARZ" "ECNIRBC.NIRB"    "ECNIRBC.NIRD"
[9] "ECNIRBC.NIRC"    "ECNIRBC.CYSG"
```

```
ec999[[1]][1:50]
```

```
[1] "a" "t" "g" "c" "c" "a" "t" "c" "a" "c" "t" "c" "a" "g" "t" "a" "a" "a"
[19] "g" "a" "a" "g" "c" "g" "g" "c" "c" "c" "t" "g" "g" "t" "t" "c" "a" "t"
[37] "g" "a" "a" "g" "c" "g" "t" "t" "a" "g" "t" "t" "g" "c"
```

This is to load the data from [39] which is available as `ec999` in the `seqinR` package. The letters `ec` are for the bacterium *Escherichia coli* and the number 999 means that there were 999 coding sequences available from this species at that time. The class of the object `ec999` is a list, which names are the coding sequence names, for instance the first coding sequence name is `ECFOLE.FOLE`. Each element of the list is a vector of character, we have listed just above the 50 first character of the first coding sequence of the list with `ec999[[1]][1:50]`, we can see that there is a start codon (ATG) at the beginning of the first coding sequence.

```
ec999.uco <- lapply(ec999, uco)
class(ec999.uco)
```

```
[1] "list"
```

```
class(ec999.uco[[1]])
```

```
[1] "table"
```

```
ec999.uco[[1]]
```

```
aaa aac aag aat aca acc acg act aga agc agg agt ata atc atg att caa cac cag
 9   5   2   4   2   8   8   1   0   2   0   4   0   9   8   6   2   3   7
cat cca ccc ccg cct cga cgc cgg cgt cta ctc ctg ctt gaa gac gag gat gca gcc
 7   1   1   6   0   1   7   1   4   1   3   13   3   12   3   1   9   1   6
gcg gct gga ggc ggg ggt gta gtc gtg gtt taa tac tag tat tca tcc tcg tct tga
 7   5   2   3   0   4   0   5   9   4   0   2   0   2   2   3   2   1   1
tgc tgg tgt tta ttc ttg ttt
 1   0   1   1   4   2   3
```

This is to compute the codon usage, that is how many times each codon is used in each coding sequence. Because `ec999` is a list, we use the function `lapply()` to apply the same function, `uco()`, to all the elements of the list and we store the result in the object `ec999.uco`. The object `ec999.uco` is a list too, and all its elements belong to the class table.

```
df <- as.data.frame(lapply(ec999.uco, as.vector))
dim(df)
```

```
[1] 64 999
```

```
df[1:5, 1:5]
```

	ECFOLE.FOLE	ECMSBAG.MSBA	ECNARZYW.C.NARV	ECNARZYW.C.NARW	ECNARZYW.C.NARY	
1	9	15	2	6	23	
2	5	18	2	4	16	
3	2	8	1	3	4	
4	4	3	2	2	4	
5	2	3	1	1	0	

This is to put the codon usage into a data.frame. Note that the codons are in row and the coding sequences are in columns. This is more convenient for the following because groups for within and between analyses are usually handled by row.

```
row.names(df) <- names(ec999.uco[[1]])
df[1:5, 1:5]
```

	ECFOLE.FOLE	ECMSBAG.MSBA	ECNARZYW.C.NARV	ECNARZYW.C.NARW	ECNARZYW.C.NARY	
aaa	9	15	2	6	23	
aac	5	18	2	4	16	
aag	2	8	1	3	4	
aat	4	3	2	2	4	
aca	2	3	1	1	0	

This is to keep a trace of codon names, just in case we would like to re-order the dataframe df. This is important because we can now play with the data at will without loosing any critical information.

```
ec999.coa <- dudi.coa(df = df, scannf = FALSE)
ec999.coa
```

```
Duality diagramm
class: coa dudi
$call: dudi.coa(df = df, scannf = FALSE)

$nf: 2 axis-components saved
$rank: 63
$eigen values: 0.05536 0.02712 0.02033 0.01884 0.01285 ...
  vector length mode content
1 $cw    999   numeric column weights
2 $lw    64    numeric row weights
3 $eig   63    numeric eigen values

  data.frame nrow ncol content
1 $tab      64   999  modified array
2 $li       64    2   row coordinates
3 $l1       64    2   row normed scores
4 $co       999   2   column coordinates
5 $c1       999   2   column normed scores
other elements: N
```

This is to run global correspondence analysis of codon usage. We have set the scannf parameter to FALSE because otherwise the eigenvalue bar plot is displayed for the user to select manually the number of axes to be kept.

```
facaa <- as.factor(translate(s2c(c2s(rownames(df)))))

facaa
```

```
[1] Lys Asn Lys Asn Thr Thr Arg Ser Arg Ser Ile Ile Met Ile Gln His
[19] Gln His Pro Pro Pro Pro Arg Arg Arg Arg Leu Leu Leu Leu Glu Asp Glu Asp
[37] Ala Ala Ala Ala Gly Gly Gly Gly Val Val Val Val Stp Tyr Stp Tyr Ser Ser
[55] Ser Ser Stp Cys Trp Cys Leu Phe Leu Phe
21 Levels: Ala Arg Asn Asp Cys Gln Glu Gly His Ile Leu Lys Met Phe ... Val
```

This is to define a factor for amino-acids. The function translate() use by default the standard genetic code and this is OK for *E. coli*.

```

ec999.syn <- within(dudi = ec999.coa, fac = facaa, scannf = FALSE)
ec999.syn

Within analysis
call: within(dudi = ec999.coa, fac = facaa, scannf = FALSE)
class: within dudi

$nf (axis saved) : 2
$rank: 43
$ratio: 0.6438642

eigen values: 0.04855 0.0231 0.01425 0.007785 0.006748 ...

  vector length mode   content
1 $eig    43    numeric eigen values
2 $lw     64    numeric row weights
3 $cw     999   numeric col weights
4 $stabw 21    numeric table weights
5 $fac    64    numeric factor for grouping

  data.frame nrow ncol content
1 $tab      64    999 array class-variables
2 $li       64    2    row coordinates
3 $l1       64    2    row normed scores
4 $co       999   2    column coordinates
5 $c1       999   2    column normed scores
6 $ls       64    2    supplementary row coordinates
7 $as       2     2    inertia axis onto within axis

```

This is to run the synonymous codon usage analysis. The value of the `ratio` component of the object `ec999.syn` shows that most of the variability is at the synonymous level, a common situation in codon usage studies.

```

ec999.btw <- between(dudi = ec999.coa, fac = facaa, scannf = FALSE)
ec999.btw

Between analysis
call: between(dudi = ec999.coa, fac = facaa, scannf = FALSE)
class: between dudi

$nf (axis saved) : 2
$rank: 20
$ratio: 0.3561358

eigen values: 0.01859 0.0152 0.01173 0.01051 0.008227 ...

  vector length mode   content
1 $eig    20    numeric eigen values
2 $lw     21    numeric group weights
3 $cw     999   numeric col weights

  data.frame nrow ncol content
1 $tab      21    999 array class-variables
2 $li       21    2    class coordinates
3 $l1       21    2    class normed scores
4 $co       999   2    column coordinates
5 $c1       999   2    column normed scores
6 $ls       64    2    row coordinates
7 $as       2     2    inertia axis onto between axis

```

This is to run the non-synonymous codon usage analysis, or amino-acid usage analysis.

```

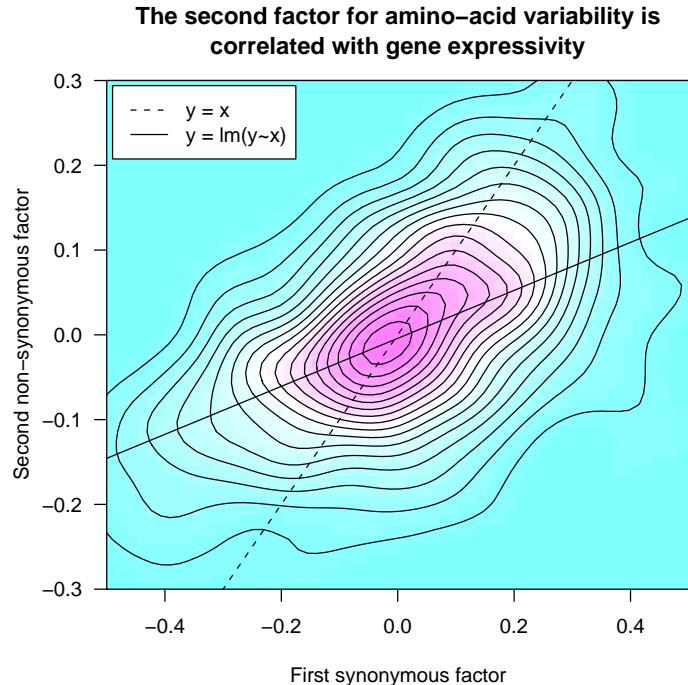
x <- ec999.syn$co[, 1]
y <- ec999.btw$co[, 2]
kxy <- kde2d(x, y, n = 100)
nlevels <- 25
breaks <- seq(from = min(kxy$z), to = max(kxy$z), length = nlevels +
  1)
col <- cm.colors(nlevels)
image(kxy, breaks = breaks, col = col, xlab = "First synonymous factor",
  ylab = "Second non-synonymous factor", xlim = c(-0.5,

```

```

0.5), ylim = c(-0.3, 0.3), las = 1, main = "The second factor for amino-acid variability is\nuncorrelated with
contour(kxy, add = TRUE, nlevels = nlevels, drawlabels = FALSE)
box()
abline(c(0, 1), lty = 2)
abline(lm(y ~ x))
legend("topleft", lty = c(2, 1), legend = c("y = x", "y = lm(y~x)"),
inset = 0.01, bg = "white")

```



This is to plot the whole thing. We have extracted the coding sequences coordinates on the first synonymous factor and the second non-synonymous factor within  $x$  and  $y$ , respectively. Because we have many points, we use the two-dimensional kernel density estimation provided by the function `kde2d()` from package MASS.

*To be completed*

## 5.3 Session Informations

This part was compiled under the following  environment:

- R version 2.4.1 (2006-12-18), i386-apple-darwin8.8.1
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: MASS 7.2-30, ade4 1.4-2, ape 1.9-3, gee 4.13-12, lattice 0.14-16, nlme 3.1-78, seqinr 1.0-7, xtable 1.4-3

	aaa	a	prec	p	h	tot	gc
1	Ala	A	pyr	1	5	12	h
2	Cys	C	3pg	7	9	25	m
3	Asp	D	oaa	1	6	13	m
4	Glu	E	akg	3	6	15	m
5	Phe	F	2 pep, eryP	13	19	52	l
6	Gly	G	3pg	2	5	12	h
7	His	H	penP	20	9	38	m
8	Ile	I	pyr, oaa	4	14	32	l
9	Lys	K	oaa, pyr	4	13	30	l
10	Leu	L	2 pyr, acCoA	3	12	27	l
11	Met	M	oaa, Cys, -pyr	10	12	34	m
12	Asn	N	oaa	3	6	15	l
13	Pro	P	akg	4	8	20	h
14	Gln	Q	akg	4	6	16	m
15	Arg	R	akg	11	8	27	h
16	Ser	S	3pg	2	5	12	m
17	Thr	T	oaa	3	8	19	m
18	Val	V	2 pyr	2	11	23	m
19	Trp	W	2 pep, eryP, PRPP, -pyr	28	23	74	m
20	Tyr	Y	eryP, 2 pep	13	18	50	l

Table 5.4: Aerobic cost of amino-acids in *Escherichia coli* and G+C classes to be loaded with `data(aacost)`.

---

---

# CHAPTER 6

---

## Nonparametric statistics

Palmeira, L.

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### 6.1 Introduction

Nonparametric statistical methods were initially developed to study variables for which little or nothing is known concerning their distribution. This makes them particularly suitable for statistical analysis of biological sequences, in particular for the study of over- and under-representation of  $k$ -letter words.

We will briefly describe two statistics for the measure of dinucleotide over- and under-representation in sequences [29, 46], which can both be computed with **seqinR**. We will subsequently use them to answer the long-time controversial question concerning the relationship between UV exposure and genomic content in bacteria [57, 2].

### 6.2 Determining dinucleotides over- and under-representation

#### 6.2.1 The *rho* statistic

The  $\rho$  statistic (`rho()`), presented in [29], measures the over- and under-representation of two-letter words:

$$\rho(xy) = \frac{f_{xy}}{f_x \times f_y}$$

where  $f_{xy}$  and  $f_x$  are respectively the frequencies of dinucleotide  $xy$  and nucleotide  $x$  in the studied sequence. The underlying model of random generation considers dinucleotides to be formed according to the specific frequencies of the two nucleotides that compose it ( $\rho_{xy} = 1$ ). Departure from this value characterizes either over- or under-representation of dinucleotide  $xy$ .

We expect the  $\rho$  statistic of a randomly generated sequence to be neither over- nor under-represented. Indeed, when we compute the  $\rho$  statistic on 500 random sequences, we can fit a normal distribution which is centered on 1 (see Fig. 6.1)

```
rhoseq <- sapply(seq(n), function(x) {
  rho(sample(s2c("acgt"), 6000, rep = TRUE))
})
hist(rhoseq[di, ], freq = FALSE, xlab = "Rho statistic", main = paste("Distribution for dinucleotide",
  toupper(labels(rhoseq)[[1]][di]), "on", n, "random sequences"),
  las = 1, col = grey(0.8), border = grey(0.5))
abline(v = 1, lwd = 2, lty = 3)
min = min(rhoseq[di, ])
max = max(rhoseq[di, ])
x = seq(min, max, length.out = 1000)
lines(x, dnorm(x, mean = mean(rhoseq[di, ])), sd = sd(rhoseq[di,
  ])), lwd = 2, lty = 2)
```

**Distribution for dinucleotide AT on 500 random sequences**

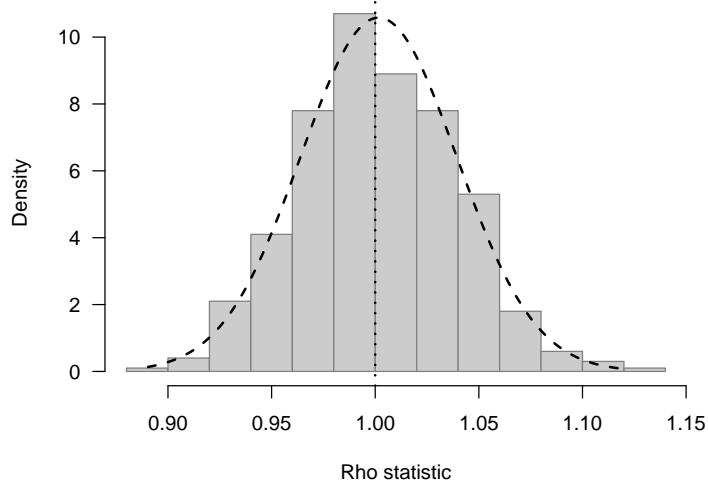


Figure 6.1: Distribution of the  $\rho$  statistic computed on 500 random sequences. The vertical dotted line is centered on 1. The dashed curve draws the fitted normal distribution.

The downside of this statistic, is that the model against which we compare the sequence under study is fixed. For several types of sequences, dinucleotides

are far from being formed by mere chance (CDS, ...). In this case, the model used in the  $\rho$  statistic becomes trivial, and the over- or under-representations measured are mainly due to the strong constraints acting on those sequences.

### 6.2.2 The `zscore` statistic

The `zscore` statistic (`zscore()`) is inspired by the  $\rho$  statistic, and is defined so that several different models can be used for the determination of over- and under-representation [46]. It allows for a finer measure of over- and under-representation in sequences, according to the chosen model.

The `zscore` is defined as follows:

$$z_{\text{score}} = \frac{\rho_{xy} - E(\rho_{xy})}{\sqrt{\text{Var}(\rho_{xy})}}$$

where  $E(\rho_{xy})$  and  $\text{Var}(\rho_{xy})$  are the expected mean and variance of  $\rho_{xy}$  according to a given model that describes the sequence.

This statistic follows the standard normal distribution, and can be computed with several different models of random sequence generation based on permutations from the original sequence (`modele` argument). More details on those models can be obtained in the documentation for the `zscore()` function, by simply typing `?zscore`.

For instance, if we want to measure the over- and under-representation of dinucleotides in CDS sequences, we can use the `codon` model, which measures the over- and under-representations existing in the studied sequence once codon usage bias has been erased. For intergenic sequences, or sequences for which no good permutation model can be established, we can use the `base` model.

### 6.2.3 Comparing statistics on a sequence

Let's have a look at what these different statistics can show. First, we will extract a CDS sequence of *Escherichia coli*'s chromosome from the Genome Reviews database. We will first make a request to retrieve all available CDS from this bacteria:

```
choosebank("greview")
query("coli", "sp=escherichia coli et t=cds et no k=partial")
sequence <- getSequence(colิ$req[[448]])
```

From the 34939 sequences annotated as CDS in the Genome Reviews database, let's choose one coding sequence: say, for instance, number 448. We can see that this CDS encodes a maltose O-acetyltransferase protein (`getAnnot(colิ$req[[448]], 30)`). We will now compare the three following nonparametric statistics:

- the  $\rho$  statistic,
- the `zscore` statistic with `base` model,
- and the `zscore` statistic with `codon` model.

```

rhocoli = rho(sequence)
zcolibase = zscore(sequence, mod = "base")
zcolicodon = zscore(sequence, mod = "codon")
par(mfrow = c(1, 3))
plot(rhocoli - 1, ylim = c(-0.5, 0.5), las = 1, ylab = "rho")
plot(zcolibase, ylim = c(-2.5, 2.5), las = 1, ylab = "zscore with base model")
plot(zcolicodon, ylim = c(-2.5, 2.5), las = 1, ylab = "zscore with codon model")

```

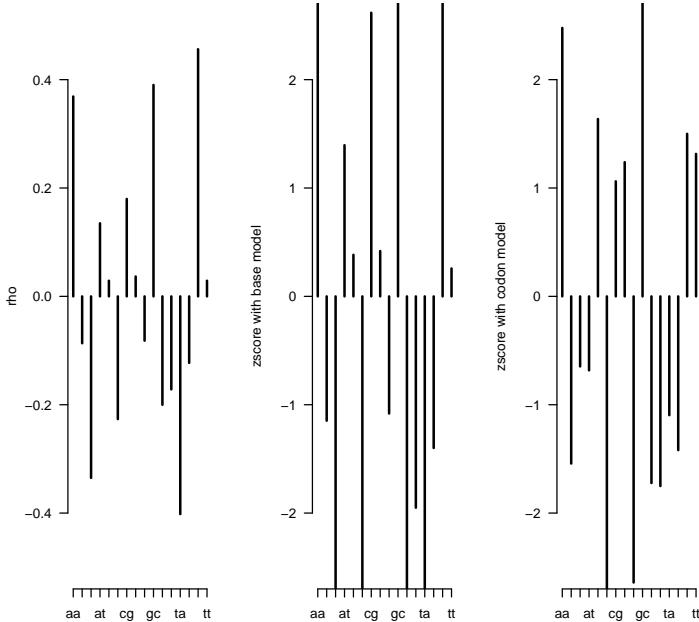


Figure 6.2: Three different non-parametric statistics (from left to right:  $\rho$ ,  $zscore$  with **base** model,  $zscore$  with **codon** model), computed on the same sequence from *Escherichia coli*. In order to make the figures easily comparable, we subtracted 1 to the `rho()` results, so that all 3 statistics are centered on 0.

The first two figures are almost identical: this is due to the way the  $zscore$  statistic has been built. The statistic computed with the **base** model is a reflection of the  $\rho$  statistic. The difference being that the  $zscore$  follows a standard normal distribution, which makes easier the comparisons between the results from the **base** model and the ones from the **codon** model. The last figure ( $zscore$  with **codon** model), is completely different: almost all over- and under-representations have been erased. We can safely say that these over- and under-representations were due to codon usage bias.

On this last figure, four dinucleotides stand out: CC and TT seem rather under-represented, CT and TC rather over-represented. This means that, in this sequence, codons ending with a given pyrimidine tend to be more frequently followed by a codon starting with the other pyrimidine than expected by chance. This is not a universal feature of *Escherichia coli*, and is probably due to the amino-acid composition of this particular sequence. It seemed a funny example,

as the following part will also relate to pyrimidine dinucleotides. However, what we see on this CDS from *Escherichia coli* has nothing to do with what follows...

## 6.3 UV exposure and dinucleotide content

In the beginning of the 1970's, two contradictory papers considered the question of the impact of UV exposure on genomic content. Both papers had strong arguments for either side, and the question remained open until recently [46].

### 6.3.1 The expected impact of UV light on genomic content

On this controversy, the known facts are: pyrimidine dinucleotides (CC, TT, CT and TC) are the major DNA target for UV-light [55]; the sensitivities of the four pyrimidine dinucleotides to UV wavelengths differ and depend on the micro-organism [55]:

	G+C content	CC (%)	CT + TC (%)	TT (%)
<i>Haemophilus influenzae</i>	62	5	24	71
<i>Escherichia coli</i>	50	7	34	59
<i>Micrococcus lysodeikticus</i>	30	26	55	19

Table 6.1: Proportion of dimers formed in the DNA of three bacteria after irradiation with 265 nm UV light. Table adapted from [55].

The hypothesis presented by Singer and Ames [57] is that pyrimidine dinucleotides are avoided in light-exposed micro-organisms. At the time, only G+C content is available, and – based exclusively on the sensitivity of the four pyrimidine dinucleotides in an *Escherichia coli* chromosome – they hypothesize that a high G+C will result in less pyrimidine target. Indeed, they find that bacteria exposed to high levels of UV have higher G+C content than the others. Bak *et al.* [2] strongly criticize their methodology, but no clear cut answer is achieved.

In an *Escherichia coli* chromosome, it is true that a sequence with a high G+C content will contain few phototargets (see Fig. 6.3).

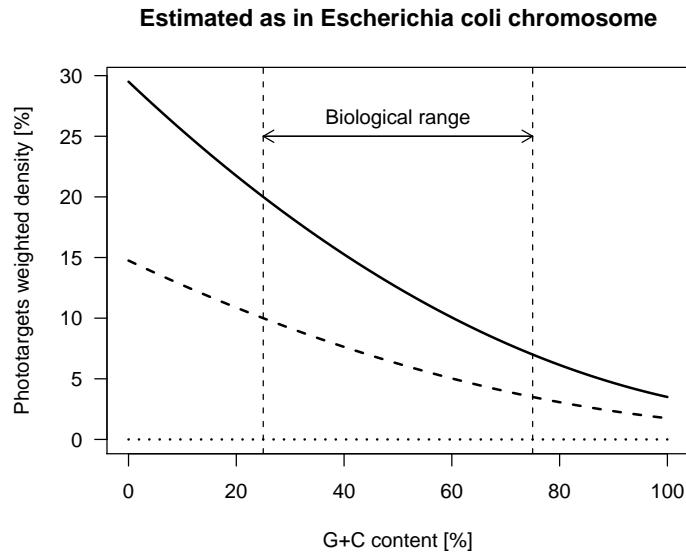


Figure 6.3: Density of phototargets, weighted by their frequency in the *Escherichia coli* chromosome, and calculated for different G+C contents and for three kinds of random genomes. The weights are as follows:  $0.59 * f_{tt} + 0.34 * (f_{tc} + f_{ct}) + 0.07 * f_{cc}$  (where  $f_{xy}$  is the frequency of dinucleotide  $xy$  in the specified genome). Three models of random genomes are analyzed. In the worst case (solid curve), the genome is the concatenation of a sequence of pyrimidines and a sequence of purines: all pyrimidines are involved in a pyrimidine dinucleotide. In the best case (dotted curve), the genome is an unbroken succession of pyrimidine-purine dinucleotides: no pyrimidine is involved in a pyrimidine dinucleotide. In the "random case" (dashed curve), the frequency of a pyrimidine dinucleotide is the result of chance ( $f_{xy} = f_x \times f_y$ ).

In a *Micrococcus lysodeikticus* sequence (see Fig. 6.4), we can see that this is no longer true...

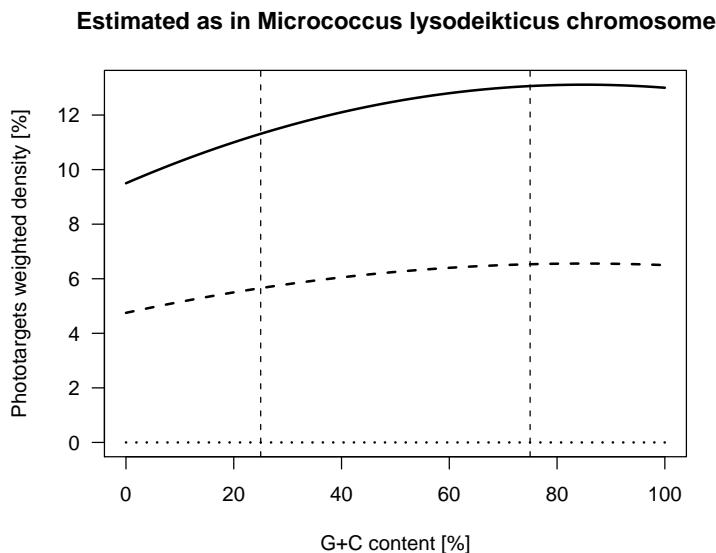


Figure 6.4: Density of phototargets, weighted by their frequency in the *Micrococcus lysodeikticus* chromosome, and calculated for different G+C contents and for three kinds of random genomes. The weights are as follows:  $0.19 * f_{tt} + 0.55 * (f_{tc} + f_{ct}) + 0.26 * f_{cc}$ . See previous figure for more details.

These two figures show that the density of phototargets depends on:

- the degree of aggregation of pyrimidine dinucleotides in the sequence,
- the sensitivities of the four pyrimidine dinucleotides.

Instead of looking at G+C content, which is an indirect measure of the impact of UV exposure on genomic content, let us look at pyrimidine dinucleotide content.

Are CC, TT, CT and TC dinucleotides avoided in light-exposed bacteria?

### 6.3.2 The measured impact of UV light on genomic content

On all available genomes (as retrieved from Genome Reviews database on June 16, 2005), we have computed the mean of the *zscores* with the **base** model on all intergenic sequences, and the mean of the *zscores* with the **codon** model on all CDS:

```
data(dinucl)
```

The results show that there is no systematic under-representation of none of the four pyrimidine dinucleotides (see Fig. 6.5).

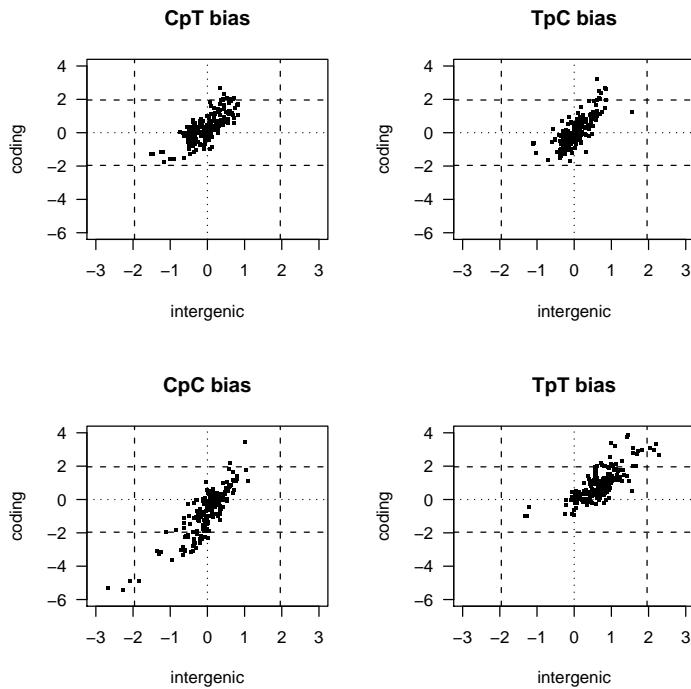


Figure 6.5: Plot of the mean *zscore* statistics for **intergenic sequences** (x-axis) and for **coding sequences** (y-axis), for each of the four pyrimidine dinucleotides. On each plot, a dot corresponds to the mean of these two statistics in a given prokaryote chromosome. The null x and y axis (dotted lines), and the 5% limits of significance for the standard normal distribution (dashed lines) are plotted as benchmarks. It should be noted that the variability within one chromosome is sometimes as great as that between different chromosomes.

However, we have little or no information on the exposure of this bacteria to UV light. In order to fully answer this question, let's do another analysis and look at *Prochlorococcus marinus* genome.

*Prochlorococcus marinus* seems to make an ideal model for investigating this hypothesis. Three completely sequenced strains are available in the Genome reviews database: two of these strains are adapted to living at a depth of more than 120 meters (accession numbers AE017126 and BX548175), and the other one at a depth of 5 meters (accession number BX548174).

Living at a depth of 5 meters, or at a depth of more than a 120 meters is totally different in terms of UV exposure: the residual intensity of 290 nm irradiation (UVb) in pure water can be estimated to 56% of its original intensity at 5 m depth and to less than 0.0001% at more than 120 m depth. For this reason, two of the *Prochlorococcus marinus* strains can be considered to be adapted to low levels of UV exposure, and the other one to much higher levels. Is pyrimidine dinucleotide content different in these three strains? And is it linked to their UV exposure?

We have computed the *zscore* with the `codon` model on all CDS from each of these three strains (as retrieved from Genome Reviews database on June 16, 2005):

```
data(prochlo)
```

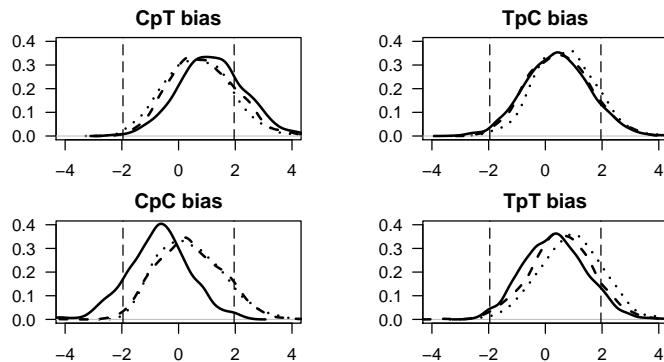


Figure 6.6: Each figure shows the distributions of the *zscore* in all **coding sequences** corresponding to each of the three strains of *Prochlorococcus marinus*. In each figure, the distribution for the MED4 (a high-light adapted strain) is shown as a solid line; the distribution for the SS120 (a low-light adapted strain) is shown as a dashed line, and the distribution for the MIT 9313 (a low-light adapted strain) is shown as a dotted line. The 5% limits of significance for the standard normal distribution (dashed vertical lines) are plotted as benchmarks.

Figure 6.6 shows that there is no difference between the relative abundances of pyrimidine dinucleotides in these three strains. We can say that pyrimidine dinucleotides are not avoided, and that the hypothesis by Singer and Ames [57] no longer stands [46].

## 6.4 Session Informations

This part was compiled under the following environment:

- R version 2.4.1 (2006-12-18), i386-apple-darwin8.8.1
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: MASS 7.2-30, ade4 1.4-2, ape 1.9-3, gee 4.13-12, lattice 0.14-16, nlme 3.1-78, seqinr 1.0-7, xtable 1.4-3



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## APPENDIX A

---

### FAQ: Frequently Asked Questions

Lobry, J.R.

#### A.1 How do I compute a score on my sequences?

In the example below we want to compute the G+C content in third codon positions for complete ribosomal CDS from *Escherichia coli*:

```
choosebank("emblTP")
query("ecrivo", "sp=escherichia coli ET t=cds ET k=ribosom0 ET NO k=partial")
myseqs <- sapply(ecrivo$req, getSequence)
(gc3 <- sapply(myseqs, GC3))
```

```
[1] 0.4946237 0.6046512 0.5000000 0.6194030 0.5772727 0.4838710 0.5980066
[8] 0.4974359 0.5031250 0.4324324 0.5000000 0.5113636 0.5290520 0.6142857
[15] 0.4904762 0.5714286 0.6191860 0.5906040 0.4880000 0.4880000 0.4946237
[22] 0.6046512 0.5000000 0.3522727 0.5076923 0.4343434 0.6194030 0.5522388
[29] 0.6104651 0.5661157 0.4946237 0.4946237 0.6079734 0.5000000 0.6343284
[36] 0.4659091 0.5789474 0.4946237 0.5000000 0.4974359 0.5689655 0.4611111
[43] 0.4611111 0.5303030 0.5303030 0.4482759 0.4201681 0.5915493 0.5000000
[50] 0.3829787 0.4519231 0.4302326 0.5696203 0.4285714 0.5689655 0.5000000
[57] 0.5224417 0.5661157 0.6057692 0.4444444 0.4659091 0.4130435 0.4946237
[64] 0.5661157 0.4946237 0.5680272
```

At the amino-acid level, we may get an estimate of the isoelectric point of the proteins this way:

```
sapply(sapply(myseqs, getTrans), computePI)
```

```
[1] 6.624309 7.801329 10.864793 5.931989 7.830476 6.624309 7.801329
[8] 9.203410 9.826485 5.674672 7.154423 6.060457 6.313741 5.571446
[15] 9.435422 4.310745 6.145496 4.876054 11.006424 10.876041 6.624309
[22] 7.801329 10.864793 9.346289 9.203410 5.877050 5.931989 9.934988
[29] 5.920490 6.612505 6.624309 6.624309 7.801329 10.864793 5.931989
[36] 11.182505 9.598944 6.624309 10.864793 9.203410 11.031943 5.858421
[43] 5.858421 11.777511 11.777511 10.619175 11.365738 9.460987 10.864793
[50] 13.002381 9.845859 10.584862 11.421252 10.248320 11.031937 10.402075
[57] 4.863862 6.612505 9.681066 11.150304 11.182499 11.043602 6.624309
[64] 6.612505 6.624309 4.310747
```

Note that some pre-defined vectors to compute linear forms on sequences are available in the EXP data.

As a matter of convenience, you may encapsulate the computation of your favorite score within a function this way:

```
GC3m <- function(list, ind = 1:list$nelem) sapply(sapply(list$req[ind],
  getSequence), GC3)
GC3m(ecribo)
```

```
[1] 0.4946237 0.6046512 0.5000000 0.6194030 0.5772727 0.4838710 0.5980066
[8] 0.4974359 0.5031250 0.4324324 0.5000000 0.5113636 0.5290520 0.6142857
[15] 0.4904762 0.5714286 0.6191860 0.5906040 0.4880000 0.4880000 0.4946237
[22] 0.6046512 0.5000000 0.3522727 0.5076923 0.4343434 0.6194030 0.5522388
[29] 0.6104651 0.5661157 0.4946237 0.4946237 0.6079734 0.5000000 0.6343284
[36] 0.4659091 0.5789474 0.4946237 0.5000000 0.4974359 0.5689655 0.4611111
[43] 0.4611111 0.5303030 0.5303030 0.4482759 0.4201681 0.5915493 0.5000000
[50] 0.3829787 0.4519231 0.4302326 0.5696203 0.4285714 0.5689655 0.5000000
[57] 0.5224417 0.5661157 0.6057692 0.4444444 0.4659091 0.4130435 0.4946237
[64] 0.5661157 0.4946237 0.5680272
```

```
GC3m(ecribo, 1:10)
```

```
[1] 0.4946237 0.6046512 0.5000000 0.6194030 0.5772727 0.4838710 0.5980066
[8] 0.4974359 0.5031250 0.4324324
```

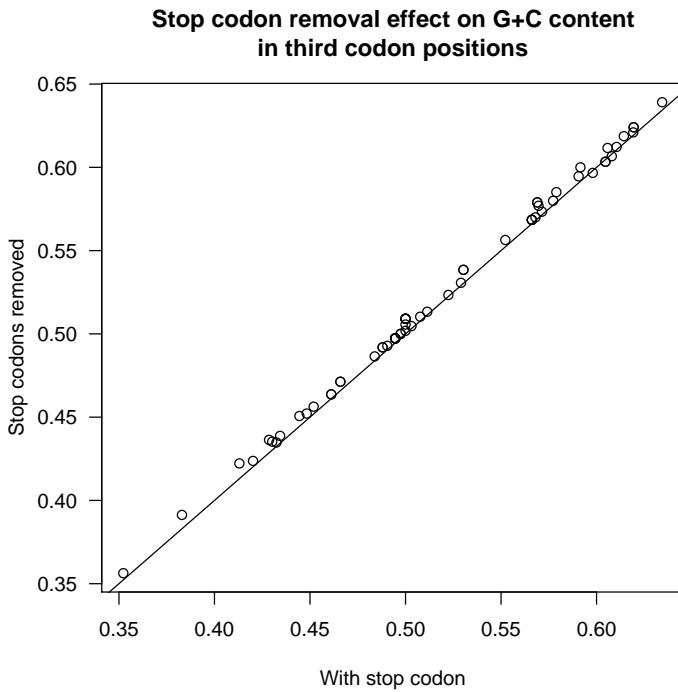
## A.2 Why do I have not exactly the same G+C content as in codonW?

This question was raised (and solved) by Oliver Clay in an e-mail (23-AUG-2006). The program codonW was written in C as part of John Peden's PhD thesis on Codon Usage [48] and is available at <http://codonw.sourceforge.net/>. The reason for the small differences in G+C content between the two programs is that the default behavior in codonW is to remove the stop codon before computations. Here is one way of removing the stop codon under R:

```
gc3nos <- sapply(myseqs, function(s) GC3(s[1:(length(s) -
  3)]))
```

As compared with the previous result, the difference is small but visible:

```
plot(x = gc3, y = gc3nos, las = 1, main = "Stop codon removal effect on G+C content\nin third codon position",
  xlab = "With stop codon", ylab = "Stop codons removed")
abline(c(0, 1))
```



CodonW was released with a test file called `input.dat`, here are the first 10 lines of the file copied from `CodonWSourceCode_1_4_4`:

```
inputdatfile <- system.file("sequences/input.dat", package = "seqinr")
cat(readLines(inputdatfile, n = 10), sep = "\n")

>YCG9 Probable      1377 residues Pha O Code 0
ATGAATATGCTCATTTGCGTAGAGTTGTTCTAGTGTTGGGGAAACGGGACTTCAAACG
CTTGCTTGTATTGGTTACGATGGTTGGTGAAGGTACGTCATTGGTGATTCC
ATCCTAAGTTGCTATTGGCTAGCTGCTATCGTGGTCTATAATCGGAGGTGCCCTT
ACAACCCATGTTACCTGGAGGTGGCTTCTATATCAATCTCCATCGGTGTCCTGCC
ATTATTATGTTTACTCACATATAAGGCCGAGAATAAGGTATACTTCAACAAATTAAA
GATGCTATAGGAACAATCTGAGCTTACTTTAGTAAGGTCAGACACCAAGTTAATTTT
AAAAGACTTATGAATGGCATATCTCAAGTTGACTCTTGGTTTGCCTCTGCTCT
GCAGGGCTGGCCTTCCACTGGGCTAACCTTGGTGGTAATAATAGTTGGAAC
TCTGGCCAAGTCATCGCATATTGGTTGGTCTTACTTTTATTTCATTGGT
```

This is a FASTA file that we import under with:

```
input <- read.fasta(file = inputdatfile)
names(input)

[1] "YCG9"      "YCG8"      "ALPHA2"    "ALPHA1"    "CHA1"      "KRR1"
[7] "PRD1"      "KAR4"      "PBN1"      "LRE1"      "APA1"      "YCE9"
[13] "YCE8"      "YCE7"      "YCE5"      "YCE6"      "YCE4"      "PDI1"
[19] "GLK1"      "YCD8"      "SRD9"      "YCD6"      "YCD5"      "YCD3"
[25] "STE50"      "HIS4"      "BIK1"      "FUS1"      "YCO8"      "AGP1"
[31] "LEU2"      "NFS1"      "BUD3"      "GBP2"      "ILV6"      "CWH36"
[37] "PEL1"      "RER1"      "CDC10"     "MRLP32"    "YCP4"      "CIT2"
[43] "YCP7"      "SAT4"      "RVS161"    "YCO0"      "ADP1"      "PGK1"
[49] "POL4"      "YCQ7"      "SRD1"      "MAK32"     "PET18"     "MAK31"
[55] "HSP30"      "YCR3"      "SYN"       "YCR6"      "GNS1"      "FEN2"
[61] "RIM1"      "CRY1"      "YCS2"      "YCS3"      "GNS1"      "RBK1"
[67] "PHO87"      "BUD5"      "MATALPHA2" "MATALPHA1" "TSM1"      "YCT5"
[73] "PETCR46"    "YCT7"      "YCT9"      "ARE1"      "RSC6"      "THR4"
[79] "CTR86"      "PWP2"      "YCU9"      "YCV1"      "G10"       "HCM1"
[85] "RAD18"      "CYPR"      "YCW1"      "YCW2"      "SSK22"     "SOL2"
[91] "ERS1"      "PAT1"      "SRB8"      "YCX3"      "TUP1"      "YC16"
[97] "ABP1"      "KIN82"     "MSH3"      "CDC39"    "YCY4"      "A2"
[103] "GIT1"      "YCW0"      "YCZ1"      "YCZ2"      "YCZ3"      "PAU3"
[109] "YCW5"      "YCW6"      "YCW7"
```

The file `input.out` contains the values obtained with `codonW` for the GC content and GC3s content:

```
inputoutfile <- system.file("sequences/input.out", package = "seqinr")
cat(readLines(inputoutfile, n = 10), sep = "\n")
```

title	GC3s	GC
YCG9_Probable_-----13	0.335	0.394
YCG8_-----573_residues_	0.439	0.446
ALPHA2_-----633_residue	0.328	0.351
ALPHA1_-----528_residue	0.345	0.379
CHA1_-----1083_residue	0.328	0.394
KRR1_-----951_residue	0.364	0.384
PRD1_-----2139_residue	0.430	0.397
KAR4_-----1008_residue	0.354	0.383
PBN1_-----1251_residue	0.330	0.386

```
input.res <- read.table(inputoutfile, header = TRUE)
head(input.res)
```

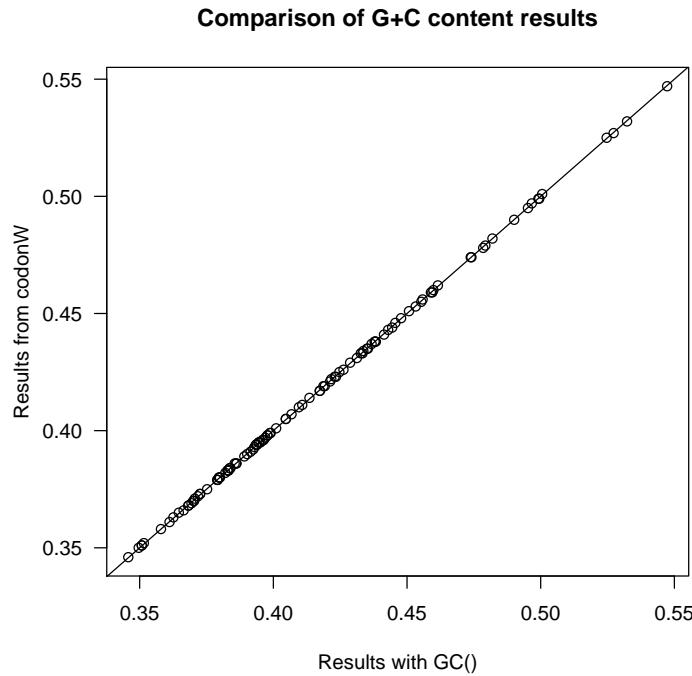
title	GC3s	GC
1 YCG9_Probable_-----13	0.335	0.394
2 YCG8_-----573_residues_	0.439	0.446
3 ALPHA2_-----633_residue	0.328	0.351
4 ALPHA1_-----528_residue	0.345	0.379
5 CHA1_-----1083_residue	0.328	0.394
6 KRR1_-----951_residue	0.364	0.384

Let's try to reproduce the results for the G+C content, we know that we have to remove the last stop codon:

```
input.gc <- sapply(input, function(s) GC(s[1:(length(s) -
  3)]))
max(abs(input.gc - input.res$GC))
```

```
[1] 0.0004946237
```

```
plot(x = input.gc, y = input.res$GC, las = 1, xlab = "Results with GC()", 
  ylab = "Results from codonW", main = "Comparison of G+C content results")
abline(c(0, 1))
```



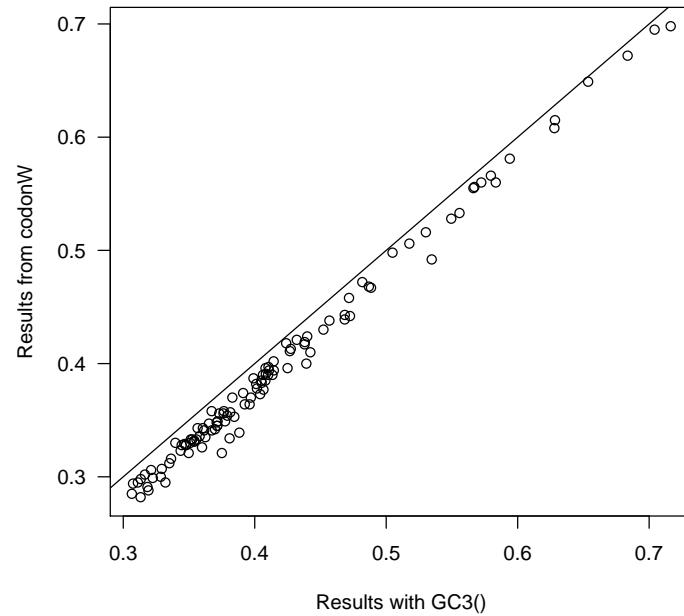
The results are consistent if we consider that we have 3 significant digits in the file `input.out`. Now, let's try to reproduce the results for G+C in third codon positions:

```
input.gc3 <- sapply(input, function(s) GC3(s[1:(length(s) - 3)]))
max(abs(input.gc3 - input.res$GC3s))

[1] 0.054

plot(x = input.gc3, y = input.res$GC3s, las = 1, xlab = "Results with GC3()", ylab = "Results from codonW", main = "Comparison of G+C content in third codon positions results")
abline(c(0, 1))
```

### Comparison of G+C content in third codon positions results

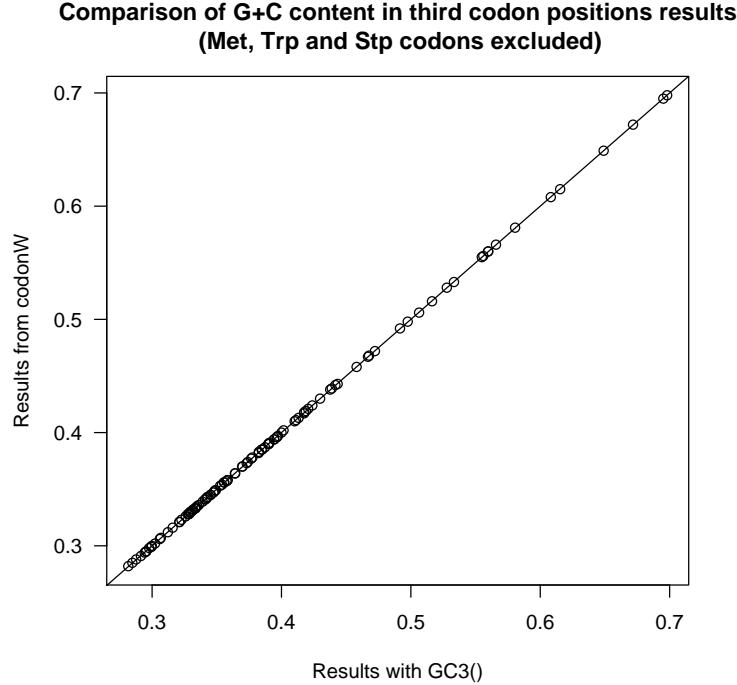


There is clearly a problem here. Looking into the documentation of `codonW`, `GC3s` is the G+C content in third codon position after removing non-synonymous and stop codons (those corresponding to Met, Trp, Stp). Let's remove these codons:

```
codons <- words()
names(codons) <- sapply(codons, function(c) aaa(translate(s2c(c),
  numcode = 1)))
okcodons <- codons[!names(codons) %in% c("Met", "Trp", "Stop")]
gc3s <- function(s) {
  tmp <- splitseq(s)
  tmp <- tmp[tmp %in% okcodons]
  tmp <- s2c(paste(tmp, collapse = ""))
  GC3(tmp)
}
input.gc3s <- sapply(input, gc3s)
max(abs(input.gc3s - input.res$GC3s))
```

```
[1] 0.0004980843
```

```
plot(x = input.gc3s, y = input.res$GC3s, las = 1, xlab = "Results with GC3()", 
  ylab = "Results from codonW", main = "Comparison of G+C content in third codon positions results\nM abline(c(0, 1))
```



The results are now consistent. But thinking more about it there is still a problem with the codons for Ile:

```
codons[names(codons) == "Ile"]
```

```
Ile    Ile    Ile
"ata" "atc" "att"
```

There are three codons for Ile. If the distribution of the four bases was uniform and selectively neutral in third codon position of synonymous codons, then we would expect to get a G+C of 50% in quartet and duet codons at third codons positions because they all have the same number of W (A or T) and S (C or G) bases in third position. But for Ile we have two codons ending in W versus only one in S so that we would get a G+C of  $\frac{1}{3}$  instead of  $\frac{1}{2}$ . This point was clearly stated [59] by Sueoka in 1988:

**G + C Content of the Three Codons Positions.** In the present analysis, observed G + C contents of the first, second, and third codon positions ( $P_1$ ,  $P_2$ , and  $P_3$ , respectively) are corrected average G + C contents of the three codon positions that are calculated from 56 triplets out of 64. Because of the inequality of  $\alpha$  and  $\gamma$  at the third codon position, the three stop codons (TAA, TAG, and TGA) and the three codons for isoleucine (ATT, ATC, and ATA) were excluded in calculation of  $P_3$ , and two single codons for methionine (ATG) and tryptophan (TGG) were excluded in all three ( $P_1$ ,  $P_2$ , and  $P_3$ )

Let's compute  $P_3$  and compare it with GC3s:

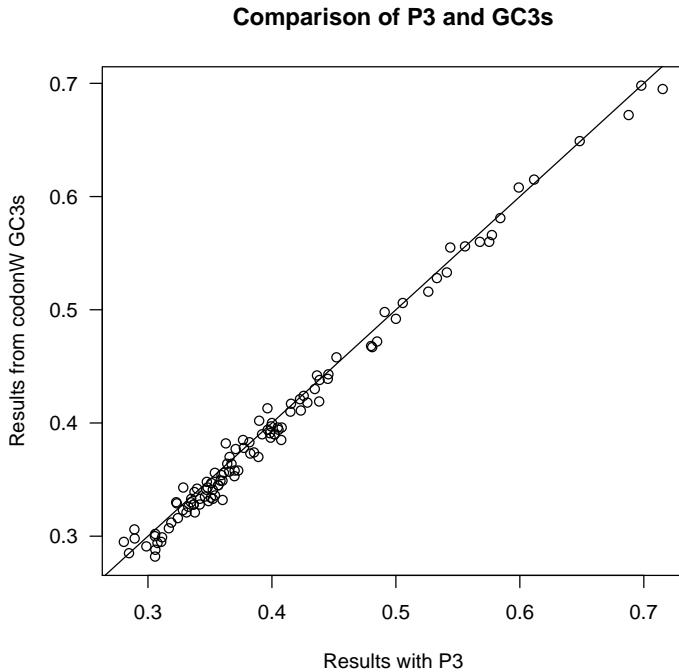
```

P3codons <- codons[!names(codons) %in% c("Met", "Trp", "Ile",
                                         "Stop")]
P3 <- function(s) {
  tmp <- splitseq(s)
  tmp <- tmp[tmp %in% P3codons]
  tmp <- s2c(paste(tmp, collapse = ""))
  GC3(tmp)
}
input.P3 <- sapply(input, P3)
max(abs(input.P3 - input.res$GC3s))

[1] 0.02821505

plot(x = input.P3, y = input.res$GC3s, las = 1, xlab = "Results with P3",
      ylab = "Results from codonW GC3s", main = "Comparison of P3 and GC3s")
abline(c(0, 1))

```



This is not exactly the same, the maximum observed difference here is about 3%. In practice,  $P_3$ , GC3, and GC3s are only slightly different [60].

### A.3 How do I get a sequence from its name?

This question is adapted from an e-mail (22 Jun 2006) by Gang Xu. I know that the UniProt (SwissProt) entry of my protein is P08758, if I know its name<sup>1</sup>, how can I get the sequence?

```

choosebank("swissprot")
query("myprot", "AC=P08758")
getSequence(myprot$req[[1]])

```

---

<sup>1</sup> More exactly, this is the accession number. Sequence names are not stable over time, it's always better to use the accession numbers.

```
[1] "M" "A" "Q" "V" "L" "R" "G" "T" "V" "T" "D" "F" "P" "G" "F" "D" "E" "R"
[19] "A" "D" "A" "E" "T" "L" "R" "K" "A" "M" "K" "G" "L" "G" "T" "D" "E" "E"
[37] "S" "I" "L" "T" "L" "L" "T" "S" "R" "S" "N" "A" "Q" "R" "Q" "E" "I" "S"
[55] "A" "A" "F" "K" "T" "L" "F" "G" "R" "D" "L" "L" "D" "D" "L" "K" "P" "S" "R" "L"
[73] "L" "T" "G" "K" "F" "E" "K" "I" "V" "A" "L" "M" "K" "P" "S" "R" "L"
[91] "Y" "D" "A" "Y" "E" "L" "K" "H" "A" "L" "K" "G" "A" "G" "T" "N" "E" "K"
[109] "V" "L" "T" "E" "I" "A" "S" "R" "T" "P" "E" "E" "L" "R" "A" "I" "K"
[127] "Q" "V" "Y" "E" "E" "Y" "G" "S" "S" "L" "E" "D" "D" "V" "V" "G" "D"
[145] "T" "S" "G" "Y" "Y" "Q" "R" "M" "L" "V" "V" "L" "L" "Q" "A" "N" "R" "D"
[163] "P" "D" "A" "G" "I" "D" "E" "A" "Q" "V" "E" "Q" "D" "A" "Q" "A" "L" "F"
[181] "Q" "A" "G" "E" "L" "K" "W" "G" "T" "D" "E" "E" "K" "F" "I" "T" "I" "F"
[199] "G" "T" "R" "S" "V" "S" "H" "L" "R" "K" "V" "F" "D" "K" "Y" "M" "T" "I"
[217] "S" "G" "F" "Q" "I" "E" "E" "T" "I" "D" "R" "E" "T" "S" "G" "N" "L" "E"
[235] "Q" "L" "L" "L" "A" "V" "V" "K" "S" "I" "R" "S" "I" "P" "A" "Y" "L" "A"
[253] "E" "T" "L" "Y" "Y" "A" "M" "K" "G" "A" "G" "T" "D" "D" "H" "T" "L" "I"
[271] "R" "V" "M" "V" "S" "R" "S" "E" "I" "D" "L" "F" "N" "I" "R" "K" "E" "F"
[289] "R" "K" "N" "F" "A" "T" "S" "L" "Y" "S" "M" "I" "K" "G" "D" "T" "S" "G"
[307] "D" "Y" "K" "K" "A" "L" "L" "L" "C" "G" "E" "D" "D"
```

## A.4 Session Informations

This part was compiled under the following  environment:

- R version 2.4.1 (2006-12-18), i386-apple-darwin8.8.1
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: MASS 7.2-30, ade4 1.4-2, ape 1.9-3, gee 4.13-12, lattice 0.14-16, nlme 3.1-78, seqinr 1.0-7, xtable 1.4-3



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# APPENDIX B

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---

# APPENDIX C

---

## Genetic codes

Lobry, J.R.

### C.1 Standard genetic code

The standard genetic code given in table C.1 was produced with the following `R` code and inserted with `\input{../tables/stdcode.tex}` within this `LATEX` document and referenced as `\ref{stdcode}` in the text.

```
tablecode(latexfile = "../tables/stdcode.tex", label = "stdcode",
          size = "small")
```

### C.2 Available genetic code numbers

The genetic code numbers are those from the NCBI<sup>1</sup> (<http://130.14.29.110/Taxonomy/Utils/wprintgc.cgi?mode=c>). This compilation from Andrzej (Anjay) Elzanowski, Jim Ostell, Detlef Leipe, and Vladimir Sossov is based primarily on two previous reviews [45, 27].

```
codes <- SEQINR.UTIL$CODES.NCBI
availablecodes <- which(codes$CODES != "deleted")
codes[availablecodes, "ORGANISMES", drop = FALSE]

                                     ORGANISMES
1             standard
2      vertebrate.mitochondrial
3          yeast.mitochondrial
4 protozoan.mitochondrial+mycoplasma
5      invertebrate.mitochondrial
6          ciliate+dasycladacean
9   echinoderm+flatworm.mitochondrial
10            euplotid
11      bacterial+plantplastid
12          alternativeyeast
13          ascidian.mitochondrial
14 alternativeflatworm.mitochondrial
15            blepharism
```

---

<sup>1</sup> National Center for Biotechnology Information, Bethesda, Maryland, U.S.A.

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	TGA	Stop
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table C.1: Genetic code number 1: standard.

```

16      chlorophycean.mitochondrial
21      trematode.mitochondrial
22      scenedesmus.mitochondrial
23      hraustochytrium.mitochondria

```

The tables of variant genetic codes outlining the differences were produced with the following  code:

```

cdorder <- paste(rep(s2c("tcag"), each = 16), s2c("tcag"),
  sep = ""), rep(s2c("tcag"), each = 4), sep = "")
stdcode <- sapply(lapply(cdorder, s2c), translate, numcode = 1)
for (cd in availablecodes[-1]) {
  Tfile <- paste("../tables/codnum", cd, ".tex", sep = "")
  preemph <- "\\textcolor{red}{\\textbf{"
  postemph <- "}}"
  stcodon <- (stdcode == sapply(lapply(cdorder, s2c), translate,
    numcode = cd))
  pre <- ifelse(stcodon, "", preemph)
  post <- ifelse(stcodon, "", postemph)
  tablecode(numcode = cd, latexfile = Tfile, size = "small",
    preaa = pre, postaa = post)
  cat(paste("\\input", Tfile, "}"), sep = ""), sep = "\n")
}

```

### C.3 Session Informations

This part was compiled under the following  environment:

- R version 2.4.1 (2006-12-18), i386-apple-darwin8.8.1
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	<b>TGA</b>	<b>Trp</b>
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
<b>ATA</b>	<b>Met</b>	ACA	Thr	AAA	Lys	<b>AGA</b>	<b>Stop</b>
ATG	Met	ACG	Thr	AAG	Lys	<b>AGG</b>	<b>Stop</b>
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

Table C.2: Genetic code number 2: vertebrate.mitochondrial.

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	<b>TGA</b>	<b>Trp</b>
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
<b>CTT</b>	<b>Thr</b>	CCT	Pro	CAT	His	CGT	Arg
<b>CTC</b>	<b>Thr</b>	CCC	Pro	CAC	His	CGC	Arg
<b>CTA</b>	<b>Thr</b>	CCA	Pro	CAA	Gln	CGA	Arg
<b>CTG</b>	<b>Thr</b>	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
<b>ATA</b>	<b>Met</b>	ACA	Thr	AAA	Lys	<b>AGA</b>	<b>Arg</b>
ATG	Met	ACG	Thr	AAG	Lys	<b>AGG</b>	<b>Arg</b>
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

Table C.3: Genetic code number 3: yeast.mitochondrial.

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	<b>TGA</b>	<b>Trp</b>
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table C.4: Genetic code number 4: protozoan.mitochondrial+mycoplasma.

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	<b>TGA</b>	<b>Trp</b>
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
<b>ATA</b>	<b>Met</b>	ACA	Thr	AAA	Lys	<b>AGA</b>	<b>Ser</b>
ATG	Met	ACG	Thr	AAG	Lys	<b>AGG</b>	<b>Ser</b>
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table C.5: Genetic code number 5: invertebrate.mitochondrial.

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	<b>TAA</b>	<b>Gln</b>	TGA	Stop
TTG	Leu	TCG	Ser	<b>TAG</b>	<b>Gln</b>	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

Table C.6: Genetic code number 6: ciliate+dasycladacean.

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	<b>TGA</b>	<b>Trp</b>
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	<b>AAA</b>	<b>Asn</b>	<b>AGA</b>	<b>Ser</b>
ATG	Met	ACG	Thr	AAG	Lys	<b>AGG</b>	<b>Ser</b>
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

Table C.7: Genetic code number 9: echinoderm+flatworm.mitochondrial.

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	<b>TGA</b>	<b>Cys</b>
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table C.8: Genetic code number 10: euplotid.

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	TGA	Stop
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table C.9: Genetic code number 11: bacterial+plantplastid.

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	TGA	Stop
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
<b>CTG</b>	<b>Ser</b>	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

Table C.10: Genetic code number 12: alternativeyeast.

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	<b>TGA</b>	<b>Trp</b>
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
<b>ATA</b>	<b>Met</b>	ACA	Thr	AAA	Lys	<b>AGA</b>	<b>Gly</b>
ATG	Met	ACG	Thr	AAG	Lys	<b>AGG</b>	<b>Gly</b>
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

Table C.11: Genetic code number 13: ascidian.mitochondrial.

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	<b>TAA</b>	<b>Tyr</b>	<b>TGA</b>	<b>Trp</b>
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	<b>AAA</b>	<b>Asn</b>	<b>AGA</b>	<b>Ser</b>
ATG	Met	ACG	Thr	AAG	Lys	<b>AGG</b>	<b>Ser</b>
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table C.12: Genetic code number 14: alternativeflatworm.mitochondrial.

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	TGA	Stop
TTG	Leu	TCG	Ser	<b>TAG</b>	<b>Gln</b>	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	<b>AAA</b>	Lys	<b>AGA</b>	<b>Arg</b>
ATG	Met	ACG	Thr	AAG	Lys	<b>AGG</b>	<b>Arg</b>
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table C.13: Genetic code number 15: blepharism.

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	TGA	Stop
TTG	Leu	TCG	Ser	<b>TAG</b>	<b>Leu</b>	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table C.14: Genetic code number 16: chlorophycean.mitochondrial.

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	TCA	Ser	TAA	Stop	<b>TGA</b>	<b>Trp</b>
TTG	Leu	TCG	Ser	TAG	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
<b>ATA</b>	<b>Met</b>	ACA	Thr	<b>AAA</b>	<b>Asn</b>	<b>AGA</b>	<b>Ser</b>
ATG	Met	ACG	Thr	AAG	Lys	AGG	Ser
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table C.15: Genetic code number 21: trematode.mitochondrial.

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
TTA	Leu	<b>TCA</b>	<b>Stop</b>	TAA	Stop	TGA	Stop
TTG	Leu	TCG	Ser	<b>TAG</b>	<b>Leu</b>	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table C.16: Genetic code number 22: *scenedesmus.mitochondrial*.

---

TTT	Phe	TCT	Ser	TAT	Tyr	TGT	Cys
TTC	Phe	TCC	Ser	TAC	Tyr	TGC	Cys
<b>TTA</b>	<b>Stop</b>	<b>TCA</b>	<b>Ser</b>	TAA	Stop	TGA	Stop
TTG	Leu	TCG	Ser	<b>TAG</b>	Stop	TGG	Trp
CTT	Leu	CCT	Pro	CAT	His	CGT	Arg
CTC	Leu	CCC	Pro	CAC	His	CGC	Arg
CTA	Leu	CCA	Pro	CAA	Gln	CGA	Arg
CTG	Leu	CCG	Pro	CAG	Gln	CGG	Arg
ATT	Ile	ACT	Thr	AAT	Asn	AGT	Ser
ATC	Ile	ACC	Thr	AAC	Asn	AGC	Ser
ATA	Ile	ACA	Thr	AAA	Lys	AGA	Arg
ATG	Met	ACG	Thr	AAG	Lys	AGG	Arg
GTT	Val	GCT	Ala	GAT	Asp	GGT	Gly
GTC	Val	GCC	Ala	GAC	Asp	GGC	Gly
GTA	Val	GCA	Ala	GAA	Glu	GGA	Gly
GTG	Val	GCG	Ala	GAG	Glu	GGG	Gly

---

Table C.17: Genetic code number 23: *hraustochytrium.mitochondria*.

- Other packages: MASS 7.2-30, ade4 1.4-2, ape 1.9-3, gee 4.13-12, lattice 0.14-16, nlme 3.1-78, seqinr 1.0-7, xtable 1.4-3



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## APPENDIX D

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### Release notes

Lobry, J.R. Necșulea, A. Palmeira, L. Penel, S.

#### D.1 release 1.0-7

- A new *experimental* function `extractseqs()` to download sequences thru zlib compressed sockets from an ACNUC server is released. Preliminary tests suggest that working with about 100,000 CDS is possible with a home ADSL connection. See the manual for some `system.time()` examples.
- As pointed by e-mail on 16 Nov 2006 by Emmanuel Prestat the URL used in `dia.bactgensize()` was no more available, this has been fixed in the current version.
- As pointed by e-mail on 16 Nov 2006 by Guy Perrière, the function `oriloc()` was no more compatible with glimmer<sup>1</sup> 3.0 outputs. The function has gained a new argument `glimmer.version` defaulting to 3, but the value 2 is still functional for backward compatibility with old glimmer outputs.
- As pointed by e-mail on 24 Oct 2006 by Lionel Guy (<http://pbil.univ-lyon1.fr/seqinr/seqinrhtmlannuel/03/0089.html>) there was no default value for the `as.string` argument in the `getSequence.SeqFastadna()`. A default `FALSE` value is now present for backward compatibility with older code.
- New utility vectorized function `stresc()` to escape L<sup>A</sup>T<sub>E</sub>X special characters present in a string.
- New low level function `readsmj()` available.
- A new function `readfirstrec()` to get the record count of the specified ACNUC index file is now available.

---

<sup>1</sup> Glimmer is a program to predict coding sequences in microbial genomes [54, 6].

- Function `getType()` called without arguments will now use the default ACNUC database to return available subsequence types.
- Function `read.alignment()` now also accepts `file` in addition to `File` as argument.
- A new function `rearranged.oriloc()` is available. This method, based on `oriloc()`, can be used to detect the effect of the replication mechanism on DNA base composition asymmetry, in prokaryotic chromosomes.
- New function `extract.breakpoints()`, used to extract breakpoints in rearranged nucleotide skews. This function uses the `segmented` package to define the position of the breakpoints.
- New function `draw.rearranged.oriloc()` available, to plot nucleotide skews on artificially rearranged prokaryotic chromosomes.
- New function `gbk2g2.euk()` available. Similarly to `gbk2g2()`, this function extracts the coding sequence annotations from a GenBank format file. This function is specifically designed for eukaryotic sequences, *i.e.* with introns. The output file will contain the coordinates of the exons, along with the name of the CDS to which they belong.
- After an e-mail by Marcelo Bertalan on 26 Mar 2007, a bug in `oriloc()` when the `gbk` argument was `NULL` was found and fixed by Anamaria Necșulea.
- Functions `translate()` and `getTrans()` have gained a new argument `NAstring` to represent untranslatable amino-acids, defaulting to character "X".
- There was a typo for the total number of printed bases in the ACNUC books [13, 14] : 474,439 should be 526,506.
- Function `invers()` has been deleted.
- Functions `translate()`, `getTrans()` and `comp()` have gained a new argument `ambiguous` defaulting to `FALSE` allowing to handle ambiguous bases. If `TRUE`, ambiguous bases are taken into account so that for instance GGN is translated to Gly in the standard genetic code.
- New function `amb()` to return the list of nucleotide matching a given IUPAC nucleotide symbol.

## D.2 release 1.0-6

Release 1.0-6 is a minor release to fix a problem found and solved by Kurt Hornik (namely a change from `SET_ELEMENT` to `SET_STRING_elt` in C code for `s2c()` in file `util.c`). The few changes are as follows.

- More typographical option for the output L<sup>A</sup>T<sub>E</sub>X table of `tablecode()` are now available to outline deviations from the standard genetic code (see example in the appendix "genetic codes" of the manual).

- A new dataset `aaindex` extracted from the aaindex database [30, 61, 44] is now available. It contains a list of 544 physicochemical and biological properties for the 20 amino-acids
- The default value for argument `dia` is now `FALSE` in function `tablecode()`.
- The example code for `data(chargaff)` has been changed.

## D.3 release 1.0-5

- A new function `dotPlot()` is now available.
- A new function `crelistfromclientdata()` is now available to create a list on the server from a local file of sequence names, sequence accession numbers, species names, or keywords names.
- A new function `pmw()` to compute the molecular weight of a protein is now available.
- A new function `reverse.align()` contributed by Anamaria Necșulea is now available to align CDS at the protein level and then reverse translate this at the nucleic acid level from a `clustalw` output. This can be done on the fly if `clustalw` is available on your platform.
- An undocumented behavior was reported by Guy Perrière for `uco()` when computing RSCU on sequences where an amino-acid is missing. There is now a new argument `NA.rscu` that allows the user to force the missing values to his favorite magic value.
- There was a bug in `read.fasta()`: some sequence names were truncated, this is now fixed (thanks to Marcus G. Daniels for pointing this). In order to be more consistent with standard functions such as `read.table()` or `scan()`, the file argument starts now with a lower case letter (`file`) in function `read.fasta()`, but the old-style `File` is still functional for forward-compatibility. There is a new logical argument in `read.fasta()` named `as.string` to allow sequences to be returned as strings instead of vector of single characters. The automatic conversion of DNA sequences into lower case letters can now be disabled with the new logical argument `forceDNAtolower`. It is also possible to disable the automatic attributes settings with the new logical argument `set.attributes`.
- A new function `write.fasta()` is now available.
- The function `kaks()` now forces character in sequences to upper case. This default behavior can be neutralized in order to save time by setting the argument `forceUpperCase` to `FALSE`.

## D.4 release 1.0-4

- The scaling factor  $n_{\bullet\bullet}$  was missing in equation 5.3.

- The files `louse.fasta`, `louse.names`, `gopher.fasta`, `gopher.names` and `ortho.fasta` that were used for examples in the previous version of this document are no more downloaded from the internet since they are now distributed in the `sequences/` folder of the package.
- An example of synonymous and non synonymous codon usage analysis was added to the vignette along with two toy data sets (`toyaa` and `toycodon`).
- A FAQ section was added to the vignette.
- A bug in `getAnnot()` when the number of lines was zero is now fixed.
- There is now a new argument, `latexfile`, in `tablecode()` to export genetic codes tables in a L<sup>A</sup>T<sub>E</sub>X document, for instance table 1.2 and table 1.3 here.
- There is now a new argument, `freq`, in `count()` to compute word frequencies instead of counts.
- Function `splitseq()` has been entirely rewritten to improve speed.
- Functions computing the G+C content: `GC()`, `GC1()`, `GC2()`, `GC3()` were rewritten to improve speed, and their document files were merged to facilitate usage.
- The following new functions have been added:
  - `syncodons()` returns all synonymous codons for a given codon. Argument `numcode` specifies the desired genetic code.
  - `ucoweight()` returns codon usage bias on a sequence as the number of synonymous codons present in the sequence for each amino acid.
  - `synsequence()` generates a random coding sequence which is synonymous to a given sequence and has a chosen codon usage bias.
  - `permutation()` generates a new sequence from a given sequence, while maintaining some constraints from the given sequence such as nucleotide frequency, codon usage bias, ...
  - `rho()` computes the rho statistic on dinucleotides as defined in [29].
  - `zscore()` computes the zscore statistic on dinucleotides as defined in [46].
- Two datasets (`dinucl` and `prochlo`) were added to illustrate these new functions.

## D.5 release 1.0-3

- The new package maintainer is Dr. Simon Penel, PhD, who has now a fixed position in the laboratory that issued **seqinR** (`penel@biomserv.univ-lyon1.fr`). Delphine Charif was successful too to get a fixed position in the same lab, with now a different research task (but who knows?). Thanks to the close vicinity of our pioneering maintainers the transition was sweet. The DESCRIPTION file of the **seqinR** package has been updated to take this into account.

- The reference paper for the package is now *in press*. We do not have the full reference for now, you may use `citation("seqinr")` to check if it is complete now:

```
citation("seqinr")
```

To cite seqinR in publications use:

in the body of the text (J.R. Lobry, personal communication), or  
wait for the exact complete reference.

A BibTeX entry for LaTeX users is

```
@incollection{,
  author = {D. Charif and J.R. Lobry},
  title = {SeqinR 1.0-2: a contributed package to the R project for statistical computing devoted to biological sequence analysis},
  booktitle = {Structural approaches to sequence evolution: Molecules, networks, populations},
  year = {2006},
  editor = {U. Bastolla, M. Porto, H.E. Roman and M. Vendruscolo},
  volume = {NA},
  series = {Biological and Medical Physics, Biomedical Engineering},
  pages = {NA},
  address = {New York},
  month = {NA},
  organization = {NA},
  publisher = {Springer Verlag},
  note = {in press},
}
```

Note that the original article and updates are available in the  
./.../.../.../seqinr.Rcheck/seqinr/doc/ folder in PDF format

- There was a bug when sending a `gfrag` request to the server for long (Mb range) sequences. The length argument was converted to scientific notations that are not understood by the server. This is now corrected and should work up to the Gb scale.
- The `query()` function has been improved by de-looping list element info request, there are now download at once which is much more efficient. For example, a query from a researcher-home ADSL connection with a list with about 1000 elements was 60 seconds and is now only 4 seconds (*i.e.* 15 times faster now).
- A new parameter `virtual` has been added to `query()` so that long lists can stay on the server without trying to download them automatically. A query like `query(s$socket, "allcds", "t=cds", virtual = TRUE)` is now possible.
- Relevant genetic codes and frames are now automatically propagated.
- SeqinR** sends now its name and version number to the server.
- Strict control on ambiguous DNA base alphabet has been relaxed.
- Default value for parameter `invisible` of function `query()` is now `TRUE`.

## D.6 Session Informations

This part was compiled under the following  environment:

- R version 2.4.1 (2006-12-18), i386-apple-darwin8.8.1

- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: MASS 7.2-30, ade4 1.4-2, ape 1.9-3, gee 4.13-12, lattice 0.14-16, nlme 3.1-78, seqinr 1.0-7, xtable 1.4-3

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## APPENDIX E

---

### Informations about databases available at pbil

Lobry, J.R.

#### E.1 Introduction

This section was compiled on April 15, 2007. The list of available database at pbil (<http://pbil.univ-lyon1.fr/>) was:

```
banknames <- choosebank()
banknames

[1] "genbank"      "embl"          "emblwgs"       "swissprot"
[5] "ensembl"     "ensembl34"    "refseq"        "nrsub"
[9] "nbrf"         "hobacnucl"    "hobacprot"    "hovernucl"
[13] "hoverprot"   "hogennucl"    "hogenprot"    "hoverclnu"
[17] "hoverclpr"   "homolensprot" "homolensnucl" "greview"
[21] "emglib"       "HAMAPnucl"    "HAMAPprot"    "hoppigen"
[25] "nurebnucl"   "nurebprot"    "taxobacgen"
```

This L<sup>A</sup>T<sub>E</sub>X file was automatically generated by the following  code:

```
for (b in banknames) {
  cat(paste("\\\\section{", b, "}}"), sep = "\\n")
  choosebank(b)
  bankdetails <- sapply(banknameSocket$details, stresc,
    USE.NAMES = FALSE)
  cat("\\\\subsection{Bank details}", sep = "\\n")
  cat(bankdetails, sep = "\\\\\\\\\\n")
  cat("\\n")
  cat("\\\\subsection{Type names}", sep = "\\n")
  types <- getType()
  if (is.null(nrow(types))) {
    cat("There are no subsequence type in this database",
        sep = "\\n")
  }
  else {
    cat("\\\\noindent\\\\begin{tabular}{ll}", sep = "\\n")
    cat("\\\\hline \\\\hline", sep = "\\n")
    cat("name & description\\\\\\\\", sep = "\\n")
```

```

cat("\\\\hline", sep = "\\n")
for (i in 1:nrow(types)) cat(paste(strec(types[i,
  "sname"])), "&", strec(types[i, "libel"]), "\\\\\\"),
  sep = "\\n")
cat("\\\\hline \\\\hline", sep = "\\n")
cat("\\\\end{tabular}", sep = "\\n")
cat("\\n")
}
closebank()
}

```

## E.2 genbank

### E.2.1 Bank details

GenBank Rel. 158 (15 February 2007) Last Updated: Mar 2, 2007  
 71,832,523,193 bases; 67,564,553 sequences; 3,798,216 subseqs; 454,417 refers.  
 Software by M. Gouy, Lab. Biometrie et Biologie Evolutive, Universite Lyon I

### E.2.2 Type names

name	description
CDS	.PE protein coding region
LOCUS	sequenced DNA fragment
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR mature ribosomal RNA
SCRNA	.SC small cytoplasmic RNA
SNRNA	.SN small nuclear RNA
TRNA	.TR mature transfer RNA

## E.3 embl

### E.3.1 Bank details

EMBL Library Release 89 (December 2006) Last Updated: Mar 2, 2007  
 72,211,385,066 bases; 67,564,144 sequences; 3,798,254 subseqs; 451,586 refers.  
 Software by M. Gouy, Laboratoire de biometrie, Universite Lyon I

### E.3.2 Type names

name	description
CDS	.PE protein coding region
ID	Locus entry
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR Ribosomal RNA coding gene
SCRNA	.SC small cytoplasmic RNA
SNRNA	.SN small nuclear RNA
TRNA	.TR Transfer RNA coding gene

## E.4 emblwgs

### E.4.1 Bank details

EMBL Whole Genome Shotgun sequences Release 89 (December 2006)  
 81,521,656,204 bases; 18,535,353 sequences; 1,821,700 subseqs; 269 refers.  
 Software by M. Gouy & M. Jacobzone, Laboratoire de biometrie, Universite Lyon I

### E.4.2 Type names

name	description
CDS	.PE protein coding region
ID	EMBL sequence data library entry
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.5 swissprot

### E.5.1 Bank details

UniProt Rel. 9 (SWISS-PROT 51 + TrEMBL 34): Last Updated: Feb 19, 2007  
 1,312,031,657 amino acids; 4,003,765 sequences; 264,298 references.  
 Non-redundant compilation of SWISS-PROT + TrEMBL  
 Software by M. Gouy & L. Duret, Laboratoire de biometrie, Universite Lyon I.

### E.5.2 Type names

There are no subsequence type in this database

## E.6 ensembl

### E.6.1 Bank details

Ensembl databases release 41  
 Espece Release/#CDS(1)/STOP(2)/N(3)/miss(4)  
 Aedes aegypti 41\_1a 11360/0/2/0 (0%/0%/0%)  
 Anopheles gambiae 41\_3d 13510/0/31/0 (0%/0%/0%)  
 Apis mellifera 38\_2d 27755/1/269/0 (0%/0%/0%)  
 Bos taurus 41\_2 32556/7/620/12 (0%/1%/0%)  
 Caenorhabditis elegans 41\_160 25218/1/0/0 (0%/0%/0%)  
 Canis familiaris 41\_1j 29813/0/0/0 (0%/0%/0%)  
 Caenorhabditis briggsae 25 14712/0/23/1 (0%/0%/0%)  
 Ciona intestinalis 41\_2c 20000/0/128/0 (0%/0%/0%)  
 Ciona savignyi 41\_2b 20150/1/27/0 (0%/0%/0%)  
 Danio rerio 41\_6b 36065/5/361/0 (0%/1%/0%)  
 Dasypus novemcinctus 40\_1 13567/12/8857/0 (0%/65%/0%)

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Drosophila melanogaster 41\_43 19577/33/1/0 (0%/0%/0%)  
 Echinops telfairi 40\_1 14309/8/9348/0 (0%/65%/0%)  
 Gallus gallus 41\_1p 20667/13/455/0 (0%/2%/0%)  
 Gasterosteus aculeatus 41\_1a 27181/13/138/0 (0%/0%/0%)  
 Homo sapiens 41\_36c 47004/41/6/0 (0%/0%/0%)  
 Loxodonta africana 40\_1 14366/10/9618/0 (0%/66%/0%)  
 Macaca mulatta 41\_10a 36446/14/491/0 (0%/1%/0%)  
 Monodelphis domestica 41\_3a 30358/0/80/0 (0%/0%/0%)  
 Mus musculus 41\_36b 29026/34/2/0 (0%/0%/0%)  
 Oryctolagus cuniculus 41\_1a 13705/4/8615/0 (0%/62%/0%)  
 Oryzias latipes 41\_1 25880/0/546/0 (0%/2%/0%)  
 Pan troglodytes 41\_21 32667/4/739/0 (0%/2%/0%)  
 Rattus norvegicus 41\_34k 32996/34/686/0 (0%/2%/0%)  
 Saccharomyces cerevisiae 41\_1d 4767/2/0/0 (0%/0%/0%)

### E.6.2 Type names

name	description
3'INT	.3I 3'intron
3'NCR	.3F 3'-non coding region
5'INT	.5I 5'intron
5'NCR	.5F 5'-non coding region
CDS	.PE protein coding region
ID	EMBL sequence data library entry
INT_INT	.IN internal intron
MISC_RNA	.RN other structural RNA coding region
MRNA	.RN mRNA
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.7 ensembl34

### E.7.1 Bank details

Ensembl databases release 34  
 Espece #CDS(1)/STOP(2)/N(3)/miss(4)  
 Apis mellifera 27736/1/269 (0%/0%/0%)  
 Caenorhabditis briggsae 14712/0/23 (0%/0%/0%)  
 Caenorhabditis elegans 25797/1/0 (0%/0%/0%)  
 Gallus gallus 28392/20/298 (0%/1%/0%)  
 Pan troglodytes 39538/6129/770 (15%/1%/0%)  
 Ciona intestinalis 21574/0/58 (0%/0%/0%)  
 Bos taurus 32647/7/617 (0%/1%/0%)  
 Canis familiaris 29998/0/0 (0%/0%/1%)  
 Drosophila melanogaster 19350/18/1 (0%/0%/0%)  
 Fugu rubripes 22099/0/283 (0%/1%/0%)  
 Homo sapiens 36919/48/24 (0%/0%/2%)

Macaca mulatta 31370/94/8360 (0%/26%/0%)  
 Anopheles gambiae 15799/0/19 (0%/0%/0%)  
 Mus musculus 35075/36/60 (0%/0%/1%)  
 Monodelphis domestica 13249/0/59 (0%/0%/0%)  
 Rattus norvegicus 32241/25/607 (0%/1%/2%)  
 Tetraodon nigroviridis 16275/1/233 (0%/1%/0%)  
 Xenopus tropicalis 52684/1/906 (0%/1%/0%)  
 Saccharomyces cerevisiae 6680/22/0 (0%/0%/0%)  
 Danio rerio 32109/0/281 (0%/0%/0%)  
 1:# of CDS;2:CDS with internal stop;3:CDS with undetermined codon;4:missing CDS  
 warning : cds located on contigs were removed  
 29,605,509,937 bases; 368,619 sequences; 5,302,323 subseqs; 0 refers.  
 Software by M. Gouy & M. Jacobzone, Laboratoire de biometrie, Universite Lyon I

### E.7.2 Type names

name	description
3'INT	.3I 3'intron
3'NCR	.3F 3'-non coding region
5'INT	.5I 5'intron
5'NCR	.5F 5'-non coding region
CDS	.PE protein coding region
ID	EMBL sequence data library entry
INT_INT	.IN internal intron
MISC_RNA	.RN other structural RNA coding region
MRNA	.RN mRNA
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.8 refseq

### E.8.1 Bank details

RefSeq 15.0 (1 January 2006) Last Updated: Jan 23, 2006  
 1,055,245,496 bases; 625,928 sequences; 254,162 subseqs; 205,831 refers.  
 Software by M. Gouy & M. Jacobzone, Laboratoire de biometrie, Universite Lyon I

### E.8.2 Type names

name	description
3'INT	.3I 3'intron
3'NCR	.3F 3'-non coding region
5'INT	.5I 5'intron
5'NCR	.5F 5'-non coding region
CDS	.PE protein coding region
INT_INT	.IN internal intron
LOCUS	sequenced DNA fragment
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.9 nrsub

### E.9.1 Bank details

NRSub database release 10.1 (December 1997)  
 Bacillus subtilis complete genome  
 Sequence data taken from the SubtiList database  
 Institut Pasteur - Unite de Regulation de l'Expression Genetique  
 Extra annotations provided by G. Perriere  
 Laboratoire BGBP - Universite Claude Bernard, Lyon 1

### E.9.2 Type names

name	description
CDS	.PE protein coding region
ID	EMBL sequence data library entry
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.10 nbrf

### E.10.1 Bank details

NBRF-PIR Release 80 (December 31, 2004)  
 96,216,763 aminoacids; 283,416 sequences; 60,847 references.  
 Retrieval software by M. Gouy, Laboratoire de Biometrie, Universite de Lyon

### E.10.2 Type names

There are no subsequence type in this database

## E.11 hobacnucl

### E.11.1 Bank details

HOBACGEN - genomic data - Release 10 (February 12 2002)  
 432,023,804 bases; 168,814 sequences; 293,669 subseqs; 52,735 references.

Bacteria + Archaea + *Saccharomyces cerevisiae*  
 Genomic data from EMBL Release 69 (December 2001)

### E.11.2 Type names

name	description
CDS	.PE protein coding region
ID	EMBL sequence data library entry
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.12 hobacprot

### E.12.1 Bank details

HOBACGEN - protein data - Release 10 (February 12 2002)  
 79,755,852 amino acids; 260,025 sequences; 37,383 references.

Bacteria + Archaea + *Saccharomyces cerevisiae*  
 Protein data from SWISS-PROT 40 + TrEMBL 19 + TrEMBL\_NEW: January  
 25, 2002

Software: M. Gouy & M. Jacobzone  
 Data maintenance: L. Duret & G. Perriere

Laboratoire de Biometrie et Biologie Evolutive  
 UMR CNRS 5558, Universite Claude Bernard - Lyon 1  
 43, bd du 11 Novembre 1918 F-69622 Villeurbanne Cedex

### E.12.2 Type names

There are no subsequence type in this database

## E.13 hovernucl

### E.13.1 Bank details

HOVERGEN - genomic data - Release 47 (Jul 27 2005) Last Updated: Jul 27, 2005  
 1,711,891,620 bases; 389,601 sequences; 1,204,187 subseqs; 106,597 refers.

Vertebrate (chordata)  
 Genomic data from EMBL Release 83 (June 2005)

Retrieval software by M. Gouy & M. Jacobzone, Lab. de Biometrie, UCB Lyon.  
 Data maintenance: L. Duret & S. Penel

### E.13.2 Type names

name	description
3'INT	.3I 3'intron
3'NCR	.3F 3'-non coding region
5'INT	.5I 5'intron
5'NCR	.5F 5'-non coding region
CDS	.PE protein coding region
ID	EMBL sequence data library entry
INT_INT	.IN internal intron
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.14 hoverprot

### E.14.1 Bank details

HOVERGEN - protein data - Release 47 (Jul 5 2005) Last Updated: Jul 5, 2005  
 103,023,967 amino acids; 294,410 sequences; 100,097 references.

Vertebrate (chordata)  
 Protein data from SWISS-PROT Rel. 47 + TrEMBL Rel. 30 + TrEMBL\_NEW:  
 Jul 5, 2005

Software: M. Gouy & M. Jacobzone  
 Data maintenance: L. Duret & S. Penel

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### E.14.2 Type names

There are no subsequence type in this database

## E.15 hogennucl

### E.15.1 Bank details

HOGENOM - genomic data - Release 03 (Oct 14 2005) Last Updated: Nov 7, 2005

2,538,433,251 bases; 227,950 sequences; 4,136,134 subseqs; 82,281 refers.

Fully Sequenced Organisms

Protein data from <http://www.ebi.ac.uk/proteome/> (August, 2005)

Genomic data from GenomeReview (June 2005)

and EMBL (June 2005)

( 263 fully sequenced organisms)

Retrieval software by M. Gouy & M. Jacobzone, Lab. de Biometrie, UCB Lyon.  
Data maintenance: L. Duret & S. Penel

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### E.15.2 Type names

name	description
ID	EMBL sequence data library entry
CDS	.PE protein coding region
TRNA	.TR transfer RNA coding region
RRNA	.RR ribosomal RNA coding region
MISC_RNA	.RN other structural RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
3'INT	.3I 3'intron
3'NCR	.3F 3'-non coding region
5'INT	.5I 5'intron
5'NCR	.5F 5'-non coding region
INT_INT	.IN internal intron

## E.16 hogenprot

### E.16.1 Bank details

HOGENOM - protein data - Release 03 (Oct 14 2005) Last Updated: Mar 10, 2006

339,891,443 amino acids; 950,216 sequences; 92,805 references.

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### Fully Sequenced Organisms

Protein data from <http://www.ebi.ac.uk/proteome/> (August 2005)  
( 263 fully sequenced organisms)

Retrieval software by M. Gouy & M. Jacobzone, Lab. de Biometrie, UCB Lyon.  
Data maintenance: L. Duret & S. Penel

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### E.16.2 Type names

There are no subsequence type in this database

## E.17 hoverclnu

### E.17.1 Bank details

HOVERGEN CLEAN - genomic data - Release 46 (Jun 10 2004)  
894,369,756 bases; 312,987 sequences; 796,415 subseqs; 99,342 refers.

Vertebrate (chordata)  
Genomic data from EMBL Release 78 (March 2004)

Retrieval software by M. Gouy & M. Jacobzone, Lab. de Biometrie, UCB Lyon.  
Data maintenance: L. Duret & S. Penel

### E.17.2 Type names

name	description
3'INT	.3I 3'intron
3'NCR	.3F 3'-non coding region
5'INT	.5I 5'intron
5'NCR	.5F 5'-non coding region
CDS	.PE protein coding region
ID	EMBL sequence data library entry
INT_INT	.IN internal intron
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.18 hoverclpr

### E.18.1 Bank details

HOVERGEN CLEAN - protein data - Release 46 (Jun 10 2004)  
75,885,664 amino acids; 219,552 sequences; 89,885 references.

Vertebrate (chordata)

Protein data from SWISS-PROT Rel. 43 + TrEMBL Rel. 26 + TrEMBL\_NEW:  
May 17, 2004

Software: M. Gouy & M. Jacobzone  
Data maintenance: L. Duret & S. Penel

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### E.18.2 Type names

There are no subsequence type in this database

## E.19 homolensprot

### E.19.1 Bank details

HOMOLENS 3 - Homologous genes from Ensembl Last Updated: Jan 19, 2007  
224,528,520 amino acids; 474,339 sequences; 0 references.

Ensembl 41 Organisms Translated CDS  
Aedes aegypti 41\_1a 11360/0/2/0 (0%/0%/0%)  
Anopheles gambiae 41\_3d 13510/0/31/0 (0%/0%/0%)  
Apis mellifera 38\_2d 27755/1/269/0 (0%/0%/0%)  
Bos taurus 41\_2 32556/7/620/12 (0%/1%/0%)  
Caenorhabditis elegans 41\_160 25218/1/0/0 (0%/0%/0%)  
Canis familiaris 41\_1j 29813/0/0/0 (0%/0%/0%)  
Caenorhabditis briggsae 25 14712/0/23/1 (0%/0%/0%)  
Ciona intestinalis 41\_2c 20000/0/128/0 (0%/0%/0%)  
Ciona savignyi 41\_2b 20150/1/27/0 (0%/0%/0%)  
Danio rerio 41\_6b 36065/5/361/0 (0%/1%/0%)  
Dasypus novemcinctus 40\_1 13567/12/8857/0 (0%/65%/0%)  
Drosophila melanogaster 41\_43 19577/33/1/0 (0%/0%/0%)  
Echinops telfairi 40\_1 14309/8/9348/0 (0%/65%/0%)  
Gallus gallus 41\_1p 20667/13/455/0 (0%/2%/0%)  
Gasterosteus aculeatus 41\_1a 27181/13/138/0 (0%/0%/0%)  
Homo sapiens 41\_36c 47004/41/6/0 (0%/0%/0%)  
Loxodonta africana 40\_1 14366/10/9618/0 (0%/66%/0%)  
Macaca mulatta 41\_10a 36446/14/491/0 (0%/1%/0%)  
Monodelphis domestica 41\_3a 30358/0/80/0 (0%/0%/0%)

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Mus musculus 41\_36b 29026/34/2/0 (0%/0%/0%)  
Oryctolagus cuniculus 41\_1a 13705/4/8615/0 (0%/62%/0%)  
Oryzias latipes 41\_1 25880/0/546/0 (0%/2%/0%)  
Pan troglodytes 41\_21 32667/4/739/0 (0%/2%/0%)  
Rattus norvegicus 41\_34k 32996/34/686/0 (0%/2%/0%)  
Saccharomyces cerevisiae 41\_1d 4767/2/0/0 (0%/0%/0%)  
Takifugu rubripes 41\_4c 22102/0/283/0 (0%/1%/0%)  
Tetraodon nigroviridis 41\_1g 15841/1/225/0 (0%/1%/0%)  
Xenopus tropicalis 41\_41b 28324/0/626/0 (0%/2%/0%)

Software: M. Gouy & M. Jacobzone

Data maintenance: L. Duret & S. Penel

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### E.19.2 Type names

There are no subsequence type in this database

## E.20 homolensnucl

### E.20.1 Bank details

HOMOLENS 3 Homologous genes from Ensembl 41 Last Updated: Jan 19, 2007  
32,635,729,329 bases; 81,903 sequences; 5,717,782 subseqs; 0 refers.

Aedes aegypti 41\_1a 11360/0/2/0 (0%/0%/0%)  
Anopheles gambiae 41\_3d 13510/0/31/0 (0%/0%/0%)  
Apis mellifera 38\_2d 27755/1/269/0 (0%/0%/0%)  
Bos taurus 41\_2 32556/7/620/12 (0%/1%/0%)  
Caenorhabditis elegans 41\_160 25218/1/0/0 (0%/0%/0%)  
Canis familiaris 41\_1j 29813/0/0/0 (0%/0%/0%)  
Caenorhabditis briggsae 25 14712/0/23/1 (0%/0%/0%)  
Ciona intestinalis 41\_2c 20000/0/128/0 (0%/0%/0%)  
Ciona savignyi 41\_2b 20150/1/27/0 (0%/0%/0%)  
Danio rerio 41\_6b 36065/5/361/0 (0%/1%/0%)  
Dasypus novemcinctus 40\_1 13567/12/8857/0 (0%/65%/0%)  
Drosophila melanogaster 41\_43 19577/33/1/0 (0%/0%/0%)  
Echinops telfairi 40\_1 14309/8/9348/0 (0%/65%/0%)  
Gallus gallus 41\_1p 20667/13/455/0 (0%/2%/0%)  
Gasterosteus aculeatus 41\_1a 27181/13/138/0 (0%/0%/0%)  
Homo sapiens 41\_36c 47004/41/6/0 (0%/0%/0%)  
Loxodonta africana 40\_1 14366/10/9618/0 (0%/66%/0%)  
Macaca mulatta 41\_10a 36446/14/491/0 (0%/1%/0%)  
Monodelphis domestica 41\_3a 30358/0/80/0 (0%/0%/0%)

Mus musculus 41\_36b 29026/34/2/0 (0%/0%/0%)  
 Oryctolagus cuniculus 41\_1a 13705/4/8615/0 (0%/62%/0%)  
 Oryzias latipes 41\_1 25880/0/546/0 (0%/2%/0%)  
 Pan troglodytes 41\_21 32667/4/739/0 (0%/2%/0%)  
 Rattus norvegicus 41\_34k 32996/34/686/0 (0%/2%/0%)  
 Saccharomyces cerevisiae 41\_1d 4767/2/0/0 (0%/0%/0%)  
 Takifugu rubripes 41\_4c 22102/0/283/0 (0%/1%/0%)  
 Tetraodon nigroviridis 41\_1g 15841/1/225/0 (0%/1%/0%)  
 Xenopus tropicalis 41\_41b 28324/0/626/0 (0%/2%/0%)

Software by M. Gouy & M. Jacobzone, Laboratoire de biometrie, Universite Lyon I

## E.20.2 Type names

name	description
3'INT	.3I 3'intron
3'NCR	.3F 3'-non coding region
5'INT	.5I 5'intron
5'NCR	.5F 5'-non coding region
CDS	.PE protein coding region
ID	EMBL sequence data library entry
INT_INT	.IN internal intron
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.21 greview

### E.21.1 Bank details

EBI Genome Reviews. Acnuc Release 7. Last Updated: Feb 26, 2007

1,454,629,225 bases; 757 sequences; 2,619,601 subseqs; 346 refers.

385 organisms

Software by M. Gouy & M. Jacobzone, Laboratoire de biometrie, Universite Lyon I

### E.21.2 Type names

name	description
3'INT	.3I 3'intron
3'NCR	.3F 3'-non coding region
5'INT	.5I 5'intron
5'NCR	.5F 5'-non coding region
CDS	.PE protein coding region
ID	EMBL sequence data library entry
INT_INT	.IN internal intron
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.22 emplib

### E.22.1 Bank details

EMGLib Release 5 (December 9, 2003)  
 434,648,385 bases; 174 sequences; 413,521 subseqs; 169 refers.  
 Data compiled from various sources by Guy Perriere

### E.22.2 Type names

name	description
CDS	.PE protein coding region
LOCUS	sequenced DNA fragment
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.23 HAMAPnucl

### E.23.1 Bank details

HAMAP - Acnuc Release - Nucleotides - Last Updated: Jul 12, 2006  
 262,753,516 bases; 4,077 sequences; 230,900 subseqs; 0 refers.

Microbial genomes

Nucleotide data from EMBL cross-references  
 in protein data from HAMAP Database  
[\(http://us.expasy.org/sprot/hamap/\)](http://us.expasy.org/sprot/hamap/)  
 developed by the Swiss-Prot group at the Swiss Institut of Bioinformatics

Software: M. Gouy & M. Jacobzone  
 Data maintenance: L. Duret & S. Penel

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### E.23.2 Type names

name	description
ID	EMBL sequence data library entry
CDS	.PE protein coding region
TRNA	.TR transfer RNA coding region
RRNA	.RR ribosomal RNA coding region
MISC_RNA	.RN other structural RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
3'INT	.3I 3' intron
3'NCR	.3F 3'-non coding region
5'INT	.5I 5' intron
5'NCR	.5F 5'-non coding region
INT_INT	.IN internal intron

## E.24 HAMAPprot

### E.24.1 Bank details

HAMAP - Acnuc Release -Proteins- Last Updated: Jul 12, 2006  
 39,750,000 amino acids; 125,848 sequences; 9,554 references.

Microbian genomes

Protein data from HAMAP Database  
[\(http://us.expasy.org/sprot/hamap/\)](http://us.expasy.org/sprot/hamap/)  
 developed by the Swiss-Prot group at the Swiss Institut of Bioinformatics

Software: M. Gouy & M. Jacobzone  
 Data maintenance: L. Duret & S. Penel

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### E.24.2 Type names

There are no subsequence type in this database

## E.25 hoppsigen

### E.25.1 Bank details

there is no information available about the contents of this bank

### E.25.2 Type names

name	description
ID	EMBL sequence data library entry
CDS	.PE protein coding region
TRNA	.TR transfer RNA coding region
RRNA	.RR ribosomal RNA coding region
MISC_RNA	.RN other structural RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
CDE	.PS
PPGENE	.PP
3'FL	.3F
5'FL	.5F
DIRECT_REPEAT	.DR
REPEAT_REGION	.RR
POLYA_REGION	.PA
FL_REPEAT	.FR

## E.26 nurebnuc1

### E.26.1 Bank details

Nurebase 4.0 (26 September 2003) Last Updated: NOV 27, 2003

2,356,663 bases; 664 sequences; 518 subseqs; 787 refers.

Software by M. Gouy & M. Jacobzone, Laboratoire de biometrie, Universite Lyon I

### E.26.2 Type names

name	description
CDS	.PE protein coding region
ID	EMBL sequence data library entry
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.27 nurebprot

### E.27.1 Bank details

Nurebase 4.0 (26 September 2003) Last Updated: NOV 27, 2003  
 277,024 amino acids; 525 sequences; 634 references.  
 Software by M. Gouy & M. Jacobzone, Laboratoire de biometrie, Universite Lyon I

### E.27.2 Type names

There are no subsequence type in this database

## E.28 taxobacgen

### E.28.1 Bank details

TaxoBacGen Rel. 7 (September 2005)  
 1,151,149,763 bases; 254,335 sequences; 847,767 subseqs; 63,879 refers.  
 Data compiled from GenBank by Gregory Devulder  
 Laboratoire de Biometrie & Biologie Evolutive, Univ Lyon I

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This database is a taxonomic genomic database.

It results from an expertise crossing the data nomenclature database DSMZ

<http://www.dsmz.de/species/bacteria.htm> Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, D-8215

and GenBank.

- Only contains sequences described under species present in Bacterial Nomenclature Up-to-date.
  - Names of species and genus validly published according to the Bacteriological Code (names with standing in nomenclature) is added in field "DEFINITION".
  - A keyword "type strain" is added in field "FEATURES/source/strain" in GenBank format definition to easily identify Type Strain.
- Taxobacgen is a genomic database designed for studies based on a strict respect of up-to-date nomenclature and taxonomy.

### E.28.2 Type names

name	description
CDS	.PE protein coding region
LOCUS	sequenced DNA fragment
MISC_RNA	.RN other structural RNA coding region
RRNA	.RR ribosomal RNA coding region
SCRNA	.SC small cytoplasmic RNA coding region
SNRNA	.SN small nuclear RNA coding region
TRNA	.TR transfer RNA coding region

## E.29 Session Informations

This part was compiled under the following  environment:

- R version 2.4.1 (2006-12-18), i386-apple-darwin8.8.1
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: MASS 7.2-30, ade4 1.4-2, ape 1.9-3, gee 4.13-12, lattice 0.14-16, nlme 3.1-78, seqinr 1.0-7, xtable 1.4-3

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