

On the inference of complex phylogenetic networks with SnappNet

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Roadmap

- 1 Introduction
- 2 Species tree inference with the SNAPP method
- 3 Network inference
 - Our new method SNAPPNET
 - Algorithm
 - BEAST (Beauti)
 - Comparison SNAPPNET vs MCMCBiMarker
 - Rice real data
- 4 Conclusion

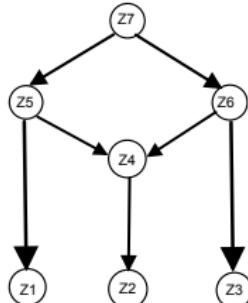
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Phylogenetic networks

Phylogenetic networks are Directed Acyclic Graphs (DAG) that allow us to detect :

- hybridizations (e.g. plants)
- introgressions (e.g. plants and animals)
- horizontal gene transfer (e.g. bacteria)

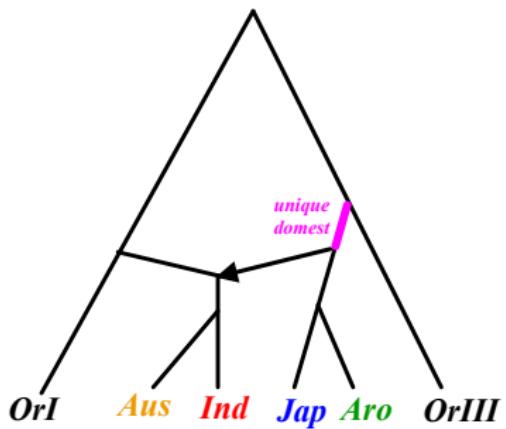


Key points :

- Edge length = evolutionary time
- Dependencies between nodes
- Reticulation nodes have 2 parents and represent reticulation events
- Our goal is to obtain a distribution of phylogenetic networks (uncertainty on clades)

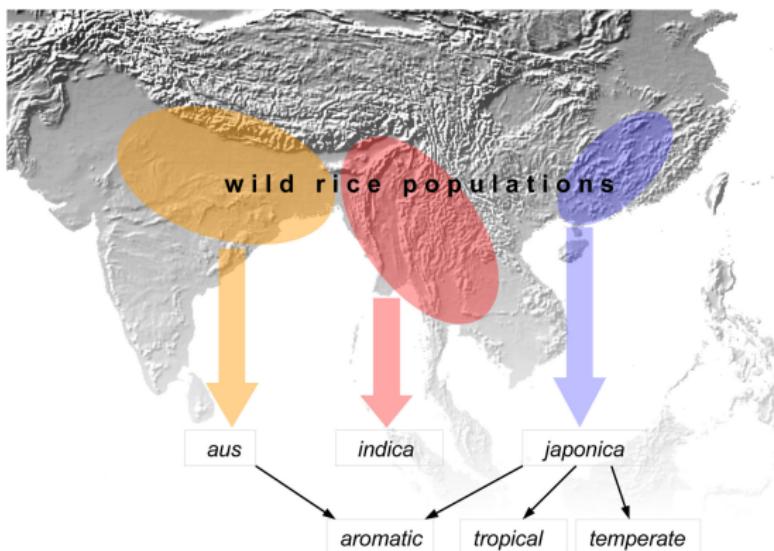
A few scenarios on the rice domestication process

- Huang et al. (Nature, 2012) : *japonica* domesticated from a wild form in the south of China, and crossed to a wild form in the south East of Asia, generating *indica*



A few scenarios on the rice domestication process

- Civan et al. (Nature Plants, 2015) : *indica*, *japonica* and *aus* domesticated separately in different locations in Asia

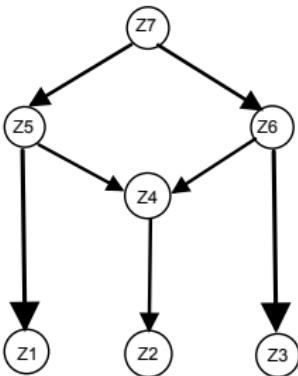


Our methodological approach

We consider a model, that handles incomplete lineage sorting, and that considers explicitly **mutations and hybridization**.

⇒ **Phylogenetic network inference** in a rich Bayesian framework

SNAPPNET = Generalization of **SNAPP** (Bryant et al. 2012) to networks

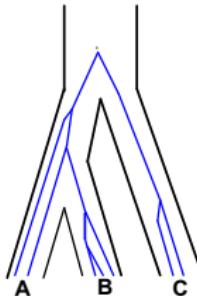


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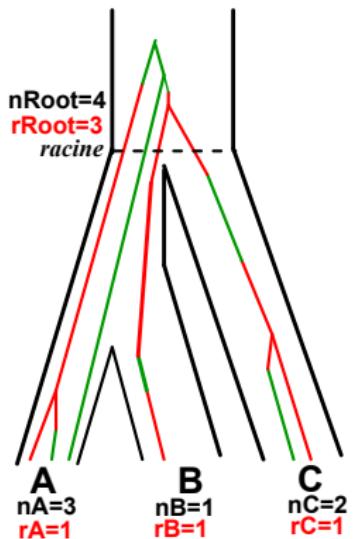
SNAPP software for the Bayesian inference of species trees (Bryant et al. 2012, MBE)

- Independent biallelic markers (SNPs) given the species tree
- Locus tree model (backward)
 - Coalescent process evolving inside a species tree (**MultiSpecies Coalescent**)
 - Process that allows the discordance between locus trees and species trees (**incomplete lineage sorting**)



Mutations happen over time

- SNP data model (forward)
 - mutation (**red** ↔ **green**) : markov model evolving along the locus tree branches
 - u : mutation rate **red** → **green**
 - v : mutation rate **green** → **red**



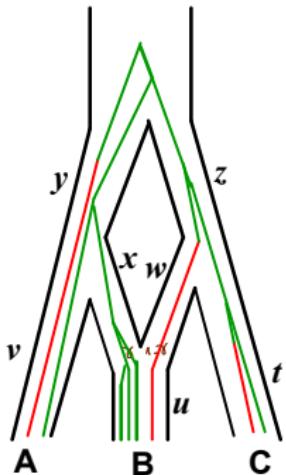
- Random variables : **rRoot**, **nRoot**, **rIntNode**, **nIntNode**, **rA**, **rB**, **rC**
- nA**, **nB**, **nC** are not random
- Data**=(**rA**, **rB**, **rC**)
- Likelihood : $\mathbb{P}(\text{Data} | S)$ with **S** species tree

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Network context

- Locus tree model (backward) :
 - Coalescent process
 - Nakhleh's model at the reticulation node
⇒ Multispecies Network Coalescent
- SNP data model (forward)



- Random Variable : **rRoot**, **nRoot**, **rIntNode**, **nIntNode**, **rA**, **rB**, **rC**
- **nA**, **nB**, **nC** are not random
- **Data=(rA, rB, rC)**
- Likelihood : $\mathbb{P}(\text{Data} | N)$ with N network

SNAPPNet : a new Bayesian method for inferring networks

- N : phylogenetic network (topology, branch lengths, population sizes, inheritance probability)
- X_i : data for locus i
- G_i : locus tree for locus i
- m loci

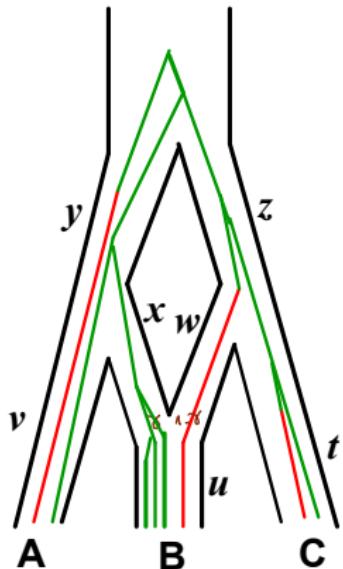
$$\begin{aligned}\mathbb{P}(N|X_1, \dots, X_m) &\propto \left(\prod_{i=1}^m \int_{\psi} \mathbb{P}(X_i|G_i)\mathbb{P}(G_i|S)dG_i \right) P(N) \\ &\propto \mathbb{P}(\text{Data} | N) P(N)\end{aligned}$$

SNAPPNET integrates over all locus trees (generalization of SNAPP, Bryant et al. MBE 2012), using new algorithms dedicated to networks

Computation of the prior $P(N)$ by the birth hybridization process of Zhang et al. (MBE 2018)

⇒ Markov Chain Monte Carlo (MCMC) in order to sample from the posterior distribution $\mathbb{P}(N|X_1, \dots, X_m)$

Specificities of phylogenetic networks

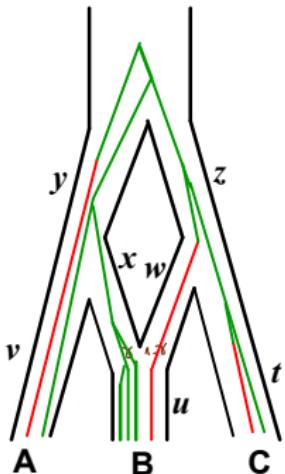


Data_z : red/green percentages
in species below branch z
Data_y : red/green percentages
in species below branch y

Data_{z^T} and *Data_{y^T}* are not independent ...

Data_{z^T} and *Data_{y^T}* contain the red and green alleles of the hybrid species

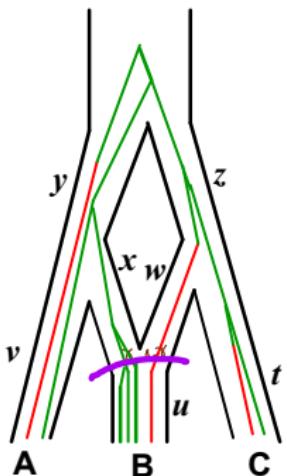
Our algorithm based on joint distributions



Quantities computed consecutively

- (1) $\mathbb{P}(\text{Data}_{uT} | n_{uT}, r_{uT})$
- (2) $\mathbb{P}(\text{Data}_{xB} \text{Data}_{wB} | n_{xB}, r_{xB}, n_{wB}, r_{wB})$
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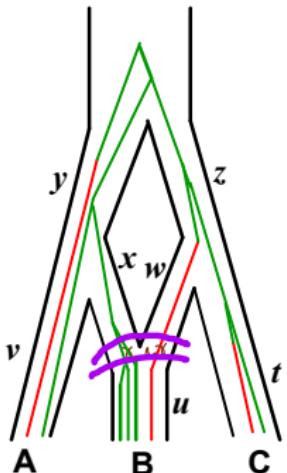
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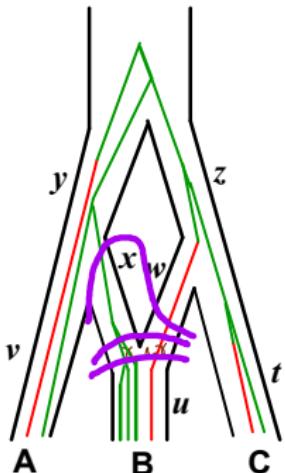
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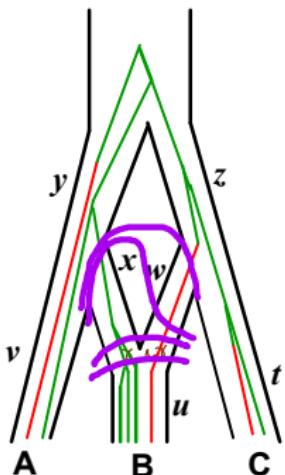
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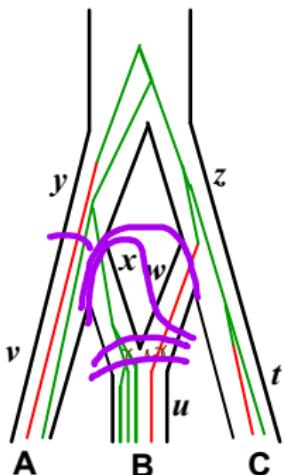
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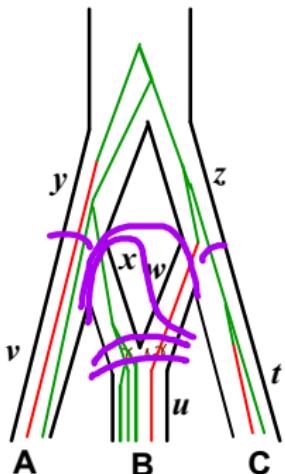
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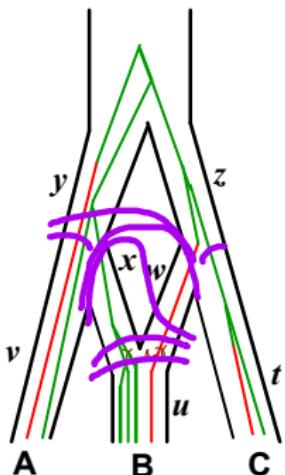
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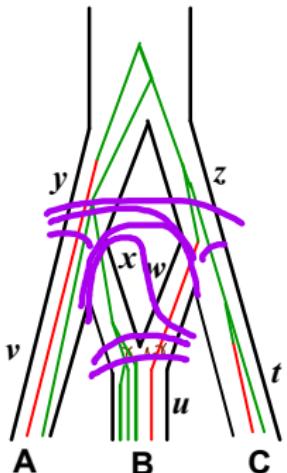
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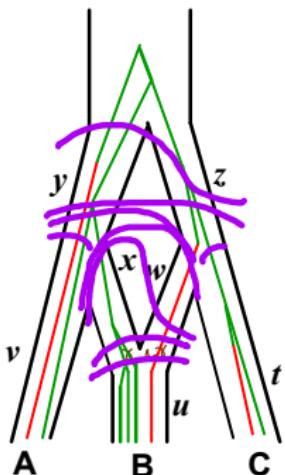
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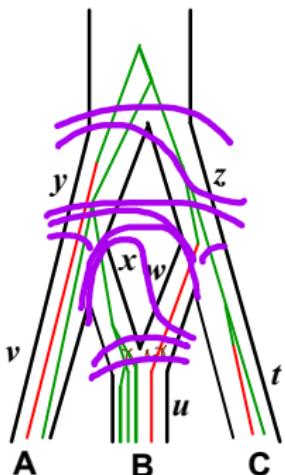
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About SNAPPNET's xml (Add On for Beast)

```
<distribution id="networkPrior"
spec="speciesnetwork.BirthHybridizationModel"
network="@network :species" netDiversification="@netDivRate :species"
turnOver="@turnOverRate :species"/>
<prior id="netDivPrior" name="distribution" x="@netDivRate :species">
<Exponential id="exponential.01" name="distr" mean="10.0"/>
</prior>
<prior id="turnOverPrior" name="distribution" x="@turnOverRate :species">
<Beta id="betadistr.01" name="distr" alpha="1.0" beta="1.0"/>
</prior>
```

How to get SNAPPNET's xml :

BEAUti — Bayesian Evolutionary Analysis Utility.

This program is used to import data, design the analysis, and generate the BEAST control file.



BEAUti : how to choose the number of reticulations

The screenshot shows the BEAUTI 2 interface with the 'Operators' tab active. On the left, a list of operators is shown, many of which have edit icons next to them. The 'addReticulation:species' operator is currently being edited, as indicated by the modal dialog box in the center. This dialog box contains fields for 'Coalescence Rate' (set to 0.01), 'maxReticulationNumber' (set to 3), and 'Weight' (set to 10.0). It also includes checkboxes for 'Sample' and 'Bound the number of reticulations'. A blue lizard icon is displayed on the left side of the dialog. The background shows the main window with other operators listed.

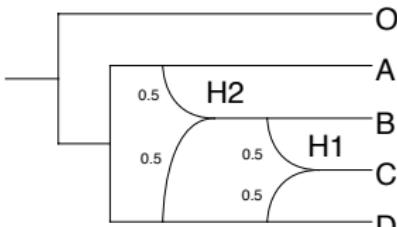
SNAPPNET available at <https://github.com/rabier/MySnappNet>

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Time required to compute the likelihood $\mathbb{P}(\text{Data} | N)$

Dataset ID	CPU time	
	SNAPPNET (in minutes)	MCMCBiMarkers (in hours)
1	5.559	35.9354
2	5.6763	34.2433
3	5.7351	32.6519
4	5.446	34.2011
5	5.5996	33.2354



Network C of level 2

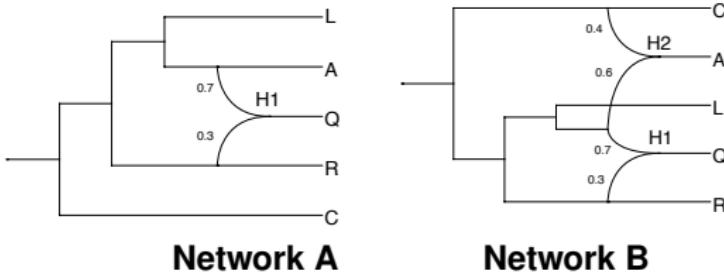
SNAPPNET vs MCMCBiMarkers

(Zhu et al., Plos Comp Biol 2018)

$O(n^8)$ vs $O(n^{12})$

SNAPPNET's ability to recover networks A and B taken from Zhu et al (Plos Comput Biol, 2018)

Number of sites	1,000	10,000	100,000
Network A	0%	100%	100%
Network B	0%	81.25%	100%

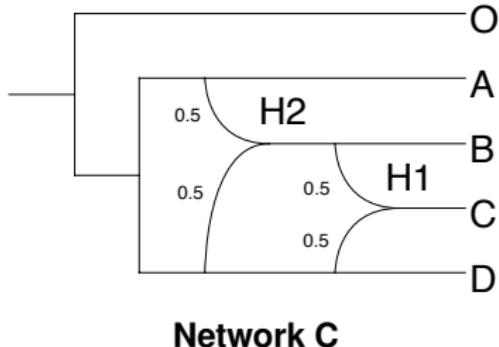


- 1000 sites : MCMCBiMarkers > SNAPPNET
- ≥ 10000 sites : SNAPPNET \approx MCMCBiMarkers
- 10 000 sites are required to infer these networks

Ability to recover the topology of network C

SNAPPNET vs MCMCBiMarkers

Number of lineages for B and for C	Number of sites		
	1,000	10,000	100,000
1	0%	7.87%	54.90%
	0%	4.84%	0%
4	0%	50.00%	49.60%
	0%	0%	0%

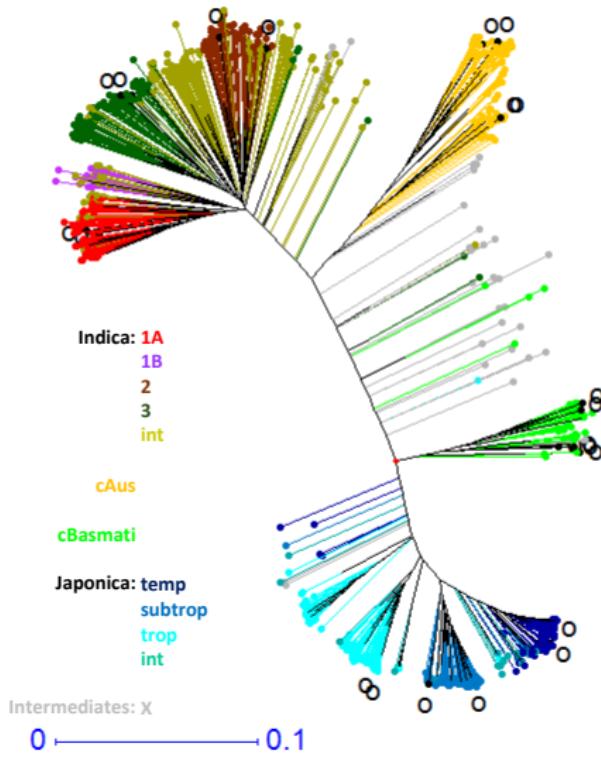


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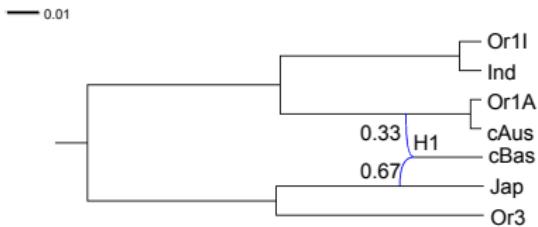
Varieties selected by J.C. Glaszmann

Neighbour joining tree based on Wang et al. (Nature, 2018), 3000 rice varieties (4.8 millions of SNPs)

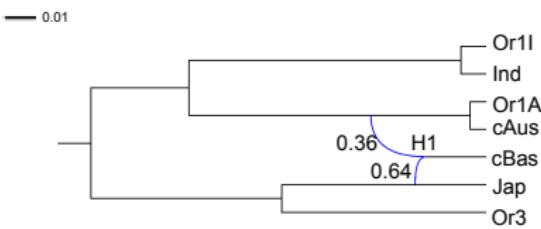


SNAPPNET in the Bayesian MCMC setting

- 2 different samplings of 10K SNPs
- 2 Markov chains per sampling
- Number of reticulations bounded by 2
- 10 millions iterations



First sampling
ESS=844, ESS=1159



Second sampling
ESS=971, ESS=535

The cultivars are associated with the expected wild forms

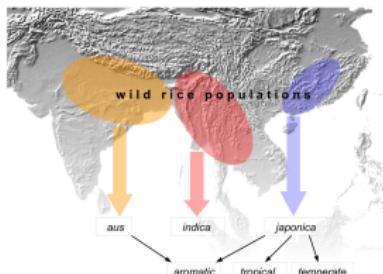
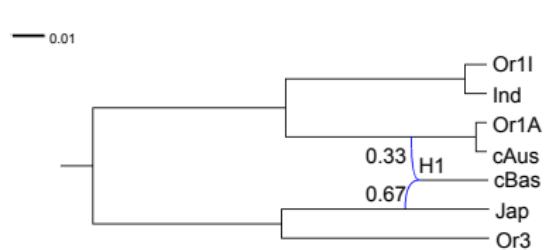
Summary of the different analyses

The inferred networks reveal stable features :

- correspondence between wild subpopulations and cultivated subpopulations
- the early divergence of Japonica, that predates the one between Indica and cAus
- the mobilisation of early Japonica cultivars to combine with the cAus pillar to produce the fourth varietal type cBas
- the indication that this hybridization may have occurred before the domestication of cAus

There is an agreement with Civan's thesis, but we observe a few differences

...



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Conclusion

- SNAPPNET available at <https://github.com/rabier/MySnappNet>
- SNAPPNET implemented within the BEAST 2 framework
- Paper : “On the inference of complex phylogenetic networks by Markov Chain Monte-Carlo”, Rabier et al. (PLoS Comput Biol, 2021)
- The gain in speed allows us to consider more complex evolutionary scenarios
- It is also possible to evaluate the likelihoods of evolutionary scenarios

Acknowledgements

Céline Scornavacca
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João D. Santos



Jean-Michel Marin



Angélique D'Hont
Manuel Ruiz

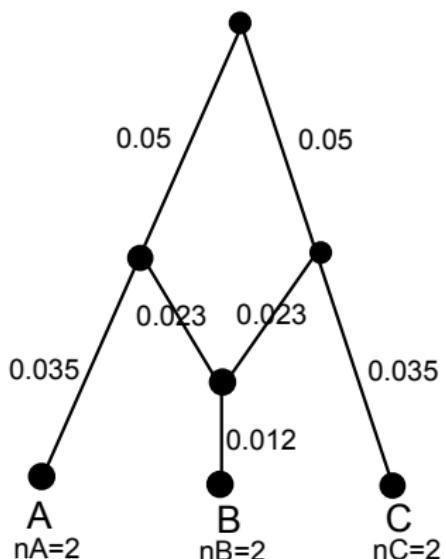


Marilyne Summo

References

- Huang et al (Nature, 2012). “A map of rice genome variation reveals the origin of cultivated rice”
- Civan et al (Nature plants, 2015). “Three geographically separate domestications of Asian rice”
- Wang et al (Nature, 2018). “Genomic variation in 3,010 diverse accessions of Asian cultivated rice”
- Bryant et al. (MBE, 2012). “Inferring species trees directly from biallelic genetic markers : bypassing gene trees in a full coalescent analysis”
- Zhang et al. (MBE, 2017). “Bayesian inference of species networks from multilocus sequence data”
- Zhu et al. (PLoS Comput Biol, 2018). “Bayesian inference of phylogenetic networks from bi-allelic genetic markers”

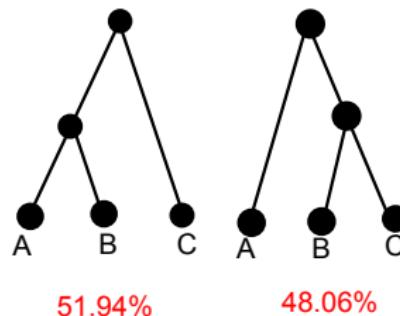
An example on simulated data



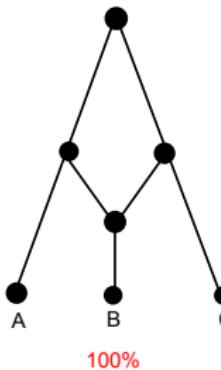
- Branch lengths in expected number of mutations per site
- $nA=2$, $nB=2$, $nC=2$
- 1 000 sites or 10 000 sites
- Population sizes θ equal to 0.005 or 0.05
- T : coalescent time for 2 lineages (in mutations par site)
 - if $\theta = 0.005$, then $\mathbb{E}(T) = 0.005/2 = 0.0025$
 - if $\theta = 0.05$, then $\mathbb{E}(T) = 0.005/2 = 0.025$

Networks sampled by MCMC

- 1 000 sites, $\theta = 0.005$



- 10 000 sites, $\theta = 0.005$
- 1 000 sites, $\theta = 0.05$
- 10 000 sites, $\theta = 0.05$

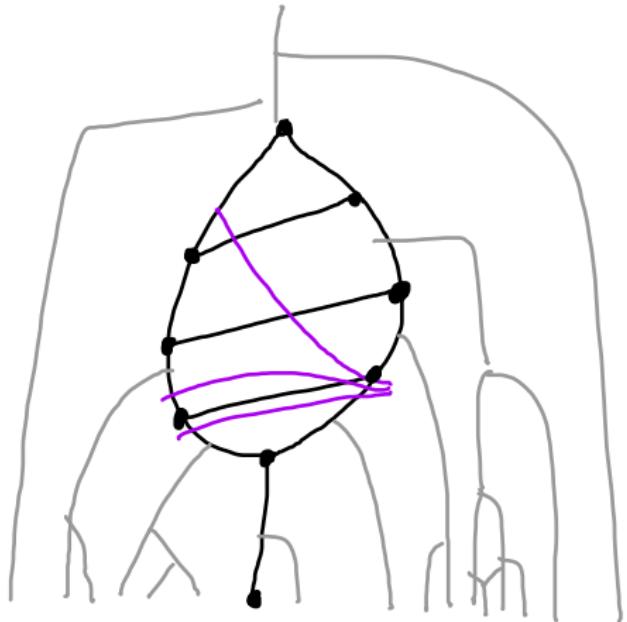


When the population size is larger,
we need more sites to recover the network

We try to minimize the number of branches simultaneously considered in our joint distributions

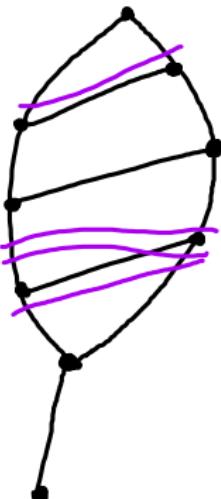
Strategy to avoid

A maximum of 5 branches



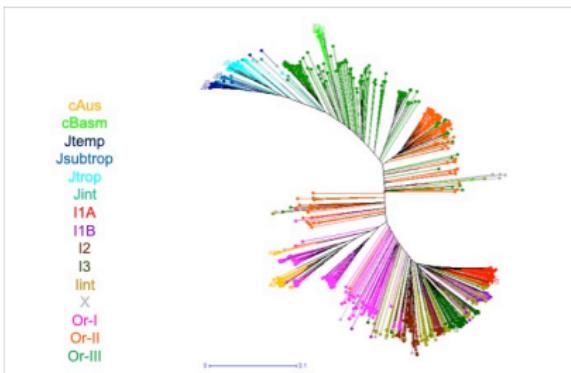
Strategy to adopt

A maximum of 3 branches



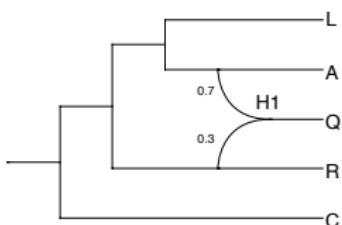
Wild forms added to our datasets

- W1559 : Or1 (Or1I), Thailand, close to Indica
- W1117 : Or1 (Or1I), India, within Indica
- W1747 : Or1 (Or1A), India, within cAus
- W0574 : Or1 (Or1A), Malaysia, close to cAus
- W3042 : Or3, China, within Japonica
- W3048 : Or3, China, between Japonica and cBasmati
- W3073 : Or3, China, other side of cBasmati

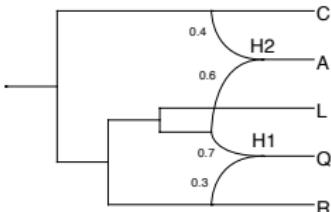


NJ tree (Glaszmann, Wang)

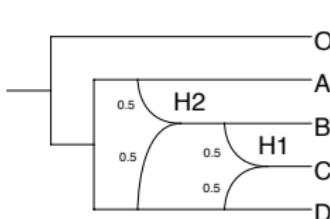
3 networks studied by simulation



Network A



Network B



Network C

Networks A and B taken from Zhu et al (Plos Comput Biol, 2018)

MCMCBiMarkers vs SNAPPNET

- Network A : $O(n^8)$ vs $O(n^6)$
- Network B and C : $O(n^{12})$ vs $O(n^8)$