

R PROFILING AND OPTIMISATION

GÜNTHER SAWITZKI

PENDING CHANGES

Warning: this is under construction.

This vignette contains experimental which may sink down to the package implementation, or vanish.

Known issues:

- Control information may be included as special stack in raw format.
- A list of profiles may become default. Only one profiling interval value per profile.
- Nodes may be implemented as *factor*.

CONTENTS

Pending changes	1
Profiling facilities in R	2
L ^A T _E X layout tools and R settings	2
1. Profiling	4
1.1. Simple regression example	5
1.1.1. R basic	6
1.1.2. Package sprof	9
1.1.3. Node classes	13
2. A better grip on profile information	18
2.1. The internal details	18
2.2. The free lunch	23
2.3. Cheap thrills	24
2.3.1. Trimming	27
2.3.2. Surgery	31
2.4. Run length	32
3. xxx – lost & found	41
3.1. Graph package	51
4. Standard output	52

Date: May 2013. *Revised:* July 2013

Typeset, with minor revisions: July 27, 2013 from SVN *Revision* : 182 2013-07-27.

Key words and phrases. R programming, profiling, optimisation, R program language.

An R vignette for package sprof.

URL: <http://sintro.r-forge.r-project.org/>

Private Version

4.1. Print	52
4.2. Summary	55
4.3. Plot	59
5. Graph	65
5.1. Example: regression	65
5.1.1. graph package	66
5.1.2. igraph package	67
5.1.3. network package	68
5.1.4. Rgraphviz package	70
5.2. Trimmed example: regression	75
5.2.1. graph package	76
5.2.2. igraph package	79
5.2.3. network package	80
5.2.4. Rgraphviz package	85
Index	90

PROFILING FACILITIES IN R

R provides the basic instruments for profiling, both for time based samplers as for event based instrumentation. Information on R profiling is in Section 3.2 “Profiling R code for speed” and section “3.3 Profiling R code for memory use”. of Writing R Extensions <http://cran.r-project.org/doc/manuals/R-exts.html>. Specific information on memory profiling is in <http://developer.r-project.org/memory-profiling.html>.

However this source of information seem to be rarely used.

Maybe the supporting tools are not adequate. The summaries provided by R reduce the information beyond necessity. Additional packages are available, but these are not sufficiently action oriented.

With package *sprof* we want to give a data representation that keeps the full profile information. Tools to answer common questions are provided. The data structure should make it easy to extend the tools as required.

The package is currently distributed at r-forge as part of the *sintro* material.

To install this package directly within R, type

```
install.packages("sprof", repos="http://r-forge.r-project.org")
```

To install the recent package from source directly within R, type

```
install.packages("sprof", repos="http://r-forge.r-project.org", type="source")
```

L^AT_EX LAYOUT TOOLS AND R SETTINGS

You may want to skip this section, unless you want to modify the vignette for your own purposes, or look at the internals.

This is the main library we are going to use.

```
library(sprof) Input
```

We want immediate warnings, if necessary. Set to level 2 to handle warnings as error.

```
message("switching options(warn=1) -- immediate warning on")
options(warn=1) Input
```

We want a second chance on errors. So install an error handler.

```
options(error = recover) Input
```

Print parameters used here:

```
options(width = 72)
options(digits = 6) Input
```

For *str* output, we generally use these settings:

```
strx <- function(x, ...){
  str(x, max.level=2, vec.len=3, nchar.max=40, list.len=6,
    width=70, strict.width="wrap",...)
} Input
```

For larger tables and data frames, we use a kludge to avoid long outputs.

ToDo: add keep3 to keep header, some middle, tail

```
xcutdata.frame <- function(df, cut, margin){
  #! keep3, to add: margin top - random center - margin bottom
  if (!is.data.frame(df)) return(df)
  nrow <- nrow(df)
  # cut a range if it is not empty.
  # Quiet noop else.
  # Does not cut single lines.
  cutrng <- function(cutfrom,cutto){
    if (cutfrom<cutto){
      df[cutfrom,] <- NA
      if (!is.null(rownames(df))) rownames(df)[cutfrom] <- "< cut >"
      if (!is.null(df$name)) df$name[cutfrom] <- ""

      cutfrom <- cutfrom+1
      df[-(cutfrom:cutto),]
    }#if
  }
  if (!missing(cut)) {df <- cutrng(cut[1],cut[2]); return(df)}
  if (!missing(margin)) {
    if (length(margin)==1) margin <- c(margin,margin)
    cut <- c(margin[1]+1,nrow-margin[2])
    df <-cutrng(cut[1],cut[2]);
```

```

    return(df)}
#   if (!missing(keep3)) { cut <- c(keep3[1]+1, keep3[1]+1,
#                                   nrow-keep3[3]-1,nrow-keep3[3]-1)
#   if (cut[3]-cut[4] > keep3[2]+2){delta<-(cut[3]-cut[2]) div 2
#   cut[3]<-0
#   browser()
#   } else df <- cutrng(cut[1],cut[4])
#   cutrng(cut[1],cut[4]) return(df)}
}

```

We use the R function `xtable()` for output and \LaTeX `longtable`. A convenient wrapper to use this in our *Sweave* source is:

ToDo: remove text
vdots from string/
name columns. Use
empty string.

```

library(xtable)
prxt <- function (x, digits=2, cut=TRUE, caption=NULL,
                  label=NULL, ...) {
  if (cut) {margin <- 10
  if (nrow(x)> 2*margin+3) x <-xcutdata.frame(x, margin=margin)}
  print(
    xtable(x, digits=digits, caption=caption,
            label=label, ...),
    floating=FALSE,
    tabular.environment="longtable",
    caption.placement="top",
    zero.print = ".",
    NA.string="\vdots")
  }

```

This is to be used with `<<print=FALSE, results =tex, label=tab:prxx>>=`

1. PROFILING

The basic information provided by all profilers is a protocol of sampled stacks. For each recorded event, the protocol has one record, such as a line with a text string showing the sampled stack.

We use profiles to provide hints on the dynamic behaviour of programs. Most often, this is used to improve or even optimise programs. Sometimes, it is even used to understand some algorithm.

Profiles represent the program flow, which is considered to be laid out by the control structure of a program. The control structure is represented by the control graph, and this leads to the common approach to (re)construct the control graph, map the profile to this graph, and used graph based methods for further analysis. The prime example for this strategy is the GNU profiler *gprof* (see <http://sourceware.org/binutils/docs/gprof/>) which is used as master plan for many common profilers.

It is only half of the truth that the control graph can serve as a base for the profiled stacks. In R, we have some peculiarities.

lazy evaluation: Arguments to functions can be passed as promises. These are only evaluated when needed, which may be at a later time, and may then lead to insertions in the stack. So we may have information resulting from the data flow, interspersed with the control flow.

memory management: Allocation of memory, and garbage collection, may interfere and leave their traces in the stack. While allocation is closely related to the visible control flow, garbage collection is a collective effect largely out of control of the code to execute.

primitives: Internal functions may escape the usual stack conventions and execute without leaving any identifiable trace on the stack.

control structures: In R, many control structures are implemented as function. Most notably, the `apply()` family appear as function calls and lead to cliques in the graph representation that do not correspond to relevant structures. Since these functions are well known, they can have a special treatment.

So while the stack follows an overall well known dynamics, in R there are exceptions from regularity.

The general approach, by `summaryRprof()` and others, is to reduce the profile to node information, or to consider single transitions.

We take a different approach. We take the stacks, as recorded in the profiles as our basic information unit. From this, we ask: what are the actions we need to answer our questions? Representation in graphs may come later, if they can help.

If the stacks would come from the control flow only, we could make use of the sequential nature of stacks. But since we have to live with the R specific interferences, we stay with the raw stacks.

In this presentation, we will use a small list of examples. Since `Rprof` is not implemented on all systems, and since the profiles tend to get very large, we use some prepared examples that are frozen in this vignette and not included in the distribution, but all the code to generate the examples is provided.

ToDo: rearrange
stacks? detect
order?

1.1. Simple regression example.

Input

```
n <- 10000
x <- runif(n)
err <- rnorm(n)
y <- 2 + 3 * x + err
reg0data <- data.frame(x=x, y=y, err=err)
rm(x,y,err)
```

We will use this example to illustrate the basics. Of course the immediate questions are the variance between varying samples, and the influence of the sample size n . We keep everything fixed, so the only issue for now is the computational performance under strict iid conditions.

Still we have parameters to choose. We can determine the profiling granularity by setting the timing interval, and we can use repeated measurements to increase precision below the timing interval.

The timing interval should depend on the clock speed. Using for example 1ms amounts to some 1000 steps on a current CPU, per kernel.

If we use repeated samples, the usual rules of statistics applies. So taking 100 runs and taking the mean reduces the standard deviation by a factor 1/10.

Following the usual R conventions, seconds are used as time base for parameters. However report will use ms as a time base.

Here is an example how to take a profile, using basic R. See section 1.1.2 on page 9 how to use *sampleRprof* in package *sprof* for an easier solution.

ToDo: Can we calibrate times to CPU rate? Introduce cpu clock cycle as a time base

Input

```

profinterval <- 0.001
simruns <- 100
Rprof(filename="RprofsRegressionExpl.out", interval = profinterval)
  for (i in 1:simruns) xxx<- summary(lm(y~x, data=reg0data))
Rprof(NULL)

```

We now have the profile data in a file *RprofsRegressionExpl.out*. For this vignette, we use a frozen version *RprofsRegressionExpl01.out*.

1.1.1. *R basic*. The basic R functions invite us to get a summary.

Input

```

sumRprofRegressionExpl <- summaryRprof("RprofsRegressionExpl01.out")
#str(profile_nodes_rle, max.level=2, vec.len=3, nchar.max=40, list.len=6)
strx(sumRprofRegressionExpl)

```

Output

```

List of 4
$ by.self : 'data.frame': 41 obs. of 4 variables:
..$ self.time : num [1:41] 0.087 0.057 0.051 0.043 0.042 0.04 0.032
  0.026 ...
..$ self.pct : num [1:41] 16.67 10.92 9.77 8.24 ...
..$ total.time: num [1:41] 0.113 0.099 0.069 0.043 0.474 0.045 0.033
  0.114 ...
..$ total.pct : num [1:41] 21.65 18.97 13.22 8.24 ...
$ by.total : 'data.frame': 62 obs. of 4 variables:
..$ total.time: num [1:62] 0.522 0.522 0.521 0.521 0.521 0.521 0.521
  0.521 ...
..$ total.pct : num [1:62] 100 100 99.8 99.8 ...
..$ self.time : num [1:62] 0.006 0 0.001 0 0 0 0 ...
..$ self.pct : num [1:62] 1.15 0 0.19 0 0 0 0 ...
$ sample.interval: num 0.001
$ sampling.time : num 0.522

```

The summary reduces the information contained in the profile to marginal statistics per node. This is provided in two data frames giving the same information, only in different order.

The file contains several spurious recordings: nodes that have been recorded only few times. It is worth noting these, but then they better be discarded. We use a time limit of 4ms, which given our sampling interval of 1ms means we require more than four observations.

```

prxt(sumRprofRegressionExpl$by.self,
      caption="summaryRprof result: by.self as final stack entry, all records",
      label="tab:prSRREbs")

```

Table 1: summaryRprof result: by.self as final stack entry, all records

	self.time	self.pct	total.time	total.pct
"lm.fit"	0.09	16.67	0.11	21.65
"[.data.frame"	0.06	10.92	0.10	18.97
"model.matrix.default"	0.05	9.77	0.07	13.22
"as.character"	0.04	8.24	0.04	8.24
"lm"	0.04	8.05	0.47	90.80
"summary.lm"	0.04	7.66	0.04	8.62
"structure"	0.03	6.13	0.03	6.32
"na.omit.data.frame"	0.03	4.98	0.11	21.84
"anyDuplicated.default"	0.02	4.21	0.02	4.21
"as.list.data.frame"	0.02	4.21	0.02	4.21
< cut >	:	:	:	:
"FUN"	0.00	0.19	0.01	1.34
"%in%"	0.00	0.19	0.00	0.77
"deparse"	0.00	0.19	0.00	0.38
"\$"	0.00	0.19	0.00	0.19
"as.list.default"	0.00	0.19	0.00	0.19
"as.name"	0.00	0.19	0.00	0.19
"coef"	0.00	0.19	0.00	0.19
"file"	0.00	0.19	0.00	0.19
"NCOL"	0.00	0.19	0.00	0.19
"terms.formula"	0.00	0.19	0.00	0.19

```

prxt(sumRprofRegressionExpl$by.total[sumRprofRegressionExpl$by.total$total.time>0.004,],
      caption="summaryRprof result: by.total, total time > 4ms",
      label="tab:prSRREbt")

```

Table 2: summaryRprof result: by.total, total time > 4ms

	total.time	total.pct	self.time	self.pct
"<Anonymous>"	0.52	100.00	0.01	1.15
"Sweave"	0.52	100.00	0.00	0.00
"eval"	0.52	99.81	0.00	0.19
"doTryCatch"	0.52	99.81	0.00	0.00

"evalFunc"	0.52	99.81	0.00	0.00
"try"	0.52	99.81	0.00	0.00
"tryCatch"	0.52	99.81	0.00	0.00
"tryCatchList"	0.52	99.81	0.00	0.00
"tryCatchOne"	0.52	99.81	0.00	0.00
"withVisible"	0.52	99.81	0.00	0.00
< cut >	⋮	⋮	⋮	⋮
"as.list"	0.02	4.41	0.00	0.00
"anyDuplicated.default"	0.02	4.21	0.02	4.21
"as.list.data.frame"	0.02	4.21	0.02	4.21
"sapply"	0.01	2.68	0.00	0.19
"match"	0.01	2.11	0.00	0.19
"[[.data.frame"	0.01	1.53	0.00	0.19
"["	0.01	1.53	0.00	0.00
"rep.int"	0.01	1.34	0.01	1.34
"FUN"	0.01	1.34	0.00	0.19
"list"	0.01	0.96	0.01	0.96

1.1.2. *Package sprof*. In contrast to the common R packages, in the *sprof* implementation we take a two step approach. First we read in the profile file to an internal representation. Analysis is done in later steps.

Input

```
sprof01<- readRprof("RprofsRegressionExpl01.out")
```

The data contain identification information for reference. This will be used in the functions of *sprof* and shown in the displays. Here is the summary of this section:

Input

```
str(sprof01$info)
```

Output

```
'data.frame':      1 obs. of  8 variables:
 $ id      : Factor w/ 1 level  "\"RprofsRegressionExpl01.out\" 2013-06-13 23:46:04": 1
 $ date    : POSIXct, format: "2013-07-27 18:07:20"
 $ nrnodes : int 62
 $ nrstacks: int 50
 $ nrrecords: int 522
 $ firstline: Factor w/ 1 level "sample.interval=1000": 1
 $ ctllines : Factor w/ 1 level "sample.interval=1000": 1
 $ ctllinenr: num 1
```

For this vignette, we change the *id* information. So in this context:

Input

```
sprof01$info$id <- "sprof01"
```

We keep this example and use the copy *sprof01* of it extensively for illustration.

Input

```
save(sprof01, file="sprof01m.RData")
```

To run the vignette with a different profile, replace *sprof01* by your example. You still have the file for reference. The data contain identification information for reference. This will be used in

Package *sprof* provides a function *sampleRprof()* to take a sample and create a profile on the fly, as in

Input

```
sprof01temp <- sampleRprof(runif(10000), runs=100)
```

The basic data structure consists of four data frames. The *info* section collects global information from the input file, such as an identification strings and various global matrix. The *nodes* section initially gives the same information marginal information as *summaryRprof*. The *stacks* section puts the node information into their calling context as found in the input profile file. The *profiles* section gives the temporal context. It is implemented as a list, but conceptually it is a data frame. Implementing it as a list allows run length encoding of variables, which unfortunately is not allowed by R in data frames.

ToDo: add sampling.interval, sampling.time for backward compability

```
strx(sprof01)
```

```
Input
```

```
List of 4
$ info : 'data.frame': 1 obs. of 8 variables:
..$ id : chr "sprof01"
..$ date : POSIXct[1:1], format: "2013-07-27 18:07:20"
..$ nrnodes : int 62
..$ nrstacks : int 50
..$ nrrecords: int 522
..$ firstline: Factor w/ 1 level "sample.interval=1000": 1
.. [list output truncated]
$ nodes : 'data.frame': 62 obs. of 5 variables:
..$ name : Factor w/ 62 levels "!", "..getNamespace", ...: 1 2 3 4 5 6 7
  8 ...
..$ self.time : num [1:62] 2 0 2 0 0 57 0 1 ...
..$ self.pct : num [1:62] 0.38 0 0.38 0 ...
..$ total.time: num [1:62] 2 1 4 26 99 99 8 8 ...
..$ total.pct : num [1:62] 0.03 0.01 0.05 0.34 1.29 1.29 0.1 0.1 ...
$ stacks : 'data.frame': 50 obs. of 7 variables:
..$ nodes :List of 50
.. .. [list output truncated]
..$ shortname : Factor w/ 50 levels
  "S<A>eFttCtCLtC0dTCwVeesleem.m..n.n...[["| __truncated__, ...: 27 17
  19 1 35 36 37 30 ...
..$ refcount : num [1:50] 1 5 26 55 13 43 51 87 ...
..$ stacklength : int [1:50] 19 20 19 21 14 15 15 14 ...
..$ stackheadnodes: int [1:50] 52 52 52 52 52 52 52 52 ...
..$ stackleafnodes: int [1:50] 27 28 41 6 39 14 38 30 ...
.. [list output truncated]
$ profiles:List of 4
..$ data : int [1:522] 1 2 2 3 4 4 5 5 ...
..$ mem : NULL
..$ malloc : NULL
..$ timesRLE:List of 2
.. ..- attr(*, "class")= chr "rle"
- attr(*, "class")= chr [1:2] "sprof" "list"
```

The nodes do not come in a specific order. Access via a permutation vector is preferred. This allows different views on the same data set. For example, table 4 on page 12 uses a permutation by total time, and a selection (compare to table 2 on page 8). The only difference is that *sprof* works uses a millisecond (ms) base, whereas R in general uses seconds as a base.

```
nodes <- sprof01$nodes[order(sprof01$nodes$self.time, decreasing=TRUE),]
prxt(nodes[nodes$self.time>4,],
caption="splot result: by.self, self time > 4ms",
label="tab:prspbtself")
```

Table 3: splot result: by.self, self time > 4ms

	name	self.time	self.pct	total.time	total.pct
30	lm.fit	87.00	16.67	113.00	1.47
6	[.data.frame	57.00	10.92	99.00	1.29
38	model.matrix.default	51.00	9.77	69.00	0.90
14	as.character	43.00	8.24	43.00	0.56
29	lm	42.00	8.05	474.00	6.16
51	summary.lm	40.00	7.66	45.00	0.59
49	structure	32.00	6.13	33.00	0.43
41	na.omit.data.frame	26.00	4.98	114.00	1.48
13	anyDuplicated.default	22.00	4.21	22.00	0.29
16	as.list.data.frame	22.00	4.21	22.00	0.29
40	na.omit	20.00	3.83	134.00	1.74
39	model.response	13.00	2.49	56.00	0.73
36	model.frame.default	12.00	2.30	168.00	2.18
46	rep.int	7.00	1.34	7.00	0.09
10	<Anonymous>	6.00	1.15	522.00	6.79
28	list	5.00	0.96	5.00	0.07

At this level, it is helpful to note the expectations, and only then inspect the timing results. Since we are using a linear model, we are not surprised to see functions related to linear models on the top of the list. We may however be surprised to see functions related to data access and to character conversion very high on the list. The sizeable amount of time spent on NA handling is another aspect that is surprising.

```

Input
nodes <- sprof01$nodes[order(sprof01$nodes$total.time, decreasing=TRUE),]
prxt(nodes[nodes$total.time>4,],
caption="splot result: by.total, total time > 4ms",
label="tab:prspbt")

```

Table 4: splot result: by.total, total time > 4ms

	name	self.time	self.pct	total.time	total.pct
10	<Anonymous>	6.00	1.15	522.00	6.79
52	Sweave	0.00	0.00	522.00	6.79
21	doTryCatch	0.00	0.00	521.00	6.78
22	eval	1.00	0.19	521.00	6.78
23	evalFunc	0.00	0.00	521.00	6.78
55	try	0.00	0.00	521.00	6.78
56	tryCatch	0.00	0.00	521.00	6.78
57	tryCatchList	0.00	0.00	521.00	6.78
58	tryCatchOne	0.00	0.00	521.00	6.78
62	withVisible	0.00	0.00	521.00	6.78
< cut >	\vdots	⋮	⋮	⋮	⋮
61	vapply	3.00	0.57	23.00	0.30
13	anyDuplicated.default	22.00	4.21	22.00	0.29

16	as.list.data.frame	22.00	4.21	22.00	0.29
47	sapply	1.00	0.19	14.00	0.18
31	match	1.00	0.19	11.00	0.14
7	[[0.00	0.00	8.00	0.10
8	[[.data.frame	1.00	0.19	8.00	0.10
25	FUN	1.00	0.19	7.00	0.09
46	rep.int	7.00	1.34	7.00	0.09
28	list	5.00	0.96	5.00	0.07

ToDo: remove text
vdots from string/
name columns

Given the sampling structure of the profiles, two aspects are common. The sampling picks up scaffolding functions with a high, nearly constant frequency. And the sampling will pick up rare recordings that are near to detection range. The display functions hide these effects by default. In our example, about half of the nodes are cleared by this garbage collector.

Common rearrangements as by total time and by self time are supplied by the display functions.

`irfunxhplotplot_nodes`, for example, currently gives a choice of four displays for nodes, and supports trimming by default. Our profile starts with 62 nodes. The defaults cut off 34 nodes as uninformative, either because they are too rare, or ubiquitous.

See fig. 1 on the next page.

Information in the scatterplots may sometimes be more accessible when using a logarithmic scale, so this is added.

If you prefer, you can have the bar charts in horizontal layout, giving more space for labels.

See fig. 2 on page 14.

We can add colour. To illustrate this, we encode the frequency of the nodes as colour. As a palette, we choose a heat map here.

ToDo: apply colour
to selection?

ToDo: spread
colour on displayed
part

```

freqrank01 <- rank(-sprof01$nodes$total.time, ties.method="random")
freqrankcol01 <- heat.colors(length(freqrank01))

```

Here is the node view using these choices:

See fig. 2 on page 14.

Colour is considered a volatile attribute. So you may need to pay some attention to keep colour indices (and colour palettes) aligned to your context. You may want to do experiments with colour, trying to find a good solution for your visual preferences. The recommended way is to use some stable colour index (the slot `icol` is reserved for this) and use this as an index to a choice of colour palettes. So `icol` becomes a part of the data structure, and the colour palette to be used is passed as a parameter.

ToDo: improve
colour: support
colour in a structure

#10

Nodes by time

total

self.sprofo1

Nodes: time as last of stack

count

sprofo1

62 nodes
25 nodes with 5 or less
total counts omitted

Nodes: total time in stack

count

sprofo1

62 nodes
25 nodes with 5 or less
total counts omitted
9 permanent nodes omitted

Nodes by time

log(total)

log(self+1)
sprofo1

FIGURE 1. Basic information on node level

The attribute *icol* is a special case which we used above. If present, it will be interpreted as an index to a colour table. For example, we can collect special well known functions in groups.

The node information is to some part arbitrary. You may achieve the same functionality by different functions, and you will see different load in the profiles. Grouping nodes may be a mean to clarify the picture.

ToDo: colour by class – redo. Bundle colour index with colour?

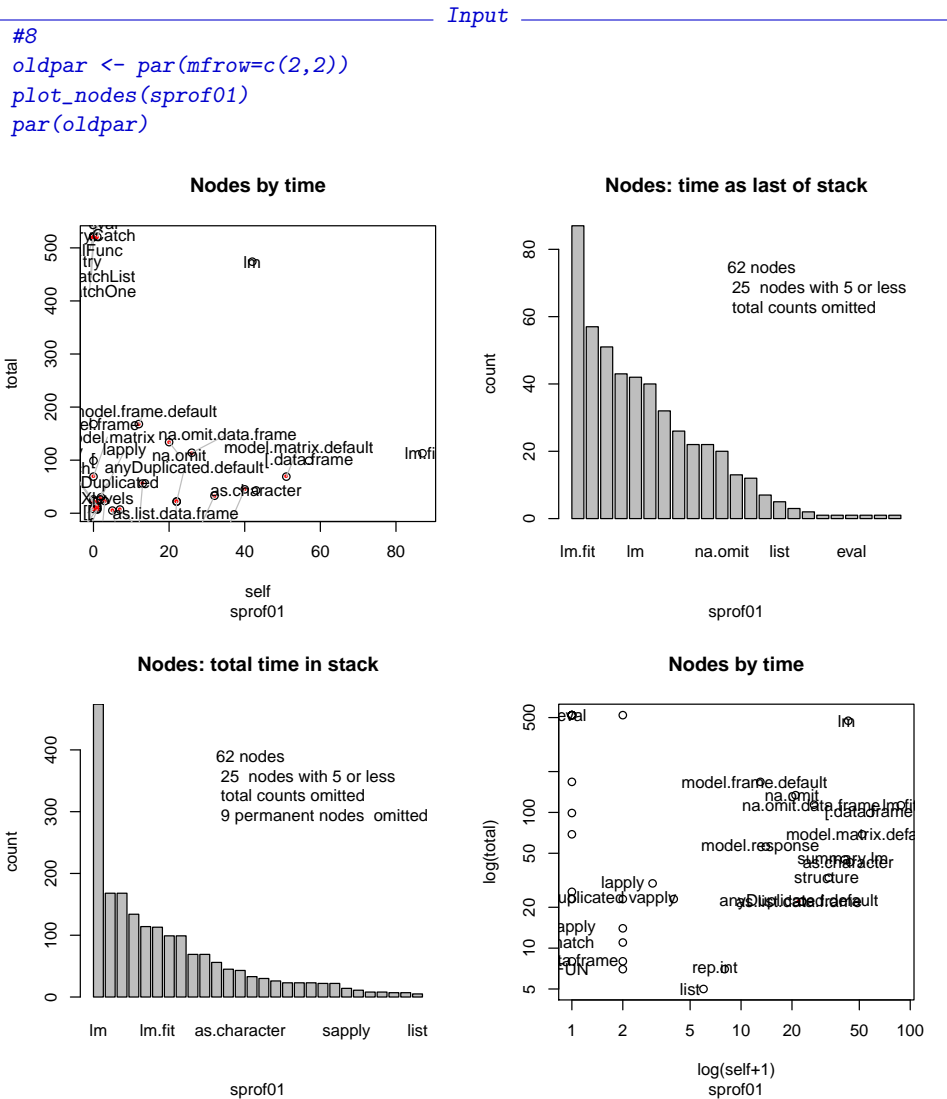


FIGURE 2. Basic information on node level

Grouping may also help you to focus your attention. “HOT” and “cold” may be very helpful tags. These can be used in a flexible way.

ToDo: Move class attributes to package code

ToDo: add class by keyword

ToDo: add class by package for all nodes

```
nodekeyword0 <- function(node)
{
}
```

Input

```
#10
sprof01$nodes$icol <- freqrank01
oldpar <- par(mfrow=c(2,2))
plot_nodes(sprof01, col=freqrankcol01)
par(oldpar)
```

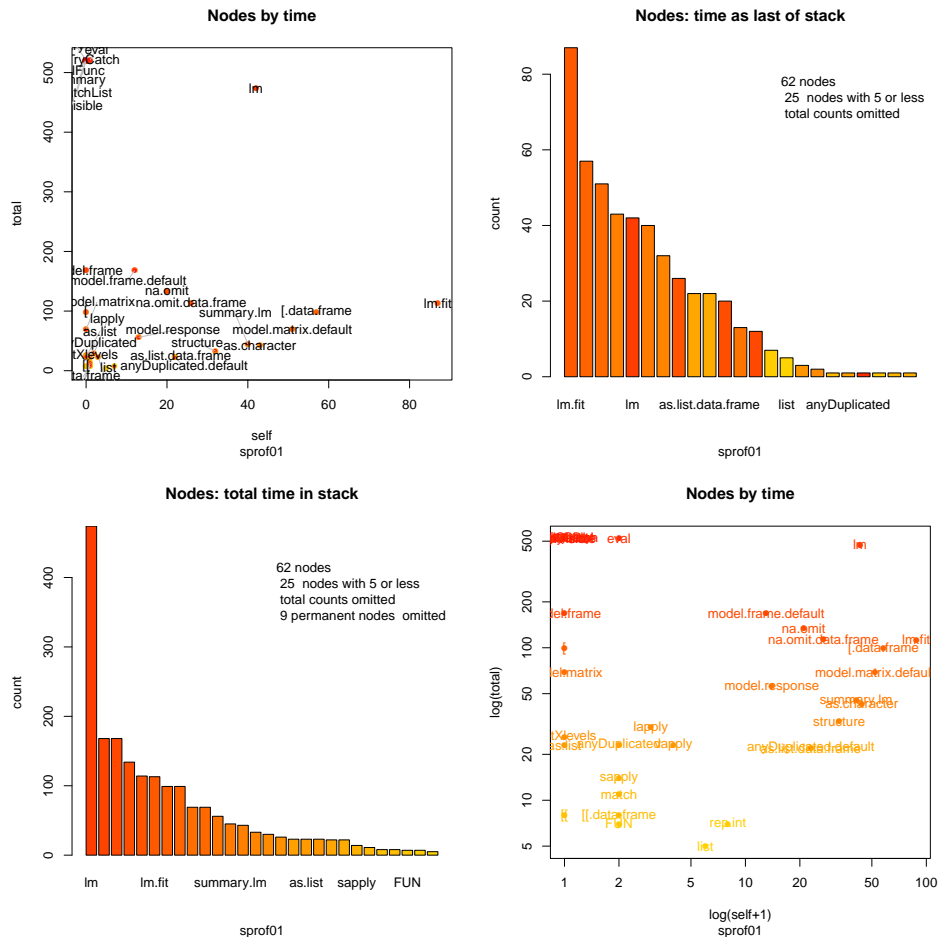


FIGURE 3. Basic information on node level, colour by total time.

```
## cf. source('~Documents/lectures/src/insider/profile_pkgs/profr_0.2/R/parse.r', chdir = TRUE)
nodepackage0 <- function(node)
{where <- getAnywhere(node)$where
  if (length(where)==0) return("<not found>") else{
    where <- strsplit(where,':')
    if (is.null(where)) return("<??>") else
      if (where[[1]][1] == "package") return(where[[1]][2]) else
        {print(paste(node,"not found. got:",where)); return("nn")}
      }
}
```

```

nodepackage<- function(x)
{
  x <- as.character(x)
  xpackage <- rep("nn",length(x))
  for (i in (1:length(x))) xpackage[i]<-nodepackage0(x[[i]])
  xpackage
}

```

```

nodepackages <- nodepackage(sprof01$nodes$name)

```

```

[1] "na.omit.data.frame not found. got: registered S3 method for na.omit from namespace stats"
[2] "na.omit.data.frame not found. got: c(\"namespace\", \"stats\")"

```

```

names(nodepackages) <- sprof01$nodes$name
table(nodepackages)

```

```

nodepackages
<not found>      base      nn      stats      utils
              6         41         1         13         1

```

```

#nodepackages <- lapply(as.character(sprof01$nodes$name),nodepackage0)

```

```

sprof01$nodes$icol <-as.factor(nodepackages)

```

See fig. 4 on the facing page.

```

x_apply <- c("apply", "lapply", "vapply", "sapply")
x_as <- c("as.list", "as.data.frame", "as.list.data.frame",
         "as.character", "as.list.default", "as.name")

```

(Extend as you need it) and then use, as for example:

```

nodeclass <- rep("x_nn", sprof01$info$nrnodes)
nodeclass[sprof01$nodes$name %in% x_apply] <- "x_apply"
nodeclass[sprof01$nodes$name %in% x_as] <- "x_as"

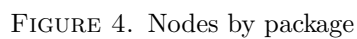
```

or use assignments on the fly

```

nodeclass[sprof01$nodes$name %in%
  c("eval", "evalFunc",
    "try", "tryCatch", "tryCatchList", "tryCatchOne",
    "doTryCatch")
] <- "x_eval"
nodeclass[sprof01$nodes$name %in%

```



```

      c("model.frame", "model.matrix.default", "model.frame.default",
        " model.response", "model.matrix", "model.response")
    ] <- "x_model"
  nodeclass[sprof01$nodes$name %in%
    c("lm", "lm.fit", "summary.lm")
    ] <- "x_lm"
  nodeclass[sprof01$nodes$name == "<Anonymous>"] <- "x_Anon"

```

```

      Input
sprof01$nodes$icol <- as.factor(nodeclass)

```

adds a sticky color attribute. To interpret, you should choose your preferred color palette, for example

ToDo: add a reference to colorbrewer

```

      Input
classcol <- c("red", "green", "blue", "yellow", "cyan", "magenta", "purple")

```

ToDo: Defaults by class

See fig. 5 on the next page.

See fig. 6 on page 20.

You can break down the frequency by classes of your choice. But beware of Simpson's paradox. The information you think you see may be strongly affected by your choices - what you see are reflections of conditional distributions. These may be very different from the global picture.

If package `wordcloud` is installed, a different view is possible. This is added in the plots above.

2. A BETTER GRIP ON PROFILE INFORMATION

The basic information provided by all profilers in R is a protocol of sampled stacks. The conventional approach is to break the information down to nodes and edges. The stacks provide more information than this. One way to access it is to use linking to pass information. This has already been used on the node level in section 1.1.2 on page 9.

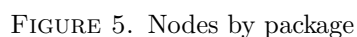
ToDo: add attributes to stacks, and discuss scope

ToDo: sorting/arranging stacks

2.1. The internal details. For each recorded event, the protocol records one line with a text string showing the sampled stack (in reverse order: most recent first). The stack lines may be preceded by header lines with event specific information. The protocol may be interspersed with control information, such as information about the timing interval used.

We know that the structural information, static information as well as dynamic information, can be represented with the help of a graph. For a static analysis, the graph representation may be the first choice. For a dynamic analysis, the stack information is our first information. A stack is a connected path in the program graph. If we start with nodes and edges, we lose information which is readily available in record of stacks.

Input



Input

```
oldpar <- par(mfrow=c(3,2))
plot_nodes(sprof01, which=1:6)
par(oldpar)
```

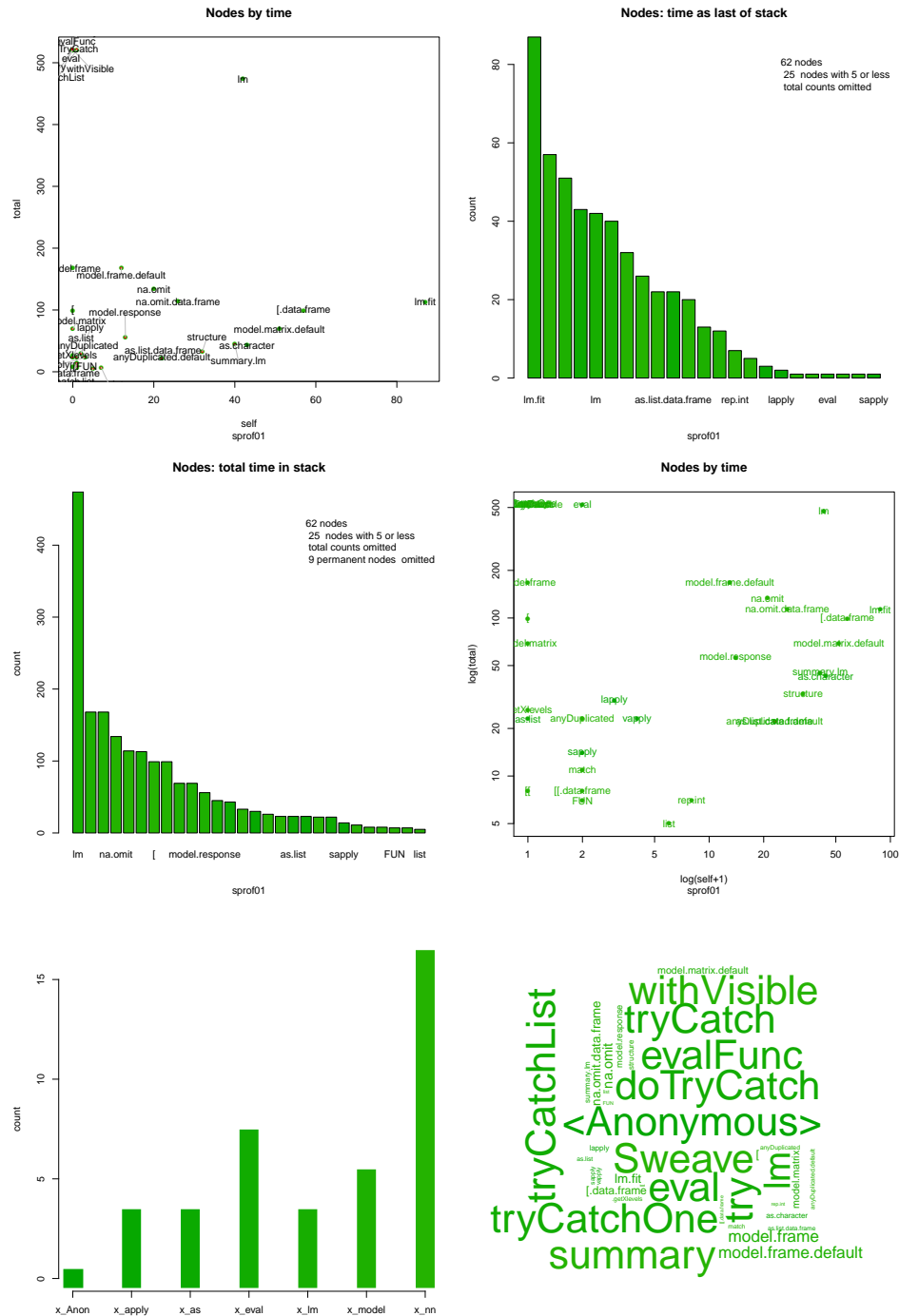


FIGURE 6. Nodes by package: default colour selection

As we know that we are working with stacks, we know that they have their peculiarities. Stacks tend to grow and shrink. Subsequent events will have extensions and shrinkages of stacks (if the recording is on a fine scale), or stack sharing common stumps (if the recording is on a coarser scale).

There have always been interrupts, and these show up in profiles. In R, this is related problem (GC)

The graph is a second instance that is (re)constructed from the stack recording.

Here is the way we represent the profile information:

The profile log file is sanitised:

- Control lines are extracted and recorded in a separate list.
- Head parts, if present, are extracted and recorded in a matrix that is kept line-aligned with the remainder
- Line content is standardised, for example by removing stray quotation marks etc.

After this, the sanitised lines are encoded as a vector of stacks, and references to this.

If necessary, these steps are done by chunks to reduce memory load.

From the vector of stacks, a vector of nodes (or rather node names) is derived.

The stacks are now encoded by references to the nodes table. For convenience, we keep the (sanitised) textual representation of the stacks.

So far, texts are in reverse order. For each stack, we record the trailing leaf, and then we reverse order. The top of stack is now on first position.

Several statistics can be accumulated easily as a side effect.

Conceptually, the data structure consist of three tables (the implementation may differ, and is subject to change).

The profiles table is the representation of the input file. Control lines are collected in a special table. With the control lines removed, the rest is a table, one row per input line. The body of the line, the stack, is encoded as a reference to a stacks table (obligatory) and header information (optional).

The stacks table contains the collected stacks, each stack encoded as a list of references to the node table. This is obligatory. This list is kept in reverse order (root at position 1). A source line representing the stack information may be kept (optional).

The nodes table keeps the names at the nodes.

Sometimes, it is more convenient to use a simple representation, such as a matrix. Several extraction routines are provided for this, and the display routines make heavy use of this. See table 5 on the following page.

We now can go beyond node level.

ToDo: complete
matrix conversion

TABLE 5. Extraction and conversion routines

<code>profiles_matrix()</code>	incidence matrix: nodes by event
<code>stacks_matrix()</code>	incidence matrix: nodes by stack
<code>list.as.matrix()</code>	fill list to equal length and convert to matrix
<code>stackstoadj()</code>	stacks to (correspondence) adjacency matrix
<code>adjacency()</code>	sprof to (correspondence) adjacency matrix

ToDo: check and stabilize color linking

This is what we get for free from the node information on our three levels: node, stack, and profile.

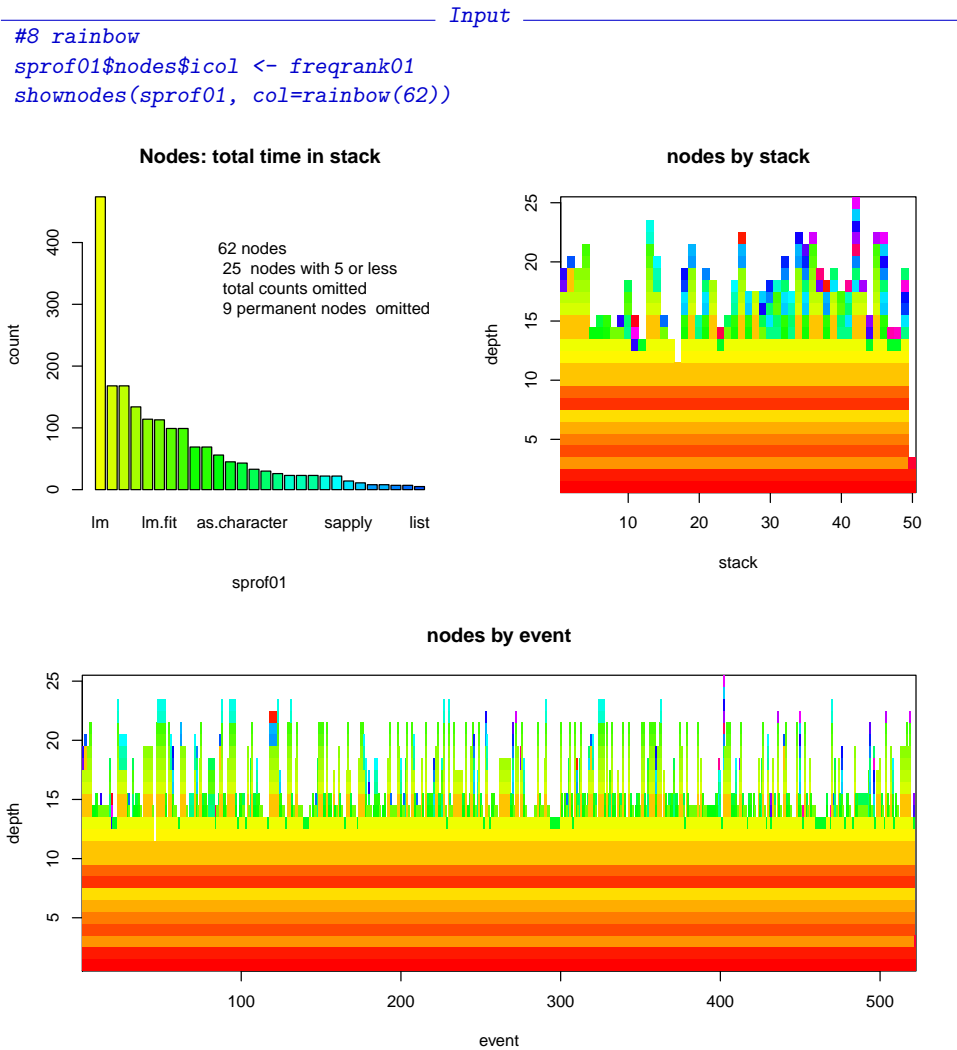


FIGURE 7. Nodes by stack and profile

See fig. 7 on the preceding page.

The obvious message is that if seen by stack level, there are different structures. Profiling usually takes place in a framework. So at the base of the stacks, we find entries that are (almost) persistent. Then usually we have some few steps where the algorithm splits, and then we have the finer details. These can be identified using information on the stack level, but of course they are not visible on the node or edge level.

Not so often, but a frequent phenomenon is to have some “burn in” or “fade out”. To identify this, we need to look at the profile level.

At a closer look, we may find stack patterns (maybe marked by specific nodes) that indicate administrative intervention and rather should be handled as separators between distinct profiles rather than as part of the general dynamics. Stable framework effect can be detected automatically. “burn in” or “fade out” may need a closer look, and special stacks need an individual inspection on low frequency stacks.

ToDo: example

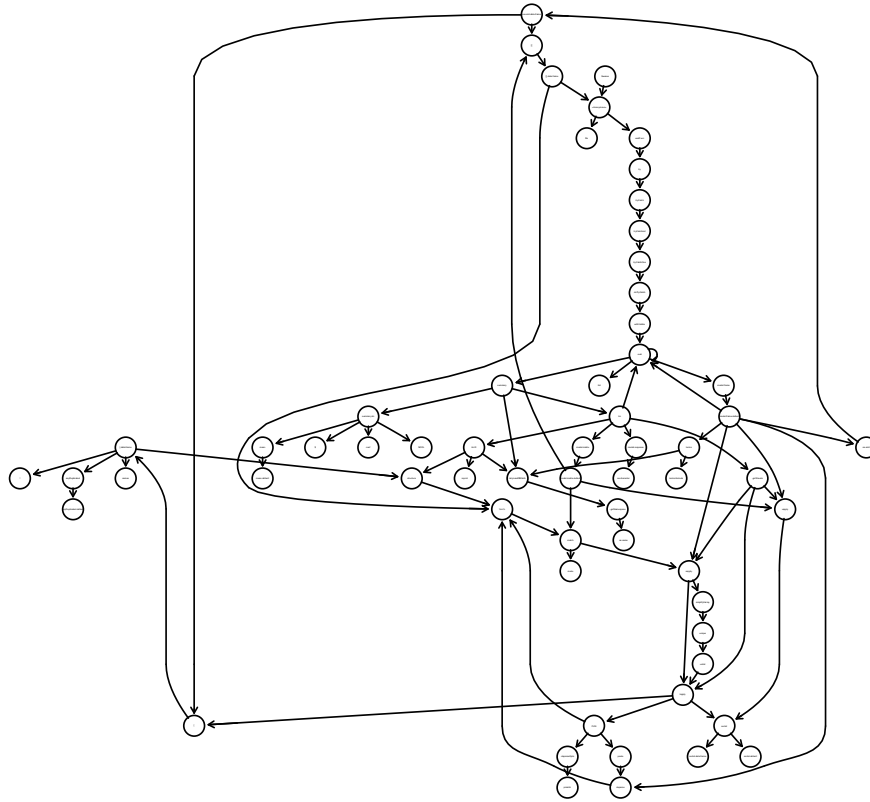
2.2. The free lunch. What you have seen so far is what you get for free when using package *sprof*. If you want to wrap up the information and look at it from a graph point of view, here is just one example. More are in section 5 on page 65. But before changing to the graph perspective, we recommend to see the next sections, not to skip them. The preview, at this point, taking package *graph* as an example:

ToDo: colours. re-colour. Propagate colour to graph.

Input

```
#6 sprofadjNEL
library(graph)
sprof01adjNEL <- as(adjacency(sprof01),"graphNEL")
plot(sprof01adjNEL, main="sprof01: graph layout example", cex.main=2)
rm(sprof01adjNEL);
# detach("package:graph")
#! sorry. still needed by Rgraphviz -- clean up
```

sprof01: graph layout example



2.3. **Cheap thrills.** Before starting additional inspection, the data better be trimmed.

```
Input
sprof02 <- sprof01
sprof02$info$id <- "sprof02"
```

```
Input
basetrim <- 13
sprof02$stacks$nodes <- sapply(sprof02$stacks$nodes,
  function (x){if (length(x)> basetrim) x[-(1:basetrim)] })
```

ToDo: handle empty stacks and zero counts gracefully

ToDo: add ?? function

At this point, it is a decision whether to adapt the timing information, or keep the original information. Since this decision does affect the structural information, it is not critical. But analysis is easier if unused nodes are eliminated.

```
Input
strx(sprof02$info)
```

Output

```
'data.frame':      1 obs. of  8 variables:
 $ id : chr "sprof02"
 $ date : POSIXct, format: "2013-07-27 18:07:20"
 $ nrnodes : int 62
 $ nrstacks : int 50
 $ nrrecords: int 522
 $ firstline: Factor w/ 1 level "sample.interval=1000": 1
 [list output truncated]
```

Input

```
print(xtable(sprof02$nodes, caption="sprof02, before update",
  label="tab:sprof02info1"),floating=FALSE,
  tabular.environment="longtable",
  caption.placement="top",
  NA.string="\\vdots")
```

Table 6: sprof02, before update

	name	self.time	self.pct	total.time	total.pct	icol
1	!	2.00	0.38	2.00	0.03	51
2	..getNamespace	0.00	0.00	1.00	0.01	53
3	.deparseOpts	2.00	0.38	4.00	0.05	41
4	.getXlevels	0.00	0.00	26.00	0.34	27
5	[0.00	0.00	99.00	1.29	18
6	[.data.frame	57.00	10.92	99.00	1.29	19
7	[[0.00	0.00	8.00	0.10	36
8	[[.data.frame	1.00	0.19	8.00	0.10	35
9	%in%	1.00	0.19	4.00	0.05	43
10	<Anonymous>	6.00	1.15	522.00	6.79	2
11	\$	1.00	0.19	1.00	0.01	60
12	anyDuplicated	1.00	0.19	23.00	0.30	30
13	anyDuplicated.default	22.00	4.21	22.00	0.29	31
14	as.character	43.00	8.24	43.00	0.56	24
15	as.list	0.00	0.00	23.00	0.30	29
16	as.list.data.frame	22.00	4.21	22.00	0.29	32
17	as.list.default	1.00	0.19	1.00	0.01	57
18	as.name	1.00	0.19	1.00	0.01	62
19	coef	1.00	0.19	1.00	0.01	55
20	deparse	1.00	0.19	2.00	0.03	49
21	doTryCatch	0.00	0.00	521.00	6.78	3
22	eval	1.00	0.19	521.00	6.78	9
23	evalFunc	0.00	0.00	521.00	6.78	7
24	file	1.00	0.19	1.00	0.01	61
25	FUN	1.00	0.19	7.00	0.09	37
26	lapply	2.00	0.38	30.00	0.39	26
27	lazyLoadDBfetch	2.00	0.38	3.00	0.04	44
28	list	5.00	0.96	5.00	0.07	39
29	lm	42.00	8.05	474.00	6.16	12
30	lm.fit	87.00	16.67	113.00	1.47	17

31	match	1.00	0.19	11.00	0.14	34
32	mean	0.00	0.00	2.00	0.03	45
33	mean.default	2.00	0.38	2.00	0.03	48
34	mode	2.00	0.38	2.00	0.03	52
35	model.frame	0.00	0.00	168.00	2.18	13
36	model.frame.default	12.00	2.30	168.00	2.18	14
37	model.matrix	0.00	0.00	69.00	0.90	21
38	model.matrix.default	51.00	9.77	69.00	0.90	20
39	model.response	13.00	2.49	56.00	0.73	22
40	na.omit	20.00	3.83	134.00	1.74	15
41	na.omit.data.frame	26.00	4.98	114.00	1.48	16
42	names	2.00	0.38	2.00	0.03	50
43	NCOL	1.00	0.19	1.00	0.01	56
44	paste	0.00	0.00	1.00	0.01	59
45	pmatch	2.00	0.38	2.00	0.03	46
46	rep.int	7.00	1.34	7.00	0.09	38
47	sapply	1.00	0.19	14.00	0.18	33
48	simplify2array	0.00	0.00	4.00	0.05	40
49	structure	32.00	6.13	33.00	0.43	25
50	summary	0.00	0.00	520.00	6.76	11
51	summary.lm	40.00	7.66	45.00	0.59	23
52	Sweave	0.00	0.00	522.00	6.79	1
53	terms	0.00	0.00	2.00	0.03	47
54	terms.formula	1.00	0.19	1.00	0.01	58
55	try	0.00	0.00	521.00	6.78	4
56	tryCatch	0.00	0.00	521.00	6.78	6
57	tryCatchList	0.00	0.00	521.00	6.78	8
58	tryCatchOne	0.00	0.00	521.00	6.78	10
59	unique	3.00	0.57	4.00	0.05	42
60	unlist	0.00	0.00	1.00	0.01	54
61	vapply	3.00	0.57	23.00	0.30	28
62	withVisible	0.00	0.00	521.00	6.78	5

Input

```
#str(sprof02)
```

Input

```
sprof02 <- updateRprof(sprof02)
sprof02$info$id <- "sprof02 updated"
```

Input

```
#xtable(summary(sprof02))
#summary(sprof02)
strx(sprof02$info)
```

Output

```
'data.frame':      1 obs. of  9 variables:
 $ id : chr "sprof02 updated"
 $ date : POSIXct, format: "2013-07-27 18:07:20"
```

```
$ nrnodes : int 62
$ nrstacks : int 50
$ nrrecords : int 522
$ firstline : Factor w/ 1 level "sample.interval=1000": 1
[list output truncated]
```

```
Input
prxt(sprof02$nodes, caption="sprof02, after update",
     label="tab:sprof02info2")
```

Table 7: sprof02, after update

	name	self.time	self.pct	total.time	total.pct	icol
1	!	2.00	0.46	2.00	0.12	51
2	..getNamespace	0.00	0.00	1.00	0.06	53
3	.deparseOpts	2.00	0.46	4.00	0.25	41
4	.getXlevels	0.00	0.00	26.00	1.61	27
5	[0.00	0.00	99.00	6.15	18
6	[[.data.frame	57.00	13.01	99.00	6.15	19
7	[[0.00	0.00	8.00	0.50	36
8	[[.data.frame	1.00	0.23	8.00	0.50	35
9	%in%	1.00	0.23	4.00	0.25	43
10	<Anonymous>	6.00	1.37	6.00	0.37	2
< cut >	\vdots	:	:	:	:	:
53	terms	0.00	0.00	2.00	0.12	47
54	terms.formula	1.00	0.23	1.00	0.06	58
55	try	0.00	0.00	0.00	0.00	4
56	tryCatch	0.00	0.00	0.00	0.00	6
57	tryCatchList	0.00	0.00	0.00	0.00	8
58	tryCatchOne	0.00	0.00	0.00	0.00	10
59	unique	3.00	0.68	4.00	0.25	42
60	unlist	0.00	0.00	1.00	0.06	54
61	vapply	3.00	0.68	23.00	1.43	28
62	withVisible	0.00	0.00	0.00	0.00	5

```
Input
```

See fig. 8 on the next page.

ToDo: trimexample

2.3.1. *Trimming.*

```
Input
trimstacks <- function(sprof, level){
  lapply(sprof$stacks$nodes, function(x) {x[-(1:level)]})
}
```

```
Input
```

#8 *Input*

`shownodes(sprof02)`

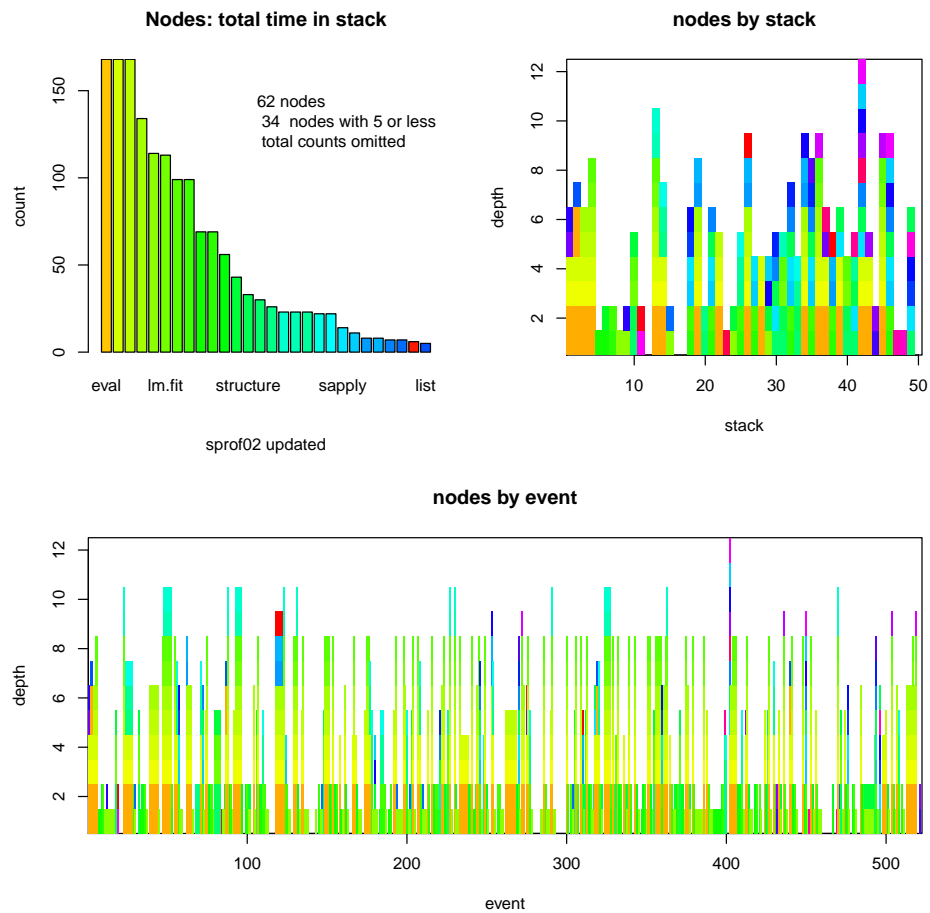


FIGURE 8. Nodes by stack and profile

```
sprof01Tr <- trimstacks(sprof01, 11)
#profile_nodesTr <- profiles_matrix(sprof01Tr)
#image(x=1:ncol(profile_nodesTr),y=1:nrow(profile_nodesTr), t(profile_nodesTr),xlab="event", ylab="
```

Input

```
nodefreq <- rep(0,length(sprof01$nodes$name))
for (i in (1:length(sprof01$stacks$nodes))) {
  nodefreq <- nodefreq +
    table( factor(sprof01$stacks$nodes[[i]],
                  levels <- 1:length(sprof01$nodes$name),
                  ordered=FALSE))
```

```

    }
    names(nodefreq) <- sprof01$nodes$name

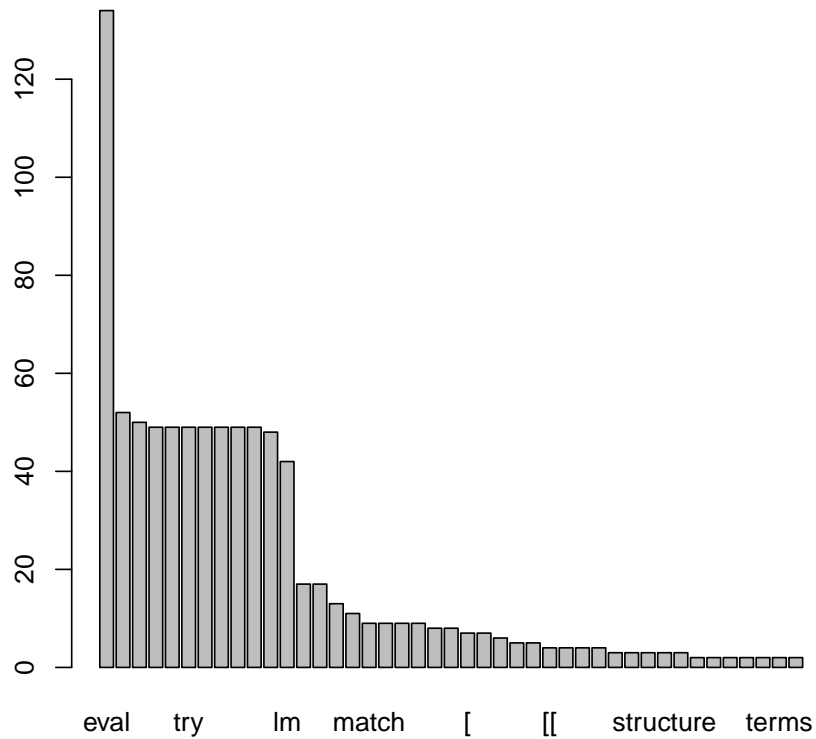
```

Top frequent nodes.

```

ndf <- nodefreq[nodefreq>1]
ondf <- order(ndf,decreasing=TRUE)
barplot(ndf[ondf])

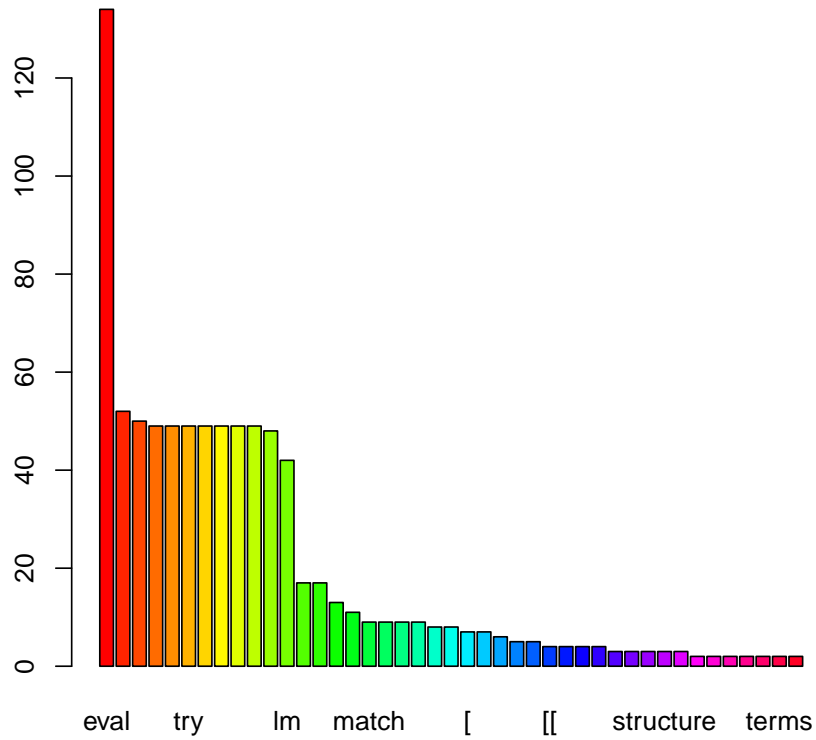
```



```

barplot(ndf[ondf], col=rainbow(length(ondf)))

```



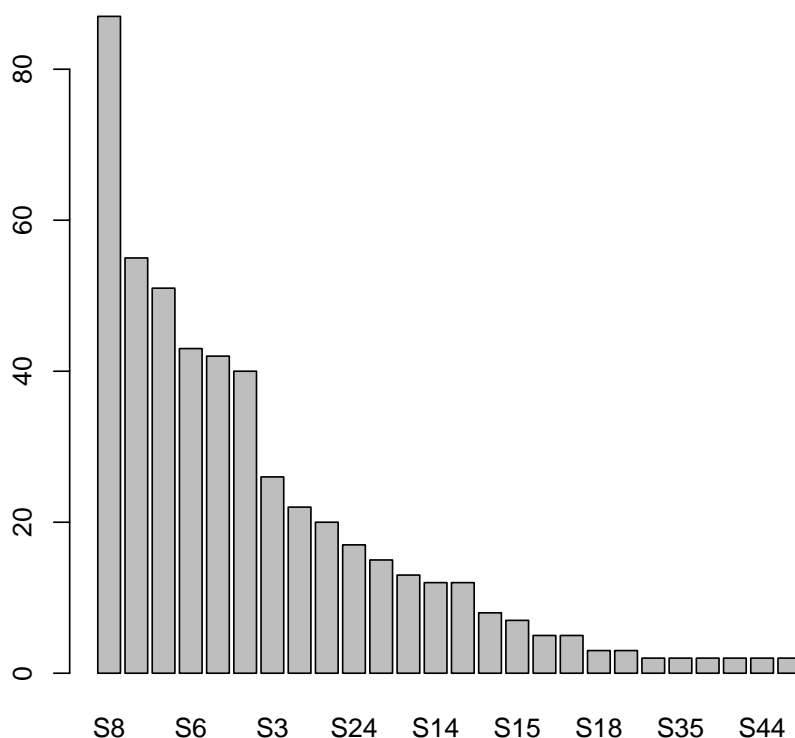
Top frequent stacks.

Input

```

x <- sprofo1
xsrc <- as.matrix(x$stacks$refcount)
rownames(xsrc) <- rownames(xsrc, do.NULL=FALSE, prefix="S")
#stf <- x$stacks$refcount[x$stacks$refcount>1]
#names(stf) <- x$stacks$shortname[x$stacks$refcount>1]
stf <- xsrc[xsrc>1]
names(stf) <- rownames(xsrc)[xsrc>1]
ostf <- order(stf, decreasing=TRUE)
barplot(stf[ostf])

```



There is no statistics on profiles. Profiling are our elementary data. However we can link to our derived data to get a more informative display. For example, going one step back we can encode stacks and use these color codes in the display of a profile.

Or going two steps back, we can encode nodes in color, giving colored stacks, and use these in the display of profile data.

2.3.2. Surgery. Looking at nodes gives you a point-wise horizon. Looking at edges gives you a one step horizon. The stacks give a wider horizon, typically a step size of 10 or more. The stacks we get from R have peculiarities, and we can handle with this broader perspective. These are not relevant if we look point-wise, but may become dominating if we try to get a global picture. We take a look ahead (details to come in section 5 on page 65 and have a preview how our example is represented as a graph. Left is the original graph as recovered from the edge information, right the graph after we have cut off the scaffold effects. Control structures may be represented in R as function, and these may lead to concentration points. Using information from the stacks, we can avoid these by introducing substitute nodes on the stack level. For example, `"[" "lapply" ".getXlevels" -> "<.getXlevels_["` If the node

ToDo: cut next level

ToDo: function
addnode to be
added

ToDo: Implement.
Currently best han-
dled on source=text
level

does not exist, we want to add it to our global variable. For now, we do it using expressions on the basic level.

```

sprof <- sprof01
node <- "<.getXlevels_>"
#nodei <- function(sprofx, node, warn = TRUE)
{
  i <- match(node, sprof$nodes$name, nomatch=0)
  if (i==0){
    sprof$nodes$name <- as.character(sprof$nodes$name)
    sprof$nodes <- rbind(sprof$nodes,NA)
    i <- length(sprof$nodes$name)
    sprof$nodes$name[i] <- node
    if (as.logical(options("warn"))){
      message("node added. An updateRprof() may be necessary.")
    }
    nodei <- i
  }
}
# sprof <- sprof01; nodei(sprof,"kiki"); sprof$nodes

```

No we have to identify the stacks that may get a replacement. First find the candidates.

```

targeti <- match(".getXlevels", sprof$nodes$name, nomatch=0)
found <- lapply(sprof$stacks$nodes, function(X) match(targeti,X))

```

ToDo: implement replacement on the stack level.

ToDo: implement

For now, these are just candidates. "as.list" "vapply" "model.frame.default" -> "<model_as.list>"

"as.list" "vapply" "model.matrix.default" -> "<model_matrix_as.list>"

```

newchopnode <- function(nodenames, chop) {
  tmpname <- paste("<",as.character(nodenames[chop]),">")
  # chec for existing.
  # add if necessary
  tmpname
}
chopstack <- function(x , chop, replacement)
{
  # is chop in x`
  # y: cut x.
  # merge x <- head + replacement + tiail
  return(x)
}

```

ToDo: needs serious revision

2.4. Run length. For a visual inspection, runs of the same node and level in the profile are easily perceived. For an analytical inspection, we have to reconstruct the runs from the data. In stacks, runs are organized hierarchically. On the root level, runs are just ordinary runs. On the next levels, runs have to be defined given

(within) the previous runs. So we need a recursive version of `rle`, applied to the profile information. This gives a detailed information about the presence time of each node, by stack level.

Input

```
profile_nodes <- profiles_matrix(sprof02)
profile_nodes_rle <- rrle(profile_nodes)
strx(profile_nodes_rle)
```

Output

```
List of 12
 $ :List of 2
  ..$ lengths: int [1:361] 6 3 1 7 1 1 1 1 ...
  ..$ values : int [1:361] 22 39 37 30 4 2 NA NA ...
  ..- attr(*, "class")= chr "rle"
 $ :List of 2
  ..$ lengths: int [1:407] 6 1 1 1 1 1 1 1 ...
  ..$ values : int [1:407] 22 NA NA 14 38 NA 27 NA ...
  ..- attr(*, "class")= chr "rle"
 $ :List of 2
  ..$ lengths: int [1:427] 6 1 1 1 1 1 1 1 ...
  ..$ values : int [1:427] 35 NA NA NA NA NA NA NA ...
  ..- attr(*, "class")= chr "rle"
 $ :List of 2
  ..$ lengths: int [1:427] 6 1 1 1 1 1 1 1 ...
  ..$ values : int [1:427] 36 NA NA NA NA NA NA NA ...
  ..- attr(*, "class")= chr "rle"
 $ :List of 2
  ..$ lengths: int [1:450] 1 2 3 1 1 1 1 1 ...
  ..$ values : int [1:450] 53 22 40 NA NA NA NA NA ...
  ..- attr(*, "class")= chr "rle"
 $ :List of 2
  ..$ lengths: int [1:466] 1 2 3 1 1 1 1 1 ...
  ..$ values : int [1:466] 27 22 41 NA NA NA NA NA ...
  ..- attr(*, "class")= chr "rle"
 [list output truncated]
```

On a given stack level, the run length is the best information on the time used per call, and the run count of a node is the best information on the number of calls. So this is a prime starting point for in-depth analysis.

Input

```
# side effect: NAs are removed
profile_nodes_rlet <- lapply(profile_nodes_rle,
  function(x) table(x,dnn=c("run length","node"))) )
invisible(lapply(profile_nodes_rlet,
  function(x) print.table(x,zero.print = ".") ))
```

Output

```
      node
run length  2  4 11 19 22 30 32 37 39 43
      1  1 17  1  1 40 46  2 55 35  1
      2  .  1  .  . 17 18  .  4  3  .
```

ToDo: keep as factor. This is a sparse cube with margins node, stack level, run length. Nodes are mostly concentrated on few levels. **ToDo:** Warning: data structure still under discussion

```

3 . 1 . . 6 3 . 2 3 .
4 . 1 . . 4 1 . . . .
5 . . . . 2 1 . . . .
6 . . . . 6 1 . . 1 .
7 . . . . 2 1 . . . .
node
run length 14 18 22 26 27 33 38 46 47 49 61
1 34 1 40 16 1 2 55 7 3 10 .
2 3 . 17 . . . 4 . . 1 .
3 1 . 6 . . . 2 . . . 1
4 . . 4 1 . . . . . .
5 . . 2 . . . . . . .
6 . . 6 . . . . . 1 .
7 . . 2 . . . . . . .
node
run length 5 7 9 15 26 31 35 48 61
1 14 1 1 2 2 9 40 1 6
2 . . . . . 17 . 1
3 . . . . . 6 . .
4 1 . . . . 4 . .
5 . . . . . 2 . .
6 . . . . . 6 . .
7 . . . . . 2 . .
node
run length 6 8 15 16 25 31 36 47 59
1 14 1 7 2 1 1 40 9 1
2 . . 1 . . . 17 . .
3 . . . . . 6 . .
4 1 . . . . 4 . .
5 . . . . . 2 . .
6 . . . . . 6 . .
7 . . . . . 2 . .
node
run length 3 10 12 16 17 20 22 26 40 47 48 49 53 60 61
1 1 1 1 6 1 1 3 5 46 2 3 11 2 1 7
2 . . . 1 . . 1 . 10 . . . . .
3 . . . . . . . 5 . . . . .
4 . . . . . . . 4 . . 1 . . .
5 . . . . . . . 1 . . . . 1
6 . . . . . . . 3 . . . . .
7 . . . . . . . 2 . . . . .
node
run length 15 22 25 26 27 41 54 59
1 7 3 5 3 1 43 1 3
2 . 1 . . . 7 . .
3 . . . . . 4 . .
4 . . . . . 5 . .
5 1 . . . . . .
6 . . . . . 3 . .
7 . . . . . 1 . .
node
run length 3 5 7 9 16 25 28
1 3 46 2 1 7 1 3

```

```

      2 . 4 . . . . 1
      3 . 3 . . . . .
      4 . 3 . . . . .
      5 . . 1 . 1 . .
      6 . 1 . . . . .
      node
run length 6 8 31 44 45
      1 46 2 1 1 2
      2 4 . . . .
      3 3 . . . .
      4 3 . . . .
      5 . 1 . . .
      6 1 . . . .
      node
run length 1 9 10 12 20 34 42
      1 2 1 . 9 1 1 2
      4 . . . 2 . . .
      5 . . 1 1 . . .
      node
run length 9 13
      1 1 9
      4 . 2
      5 . 1
      node
run length 31
      1 1
      node
run length 34
      1 1

```

This is a poor first attempt to tame `profile_nodes_rlet`.

```

Input
-----
maxnode <-0
maxlen <-0
maxlevel <-length(profile_nodes_rle)
for (lev in (1:maxlevel) ) {
  proflev <- profile_nodes_rle[[lev]]
  if (!is.null(proflev)) {
    maxn <- max(proflev$values, na.rm=TRUE)
    if (maxn>maxnode) maxnode <- maxn
    maxl <- max(proflev$lengths, na.rm=TRUE)
    if (maxl>maxlen) maxlen <- maxl
    # cat("Level ",lev,maxn," Length:",maxl,"\n")
  }
}
## collapse profile_nodes_rle to 3d array. Allocate memory first.
profile_nodes_rlearray <- array(0,
  dim=c(maxnode,length(profile_nodes_rle), maxlen),
  dimnames= list("node"=sprof02$nodes$name[1:maxnode],
    "level"=1:length(profile_nodes_rle),
    "run_length"=1:maxlen))
strx(profile_nodes_rlearray)

```

ToDo: some mess in the code below – not working

ToDo: replace by decent vector/array based implementation

ToDo: add names for node dimension

```

num [1:61, 1:12, 1:7] 0 0 0 0 0 0 0 0 ...
- attr(*, "dimnames")=List of 3
..$ node : chr [1:61] "!" "..getNamespace" ".deparseOpts" ...
..$ level : chr [1:12] "1" "2" "3" ...
..$ run_length: chr [1:7] "1" "2" "3" ...

```

```

Input
for (lev in (1:maxlevel) ) {
  proflev <- profile_nodes_rle[[lev]]
  if (!is.null(proflev)) {
    for (j in (1: length(proflev$lengths))) {
      if (!is.na(proflev$values[j])) {
        profile_nodes_rlearray[proflev$values[j],lev,proflev$lengths[j]] <-
        profile_nodes_rlearray[proflev$values[j],lev,proflev$lengths[j]] +1
        #cat(lev,j,":",proflev$values[j],lev,proflev$lengths[j],"\n")
      }#if (!is.na
    }#for j
  }
}

```

ToDo: add
marginals and
conditionals.
Provide function
node_summary.

This allows us to extract marginal from `proflev[node, level, run length]`.

```

Input
node=41
cat(sprof02$nodes$name[node],"\n")

```

```

Output
41

```

```

Input
nn <- profile_nodes_rlearray[node, , ]
#dimnames(nn)<- list( "level", "run_length")
nn

```

```

Output
      run_length
level 1 2 3 4 5 6 7
  1    0 0 0 0 0 0 0
  2    0 0 0 0 0 0 0
  3    0 0 0 0 0 0 0
  4    0 0 0 0 0 0 0
  5    0 0 0 0 0 0 0
  6   43 7 4 5 0 3 1
  7    0 0 0 0 0 0 0
  8    0 0 0 0 0 0 0
  9    0 0 0 0 0 0 0
 10    0 0 0 0 0 0 0
 11    0 0 0 0 0 0 0
 12    0 0 0 0 0 0 0

```

```

Input

```

```
print.table(addmargins(nn), zero.print = ".")
```

Output								
	run_length							
level	1	2	3	4	5	6	7	Sum
1
2
3
4
5
6	43	7	4	5	.	3	1	63
7
8
9
10
11
12
Sum	43	7	4	5	.	3	1	63

```
mt <- margin.table(profile_nodes_rlearray, margin = c(1,3))
amt <- addmargins(mt)
amt <- amt[amt[, "Sum"] > 0,]
print.table(amt, zero.print = ".")
```

Output								
	run_length							
node	1	2	3	4	5	6	7	Sum
!	2	2
..getNamespace	1	1
.deparseOpts	4	4
.getXlevels	17	1	1	1	.	.	.	20
[60	4	3	4	.	1	.	72
[.data.frame	60	4	3	4	.	1	.	72
[[3	.	.	.	1	.	.	4
[[.data.frame	3	.	.	.	1	.	.	4
%in%	4	4
<Anonymous>	1	.	.	.	1	.	.	2
\$	1	1
anyDuplicated	10	.	.	2	1	.	.	13
anyDuplicated.default	9	.	.	2	1	.	.	12
as.character	34	3	1	38
as.list	16	1	.	.	1	.	.	18
as.list.data.frame	15	1	.	.	1	.	.	17
as.list.default	1	1
as.name	1	1
coef	1	1
deparse	2	2
eval	86	36	12	8	4	12	4	162
FUN	7	7
lapply	26	.	.	1	.	.	.	27
lazyLoadDBfetch	2	2
list	3	1	4

ToDo: rescale to application scale

ToDo: replace sum by weighted sum

ToDo: allow sorting, e.g. by marginals

lm.fit	46	18	3	1	1	1	1	71
match	12	12
mean	2	2
mean.default	2	2
mode	2	2
model.frame	40	17	6	4	2	6	2	77
model.frame.default	40	17	6	4	2	6	2	77
model.matrix	55	4	2	61
model.matrix.default	55	4	2	61
model.response	35	3	3	.	.	1	.	42
na.omit	46	10	5	4	1	3	2	71
na.omit.data.frame	43	7	4	5	.	3	1	63
names	2	2
NCOL	1	1
paste	1	1
pmatch	2	2
rep.int	7	7
sapply	14	14
simplify2array	4	4
structure	21	1	.	1	.	1	.	24
terms	2	2
terms.formula	1	1
unique	4	4
unlist	1	1
vapply	13	1	1	.	1	.	.	16
Sum	820	133	52	41	18	35	12	1111

These are some attempts to recover the factor structures.

Input

```

xfi <- levels(sprof02$nodes$name)
profile_nodes_rlefac <- lapply(profile_nodes_rle,
  function(xl) {xl$values <- factor(xl$values,
    levels=1:62,
    labels=xfi); xl}) # seems ok
profile_nodes_rletfac <- lapply(profile_nodes_rle,
  function(x) table(x,dnn=c("run length","node")) ) #factors lost again
colnames(profile_nodes_rletfac[[1]]) <-
  sprof02$nodes$name[ as.integer(colnames(profile_nodes_rletfac[[1]]))]
profile_nodes_rletfac1 <- lapply(profile_nodes_rletfac,
  function(xl) {colnames(xl) <-
    sprof02$nodes$name[ as.integer(colnames(xl))];
    xl} )
invisible(lapply(profile_nodes_rletfac1,
  function(x) print.table(t(x),zero.print = ".") ))

```

Output

```

      run length
node  1  2  3  4  5  6  7
<NA>  1  .  .  .  .  .  .
<NA> 17  1  1  1  .  .  .
<NA>  1  .  .  .  .  .  .
<NA>  1  .  .  .  .  .  .

```

```

<NA> 40 17 6 4 2 6 2
<NA> 46 18 3 1 1 1 1
<NA> 2 . . . . .
<NA> 55 4 2 . . . .
<NA> 35 3 3 . . 1 .
<NA> 1 . . . . .

run length
node      1 2 3 4 5 6 7
as.character 34 3 1 . . . .
as.name      1 . . . . .
eval         40 17 6 4 2 6 2
lapply       16 . . 1 . . .
lazyLoadDBfetch 1 . . . . .
mean.default 2 . . . . .
model.matrix.default 55 4 2 . . . .
rep.int       7 . . . . .
sapply        3 . . . . .
structure     10 1 . . . 1 .
vapply        . . 1 . . . .

run length
node      1 2 3 4 5 6 7
[         14 . . 1 . . .
[[        1 . . . . .
%in%      1 . . . . .
as.list   2 . . . . .
lapply    2 . . . . .
match     9 . . . . .
model.frame 40 17 6 4 2 6 2
simplify2array 1 . . . . .
vapply    6 1 . . . . .

run length
node      1 2 3 4 5 6 7
[.data.frame 14 . . 1 . . .
[.data.frame 1 . . . . .
as.list      7 1 . . . . .
as.list.data.frame 2 . . . . .
FUN          1 . . . . .
match        1 . . . . .
model.frame.default 40 17 6 4 2 6 2
sapply        9 . . . . .
unique        1 . . . . .

run length
node      1 2 3 4 5 6 7
.deparseOpts 1 . . . . .
<Anonymous> 1 . . . . .
anyDuplicated 1 . . . . .
as.list.data.frame 6 1 . . . .
as.list.default 1 . . . . .
deparse       1 . . . . .
eval          3 1 . . . . .
lapply        5 . . . . .
na.omit       46 10 5 4 1 3 2
sapply        2 . . . . .

```

```

simplify2array      3 . . . . .
structure          11 . . 1 . . .
terms              2 . . . . .
unlist             1 . . . . .
vapply             7 . . . 1 . .

run length
node              1 2 3 4 5 6 7
as.list           7 . . . 1 . .
eval             3 1 . . . . .
FUN             5 . . . . .
lapply           3 . . . . .
lazyLoadDBfetch  1 . . . . .
na.omit.data.frame 43 7 4 5 . 3 1
terms.formula     1 . . . . .
unique            3 . . . . .

run length
node              1 2 3 4 5 6
.deparseOpts      3 . . . . .
[                46 4 3 3 . 1
[[               2 . . . 1 .
%in%             1 . . . . .
as.list.data.frame 7 . . . 1 .
FUN              1 . . . . .
list             3 1 . . . .

run length
node              1 2 3 4 5 6
[.data.frame     46 4 3 3 . 1
[[.data.frame    2 . . . 1 .
match            1 . . . . .
paste            1 . . . . .
pmatch          2 . . . . .

run length
node              1 4 5
!                2 . .
%in%             1 . .
<Anonymous>     . . 1
anyDuplicated    9 2 1
deparse         1 . .
mode            1 . .
names           2 . .

run length
node              1 4 5
%in%             1 . .
anyDuplicated.default 9 2 1

run length
node      1
match 1

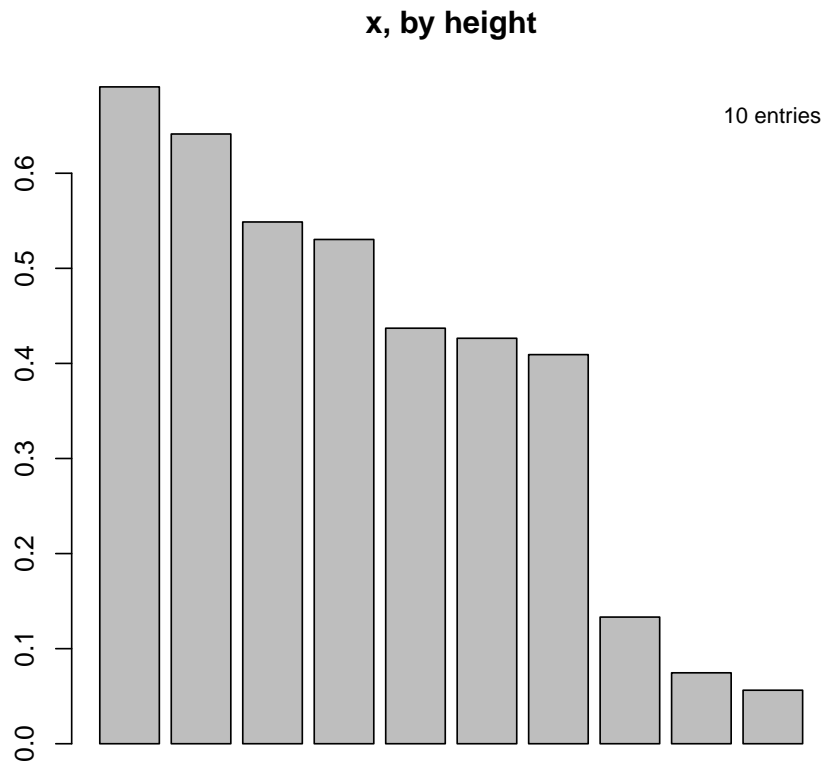
run length
node      1
mode 1

```


3. XXX – LOST & FOUND

Input

```
barplot_s(runif(10))
```



ToDo: add current

ToDo: generate a coplot representation

ToDo: add time per call information: add marginals statistics run time by node

ToDo: table: node #runs min median run length max

Input

```
# xtable cannot handle posix, so we use print output here
# str(profile_nodes_rle, max.level=2, vec.len=3, nchar.max=40, list.len=6)
strx(sprof01$info)
```

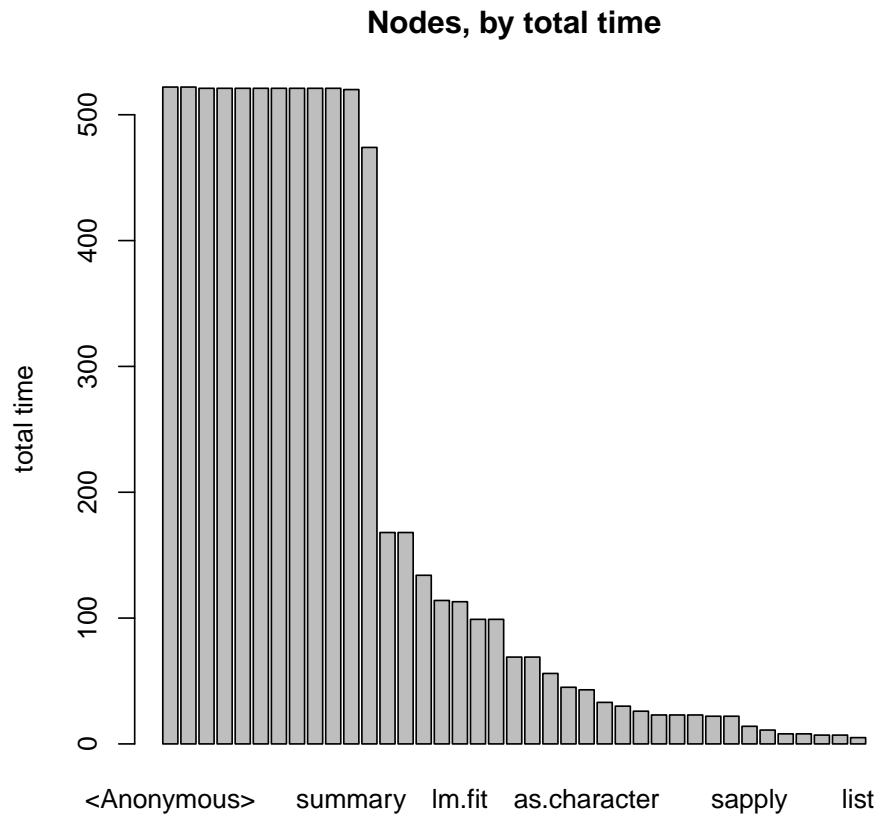
Output

```
'data.frame':      1 obs. of  8 variables:
 $ id : chr "sprof01"
 $ date : POSIXct, format: "2013-07-27 18:07:20"
 $ nrnodes : int 62
 $ nrstacks : int 50
 $ nrrecords: int 522
```

```
$ firstline: Factor w/ 1 level "sample.interval=1000": 1
[list output truncated]
```

Selections are recorded as selection vectors, with reference to the original order. This needs some caution to align them with the order choices.

```
Input
rownames(sprof01$nodes) <- sprof01$nodes$names
nodesperm <- order(sprof01$nodes$total.time,decreasing=TRUE)
nodesnrobsok <- sprof01$nodes$total.time > 4
sp <- sprof01$nodes$total.time[nodesperm][nodesnrobsok[nodesperm]]
names(sp) <- sprof01$nodes$name[nodesperm][nodesnrobsok[nodesperm]]
barplot(sp,
  main="Nodes, by total time", ylab="total time")
```

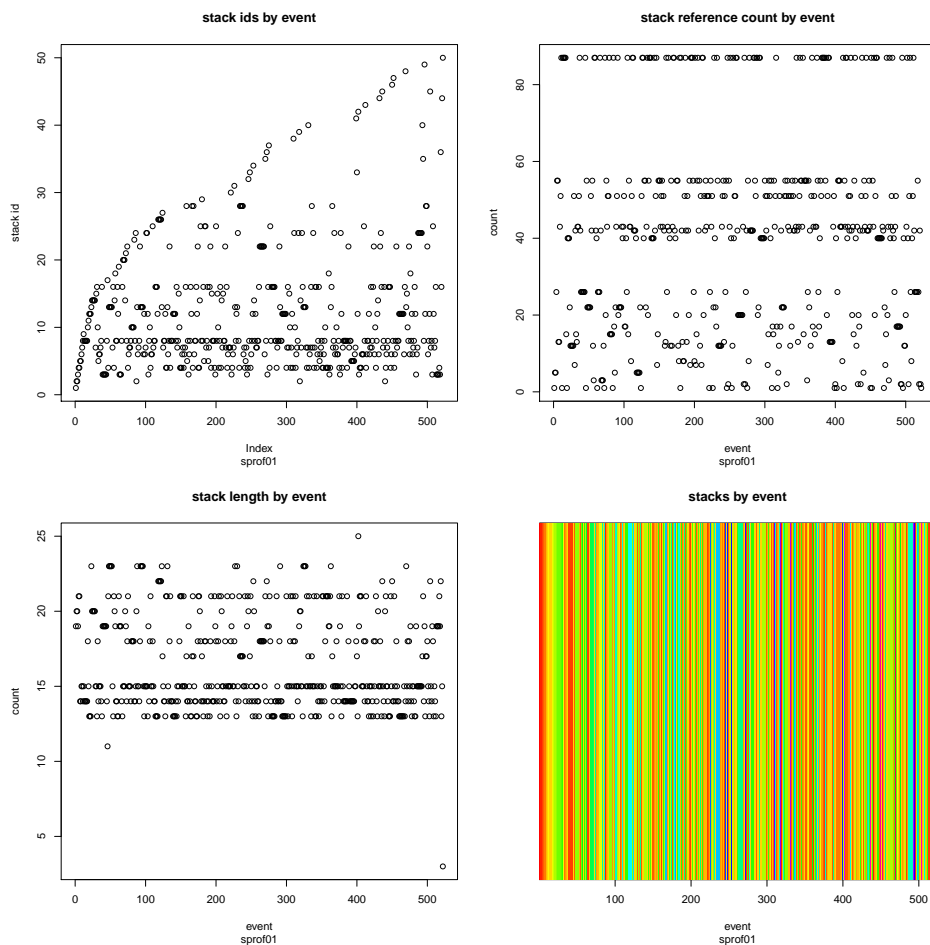


On the first look, information on the profile level is not informative. Profile records are just recordings of some step, taken at regular intervals. We get a minimal information, if we encode the stacks in colour.

ToDo: use stack colours

Input

```
oldpar<-par(mfrow=c(2,2))
plot_profiles(sprof01)
par(oldpar)
```

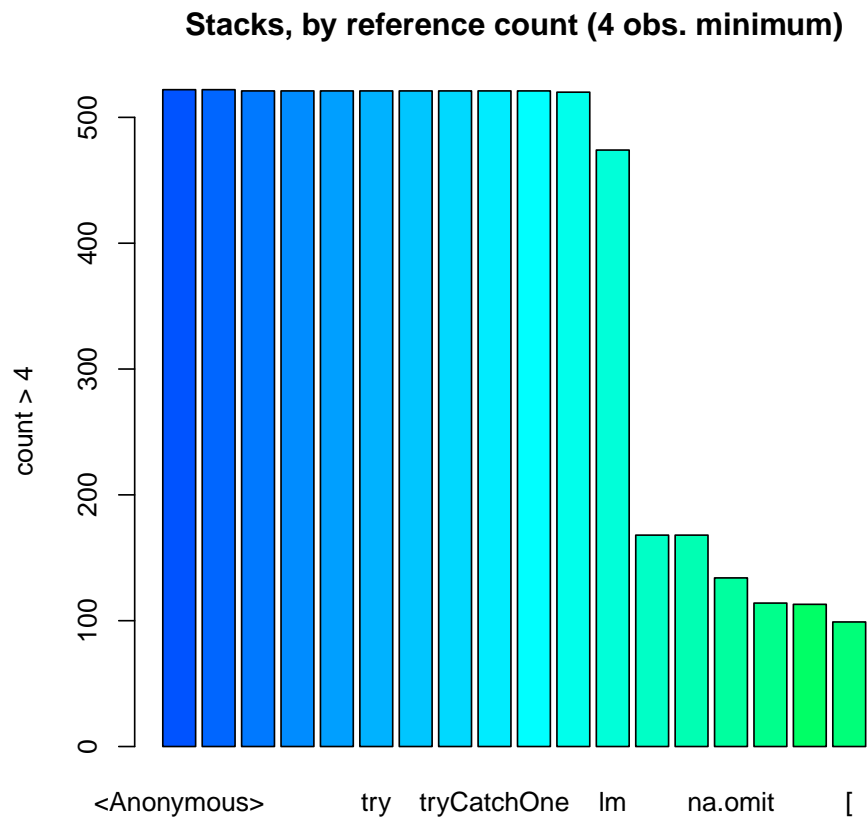


We now do a step down analysis. Aggregating the information from the profiling events, we have the frequency of stack references. On the stack level, we encode the frequency in color, and linking propagates this to the profile level.

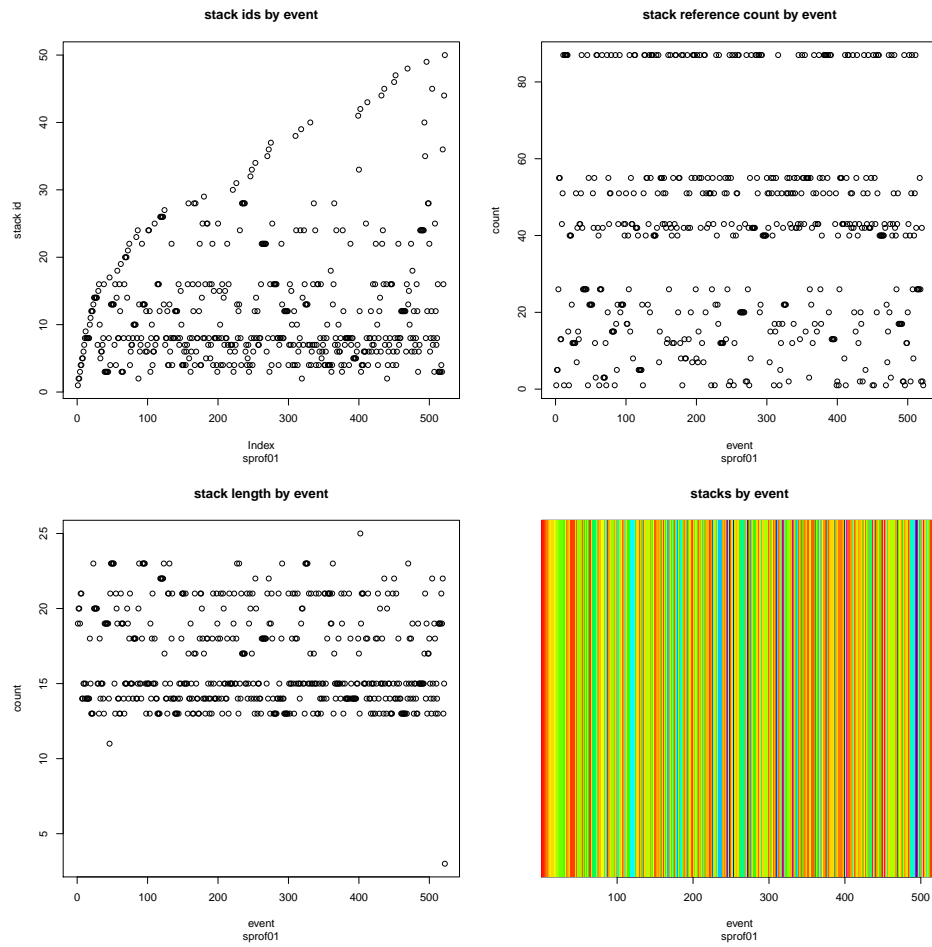
Input

```
stackfreqscore <- rank(sprof01$stacks$refcount,ties.method="random")
stacksperm <- order(sprof01$stacks$refcount,decreasing=TRUE)
stacksnrobsok <- sprof01$stacks$refcount > 4
stackfreqscore4<- stackfreqscore[stacksperm][stacksnrobsok[stacksperm]]
barplot(sp[stacksnrobsok[stacksperm]]),
```

```
main="Stacks, by reference count (4 obs. minimum)", ylab="count > 4",
col=rainbow(80)[stackfreqscore4])
```



```
oldpar<- par(mfrow=c(2,2))      Input
plot_profiles(sprof01)
par(oldpar)
```



Input

```
prxt(sprof01$nodes,
     caption="nodes",
     label="tab:prSREnodes",
     max.level=2, vec.len=3,nchar.max=40)
```

Table 8: nodes

	name	self.time	self.pct	total.time	total.pct	icol
1	!	2.00	0.38	2.00	0.03	51
2	..getNamespace	0.00	0.00	1.00	0.01	53
3	.deparseOpts	2.00	0.38	4.00	0.05	41
4	.getXlevels	0.00	0.00	26.00	0.34	27
5	[0.00	0.00	99.00	1.29	18
6	[.data.frame	57.00	10.92	99.00	1.29	19
7	[[0.00	0.00	8.00	0.10	36
8	[[.data.frame	1.00	0.19	8.00	0.10	35
9	%in%	1.00	0.19	4.00	0.05	43
10	<Anonymous>	6.00	1.15	522.00	6.79	2
< cut >	\vdots	⋮	⋮	⋮	⋮	⋮
53	terms	0.00	0.00	2.00	0.03	47
54	terms.formula	1.00	0.19	1.00	0.01	58
55	try	0.00	0.00	521.00	6.78	4
56	tryCatch	0.00	0.00	521.00	6.78	6
57	tryCatchList	0.00	0.00	521.00	6.78	8
58	tryCatchOne	0.00	0.00	521.00	6.78	10
59	unique	3.00	0.57	4.00	0.05	42
60	unlist	0.00	0.00	1.00	0.01	54
61	vapply	3.00	0.57	23.00	0.30	28
62	withVisible	0.00	0.00	521.00	6.78	5

```

#str(sprof01$stacks, max.level=2, vec.len=6,
#      nchar.max=40, list.len=20,width=70, strict.width="wrap"
strx(sprof01$stacks)

```

```

'data.frame':      50 obs. of  7 variables:
 $ nodes :List of 50
 ..$ : int 52 10 23 55 56 57 58 21 ...
 ..$ : int 52 10 23 55 56 57 58 21 ...
 ..$ : int 52 10 23 55 56 57 58 21 ...
 ..$ : int 52 10 23 55 56 57 58 21 ...
 ..$ : int 52 10 23 55 56 57 58 21 ...
 ..$ : int 52 10 23 55 56 57 58 21 ...
 .. [list output truncated]
 $ shortname : Factor w/ 50 levels
   "S<A>eFttCtCLtCOdTCwVeesleem.m..n.n...["| __truncated__,...: 27 17
   19 1 35 36 37 30 ...
 $ refcount : num 1 5 26 55 13 43 51 87 ...
 $ stacklength : int 19 20 19 21 14 15 15 14 ...
 $ stackheadnodes: int 52 52 52 52 52 52 52 52 ...
 $ stackleafnodes: int 27 28 41 6 39 14 38 30 ...
 [list output truncated]

```

A summary is provided on request.

```

sumsprof01 <- summary.sprof(sprof01)

```

```

$id
[1] "Profile Summary Sat Jul 27 18:07:23 2013"

$len
[1] 522

$uniquestacks
[1] 50

$nr_runs
[1] 396

$nrstacks
[1] 50

$stacklength
[1] 3 25

$nrnodesperlevel
[1] 1 1 2 1 1 1 1 1 1 1 1 1 3 10 11 9 9 15 8 7 5 7
[23] 2 1 1

          shortname root leaf self.time  self.pct
!              !      - LEAF          2 0.383142

```

..getNamespace	..gN	-	-	0	0.000000
.deparseOpts	.dp0	-	LEAF	2	0.383142
.getXlevels	.gtX	-	-	0	0.000000
[[-	-	0	0.000000
[.data.frame	[.d.	-	LEAF	57	10.919540
[[[[-	-	0	0.000000
[[.data.frame	[[..	-	LEAF	1	0.191571
%in%	%in%	-	LEAF	1	0.191571
<Anonymous>	<An>	-	LEAF	6	1.149425
\$	\$	-	LEAF	1	0.191571
anyDuplicated	anyD	-	LEAF	1	0.191571
anyDuplicated.default	anD.	-	LEAF	22	4.214559
as.character	as.c	-	LEAF	43	8.237548
as.list	as.l	-	-	0	0.000000
as.list.data.frame	a...	-	LEAF	22	4.214559
as.list.default	as..	-	LEAF	1	0.191571
as.name	as.n	-	LEAF	1	0.191571
coef	coef	-	LEAF	1	0.191571
deparse	dprs	-	LEAF	1	0.191571
doTryCatch	dTrC	-	-	0	0.000000
eval	eval	-	LEAF	1	0.191571
evalFunc	evlF	-	-	0	0.000000
file	file	-	LEAF	1	0.191571
FUN	FUN	-	LEAF	1	0.191571
lapply	lppl	-	LEAF	2	0.383142
lazyLoadDBfetch	lLDB	-	LEAF	2	0.383142
list	list	-	LEAF	5	0.957854
lm	lm	-	LEAF	42	8.045977
lm.fit	lm.f	-	LEAF	87	16.666667
match	mtch	-	LEAF	1	0.191571
mean	mean	-	-	0	0.000000
mean.default	mn.d	-	LEAF	2	0.383142
mode	mode	-	LEAF	2	0.383142
model.frame	mdl.f	-	-	0	0.000000
model.frame.default	mdl.f.	-	LEAF	12	2.298851
model.matrix	mdl.m	-	-	0	0.000000
model.matrix.default	mdl.m.	-	LEAF	51	9.770115
model.response	mdl.r	-	LEAF	13	2.490421
na.omit	n.mt	-	LEAF	20	3.831418
na.omit.data.frame	n...	-	LEAF	26	4.980843
names	nams	-	LEAF	2	0.383142
NCOL	NCOL	-	LEAF	1	0.191571
paste	past	-	-	0	0.000000
pmatch	pmtc	-	LEAF	2	0.383142
rep.int	rp.n	-	LEAF	7	1.340996
sapply	sppl	-	LEAF	1	0.191571
simplify2array	smp2	-	-	0	0.000000
structure	strc	-	LEAF	32	6.130268
summary	smmr	-	-	0	0.000000
summary.lm	smm.	-	LEAF	40	7.662835
Sweave	Swev	ROOT	-	0	0.000000
terms	trms	-	-	0	0.000000
terms.formula	trm.	-	LEAF	1	0.191571

try	try	-	-	0	0.000000
tryCatch	tryC	-	-	0	0.000000
tryCatchList	trCL	-	-	0	0.000000
tryCatchOne	trCO	-	-	0	0.000000
unique	uniq	-	LEAF	3	0.574713
unlist	unls	-	-	0	0.000000
vapply	vppl	-	LEAF	3	0.574713
withVisible	wthV	-	-	0	0.000000
	total.time		total.pct		
!	2		0.383142		
..getNamespace	1		0.191571		
.deparseOpts	4		0.766284		
.getXlevels	26		4.980843		
[99		18.965517		
[[.data.frame	99		18.965517		
[[8		1.532567		
[[.data.frame	8		1.532567		
%in%	4		0.766284		
<Anonymous>	522		100.000000		
\$	1		0.191571		
anyDuplicated	23		4.406130		
anyDuplicated.default	22		4.214559		
as.character	43		8.237548		
as.list	23		4.406130		
as.list.data.frame	22		4.214559		
as.list.default	1		0.191571		
as.name	1		0.191571		
coef	1		0.191571		
deparse	2		0.383142		
doTryCatch	521		99.808429		
eval	521		99.808429		
evalFunc	521		99.808429		
file	1		0.191571		
FUN	7		1.340996		
lapply	30		5.747126		
lazyLoadDBfetch	3		0.574713		
list	5		0.957854		
lm	474		90.804598		
lm.fit	113		21.647510		
match	11		2.107280		
mean	2		0.383142		
mean.default	2		0.383142		
mode	2		0.383142		
model.frame	168		32.183908		
model.frame.default	168		32.183908		
model.matrix	69		13.218391		
model.matrix.default	69		13.218391		
model.response	56		10.727969		
na.omit	134		25.670498		
na.omit.data.frame	114		21.839080		
names	2		0.383142		
NCOL	1		0.191571		
paste	1		0.191571		

pmatch	2	0.383142
rep.int	7	1.340996
sapply	14	2.681992
simplify2array	4	0.766284
structure	33	6.321839
summary	520	99.616858
summary.lm	45	8.620690
Sweave	522	100.000000
terms	2	0.383142
terms.formula	1	0.191571
try	521	99.808429
tryCatch	521	99.808429
tryCatchList	521	99.808429
tryCatchOne	521	99.808429
unique	4	0.766284
unlist	1	0.191571
vapply	23	4.406130
withVisible	521	99.808429

Input

```
#str(profile_nodes_rle, max.level=2, vec.len=3, nchar.max=40, list.len=6)
strx(sumsprof01)
```

Output

```
'data.frame':      62 obs. of  7 variables:
 $ shortname : Factor w/ 62 levels "!", "..gN", ".dp0", ...: 1 2 3 4 5 6 7
   8 ...
 $ root      : Factor w/ 2 levels "-", "ROOT": 1 1 1 1 1 1 1 ...
 $ leaf      : Factor w/ 2 levels "-", "LEAF": 2 1 2 1 1 2 1 2 ...
 $ self.time : num 2 0 2 0 0 57 0 1 ...
 $ self.pct  : num 0.383 0 0.383 0 ...
 $ total.time: num 2 1 4 26 99 99 8 8 ...
 [list output truncated]
```

Input

```
#str(sumsprof01, max.level=2, vec.len=3,
#      nchar.max=40, list.len=6,
#      width=70, strict.width="wrap")
```

The classical approach hides the work that has been done. Actually it breaks down the data to record items. This figure is not reported anywhere. In our case, it can be reconstructed. The profile data have 8456 words in 524 lines.

In our approach, we break down the information. Two lines of control information are split off. We have 522 lines of profile with 50 unique stacks, referencing 62 nodes. Instead of reducing it to a summary, we keep the full information. Information is always kept on its original level.

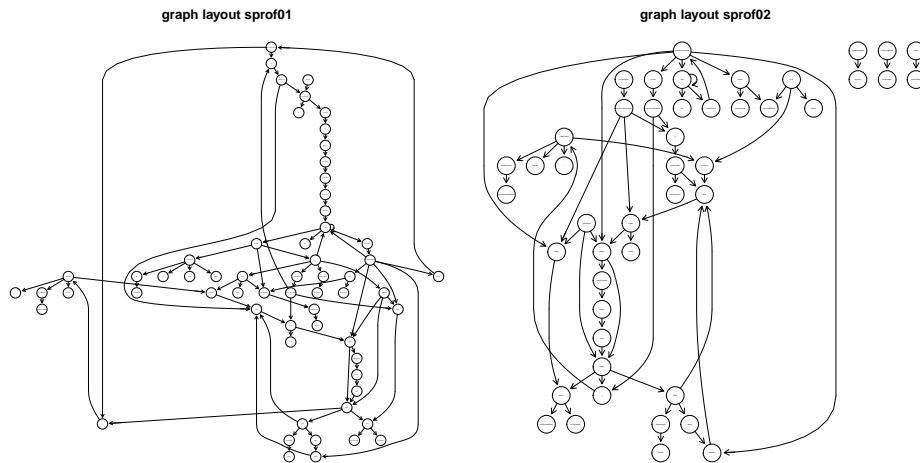
On the profiles level, we know the sample interval length, and the id of the stack recorded. On the stack level, for each stack we have a reference count, with the sample interval lengths used as weights. This reference count is added up for each node in the stack to give the node timings.

Cheap statistics are collected as they come by. For example, from the stacks table it is cheap to identify root and leaf nodes, and this mark is propagated to the nodes table.

3.1. Graph package.

Input

```
oldpar <- par(mfrow=c(1,2))
library(graph)
plot(as(adjacency(sprof01),"graphNEL"),
     main="graph layout sprof01", cex.main=2)
plot(as(adjacency(sprof02),"graphNEL"),
     main="graph layout sprof02", cex.main=2)
par(oldpar)
```



R is function based, and control structures in general are implemented as functions. In a graph representation, they appear as nodes, concentrating and seeding to unrelated paths. We can detect these on the stack level and replace them by surrogates, introducing new nodes.

ToDo: fix null name

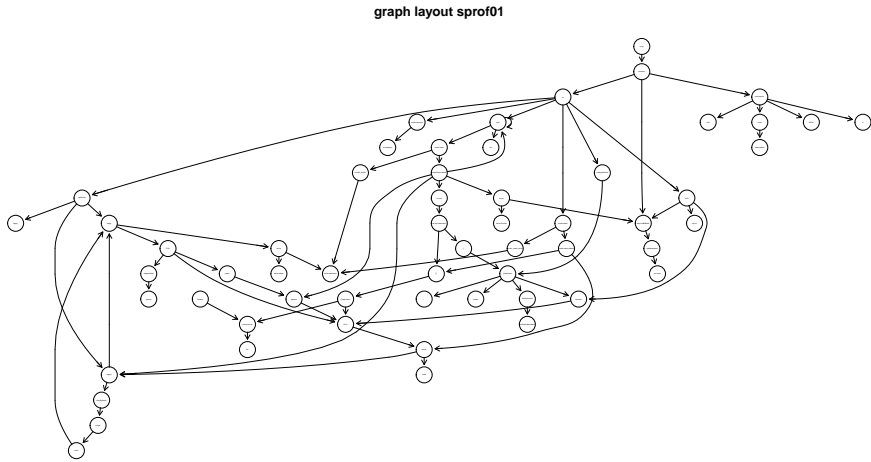
Input

```
sprof03 <- readRprof("RprofsRegressionExp103.out")
sprof03$info$id <- "sprof03"
#sprof03$nodes$name[1] <- sprof03$nodes$name[2]
#sprof03$nodes$name[1] <- "<noop>"??"
```

ToDo: cut top levels

Input

```
library(graph)
a03<-adjacency(sprof03)
rnames <- rownames(a03)
rnames[1]<-"noop";rownames(a03) <- rnames; colnames(a03) <- rnames;
plot(as(a03,"graphNEL"), main="graph layout sprof01", cex.main=2)
```



4. STANDARD OUTPUT

For a reference, here are complete outputs of the standard function.

`sprof <- sprof01`

Input

ToDo: Clarify: "print prints its argument and returns it invisibly (via invisible(x))." Return the argument, or some print representation?

ToDo: is there a print=FALSE variant to postpone printing to e.g. xtable?

4.1. **Print.** We omit the (lengthy) print output here and just give the commands as a reference.

`print_nodes(sprof)`

Input

	Output		self.time	self.pct
	shortname	root leaf		
!	!	- LEAF	2	0.383142
..getNamespace	..gN	- -	0	0.000000
..deparseOpts	..dp0	- LEAF	2	0.383142
..getXlevels	..gtX	- -	0	0.000000
[[- -	0	0.000000
[.data.frame	[.d.	- LEAF	57	10.919540
[[[[- -	0	0.000000
[[.data.frame	[[..	- LEAF	1	0.191571
%in%	%in%	- LEAF	1	0.191571
<Anonymous>	<An>	- LEAF	6	1.149425
\$	\$	- LEAF	1	0.191571
anyDuplicated	anyD	- LEAF	1	0.191571
anyDuplicated.default	anD.	- LEAF	22	4.214559
as.character	as.c	- LEAF	43	8.237548
as.list	as.l	- -	0	0.000000
as.list.data.frame	a...	- LEAF	22	4.214559
as.list.default	as..	- LEAF	1	0.191571
as.name	as.n	- LEAF	1	0.191571
coef	coef	- LEAF	1	0.191571
deparse	dprs	- LEAF	1	0.191571

doTryCatch	dTrC	-	-	0	0.000000
eval	eval	-	LEAF	1	0.191571
evalFunc	evlF	-	-	0	0.000000
file	file	-	LEAF	1	0.191571
FUN	FUN	-	LEAF	1	0.191571
lapply	lppl	-	LEAF	2	0.383142
lazyLoadDBfetch	lLDB	-	LEAF	2	0.383142
list	list	-	LEAF	5	0.957854
lm	lm	-	LEAF	42	8.045977
lm.fit	lm.f	-	LEAF	87	16.666667
match	mtch	-	LEAF	1	0.191571
mean	mean	-	-	0	0.000000
mean.default	mn.d	-	LEAF	2	0.383142
mode	mode	-	LEAF	2	0.383142
model.frame	mdl.f	-	-	0	0.000000
model.frame.default	mdl.f.	-	LEAF	12	2.298851
model.matrix	mdl.m	-	-	0	0.000000
model.matrix.default	mdl.m.	-	LEAF	51	9.770115
model.response	mdl.r	-	LEAF	13	2.490421
na.omit	n.mt	-	LEAF	20	3.831418
na.omit.data.frame	n...	-	LEAF	26	4.980843
names	nams	-	LEAF	2	0.383142
NCOL	NCOL	-	LEAF	1	0.191571
paste	past	-	-	0	0.000000
pmatch	pmtc	-	LEAF	2	0.383142
rep.int	rp.n	-	LEAF	7	1.340996
sapply	sppl	-	LEAF	1	0.191571
simplify2array	smp2	-	-	0	0.000000
structure	strc	-	LEAF	32	6.130268
summary	smmr	-	-	0	0.000000
summary.lm	summ.	-	LEAF	40	7.662835
Sweave	Swev	ROOT	-	0	0.000000
terms	trms	-	-	0	0.000000
terms.formula	trm.	-	LEAF	1	0.191571
try	try	-	-	0	0.000000
tryCatch	tryC	-	-	0	0.000000
tryCatchList	trCL	-	-	0	0.000000
tryCatchOne	trCO	-	-	0	0.000000
unique	uniq	-	LEAF	3	0.574713
unlist	unls	-	-	0	0.000000
vapply	vppl	-	LEAF	3	0.574713
withVisible	wthV	-	-	0	0.000000
	total.time		total.pct		
!	2		0.383142		
..getNamespace	1		0.191571		
.deparseOpts	4		0.766284		
.getXlevels	26		4.980843		
[99		18.965517		
[.data.frame	99		18.965517		
[[8		1.532567		
[[.data.frame	8		1.532567		
%in%	4		0.766284		
<Anonymous>	522		100.000000		

\$	1	0.191571
anyDuplicated	23	4.406130
anyDuplicated.default	22	4.214559
as.character	43	8.237548
as.list	23	4.406130
as.list.data.frame	22	4.214559
as.list.default	1	0.191571
as.name	1	0.191571
coef	1	0.191571
deparse	2	0.383142
doTryCatch	521	99.808429
eval	521	99.808429
evalFunc	521	99.808429
file	1	0.191571
FUN	7	1.340996
lapply	30	5.747126
lazyLoadDBfetch	3	0.574713
list	5	0.957854
lm	474	90.804598
lm.fit	113	21.647510
match	11	2.107280
mean	2	0.383142
mean.default	2	0.383142
mode	2	0.383142
model.frame	168	32.183908
model.frame.default	168	32.183908
model.matrix	69	13.218391
model.matrix.default	69	13.218391
model.response	56	10.727969
na.omit	134	25.670498
na.omit.data.frame	114	21.839080
names	2	0.383142
NCOL	1	0.191571
paste	1	0.191571
pmatch	2	0.383142
rep.int	7	1.340996
sapply	14	2.681992
simplify2array	4	0.766284
structure	33	6.321839
summary	520	99.616858
summary.lm	45	8.620690
Sweave	522	100.000000
terms	2	0.383142
terms.formula	1	0.191571
try	521	99.808429
tryCatch	521	99.808429
tryCatchList	521	99.808429
tryCatchOne	521	99.808429
unique	4	0.766284
unlist	1	0.191571
vapply	23	4.406130
withVisible	521	99.808429

print_stacks(sprof) Input

Output

```
$nrstacks
[1] 50
```

```
$stacklength
[1] 3 25
```

```
$nrnodesperlevel
[1] 1 1 2 1 1 1 1 1 1 1 1 1 3 10 11 9 9 15 8 7 5 7
[23] 2 1 1
```

print_profiles(sprof) Input

Output

```
$id
[1] "Profile Summary Sat Jul 27 18:07:23 2013"
```

```
$len
[1] 522
```

```
$uniquestacks
[1] 50
```

```
$nr_runs
[1] 396
```

The `print()` method for *sprof* objects concatenates these three functions.

4.2. Summary.

summary_nodes(sprof) Input

	shortname	root	leaf	self.time	self.pct
!	!	-	LEAF	2	0.383142
..getNamespace	..gN	-	-	0	0.000000
.deparseOpts	.dpO	-	LEAF	2	0.383142
.getXlevels	.gtX	-	-	0	0.000000
[[-	-	0	0.000000
[/data.frame	[/d.	-	LEAF	57	10.919540
[[[[-	-	0	0.000000
[[/data.frame	[[/.	-	LEAF	1	0.191571
%in%	%in%	-	LEAF	1	0.191571
<Anonymous>	<An>	-	LEAF	6	1.149425
\$	\$	-	LEAF	1	0.191571
anyDuplicated	anyD	-	LEAF	1	0.191571
anyDuplicated.default	anD.	-	LEAF	22	4.214559
as.character	as.c	-	LEAF	43	8.237548
as.list	as.l	-	-	0	0.000000

as.list.data.frame	a...	- LEAF	22	4.214559
as.list.default	as..	- LEAF	1	0.191571
as.name	as.n	- LEAF	1	0.191571
coef	coef	- LEAF	1	0.191571
deparse	dprs	- LEAF	1	0.191571
doTryCatch	dTrC	- -	0	0.000000
eval	eval	- LEAF	1	0.191571
evalFunc	evlF	- -	0	0.000000
file	file	- LEAF	1	0.191571
FUN	FUN	- LEAF	1	0.191571
lapply	lppl	- LEAF	2	0.383142
lazyLoadDBfetch	lLDB	- LEAF	2	0.383142
list	list	- LEAF	5	0.957854
lm	lm	- LEAF	42	8.045977
lm.fit	lm.f	- LEAF	87	16.666667
match	mtch	- LEAF	1	0.191571
mean	mean	- -	0	0.000000
mean.default	mn.d	- LEAF	2	0.383142
mode	mode	- LEAF	2	0.383142
model.frame	mdl.f	- -	0	0.000000
model.frame.default	mdl.f.	- LEAF	12	2.298851
model.matrix	mdl.m	- -	0	0.000000
model.matrix.default	mdl.m.	- LEAF	51	9.770115
model.response	mdl.r	- LEAF	13	2.490421
na.omit	n.mt	- LEAF	20	3.831418
na.omit.data.frame	n...	- LEAF	26	4.980843
names	nams	- LEAF	2	0.383142
NCOL	NCOL	- LEAF	1	0.191571
paste	past	- -	0	0.000000
pmatch	pmtc	- LEAF	2	0.383142
rep.int	rp.n	- LEAF	7	1.340996
sapply	sppl	- LEAF	1	0.191571
simplify2array	smp2	- -	0	0.000000
structure	strc	- LEAF	32	6.130268
summary	smmr	- -	0	0.000000
summary.lm	smm.	- LEAF	40	7.662835
Sweave	Swev	ROOT -	0	0.000000
terms	trms	- -	0	0.000000
terms.formula	trm.	- LEAF	1	0.191571
try	try	- -	0	0.000000
tryCatch	tryC	- -	0	0.000000
tryCatchList	trCL	- -	0	0.000000
tryCatchOne	trCO	- -	0	0.000000
unique	uniq	- LEAF	3	0.574713
unlist	unls	- -	0	0.000000
vapply	vppl	- LEAF	3	0.574713
withVisible	wthV	- -	0	0.000000
	total.time	total.pct		
!	2	0.383142		
..getNamespace	1	0.191571		
.deparseOpts	4	0.766284		
.getXlevels	26	4.980843		
[99	18.965517		

[.data.frame	99	18.965517
[[8	1.532567
[[.data.frame	8	1.532567
%in%	4	0.766284
<Anonymous>	522	100.000000
\$	1	0.191571
anyDuplicated	23	4.406130
anyDuplicated.default	22	4.214559
as.character	43	8.237548
as.list	23	4.406130
as.list.data.frame	22	4.214559
as.list.default	1	0.191571
as.name	1	0.191571
coef	1	0.191571
deparse	2	0.383142
doTryCatch	521	99.808429
eval	521	99.808429
evalFunc	521	99.808429
file	1	0.191571
FUN	7	1.340996
lapply	30	5.747126
lazyLoadDBfetch	3	0.574713
list	5	0.957854
lm	474	90.804598
lm.fit	113	21.647510
match	11	2.107280
mean	2	0.383142
mean.default	2	0.383142
mode	2	0.383142
model.frame	168	32.183908
model.frame.default	168	32.183908
model.matrix	69	13.218391
model.matrix.default	69	13.218391
model.response	56	10.727969
na.omit	134	25.670498
na.omit.data.frame	114	21.839080
names	2	0.383142
NCOL	1	0.191571
paste	1	0.191571
pmatch	2	0.383142
rep.int	7	1.340996
sapply	14	2.681992
simplify2array	4	0.766284
structure	33	6.321839
summary	520	99.616858
summary.lm	45	8.620690
Sweave	522	100.000000
terms	2	0.383142
terms.formula	1	0.191571
try	521	99.808429
tryCatch	521	99.808429
tryCatchList	521	99.808429
tryCatchOne	521	99.808429

unique	4	0.766284
unlist	1	0.191571
vapply	23	4.406130
withVisible	521	99.808429

summary_stacks(sprof) *Input*

Output

```
$nrstacks
[1] 50
```

```
$stacklength
[1] 3 25
```

```
$nrnodesperlevel
[1] 1 1 2 1 1 1 1 1 1 1 1 1 3 10 11 9 9 15 8 7 5 7
[23] 2 1 1
```

summary_profiles(sprof) *Input*

Output

```
$id
[1] "Profile Summary Sat Jul 27 18:07:23 2013"
```

```
$len
[1] 522
```

```
$uniquestacks
[1] 50
```

```
$nr_runs
[1] 396
```

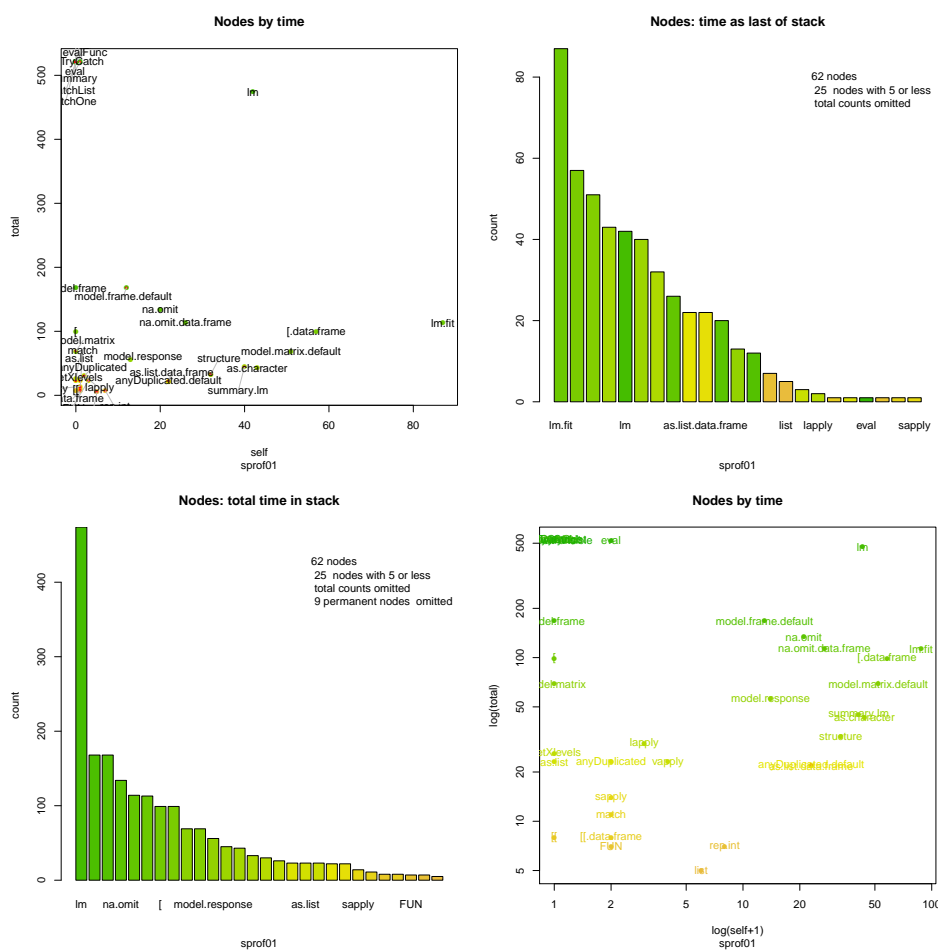
The `summary()` method for `sprof` objects concatenates these three functions.

4.3. **Plot.** Looking at lists of numbers is not too informative. We get a first impression by plotting the data. Examples of the plot output is not given here. Please run the examples, or see the *sprof* package reference.

```

#plot_nodes(sprof01, col=nodescol[nodescore])
oldpar <- par(mfrow=c(2,2))
plot_nodes(sprof01)
par(oldpar)

```

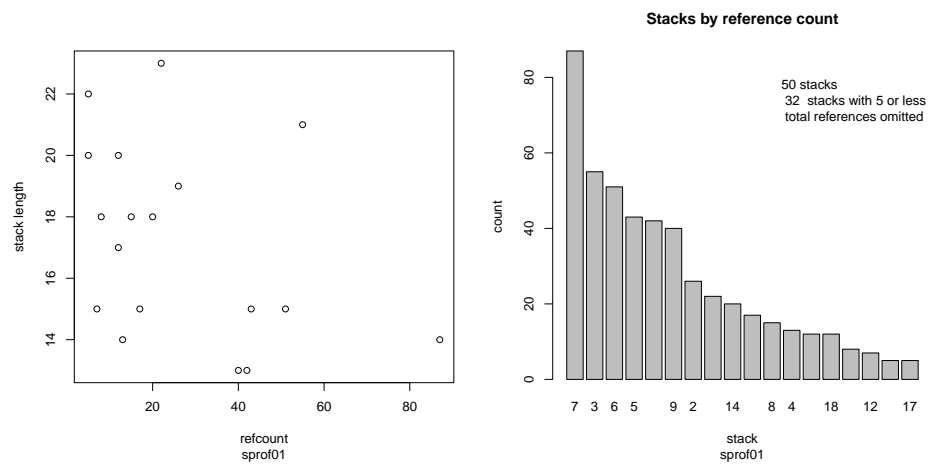


```

oldpar <- par(mfrow=c(1,2))
plot_stacks(sprof01)
par(oldpar)

```

Input

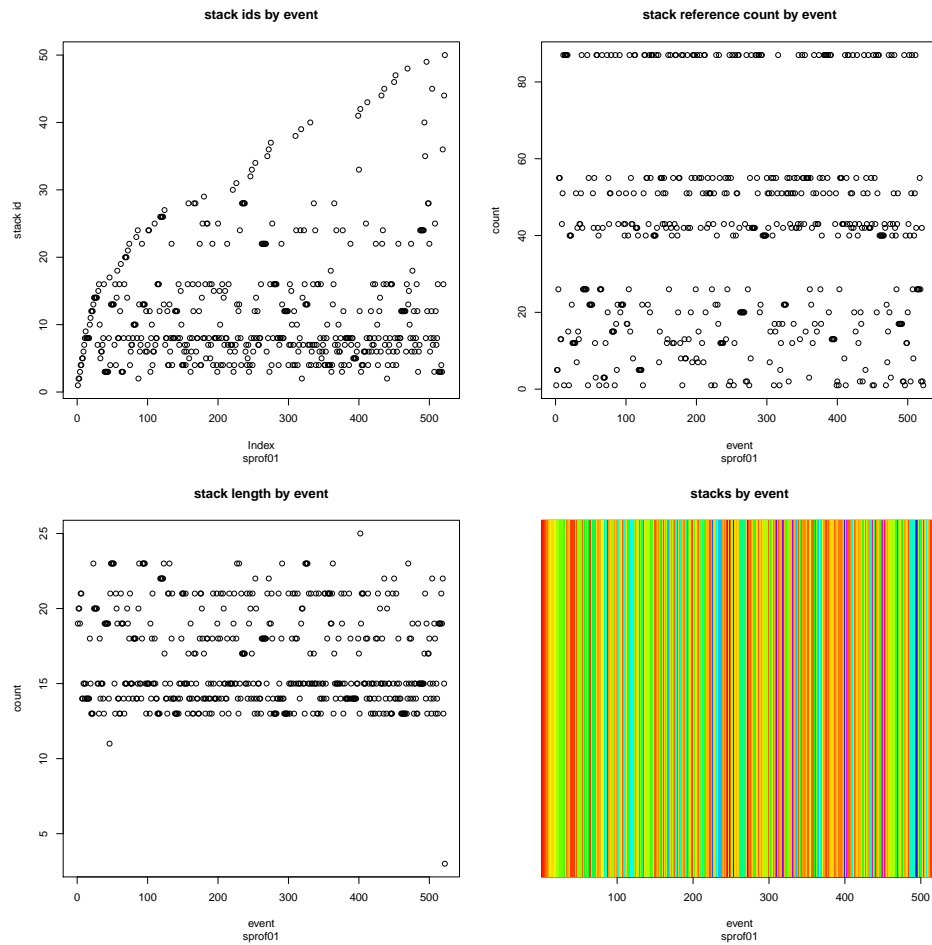


Input

```

oldpar <-par(mfrow=c(2,2))
plot_profiles(sprof01)
par(oldpar)

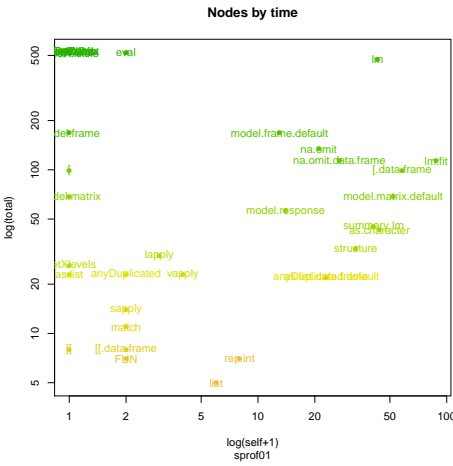
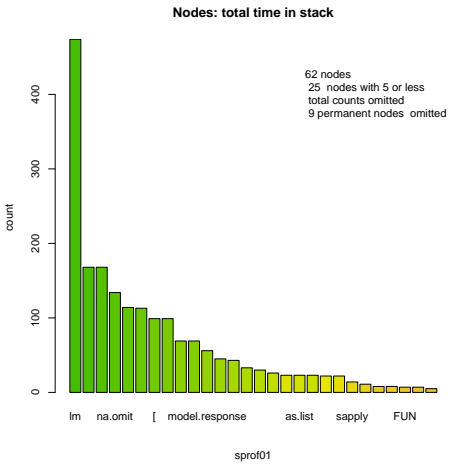
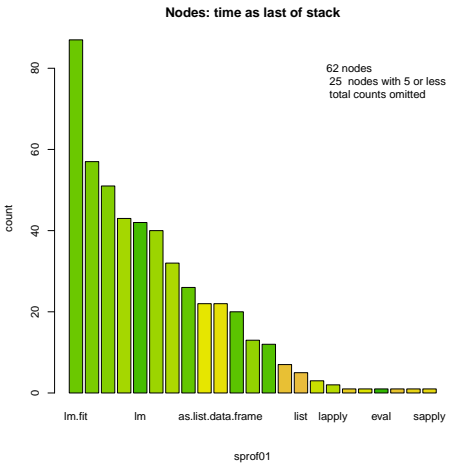
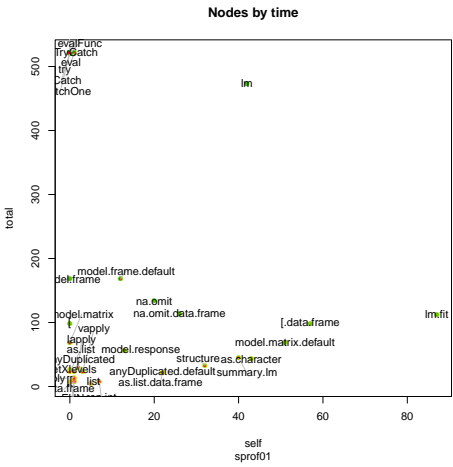
```



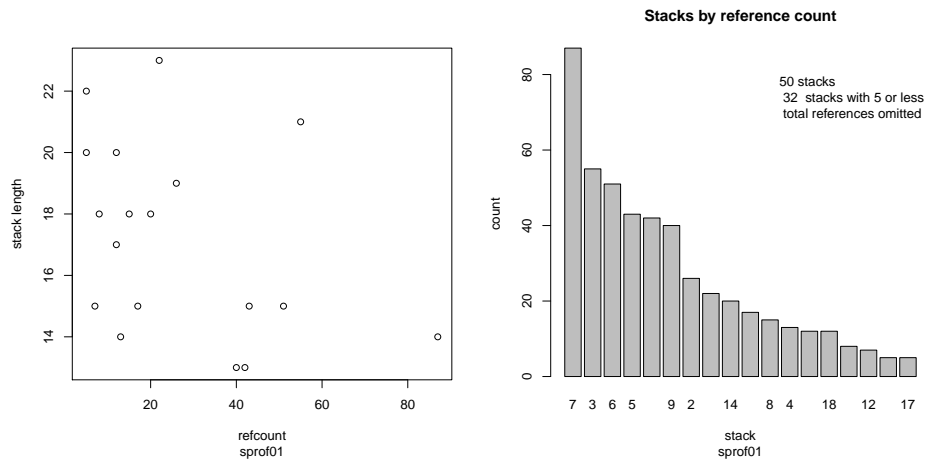
The `plot()` method for `sprof` objects concatenates these three functions.

Input

```
oldpar<- par(mfrow=c(2,2))
plot_nodes(sprof)
par(oldpar)
```

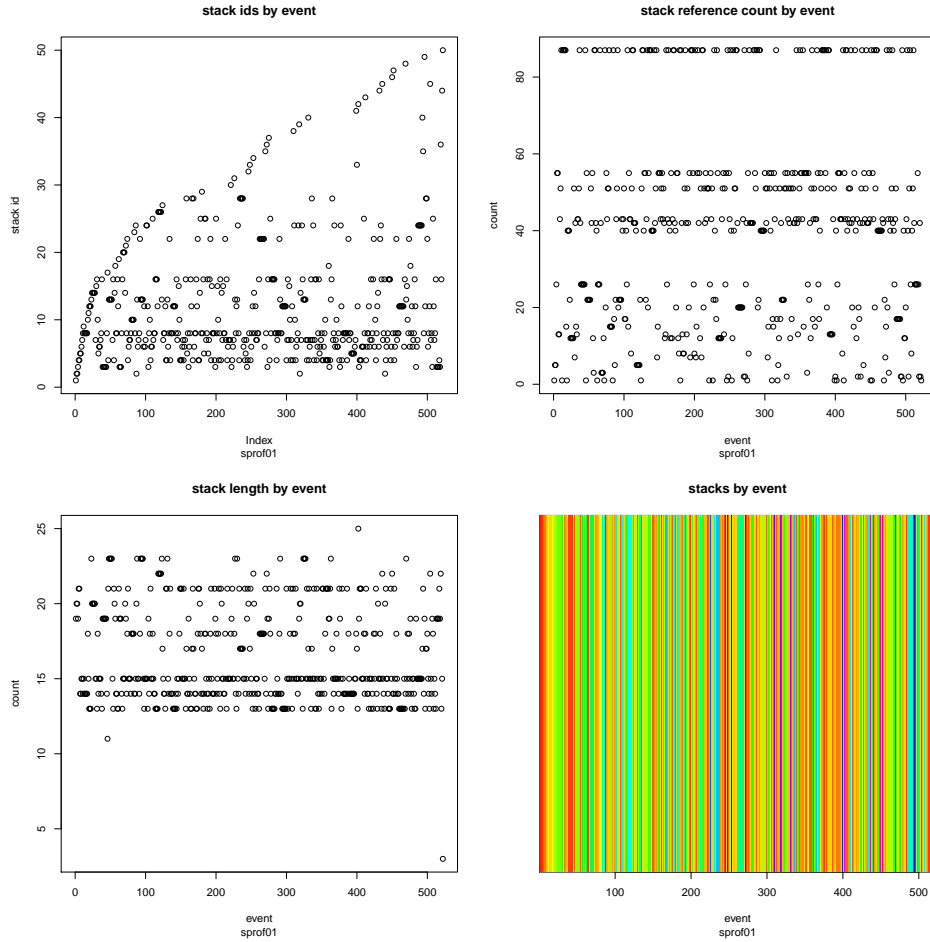


```
oldpar<- par(mfrow=c(1,2)) Input
plot_stacks(sprof)
par(oldpar)
```



 Input

```
oldpar<- par(mfrow=c(2,2))
plot_profiles(sprof)
par(oldpar)
```



The `plot()` method for `sprof` objects concatenates these three functions.

5. GRAPH

Graph layout is a theme of its own. Proposals are readily available, as are their implementation. For some of them, there are R interfaces or re-implementations in R. Their usefulness in our context has to be explored, and the answers will vary with personal preferences.

For some graph layout packages we illustrate an interface here and show a sample result. We use the original profile data here. This is a nasty graph with some R stack peculiarities. The corresponding results for the trimmed profile data are shown in the next section section 5.2 on page 75. This is a more realistic example of the kind of graphs you will have to work with.

5.1. Example: regression. In this section, we use the recent version of our example, *sprof02* for demonstration. You can re-run it, using your *sprof* data by modifying this instruction:

sprof <- *sprof02* *Input*

To interface *sprof* to a graph handling package, *adjacency()* can extract the adjacency matrix from the profile.

There are various packages for finding a graph layout, and the choice is open to your preferences. The R packages for most of these are just wrapper

sprofadj <- *adjacency(sprof)* *Input*

This is a format any graph package can handle (maybe).

ToDo: by graph package: preferred input format?

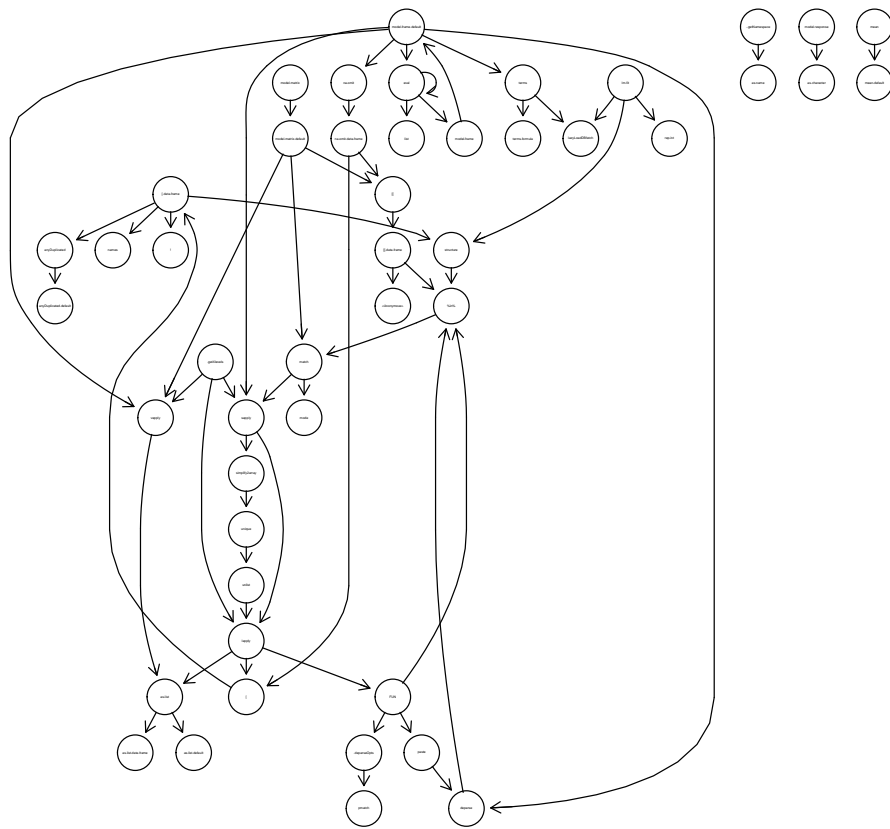
ToDo: use attributes. Edge with should be easy.

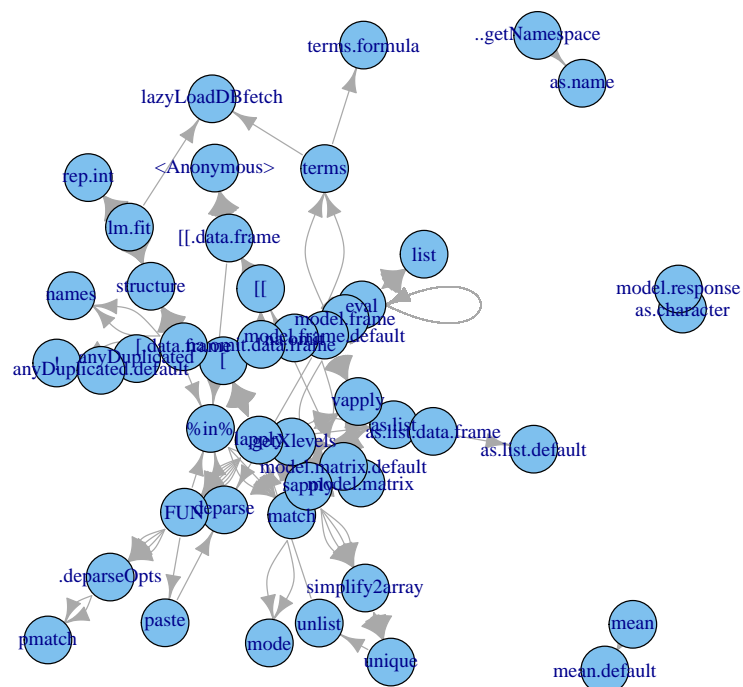
ToDo: include information from stack connectivity.

5.1.1. *graph* package.

```
library(graph)
sprofadjNEL <- as(sprofadj, "graphNEL")
```

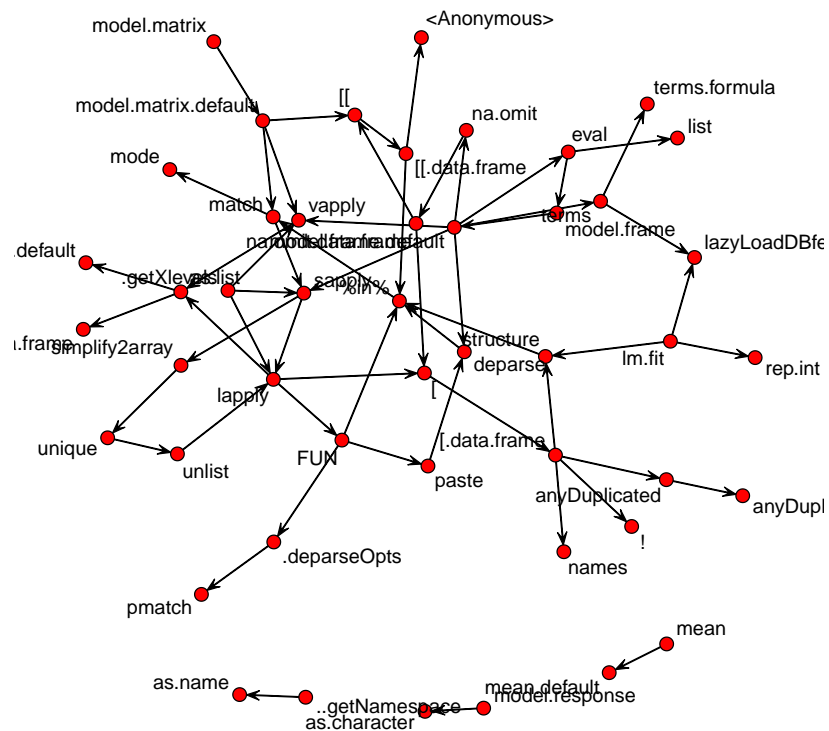
```
plot(sprofadjNEL, main="sprof01: graph layout", cex.main=2)
#detach("package:graph")
```

sprof01: graph layout



5.1.3. *network* package.

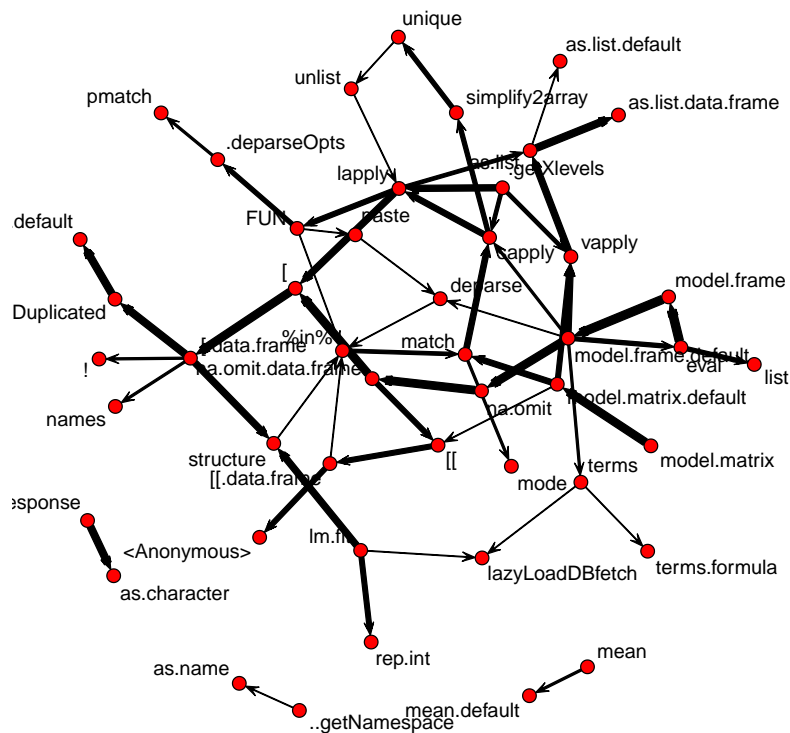
```
library(network)
nwsprofadj <- as.network(sprofadj) # names is not imported
network.vertex.names(nwsprofadj) <- rownames(sprofadj) # not honoured by plot
plot(nwsprofadj, label=rownames(sprofadj), main="sprof01: network layout", cex.main=2)
```

sprof01: network layout

Experiments to include weight.

Input

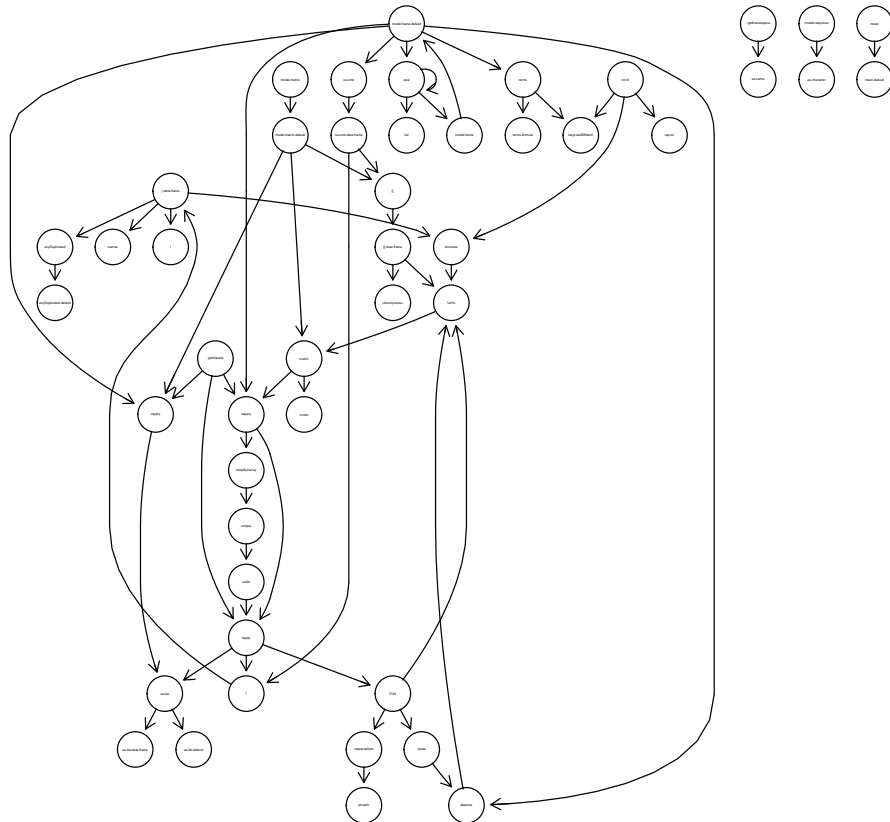
sprof01: network layout



5.1.4. *Rgraphviz* package.

```
library(Rgraphviz)
sprofadjRag <- agopen(sprofadjNEL, name="Rprof Example")
```

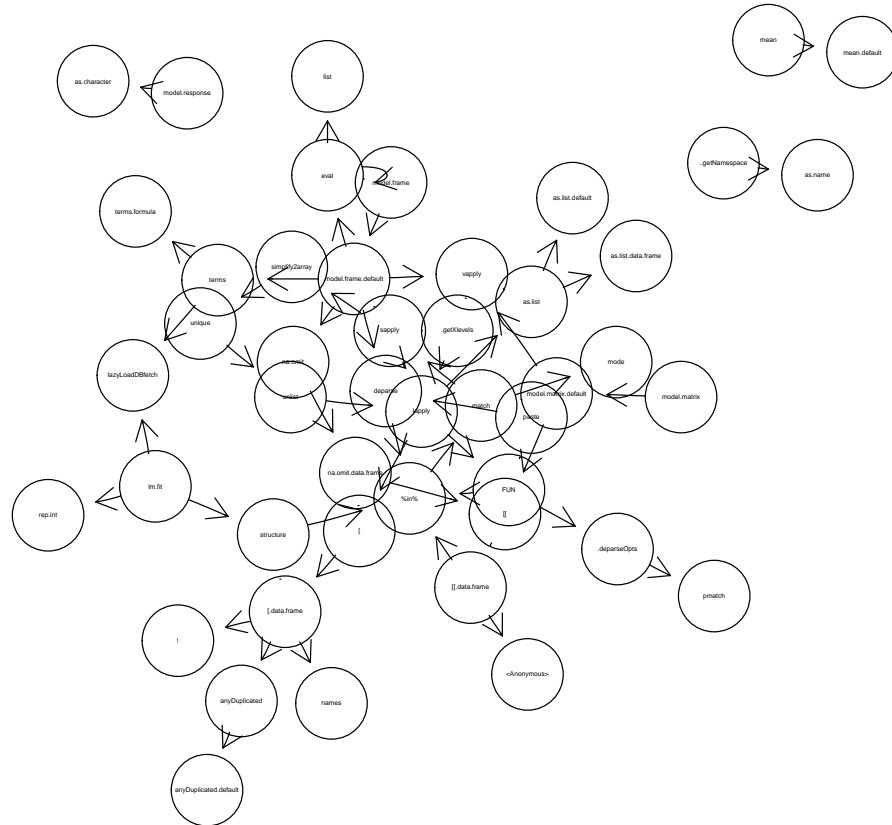
```
plot(sprofadjRag, main="sprof01: Graphviz dot layout", cex.main=2)
```

sprof01: Graphviz dot layout

#8 *Input*

```
plot(sprofadjRag,"neato", main="sprof01: Graphviz neato layout", cex.main=2)
```

sprof01: Graphviz neato layout

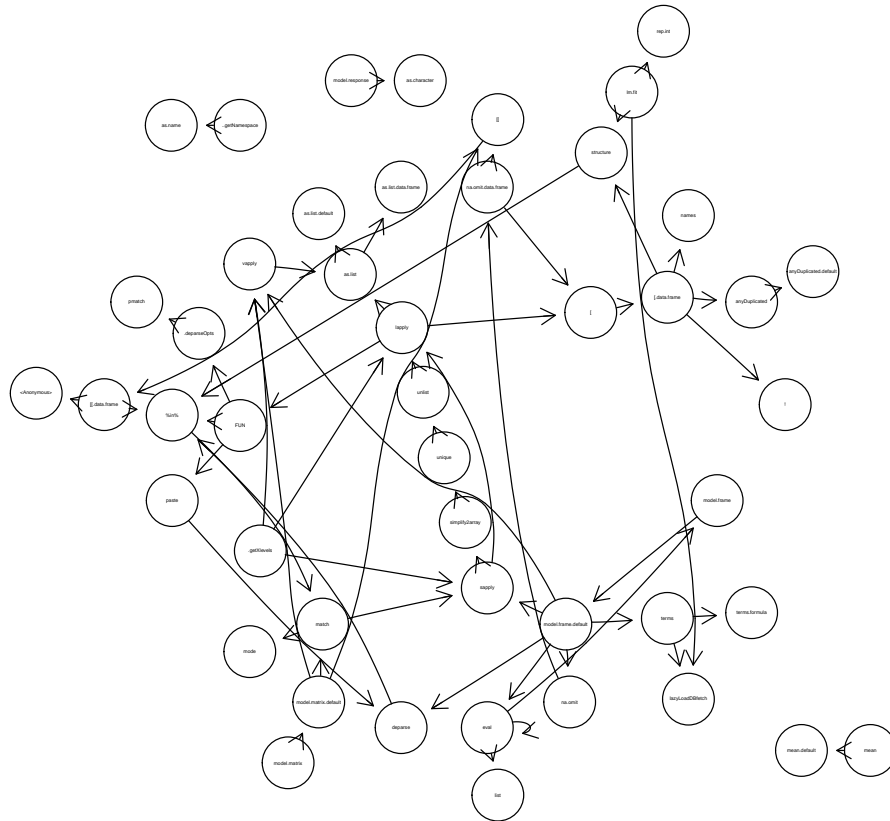


 Input

#6

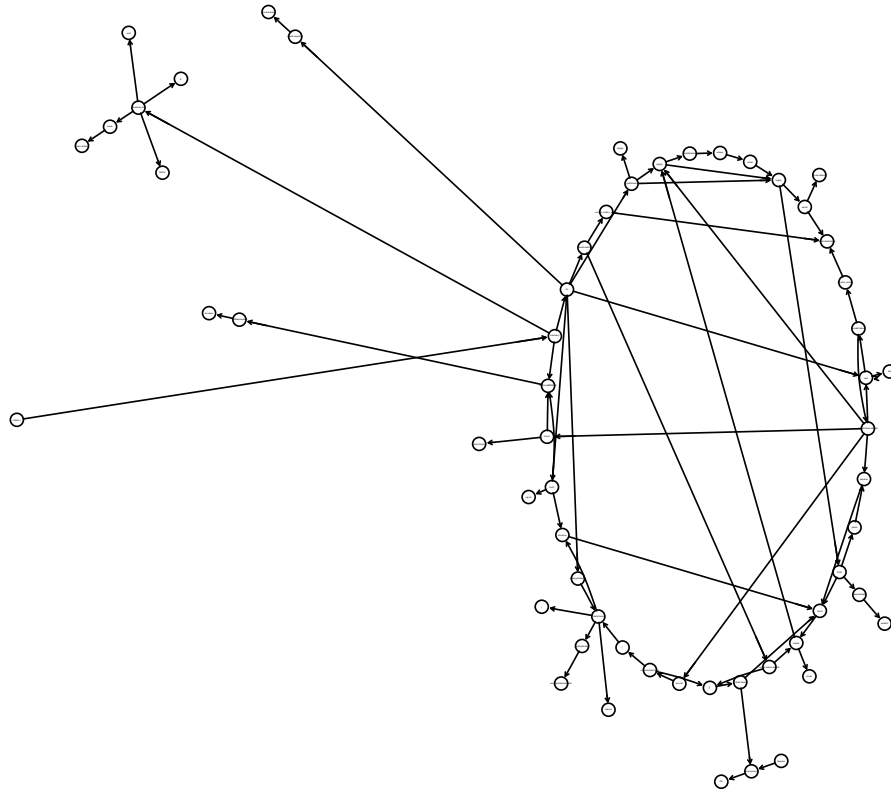
```
plot(sprofadjRag,"twopi", main="sprof01: Graphviz twopi layout", cex.main=2)
```

sprof01: Graphviz twopi layout



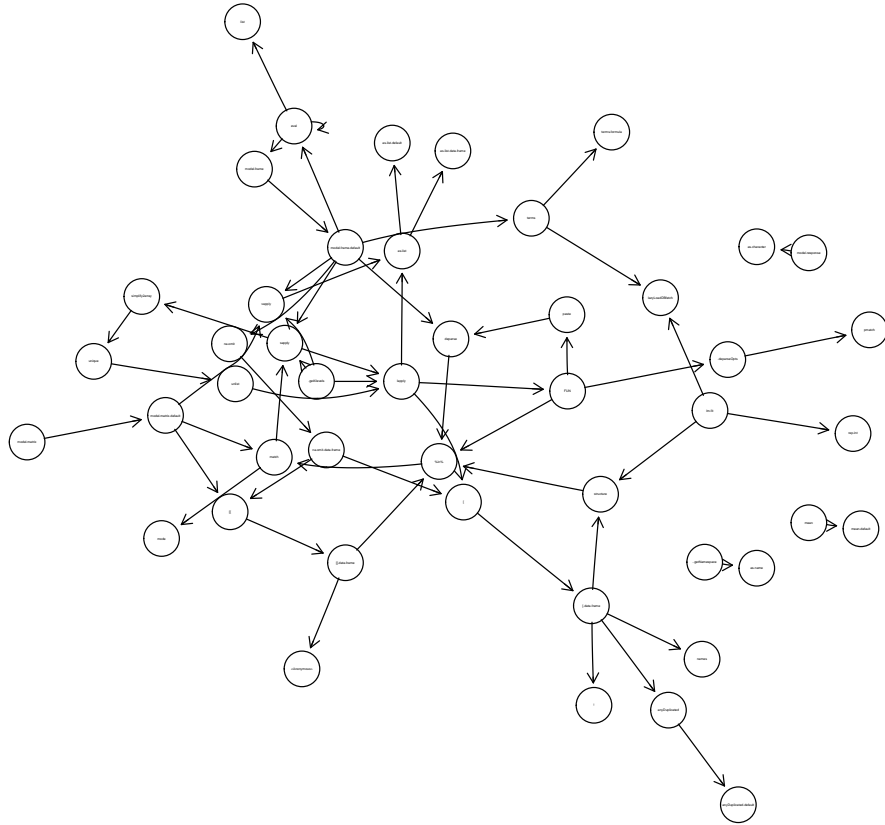
Input
`plot(sprofadjRag,"circo", main="sprof01: Graphviz circo layout", cex.main=2)`

sprof03: Graphviz circo layout



Input
`plot(sprofadjRag,"fdp", main="sprof01: Graphviz fdp layout", cex.main=2)`

sprof01: Graphviz fdp layout



5.2. Trimmed example: regression. In this section, we use the reduced version of our example, *sprof03* for demonstration. Except for the change of the data set, this is just a copy of the previous chapter, collecting the various layouts for easy reference.

Some experiments may have found their way to this chapter. They will be expelled.

You can re-run it, using your *sprof* data by modifying this instruction:

```
sprof <- sprof03
```

To interface *sprof* to a graph handling package, *until()* can extract the adjacency matrix from the profile.

```
sprofadj <- adjacency(sprof)  
adjname <- colnames(sprofadj)  
adjname[adjname==""] <- "<NULL>"  
colnames(sprofadj) <- adjname  
rownames(sprofadj) <- adjname
```

This is a format any graph package can handle (maybe).

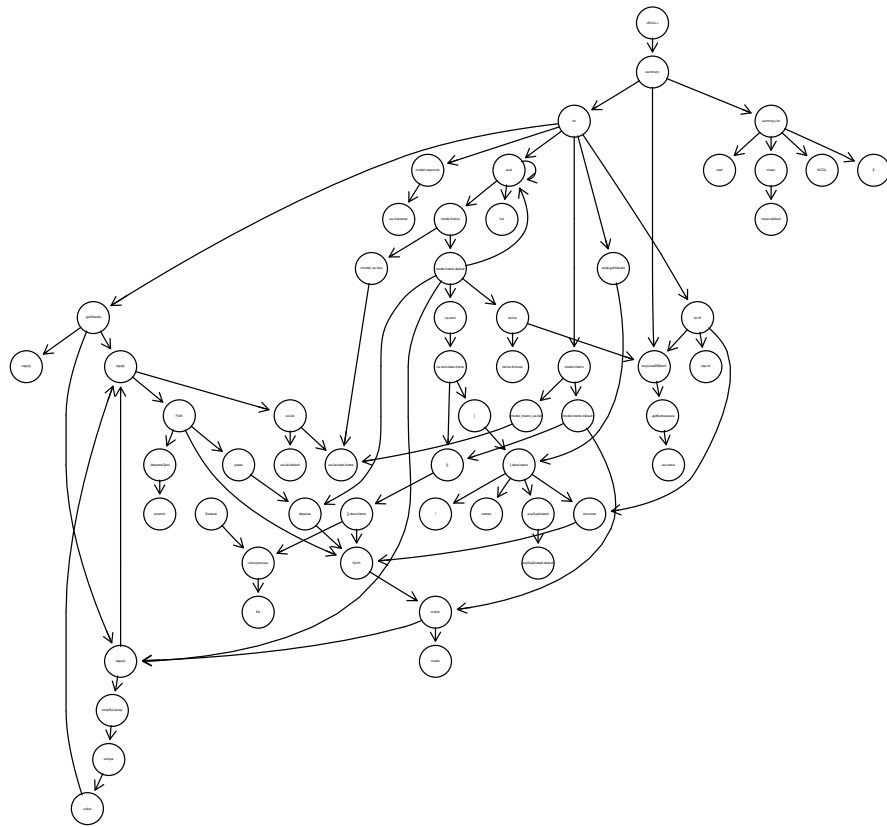
5.2.1. *graph* package.

Input

```
library(graph)
sprofadjNEL <- as(sprofadj, "graphNEL")
```

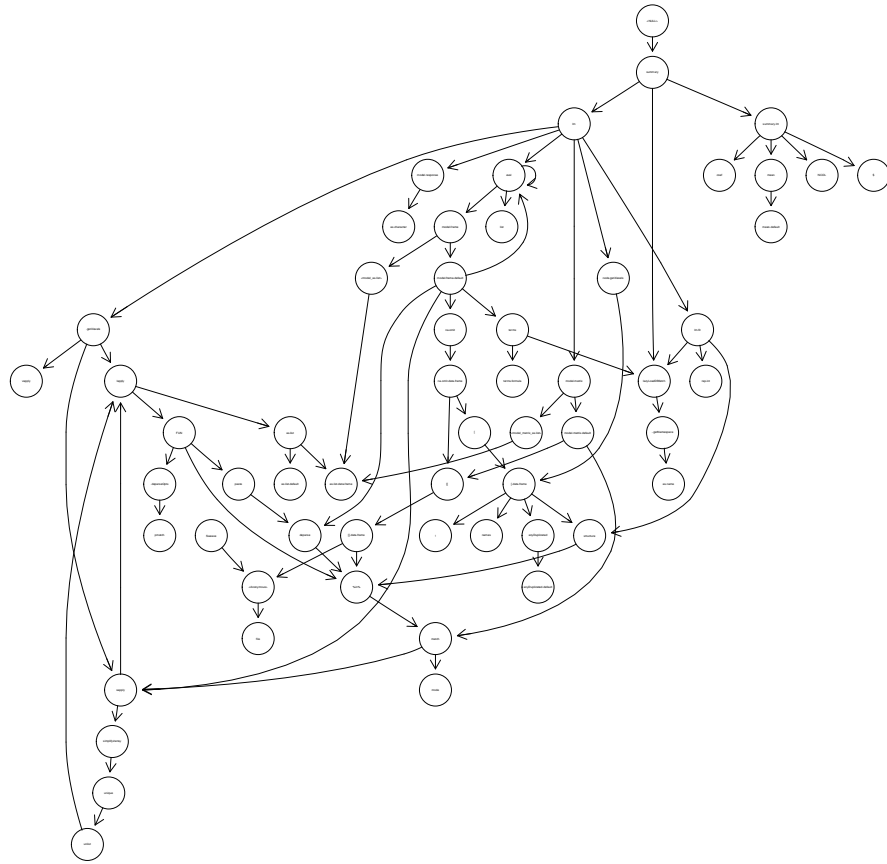
Input

```
#24
plot(sprofadjNEL, main="sprof03: graph layout", cex.main=2)
#detach("package:graph")
```

sprof03: graph layout

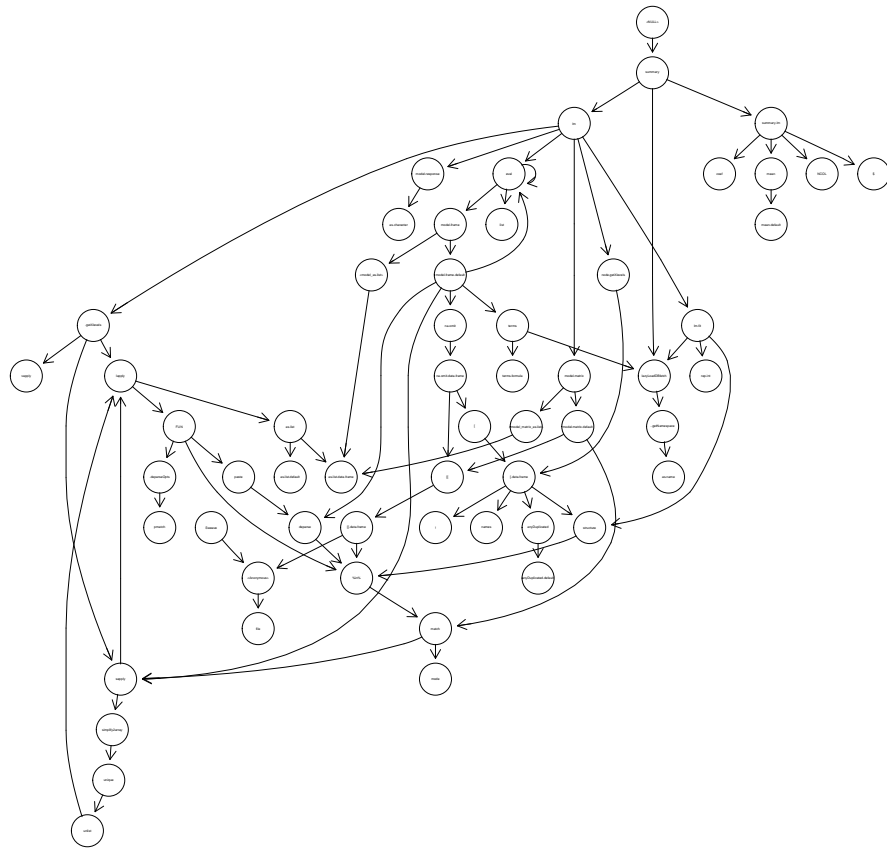
Input

```
#18
plot(sprofadjNEL, main="sprof03: graph layout", cex.main=2)
#detach("package:graph")
```

sprof03: graph layout

 Input

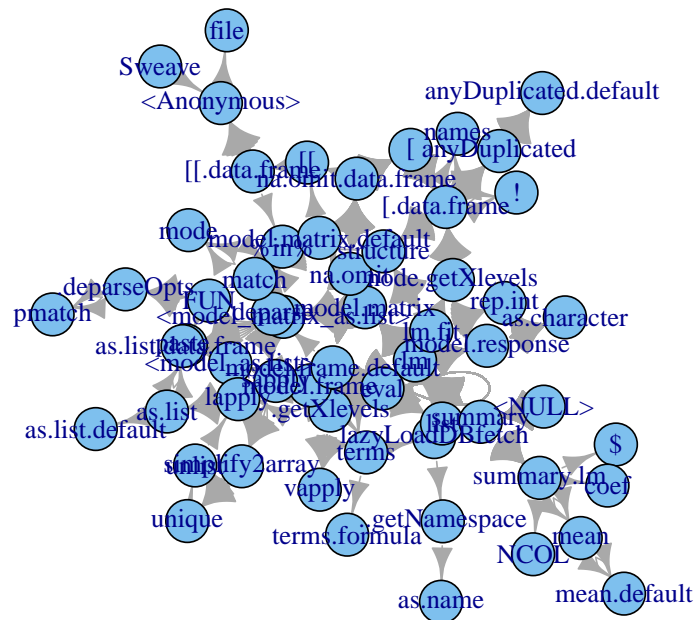
```
#12
plot(sprofadjNEL, main="sprof03: graph layout", cex.main=2)
#detach("package:graph")
```

sprof03: graph layout

5.2.2. *igraph* package.

```
library(igraph)
sprof03 <- graph.adjacency(sprofadj)
```

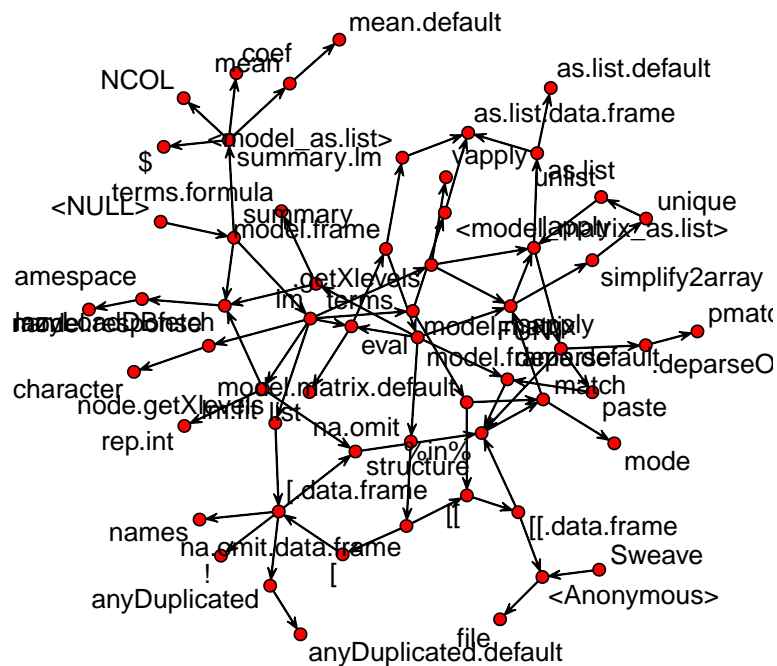
```
#6
#plot(sprof03, main="sprof03: igraph layout", cex.main=2)
plot(sprof03, main="sprof03: igraph layout: trimmed data")
detach("package:igraph")
```

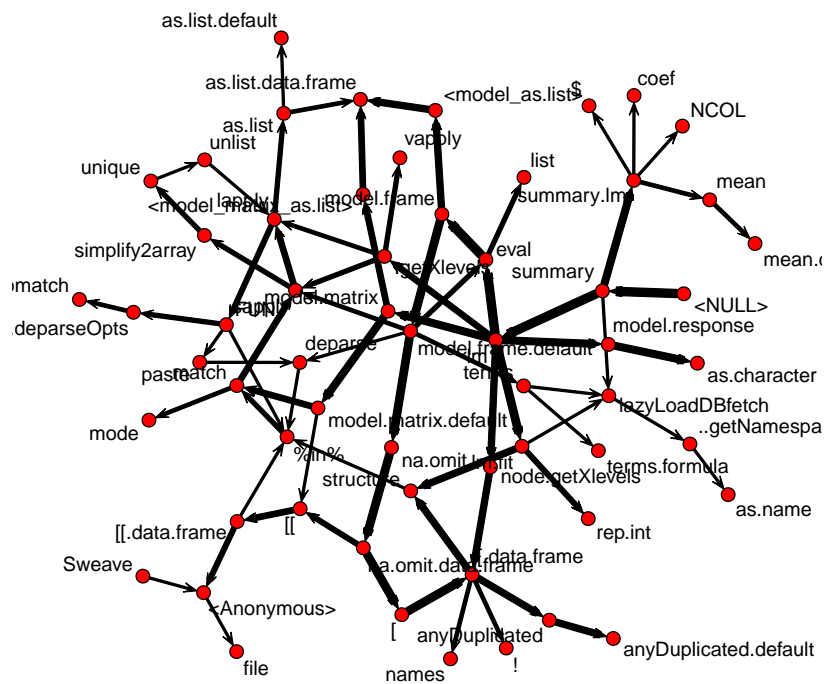
sprof03: igraph layout: trimmed data

5.2.3. *network* package.

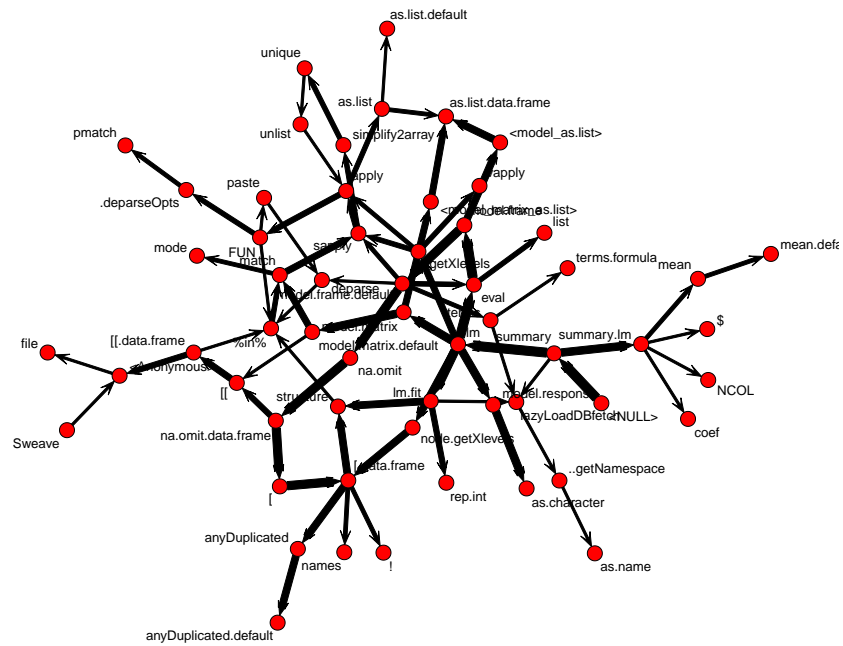
```
#6
library(network)
nwsprowfadj <- as.network(sprofadj) # names is not imported
network.vertex.names(nwsprowfadj) <- rownames(sprofadj) # not honoured by plot
plot(nwsprowfadj, label=rownames(sprofadj),
     main="sprof03: network layout: trimmed data", cex.main=2)
```

sprof03: network layout: trimmed dat



sprof03: network layout: trimmed data

#12

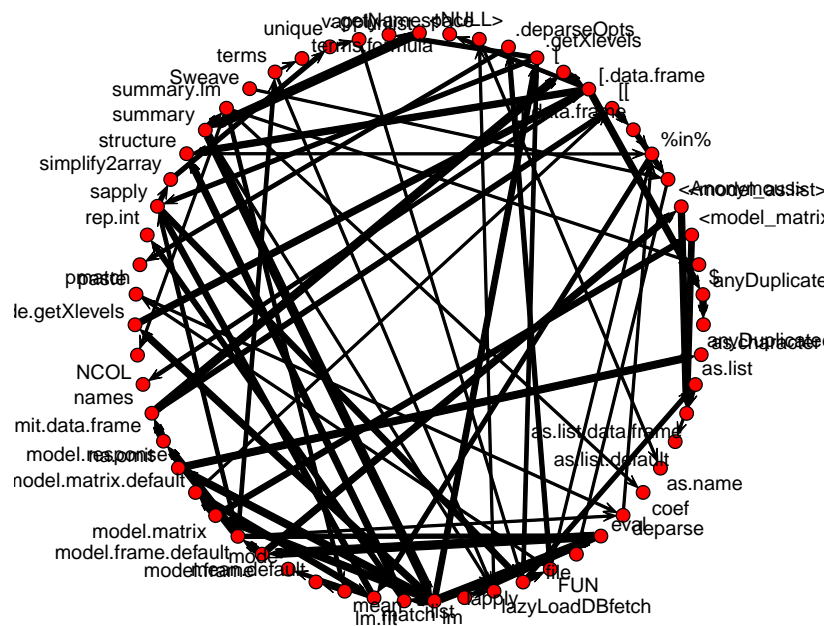
**sprof03: network kamadakawai layout:
trimmed data**

```

Input
plot(nwsprofadj, label=rownames(sprofadj),
     main="sprof03: network circle layout: \n trimmed data",
     mode="circle",
     cex.main=2, edge.lwd=edge.lwd)

```

sprof03: network circle layout: trimmed data

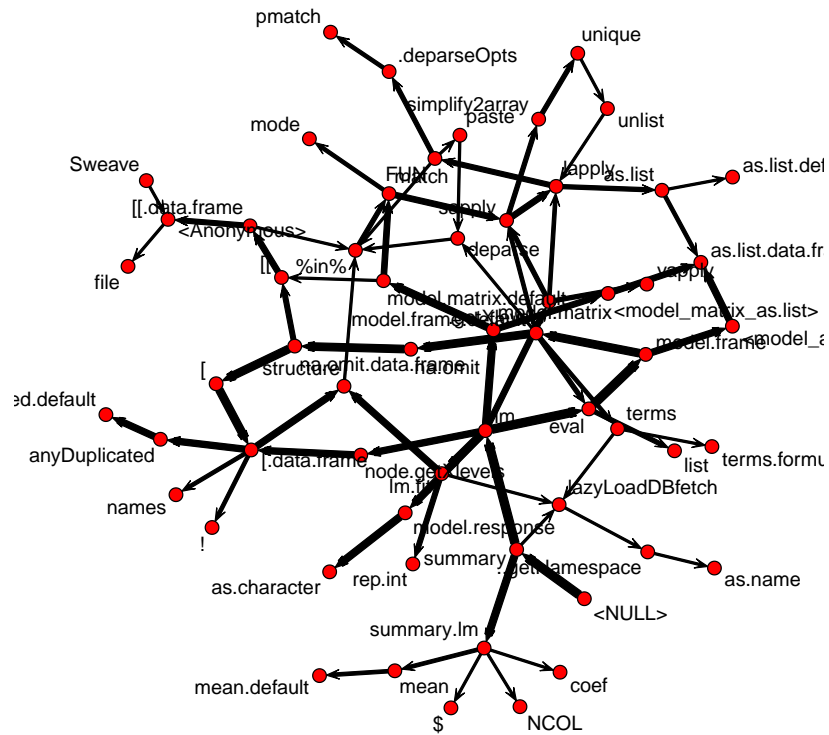


```

Input
plot(nwsprofadj, label=rownames(sprofadj),
     main="sprof03: network fruchtermanreingold layout: \n trimmed data",
     mode="fruchtermanreingold",
     cex.main=2, edge.lwd=edge.lwd)
detach("package:network")

```

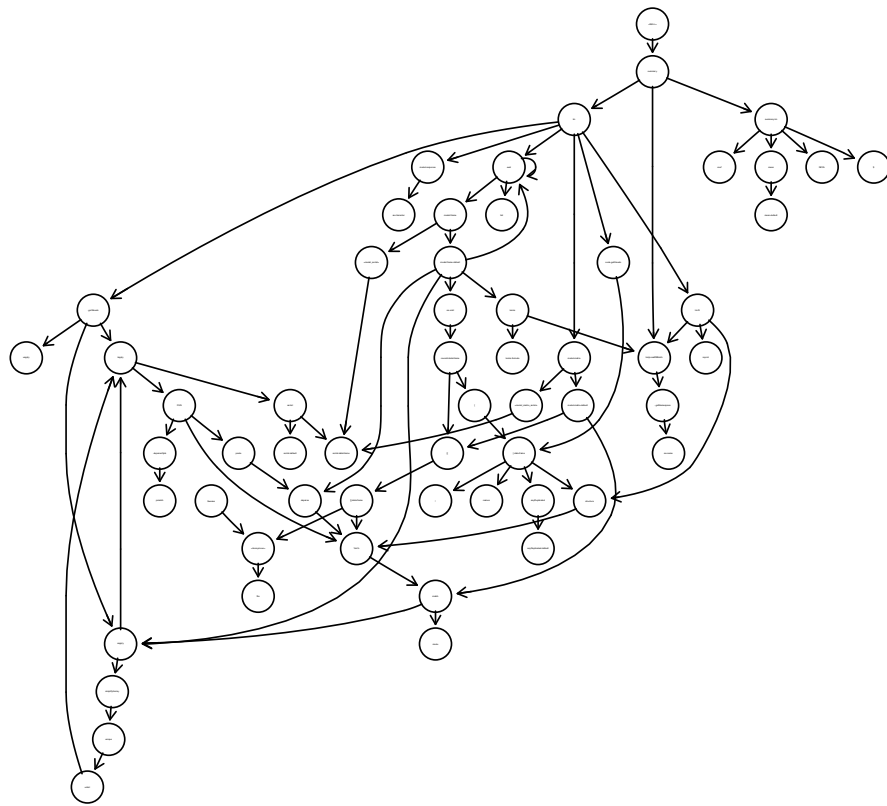
**sprof03: network fruchtermanreingold layout:
trimmed data**



5.2.4. *Rgraphviz* package.

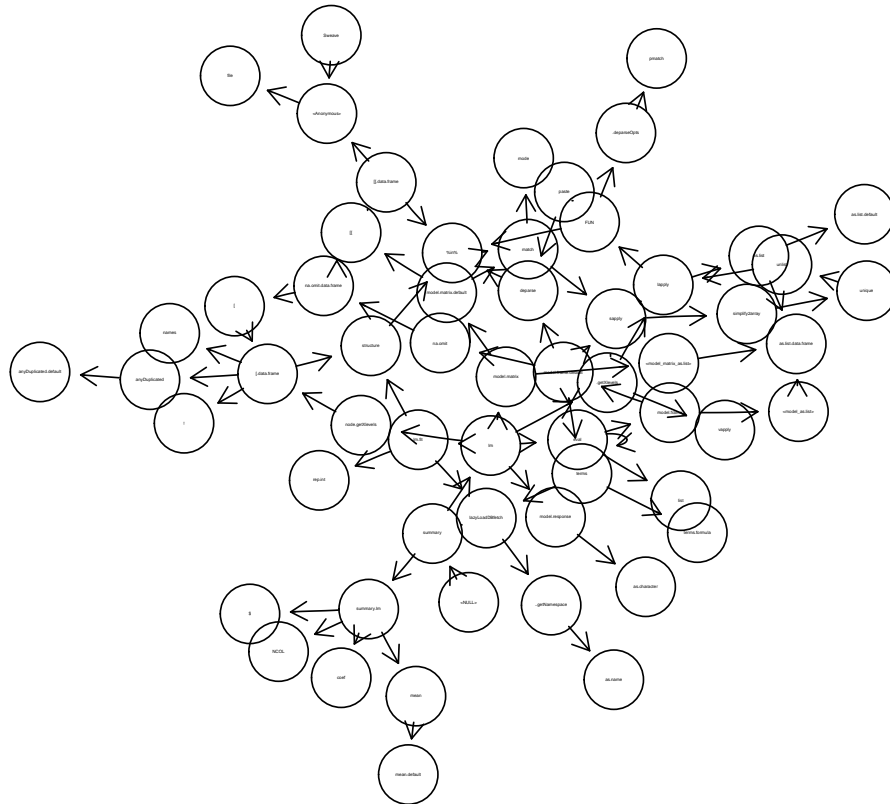
```
library(Rgraphviz)
sprofadjRag <- agopen(sprofadjNEL, name="Rprof Example")
```

```
#6
plot(sprofadjRag, main="sprof03: Graphviz dot layout", cex.main=2)
```

sprof03: Graphviz dot layout

#6

sprof03: Graphviz neato layout

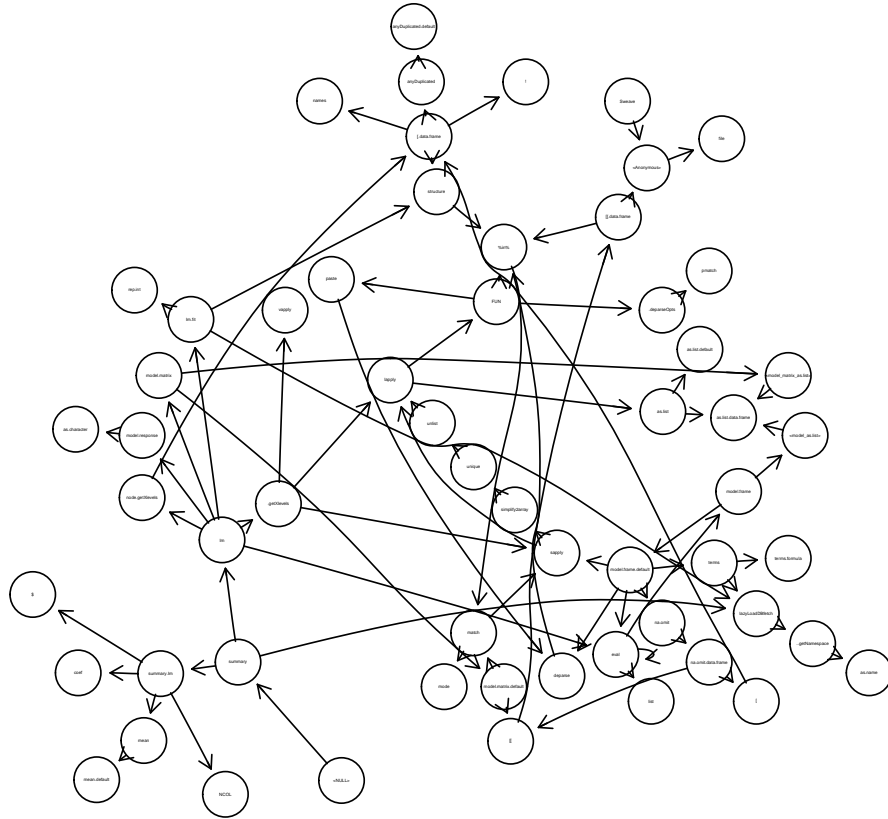


Input

#6

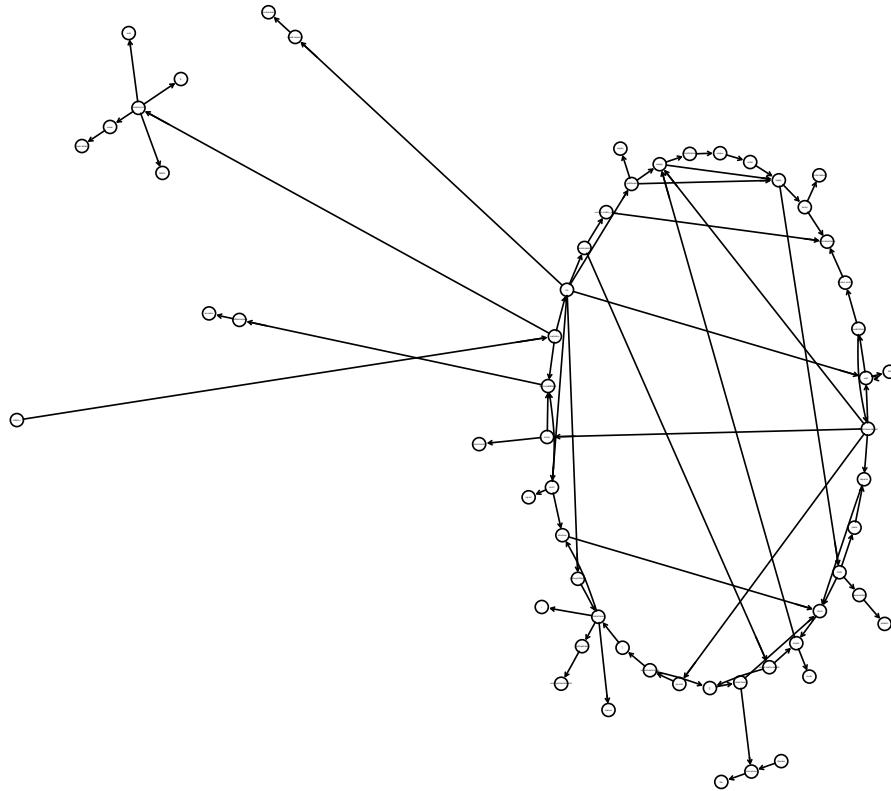
```
plot(sprofadjRag,"twopi", main="sprof03: Graphviz twopi layout")
```

sprof03: Graphviz twopi layout



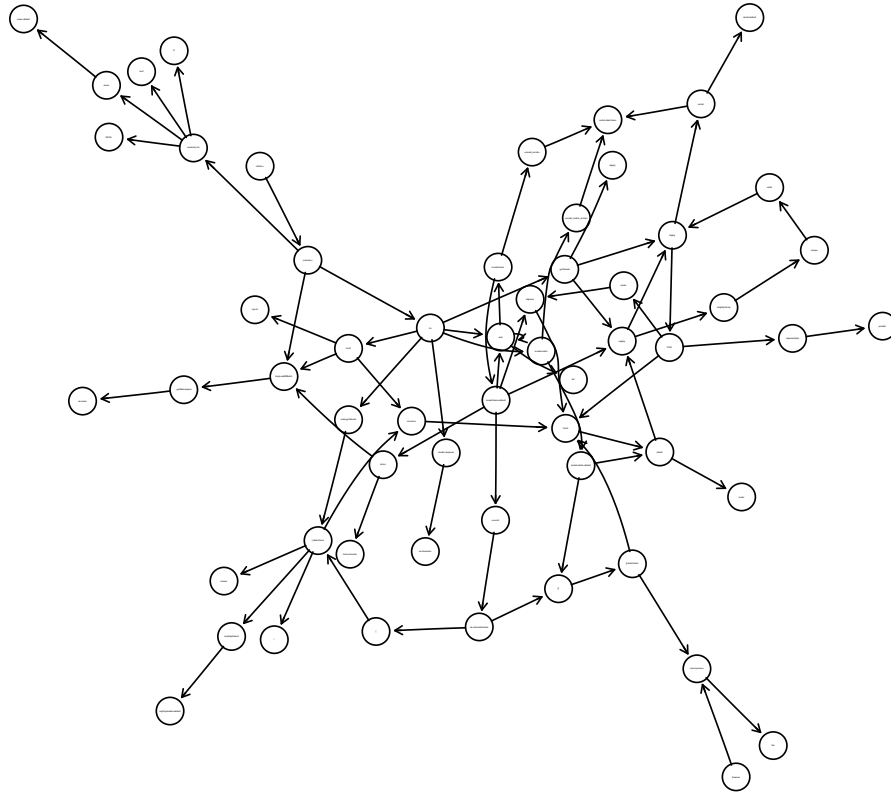
#6
plot(sprofadjRag,"circo", main="sprof03: Graphviz circo layout", cex.main=2)

sprof03: Graphviz circo layout



```
#6  
plot(sprofadjRag,"fdp", main="sprof03: Graphviz fdp layout", cex.main=2)
```

sprof03: Graphviz fdp layout



INDEX

Topic **adjacency**

until, 75

Topic **manip**

adjacency, 22

list.as.matrix, 22

profiles_matrix, 22

stacks_matrix, 22

stackstoadj, 22

Topic **misc**

apply, 5

plot, 61, 64

print, 55

sampleRprof, 9

summary, 58

summaryRprof, 5

Topic **util**

adjacency, 65

ToDo

0: add keep3 to keep header, some middle, tail, 3

0: remove text vdots from string/name columns. Use empty string., 4

1: Can we calibrate times to CPU rate? Introduce cpu clock cycle as a time base, 6

1: Defaults by class, 18

1: Move class attributes to package code, 14

1: add a reference to colorbrewer, 18

1: add class by keyword, 14

1: add class by package for all nodes, 14

1: add sampling.interval, sampling.time for backward compability, 9

1: apply colour to selection?, 12

1: colour by class – redo. Bundle colour index with colour?, 13

1: improve colour: support colour in a structure, 12

1: rearrange stacks? detect order?, 5

1: remove text vdots from string/name columns, 12

1: spread colour on displayed part, 12

2: Implement. Currently best handled on source=text level, 31

2: Warning: data structure still under discussion, 33

2: add ?? function, 24

2: add attributes to stacks, and discuss scope, 18

2: add current level, 41

2: add marginals and conditionals.

Provide function node.summary., 36

2: add names for node dimension, 35

2: add time per call information: add marginals statistics run time by node, 41

2: allow sorting, e.g. by marginals, 37

2: check and stabilize color linking, 22

2: colours. recolour. Propagate colour to graph., 23

2: complete matrix conversion, 21

2: cut next level, 31

2: example, 23

2: function addnode to be added, 31

2: generate a coplot representation, 41

2: handle empty stacks and zero counts gracefully, 24

2: implement, 32

2: implement replacement on the stack level., 32

2: keep as factor. This is a saprse cube with margins node, stack level, run length. Nodes are mostly concentrated on few levels., 33

2: needs serious revision, 32

2: replace by decent vector/array based implementation, 35

2: replace sum by weighted sum, 37

2: rescale to application scale, 37

2: some mess in the code below – not working, 35

2: sorting/arranging stacks, 18

2: table: node #runs min median run length max, 41

2: trimexample, 27

3: cut top levels, 51

3: fix null name, 51

3: use stack colours, 43

4: Clarify: print prints its argument and returns it invisibly (via invisible(x)). Return the argument, or some print represntation?, 52

4: is there a print=FALSE variant to postpone printing to e.g. xtable?, 52

5: by graph package: preferred input format?, 65

5: include information from stack connectivity., 65

5: maximum edge.lwd?, 69

5: use attributes. Edge with should be easy., 65

adjacency, 22, 65

apply, 5

Index01, 89

list.as.matrix, 22

plot, 61, 64

`print`, 55
`profiles_matrix`, 22

`sampleRprof`, 9
`stacks_matrix`, 22
`stackstoadj`, 22
`summary`, 58
`summaryRprof`, 5

`until`, 75

R session info:

- R version 3.0.1 (2013-05-16), x86_64-apple-darwin10.8.0
- Locale: en_GB.UTF-8/en_GB.UTF-8/en_GB.UTF-8/C/en_GB.UTF-8/en_GB.UTF-8
- Base packages: base, datasets, graphics, grDevices, grid, methods, stats, utils
- Other packages: graph 1.38.2, RColorBrewer 1.0-5, Rcpp 0.10.3, Rgraphviz 2.4.0, sna 2.3-1, sprof 0.0-5, wordcloud 2.4, xtable 1.7-1
- Loaded via a namespace (and not attached): BiocGenerics 0.6.0, igraph 0.6.5-2, network 1.7.2, parallel 3.0.1, slam 0.1-28, stats4 3.0.1, tools 3.0.1

L^AT_EX information: textwidth: 4.9823in linewidth:4.9823in

textheight: 8.0824in

Svn repository information:

```
$HeadURL: svnssh://gsawitzki@svn.r-forge.r-project.org/svnroot/sintro/pkg/sprof/vignettes/sprofiling.Rnw +
$Source: /u/math/j40/cvsroot/lectures/src/insider/profile/Rnw/profile.Rnw,v $
$Id: sprofiling.Rnw 182 2013-07-27 15:52:16Z gsawitzki $
$Revision: 182 $
$Date: 2013-07-27 17:52:16 0200(Sat, 27 Jul 2013)+
$name: $
$Author: gsawitzki $
```

GÜNTHER SAWITZKI
STATLAB HEIDELBERG
IM NEUENHEIMER FELD 294
D 69120 HEIDELBERG

E-mail address: gs@statlab.uni-heidelberg.de

URL: <http://sintro.r-forge.r-project.org/>