

Unsupervised Learning Mini-Project

Core functions:

```
read.csv("YourFileName")
```

```
prcomp(x, scale = TRUE)
```

```
kmeans(x, centers = ?)
```

```
hclust(dist(x))
```

```
# Make Available Data for Project
wisc.df <- read.csv("WisconsinCancer.csv", row.names = 1)
# Separate data and physician diagnosis
wisc.data <- (wisc.df[, -1])
diagnosis <- (wisc.df[, 1])
```

Exploratory Data Analysis.

#Q1. There are 569 observations in this dataset.

```
#Check how many observations have a malignant diagnosis.
x = 0
for(i in 1:length(diagnosis)) {
  if(diagnosis[i] == "M") {x <- x+1}
}
x
```

```
## [1] 212
```

```
#also 'table(diagnosis)'
#also 'sum(diagnosis == "M")'
```

#Q2. There are 212 observations with a malignant diagnosis.

```
# Check how many variables end with "_mean".
length(grep(pattern = "*_mean", x = colnames(wisc.data)))
```

```
## [1] 10
```

#Q3. There are 10 variables suffixed with “_mean”.

#Principal Component Analysis.

```
# Check column means and standard deviations (1 = rows, 2 = cols)
apply(wisc.data, 2, mean)
```

##	radius_mean	texture_mean	perimeter_mean
##	1.412729e+01	1.928965e+01	9.196903e+01
##	area_mean	smoothness_mean	compactness_mean
##	6.548891e+02	9.636028e-02	1.043410e-01
##	concavity_mean	concave.points_mean	symmetry_mean
##	8.879932e-02	4.891915e-02	1.811619e-01
##	fractal_dimension_mean	radius_se	texture_se
##	6.279761e-02	4.051721e-01	1.216853e+00
##	perimeter_se	area_se	smoothness_se
##	2.866059e+00	4.033708e+01	7.040979e-03
##	compactness_se	concavity_se	concave.points_se
##	2.547814e-02	3.189372e-02	1.179614e-02
##	symmetry_se	fractal_dimension_se	radius_worst
##	2.054230e-02	3.794904e-03	1.626919e+01
##	texture_worst	perimeter_worst	area_worst
##	2.567722e+01	1.072612e+02	8.805831e+02
##	smoothness_worst	compactness_worst	concavity_worst
##	1.323686e-01	2.542650e-01	2.721885e-01
##	concave.points_worst	symmetry_worst	fractal_dimension_worst
##	1.146062e-01	2.900756e-01	8.394582e-02

```
apply(wisc.data, 2, sd)
```

##	radius_mean	texture_mean	perimeter_mean
##	3.524049e+00	4.301036e+00	2.429898e+01
##	area_mean	smoothness_mean	compactness_mean
##	3.519141e+02	1.406413e-02	5.281276e-02
##	concavity_mean	concave.points_mean	symmetry_mean
##	7.971981e-02	3.880284e-02	2.741428e-02
##	fractal_dimension_mean	radius_se	texture_se
##	7.060363e-03	2.773127e-01	5.516484e-01
##	perimeter_se	area_se	smoothness_se
##	2.021855e+00	4.549101e+01	3.002518e-03
##	compactness_se	concavity_se	concave.points_se
##	1.790818e-02	3.018606e-02	6.170285e-03
##	symmetry_se	fractal_dimension_se	radius_worst
##	8.266372e-03	2.646071e-03	4.833242e+00
##	texture_worst	perimeter_worst	area_worst
##	6.146258e+00	3.360254e+01	5.693570e+02
##	smoothness_worst	compactness_worst	concavity_worst
##	2.283243e-02	1.573365e-01	2.086243e-01
##	concave.points_worst	symmetry_worst	fractal_dimension_worst
##	6.573234e-02	6.186747e-02	1.806127e-02

```
# Perform PCA on wisc.data by completing the following code
wisc.pr <- prcomp(wisc.data, scale=TRUE)
summary(wisc.pr)
```

```
## Importance of components:
##
```

	PC1	PC2	PC3	PC4	PC5	PC6	PC7
## Standard deviation	3.6444	2.3857	1.67867	1.40735	1.28403	1.09880	0.82172
## Proportion of Variance	0.4427	0.1897	0.09393	0.06602	0.05496	0.04025	0.02251
## Cumulative Proportion	0.4427	0.6324	0.72636	0.79239	0.84734	0.88759	0.91010

```
##
```

	PC8	PC9	PC10	PC11	PC12	PC13	PC14
## Standard deviation	0.69037	0.6457	0.59219	0.5421	0.51104	0.49128	0.39624
## Proportion of Variance	0.01589	0.0139	0.01169	0.0098	0.00871	0.00805	0.00523
## Cumulative Proportion	0.92598	0.9399	0.95157	0.9614	0.97007	0.97812	0.98335

```
##
```

	PC15	PC16	PC17	PC18	PC19	PC20	PC21
## Standard deviation	0.30681	0.28260	0.24372	0.22939	0.22244	0.17652	0.1731
## Proportion of Variance	0.00314	0.00266	0.00198	0.00175	0.00165	0.00104	0.0010
## Cumulative Proportion	0.98649	0.98915	0.99113	0.99288	0.99453	0.99557	0.9966

```
##
```

	PC22	PC23	PC24	PC25	PC26	PC27	PC28
## Standard deviation	0.16565	0.15602	0.1344	0.12442	0.09043	0.08307	0.03987
## Proportion of Variance	0.00091	0.00081	0.0006	0.00052	0.00027	0.00023	0.00005
## Cumulative Proportion	0.99749	0.99830	0.9989	0.99942	0.99969	0.99992	0.99997

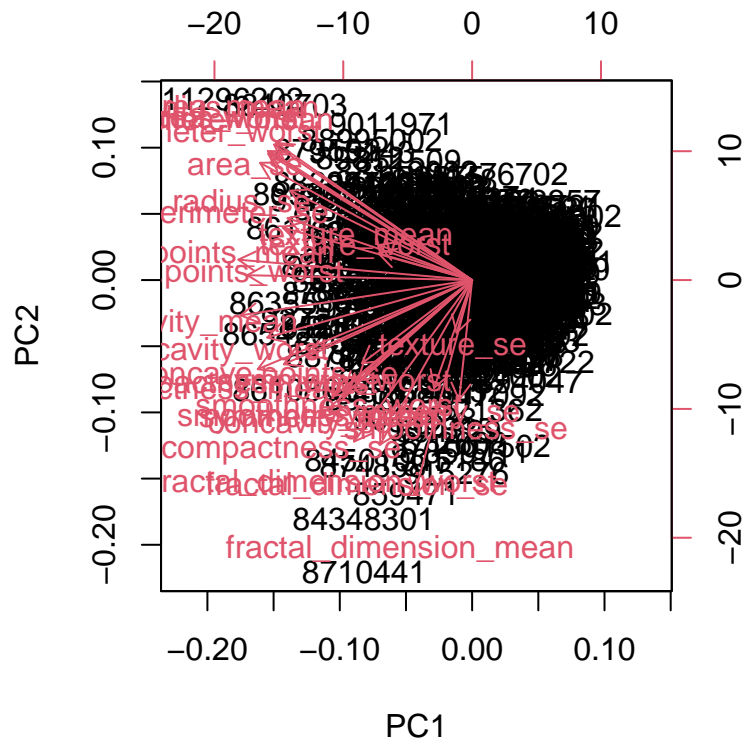
```
##
```

	PC29	PC30
## Standard deviation	0.02736	0.01153
## Proportion of Variance	0.00002	0.00000
## Cumulative Proportion	1.00000	1.00000

The more PCs you need to describe the data, the more all-over-the-place it is...

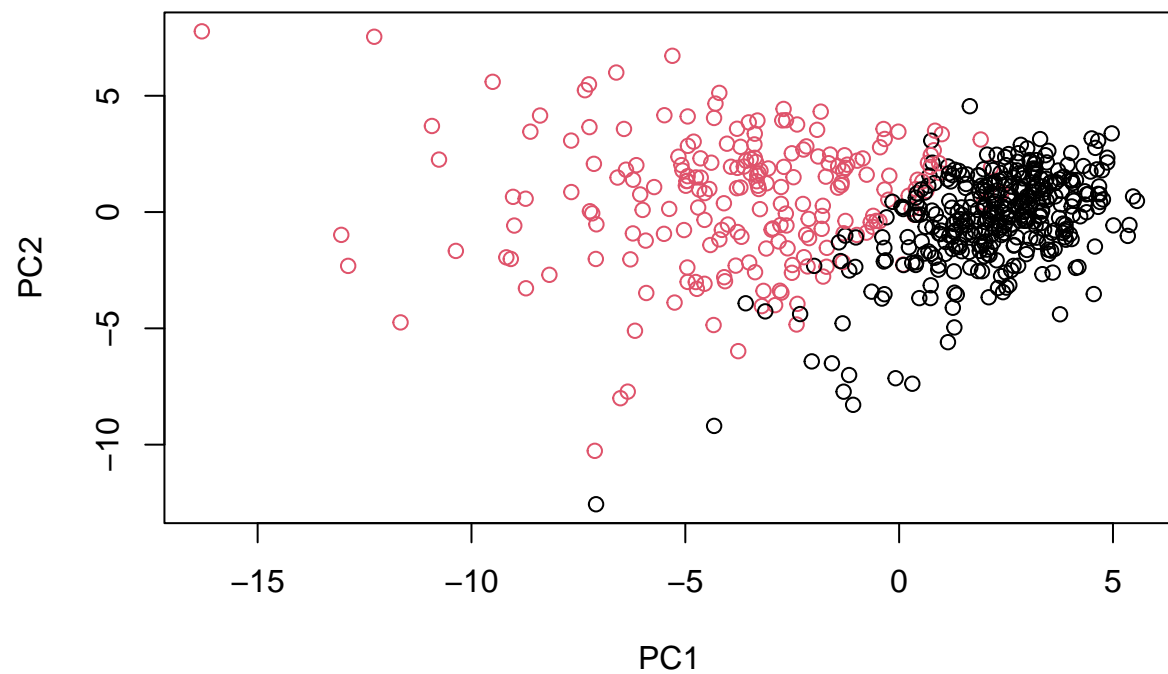
#Q4. The first principal component captures 44% of the variance in the data. #Q5. At least 3 PCs to describe >70% of the variance. #Q6. At least 7 PCs to describe >90% of the variance.

```
# Visualize PCA Results
biplot(wisc.pr)
```

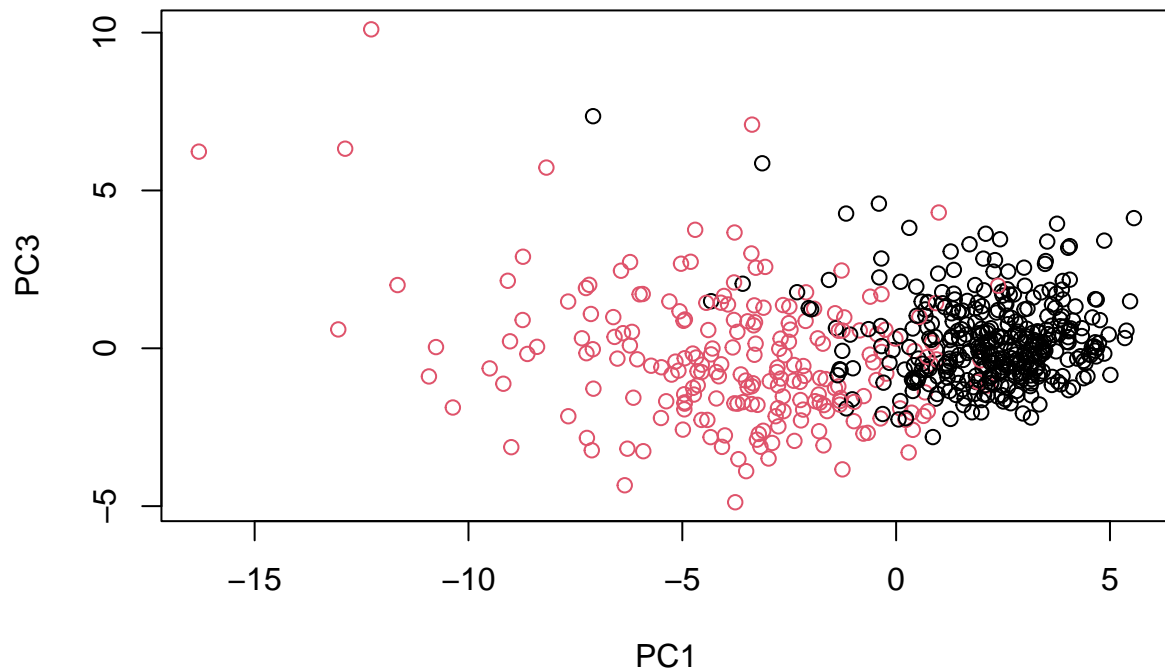


#Q7. The plot contains a vast amount of data, but there seems to be an overall leftward trend. The plot is not at all easily interpretable.

```
# Visualize PCA Results, but better
factor_diagnosis <- as.factor(diagnosis)
plot(wisc.pr$x[,1:2] , col=factor_diagnosis)
```



```
plot(wisc.pr$x[,1], wisc.pr$x[,3], col=factor_diagnosis,  
     xlab="PC1", ylab="PC3")
```



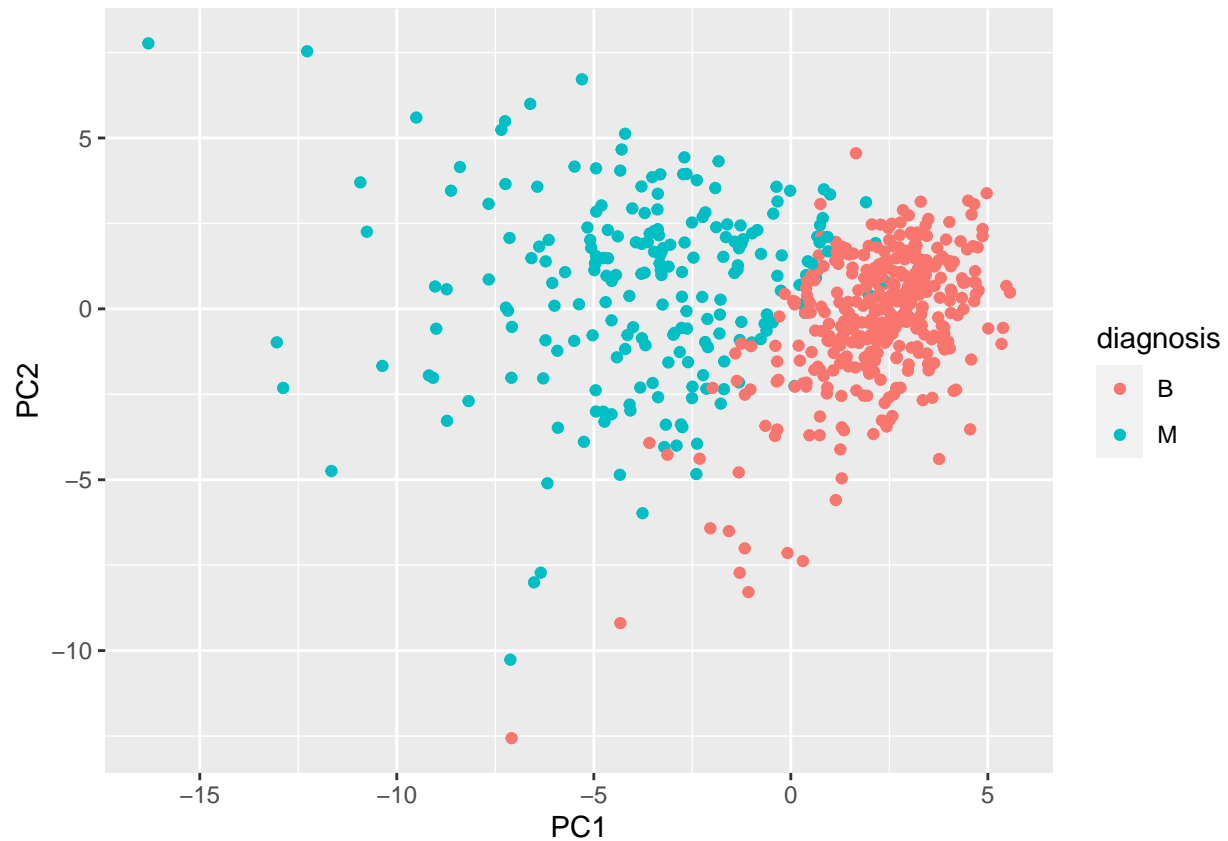
#Q8. The plots are largely similar. The only notable differences are that the P1/P2 plot seems to have negative outliers, whereas the P1/P3 plot has positive ones. These outliers change the y-scale of the plot, and the separation seems to be a little clearer in the P1/P2 plot, as the black values in the P1/P3 plot cross far into the red cluster territory.

```
# Visualize data in ggplot

# Create a data.frame for ggplot
df <- as.data.frame(wisc.pr$x)
df$diagnosis <- factor_diagnosis

# Load the ggplot2 package
library(ggplot2)

# Make a scatter plot colored by diagnosis
ggplot(df) +
  aes(PC1, PC2, col=diagnosis) +
  geom_point()
```



```
# Visualize Variance Capturing by PCs
```

```
# Calculate variance from standard deviation
```

```
pr.var <- wisc.pr$sdev^2
```

```
pr.var
```

```
## [1] 1.328161e+01 5.691355e+00 2.817949e+00 1.980640e+00 1.648731e+00
```

```
## [6] 1.207357e+00 6.752201e-01 4.766171e-01 4.168948e-01 3.506935e-01
```

```
## [11] 2.939157e-01 2.611614e-01 2.413575e-01 1.570097e-01 9.413497e-02
```

```
## [16] 7.986280e-02 5.939904e-02 5.261878e-02 4.947759e-02 3.115940e-02
```

```
## [21] 2.997289e-02 2.743940e-02 2.434084e-02 1.805501e-02 1.548127e-02
```

```
## [26] 8.177640e-03 6.900464e-03 1.589338e-03 7.488031e-04 1.330448e-04
```

```
# Variance explained by each principal component: pve
```

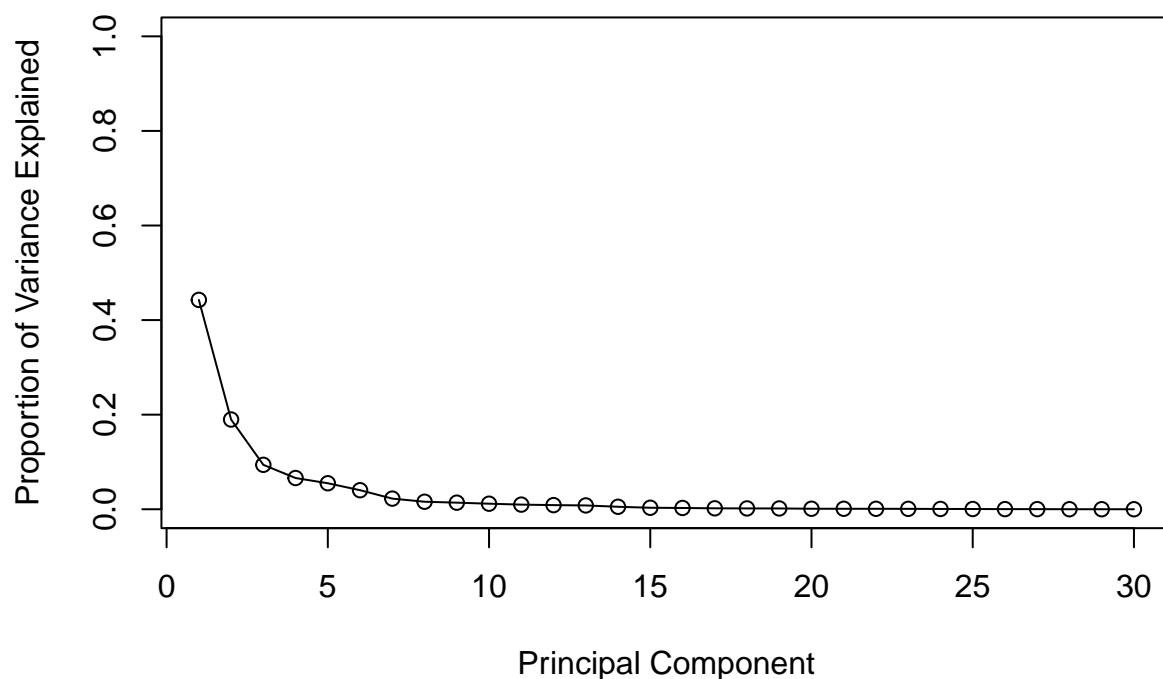
```
pve <- pr.var / sum(pr.var)
```

```
# Plot variance explained for each principal component
```

```
plot(pve, xlab = "Principal Component",
```

```
      ylab = "Proportion of Variance Explained",
```

```
      ylim = c(0, 1), type = "o")
```



```
# Communicating PCA Results
wisc.pr$rotation["concave.points_mean",1]
```

```
## [1] -0.2608538
```

#Q9. The component of the loading vector for “concave.points_mean” is -0.26. #Q10. At least 5 PCs to describe >80% of the variance.

Hierarchical Clustering.

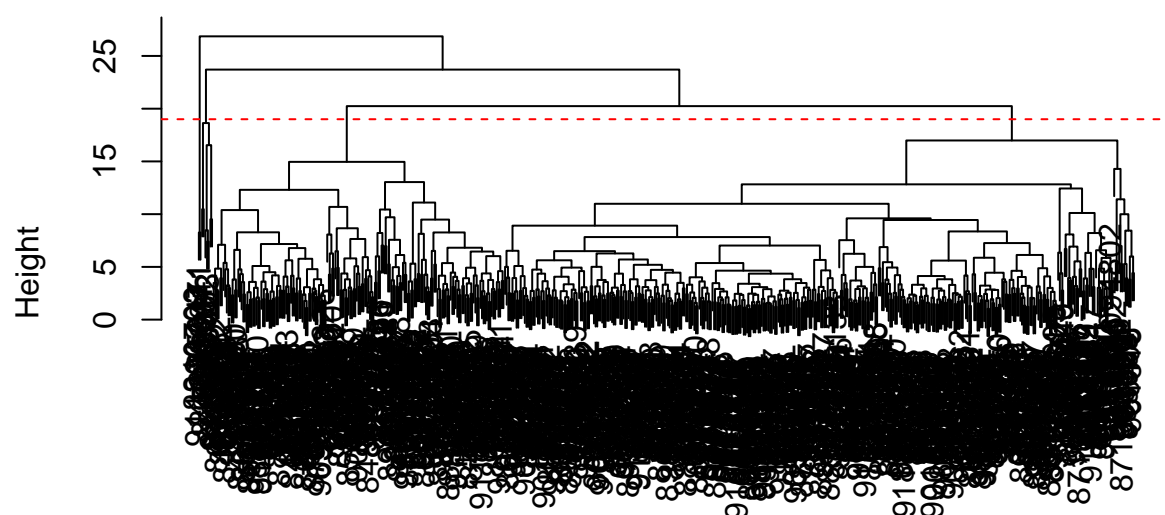
```
# Try 'complete' clustering method

# Calculate scaled Euclidean distances of data points
wisc.dist <- dist(scale(wisc.data))

# Cluster w/ 'hclust()'
wisc.hclust <- hclust(wisc.dist, method="complete")

# Visualize
plot(wisc.hclust)
abline(h=19, col="red", lty=2)
```


Cluster Dendrogram



```
wisc.dist
hclust (*, "complete")
```

#Q11. The abline must be placed at h=19 to cut the tree into 4 groups.

```
# Extract Data
ans <- NULL
for(i in 2:10) {
  x <- cutree(wisc.hclust, k=i)
  ans <- rbind(ans, x)
}
#ans

for(i in 1:9) {
  print(table(ans[i,], diagnosis))
}
```

```
##      diagnosis
##      B  M
##  1 357 210
##  2   0   2
##      diagnosis
##      B  M
##  1 355 205
##  2   2   5
##  3   0   2
##      diagnosis
##      B  M
##  1  12 165
```

```

## 2 2 5
## 3 343 40
## 4 0 2
## diagnosis
## B M
## 1 12 165
## 2 0 5
## 3 343 40
## 4 2 0
## 5 0 2
## diagnosis
## B M
## 1 12 165
## 2 0 5
## 3 331 39
## 4 2 0
## 5 12 1
## 6 0 2
## diagnosis
## B M
## 1 12 165
## 2 0 3
## 3 331 39
## 4 2 0
## 5 12 1
## 6 0 2
## 7 0 2
## diagnosis
## B M
## 1 12 86
## 2 0 79
## 3 0 3
## 4 331 39
## 5 2 0
## 6 12 1
## 7 0 2
## 8 0 2
## diagnosis
## B M
## 1 12 86
## 2 0 79
## 3 0 3
## 4 331 39
## 5 2 0
## 6 12 0
## 7 0 2
## 8 0 2
## 9 0 1
## diagnosis
## B M
## 1 12 86
## 2 0 59
## 3 0 3
## 4 331 39

```

```
## 5 0 20
## 6 2 0
## 7 12 0
## 8 0 2
## 9 0 2
## 10 0 1
```

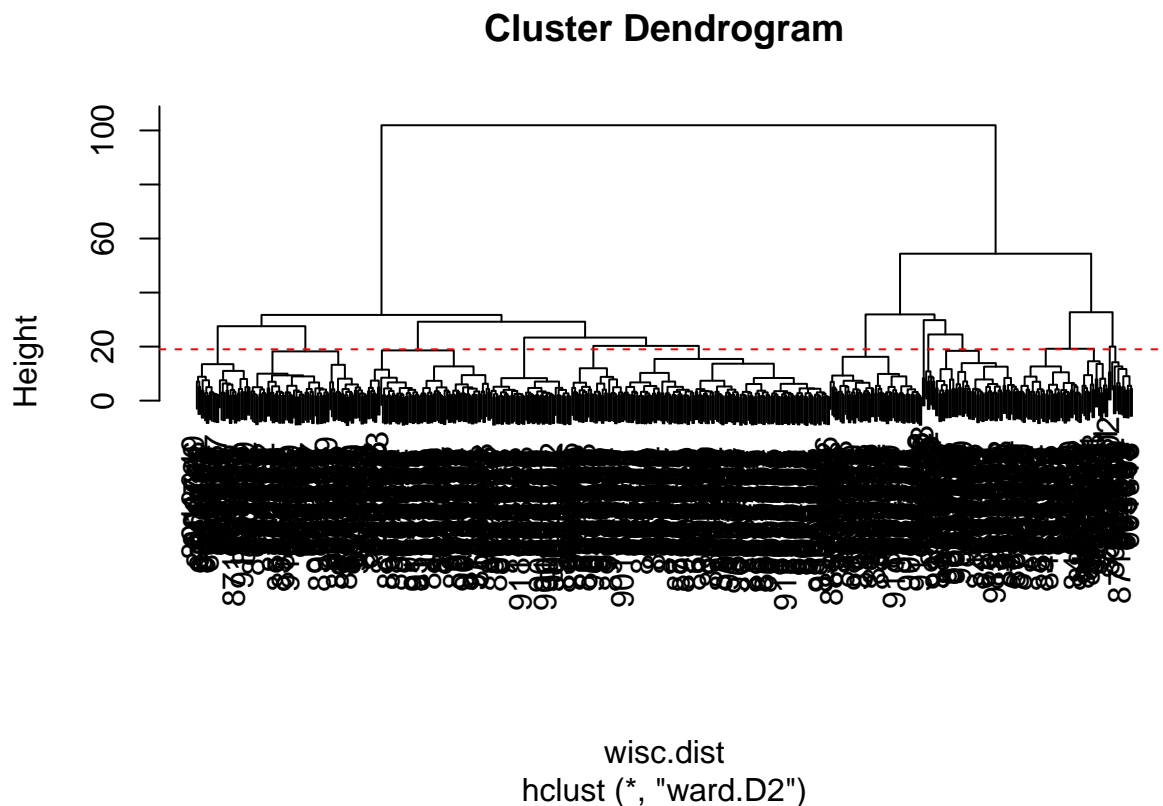
#Q12. No, cutting the tree into any less groups ignores the split between the two large groups of patients, and cutting it into any more groups than 4 unnecessarily divides groups.

```
# Try 'ward.D2' method

# Calculate scaled Euclidean distances of data points
wisc.dist <- dist(scale(wisc.data))

# Cluster w/ 'hclust()'
wisc.hclust <- hclust(wisc.dist, method="ward.D2")

# Visualize
plot(wisc.hclust)
abline(h=19, col="red", lty=2)
```



#Q13. Neither the 'single' nor the 'average' method come up with any kind of clustering that separates the malignant and benign groups, but I don't think I can pick a favorite between the 'complete' and 'ward.D2' methods. While 'ward.D2' returns satisfactory results with separation into just two clusters, when 'complete' works, it separates more accurately (less erroneous results) than 'ward.D2'.

```
# See if k-means clustering gives same result

wisc.km <- kmeans(wisc.data, centers= 2, nstart= 20)

# Check result with table
table(wisc.km$cluster, diagnosis) #kmeans clustering result
```

```
##      diagnosis
##           B    M
##    1     1 130
##    2    356  82
```

```
table(ans [3,], diagnosis) #hclust result
```

```
##      diagnosis
##           B    M
##    1     12 165
##    2      2   5
##    3    343  40
##    4      0   2
```

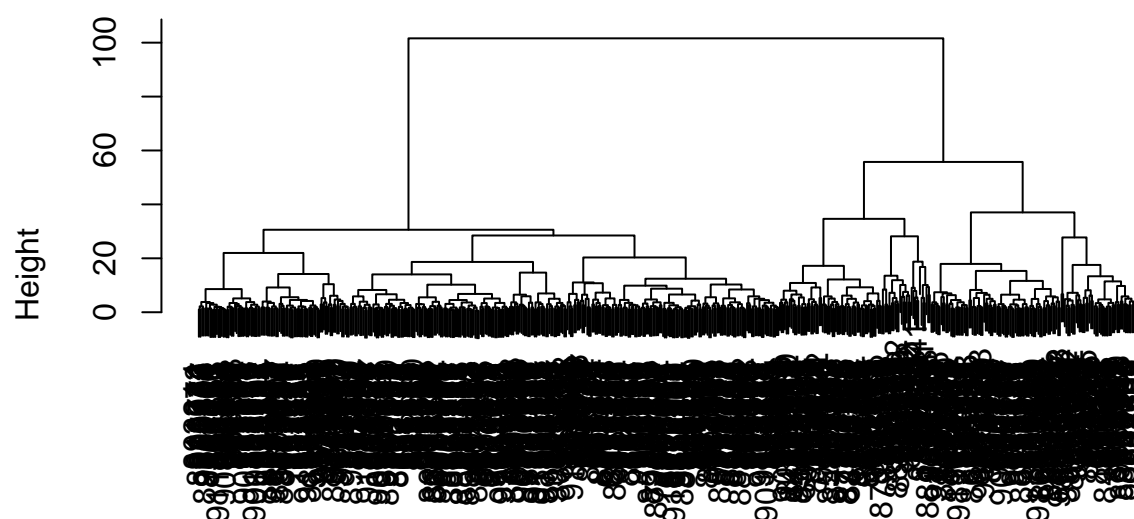
#Q14. K-means satisfactorily separates the diagnoses. It is not as accurate as hclust, but it is close behind. There are clearly two groups

Combining Methods

```
# Apply hierarchical clustering to PCA Analysis
wisc.pr.hclust <- hclust(dist(wisc.pr$x[,1:7]), method="ward.D2")

# Visualize
plot(wisc.pr.hclust)
```

Cluster Dendrogram



```
dist(wisc.pr$x[, 1:7])
hclust (*, "ward.D2")
```

```
# Split up the tree into groups
wisc.pr.hclust.cut <- cutree(wisc.pr.hclust, k=2)
table(wisc.pr.hclust.cut, diagnosis)
```

```
##           diagnosis
## wisc.pr.hclust.cut  B  M
##           1  28 188
##           2 329  24
```

#Q15. This new model has an error rate of 9%. It is a slight improvement over the hierarchical clustering we did on the raw data. (52 errors now, over 61 errors previously).

```
# Compare all Results

## K-means Clustering
table(wisc.km$cluster, diagnosis)
```

```
##      diagnosis
##        B  M
##      1   1 130
##      2 356  82
```

```
## Hierarchical Clustering on Data
table(ans [3,], diagnosis)
```

```
##      diagnosis
##      B    M
##    1 12 165
##    2  2   5
##    3 343  40
##    4  0   2
```

```
## Hierarchical Clustering on PCA Analysis
table(wisc.pr.hclust.cut, diagnosis)
```

```
##              diagnosis
## wisc.pr.hclust.cut  B    M
##              1  28 188
##              2 329  24
```

#Q16. Accuracy-wise overall, hierarchical clustering on PCA Analysis takes the win. It also identifies the lowest number of false negatives, which makes this the safest method for its applicaiton.

Sensitivity/ Specificity

#Q17. Total Malignant: 212, Total Benign: 357

K-means Clustering

Sensitivity: $130/212 = 61.3\%$

Specificity: $356/357 = 99.7\%$

Hierarchical Clustering

Sensitivity: $165/212 = 77.8\%$

Specificity: $343/357 = 96.1\%$

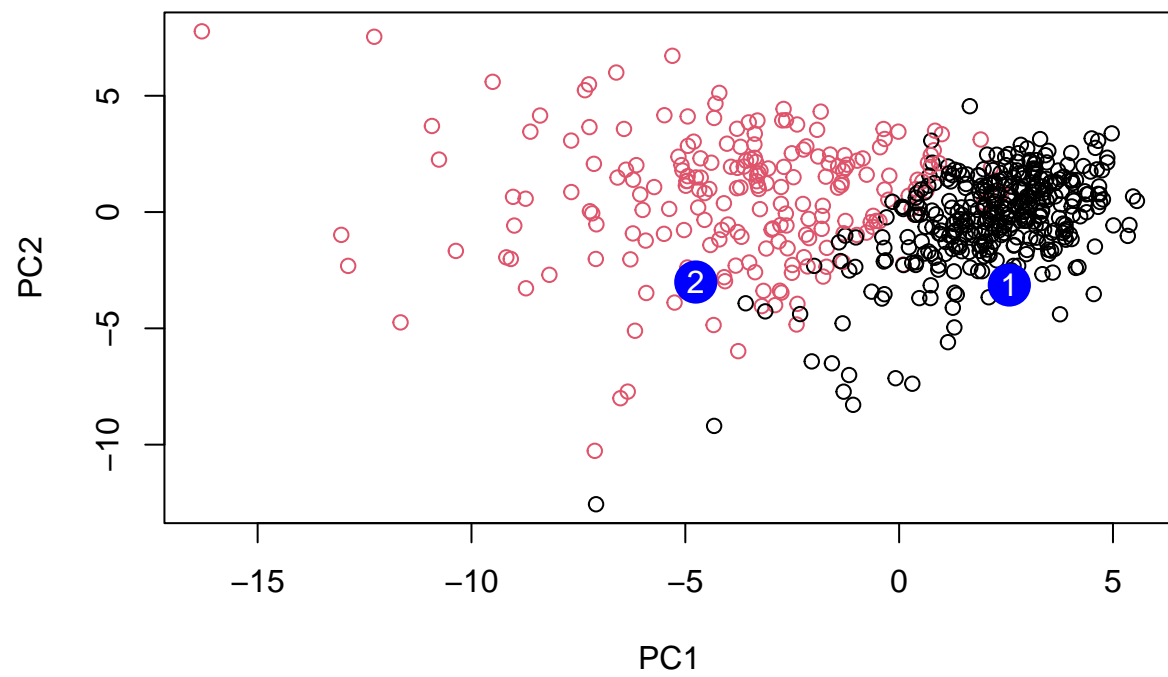
Hierarchical Clustering on Principal Components

Sensitivity: $188/212 = 88.7\%$

Specificity: $329/357 = 92.2\%$

```
# Import New Data Points
new <- read.csv("new_samples.csv")
npc <- predict(wisc.pr, newdata=new)

# Plot Old Data with Two New Points Overlay
plot(wisc.pr$x[,1:2], col=factor_diagnosis)
points(npc[,1], npc[,2], col="blue", pch=16, cex=3)
text(npc[,1], npc[,2], c(1,2), col="white")
```



#Q18. Patient 2 should be prioritized as they probably have the malignant tumor.