R PROFILING AND OPTIMISATION

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PENDING CHANGES

Warning: this is under construction.

This vignette contains experimental material which may sink down to the package implementation, or vanish.

Known issues:

- Control information may be included as special stack in raw format.
- A list of profiles may become default. Only one profiling interval value per profile.
- Nodes may be implemented as factor. Work-around for the R factor handling needs to be added, i.e. factor as a data structure.
- changing timing interval is too expensive, as rle is not transparent to data frames. Implement profiles as a list, with a time interval attribute per list element.

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An R vignette for package sprof.

URL: http://sintro.r-forge.r-project.org/

Private Version

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Profiling facilities in R

For the impatient: table 1 on page 9 and table 2 on page 10 give you information from profiling 100 runs of a simple linear regression. You get another view of the same information by fig. 15 on page 60. If you want to know more, please have some patience.

R provides the basic instruments for profiling, both for time based samplers as for event based instrumentation. Information on R profiling is in section 3.2 "Profiling R code for speed" and section 3.3. "Profiling R code for memory use" of "Writing R Extensions" http://cran.r-project.org/doc/manuals/R-exts.html. Specific information on memory profiling is in

http://developer.r-project.org/memory-profiling.html.

However this source of information seems to be rarely used.

Maybe the supporting tools are not adequate. The summaries provided by R reduce the information beyond necessity. Additional packages are available, but these are not sufficiently action oriented.

With package **sprof** we want to give a data representation that keeps the full profile information. Tools to answer common questions are provided. The data structure should make it easy to extend the tools as required.

The package is currently distributed at r-forge as part of the sintro material.

To install this package directly within R, type install.packages("sprof",repos="http://r-forge.r-project.org")

To install the recent package from source directly within R, type install.packages("sprof",repos="http://r-forge.r-project.org",type="source")

```
LATEX LAYOUT TOOLS AND R SETTINGS
```

You may want to skip this section, unless you want to modify the vignette for your own purposes, or look at the internals.

To make sure we do not depend on packages collected along the way, we clean up packages. Calls to this funtion will be hidden.

```
cleanpackages <- function()
{
    # make sure we do not get inherited methods or garbage here.
    try(detach("package:sna"), silent=TRUE)
    try(detach("package:igraph"), silent=TRUE)
    try(detach("package:network"), silent=TRUE)
    try(detach("package:graph"), silent=TRUE)
    try(detach("package:Rgraphviz"), silent=TRUE)
    try(detach("package:Rgraphviz"), silent=TRUE)
}</pre>
```

The main library we are going to use is **sprof**.

```
library(sprof)
```

We want immediate warnings, if necessary. So we set warn to level 1. Set warn to level 2 if you want to handle warnings as error.

```
______Input ______
message("switching options(warn=1) -- immediate warning on")
options(warn=1)
```

We want a second chance on errors. So we install an error handler.

```
#options(error = recover)
```

Print parameters used here:

```
options(width = 72)
options(digits = 6)
```

For str output, we generally use these settings:

```
strx <- function(x,
    max.level=2, vec.len=3,
    nchar.max=40,
    list.len=12,</pre>
```

For larger tables and data frames, we use a kludge to avoid long outputs.

```
- Input
xcutrows <- function(df, cut, margin=5)
        if (!is.data.frame(df)) return(df)
        nrow <- nrow(df)</pre>
         # cut a range if it is not empty.
         # Quiet noop else.
         # Does not cut single lines.
         cutrng <- function(cutfrom,cutto, cutname="<cut>"){
                  if (cutfrom<cutto){</pre>
                 df[cutfrom,] <- NA</pre>
                 if (!is.null(rownames(df))) rownames(df)[cutfrom] <- cutname</pre>
                 if (!is.null(df$name)) df$name[cutfrom] <- ""</pre>
                 cutfrom <- cutfrom+1</pre>
                 df[-(cutfrom:cutto),]
                 }#if
        }
         if (!missing(cut)) {df <- cutrng(cut[1],cut[2]); return(df)}</pre>
         if (length(margin)==0) return(df)
        if (length(margin)==1) margin <- c(margin,margin)</pre>
        if (length(margin)==2) {
                 cut <- c(margin[1]+1,nrow-margin[2])</pre>
                 df <-cutrng(cut[1],cut[2]);</pre>
                 return(df)
        delta <- (nrow-sum(margin)) %/% 2</pre>
        if (delta<2) return(df)</pre>
        c1 <- margin[1]+1</pre>
        c2 <- c1 + delta -1
        c4 <- nrow- margin[3]</pre>
         c3 \leftarrow c4 - delta
        df <- cutrng(c3,c4, cutname="<cut hi>")
        df <- cutrng(c1,c2, cutname="<cut low>")
        return(df)
}
```

We use the R function xtable() for output and LATEX longtable. A convenient wrapper to use this in our Sweave source is given here. Among others, it adds zero.print.

```
ToDo: remove text vdots from string/-name columns. Note: this may be a factor. Use empty string.
```

```
_ Input
library(xtable)
prxt <- function (x, digits=2, cut=TRUE, caption=NULL,</pre>
        label=NULL, zero.print=NULL, print.results=TRUE,...) {
        if (cut) {margin <- 10
        if (nrow(x) > 2*margin+3) x <-xcutrows(x, margin=margin)}
        pr <- print(</pre>
                 xtable(x, digits=digits, caption=caption,
                                 label=label, ...),
                 floating=FALSE,
                 tabular.environment="longtable",
                 caption.placement="top",
                 zero.print = ".",
                 NA.string="\\vdots",
                 print.results=FALSE)
                  NA.string="", #NA.string="\\vdots",
                 pr <- gsub( "$\backslash$vdots","\vdots",x=pr, fixed=TRUE)</pre>
                 if(!is.null(zero.print))
                         pr <- gsub( " 0 ",zero.print, x=pr, fixed=TRUE)</pre>
                 if (print.results) cat(pr)
                 invisible(pr)
        }
```

This is to be used with <<pre>cprint=FALSE, results =tex, label=tab:prxx>>=

The graph visualisation family is not friendly. We try to get control by using a wrapper which is at least used to the members of the *graphviz* clan. This will be used in later sections.

```
plotviz <- function(x, layout="dot", main=NULL, sub=NULL,...)
{
    xid <- deparse(substitute(x))
    xsubid <- NULL
    class1 <- NULL
    if (inherits(x, "sprof")) {
        class1 <- class1 <- pasteO("class orig:",class(x))
        xsubid <- x$info$id
        sub <- pasteO(sub, " ",x$info$id)
        x <- as(adjacency(x), "graphNEL")
}

if (!is.null(sub)) sub <- as.character(sub)</pre>
```

```
if (is.null(main))
                main <-paste0("plotviz( ", xid ,", ", layout, " )\n", xsubid) else</pre>
                main=paste0(main, "\n plotviz( ", xid ,", ", layout, " )\n", xsubid)
        if (inherits(x, "Ragraph")) {
        plot(x=x, y=layout,
                        cex.main=1.2,
                main=main,sub=sub,...)
        } else {
                plot(x=x, y=layout,
                attrs=list(
                        node=list(cex=4, fontsize=40, shape="ellipse")),
                cex.main=1.2,
                main=main, sub=sub,...)
        }
         title(sub = paste0(sub, " ", as.character(class(x)))
#
                                                                       )
        legend("topleft",
                legend=c(class1,
                        paste0("class: ",class(x)),
                        paste0("layout: ",layout)),
                bg="#FFFE040",
                seg.len=0
                )#"lightyellow" =#FFFFE0
}
```

1. Profiling

The basic information provided by all profilers is a protocol of sampled stacks. For each recorded event, the protocol has one record, such as a line with a text string showing the sampled stack.

We use profiles to provide hints on the dynamic behaviour of programs. Most often, this is used to improve or even optimise programs. Sometimes, it is even used to understand some algorithm.

Profiles represent the program flow, which is considered to be laid out by the control structure of a program. The control structure is represented by the control graph, and this leads to the common approach to (re)construct the control graph, map the profile to this graph, and used graph based methods for further analysis. The prime example for this strategy is the GNU profiler <code>gprof</code> (see http://sourceware.org/binutils/docs/gprof/) which is used as master plan for many common profilers.

It is only half of the truth that the control graph can serve as a base for the profiled stacks. In R, we have some peculiarities.

lazy evaluation: Arguments to functions can be passed as promises. These are only evaluated when needed, which may be at a later time, and may

then lead to insertions in the stack. So we may have information resulting from the data flow, interspersed with the control flow.

memory management: Allocation of memory, and garbage collection, may interfere and leave their traces in the stack. While allocation is closely related to the visible control flow, garbage collection is a collective effect largely out of control of the code to execute.

primitives: Internal functions may escape the usual stack conventions and execute without leaving any identifiable trace on the stack.

control structures: In R, many control structures are implemented as function. Most notably, the <code>apply()</code> family appears as function calls and can lead to cliques in the graph representation that do not correspond to relevant structures. Since these functions are well know, they can have a special treatment.

So while the stack follows an overall well known dynamics, in R there are exceptions from regularity.

The general approach, by **summaryRprof()** and others, is to reduce the profile to node information, or to consider single transitions.

We take a different approach. We take the stacks, as recorded in the profiles as our basic information unit. From this, we ask: what are the actions we need to answer our questions? Representation in graphs may come later, if they can help.

If the stacks would come from the control flow only, we could make use of the sequential nature of stacks. But since we have to live with the R specific interferences, we stay with the raw stacks.

In this presentation, we will use a small list of examples. Since Rprof is not implemented on all systems, and since the profiles tend to get very large, we use some prepared examples that are frozen in this vignette and not included in the distribution, but all the code to generate the examples is provided.

ToDo: stacks? order?

rearrange detect

1.1. Simple regression example.

```
Input

n <- 10000
x <- runif(n)
err <- rnorm(n)
y <- 2+ 3 * x + err
reg0data <- data.frame(x=x, y=y, err=err)
rm(x,y,err)</pre>
```

We will use this example to illustrate the basics. Of course the immediate questions are the variance between varying samples, and the influence of the sample size n. We keep everything fixed, so the only issue for now is the computational performance under strict iid conditions.

Still we have parameters to choose. We can determine the profiling granularity by setting the timing interval, and we can use repeated measurements to increase precision below the timing interval.

ToDo: Can we calibrate times to CPU rate? Introduce cpu clock cycle as a time base

The timing interval should depend on the clock speed. Using for example 1ms amounts to some 1000 steps on a current CPU, per kernel.

If we use repeated samples, the usual rules of statistics applies. So taking 100 runs and taking the mean reduces the standard deviation by a factor 1/10.

By the usual R conventions, seconds are used as time base for parameters. However report will use ms as a time base.

Here is an example how to take a profile, using basic R. See section 1.1.2 on page 11 how to use **sampleRprof** in package **sprof** for an easier solution.

```
profinterval <- 0.001
simruns <- 100
Rprof(filename="RprofsRegressionExpl.out", interval = profinterval)
for (i in 1:simruns) xxx<- summary(lm(y~x, data=reg0data))
Rprof(NULL)
```

We now have the profile data in a file <code>RprofsRegressionExpl.out</code>. For this vignette, we use a frozen version <code>RprofsRegressionExpl01.out</code>.

1.1.1. R basic. The basic R functions invite us to get a summary.

```
Input
 sumRprofRegressionExpl <- summaryRprof("RprofsRegressionExpl01.out")</pre>
 #str(profile_nodes_rle, max.level=2, vec.len=3, nchar.max=40, list.len=6)
 strx(sumRprofRegressionExpl)
                                    Output -
##strx: sumRprofRegressionExpl
List of 4
$ by.self :'data.frame': 41 obs. of 4 variables:
..$ self.time : num [1:41] 0.087 0.057 0.051 0.043 0.042 0.04 0.032
   0.026 ...
..$ self.pct : num [1:41] 16.67 10.92 9.77 8.24 ...
..$ total.time: num [1:41] 0.113 0.099 0.069 0.043 0.474 0.045 0.033
   0.114 ...
..$ total.pct : num [1:41] 21.65 18.97 13.22 8.24 ...
$ by.total :'data.frame': 62 obs. of 4 variables:
..$ total.time: num [1:62] 0.522 0.522 0.521 0.521 0.521 0.521 0.521
   0.521 ...
..$ total.pct : num [1:62] 100 100 99.8 99.8 ...
..$ self.time : num [1:62] 0.006 0 0.001 0 0 0 0 0 ...
..$ self.pct : num [1:62] 1.15 0 0.19 0 0 0 0 0 ...
$ sample.interval: num 0.001
$ sampling.time : num 0.522
```

The summary reduces the information contained in the profile to marginal statistics per node. This is provided in two data frames giving the same information, only in different order

The file contains several spurious recordings: nodes that have been recorded only few times. It is worth noting these, but then they better be discarded. We use a

time limit of 4ms, which given our sampling interval of 1ms means we require more than four observations.

Table 1: summary Rprof result: by.self as final stack entry, all records

	self.time	self.pct	total.time	total.pct
"lm.fit"	0.09	16.67	0.11	21.65
"[.data.frame"	0.06	10.92	0.10	18.97
"model.matrix.default"	0.05	9.77	0.07	13.22
"as.character"	0.04	8.24	0.04	8.24
"lm"	0.04	8.05	0.47	90.80
"summary.lm"	0.04	7.66	0.04	8.62
"structure"	0.03	6.13	0.03	6.32
"na.omit.data.frame"	0.03	4.98	0.11	21.84
"anyDuplicated.default"	0.02	4.21	0.02	4.21
"as.list.data.frame"	0.02	4.21	0.02	4.21
Zout >	:	:	:	:
<cut></cut>		0.10	. 0.01	1.04
"FUN"	0.00	0.19	0.01	1.34
"%in%"	0.00	0.19	0.00	0.77
"deparse"	0.00	0.19	0.00	0.38
"\$"	0.00	0.19	0.00	0.19
"as.list.default"	0.00	0.19	0.00	0.19
"as.name"	0.00	0.19	0.00	0.19
"coef"	0.00	0.19	0.00	0.19
"file"	0.00	0.19	0.00	0.19
"NCOL"	0.00	0.19	0.00	0.19
"terms.formula"	0.00	0.19	0.00	0.19

```
prxt(

sumRprofRegressionExpl$by.total[

sumRprofRegressionExpl$by.total$total.time>0.004,],

caption="summaryRprof result: by.total, total time > 4ms",

label="tab:prSRREbt")
```

Table 2: summary Rprof result: by.total, total time > 4 ms

	total.time	total nct	self time	self nct
22 . 4 . 22		-		
" <anonymous>"</anonymous>	0.52	100.00	0.01	1.15
"Sweave"	0.52	100.00	0.00	0.00
"eval"	0.52	99.81	0.00	0.19
"doTryCatch"	0.52	99.81	0.00	0.00

"evalFunc"	0.52	99.81	0.00	0.00
"try"	0.52	99.81	0.00	0.00
"tryCatch"	0.52	99.81	0.00	0.00
"tryCatchList"	0.52	99.81	0.00	0.00
"tryCatchOne"	0.52	99.81	0.00	0.00
"withVisible"	0.52	99.81	0.00	0.00
<cut></cut>	:	:	:	:
"as.list"	0.02	4.41	0.00	0.00
"anyDuplicated.default"	0.02	4.21	0.02	4.21
"as.list.data.frame"	0.02	4.21	0.02	4.21
"sapply"	0.01	2.68	0.00	0.19
"match"	0.01	2.11	0.00	0.19
"[[.data.frame"	0.01	1.53	0.00	0.19
"[["	0.01	1.53	0.00	0.00
"rep.int"	0.01	1.34	0.01	1.34
"FUN"	0.01	1.34	0.00	0.19
"list"	0.01	0.96	0.01	0.96

1.1.2. Package sprof. In contrast to the common R packages, in the **sprof** implementation we take a two step approach. First we read in the profile file to an internal representation. Analysis is done in later steps.

```
______Input ______sprof01<- readRprof("RprofsRegressionExpl01.out")
```

The data contain identification information for reference. This will be used in the functions of **sprof** and shown in the displays. Here is the summary of this section:

```
_ Input _
str(sprof01$info)
                                   Output
'data.frame':
                    1 obs. of 9 variables:
                  : chr "RprofsRegressionExpl01.out 2013-06-13 23:46:04"
s id
$ date
                  : POSIXct, format: "2013-08-22 21:01:57"
$ nrnodes
                  : int 62
$ nrstacks
                  : int 50
$ nrrecords
                  : int 522
$ sample.interval: num 0.001
$ sampling.time : num 0.522
                  : chr "sample.interval=1000"
$ ctllines
$ ctllinenr
                  : num 1
```

For this vignette, we change the id information. So in this context:

```
sprof01$info$id <- "sprof01"
```

We keep this example and use the copy **sprof01** of it extensively for illustration.

```
save(sprof01, file="sprof01lm.RData")
```

To run the vignette with a different profile, replace **sprof01** by your example. You still have the file for reference.

Package **sprof** provides a function **sampleRprof()** to take a sample and create a profile on the fly, as in

```
sprof01temp <- sampleRprof(runif(10000), runs=100)
```

The basic data structure consists of four data frames. The <code>info</code> section collects global information from the input file, such as an identification strings and various global matrix. The <code>nodes</code> section initially gives the same information marginal information as <code>summaryRprof</code>. The <code>stacks</code> section puts the node information into their calling context as found in the input profile file. The <code>profiles</code> section gives the temporal context. It is implemented as a list, but conceptually it is a data frame. Implementing it as a list allows run length encoding of variables, which unfortunately is not allowed by R in data frames.

```
strx(sprof01)

##strx: sprof01
```

```
##strx: sprof01
List of 4
$ info :'data.frame': 1 obs. of 9 variables:
..$ id : chr "sprof01"
..$ date : POSIXct[1:1], format: "2013-08-22 21:01:57"
..$ nrnodes : int 62
..$ nrstacks : int 50
..$ nrrecords : int 522
..$ sample.interval: num 0.001
..$ sampling.time : num 0.522
..$ ctllines : chr "sample.interval=1000"
..$ ctllinenr : num 1
$ nodes :'data.frame': 62 obs. of 5 variables:
..$ name : Factor w/ 62 levels "!","..getNamespace",..: 1 2 3 4 5 6 7
   8 ...
..$ self.time : num [1:62] 2 0 2 0 0 57 0 1 ...
..$ self.pct : num [1:62] 0.38 0 0.38 0 ...
..$ total.time: num [1:62] 2 1 4 26 99 99 8 8 ...
..$ total.pct : num [1:62] 0.03 0.01 0.05 0.34 1.29 1.29 0.1 0.1 ...
$ stacks :'data.frame': 50 obs. of 7 variables:
..$ nodes :List of 50
.. .. [list output truncated]
..$ shortname : Factor w/ 50 levels
   "S<A>eFttCtCLtCOdTCwVeesleem.m..n.n...[["| __truncated__,..: 27 17
   19 1 35 36 37 30 ...
..$ refcount : num [1:50] 1 5 26 55 13 43 51 87 ...
..$ stacklength : int [1:50] 19 20 19 21 14 15 15 14 ...
..$ stackheadnodes: int [1:50] 52 52 52 52 52 52 52 ...
..$ stackleafnodes: int [1:50] 27 28 41 6 39 14 38 30 ...
..$ stackssrc : Factor w/ 50 levels "! [.data.frame [
   na.omit.data.frame na."| __truncated__,..: 27 28 39 5 37 13 36 30
profiles:List of 4
..$ data : int [1:522] 1 2 2 3 4 4 5 5 ...
..$ mem : NULL
.. $ malloc : NULL
..$ timesRLE:List of 2
.. ..- attr(*, "class")= chr "rle"
- attr(*, "class")= chr [1:2] "sprof" "list"
```

The nodes do not come in a specific order. Access via a permutation vector is preferred. This allows different views on the same data set. For example, table 4 on page 14 uses a permutation by total time, and a selection (compare to table 2 on page 10). The only difference is that **sprof** works uses a millisecond (ms) base, whereas R in general uses seconds as a base.

Table 3:	splot	result:	by.self.	self	time	>	$4 \mathrm{ms}$

		10	1.0	1	1
	name	self.time	self.pct	total.time	total.pct
30	$\operatorname{lm.fit}$	87.00	16.67	113.00	1.47
6	[.data.frame	57.00	10.92	99.00	1.29
38	model.matrix.default	51.00	9.77	69.00	0.90
14	as.character	43.00	8.24	43.00	0.56
29	lm	42.00	8.05	474.00	6.16
51	summary.lm	40.00	7.66	45.00	0.59
49	structure	32.00	6.13	33.00	0.43
41	na.omit.data.frame	26.00	4.98	114.00	1.48
13	any Duplicated. default	22.00	4.21	22.00	0.29
16	as.list.data.frame	22.00	4.21	22.00	0.29
40	na.omit	20.00	3.83	134.00	1.74
39	model.response	13.00	2.49	56.00	0.73
36	model.frame.default	12.00	2.30	168.00	2.18
46	rep.int	7.00	1.34	7.00	0.09
10	<anonymous></anonymous>	6.00	1.15	522.00	6.79
28	list	5.00	0.96	5.00	0.07

At this level, it is helpful to note the expectations, and only then inspect the timing results. Since we are using a linear model, we are not surprised to see functions related to linear models on the top of the list. We may however be surprised to see functions related to data access and to character conversion very high on the list. The sizeable amount of time spent on NA handling is another aspect that is surprising.

Table 4: splot result: by.total, total time > 4ms

	name	self.time	self.pct	total.time	total.pct
10	<anonymous></anonymous>	6.00	1.15	522.00	6.79
52	Sweave	0.00	0.00	522.00	6.79
21	doTryCatch	0.00	0.00	521.00	6.78
22	eval	1.00	0.19	521.00	6.78
23	evalFunc	0.00	0.00	521.00	6.78
55	try	0.00	0.00	521.00	6.78
56	tryCatch	0.00	0.00	521.00	6.78
57	tryCatchList	0.00	0.00	521.00	6.78

58	tryCatchOne	0.00	0.00	521.00	6.78
62	withVisible	0.00	0.00	521.00	6.78
<cut></cut>	\vdots	:	:	:	:
61	vapply	3.00	0.57	23.00	0.30
13	anyDuplicated.default	22.00	4.21	22.00	0.29
16	as.list.data.frame	22.00	4.21	22.00	0.29
47	sapply	1.00	0.19	14.00	0.18
31	match	1.00	0.19	11.00	0.14
7	[[0.00	0.00	8.00	0.10
8	[[.data.frame	1.00	0.19	8.00	0.10
25	FUN	1.00	0.19	7.00	0.09
46	rep.int	7.00	1.34	7.00	0.09
28	list	5.00	0.96	5.00	0.07

<u>ToDo:</u> remove text vdots from string/name columns

Given the sampling structure of the profiles, two aspect are common. The sampling picks up scaffold functions with a high, nearly constant frequency. And the sampling will pick up rare recordings that are near to detection range. The display functions hide these effects by default. In our example, about half of the nodes are cleared by this garbage collector.

Common rearrangements as by total time and by self time are supplied by the display functions.

<code>plot_nodes()</code>, for example, currently gives a choice of four displays for nodes, and supports trimming by default. Our profile starts with 62 nodes. The defaults cut off 34 nodes as uninformative, either because they are too rare, or ubiquitous.

```
#8
oldpar <- par(mfrow=c(2,2))
plot_nodes(sprof01)
par(oldpar)
```

Basic information on node level: see fig. 1 on the facing page.

Information in the time scatterplots may sometimes more accessible when using a logarithmic scale, so this is added.

If you prefer, you can have the bar charts in horizontal layout, giving more space for labels (Basic information on node level - horizontal bars: see fig. 2 on page 16).

```
#8
oldpar <- par(mfrow=c(2,2))
plot_nodes(sprof01, horiz=TRUE, cex.names=0.5)
par(oldpar)
```

We can add colour. To illustrate this, we encode the frequency of the nodes as colour. As a palette, we choose a heat map here.

<u>ToDo:</u> apply colour to selection?

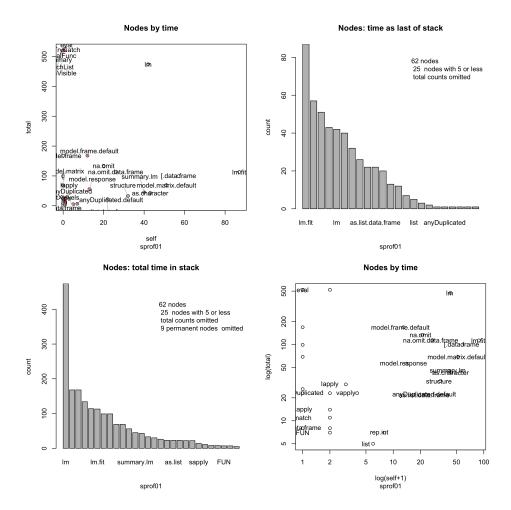


FIGURE 1. Basic information on node level

Note: we chose a colour palette for the full data set. If we restrict a display to a selection, only a fraction of the palette may be visible. As an alrernative, you can adjust the aplette to focus on your selection.

```
Input _____
freqrank01 <- rkindex(-sprof01$nodes$total.time, ties.method="random")
freqrankcol01 <- heat.colors(length(freqrank01))</pre>
```

Here is the node view using these choices (Basic information on node level, colour by total time see fig. 3 on the facing page).

```
#10
sprof01$nodes$icol <- freqrank01
oldpar <- par(mfrow=c(2,2))
```

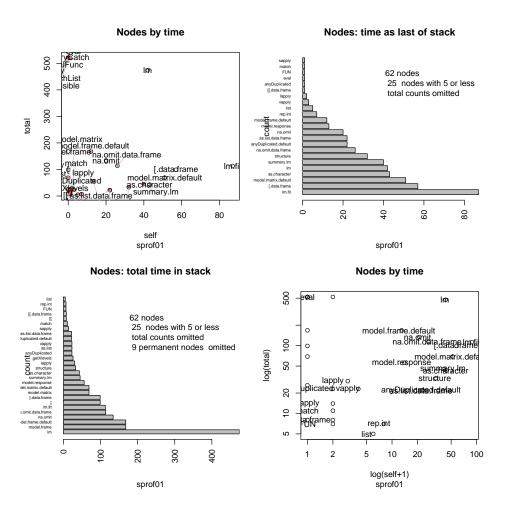


FIGURE 2. Basic information on node level - horizontal bars

plot_nodes(sprof01, col=freqrankcol01)
par(oldpar)

Colour is considered a volatile attribute. So you may need to pay some attention to keep colour indices (and colour palettes) aligned to your context. You may want to do experiments with colour, trying to find a good solution for your visual preferences. The recommended way is to use some stable colour index (the slot icol is reserved for this) and use this as an index to a choice of colour palettes. So icol becomes a part of the data structure, and the colour palette to be used is passed as a parameter. Package RColorBrewer may be a helpful source for colour palettes. You find more information and references on this in the vignette "Bertin matrices" of package bertin, http://bertin.r-forge.r-project.org.

<u>ToDo:</u> improve colour: support colour in a structure

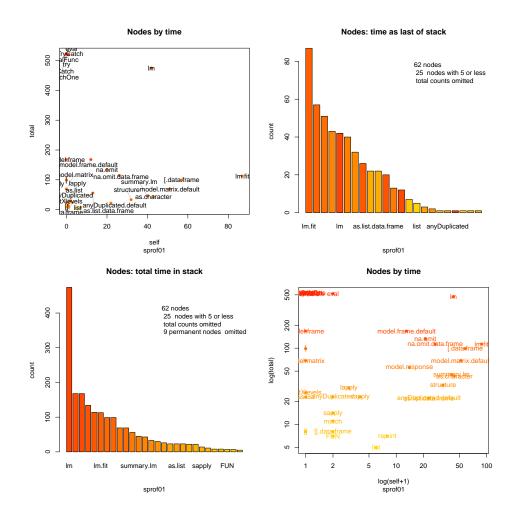


FIGURE 3. Basic information on node level, colour by total time.

1.1.3. *Node classes*. We can add attributes to the plots. But we can also add attributes to the nodes, and use these in the plots. In principle, this has been alway available. We are now making explicit use of this possibility.

The attribute <code>icol</code> is a special case which we used above. If present, it will be interpreted as an index to a colour table. For example, we can collect special well known functions in groups.

The node information is to some part arbitrary. You may achieve the same functionality by different functions, and you will see different load in the profiles. Grouping nodes may be a mean to clarify the picture.

Grouping may also help you to focus your attention. "HOT" and "cold" may be ver helpful tags. These can be used in a flexible way.

<u>ToDo:</u> colour by class – redo. Bundle colour index with colour?

<u>ToDo:</u> Move class attributes to package code <u>ToDo:</u> add class by

keyword

```
nodekeyword0 <- function(node)</pre>
 {
 }
 nodepackages <- nodepackage(sprof01$nodes$name)</pre>
 names(nodepackages) <- sprof01$nodes$name</pre>
 table(nodepackages)
                                   _ Output _
nodepackages
<not found>
                                stats
                                            utils
                    base
                      41
                                  14
                                     Input _
 sprof01$nodes$icol <-as.factor(nodepackages)</pre>
Nodes by package, colours by RColorBrewer: see fig. 4 on the next page.
x_apply <- c("apply", "lapply", "vapply", "sapply")
 x_as <- c("as.list", "as.data.frame", "as.list.data.frame",</pre>
         "as.character", "as.list.default", "as.name")
(Extend as you need it) and then use, as for example:
 nodeclass <- rep("x_nn", sprof01$info$nrnodes)
 nodeclass[sprof01\$nodes\$name \ \%in\% \ x\_apply] <- \ "x\_apply"
 nodeclass[sprof01$nodes$name %in% x_as] <- "x_as"</pre>
or use assignments on the fly
                                      Input _
 nodeclass[sprof01$nodes$name %in%
         c("eval", "evalFunc",
                  "try", "tryCatch", "tryCatchList", "tryCatchOne",
                  "doTryCatch", "withVisible")
                  ] <- "x_eval"
 nodeclass[sprof01$nodes$name %in%
         c("model.frame", "model.matrix.default", "model.frame.default",
          " model.response", "model.matrix", "model.response")
                  ] <- "x_model"
 nodeclass[sprof01$nodes$name %in%
         c("lm", "lm.fit", "summary.lm")
                  ] <- "x_1m"
 nodeclass[sprof01$nodes$name =="<Anonymous>"] <- "x_Anon"</pre>
 nodeclass[sprof01$nodes$name == "Sweave"] <- "x_Sweave"</pre>
 nodeclass[sprof01$nodes$name %in% c( "summary" , "summary.lm")] <-</pre>
           "x_summary"
```

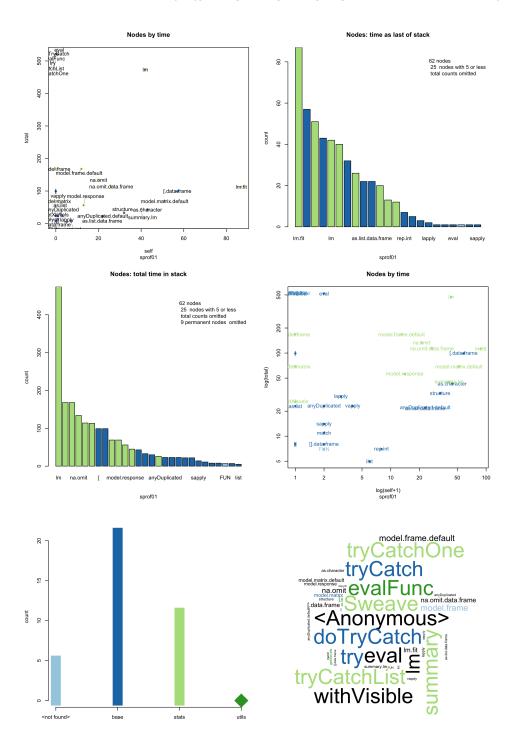


FIGURE 4. Nodes by package, colours by RColorBrewer.

```
sprof01$nodes$icol <-as.factor(nodeclass)</pre>
```

adds a sticky colour attribute. For interpretation, you should choose your preferred colour palette, for example

```
nodeclasscol <- c("red", "green", "blue", "yellow",

"cyan", "magenta", "purple",

"brown", "aquamarine")
```

<u>ToDo:</u> Defaults by class

gold cyan4 aquamarine pink violet orchid hotpink salmon turquoise1

```
#8 12
oldpar <- par(mfrow=c(3,2))
plot_nodes(sprof01, which=1:6, col=nodeclasscol)
par(oldpar)
```

<u>**ToDo:**</u> classes need separate colour palette

Nodes by class, user defined colors: see fig. 5 on the facing page.

```
#8 12
oldpar <- par(mfrow=c(3,2))
plot_nodes(sprof01, which=1:6)
par(oldpar)
```

Nodes by class: default colour selection: see fig. 6 on page 22.

You can break down the frequency by classes of your choice. But beware of Simpson's paradox. The information you think you see may be strongly affected by your choices - what you see are reflections of conditional distributions. These may be very different from the global picture.

If package wordcloud is installed, a different view is possible. This is added in the plots above.

2. A BETTER GRIP ON PROFILE INFORMATION

The basic information provided by all profilers in R is a protocol of sampled stacks. The conventional approach is to break the information down to nodes and edges. The stacks provide more information than this. One way to access it is to use linking to pass information. This has already been used on the node level in section 1.1.2 on page 11.

ToDo: add attributes to stacks, and discuss scope
ToDo: sorting/arranging stacks

2.1. **The internal details.** For each recorded event, the protocol records one line with a text string showing the sampled stack (in reverse order: most recent first). The stack lines may be preceded by header lines with event specific information. The protocol may be interspersed with control information, such as information about the timing interval used. ?? on page ??.

We know that the structural information, static information as well as dynamic information, can be represented with the help of a graph. For a static analysis,

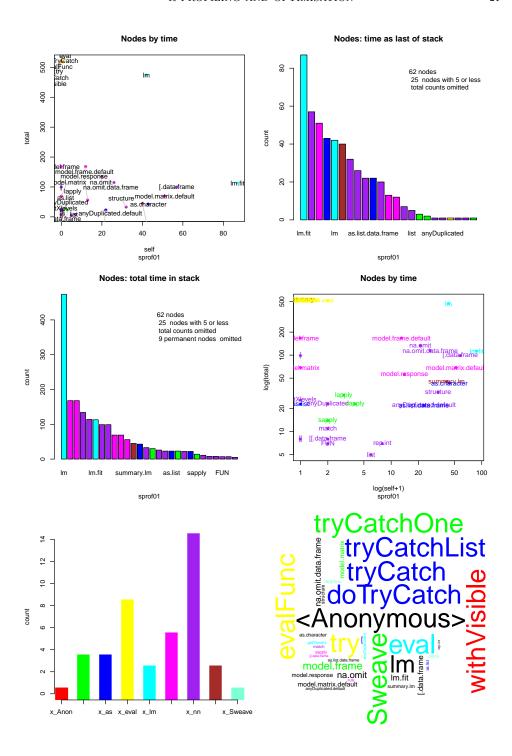


Figure 5. Nodes by class. User defined colors.

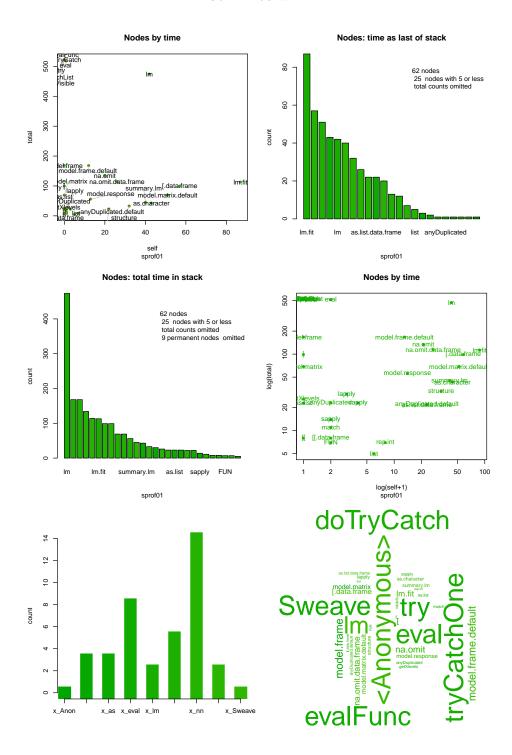


Figure 6. 8 12 Nodes by class: default colour selection

the graph representation may be the first choice. For a dynamic analysis, the stack information is our first information. A stack is a connected path in the program graph. If we start with nodes and edges, we loose information which is readily available in record of stacks.

As we know that we are working with stacks, we know that they have their peculiarities. Stacks tend to grow and shrink. Subsequent events will have extensions and shrinkages of stacks (if the recording is on a fine scale), or stack sharing common stumps (if the recording is on a coarser scale). We could exploit this information, but it does not seem worth the effort.

There have always been interrupts, and these show up in profiles. In R, there is a related problem: garbage collection (GC) may interfere and leave traces in the stack.

Stack information is first. The call graph is a second instance that is (re)constructed from the stack recording. The graph represents cumulated one-step information. Longer scale information contained in the stacks is lost in the graph.

Here is the way we represent the profile information:

The profile log file is sanitised:

- Control lines are extracted and recorded in a separate list.
- Head parts, if present, are extracted and recorded in a matrix that is kept line-aligned with the remainder
- Line content is standardised, for example by removing stray quotation marks etc.

After this, the sanitised lines are encoded as a vector of stacks, and references to this.

Note: after sanitising, stacks may have an empty node list. However they should not be removed as long as they may still be present in the profile.

If necessary, these steps are done by chunks to reduce memory load.

From the vector of stacks, a vector of nodes (or rather node names) is derived.

The stacks are now encoded by references to the nodes table. For convenience, we keep the (sanitised) textual representation of the stacks. (This may change.)

So far, texts are in reverse order. For each stack, we record the trailing leaf, and then we reverse order. The top of stack is now on first position.

Several statistics can be accumulated easily as a side effect.

Conceptually, the data structure consist of three tables (the implementation may differ, and is subject to change).

The profiles table is the representation of the input file. Control lines are collected in a special table. With the control lines removed, the rest is a table, one row per input line. The body of the line, the stack, is encoded as a reference to a stacks table (obligatory) and header information (optional).

<u>ToDo:</u> re-think: sort stacks

The stacks table contains the collected stacks, each stack encoded as a list of references to the node table. This is obligatory. This list is kept in reverse order (root at position 1). A source line representing the stack information may be kept (optional).

The nodes table keeps the names at the nodes.

Sometimes, it is more convenient to use a simple representation, such as a matrix. Several extraction routines are provided for this, and the display routines make heavy use of this. See table 5.

<u>ToDo:</u> complete matrix conversion

Table 5. Extraction and conversion routines

<pre>profiles_matrix()</pre>	incidence matrix: nodes by event
stacks_matrix()	incidence matrix: nodes by stack
list.as.matrix()	fill list to equal length and convert to matrix
stackstoadj()	stacks to (correspondence) adjacency matrix
adjacency()	sprof to (correspondence) adjacency matrix

We now can go beyond node level.

This is what we get for free from the node information on our three levels: node, stack, and profile.

```
ToDo: check and stabilise colour linking
ToDo: clean up colour handling
```

```
#8 rainbow
sprof01$nodes$icol <- freqrank01; freqrankcol <- rainbow(62)
shownodes(sprof01, col=freqrankcol)
```

See fig. 7 on the facing page for a summary of nodes by stack and profile.

The obvious message is that if seen by stack level, there are different structures. Profiling usually takes place in a framework. So at the base of the stacks, we find entries that are (almost) persistent. Then usually we have some few steps where the algorithm splits, and then we have the finer details. These can be identified using information on the stack level, but of course they are not visible on the node or edge level in a graph representation. On the stack level, we see a socket. If we want a statistic, we can look at number of different nodes by level.

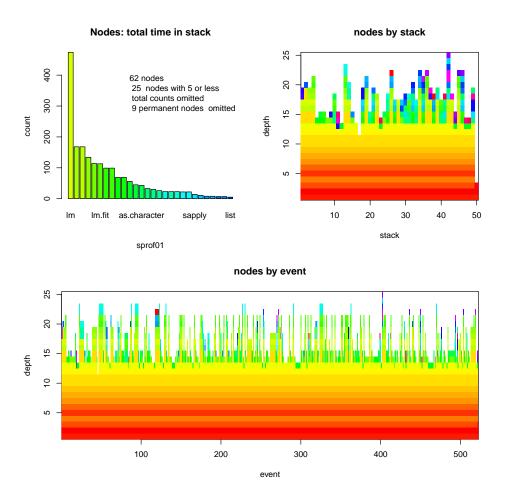


FIGURE 7. Nodes by stack and profile

Nr. of unique nodes by stack level: See fig. 8 on the next page.

We will come to finer tools in section 2.4 on page 46 but for the moment the rough information should suffice to take a decision. In our example, it is only a matter of taste whether we cut off 12 levels, or we want to work with five components after cutting 13 levels three leaves us to start with five roots on the next level in our example.

Not so often, but a frequent phenomenon is to have some "burn in" or "fade out". To identify this, we need to look at the profile level. The indicator to check is

<u>ToDo:</u> check and synchronise

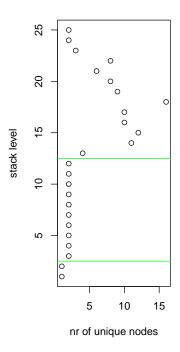


FIGURE 8. Nr of unique nodes by stack level.

to whether we have very low frequency stacks at the beginning or the end of our recording. The counts to be takes as reference can be seen from the summary.

summary(s	prof01	\$stacks\$r	refcount)	_ Input		
				_ Output		
Min. 1s	t Qu.	Median	Mean 3r			
1.0	1.0	2.0	10.4	12.0	87.0	

<u>ToDo:</u> we could do smart smoothing of the stacks here

This summary has to be taken with caution. As the program runs, the stacks are build up und teared down, and we only take random samples. So in dynamic parts, we see images with some fluctuation, as one stack may be a snapshot of an other under construction. A better information is to cut off fluctuations and use this summary as a reference.

Input								
<pre>summary(sprof01\$stacks\$refcount[sprof01\$stacks\$refcount>2])</pre>								
					Output			
Min.	1st	Qu.	Median	Mean	3rd Qu.			
3 00	-	7 75	16 00	24 30	10 50	97 00		

```
__ Input _
```

Table 6: Stacks by event: burn in/fade out

	stack	count
1	1	1
2	2	5
3	2	5
4	3	26
5	4	55
6	4	55
7	5	13
8	5	13
9	6	43
10	7	51
<cut></cut>	:	:
513	3	26
514	3	26
515	3	26
516	3	26
517	4	55
518	3	26
519	36	2
520	16	42
521	44	2
522	50	1

Here at least one recording on either side is a candidate to be off. We may have a look at the next recordings and decide to go beyond and cut off events 1:3 and 519:522.

At a closer look, we may find stack patterns (maybe marked by specific nodes) that indicate administrative intervention and rather should be handled as separators between distinct profiles rather than as part of the general dynamics. Again we may use some indicator nodes to be used as marker for special stacks.

Stable framework effects sometimes are obvious and can be detected automatically. "burn in" or "fade out' may need a closer look, and special stacks need and individual inspection on low frequency stacks. Tools for trimming are in section 2.3.1 on page 32.

ToDo: example

<u>**ToDo:**</u> colours. recolour. Propagate colour to graph.

2.2. **The free lunch.** What you have seen so far is what you get for free when using package *sprof*.

If you want to wrap up the information and look at it from a graph point of view, here is just one example. More are in section 3 on page 51 and vrefsec:moregraph. But before changing to the graph perspective, we recommend to see the next sections, not to skip them.

The preview, at this point, taking package <code>graph</code> as an example ¹. <code>graph</code> on its side has an undocumented feature: it needs <code>Rgraphviz</code> to handle graph attributes². We have to take two steps. We extract the graph information from <code>sprof</code>. Using an adjacency matrix is a simple solution here. This is then converted to the <code>"graph-NEL"</code> format which is shared by <code>graph</code> and <code>Rgraphviz</code>. <code>Rgraphviz</code> is hidden in the use of <code>plot()</code>. So here is a bare foot approach. A more sophisticated function implementation is in section 3 on page 51.

¹Package 'graph' was removed from the CRAN repository.

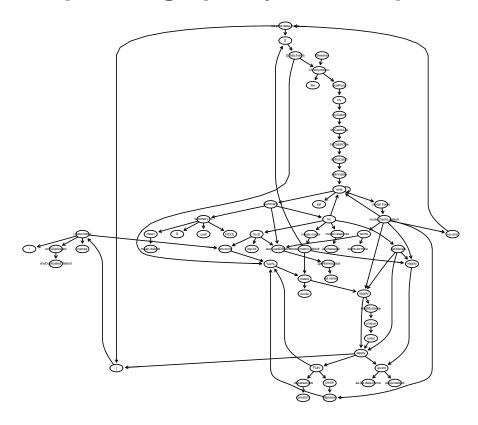
This package is now available from Bioconductor only, see http://www.bioconductor.org/packages/release/bioc/html/graph.html.

²Package 'Rgraphviz' was removed from the CRAN repository.

This package is now available from Bioconductor only.

See http://www.bioconductor.org/packages/release/bioc/html/Rgraphviz.html.

sprof01: graph layout example



2.3. Cheap thrills. Before starting additional inspection, the data better be trimmed. Trimming routines are in section 2.3.1 on page 32, but the data structure is robust enough to allow manual intervention as used here.

```
sprof02 <- sprof01; sprof02$info$id <- "sprof02: trimmed"
```

On the stack level, we take brute force to cut off the basic stacks.

```
basetrim <- 13
sprof02$stacks$nodes <- sapply(sprof02$stacks$nodes,
    function (x){if (length(x)> basetrim) x[-(1:basetrim)] })
```

We have noted burn in/fade out. This is on the profile level. Taking the big knife is not advisable, since time information and stack data must be synchronised. So we are more cautious.

ToDo: updateRprof needs careful checking. For now, we are including long listings here to provide the necessary information

ToDo: handle empty stacks and zero counts gracefully

<u>ToDo:</u> add a purge function

```
sprof02$profiles$data[1:3] <- NA
sprof02$profiles$data[519:522] <- NA</pre>
```

At this point, it is a decision whether to adapt the timing information, or keep the original information. Since this decision does affect the structural information, it is not critical. But analysis is easier if unused nodes are eliminated. The <code>info</code> section is inconsistent at this point. Another reason to call <code>updateRprof()</code>.

```
____ Input _
 strx(sprof02$info)
                                __ Output _
##strx: sprof02$info
'data.frame': 1 obs. of 9 variables:
$ id : chr "sprof02: trimmed"
$ date : POSIXct, format: "2013-08-22 21:01:57"
$ nrnodes : int 62
$ nrstacks : int 50
$ nrrecords : int 522
$ sample.interval: num 0.001
$ sampling.time : num 0.522
$ ctllines : chr "sample.interval=1000"
$ ctllinenr : num 1
                         _____ Input __
 prxt(sprof02$nodes,
         caption="sprof02, before update",
         label="tab:sprof02info1",
         digits=c(0,0,0,2,0,2,0),
         zero.print=" . "
         )
```

Table 7: sprof02, before update

	name	self.time	self.pct	total.time	total.pct	icol
1	!	2	0.38	2	0.03	50
2	get Name space	•	0.00	1	0.01	55
3	. departs = Opts	2	0.38	4	0.05	41
4	.get X levels		0.00	26	0.34	28
5	[•	0.00	99	1.29	20
6	[.data.frame	57	10.92	99	1.29	19
7	[[•	0.00	8	0.10	36
8	[[.data.frame	1	0.19	8	0.10	35
9	%in $%$	1	0.19	4	0.05	40
10	<anonymous></anonymous>	6	1.15	522	6.79	2
<cut></cut>	\vdots	:	:	:	:	:
53	terms	•	0.00	2	0.03	45
54	terms.formula	1	0.19	1	0.01	54
55	try	•	0.00	521	6.78	7
56	tryCatch	·	0.00	521	6.78	4
57	tryCatchList		0.00	521	6.78	6

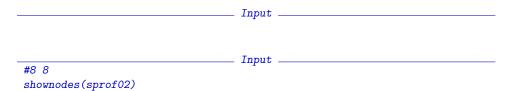
58	tryCatchOne		0.00	521	6.78	8
59	unique	3	0.57	4	0.05	43
60	unlist		0.00	1	0.01	56
61	vapply	3	0.57	23	0.30	30
62	withVisible		0.00	521	6.78	10

```
_____ Input ____
                                _ Input _
 sprof02 <- updateRprof(sprof02)</pre>
 sprof02$info$id <- "sprof02 updated"</pre>
                           _____ Input __
strx(sprof02$info)
                      _____ Output _____
##strx: sprof02$info
'data.frame': 1 obs. of 10 variables:
$ id : chr "sprof02 updated"
$ date : POSIXct, format: "2013-08-22 21:01:57"
$ nrnodes : int 62
$ nrstacks : int 50
$ nrecords : int 522
$ sample.interval: num 0.001
$ sampling.time : num 0.522
$ ctllines : chr "sample.interval=1000"
$ ctllinenr : num 1
$ date_updated : POSIXct, format: "2013-08-22 21:02:04"
                      _____ Input __
prxt(sprof02$nodes,
        caption="sprof02, after update",
        label="tab:sprof02info2",
        digits=c(0,0,0,2,0,2,0),
        zero.print=" . "
```

Table 8: sprof02, after update

	name	self.time	self.pct	total.time	total.pct	icol
1	!	1	0.23	1	0.06	50
2	getNamespace		0.00	1	0.06	55
3	. departseOpts	2	0.46	4	0.25	41
4	.get X levels		0.00	26	1.64	28
5	[0.00	98	6.17	20
6	[.data.frame	57	13.16	98	6.17	19
7	[[0.00	8	0.50	36
8	[[.data.frame	1	0.23	8	0.50	35
9	%in $%$	1	0.23	4	0.25	40

10	<anonymous></anonymous>	6	1.39	6	0.38	2
<cut></cut>	\vdots	÷	:	÷	:	:
53	terms		0.00	1	0.06	45
54	terms.formula	1	0.23	1	0.06	54
55	try		0.00		0.00	7
56	tryCatch		0.00		0.00	4
57	tryCatchList		0.00		0.00	6
58	tryCatchOne		0.00		0.00	8
59	unique	3	0.69	4	0.25	43
60	unlist		0.00	1	0.06	56
61	vapply	3	0.69	23	1.45	30
62	withVisible		0.00		0.00	10



Nodes by stack and profile: see fig. 9 on the next page.

2.3.1. Trimming. Note: trimming may be supported by the graph packages. If you are more familiar with your graph package, you may prefer to handle trimming there.

To decide about trimming, we can use the displays shown above, or we can use some statistics. Global trimming works on the stack level. To make life easy, we can imbed the stack information in a matrix and use marginals.

```
stackm01 <- list.as.matrix(sprof01$stacks$nodes)</pre>
nr_uniquenodes01 <- apply(stackm01,1, function(x) { length(unique(x))})</pre>
names( nr_uniquenodes01)<- rownames(nr_uniquenodes01,FALSE,"L")</pre>
nr_uniquenodes01
                                 Output
L1 L2
        L3 L4 L5 L6 L7
                           L8 L9 L10 L11 L12 L13 L14 L15 L16 L17 L18
    1
         2
            2
                2
                    2
                        2
                            2
                               2 2 2 2 4 11 12 10 10 16
L19 L20 L21 L22 L23 L24 L25
    8
         6
             8 3
                   2
                                 _ Input -
rm(stackm01, nr_uniquenodes01)
```

<u>ToDo:</u> This section needs to be reworked

```
rsprof01Tr <- trimstacks(sprof01, level=11)
```

After trimming, it is worth to inspect the result and to see whether additional trimming is helpful. Function $roots_sprof$ is handy.

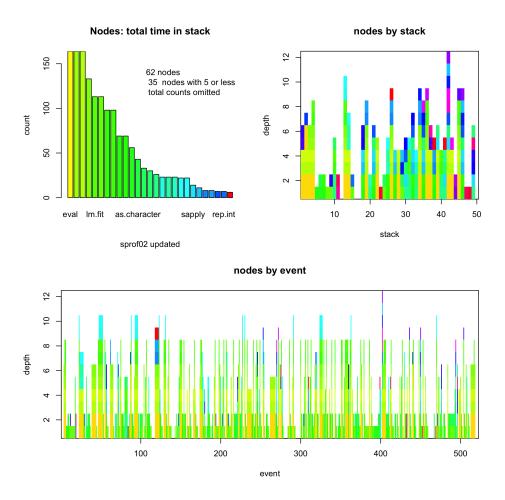


FIGURE 9. sprof02: Nodes by stack and profile

		Input	
roots_sp	rof(sprof01,	stacks=rsprof01Tr)	
- •	•	•	
		Output	
summary	<na></na>	•	
50	NA		

Here after trimming we have one root node. This may be esthetically pleasing, but it is more informative to note that *summary* is a common root, and then cut it off (thus fragmenting the graph into components).

```
rsprof02Tr <- trimstacks(sprof01, level=12)
roots_sprof(sprof01, stacks=rsprof02Tr)

Output
```

lm lazyLo	oadDBfetch	summary.lm	<na></na>
29	27	51	NA

Now this is revealing. We know how our test example was contructed. So we are prepared to find 1m and summary.1m at prominent positions. But it is surprising to find lazyLoadDBfetch as dominant node, and with a frequency comparable to that of 1m.

<u>ToDo:</u> move to other section

There is no statistics on profiles. Profiles are our elementary data. However we can link to our derived data to get a more informative display. For example, going one step back we can encode stacks and use these colour codes in the display of a profile.

Or going two steps back, we can encode nodes in colour, giving coloured stacks, and use these in the display of profile data.

2.3.2. Surgery. Note: surgery may be supported by the graph packages. Use the implementation you are more familiar with. Surgery can also been done conveniently on the source level, using simple replace statements.

Looking at nodes gives you a point-wise horizon. Looking at edges gives you a one step horizon. The stacks give a wider horizon, typically a step size of 10 or more. The stacks we get from R have peculiarities, and we can handle with this broader perspective. These are not relevant if we look point- wise, but may become dominating if we try to get a global picture. We take a look ahead (details to come in section 3 on page 519 and have a preview how our example is represented as a graph. Left is the original graph as recovered from the edge information, right the graph after we have cut off the scaffold effects.

ToDo: cut next level

Control structures may be represented in R as function, and these may lead to concentration points. Using information from the stacks, we can avoid these by introducing substitute nodes on the stack level. For example, <code>lapply</code> is appearing in various contexts and may be confusing any graph representation. We can avoid this by replacing a short sequence.

```
ToDo: Implement. "["
Currently best han-
```

dled on source=text level

ToDo: function addnode using "call by reference" to be added

```
"[" "lapply" ".getXlevels" -> "<.getXlevels_[>"
```

If the node does not exist, we want to add it to our global variable. For now, we do it using expressions on the R basic level and avoid tricks like simulating "call by reference".

```
sprof03 <- sprof02; sprof03$info$id <- "sprof03: surgery"
node <- "<.getXlevels_[>"
#nodei <- function(sprofx, node, warn = TRUE)
{
    i <- match(node, sprof03$nodes$name, nomatch=0)
    if (i==0) {
        sprof03$nodes$name <- as.character(sprof03$nodes$name)
        sprof03$nodes <- rbind(sprof03$nodes,NA)
        i <- length(sprof03$nodes$name)
        sprof03$nodes$name[i] <- node
    if (as.logical(options("warn")))</pre>
```

```
message("node added. An updateRprof() may be necessary.")
}
nodei <- i
}
# sprof <- sprof01; nodei(sprof, "kiki"); sprof$nodes</pre>
```

Now we have to identify the stacks that may get a replacement. First find the candidates.

```
Input

targeti <- match("lapply", sprof03$nodes$name, nomatch=0)

found <- lapply(sprof03$stacks$nodes, function(X) match(targeti,X))

found <- data.frame(stack=1:length(found), position=as.matrix(found))

found[!is.na(found$position),]
```

```
_ Output _
   stack position
10
     10
                 2
21
      21
                 5
30
      30
                 3
31
      31
                 2
32
      32
                 5
33
      33
                 2
35
      35
                 5
39
      39
                 6
                 2
40
      40
                 3
41
      41
                 6
42
      42
                 5
46
      46
      49
                 6
```

For now, these are just candidates.

```
Other candidates are:
```

```
"as.list" "vapply" "model.frame.default" -> "<model_as.list>"
or
"as.list" "vapply" "model.matrix.default" -> "<model_matrix_as.list>"
```

ToDo: xreplacenodes: improve implement

```
newchopnode <- function(nodenames, chop) {
tmpname <- paste("<",as.character(nodenames[chop]),">")
# chec for existing.
# add if necessary
tmpname
}
chopstack <- function(x , chop, replacement)
{
# is chop in x`
# y: cut x.
# merge x <- head + replacement + tiail
return(x)</pre>
```

```
}
x <-sprof01$stacks$nodes[1]</pre>
```

<u>ToDo:</u> clean up factor handling

So far, we use factor indices only.

```
firstpos <- 1; lastpos <- length(x) - length(oldseq) +1
        if (lastpos <1) return(0)</pre>
        1 <- length(oldseq)-1</pre>
        while (firstpos <= lastpos) {</pre>
                 if ( isTRUE(all.equal(oldseq , x[firstpos:(firstpos+1)])) )
                         {return(firstpos) }
                 firstpos <- firstpos+1
        return(0)
}
                                   _ Input
xreplace <- function(oldseq, newseq, x){</pre>
#! handle multiple replacements
        wh <- xwhere(oldseq,x)</pre>
        if (wh) {
        1 <- length(oldseq)-1</pre>
        if (wh>1) x1 \leftarrow c(x[1:(wh-1)], newseq) else x1 \leftarrow newseq
        if (wh+1 < length(x)) x <- c(x1, x[-(1:(wh + 1))]) else x <- x1
        return(x)
        } else return(x)
}
                                   - Input
nodenames2index <- function(names, sprof){</pre>
if (is.character(names))
                 { sapply(names, function(x) {match(x,sprof$nodes$name)})
                   #if (is.na(trimnode)) return(ts)
                 } else names
}
nodenames2index(c("[","lapply",".getXlevels"), sprof01)
                                   Output .
         lapply .getXlevels
         5
                     26
                                    Input
xreplacenodes <- function(sprof, oldseq, newseq)</pre>
        if (is.character(oldseq)) oldseq <- nodenames2index(oldseq,sprof)</pre>
        if (is.character(newseq)) newseq <- nodenames2index(newseq,sprof)</pre>
        stacks <- sprof$stacks$nodes
```

sapply(stacks, function(x){xreplace(oldseq, newseq, x)})

}

xrep	lace	node	s(sp	rof0	1, c	(10,	23,	I: 55)	nput ,999)							
								O11	tput								
[[1]]	52	999	56	57	58	21	62		_			22	22	35	36	53	27
[[2]] [1] [18]	52 28	999	56	57	58	21	62	22	22	50	29	22	22	35	36	22	22
[[3]] [1]	52	999	56	57	58	21	62	22	22	50	29	22	22	35	36	40	41
[[4]] [1] [18]	52 5	999 6	56	57	58	21	62	22	22	50	29	22	22	35	36	40	41
[[5]] [1]	52	999	56	57	58	21	62	22	22	50	29	39					
[[6]] [1]	52	999	56	57	58	21	62	22	22	50	29	39	14				
[[7]] [1]	52	999	56	57	58	21	62	22	22	50	29	37	38				
[[8]] [1]	52	999	56	57	58	21	62	22	22	50	29	30					
[[9]] [1]	52	999	56	57	58	21	62	22	22	50	29	30	27				
[[10] [1]		999	56	57	58	21	62	22	22	50	29	4	26	5	6	49	
[[11] [1]		999	56	57	58	21	62	22	22	50	27	2	18				
[[12] [1]		999	56	57	58	21	62	22	22	50	51						
[[13] [1] [18]	52				58	21	62	22	22	50	29	22	22	35	36	40	41
[[14] [1] [18]	52	999	56	57	58	21	62	22	22	50	29	22	22	35	36	61	15

[[31]]

[1] 52 999 56 57 58 21 62 22 22 50 29 4 26 5 6 12

[[32]] [1] 52 999 [18] 3	56	57	58	21	62	22	22	50	29	37	38	31	47	26	25
[[33]] [1] 52 999	56	57	58	21	62	22	22	50	29	4	26	15	16		
[[34]] [1] 52 999 [18] 7 8	56 9	57	58	21	62	22	22	50	29	22	22	35	36	40	41
[[35]] [1] 52 999 [18] 3 45	56	57	58	21	62	22	22	50	29	37	38	31	47	26	25
[[36]] [1] 52 999 [18] 5 6	56 1	57	58	21	62	22	22	50	29	22	22	35	36	40	41
[[37]] [1] 52 999	56	57	58	21	62	22	22	50	29	22	22	35	36	53	54
[[38]] [1] 52 999	56	57	58	21	62	22	22	50	29	37	38	7	8	10	
[[39]] [1] 52 999	56	57	58	21	62	22	22	50	29	22	22	35	36	47	26
[[40]] [1] 52 999	56	57	58	21	62	22	22	50	29	4	26	5	6		
[[41]] [1] 52 999	56	57	58	21	62	22	22	50	29	4	47	26	15	17	
[[42]] [1] 52 999 [18] 25 44	56 20	57 9	58 31	21 34	62	22	22	50	29	22	22	35	36	47	26
[[43]] [1] 52 999	56	57	58	21	62	22	22	50	29	22	22	35	36	20	
[[44]] [1] 52 999	56	57	58	21	62	22	22	50	51	32	33				
[[45]] [1] 52 999 [18] 5 6	56 42	57	58	21	62	22	22	50	29	22	22	35	36	40	41
[[46]] [1] 52 999 [18] 9 31		57	58	21	62	22	22	50	29	37	38	31	47	26	25
[[47]]															

```
[1] 52 999 56 57 58 21 62 22 22 50 51 43
[[48]]
[1] 52 999 56 57 58 21 62 22 22 50 51 19
[[49]]
[1] 52 999 56 57 58 21 62 22 22 50 29 4 47 48 59 60 26
[[50]]
[1] 52 10 24
Input ______ Input _____ xreplacenodes(sprof01, c("as.list","vapply", "model.matrix.default"), 999)
                                _ Output _
[[1]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 53 27
[[2]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 22 22 28
[[3]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41
[[4]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41 5 6
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 39
[[6]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 39 14
[[7]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 30
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 30 27
[[10]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 26 5 6 49
[[11]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 27 2 18
[1] 52 10 23 55 56 57 58 21 62 22 22 50 51
[[13]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41 5 6 12
```

[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 47 26 25 3

[[30]]

[[48]]

```
[[31]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 26 5 6 12
[[32]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 31 47 26 25 3
[[33]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 26 15 16
[[34]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41 7 8 9
[[35]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 31 47 26 25 3 45
[[36]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41 5 6 1
[[37]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 53 54
[[38]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 7 8 10
[[39]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 47 26
ΓΓ4011
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 26 5 6
[[41]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 47 26 15 17
[[42]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 47 26 25 44 20
[23] 9 31 34
[[43]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 20
[[44]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 51 32 33
[[45]]
 \begin{bmatrix} 1 \end{bmatrix} \ 52 \ 10 \ 23 \ 55 \ 56 \ 57 \ 58 \ 21 \ 62 \ 22 \ 22 \ 50 \ 29 \ 22 \ 22 \ 35 \ 36 \ 40 \ 41 \ \ 5 \ \ 6 \ 42  
ΓΓ46]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 31 47 26 25 9 31 34
[[47]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 51 43
```

```
[1] 52 10 23 55 56 57 58 21 62 22 22 50 51 19
[[49]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 47 48 59 60 26
[[50]]
[1] 52 10 24
replacenodes(sprof01, c("[","lapply",".getXlevels"),1000)
                             _____ Output _
[[1]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 53 27
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 22 22 28
[[3]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41
[[4]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41 5 6
[[5]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 39
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 39 14
[[7]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38
[[8]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 30
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 30 27
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 26 5 6 49
[[11]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 27 2 18
[[12]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 51
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41 5 6 12
[23] 13
[[14]]
```

```
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 61 15 16
[[15]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 30 46
[[16]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29
[[17]]
[1] 52 10 23 55 56 57 58 21 62 22 22
[[18]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 31 47 48 59
[[19]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41 7 8
[[20]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 61
[[21]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 31 47 26 25
[[22]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40
[[23]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 51 11
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 30 49
[[25]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 61 15 16
[[26]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41 7 8 10
[[27]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 31 47
[[28]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36
[[29]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 30 49 9 31
[[30]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 47 26 25 3
[[31]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 26 5 6 12
```

```
[[32]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 31 47 26 25 3
[[33]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 26 15 16
[[34]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41 7 8 9
[[35]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 31 47 26 25 3 45
[[36]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41 5 6 1
[[37]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 53 54
[[38]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 7 8 10
[[39]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 47 26
[[40]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 26 5 6
Γ[41]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 47 26 15 17
[[42]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 23 53 647 26 25 44 20
[23] 9 31 34
[[43]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 20
[[44]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 51 32 33
[[45]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 22 22 35 36 40 41 5 6 42
[[46]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 37 38 31 47 26 25 9 31 34
ΓΓ4711
[1] 52 10 23 55 56 57 58 21 62 22 22 50 51 43
[[48]]
[1] 52 10 23 55 56 57 58 21 62 22 22 50 51 19
[[49]]
```

```
[1] 52 10 23 55 56 57 58 21 62 22 22 50 29 4 47 48 59 60 26
[[50]]
[1] 52 10 24
```

ToDo: fix null name

We use a prepared sanitised version of our data set.

```
Input
sprof04 <- readRprof("RprofsRegressionExpl03.out", id="sprof04")
```

Applying our old classification:

```
Input -
 nodeclass <- rep("x_nn", length(sprof04$nodes$name))</pre>
 nodeclass[sprof04$nodes$name %in% x_apply] <- "x_apply"</pre>
 nodeclass[sprof04$nodes$name %in% x_as] <- "x_as"
 nodeclass[sprof04$nodes$name %in%
         c("eval", "evalFunc",
                  "try", "tryCatch", "tryCatchList", "tryCatchOne",
                  "doTryCatch", "withVisible")
                  ] <- "x_eval"
 nodeclass[sprof04$nodes$name %in%
         c("model.frame", "model.matrix.default", "model.frame.default",
           " model.response", "model.matrix", "model.response")
                  ] <- "x_model"
 nodeclass[sprof04$nodes$name %in%
         c("lm", "lm.fit", "summary.lm")
                 ] <- "x_lm"
 nodeclass[sprof04$nodes$name =="<Anonymous>"] <- "x_Anon"</pre>
 nodeclass[sprof04$nodes$name == "Sweave"] <- "x_Sweave"</pre>
 nodeclass[sprof04$nodes$name %in% c( "summary" , "summary.lm")] <-</pre>
          "x_summary"
 sprof04$nodes$icol <-as.factor(nodeclass)</pre>
now gives
                                   _ Input _
 #8 12
 oldpar <- par(mfrow=c(3,2))</pre>
```

plot_nodes(sprof04, which=1:6, col=nodeclasscol) par(oldpar)

Sanitised nodes by class, user defined colours: see fig. 10 on the next page.

2.4. Run length. For a visual inspection, runs of the same node and level in the profile are easily perceived. For an analytical inspection, we have to reconstruct the runs from the data. In stacks, runs are organised hierarchically. On the root level, runs are just ordinary runs. On the next levels, runs have to be defined given (within) the previous runs. So we need rrle(), a recursive version of rle, applied to the profile information. This gives a detailed information about the presence time of each node, by stack level.

ToDo: use sprof02 or sprof03?

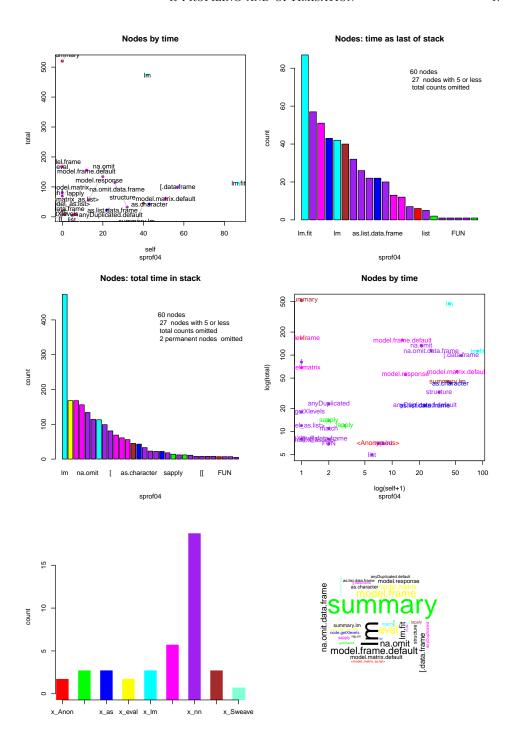


Figure 10. Sanitised. Nodes by class. User defined colours.

```
profile_nodes <- profiles_matrix(sprof02)
profile_nodes_rle<- rrle(profile_nodes, collapseNA=FALSE)
#!NA needs special case in run length handling.</pre>
```

strx(profile_nodes_rle, list.len=5)

```
_{-} Output _{-}
##strx: profile_nodes_rle
List of 12
$ :List of 2
..$ lengths: int [1:365] 1 1 1 3 3 1 7 1 ...
..$ values : int [1:365] NA NA NA 22 39 37 30 4 ...
..- attr(*, "class")= chr "rle"
$ :List of 2
..$ lengths: int [1:411] 1 1 1 3 1 1 1 1 ...
..$ values : int [1:411] NA NA NA 22 NA NA 14 38 ...
..- attr(*, "class")= chr "rle"
$ :List of 2
..$ lengths: int [1:431] 1 1 1 3 1 1 1 1 ...
..$ values : int [1:431] NA NA NA 35 NA NA NA NA ...
..- attr(*, "class")= chr "rle"
$ :List of 2
..$ lengths: int [1:431] 1 1 1 3 1 1 1 1 ...
..$ values : int [1:431] NA NA NA 36 NA NA NA NA ...
..- attr(*, "class")= chr "rle"
$ :List of 2
..$ lengths: int [1:452] 1 1 1 3 1 1 1 1 ...
..$ values : int [1:452] NA NA NA 40 NA NA NA NA ...
..- attr(*, "class")= chr "rle"
[list output truncated]
```

On a given stack level, the run length is the best information on the time used per call, and the run count of a node is the best information on the number of calls. So this is a prime starting point for in-depth analysis.

_ Input _

If you need it, you can represent the run length information by level as a matrix. This is expanding a sparse matrix to full and should be avoided.

```
profile_nodes_rlearray <- nodesprofile(sprof02)
strx(profile_nodes_rlearray)

##strx: profile_nodes_rlearray
num [1:61, 1:12, 1:7] 0 1 0 17 0 0 0 0 ...
- attr(*, "dimnames")=List of 3
..$ node : chr [1:61] "!" "..getNamespace" ".deparseOpts" ...
..$ level : chr [1:12] "1" "2" "3" ...
..$ run_length: chr [1:7] "1" "2" "3" ...</pre>
```

This allows us to extract marginal from provlev[node, level, run length].

ToDo: keep as factor. This is a sparse cube with margins node, stack level, run length. Nodes are mostly concentrated on few levels.

ToDo: Warning: data structure still under discussion

ToDo: hack. replace by decent vector/array based implementation

ToDo: add names for node dimension
ToDo: add summary for NA

ToDo: add marginals and conditionals.

Provide function node_summary.

```
Input .
 nn <- profile_nodes_rlearray["model.frame", , ]</pre>
 print.table(addmargins(nn), zero.print = ".")
                                   _ Output _
     run_length
level 1 2 3 4 5 6 7 Sum
  1
  2
      40 17 7 4 2 6 1 77
  3
  5
  6
  8
  9
  10
  11
  Sum 40 17 7 4 2 6 1 77
                                    _{-} Input _{-}
 amt <- nodesrunlength(sprof02)</pre>
 # print.table(as.table(amt[,1:9]), zero.print = ".")
 ## ??? print.table changes format when col.10 is present.
                                                                                            [ needs
But print.xtable has no zero.print ...
                                                                                   ToDo:
                                                                                   to be hidden for
                                    _ Input _
                                                                                   print.xtable
 #special sanitising for xtable
 xr <- rownames(amt)</pre>
 #for (i in (1:length(xr))) \{xr[i] \leftarrow sub(xr[i],"\setminus[","$[$", fixed=TRUE)]\}
 #xr <- paste("$",xr,"$")</pre>
 xr <- gsub("[","{[}",xr, fixed=TRUE)</pre>
 xr <- gsub("_","\\_",xr, fixed=TRUE)</pre>
 xr <- gsub("^","\\^",xr, fixed=TRUE)</pre>
 rownames(amt)<- xr</pre>
 prxt(amt,
          caption=pasteO("Marginal statistics on nodes by run length, ",
                   "sorted by total time used, count > 1 only"),
         label="tab:pramt4",
         digits=c(rep(0,dim(amt)[2]),2),
         zero.print=" . ") #dim(amt)[2]-1, +1 for rownames
         Table 9: Marginal statistics on nodes by run length, sorted by total
         time used, count > 1 only
```

	1	2	3	4	5	6	7	nr_runs	$total_time$	avg_time
eval	86	34	14	8	4	12	2	160	334	2.09
model.frame	40	17	7	4	2	6	1	77	164	2.13
model.frame.default	40	17	7	4	2	6	1	77	164	2.13
na.omit	46	10	5	4	1	4	1	71	133	1.87
$_{ m lm.fit}$	46	18	3	1	1	1	1	71	113	1.59

na.omit.data.frame											
{ }-data-frame	na.omit.data.frame		7		5				63	113	
model.matrix. default 55 4 2 61 69 1.13 model.matrix. default 55 4 2 61 69 1.13 model.response 35 3 3 1 42 56 1.33 as.character 34 3 1 1 24 33 1.38 lapply 26 - 1 27 30 1.11 getXlevels 17 1 1 20 26 1.30 anyDuplicated 10 2 1 13 23 1.28 vapply 13 1 1 1 18 23 1.28 vapply 13 1 1 1 16 23 1.44 anyDuplicated default 9 2 2 1 16 23 1.44 anyDuplicated default 9 2 2 1 12 22 1.83 as.list.default			4		4						
model.matrix.default 55 4 2 . 61 69 1.13 model.response 35 3 3 . 1 42 56 1.33 as.character 34 3 1 . 24 33 1.33 structure 21 1 1 1 24 33 1.38 lapply 26 . 1 . 27 30 1.11 .getXlevels 17 1 1 1 . 20 26 1.30 anyDuplicated.defaul 10 . 2 1 13 23 1.77 as.list.data.frame 15 1 . 1 16 23 1.44 anyDuplicated.default 9 . 2 1 12 22 1.83 as.list.defame 15 1 . 1 1 4 8 2.00 {[]{{[}{{[}{}}}} 3 .		59	4		4		1				
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$\begin{array}{cccccccccccccccccccccccccccccccccccc$		1							1	1	1.00
terms 1	NCOL	1							1	1	1.00
terms.formula 1	paste	1							1		1.00
		1							1	1	1.00
<u>unlist 1 1 1 1.00</u>	terms.formula	1							1	1	1.00
	unlist	1							1	1	1.00

<u>ToDo:</u> hack. keep See table 9: Marginal statistics on nodes by run length. length in nodesrunlength

We can add this information to our base and use it to enhance graphical displays. See: nodes marked by run length and run count, fig. 15 on page 60.

eval is the base of all evaluation in R, so we should not be surprised to find it at the top of the list. The next two entries, model.frame and model.frame.default are seen 77 times. We know that in our example we had a loop with 100 repetitions, so a frequency of 100 would not be surprising. A closer look shows that both functions occur as isolated events 40 times, but frequency decreases with run length until we see another high at run length 6. We are sampling, and of course sampling can miss some function calls. But this pattern is the oopposite. This is what if occurs if we do not miss a function call, but we miss a gap. So several function calls are joined and appear as some longer runs, typically a multiple of the original run length.

We can improve the hit rate by increasing the sampling rate. But of course this is at the expense of using more space for the log files, and increased time for the overhead. In our case, 1 ms seems to be a good compromise, but 0.1 ms might be another feasible choice.

As we walk down the list, na.omit is next, followed closely by na.omit.data.frame. In our problem, there are no missing data. But R does not have something like a "vanilla mode". The overhead used for the handling of potentially missing values would need a restructuring of the linear model algorithm, e.g. by introducing a "has.na" flag.

Walking further down, we see other candidates such as **as.character** that may be avoidable in this problem.

```
Input

nr <- nodesrunlength(sprof02, clean=FALSE)

#nr <- rbind(nr,0) ## hack. keep length in nodesrunlength

rownames(nr) <- sprof02$nodes$name

sprof02$nodes <- cbind(sprof02$nodes, nr)

rownames(sprof02$nodes) <- sprof02$nodes$name
```

3. Graph Package

What we have achieved so far can be seen from the graph representations. For our purposes, an edge table is most convenient. To allow for edge attibutes, we can use an R data.frame as provided by

We can make use of any graph mapping package. Unfortunately, each seem to have its own calling convention. So we habe to do some translation.

```
Input

library(graph)
search()

Output

[1] ".GlobalEnv" "package:graph"
[3] "package:grid" "package:wordcloud"
[5] "package:RColorBrewer" "package:Rcpp"
[7] "package:xtable" "package:sprof"
```

ToDo: table: node #runs min median run length max

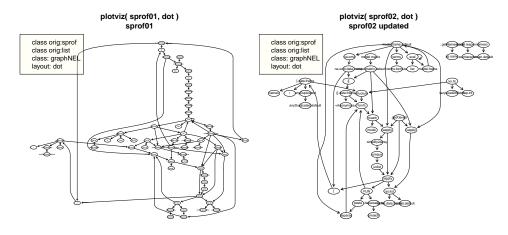


FIGURE 11. Sprof graph, before and after trimming.

See fig. 11 for a comparison before and after trimming. The scaffold effect are removed from the picture on the right side. This cuts off the uninformative spine, and induces minor changes in the body of the graph. You can do additional trimming, if you want.

R is function based, and control structures in general are implemented as functions. In a graph representation, they appear as nodes, concentrating and seeding to unrelated paths. We can detect these on the stack level and replace them by surrogates, introducing new nodes. This is a case for surgery.

ToDo: fix null name We use a prepared sanitised version of our data set.

```
Input

sprof04 <- readRprof("RprofsRegressionExpl03.out", id="sprof04")

#12 6
oldpar <- par(mfrow=c(1,2))
plotviz(sprof02)
plotviz(sprof04)
par(oldpar)
```

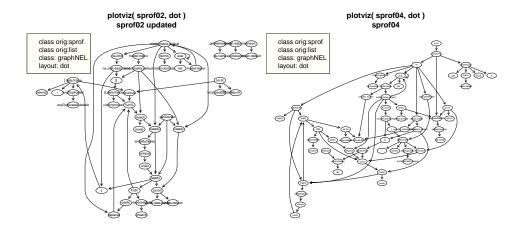


FIGURE 12. Sprof graph, after trimming, and after trimming and some sanitising.

ToDo: cut top levels

Now the structure becomes obvious. Cutting off may be taken two levels deeper. This would completely separate the lm() branch from the summary.lm() In the lm() branch, there are tree nodes (lapply(), %in% and [.data.frame) that are a cases for additional surgery to avoid focusing affects in the graph display. It is your choice to remove them, or live with them.

We know ho to create standard graph displays from this. The next step is to encode additional information we have from the profiles as attribute to the graph.

The derived edge frequency is the first bit of information. Implicitly, it can be used as weight in the graph placement routines. We make this explicit by giving a choice whether to use it or not. Irrespective of this choice, we encode reference counts as line width of the edges.

The functions in this chapter are not included in the package to avoid dependency of **sprof** on **graph** and other graph packages.

```
library(graph)
library(Rgraphviz)
```

This is a common routine for the graph and Rgraphviz package.

```
as_graphNEL_sprof <- function(sprof, weight=TRUE){

a04<-adjacency(sprof)

rnames <- rownames(a04)

if (!weight) {
    dimold <- dim(a04);a04 <- as.numeric(a04); dim(a04) <- dimold
    rownames(a04)<- rnames; colnames(a04)<- rnames;
```

ToDo: Unify the plot functions in this

{

chapter.

```
} #! define lwd first
        e104 <- edgedf(a04)
        el04$lwd <- rkindex(el04$count, maxindex=6, ties.method="min")</pre>
        a04NEL <- as(a04,"graphNEL")</pre>
        nodeDataDefaults(a04NEL, "shape") <- "ellipse"</pre>
        nodeDataDefaults(a04NEL, "cex") <- 0.6</pre>
        nodeDataDefaults(a04NEL, "weigth") <- 1</pre>
        nodeDataDefaults(a04NEL, "fill") <- "green"</pre>
        nodeDataDefaults(a04NEL, "col") <- "yellow"</pre>
        a04NEL <- layoutGraph(a04NEL)</pre>
        nodeRenderInfo(a04NEL) <- list(shape="ellipse")</pre>
        nodeRenderInfo(a04NEL) <- list(cex=0.6, shape="ellipse")</pre>
        nodeRenderInfo(a04NEL) <- list(weight=1)</pre>
        #nodeRenderInfo(a04NEL) <- list(color="yellow")</pre>
        nodeRenderInfo(a04NEL) <- list(fill="yellow", col="blue")</pre>
        edgeDataDefaults(a04NEL,"lwd") <- 1
        edgeDataDefaults(a04NEL,"col") <- "grey"</pre>
        #nodeRenderInfo(a04NEL) <- list(weight=1)</pre>
        #edgeRenderInfo(a04NEL) <- list(lwd=e104$1wd)</pre>
        #edgeRenderInfo(a04NEL)$lwd <- e104$lwd</pre>
        for (i in 1:length(el04$lwd))
        {edgeRenderInfo(a04NEL)$lwd[i] <- e104$lwd[i]}</pre>
        a04NEL
                                      Input
plot_graphNEL_sprof <- function(graphNEL,</pre>
        mode="dot",
        fill_list=NULL,
        main=NULL, sub=NULL,...)
        main1 <- deparse(substitute(graphNEL))</pre>
        xsubid <- NULL
        y<-NULL
        class1 <- NULL</pre>
        if (inherits(graphNEL, "sprof")) {
                 class1 <- paste0("class orig:",class(graphNEL))</pre>
                 main1 <- graphNEL$info$id</pre>
                 graphNEL <-
                          as_graphNEL_sprof(graphNEL, weight=FALSE)
                 # layout(graphNEL,mode)
                 y <- mode
        }
```

nodeDataDefaults(graphNEL, "shape") <- "ellipse"</pre>

```
graphNEL <- layoutGraph(graphNEL)</pre>
        nodeRenderInfo(graphNEL) <- list( #lwd=lwd_list,</pre>
                fill=fill_list,
                col="#0000FF80",
                shape="ellipse",
                weight=1)
        edgeRenderInfo(graphNEL) <- list(col= "#80808080")</pre>
        renderGraph(graphNEL)
        if (!is.null(sub)) sub <- as.character(sub)</pre>
        if (is.null(main))
                main <-paste0("plot_graphNEL_sprof( ",</pre>
                         main1 ,", ",
                         mode, " )\n", xsubid) else
                main=paste0(main, "\n plot_graphNEL_sprof( ",
                        main1 ,", ",
                        mode, " )\n", xsubid)
        title(main=main, sub=sub)
        legend("topleft",
                legend=c( class1, paste0("class: ", class(graphNEL)),
                         paste0("mode: ", mode)),
                bg="#FFFE040",
                seg.len=0
                )#"lightyellow" =#FFFFE0
        legend("bottomright",
                legend= c( #"node color: self.time, pwr=0.5",
                #"node lwd: total.time, pwr=0.5",
                "edge lwd: frequency"),
                bty="n",
                text.font=3,
                bg="#00004040",
                seg.len=0)
}
a04NEL <- as_graphNEL_sprof(sprof04)
```

```
plot_graphNEL_sprof(a04NEL)
# plot_graphNEL_sprof(sprof04)
```

Rgraphviz/graph basic plot: see fig. 13 on the next page.

To use attributes on nodes and edges, we need Rgraphviz.

```
_ Input _
library(graph)
# see help(nodeRenderInfo) for attributes.
plot_graphNEL_sprof1 <- function(sprof04)</pre>
```

ToDo: remove global colour; implement local colour **ToDo:** merge with as_graphNEL_sprof

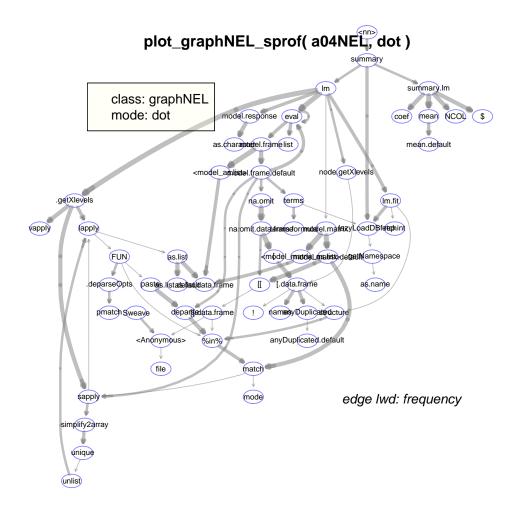


FIGURE 13. Rgraphviz/graph basic plot

```
lwd_list <- rkindex(sprof04$nodes$total.time,</pre>
        pwr=0.5, maxindex=6, ties.method="min")
names(lwd_list) <- sprof04$nodes$name</pre>
#strx(nodeRenderInfo(a04NEL0))
nDD3 <- (nodeRenderInfo(a04NEL0))</pre>
nodeRenderInfo(a04NEL0) <- list(lwd=lwd_list,</pre>
        fill=fill_list,
        col="#0000FF80",
        shape="ellipse",
        weight=1)
#strx(nodeRenderInfo(a04NEL0))
# edge attributes
# strx(edgeRenderInfo(a04NEL0))
nER01 <- edgeRenderInfo(a04NEL0)</pre>
edgeRenderInfo(a04NEL0) <- list(col= "#80808080")</pre>
# strx(edgeRenderInfo(a04NEL0))
renderGraph(a04NEL0)
title(main="nodes (time)", sub=sprof04$info$id, col.sub=grey(0.5))
legend("bottomright",legend= c("node color: self.time, pwr=0.5",
        "node lwd: total.time, pwr=0.5",
        "edge lwd: frequency"),
        bty="n",
        text.font=3,
        bg="#00004040",
        seg.len=0)
legend("topleft",
        legend=c(paste0("class: ",class(a04NEL0)),"dot"),
        bg="#FFFE040",
        seg.len=0)
}
                                 ___ Input _
#6 6
plot_graphNEL_sprof1(sprof04)
```

Rgraphviz/graph plot with attributes: see fig. 14 on the following page. This plot gives us the traditional view, highlighting nodes and edges with overall time presence.

```
detach(package:Rgraphviz)

Input ______
```

We should not overload the plot. We could use colour encoding for the edges, but this would conflict visually with the colour encoding of the nodes. We could use different colours for classes of nodes, but then we would need an additional display to explain the shape information.

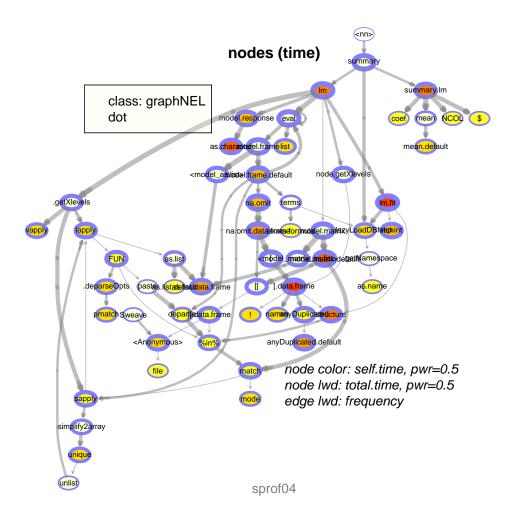


FIGURE 14. Rgraphviz/graph plot with attributes

But within the choice of attributes used, we still can select the information shown. To close this round, instead of showing the node information from the rough summary, we can show the information from the run length discussed in section 2.4 on page 46.

```
#6 6
amt04 <- nodesrunlength(sprof04)
library(graph)
library(Rgraphviz)
# see help(nodeRenderInfo) for attributes.

a04NELOrle <- as_graphNEL_sprof(sprof04, weight=FALSE)
#nDDO <- (nodeRenderInfo(a04NELOrle))
#a04NELOrle <- layoutGraph(a04NELOrle)
#nDD1 <- (nodeRenderInfo(a04NELOrle))# fill & col corrupted
```

```
nodeDataDefaults(a04NELOrle, "shape") <- "ellipse"</pre>
#nodeDataDefaults(a04NELOrle, "cex") <- 1.0</pre>
#nodeDataDefaults(a04NELOrle, "weigth") <- 1</pre>
#nodeDataDefaults(a04NELOrle, "fill") <- "green"</pre>
#nodeDataDefaults(a04NELOrle, "col") <- "yellow"</pre>
nDD2 <- (nodeRenderInfo(a04NELOrle))</pre>
a04NELOrle <- layoutGraph(a04NELOrle)
# node attributes
#sprof04$nodes$self.time -> fill
fill_list <- heat.colors(12)[rkindex(-amt04[,"avg_time"], pwr=0.5, maxindex=12, ties.method="min")]
names(fill_list) <- sprof04$nodes$name</pre>
#sprof04$nodes$total.time -> lwd
lwd_list <- rkindex(amt04[,"nr_runs"], pwr=0.5,maxindex=6, ties.method="min")</pre>
names(lwd_list) <- sprof04$nodes$name
#strx(nodeRenderInfo(a04NELOrle))
nodeRenderInfo(a04NELOrle) <- list(lwd=lwd_list,</pre>
        fill=fill_list,
        col="#0000FF80",
        shape="ellipse",
        weight=1)
#strx(nodeRenderInfo(a04NELOrle))
# edge attributes
# strx(edgeRenderInfo(a04NELOrle))
edgeRenderInfo(a04NELOrle) <- list(col= "#80808080")</pre>
# strx(edgeRenderInfo(a04NELOrle))
renderGraph(a04NELOrle)
title(main="nodes (run length)", sub=sprof04$info$id, col.sub=grey(0.5))
legend("bottomright",
        legend= c("node lwd: nr runs, pwr=0.5",
        "node color: avg time, pwr=0.5",
        "edge lwd: frequency"),
                        bg="#FFFE040",
                seg.len=0,
        bty="n",
        text.font=3)
legend("topleft",
                legend=paste0("class: ",class(a04NELOrle)),
                bg="#FFFFE040",
                seg.len=0
detach(package:Rgraphviz)
```

Nodes marked by run length and run count: see fig. 15 on the following page. This plot gives us a more informative view, highlighting nodes by number of runs and average run length and edges with overall time presence. It puts the information in place, showing us the context where the resources are consumed. Some are consumed with good reason, since here work is done. Others are very doubtful and seem to be mere administration. These are candidates for improvement.

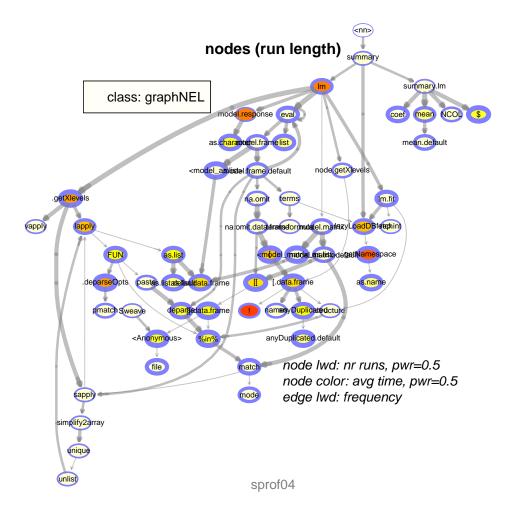


FIGURE 15. nodes (run length)

4. Standard output

For a reference, here are the standard functions.

sprof <- sprof01

4.1. **Print.** We omit the (lengthy) print output here and just give the commands as a reference.

print_nodes(sprof)

Input

print_stacks(sprof)

ToDo: Clarify:"print prints its argument and returns it invisibly (via invisible(x))." Return the argument, or some print representation?

ToDo: is there a print=FALSE variant to postpone printing to e.g. xtable?

	Input
<pre>print_profiles(sprof)</pre>	
The print() method for sprof	objects concatenates these three functions.
4.2. Summary.	
summary_nodes(sprof)	Input
summary_stacks(sprof)	Input
summary_profiles(sprof)	Input

The summary() method for sprof objects concatenates these three functions.

4.3. Plot. Examples are given in the reference manual for sprof.

```
plot_nodes(sprof)

Input

plot_stacks(sprof)

Input

plot_profiles(sprof)
```

The plot.sprof() method for sprof objects concatenates these three functions, see fig. 16. Using the plot functions above allows better control and will be preferred. shownodes() may be a sufficient summary, see fig. 7 on page 25.

```
#12 16
oldpar <- par(mfrow=c(3,4))
plot.sprof(sprof04)
par(oldpar)
```

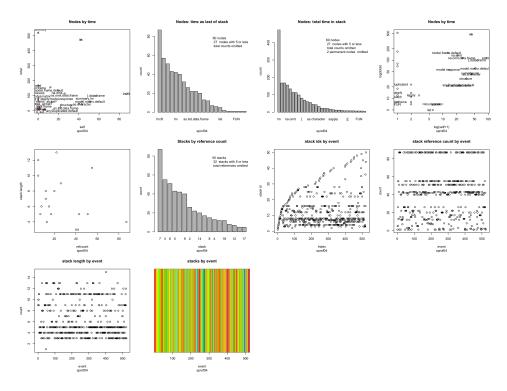


FIGURE 16. plot.sprof(sprof04)

5. More Graphs

Note: This section is collecting experiments with various graph packages. It is only of interest for you, if you have a preference for some graphic package and want to look up a wrapper here. On the other side: contributions and suggestions are welcome.

Graph layout is a theme of its own. Proposals are readily available, as are their implementation. For some of them, there are R interfaces or re-implementations in R. Their usefulness in our context has to be explored, and the answers will vary with personal preferences.

For some graph layout packages we illustrate an interface here and show a sample result. We use the original profile data here. This is a nasty graph with some R stack peculiarities. The corresponding results for the trimmed profile data are shown in the next section 6 on page 89. This is a more realistic example of the kind of graphs you will have to work with.

5.1. **Example: regression.** In this section, we use the recent version of our example, *sprof02* for demonstration. You can re-run it, using your *sprof* data by modifying this instruction by replacing *sprof02* in the next statement with your profile information.

sprof <- sprof02

To interface **sprof** to a graph handling package, **adjacency()** can extract the adjacency matrix from the profile.

There are various packages for finding a graph layout, and the choice is open to your preferences. The R packages for most of these are just wrapper

sprofadj02 <- adjacency(sprof)

This is a format any graph package can handle (maybe). To be on the save side, we provide an (extended) edge list. The added component lwd is a proposal for the line width in the graph rendering.

sprofedgel02 <- edgedf(sprofadj02)

sprofedgel02\$lwd <- rkindex(sprofedgel02\$count,

maxindex=12,

ties.method="min")

ToDo: by graph package: preferred input format?

ToDo: use attributes. Edge width should be easy.

<u>ToDo:</u> include information from stack connectivity.

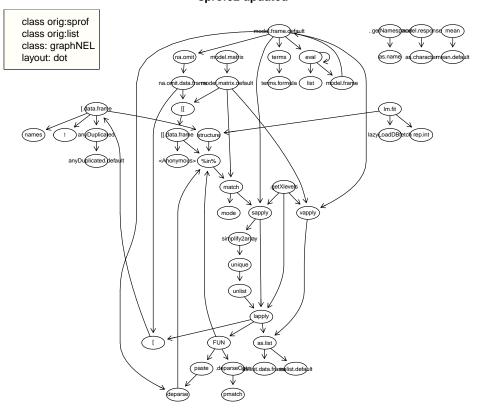
<u>ToDo:</u> add usage of sprofedgel02

5.1.1. graph package. Package 'graph' was removed from the CRAN repository.

This package is now available from Bioconductor only, see http://www.bioconductor.org/packages/release/bioc/html/graph.html.

library(graph)
#search()
plotviz(sprof)
detach("package:graph")

plotviz(sprof, dot) sprof02 updated



```
5.1.2. igraph package. Attributes for igraph are documented in help(igraph.plotting).
Layouts available for igraph:
layout.auto(graph, dim=2, ...)
layout.random(graph, params, dim=2)
layout.circle(graph, params)
layout.sphere(graph, params)
layout.fruchterman.reingold(graph, ..., dim=2, params)
layout.kamada.kawai(graph, ..., dim=2, params)
layout.spring(graph, ..., params)
layout.reingold.tilford(graph, ..., params)
layout.fruchterman.reingold.grid(graph, ..., params)
layout.lgl(graph, ..., params)
layout.graphopt(graph, ..., params=list())
layout.svd(graph, d=shortest.paths(graph), ...)
layout.norm(layout, xmin = NULL, xmax = NULL, ymin = NULL, ymax = NULL,
zmin = NULL, zmax = NULL)
                                  _{-} Input _{-}
library(igraph)
 search()
                                  Output
[1] ".GlobalEnv"
                            "package:igraph"
 [3] "package:grid"
                            "package:wordcloud"
 [5] "package:RColorBrewer" "package:Rcpp"
 [7] "package:xtable"
                            "package:sprof"
[9] "package:stats"
                            "package:graphics"
[11] "package:grDevices"
                            "package:utils"
[13] "package:datasets"
                            "package:methods"
[15] "Autoloads"
                            "package:base"
as_igraph_sprof <- function(sprof, layoutfun, params=NULL,...){
        adj <- adjacency(sprof)</pre>
         adj[adj!=0] <-1
         sprof_igraph <- graph.adjacency(adj)</pre>
         sprof_igraph <- set.graph.attribute(sprof_igraph, "layout",</pre>
                 layoutfun(sprof_igraph,params=params,...),...)
         V(sprof_igraph)$color <- "yellow"</pre>
         E(sprof_igraph)$color <- "#0000FF20"</pre>
         E(sprof_igraph)$width <- c(1,2)</pre>
        return(sprof_igraph)
 }
```

```
sprof_igraph <- graph.adjacency(adjacency(sprof))</pre>
sprof_igraph <- set.graph.attribute(sprof_igraph, "layout", layout.kamada.kawai(sprof_igraph))</pre>
V(sprof_igraph)$color <- "yellow"</pre>
#E(sprof_igraph)$width <- c(1,2)</pre>
E(sprof_igraph)$color <- "#0000FF40"</pre>
E(sprof_igraph)$width <- c(1,2)
# see
                                      Input
plot(sprof_igraph, main=paste0("igraph kamada.kawai layout\n", sprof$info$id))
         legend("topleft",
                  legend= paste0("class: ",class(sprof_igraph)),
                  bg="#FFFE040",
                  seg.len=0
                              igraph kamada.kawai layout
                                    sprof02 updated
           class: igraph
                                     lazyLoadDB
                                                          <An<mark>onym</mark>ous>
                         anyDup(icate)
                                                            terms.formula
```

Experiments with various layouts for igraph follow.

```
#8 8

sprof_ig_auto <- as_igraph_sprof(sprof, layout.auto)

plot(sprof_ig_auto, main=paste0("igraph auto layout\n", sprof$info$id))

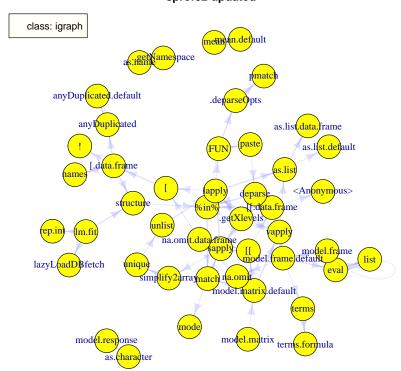
legend("topleft",

legend= paste0("class: ",class(sprof_ig_auto)),

bg="#FFFFE040",

seg.len=0
)
```

igraph auto layout sprof02 updated



```
#6 6

sprof_ig_random <- as_igraph_sprof(sprof, layout.random)

plot(sprof_ig_random, main=paste0("igraph random layout\n", sprof$info$id))

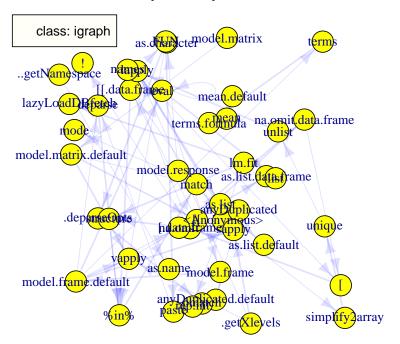
legend("topleft",

legend= paste0("class: ",class(sprof_ig_random)),

bg="#FFFFE040",

seg.len=0
)
```

igraph random layout sprof02 updated



```
#6 6

sprof_ig_circle <- as_igraph_sprof(sprof, layout.circle)

plot(sprof_ig_circle, main=paste0("igraph circle layout\n", sprof$info$id))

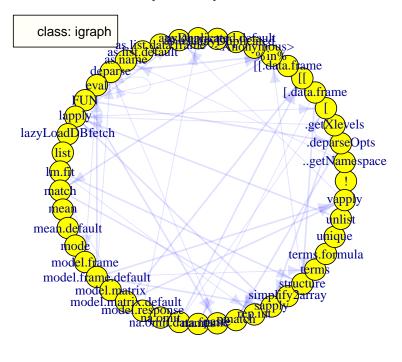
legend("topleft",

legend= paste0("class: ",class(sprof_ig_circle)),

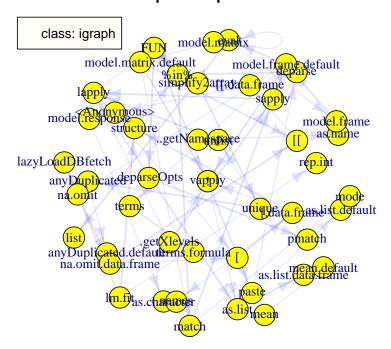
bg="#FFFFE040",

seg.len=0
```

igraph circle layout sprof02 updated



igraph sphere layout sprof02 updated



```
#6 6

sprof_ig_auto <- as_igraph_sprof(sprof, layout.fruchterman.reingold)

plot(sprof_ig_auto, main=paste0("igraph fruchterman.reingold layout\n", sprof$info$id))

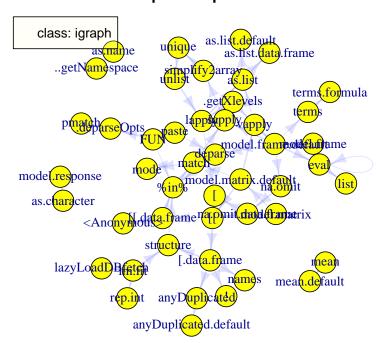
legend("topleft",

legend= paste0("class: ",class(sprof_ig_auto)),

bg="#FFFFE040",

seg.len=0
)
```

igraph fruchterman.reingold layout sprof02 updated



```
#6 6

sprof_ig_auto <- as_igraph_sprof(sprof, layout.kamada.kawai)

plot(sprof_ig_auto, main=paste0("igraph kamada.kawai layout\n", sprof$info$id))

legend("topleft",

legend= paste0("class: ",class(sprof_ig_auto)),

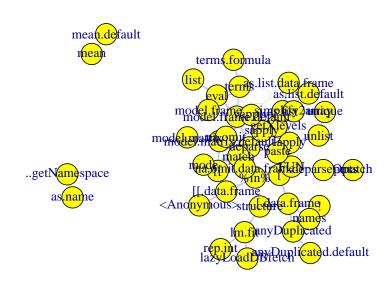
bg="#FFFFE040",

seg.len=0
)
```

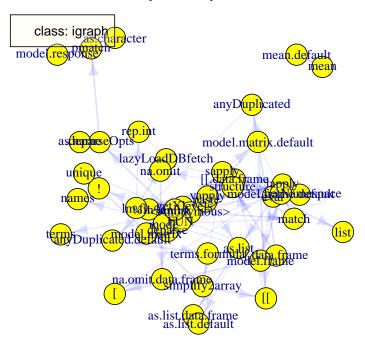
igraph kamada.kawai layout sprof02 updated

class: igraph





igraph spring layout sprof02 updated



```
#6 6

sprof_ig_auto <- as_igraph_sprof(sprof, layout.fruchterman.reingold.grid)

plot(sprof_ig_auto, main=paste0("igraph fruchterman.reingold.grid layout\n", sprof$info$id))

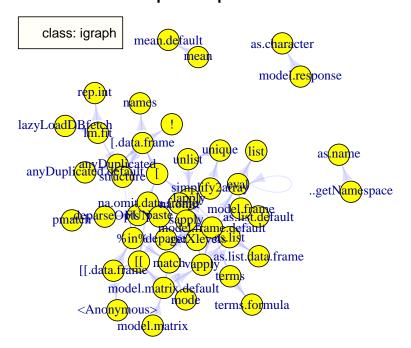
legend("topleft",

legend= paste0("class: ",class(sprof_ig_auto)),

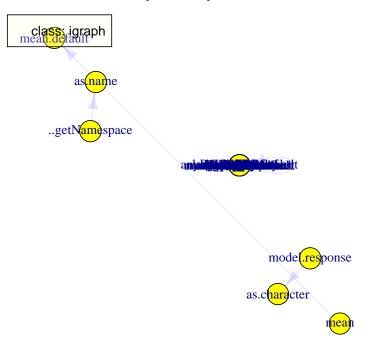
bg="#FFFFE040",

seg.len=0
)
```

igraph fruchterman.reingold.grid layout sprof02 updated



igraph Igl layout sprof02 updated



```
#6 6

sprof_ig_auto <- as_igraph_sprof(sprof, layout.graphopt)

plot(sprof_ig_auto, main=paste0("igraph graphopt layout\n", sprof$info$id))

legend("topleft",

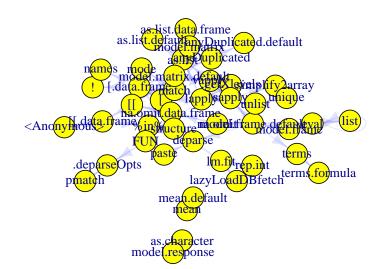
legend= paste0("class: ",class(sprof_ig_auto)),

bg="#FFFFE040",

seg.len=0
)
```

igraph graphopt layout sprof02 updated





```
#6 6

sprof_ig_auto <- as_igraph_sprof(sprof, layout.svd)

plot(sprof_ig_auto, main=paste0("igraph svd layout\n", sprof$info$id))

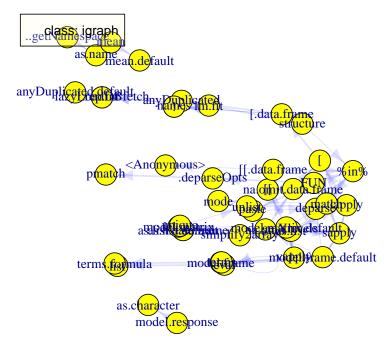
legend("topleft",

legend= paste0("class: ",class(sprof_ig_auto)),

bg="#FFFFE040",

seg.len=0
)
```

igraph svd layout sprof02 updated



ToDo: propagate

5.1.3. network package.

```
detach("package:igraph")
 #detach("package:sna")
 search()
                                   _{-} Output
[1] ".GlobalEnv"
                             "package:grid"
 [3] "package:wordcloud"
                             "package: RColorBrewer"
 [5] "package:Rcpp"
                             "package:xtable"
 [7] "package:sprof"
                             "package:stats"
[9] "package:graphics"
                             "package:grDevices"
[11] "package:utils"
                             "package:datasets"
[13] "package:methods"
                             "Autoloads"
[15] "package:base"
                                    _ Input _
 library(network)
 search()
                                    Output
[1] ".GlobalEnv"
                             "package:network"
 [3] "package:grid"
                             "package:wordcloud"
 [5] "package:RColorBrewer" "package:Rcpp"
 [7] "package:xtable"
                             "package:sprof"
 [9] "package:stats"
                             "package:graphics"
[11] "package:grDevices"
                             "package:utils"
[13] "package:datasets"
                             "package:methods"
[15] "Autoloads"
                             "package:base"
as_network_sprof <- function(sprof) {
         sprofadj02 <- adjacency(sprof)</pre>
         nwsprof02 <- as.network(sprofadj02)</pre>
         network.vertex.names(nwsprof02) <-</pre>
                 rownames(sprofadj02) # not honoured by plot
         return(nwsprof02)
 }
plot_network_sprof <- function( nwsprof,</pre>
         mode = "fruchtermanreingold", main=NULL, label=NULL, sub=NULL,...)
 {
         classnwsprof <- class(nwsprof)</pre>
         xid <- deparse(substitute(nwsprof))</pre>
         xsubid <- NULL
         if (inherits(nwsprof, "sprof")) {
                 xsubid <- nwsprof$info$id</pre>
                 nwsprof <-
```

as_network_sprof(nwsprof)

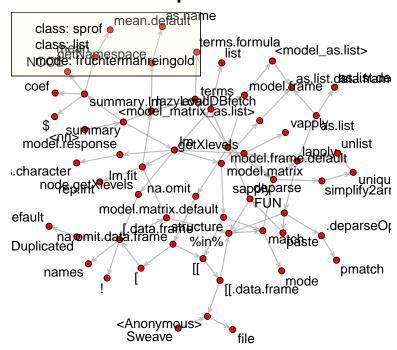
}

```
}
if (!is.null(label))
        warning("explicit label supplied, but will use vertex names")
if (!is.null(sub)) sub <- as.character(sub)</pre>
if (is.null(main))
       paste0("plot_network_sprof( ", xid ,", ", mode, " )\n", xsubid) else
main <-</pre>
                pasteO(main, "\n plot_network_sprof( ", xid ,", ", mode, " )\n", xsubid)
plot( nwsprof,
        label = network.vertex.names(nwsprof),
        main= main,
        mode = mode,
        edge.len=2,
        edge.col="#80808080",
        sub= sub,
        cex.main=1.5,...)
legend("topleft",
        legend=c(paste0("class: ",classnwsprof),paste0("mode: ",mode)),
                                                                                         bg='
        seg.len=0
```

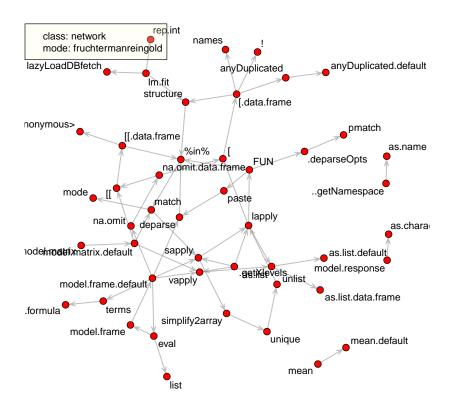
_ Input -

#6 6
plot_network_sprof(sprof04)

plot_network_sprof(sprof04, fruchtermanreingold sprof04



plot_network_sprof(nwsprof02, fruchtermanreingold)

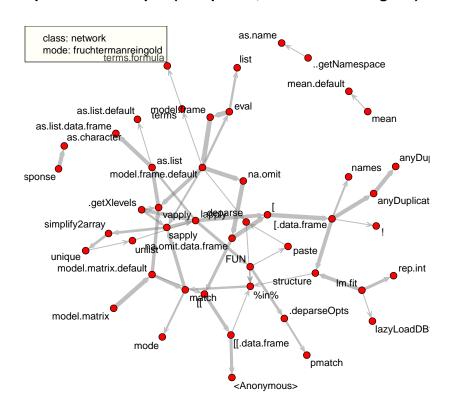


Experiments to include weight.

ToDo: maximum edge.lwd?

```
edge.lwd<- sprofadj02
edge.lwd[edge.lwd>0]<- rkindex(edge.lwd[edge.lwd>0], maxindex=12, ties.method="min")
plot_network_sprof(nwsprof02, label=rownames(sprofadj02),
edge.lwd=edge.lwd)
detach("package:network")
```

plot_network_sprof(nwsprof02, fruchtermanreingold)



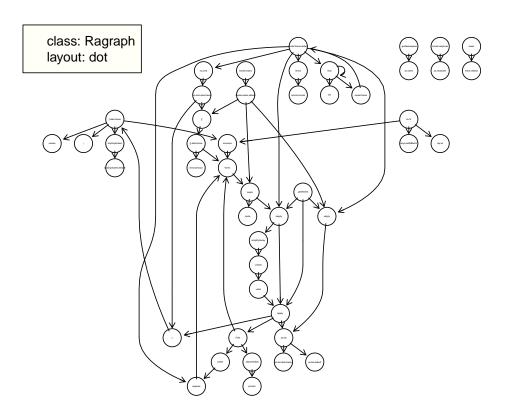
 $5.1.4.\ Rgraphviz$ package 'Rgraphviz' was removed from the CRAN repository.

This package is now available from Bioconductor only, see http://www.bioconductor.org/packages/release/bioc/html/Rgraphviz.html.

```
_ Input _
library(Rgraphviz)
search()
                                   Output
[1] ".GlobalEnv"
                            "package:Rgraphviz"
[3] "package:graph"
                            "package:grid"
[5] "package:wordcloud"
                            "package:RColorBrewer"
[7] "package:Rcpp"
                            "package:xtable"
[9] "package:sprof"
                            "package:stats"
[11] "package:graphics"
                            "package:grDevices"
                            "package:datasets"
[13] "package:utils"
[15] "package:methods"
                            "Autoloads"
[17] "package:base"
sprofadjNEL02 <- as(sprofadj02,"graphNEL")
sprofadjRag02 <- agopen(sprofadjNEL02, name="Rprof Example")</pre>
```

6 6
plotviz(sprofadjRag02,sub=sprof\$info\$id)

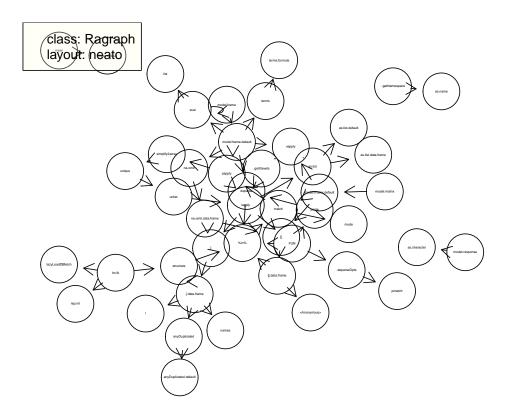
plotviz(sprofadjRag02, dot)



_____ Input _

#6 6

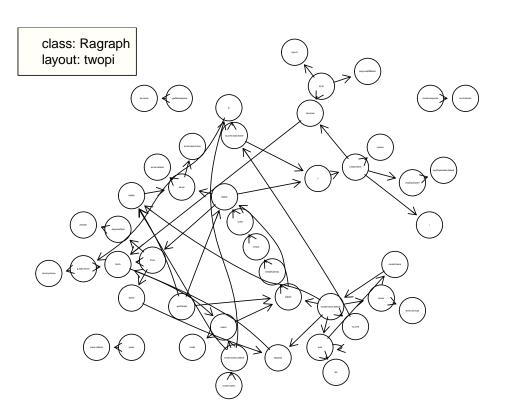
plotviz(sprofadjRag02, neato)



_ Input _

#6 6

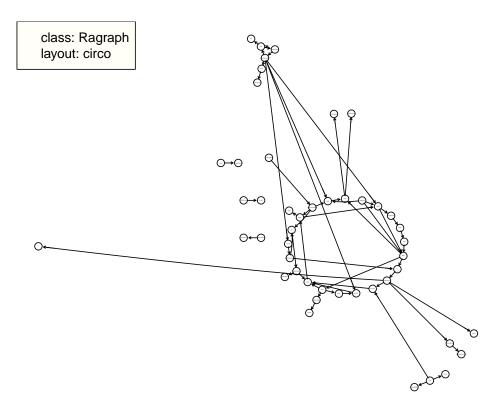
plotviz(sprofadjRag02, twopi)



____ Input _

#6 6

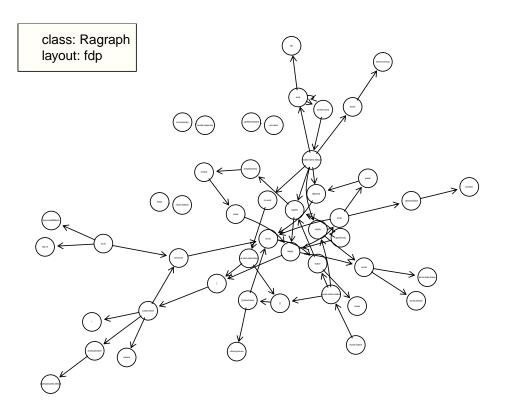
plotviz(sprofadjRag02, circo)



_ Input _

plotviz(sprofadjRag02,"fdp", sub=sprof\$info\$id)

plotviz(sprofadjRag02, fdp)



6. Trimmed example: regression

In this section, we use the reduced version of our example, *sprof04* for demonstration. Except for the change of the data set, this is just a copy of the previous chapter, collecting the various layouts for easy reference.

Note: This section will be replaced by the template section. For internal use only. Skip it.

Some experiments may have found their way to this chapter. They will be expelled.

You can re-run it, using your sprof data by modifying this instruction:

sprof <- sprof04

To interface sprof to a graph handling package, until() can extract the adjacency matrix from the profile.

sprofadj02 <- adjacency(sprof)

This is a format any graph package can handle (maybe).

6.0.5. graph package.

```
library(graph)
search()
```

```
Output

[1] ".GlobalEnv" "package:graph"

[3] "package:grid" "package:wordcloud"

[5] "package:RColorBrewer" "package:Rcpp"

[7] "package:xtable" "package:sprof"

[9] "package:stats" "package:graphics"

[11] "package:grDevices" "package:utils"

[13] "package:datasets" "package:methods"

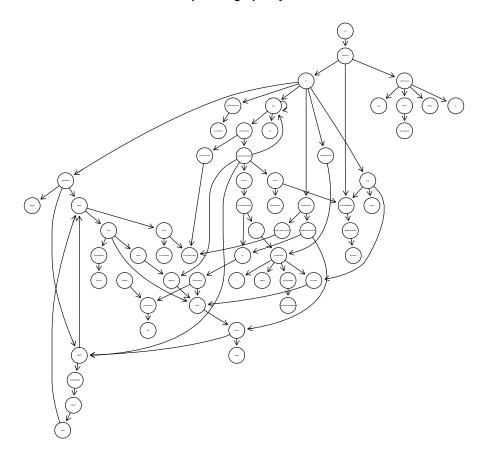
[15] "Autoloads" "package:base"
```

Some tests for scaling \dots

```
sprofadjNEL02 <- as(sprofadj02,"graphNEL")
```

plot(sprofadjNEL02, main="sprof04: graph layout",)
#detach("package:graph")

sprof04: graph layout

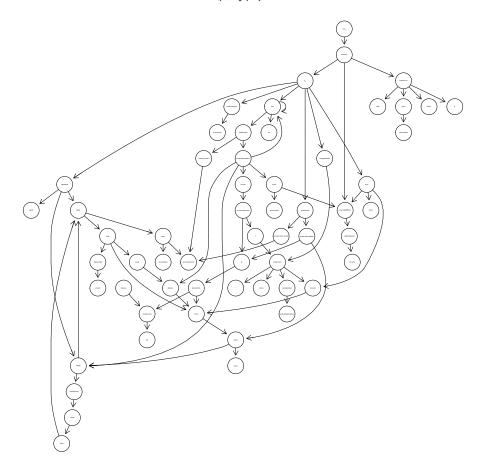


_ Input _

#18

plot(sprofadjNEL02, main="sprof04: graph layout",)
#detach("package:graph")

sprof04: graph layout

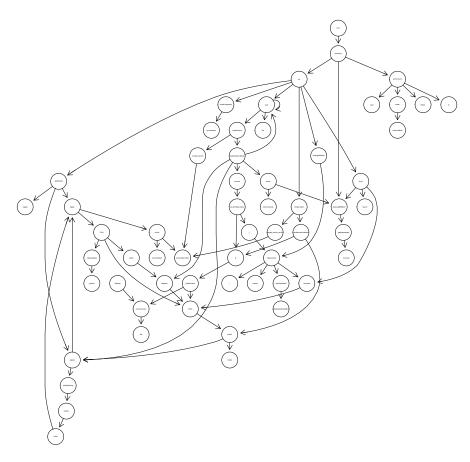


__ Input _____

#12

plot(sprofadjNEL02, main="sprof04: graph layout")
#detach("package:graph")

sprof04: graph layout



6.0.6. igraph package.

```
library(igraph)
search()
```

```
Output

[1] ".GlobalEnv" "package:igraph"

[3] "package:RColorBrewer" "package:Rcpp"

[7] "package:xtable" "package:sprof"

[9] "package:stats" "package:graphics"

[11] "package:grDevices" "package:utils"

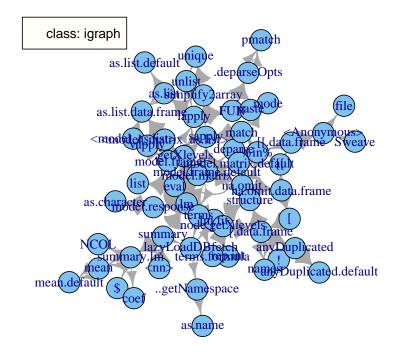
[13] "package:datasets" "package:methods"

[15] "Autoloads" "package:base"
```

```
sprof_igraph <- graph.adjacency(sprofadj02)
```

```
#6
plot(sprof_igraph, main="sprof04: igraph layout: trimmed data")
legend("topleft",
legend=c(paste0("class: ",class(sprof_igraph))),
bg="#FFFFE040",
seg.len=0
)#"lightyellow" =#FFFFE0
detach("package:igraph")
```

sprof04: igraph layout: trimmed data



 $6.0.7.\ network\ package.$

library(network)
search()

Output

[1] ".GlobalEnv" "package:network"

[3] "package:grid" "package:wordcloud"

[5] "package:RColorBrewer" "package:Rcpp"

[7] "package:xtable" "package:sprof"

[9] "package:stats" "package:graphics"

[11] "package:grDevices" "package:utils"

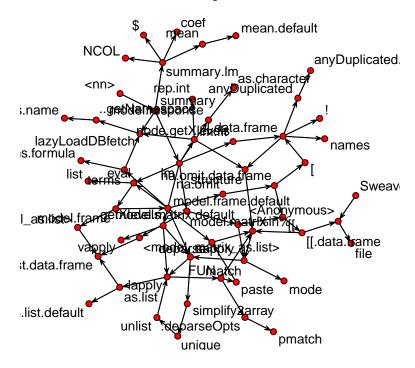
[13] "package:datasets" "package:methods"

[15] "Autoloads" "package:base"

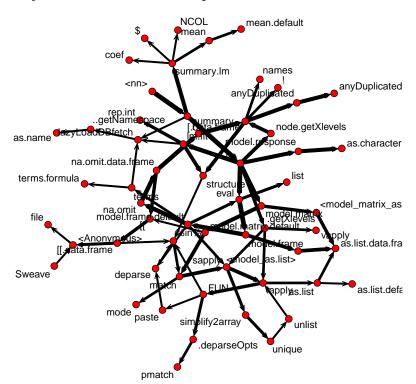
_ Input _____

#6

sprof04: network layout: trimmed dat

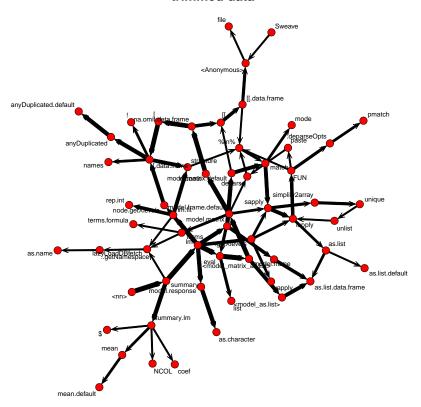


sprof04: network layout: trimmed data



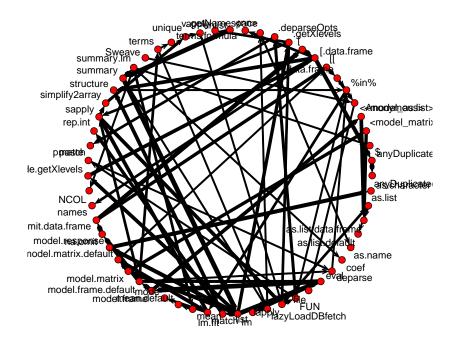
```
#12
plot(nwsprof02, label=rownames(sprofadj02),
main="sprof04: network kamadakawai layout: \n trimmed data",
mode="kamadakawai",
cex.main=2, edge.lwd=edge.lwd)
```

sprof04: network kamadakawai layout: trimmed data

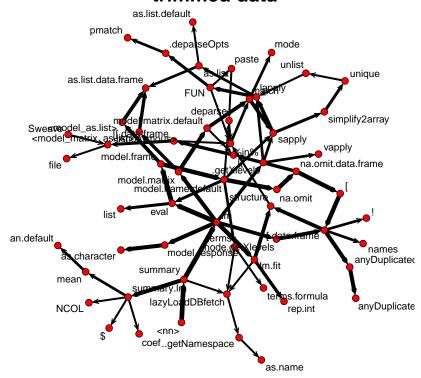


```
plot(nwsprof02, label=rownames(sprofadj02),
    main="sprof04: network circle layout: \n trimmed data",
    mode="circle",
    cex.main=2, edge.lwd=edge.lwd)
```

sprof04: network circle layout: trimmed data



sprof04: network fruchtermanreingold layout: trimmed data



 $6.0.8.\ Rgraphviz\ package.$

library(Rgraphviz)
search()

Output

[1] ".GlobalEnv" "package:Rgraphviz"

[3] "package:graph" "package:grid"

[5] "package:wordcloud" "package:RColorBrewer"

[7] "package:Rcpp" "package:xtable"

[9] "package:sprof" "package:stats"

[11] "package:graphics" "package:grDevices"

[13] "package:utils" "package:datasets"

[15] "package:methods" "Autoloads"

[17] "package:base"

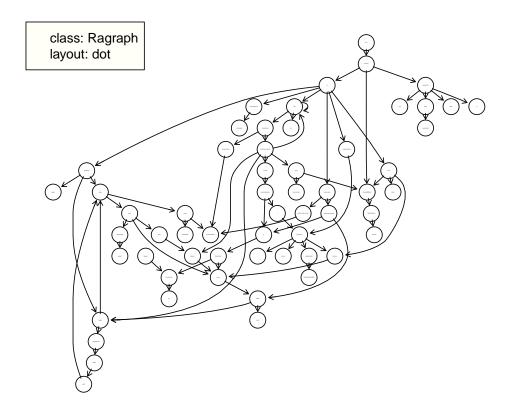
sprofadjRag02 <- agopen(sprofadjNEL02, name="Rprof Example")

__ Input _____

#6 6

plotviz(sprofadjRag02, sub=as.character(sprof\$info\$id))

plotviz(sprofadjRag02, dot)

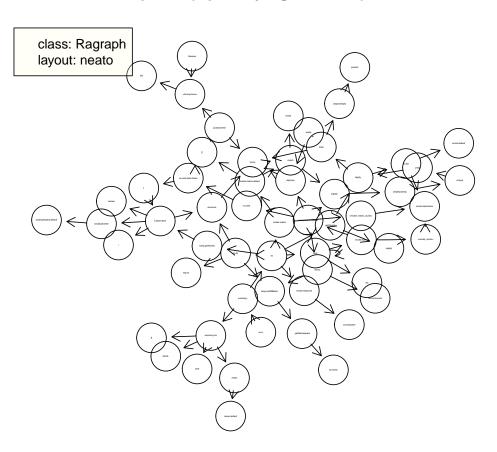


#6

_ Input _

plotviz(x=sprofadjRag02, layout="neato")

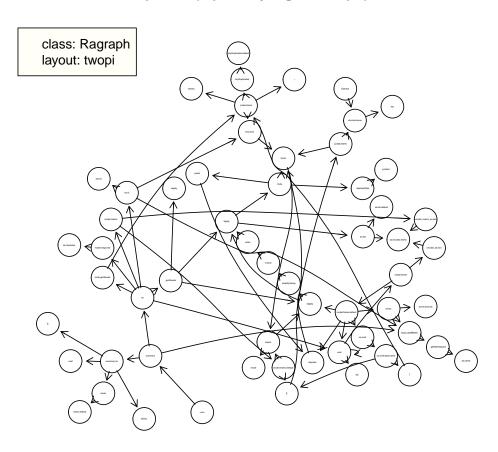
plotviz(sprofadjRag02, neato)



_____ Input _

plotviz(sprofadjRag02, layout="twopi")

plotviz(sprofadjRag02, twopi)

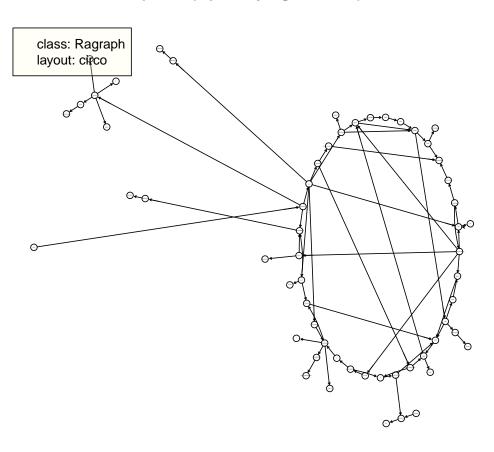


#6

_____ Input _

plotviz(sprofadjRag02, layout="circo")

plotviz(sprofadjRag02, circo)

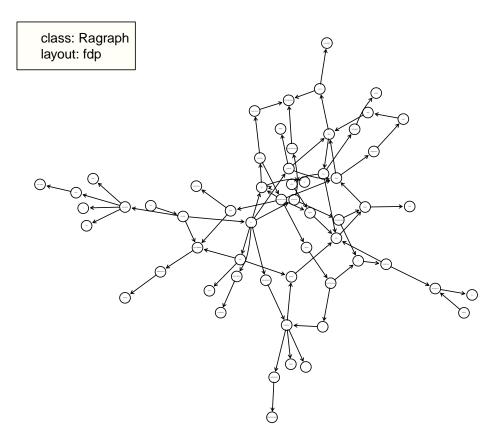


_ Input _

#6

plotviz(sprofadjRag02, layout="fdp")

plotviz(sprofadjRag02, fdp)



7. Template

- Run a profiling routine to profile your functions. You can do it on the fly
- $\bullet\,$ Read in the profile
- Get a survey
- Trim base level and burn-in/fade-out
- \bullet Get a revised survey
- $\bullet\,$ Use a graph display
- \bullet Think!

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R session info:

- R version 3.0.1 (2013-05-16), x86_64-apple-darwin10.8.0
 Locale: en_GB.UTF-8/en_GB.UTF-8/en_GB.UTF-8/C/en_GB.UTF-8/en_GB.UTF-8
 Base packages: base, datasets, graphics, grDevices, grid, methods, stats, utils
 Other packages: graph 1.38.3, RColorBrewer 1.0-5, Rcpp 0.10.3, Rgraphviz 2.4.0, sprof 0.0-6, wordcloud 2.4, xtable 1.7-1
- Loaded via a namespace (and not attached): BiocGenerics 0.6.0, igraph 0.6.5-2, network 1.7.2, parallel 3.0.1, slam 0.1-28, sna 2.3-1, stats4 3.0.1, tools 3.0.1

LATEX information: textwidth: 4.9823in linewidth:4.9823in

textheight: 8.0824in

Svn repository information:

\$Source: /u/math/j40/cvsroot/lectures/src/insider/profile/Rnw/profile.Rnw,v \$

\$Id: sprofiling.Rnw 215 2013-08-21 18:52:29Z gsawitzki \$

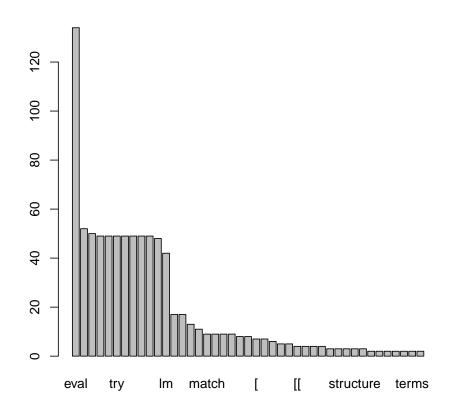
\$Revision: 215 \$

\$Date: 2013-08-21 20:52:29 0200(Wed,21Aug2013)+

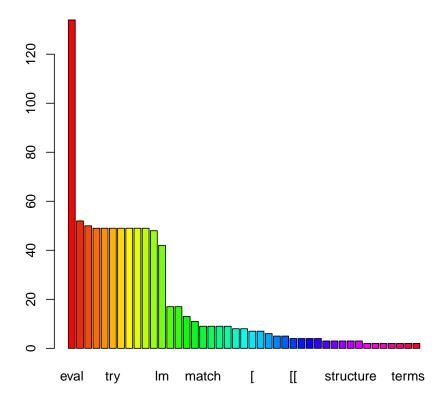
\$name: \$

\$Author: gsawitzki \$

8. XXX - LOST & FOUND

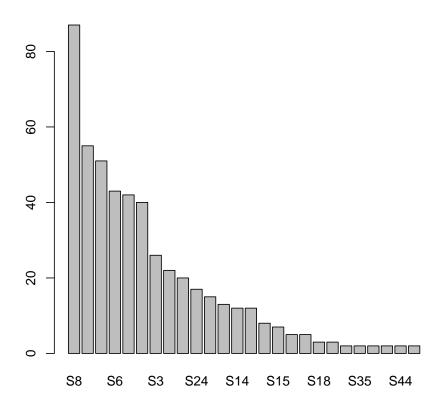


barplot(ndf[ondf], col=rainbow(length(ondf)))



Top frequent stacks.

```
x <- sprof01
xsrc <- as.matrix(x$stacks$refcount)
rownames(xsrc) <- rownames(xsrc, do.NULL=FALSE, prefix="S")
#stf <- x$stacks$refcount[x$stacks$refcount>1]
#names(stf) <- x$stacks$shortname[x$stacks$refcount>1]
stf <- xsrc[xsrc>1]
names(stf) <- rownames(xsrc)[xsrc>1]
ostf <- order(stf,decreasing=TRUE)
barplot(stf[ostf])</pre>
```



xtable cannot handle posix, so we use print output here
str(profile_nodes_rle, max.level=2, vec.len=3, nchar.max=40, list.len=6)
strx(sprof01\$info)

```
#stry: gprof01$info
```

##strx: sprof01\$info

'data.frame': 1 obs. of 9 variables:

\$ id : chr "sprof01"

\$ date : POSIXct, format: "2013-08-22 21:01:57"

\$ nrnodes : int 62
\$ nrstacks : int 50
\$ nrrecords : int 522
\$ sample.interval: num 0.001
\$ sampling.time : num 0.522

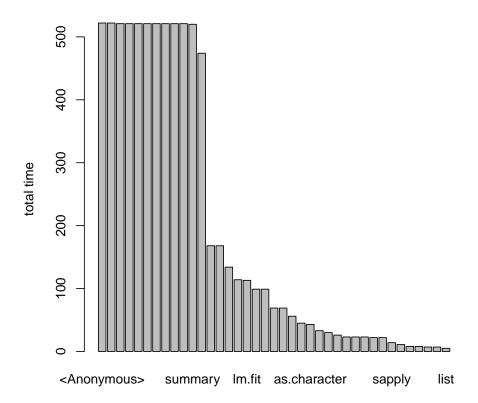
\$ ctllines : chr "sample.interval=1000"

\$ ctllinenr : num 1

Selections are recorded as selection vectors, with reference to the original order. This needs some caution to align them with the order choices.

```
rownames(sprof01$nodes) <- sprof01$nodes$names
nodesperm <- order(sprof01$nodes$total.time,decreasing=TRUE)
nodesnrobsok <- sprof01$nodes$total.time > 4
sp <- sprof01$nodes$total.time[nodesperm][nodesnrobsok[nodesperm]]
names(sp) <- sprof01$nodes$name[nodesperm][nodesnrobsok[nodesperm]]
barplot(sp,
main="Nodes, by total time", ylab="total time")
```

Nodes, by total time



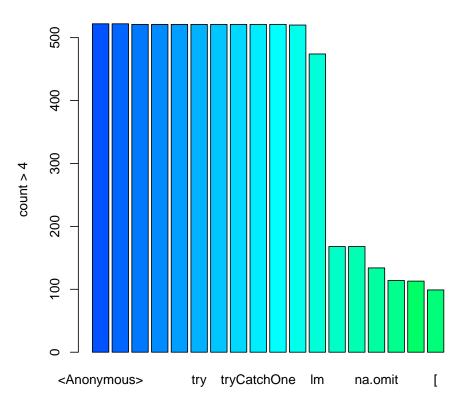
On the first look, information on the profile level is not informative. Profile records are just recordings of some step, taken at regular intervals. We get a minimal information, if we encode the stacks in colour.

We now do a step down analysis. Aggregating the information from the profiling events, we have the frequency of stack references. On the stack level, we encode the frequency in colour, and linking propagates this to the profile level.

<u>ToDo:</u> use stack colours

ToDo: use as_rkindex

Stacks, by reference count (4 obs. minimum)



sprof01

```
prxt(sprof01$nodes,
caption="nodes",
label="tab:prSREnodes",
max.level=2, vec.len=3,nchar.max=40,
digits=c(0,0,0,2,0,2,0))
```

Table 10: nodes

-	name	self.time	self.pct	total.time	total.pct	icol
1	!	2	0.38	2	0.03	50
2	get Name space	0	0.00	1	0.01	55
3	. departs = Opts	2	0.38	4	0.05	41
4	.get X levels	0	0.00	26	0.34	28
5	[0	0.00	99	1.29	20
6	[.data.frame	57	10.92	99	1.29	19
7	[[0	0.00	8	0.10	36
8	[[.data.frame	1	0.19	8	0.10	35
9	$\% \mathrm{in}\%$	1	0.19	4	0.05	40
10	<anonymous></anonymous>	6	1.15	522	6.79	2
<cut></cut>	$\setminus vdots$:	÷	:	:	:
53	terms	0	0.00	2	0.03	45
54	terms.formula	1	0.19	1	0.01	54
55	try	0	0.00	521	6.78	7
56	tryCatch	0	0.00	521	6.78	4
57	tryCatchList	0	0.00	521	6.78	6
58	tryCatchOne	0	0.00	521	6.78	8
59	unique	3	0.57	4	0.05	43
60	unlist	0	0.00	1	0.01	56
61	vapply	3	0.57	23	0.30	30
62	withVisible	0	0.00	521	6.78	10

```
#str(sprof01$stacks, max.level=2, vec.len=6,
 # nchar.max=40, list.len=20,width=70, strict.width="wrap"
 strx(sprof01$stacks)
                              _____ Output _
##strx: sprof01$stacks
'data.frame': 50 obs. of 7 variables:
$ nodes :List of 50
..$ : int 52 10 23 55 56 57 58 21 ...
..$ : int 52 10 23 55 56 57 58 21 ...
..$ : int 52 10 23 55 56 57 58 21 ...
..$ : int 52 10 23 55 56 57 58 21 ...
..$ : int 52 10 23 55 56 57 58 21 ...
..$ : int 52 10 23 55 56 57 58 21 ...
..$ : int 52 10 23 55 56 57 58 21 ...
..$ : int 52 10 23 55 56 57 58 21 ...
..$ : int 52 10 23 55 56 57 58 21 ...
..$ : int 52 10 23 55 56 57 58 21 ...
..$ : int 52 10 23 55 56 57 58 21 ...
..$ : int 52 10 23 55 56 57 58 21 ...
.. [list output truncated]
$ shortname : Factor w/ 50 levels
   "S<A>eFttCtCLtCOdTCwVeesleem.m..n.n...[["| __truncated__,..: 27 17
   19 1 35 36 37 30 ...
$ refcount : num 1 5 26 55 13 43 51 87 ...
$ stacklength : int 19 20 19 21 14 15 15 14 ...
$ stackheadnodes: int 52 52 52 52 52 52 52 ...
$ stackleafnodes: int 27 28 41 6 39 14 38 30 ...
$ stackssrc : Factor w/ 50 levels "! [.data.frame [
  na.omit.data.frame na." | __truncated__,..: 27 28 39 5 37 13 36 30
A summary is provided on request.
                                    Input
 sumsprof01 <- summary.sprof(sprof01)</pre>
                                 Output -
$id
[1] "Profile Summary Thu Aug 22 21:02:34 2013"
$len
[1] 522
$uniquestacks
[1] 50
$nr_runs
[1] 396
$nrstacks
Γ17 50
```

\$stacklength

[1] 3 25

\$nrnodesperlevel

[1] 1 1 2 1 1 1 1 1 1 1 1 3 10 11 9 9 15 8 7 5 7 [23] 2 1 1

shortname root leaf self.time self.pct ! - LEAF 2 0.383142 0 0.000000 ..gN ..getNamespace - LEAF 2 0.383142 .deparseOpts .dpO 0 0.000000 - -.getXlevels .gtX 0 0.000000 [57 10.919540 [.data.frame ſ.d. - LEAF]] 11 0 0.000000 - LEAF [[.data.frame [[.. 1 0.191571 %in% %in% - LEAF 1 0.191571 6 1.149425 <Anonymous> <An>- LEAF 1 0.191571 - LEAF \$ - LEAF 1 0.191571 anyDuplicated anyD22 4.214559 - LEAF anyDuplicated.default anD. - LEAF 43 8.237548 as.character as.c as.list - -0.000000 as.l - LEAF 22 4.214559 as.list.data.frame a... - LEAF 1 0.191571 as.list.default as.. as.n - LEAF 1 0.191571 as.name coef coef - LEAF 1 0.191571 deparse dprs - LEAF 1 0.191571 dTrC - -0 0.000000 doTryCatch - LEAF eval eval 1 0.191571 0 0.000000 evlF - evalFunc 1 0.191571 file - LEAF file 1 0.191571 FUN FUN - LEAF - LEAF 2 0.383142 lapply lppl lazyLoadDBfetch - LEAF 2 0.383142 1LDB list - LEAF 5 0.957854 list 42 8.045977 lm lm - LEAF lm.fit lm.f - LEAF 87 16.666667 1 0.191571 match mtch- LEAF 0 0.000000 mean mean - -- LEAF mean.default 2 0.383142 mn.d- LEAF 2 0.383142 mode mode 0 0.000000 - model.frame mdl.f model.frame.default mdl.f. - LEAF 12 2.298851 model.matrix mdl.m - -0 0.000000 - LEAF 51 9.770115 model.matrix.default mdl.m. mdl.r - LEAF 13 2.490421 model.response n.mt - LEAF 20 3.831418 na.omit na.omit.data.frame n... - LEAF 26 4.980843 nams - LEAF 2 0.383142 names NCOL - LEAF NCOL 1 0.191571 0 0.000000 paste past - LEAF 2 0.383142 pmatch pmtc

			-	
rep.int	rp.n	- LEAF	7	1.340996
sapply	sppl		1	0.191571
simplify2array	smp2		0	0.000000
structure	strc	- LEAF	32	
summary	smmr		0	
summary.lm	smm.	- LEAF	40	7.662835
Sweave	Swev I	ROOT -	0	0.000000
terms	trms		0	0.000000
terms.formula	trm.	- LEAF	1	0.191571
try	try		0	0.000000
tryCatch	tryC		0	0.000000
tryCatchList	trCL		0	0.000000
tryCatchOne	trCO		0	0.000000
unique	uniq	- LEAF	3	0.574713
unlist	unls		0	0.000000
vapply	vppl	- LEAF	3	0.574713
withVisible	wthV		0	0.000000
	total.time	total.pct		
!	2			
getNamespace	1	0.191571		
.deparseOpts		0.766284		
.getXlevels	26			
[99			
[.data.frame	99			
[[8	1.532567		
[[.data.frame	8	1.532567		
	4			
%in%	_	0.766284		
<anonymous></anonymous>		100.000000		
\$	1			
anyDuplicated	23	4.406130		
anyDuplicated.default		4.214559		
as.character	43			
as.list	23			
as.list.data.frame	22			
as.list.default	1	0.191571		
as.name	1	0.191571		
coef	1	0.191571		
deparse	2	0.383142		
doTryCatch	521	99.808429		
eval	521	99.808429		
evalFunc	521	99.808429		
file	1	0.191571		
FUN	7	1.340996		
lapply	30	5.747126		
lazyLoadDBfetch	3	0.574713		
list	5	0.957854		
lm	474	90.804598		
lm.fit	113	21.647510		
match	11			
mean	2			
mean.default	2			
mode	2			
model.frame	168			
	100	22.230000		

```
model.frame.default
                        168 32.183908
model.matrix
                         69 13.218391
                         69 13.218391
model.matrix.default
                         56 10.727969
model.response
                        134 25.670498
na.omit
na.omit.data.frame
                        114 21.839080
names
                          2 0.383142
NCOL
                          1 0.191571
                          1 0.191571
paste
                          2 0.383142
pmatch
                          7 1.340996
rep.int
                         14 2.681992
sapply
                          4 0.766284
simplify2array
structure
                          33
                              6.321839
summary
                        520 99.616858
summary.lm
                          45
                              8.620690
Sweave
                         522 100.000000
                         2 0.383142
terms
                          1 0.191571
terms.formula
                        521 99.808429
try
                        521 99.808429
tryCatch
                        521 99.808429
tryCatchList
tryCatchOne
                        521 99.808429
unique
                         4 0.766284
unlist
                          1 0.191571
                         23 4.406130
vapply
withVisible
                        521 99.808429
```

#str(profile_nodes_rle, max.level=2, vec.len=3, nchar.max=40, list.len=6)
strx(sumsprof01)

```
##strx: sumsprof01
'data.frame': 62 obs. of 7 variables:
$ shortname : Factor w/ 62 levels "!","..gN",".dp0",..: 1 2 3 4 5 6 7
8 ...
$ root : Factor w/ 2 levels "-","R00T": 1 1 1 1 1 1 1 1 1 ...
$ leaf : Factor w/ 2 levels "-","LEAF": 2 1 2 1 1 2 1 2 ...
$ self.time : num 2 0 2 0 0 57 0 1 ...
$ self.pct : num 0.383 0 0.383 0 ...
$ total.time: num 2 1 4 26 99 99 8 8 ...
$ total.pct : num 0.383 0.192 0.766 4.981 ...
```

```
#str(sumsprof01, max.level=2, vec.len=3,
# nchar.max=40, list.len=6,
# width=70, strict.width="wrap")
```

The classical approach hides the work that has been done. Actually it breaks down the data to record items. This figure is not reported anywhere. In our case, it can be reconstructed. The profile data have 8456 words in 524 lines.

In our approach, we break down the information. Two lines of control information are split off. We have 522 lines of profile with 50 unique stacks, referencing 62 nodes. Instead of reducing it to a summary, we keep the full information. Information is always kept on its original level.

On the profiles level, we know the sample interval length, and the id of the stack recorded. On the stack level, for each stack we have a reference count, with the sample interval lengths used as weights. This reference count is added up for each node in the stack to give the node timings.

Cheap statistics are collected as the come by. For example, from the stacks table it is cheap to identify root and leaf nodes, and this mark is propagated to the nodes table. These are some attempts to recover the factor structures.

```
Input -
xfi <- levels(sprof02$nodes$name)
 profile_nodes_rlefac <- lapply(profile_nodes_rle,</pre>
         function(xl) {xl$values <- factor(xl$values,</pre>
                levels=1:62.
                labels=xfi); xl}) # seems ok
 profile_nodes_rletfac <- lapply(profile_nodes_rle,</pre>
         function(x) table(x,dnn=c("run length","node")) ) #factors lost again
         colnames(profile_nodes_rletfac[[1]]) <-</pre>
                sprof02$nodes$name[ as.integer(colnames(profile_nodes_rletfac[[1]]))]
 profile_nodes_rletfac1 <- lapply(profile_nodes_rletfac,</pre>
         function(x1) {colnames(x1) <-</pre>
                sprof02$nodes$name[ as.integer(colnames(x1))];
        x1} )
 invisible(lapply(profile_nodes_rletfac1,
 function(x) print.table(t(x),zero.print = ".") ))
                               - Output -
     run length
      1 2 3 4 5 6 7
node
  <NA> 17 1 1 1 .
  <NA> 1 . . . .
  <NA> 1
  <NA> 40 17 7 4 2 6 1
  <NA> 46 18 3 1 1 1 1
  <NA> 1 .
             2 . .
  <NA> 55 4
  <NA> 35 3 3 . . 1
  <NA> 1
                     run length
                      1 2 3 4 5 6 7
node
  as.character
                      34
                          3 1
  as.name
                      1
                     40 17 7 4 2 6 1
  eval
                      16 .
  lapply
                               1
  lazyLoadDBfetch
                     1
                     1
  mean.default
  model.matrix.default 55 4 2
```

```
rep.int
                  7 . . . . . .
 sapply
                  3 .
                 10 1 . . . 1 .
 structure
 vapply
                 . . 1
           run length
node
            1 2 3 4 5 6 7
 [
            14 . . 1 . .
 ]]
             1 . . . .
            1 . . . . .
 %in%
 as.list
             2 . . . . .
             2 . . . .
 lapply
 match
             9.
 model.frame 40 17 7 4 2 6 1
 simplify2array 1 . . . . .
 vapply 6 1 . .
                run length
                 1 2 3 4 5 6 7
node
                      . 1
 [.data.frame
                14
 [[.data.frame
                1
                7 1 .
 as.list
 as.list.data.frame 2
 FUN
                 1
 match
                 1
 model.frame.default 40 17 7 4 2 6 1
         9 . . . .
 sapply
                 1 . .
 unique
               run length
node
                1 2 3 4 5 6 7
 .deparseOpts
                1 . . . . .
 <Anonymous>
                1 . . . . .
 anyDuplicated
                1 . . . .
 as.list.data.frame 6 1 . .
 as.list.default 1 . . .
 deparse
                1 . . .
 eval
                3
 lapply
                5.
               46 10 5 4 1
 na.omit
 sapply
                2 .
 simplify2array
                3
                11 .
                     . 1
 structure
 terms
                1 .
 unlist
                1
                7 . . .
 vapply
               run length
                1 2 3 4 5 6
node
 as.list
 eval
 FUN
                5 . . .
                3 .
 na.omit.data.frame 43 7 4 5 . 4
 terms.formula 1 . . . . .
                3 . . . . .
 unique
               run length
```

```
node
                    1 2 3 4 5 6
                   3 .
  .deparseOpts
                   45 4 3 3 .
  [
 ]]
                   2 . .
                            . 1
 %in%
                   1 .
 as.list.data.frame 7 . . . 1
 FUN
                    1 . . . .
                    3 .
 list
            run length
           1 2 3 4 5 6
node
 [.data.frame 45 4 3 3 . 1
 [[.data.frame 2 . . . 1 .
 \mbox{match} \qquad \qquad 1 \ \ . \ \ . \ \ . \ \ .
 paste
               1 . . . . .
              2 . .
 pmatch
            run length
node
              1 4 5
 %in%
              1 . .
 <Anonymous> . . 1
 anyDuplicated 9 2 1
 deparse 1 . .
 mode
              1 . .
             2 . .
 names
                    run length
node
                     1 4 5
 %in%
                      1 . .
 anyDuplicated.default 9 2 1
    run length
node
     1
 match 1
     run length
node 1
 mode 1
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